

Keloid Patient Plasma-Derived Exosomal hsa_circ_0020792 Promotes Normal Skin Fibroblasts Proliferation, Migration, and Fibrogenesis via Modulating miR-193a-5p and Activating TGF- β 1/Smad2/3 Signaling

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Background: Keloids are fibroproliferative disorders, which seriously affect the quality of life of patients with keloids. Additionally, circRNAs are enriched within exosomes derived from human blood samples, whereas their relationship with keloids remains largely unknown. It has been reported that hsa_circ_0020792 was abnormally upregulated in keloid tissues. However, the role of keloid patient plasma-derived exosomal hsa_circ_0020792 in the formation and development of keloids is not well understood.

Methods: Exosomes were isolated from the peripheral blood plasma of the patients with keloids (keloid patient-Exo) and healthy controls (Healthy control-Exo). The hsa_circ_0020792 and miR-193a-5p levels in keloid patient-Exo and healthy control-Exo, as well as in keloid fibroblasts and normal skin fibroblasts (NFs) were evaluated by RT-qPCR.

Results: The level of hsa_circ_0020792 was remarkably increased in keloid patient-Exo and keloid fibroblasts compared with that in Healthy control-Exo and NFs, respectively. In addition, keloid patient-Exo obviously enhanced the viability, migration, and extracellular matrix (ECM) synthesis, but reduced the apoptosis of NFs. Moreover, keloid patient-Exo notably promoted the fibrogenesis of NFs, as characterized by enhanced TGF- β signaling, increased expressions of phosphorylated Smad2/3. However, downregulation of hsa_circ_0020792 markedly reversed the promoting effects of keloid patient-Exo on cell growth, migration, and myofibroblast activation and fibrogenesis. Furthermore, downregulation of hsa_circ_0020792 significantly reduced the viability, migration, and fibrogenesis in NFs, whereas these phenomena were reversed by miR-193a-5p inhibitor.

Conclusion: Collectively, keloid patient plasma-derived exosomal hsa_circ_0020792 could promote the proliferation, migration, and fibrogenesis of NFs via modulating miR-193a-5p and activating TGF- β 1/Smad2/3 signaling.

Keywords: keloid, fibroblasts, exosomes, hsa_circ_0020792, miR-193a-5p

Introduction

Scarring in the skin after various trauma not only affects the external skin of the patient, but also seriously affects the quality of life of patients.¹⁻³ In addition, pathological scar mainly includes keloid and hypertrophic scar, which is a spontaneous and excessive skin fibro-proliferative disease after trauma.^{4,5} Clinically, keloid is considered a “benign tumor” of the skin.^{5,6} The reason is that keloids grow beyond the boundary of the original wound, and there is no sign of spontaneous decay.⁷⁻⁹ Currently, the treatments for keloid include surgery and laser therapy; however, the therapeutic effects are not satisfactory.^{10,11}

Circular RNAs (circRNAs) play important roles in mediating cell proliferation, migration, and invasion.^{12–14} CircRNAs can bind intracellular microRNAs (miRNAs) as competing endogenous RNAs (ceRNAs), thereby blocking the inhibition of miRNAs on their target genes.¹⁵ Moreover, it has been shown that circRNAs are related to the formation and development of keloid.^{16,17} For instance, circCOL5A1 could accelerate the development process of keloid by downregulating miR-7-5p.¹⁶ Besides, knockdown of circPDE7B could inhibit the progression of keloid by increasing miR-661.¹⁸

Exosomes are nano-sized membrane vesicles containing active substances (DNA, RNA, proteins, lipids, etc.).^{19,20} Additionally, exosomes are able to provide a new method for transferring effector information between cells in biological functions and normal body responses.^{21,22} For example, exosomal miRNA-21 can accelerate the proliferation of keloid fibroblasts by downregulating Smad7.²³ Exosomal miR-29a derived from mesenchymal stem cells was able to suppress the excessive proliferation of human hypertrophic scar fibroblasts.²⁴ Shi et al found that hsa_circ_0020792 is abnormally upregulated in keloid tissues.¹⁷ However, the role of keloid patient-derived exosomal hsa_circ_0020792 in the formation and development of keloid remains unclear. Thus, we aimed to explore whether keloid patient-derived exosomal hsa_circ_0020792 could regulate NFs proliferation, migration, and ECM production.

