

# Hsa\_Circ\_0001947/MiR-661/DOK7 Axis Restrains Non-Small Cell Lung Cancer Development

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Hsa\_circ\_0001947 is associated with multiple cancers, but its function in non-small cell lung cancer (NSCLC) is ambiguous and needs further research. The targeting relationship among circ\_0001947, miR-661, and downstream of tyrosine kinase 7 (DOK7) was predicted by database and further verified by dual-luciferase reporter assay, while their expressions in cancer tissues and cells were detected by quantitative real-time polymerase chain reaction (qRT-PCR). After transfection, cell biological behaviors and expressions of miRNAs, miR-661 and DOK7 were determined by cell function experiments and qRT-PCR, respectively. Circ\_0001947 was low-expressed in NSCLC tissues and cells. Circ\_0001947 knockdown intensified cell viability and proliferation, induced cell cycle arrest at S phase, suppressed apoptosis and evidently enhanced miR-510, miR-587, miR-661 and miR-942 levels, while circ\_0001947 overexpression did the opposite. MiR-661 was a target gene of circ\_0001947 that participated in the regulation of circ\_0001947 on cell biological behaviors. Furthermore, DOK7, the target gene of miR-661, partly participated in the regulation of miR-661 on cell viability. Hsa\_circ\_0001947 acts as a sponge of miR-661 to repress NSCLC development by elevating the expression of DOK7.

Keywords: Non-small cell lung cancer, hsa\_circ\_0001947, miR-661, downstream of tyrosine kinase 7

## Introduction

Lung cancer is a prevalent malignancy in the world. According to statistics, there were approximately 2.1 million new cases and 1.8 million deaths worldwide in 2018, ranking first among all cancer types [1]. Non-small cell lung cancer (NSCLC) is the most common histopathological subtype of lung cancer, accounting for 80-90% of lung cancer [2]. NSCLC is generally diagnosed at an advanced stage, and 90% of patients die from distant metastasis [3]. The five-year survival rate of patients with NSCLC could be improved to 50%-70% with early diagnosis and treatment prior to metastasis of the disease, otherwise the rate may be less than 5% [4]. In addition, the lack of relatively specific tumor markers poses challenges to the diagnosis, treatment, and prognosis of NSCLC. Therefore, it is extremely important and necessary to find avenues that are conducive to early diagnosis and treatment of NSCLC.

CircRNA is a special endogenous non-coding RNA that has long been found in RNA viruses [5]. However, due to technical limitations at that time, circRNA was considered a by-product of the splicing process and thus received little interest [6]. In recent years, with the development of sequencing technology, circRNA has been found to be widely presented and stably expressed in eukaryotic cells, playing a pivotal role in the regulation of gene expression in human cells [7]. Current researches pointed out that circRNA may serve as a molecular marker for prognostic judgment of different tumors including NSCLC [8, 9]. Studies have manifested that some circRNAs can participate in the pathogenesis of NSCLC as oncogenes or tumor suppressor genes, such as circ\_BANP, circ\_100395, and circ\_0001947 [10-12]. Among them, only one research proposed that circ\_0001947 may be closely associated with the presence and progression of NSCLC [13]. However, the exact role of circ\_0001947 in NSCLC has not been reported yet.

Moreover, it has been put forward that multiple circRNAs themselves contain at least one miRNA binding site, which can act as the "sponge" of RNA to adsorb miRNA, thus regulating miRNA-induced inhibition of downstream target genes expressions through the mechanism of competitive endogenous RNAs (ceRNAs) [14]. MiRNA controls gene expression by binding to the 3'-UTR of target gene mRNA, resulting in target gene translation inhibition or degradation [15]. More and more evidence exhibited that a series of miRNAs (miR-96-

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Copyright © 2021 by the authors. Licensee KMB. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license. 5p, miR-133a-5p, miR-182-5p, and miR-661) are closely related to the progression of NSCLC [16-19]. Gu et al. confirmed that GAPLINC accelerates the progression of NSCLC via modulating the miR-661/eEF2K axis [19]. Nevertheless, the participation of circ\_0001947 in NSCLC progression via modulating miR-661 remains to be clarified.

Based on the above evidence, we aimed to figure out the influence of the circ\_0001947/miR-661 axis on the biological behaviors of NSCLC cells and to explore whether circ\_0001947 and miR-661 have complementary binding sites to determine potential therapeutic targets for NSCLC.

### **Materials and Methods**

#### **Ethics Statement**

We obtained NSCLC tissues and adjacent non-tumor tissues from 40 patients who had been diagnosed and operated in the Sanmen People's Hospital between March 2018 and July 2019. All participants signed written informed consent. The present research was ratified by the Ethics Committee of the Sanmen People's Hospital (2018020039).

In this research, tissue specimens were acquired via surgical resection and snap-frozen in a -80°C refrigerator (TSE600V, Thermo Scientific, USA). We used the quantitative real-time polymerase chain reaction (qRT-PCR) to examine the circ\_0001947, miR-661 and DOK7 expressions.