Materials and Methods

Samples

All specimens used in this study were obtained from Shanghai Sixth People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine. The peripheral blood was collected from three patients with keloids (3 females, range 24–34 years old) and three healthy participants (3 females, range 25–32 years old). This study complies with the declaration of Helsinki and was approved by the Ethics Committee of Shanghai Jiao Tong University Affiliated Sixth People's Hospital. Informed consent was obtained from all participants.

Exosomes Extraction and Identification

The Exosome Isolation and Purification Kit (cat. no. UR52136-30T; Umibio, China) was applied to isolate exosomes from the peripheral blood plasma samples of patients with keloids and healthy participants (Keloid patient-Exo and Healthy control-Exo). Briefly, samples were centrifuged at 3000 x g for 10 min, and 10,000 x g for 10 min. Next, the supernatant was treated with Blood PureExo Solution at 4°C for 2 h, and then centrifuged at 10,000 x g for 1 h. Finally, the exosome vesicles were obtained and resuspended in PBS.

The isolated vesicles were suspended in 2% PFA and then added to a formvar-carbon copper grid. After that, the grid was treated with 1% glutaraldehyde for 5 min. Next, the grid was treated with uranium oxalate (pH = 7) for 5 min, and then treated with methylcellulose reagent for 10 min. Subsequently, a transmission electron microscopy (TEM) was used to observe the morphology of the isolated vesicles.

Next, the ZetaView analyzer was calibrated using polystyrene microspheres. Then, the vesicles were diluted by 1× PBS buffer. Subsequently, Nanoparticle Tracking Analysis (NTA) was used to observe the number and size of vesicles according to the previous report.²⁵

Western Blot Assay

The protein concentration was quantified using the BCA protein assay kit (cat. no. C503021, Sangon biotech). Later on, 10% SDS-PAGE was used to separate protein. Next, protein was transferred onto the PVDF membranes. After that, the membranes were incubated with primary antibodies against CD9 (cat. no. 20597-1-AP, Proteintech), CD63 (cat. no. 67605-1-Ig, Proteintech), CD81 (cat. no. 66866-1-Ig, Proteintech), Collagen I (cat. no. ab260043, Abcam), Collagen III (cat. no. 22734-1-AP, Proteintech), α -SMA (cat. no. 23081-1-AP, Proteintech), TGF- β 1 (cat. no. 21898-1-AP, Proteintech), p-Smad2 (cat. no. ab280888, Abcam), Smad2 (cat. no. 12570-1-AP, Proteintech), Smad3 (cat. no. 66516-1-Ig, Proteintech), p-Smad3 (cat. no. ab52903, Abcam), Bcl-2 (cat. no. 12789-1-AP, Proteintech), Bax (cat. no. 50599-2-Ig, Proteintech), cleaved caspase 3 (cat. no. ab2302, Abcam), and GAPDH (cat. no. 60004-1-1, Proteintech) overnight at 4°C. The membranes were then immersed with an HRP-conjugated secondary antibody (A0216, Beyotime) at room temperature

for 2 h. Afterwards, blot signals were observed by an ECL kit (cat. no. AS1059, ASPEN). β -actin was used as the internal standard. The western ladder (cat. no. WJ103, Shanghai Epizyme Biomedical Technology Co., Ltd) was used in this study.

Cell Culture

Fibroblasts collected from normal skin tissues were obtained from BeNa Culture Collection (China) and human keloid fibroblasts were obtained from Procell (China). Cells were cultured in DMEM (cat. no. L110, Shanghai BasalMedia Technologies Co., LTD) in 5% CO₂ at 37°C. DMEM was contained 10% FBS (cat. no. 1803122, Biological Industries-Israel), 1% penicillin and 1% streptomycin (cat. no. 15070063, Gibco).