#### Cell Culture

Four human NSCLC cell lines H1299 (CRL-5803), A549 (CCL-185), H358 (CRL-5807), and H1395 (CRL-5868) were obtained from American Type Culture Collection (ATCC, USA). A human bronchial epithelial cell line (16HBE, MXC003) was procured from MEIXUAN (China). The above cells were grown in RPMI 1640 medium (30-2001, ATCC) supplemented with 10% fetal bovine serum (FBS, 30-2020, ATCC) in a Nunc Cell Factory Incubator (120300, Thermo Scientific) at 37°C with 5% CO<sub>2</sub>.

#### **Cell Transfection**

Small interfering RNAs targeting circ\_0001947 (si-circ\_0001947#1, sequence: 5'-GATGGACATGTAAACAAATTT-3'), si-circ\_0001947#2 (sequence: 5'-TGGAACAGAGTCTACAAATAA-3') and downstream of tyrosine kinase 7 (siDOK7, sequence: 5'-GACTGGAAATAAAATGTTCTTTC-3'), and non-targeting negative control (si-NC, sequence: 5'-ACCCGTTTCCGATTCTTTAGTT-3') were constructed by YouBio and inserted into the pSilencer 4.1-CMV neo vector (VT1395, YouBio, China). The full-length sequences of circ\_0001947 and DOK7 were constructed from Invitrogen and inserted into the pcDNA3.1 vector (V87020, Invitrogen, China). Empty vector was used as the NC. MiR-661 mimic (M, miR10003324-1-5), inhibitor (I, miR20003324-1-5), mimic control (MC, miR1N000001-1-5), and inhibitor control (IC, miR2N000001-1-5) were obtained from RiboBio (China). We used the lipofectamine 2000 (12566014, Invitrogen, USA) to transfect the above plasmids into A549 and H1299 cells. After 48 h (h) of transfection, qRT-PCR was used to determine the transfection rate.

#### **Cell Processing**

First, to fathom out the role of circ\_0001947 on cells, the cells were assigned into the control group, NC group, circ\_0001947 group, si-NC group, and si-circ\_0001947#2 group. In the last four groups, cells were transfected with NC, overexpressed circ\_0001947, si-NC and si-circ\_0001947#2, respectively. Second, to ascertain the functions of si-circ\_0001947#2 and miR-661 I on cells, the cells were separated into the control group, si-NC + IC group (co-transfection with si-NC and miR-661 IC), si-circ\_0001947#2 + IC group (co-transfection with sicirc\_0001947#2 and miR-661 IC), si-circ\_0001947#2 + I group (co-transfection with si-circ\_0001947#2 and miR-661 I), and si-NC + I group (co-transfection with si-NC and miR-661 I). Third, to investigate the functions of overexpressed circ\_0001947 and miR-661 M on cells, cells were divided into the control group, NC + MC group (co-transfection with NC and miR-661 MC), circ\_0001947 + MC group (co-transfection with overexpressed circ\_0001947 and miR-661 MC), circ\_0001947 + M group (co-transfection with overexpressed circ\_0001947 and miR-661 M), and NC + M group (co-transfection with NC and miR-661 M). Fourth, to comprehend the roles of miR-661 I and siDOK7 on cells, cells were classified into the IC + siNC group (co-transfection with miR-661 IC and siNC), I + siNC group (co-transfection with miR-661 I and siNC), I + siDOK7 group (co-transfection with miR-661 I and siDOK7), and IC + siDOK7 group (co-transfection with miR-661 IC and siDOK7). Fifth, to investigate the functions of miR-661 M and overexpressed DOK7 on cells, cells were divided into the MC + NC group (co-transfection with miR-661 MC and NC), M + NC group (co-transfection with miR-661 M and NC), M + DOK7 group (co-transfection with miR-661 M and DOK7), and MC + DOK7 group (co-transfection with miR-661 MC and overexpressed DOK7).

#### qRT-PCR

RNA from lung tissues and cells was acquired using the RNA Extraction Kit (N065, China). Next, to quantify the mRNA expression, cDNA synthesis was conducted in line with instructions of the cDNA first chain synthesis kit (N118, China) and miRNA cDNA first-strand synthesis kit (KR211, China), respectively. Then, cDNA was amplified and quantitatively analyzed by the FastFire qPCR PreMix (FP207-02, Tiangen) in a C1000 Touch PCR system (E1138, Bio-Rad, USA). GAPDH and U6 were employed for normalization controls. The results were normalized by the  $2^{-\Delta\Delta Ct}$  method [20]. The primers were listed in Table 1.

Table 1.	Gene sequence primers.	
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Name	Forward primer(5'-3')	Reverse primer(5'-3')
circ_0001947	ACACTCTTGGATGGAAAACCCA	CGTGTTCTGGACTCGGTTGG
miR-510	CTTCCATACTCAGGAGAGTGGC	TATCTCCAGACCAAGAC
miR-587	CCAGGCAAGAGAGAGTTGCTG	ATGGGCTTTCCACTGGTGATG
miR-661	AATGGTGGGTGCAAATGTGG	GAAACGCATGCCAAAAAGAC
miR-942	GCGCGCTCTTCTCTGTTTTGGC	GTGCAGGGTCCGAGGT
DOK7	GACAAGTCGGAGCGTATCAAG	ATGTCCTCTAGCGTCAGGCT
U6	GATTATCGGGACCATTCCACTG	GATCTGGTTCCCAATGACTGTG
GAPDH	AATGGATTTGGACGCATTGGT	TTTGCACTGGTACGTGTTGAT

#### Cell Counting Kit-8 (CCK-8)

We used the CCK-8 kit (abs50003, absin, China) to assess cell viability. Based on the above groups, cells  $(1 \times 10^4$  per well) were transfected and then maintained in an incubator for a specified time (24 or 48 h). Next, 10  $\mu$ l CCK-8 solution was adopted to treat cells for 4 h. Then, the optical density at a wavelength of 450 nm was determined under a microplate reader (Z742711, Sigma, USA).

### **Apoptosis Analysis**

The measurement of cell apoptosis was performed using the Annexin V-FITC/propidium iodide (PI) kit (C1062S, Beyotime, China). Cells ( $5 \times 10^4$  per well) were treated according to the above grouping conditions. After treatment, 195 µl binding buffer was added to the cell suspensions. Thereafter, 5 µl Annexin V-FITC and 10 µl PI were added to the cell suspensions and mixed well. The mixture was placed in an incubator at 37°C for 15 min (min). Finally, we used a flow cytometer (DxFLEX, Beckman, USA) to examine the FITC and PI signals.

#### **Colony Formation Assay**

In this research, the cells in good growth condition were made into a single-cell suspension. Cell suspension  $(1 \times 10^4 \text{ cells/ml})$  was seeded into a 6-well plate and then transfected based on the above groups. After incubation for 14 days, we used the 4% paraformaldehyde (P0099, Beyotime) and Giemsa dye solution (G8220, Solarbio, China) to fix and stain cells, respectively. Finally, we used a microscope (Z723975-1EA, Sigma) to count the clone numbers of the cells.

#### Cell Cycle Assay

Cell Cycle and Apoptosis Analysis Kit (C1052, Beyotime) was used in this research. After treatment, cells  $(1 \times 10^5 \text{ cells/ml})$  were collected and centrifuged. After being washed, the cells were immobilized by 70% cold ethanol solution (E111991, Aladdin, China) at 4°C overnight. Then, the cells were washed and reacted with prepared 0.5 ml PI stain solution at 37°C in the dark for 30 min. Finally, the percentage of NSCLC cells at different stages of cell cycle was examined using flow cytometer.

#### Target Gene Prediction and Dual-Luciferase Reporter Assay

Binding sites between circ\_0001947 and miR-661 as well as between miR-661 and DOK7 were determined by CircInteractome (https://circinteractome.irp.nia.nih.gov/) or TargetScan v7.2 (https://www.targetscan. org/). The wild-type (WT) or mutated (MUT) circ\_0001947 and DOK7 sequences were amplified by PCR and inserted into the pmirGLO vector (LM1439, LMAI Bio, China) to construct circ\_0001947-WT, circ\_0001947-MUT, DOK7-WT, and DOK7-MUT report plasmids. Then, all report plasmids were co-transfected with miR-661 M or MC into cells by lipofectamine 2000. The role of miR-661 M on luciferase activity was assessed after 48 h of transfection using the luciferase assay kit (16181, Thermo Scientific) under the chemiluminescence instrument (GloMax 2020, Promega, USA).

#### **Statistical Analysis**

Data were gathered and analyzed by GraphPad Prism 8.0 (GraphPad Software Inc., USA). The measurement data were expressed as mean  $\pm$  standard deviation (SD). The levels of circ\_0001947, miR-661, and DOK7 in 40 NSCLC tissues and adjacent non-tumor tissues were analyzed by paired-samples *t* test. An independent sample *t* test was employed to compare the difference of two groups. The multiple group comparisons were completed through a one-way analysis of variance (ANOVA) followed by Tukey's post hoc test. *p* < 0.05 was considered as statistically significant.

#### Results

## Circ\_0001947 Was Low-Expressed in NSCLC Tissues and Cells and Si-Circ\_0001947#2 Intensified the NSCLC Cell Viability

To explore the level of circ\_0001947 in NSCLC tissues or cell lines, we checked the data from qRT-PCR and clarified that circ\_0001947 expression was lower in NSCLC tissues than that in adjacent non-tumor tissues (Fig. 1A), while the same trend was discovered in NSCLC cells when compared with that in 16HBE cells (Fig. 1B). Since circ\_0001947 expression was notably down-regulated in H1299 cells and slightly restrained in A549 cells relative to other NSCLC cells, H1299 and A549 cells were singled out for subsequent experiments. Next, we transfected si-circ\_0001947#1, si-circ\_0001947#2, circ\_0001947 overexpression plasmids, or their NCs into



Fig. 1. Circ\_0001947 was low-expressed in NSCLC tissues and cells and si-circ\_0001947#2 intensified the NSCLC cell viability. A. The level of circ\_0001947 in 40 non-small cell lung cancer (NSCLC) tissues and adjacent non-tumor tissues was analyzed by quantitative real-time polymerase chain reaction (qRT-PCR). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as the internal control. (\*\*\*\*p < 0.001 vs. normal) B. The level of circ\_0001947 in NSCLC cells and normal cells was analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs. normal) B. The level of circ\_0001947 in NSCLC cells and normal cells was analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs.16HBE) C-D. The level of circ\_0001947 in NSCLC cells transfected with small interfering RNA targeting circ\_0001947 (si-circ\_0001947#1), si-circ\_0001947#2, or overexpressed circ\_0001947 was analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs. small interfering RNA targeting negative control (si-NC) group; ^^n p < 0.001 vs. negative control (NC) group) E-F. The viability of NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 was analyzed by Cell Counting Kit-8 (CCK-8). (p < 0.05, p < 0.01 vs. si-NC group; ^n p < 0.01 vs. NC group) All experiments have been performed in triplicate and data were expressed as mean ± standard deviation (SD).

H1299 and A549 cells, confirming that circ\_0001947 overexpression plasmids largely enhanced the circ\_0001947 level, while si-circ\_0001947#1 and si-circ\_0001947#2 showed opposite results (Figs. 1C-1D). Since si-circ\_0001947#2 generated a stronger inhibitory effect on the circ\_0001947 level relative to the si-circ\_0001947#1, we chose si-circ\_0001947#2 for the following experiments. Then, the CCK-8 assay manifested that the cell viability was mightily strengthened by si-circ\_0001947#2 but notably repressed by overexpressed circ\_0001947 (Figs. 1E and 1F).

## Si-circ\_0001947#2 Intensified Cell Proliferation, Induced Cell Cycle Arrest at S Phase and Restrained Apoptosis, while Overexpressed circ\_0001947 did the Opposite

The flow cytometer and colony formation assay exhibited that si-circ\_0001947#2 markedly restrained apoptosis and enhanced cell proliferation, while overexpressed circ\_0001947 generated the inverse effects (Figs. 2A-2D). Moreover, si-circ\_0001947#2 induced cell cycle arrest at S phase whereas circ\_0001947 overexpression facilitated the entry of G0/G1 phase but restrained the entry of G2/M and S phases (Figs. 2E and 2F).

## Si-circ\_0001947#2 Enhanced the miR-510, miR-587, miR-661 and miR-942 Levels and circ\_0001947 Targeted miR-661

The online database CircInteractome helped us acquire 47 miRNAs interacting with circ\_0001947. We finally detected the levels of these miRNAs related to NSCLC and speculated to bind to circ\_0001947, including miR-510, miR-587, miR-661 and miR-942. The qRT-PCR assay corroborated that si-circ\_0001947#2 evidently enhanced the miR-510, miR-587, miR-661, and miR-942 levels, while circ\_0001947 overexpression did the opposite (Figs. 3A and 3B). Since si-circ\_0001947#2 and circ\_0001947 overexpression exerted the most significant effects on the level of miR-661, miR-661 was screened out for subsequent experiments. CircInteractome analysis was used to certify that circ\_0001947 targeted miR-661 (Fig. 3C). Since then, we further confirmed that miR-661 M greatly reduced the luciferase activity of circ\_0001947-WT rather than that of circ\_0001947-MUT in H1299 and A549 cells (Figs. 3D and 3E).

## MiR-661 Was Overexpressed in NSCLC Tissues and Cells, and Participated in the Regulation of Circ\_0001947 on Cell Viability

Our study revealed that miR-661 was overexpressed in NSCLC tissues and cells (Figs. 4A and 4B). Furthermore, we explored the role of miR-661 in the regulation of circ\_0001947 on the viability of H1299 and A549 cells. Interestingly, the miR-661 level was intensified by si-circ\_0001947#2 but was weakened by miR-661 I, while the above effects were overturned by co-transfection of si-circ\_0001947#2 and miR-661 I (Fig. 4C). Co-transfection of overexpressed circ\_0001947 and miR-661 M reversed the regulation of miR-661 M / overexpressed circ\_0001947



Fig. 2. Si-circ\_0001947#2 intensified cell proliferation, induced cell cycle arrest at S phase and restrained apoptosis, while overexpressed circ\_0001947 did the opposite. A-B. Apoptosis of NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 was analyzed by flow cytometry. ("p < 0.001 vs. si-NC group;  $^{\wedge n}p < 0.001$  vs. NC group) C-D. Proliferation of NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 was analyzed by flow cytometry. ("p < 0.001 vs. si-NC group;  $^{\wedge n}p < 0.01$  vs. NC group) E-F. Cell cycle of NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 was analyzed by flow cytometry. ("p < 0.01 vs. si-NC group;  $^{\wedge p} < 0.01$  vs. NC group) E-F. Cell cycle of NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 was analyzed by flow cytometry. ("p < 0.01 vs. si-NC group;  $^{\wedge p} < 0.05$ ,  $^{\wedge p} < 0.01$  vs. NC group)

on the miR-661 level (Fig. 4D). Moreover, cell viability was elevated by si-circ\_0001947#2 but was inhibited by miR-661 I, the two tendencies were reversed by co-transfection of si-circ\_0001947#2 and miR-661 I (Fig. 4E). However, co-transfection of overexpressed circ\_0001947 and miR-661 M partially offset the regulation of miR-661 M / overexpressed circ\_0001947 on cell viability (Fig. 4F).

## MiR-661 Was Involved in the Regulation of circ\_0001947 on Apoptosis, Proliferation, and Cell Cycle of NSCLC Cells

As delineated in Fig. 5A, the results unveiled that apoptosis was largely elevated by miR-661 I but was suppressed by si-circ\_0001947#2, which was mitigated by co-transfection of si-circ\_0001947#2 and I. Meanwhile, an increase in apoptosis induced by overexpressed circ\_0001947 or a decrease in apoptosis triggered by miR-661 M was reversed by co-transfection of overexpressed circ\_0001947 and miR-661 M (Fig. 5B). Then, we also proved that co-transfection of si-circ\_0001947#2 and miR-661 I neutralized the inhibition of miR-661 I and the promotion of si-circ\_0001947#2 on cell proliferation (Fig. 5C). Moreover, co-treatment of overexpressed



Fig. 3. Si-circ\_0001947#2 enhanced miR-510, miR-587, miR-661, and miR-942 levels and circ\_0001947 targeted miR-661. A-B. The levels of miR-510, miR-587, miR-661, and miR-942 in NSCLC cells transfected with si-circ\_0001947#2 or overexpressed circ\_0001947 were analyzed by qRT-PCR. U6 was used as the internal control. (p < 0.05, p < 0.01, p < 0.001 vs. si-NC group; p < 0.05, p < 0.01, p < 0.001 vs. NC group) C. The binding sequences of circ\_0001947 containing the miR-661 binding sites were determined by CircInteractome (https://circinteractome.irp.nia.nih.gov/). D-E. Direct interaction between circ\_0001947 and miR-661 was detected by dual-luciferase reporter assay. (p < 0.01, p < 0.001 vs. mimic control (MC) group)

circ\_0001947 and miR-661 M partially offset the regulation of miR-661 M / overexpressed circ\_0001947 on cell proliferation (Fig. 5D). The detection of cell cycle confirmed that si-circ\_0001947#2 and miR-661 M induced cell cycle arrest at S phase, while miR-661 I and overexpressed circ\_0001947 facilitated the entry of G0/G1 phase (Figs. 5E and 5F). Besides, the effect of si-circ\_0001947#2 or miR-661 I on cell cycle was partially reversed by co-transfection of si-circ\_0001947#2 and I , while the effect of overexpressed circ\_0001947 or miR-661 M was partially reversed by co-transfection of overexpressed circ\_0001947 and miR-661 M (Figs. 5E and 5F).

#### MiR-661 Targeted DOK7

We used TargetScan v7.2 analysis to discover that miR-661 targeted DOK7 (Fig. 6A). Additionally, the relative luciferase activity was lower in the miR-661 M group than that in the miR-661 MC group when the sequence of DOK7 was wild type (Figs. 6B and 6C), while there was no obvious difference between miR-661 M and MC groups when the sequence of DOK7 was mutant.

## The Expression of DOK7 Was Down-Regulated in NSCLC Tissues and Cells, Which Was Regulated by Circ\_0001947

In this study, we first tested the level of DOK7 in NSCLC tissues or cell lines and manifested its downregulation (Figs. 7A and 7B). Next, we uncovered that DOK7 expression was dramatically inhibited by si-circ\_0001947#2 and miR-661M, but was greatly promoted by overexpressed circ\_0001947 and miR-661 I (Figs. 7C and 7D). Additionally, the effect of si-circ\_0001947#2 or miR-661 I on the mRNA expression of DOK7 was partially reversed by co-transfection of si-circ\_0001947#2 and miR-661 I in A549 cells, while the effect of overexpressed





circ\_0001947 or miR-661 M on the mRNA expression of DOK7 was partially reversed by co-transfection of overexpressed circ\_0001947 and miR-661 M in H1299 cells (Figs. 7C and 7D).

### DOK7 Was Partly Involved in the Regulation of miR-661 on NSCLC Cell Viability

To figure out the role of DOK7 in the regulation of miR-661 on H1299 and A549 cells, we measured the alteration of DOK7 expression. Co-transfection of miR-6611 and siDOK7 partially reversed the regulation of miR-661 I or siDOK7 on DOK7 expression (Fig. 8A). In the meantime, the effect of miR-661 M on restraining DOK7 expression was discovered to be mitigated by overexpressed DOK7 (Fig. 8B). Besides, the promotion of cell viability induced by siDOK7 and the inhibition of that induced by miR-661 I were reversed by co-transfection of miR-661 I and siDOK7 (Fig. 8C). The enhancement of miR-661 M on cell viability was offset by overexpressed DOK7 (Fig. 8D).

### Discussion

In our study, we tested the expression of circ\_0001947 in NSCLC tissues, adjacent non-tumor tissues, and multiple NSCLC cell lines. Subsequently, we observed changes in cell viability, proliferation, apoptosis, and miRNAs expressions after circ\_0001947 overexpression or silencing in H1299 and A549 cells. On this basis, we further verified the interactions between circ\_0001947 and miR-661 as well as between miR-661 and downstream target genes, suggesting that miR-661 and downstream target genes might be the mediators of circ\_0001947 in regulating the biological behaviors of NSCLC cells. Our findings revealed the role of circ\_0001947 in modulating certain vital features of NSCLC.

First of all, we have clarified that circ\_0001947 was low-expressed in NSCLC tissues and cell lines. Moreover, early research has demonstrated that circ\_0001947 facilitates gastric cancer progression via modulating the miR-6894-5p/ANTXR1 axis [21]. Han et al. has put forward that circ\_0001947 restrains acute myeloid leukemia development by modulating the miR-329-5p/CREBRF axis [12]. Our study complemented the research deficiency of circ\_0001947 in NSCLC, which expounded for the first time that circ\_0001947 overexpression inhibited cell viability and proliferation, induced cell cycle arrest at G0/G1 phase, and promoted apoptosis, while circ\_0001947 knockdown exerted the opposite effects, manifesting the involvement of circ\_0001947 in the progression of NSCLC.

Additionally, by searching the online database CircInteractome, we detected miR-510, miR-587, miR-661, and





**Fig. 6. MiR-661 targeted DOK7. A.** The binding sequences of DOK7 containing the miR-661 binding sites were determined by TargetScan v7.2 (https://www.targetscan. org/). **B-C.** Direct interaction between DOK7 and miR-661 was detected by dual-luciferase reporter assay. (\*\*p < 0.01, \*\*\*p < 0.001 vs. MC group)



Fig. 7. DOK7 was down-expressed in NSCLC tissues and cells, whose expression was regulated by circ\_0001947. A. The level of DOK7 in 40 NSCLC tissues and adjacent non-tumor tissues was analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs. normal) B. The level of DOK7 in NSCLC cells and normal cells was analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs.16HBE) C. The effects of circ\_0001947 knockdown and miR-661 I on DOK7 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs. si-NC + IC group; \*\*\*p < 0.001 vs. si-NC + I group; ^^^ p < 0.001 vs. si-circ\_0001947#2 + IC group) D. The effects of circ\_0001947 overexpression and miR-661 M on DOK7 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. (^^p < 0.001 vs. si-Cr\_0001947 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. (\*\*\*p < 0.001 vs. si-NC + I group;  $^{0.00} p < 0.001$  vs. si-Cr\_0001947#2 + IC group) D. The effects of circ\_0001947 overexpression and miR-661 M on DOK7 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. (^^p p < 0.001 vs. circ\_0001947 + MC group;  $^{1.0} p < 0.001$  vs. NC + MC group;  $^{0.00} p < 0.001$  vs. circ\_0001947 + MC group;  $^{1.1} p < 0.001$  vs. NC + M group)

miR-942 expressions, whose functions and expressions were related to NSCLC. The qRT-PCR assay unearthed that circ\_0001947 knockdown and overexpression made profound impacts upon the level of miR-661, and miR-661 was therefore chosen for follow-up experiments. CircRNAs are important members of the ceRNA network,



**Fig. 8. DOK7 was partly involved in the regulation of miR-661 on NSCLC cell viability. A.** The effects of miR-661 I and small interfering RNA targeting DOK7 (siDOK7) on DOK7 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. ("p < 0.001 vs. IC + siNC group;  $^{\triangle \triangle} p < 0.001$  vs. I + siNC group;  $^{\pm\pm} p < 0.001$  vs. IC + siDOK7 group) **B**. The effects of miR-661 M and overexpressed DOK7 on DOK7 expression were analyzed by qRT-PCR. GAPDH was used as the internal control. ( $^{\wedge}p < 0.01$ ,  $^{\wedge \wedge}p < 0.001$  vs. MC + NC group;  $^{\pm\pm}p < 0.001$  vs. MC + DOK7 group;  $^{\oplus\Phi\Phi}p < 0.001$  vs. M + NC group) **C**. The effects of miR-661 I and siDOK7 on cell viability were analyzed by CCK-8. ( $^{\circ}p < 0.05$ , "p < 0.01, "p < 0.001 vs. IC + siNC group;  $^{\pm}p < 0.001$  vs. MC + NC group;  $^{\pm}p < 0.001$  vs. MC + DOK7 group;  $^{\pm}p < 0.05$  vs. M + NC group)

playing crucial roles as "miRNA sponges" in the "circRNA-miRNA-mRNA" axis [22]. In our research, to probe into potential downstream pathways that may be implicated in the function of circ\_0001947 in NSCLC, we performed database analysis and finally discovered that miR-661 may be a potential target of circ\_0001947 which was further supported by the dual-luciferase reporter assay. MiR-661 has been confirmed to be abnormally expressed in a variety of malignant tumors and closely associated with the initiation and progression of tumors [23, 24]. Wang, et al. has disclosed that miR-661 is overexpressed in NSCLC tissues and cells and it accelerates NSCLC development via targeting RUNX3, which is consistent with findings of our research [25]. Liu et al. has proposed that miR-661 facilitates the invasion and metastasis of NSCLC cells via restraining RB1 [26]. Then, we provided the first demonstration that miR-661 inhibitor suppressed cell viability and proliferation, induced cell cycle arrest at G0/G1 phase, and elevated apoptosis, while the above effects were overturned by co-transfection of circ\_0001947 knockdown and miR-661 inhibitor. It was clarified that miR-661 might serve as a regulator in the regulation of circ\_0001947 on the cell biological behaviors of NSCLC cells.

Furthermore, in order to explore the target of miR-661 in NSCLC, we used database prediction software to predict the target gene of miR-661 and confirmed that DOK7 might be a potential target of miR-661. Luciferase assay corroborated that miR-661 could directly bind to 3'-UTR of DOK7. The DOK7 gene, a member of the DOK family, is located at the end of the short arm of chromosome 4 [27]. Recent studies have revealed that the DOK7 gene and its family are closely related to the initiation and development of many malignant tumors, such as lung cancer, breast cancer, and leukemia [28-30], but the detailed mechanism has not been fully understood. At present, there are few studies on the specific role of DOK7 in NSCLC. Only one study confirmed that DOK7 was greatly under-expressed in lung cancer, and DOK7 overexpression suppressed the proliferation and migration of lung cancer cells [27]. Consistent with the previous study, our data proved that DOK7 was down-expressed in NSCLC tissues and cells, the expression of which was discovered to be elevated by circ\_0001947 overexpression through targeting miR-661. Meanwhile, DOK7 was partly involved in the regulation of miR-661 on NSCLC cell viability. It suggested that circ\_0001947-modulated miR-661 could further modulate the upregulation of DOK7 in NSCLC cells.

However, despite the evidence that DOK7 is the target gene of miR-661, the upstream and downstream pathways of DOK7 are not tested, which may be a defect of this study. In addition, this study lacks in vivo experiments, which will be listed in our future experimental projects, so as to further improve the accuracy of this research.

In conclusions, altogether, this research emphasized that circ\_0001947 acted as a sponge of miR-661 to restrain NSCLC progression via modulating DOK7, revealing that circ\_0001947 might be a potential new biomarker for the treatment of NSCLC.

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### **Conflict of Interest**

The authors have no financial conflicts of interest to declare.

#### References

- 1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. 2018. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: Cancer J. Clin. 68: 394-424.
- 2. Osarogiagbon RU, Veronesi G, Fang W, Ekman S, Suda K, Aerts JG, et al. 2019. Early-stage NSCLC: advances in thoracic oncology 2018. J. Thorac. Oncol. 14: 968-978.
- Gupta GP, Massagué J. 2006. Cancer metastasis: building a framework. *Cell* 127: 679-695.
  Yang P, Allen MS, Aubry MC, Wampfler JA, Marks RS, Edell ES, *et al.* 2005. Clinical features of 5,628 primary lung cancer patients: experience at Mayo Clinic from 1997 to 2003. Chest 128: 452-462.
- 5. Sanger HL, Klotz G, Riesner D, Gross HJ, Kleinschmidt AK. 1976. Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. Proc. Natl. Acad. Sci. USA 73: 3852-3856.
- 6. Cocquerelle C, Mascrez B, Hétuin D, Bailleul B. 1993. Mis-splicing yields circular RNA molecules. FASEB J. 7: 155-160.
- Xu T, Wu J, Han P, Zhao Z, Song X. 2017. Circular RNA expression profiles and features in human tissues: a study using RNA-seq 7. data. BMC Genomics 18: 680.
- 8. Zhang ZY, Gao XH, Ma MY, Zhao CL, Zhang YL, Guo SS. 2020. CircRNA\_101237 promotes NSCLC progression via the miRNA-490-3p/MAPK1 axis. Sci. Rep. 10: 9024
- Chen RX, Liu HL, Yang LL, Kang FH, Xin LP, Huang LR, et al. 2019. Circular RNA circRNA\_0000285 promotes cervical cancer 9 development by regulating FUS. Eur. Rev. Med. Pharmacol. Sci. 23: 8771-8778
- 10. Han J, Zhao G, Ma X, Dong Q, Zhang H, Wang Y, et al. 2018. CircRNA circ-BANP-mediated miR-503/LARP1 signaling contributes to lung cancer progression. Biochem. Biophys. Res. Commun. 503: 2429-2435.
- 11. Wu J, Vogel T, Gao X, Lin B, Kulwin C, Chen J. 2018. Neuroprotective effect of dexmedetomidine in a murine model of traumatic brain injury. Sci Rep. 8: 4935.
- 12. Han F, Zhong C, Li W, Wang R, Zhang C, Yang X, et al. 2020. hsa\_circ\_0001947 suppresses acute myeloid leukemia progression via targeting hsa-miR-329-5p/CREBRF axis. Epigenomics 12: 935-953
- 13. Cai X, Lin L, Zhang Q, Wu W, Su A. 2020. Bioinformatics analysis of the circRNA-miRNA-mRNA network for non-small cell lung cancer. J. Int Med. Res. 48: 300060520929167
- 14. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, et al. 2013. Natural RNA circles function as efficient microRNA sponges. Nature 495: 384-388. 15. Kozomara A, Griffiths-Jones S. 2011. miRBase: integrating microRNA annotation and deep-sequencing data. Nucleic Acids Res.
- 39: D152-157 16. Wang T, Wang X, Du Q, Wu N, Liu X, Chen Y, et al. 2019. The circRNA circP4HB promotes NSCLC aggressiveness and metastasis by
- sponging miR-133a-5p. Biochem. Biophys. Res. Commun. 513: 904-911. 17. Wei S, Zheng Y, Jiang Y, Li X, Geng J, Shen Y, et al. 2019. The circRNA circPTPRA suppresses epithelial-mesenchymal transitioning
- and metastasis of NSCLC cells by sponging miR-96-5p. EBioMedicine 44: 182-193. 18. Li C, Fei K, Tian F, Gao C, Yang S. 2019. Adipose-derived mesenchymal stem cells attenuate ischemic brain injuries in rats by
- modulating miR-21-3p/MAT2B signaling transduction. Croat. Med. J. 60: 439-448.
- 19. Gu H, Chen J, Song Y, Shao H. 2018. Gastric adenocarcinoma predictive long intergenic non-coding RNA promotes tumor occurrence and progression in non-small cell lung cancer via regulation of the miR-661/eEF2K signaling pathway. Cell. Physiol. Biochem. 51: 2136-2147.
- 20. Yu W, Sun Z, Yang L, Han Y, Yue L, Deng L, et al. 2019. IncRNA PTAR promotes NSCLC cell proliferation, migration and invasion by sponging microRNA-101. Mol. Md. Rep. 20: 4168-4174.
- 21. Bu X, Chen Z, Zhang A, Zhang X, Yuan H, Zhang Y, et al. 2021. Circular RNA circAFF2 accelerates gastric cancer development by activating miR-6894-5p and regulating ANTXR 1 expression. Clin. Res. Hepatol. Gastroenterol. 45: 101671.
- 22. Chen X, Chen RX, Wei WS, Li YH, Feng ZH, Tan L, et al. 2018. PRMT5 circular RNA promotes metastasis of urothelial carcinoma of the bladder through sponging miR-30c to induce epithelial-mesenchymal transition. Clin. Cancer Res. 24: 6319-6330.
- 23. Zhu T, Yuan J, Wang Y, Gong C, Xie Y, Li H. 2015. MiR-661 contributed to cell proliferation of human ovarian cancer cells by repressing INPP5J expression. Biomed. Pharmacother. 75: 123-128.
- 24. Li Z, Liu YH, Diao HY, Ma J, Yao YL. 2015. MiR-661 inhibits glioma cell proliferation, migration and invasion by targeting hTERT. Biochem. Biophys. Res. Commun. 468: 870-876.
- Wang ZM, Wan XH, Sang GY, Zhao JD, Zhu QY, Wang DM. 2017. miR-15a-5p suppresses endometrial cancer cell growth via Wnt/ 25. beta-catenin signaling pathway by inhibiting WNT3A. Eur. Rev. Med. Pharmacol. Sci. 21: 4810-4818.
- 26. Liu F, Cai Y, Rong X, Chen J, Zheng D, Chen L, et al. 2017. MiR-661 promotes tumor invasion and metastasis by directly inhibiting RB1 in non small cell lung cancer. Mol. Cancer 16: 122.
- 27. Chen G, Yu H, Satherley L, Zabkiewicz C, Resaul J, Zhao H, et al. 2017. The downstream of tyrosine kinase 7 is reduced in lung cancer and is associated with poor survival of patients with lung cancer. Oncol. Rep. 37: 2695-2701.
- 28. Berger AH, Niki M, Morotti A, Taylor BS, Socci ND, Viale A, et al. 2010. Identification of DOK genes as lung tumor suppressors. Nat. Genet. 42: 216-223
- 29. Heyn H, Carmona FJ, Gomez A, Ferreira HJ, Bell JT, Sayols S, et al. 2013. DNA methylation profiling in breast cancer discordant identical twins identifies DOK7 as novel epigenetic biomarker. Carcinogenesis 34: 102-108.
- 30. Niki M, Di Cristofano A, Zhao M, Honda H, Hirai H, Van Aelst L, et al. 2004. Role of Dok-1 and Dok-2 in leukemia suppression. J. Exp. Med. 200: 1689-1695.