Cell Transfection

NFs was transfected with siRNA negative control (siR-NC), hsa_circ_0020792 siRNA1 (siR-circ-1), hsa_circ_0020792 siRNA2 (siR-circ-2) or hsa_circ_0020792 siRNA3 (siR-circ-3), miR-193a-5p inhibitor (miR-inhibitor) and miRNA negative control (miR-NC) using Lipofectamine 2000 (Thermo Fisher Scientific), respectively. The sequence as follows: siRNA-NC, ATTACAAGCGTTCACATTA; siR-circ-1, AAGCACAGCAGCATCTTCAAA; siR-circ-2, AAGCAC TCATACTTTATGCAT; siR-circ-3, AAGCTCAGAAATTGGCTTTAA; miR-inhibitor, 5'-UCAUCUCGCCCGCAAAG ACCCA-3'.

Reverse Transcription-Quantitative PCR (RT-qPCR)

The total RNA was extracted using Trizol reagent (cat. no. 15596026, Invitrogen). Next, the corresponding cDNAs were synthesized using the ReverTra Ace qPCR RT Kit (cat. no. FSQ-101, TOYOBO). Later on, qPCR was performed on the PCR System (ABI) using a SYBR Green PCR MASTER kit (cat. no. 4368708, Applied Biosystems). The primer sequence as follows: hsa_circ_0020792, forward, 5'-GGCCCCGTGTTTACTCAACT-3' and reverse, 5'-CTGGGGAAGTTGTCGAAGATCA-3'; GAPDH, forward, 5'-GGAGCGAGATCCCTCCAAAAT-3' and reverse, 5'-GGCTGTTGTCATACTTCTCATGG-3'; miR-193a-5p, forward, 5'-TATATGGGTCTTTGCGGGCG-3' and reverse, 5'-GTGCAGGGTCCGAGGT-3'; U6, forward, 5'-CTCGCTTCGGCAGCACAT-3' and reverse, 5'-AACGCTTACGAATTTGCGT-3'.

Cell Counting Kit-8 (CCK-8) Assay

CCK-8 assay was applied to determine cell viability. NFs were seeded into 96-well plates (5×10^3 cells per well) overnight, and then treated with indicated exosomes (10 μ g/mL) for 24 h. After that, cells were mixed with 10 μ L of CCK-8 reagent (cat. no. MA0218-T, meilunbio[®]) for 2 h at 37°C. Next, the absorbance was detected at 450 nm with a microplate reader (Infinite M Nano, TECAN).

Flow Cytometry Assay

The Annexin V-FITC apoptosis detection kit (cat. no. C1062S, Beyotime) was used for evaluating cell apoptosis. NFs were incubated with indicated exosomes (10 μ g/mL) for 24 h. After that, cells were stained with 5 μ L annexin V-FITC and PI reagent for 15 min. Subsequently, a flow cytometer (BD Bioscience) was used to detect the apoptotic cells.

Transwell Migration Assay

Transwell assay was used for assessing cell migratory ability. The 24-well transwell chamber (cat. no. 3422, Corning) was used to perform transwell migration assay. NFs (1×10^5 cells per well) were plated onto the upper chamber and cultured with serum free medium. Meanwhile, 500 μ L of medium with 10% FBS was loaded into the lower chamber. Later on, cells that had migrated from the upper part of the membrane to the lower part were stained with 0.1% crystal violet (cat. no. AS1086, ASPEN) at 24 h. Next, a microscope (ECLIPSE TS2, Nikon) was used to observe the stained migrated cells.

Statistical Analysis

The results data were analyzed using GraphPad Prism using One-way analysis of variance (ANOVA) and Tukey's tests. Data were presented as mean \pm SEM. *P < 0.05 was considered significant. All data were repeated in triplicate.

Results

Exosomes are Successfully Collected from Peripheral Blood of Healthy Participants and Patients with Keloid

To explore the role of exosomes in the formation and development of keloid, exosomes were collected from the peripheral blood of the patients with keloid (keloid patient-Exo) and healthy controls (Healthy control-Exo). The results of TEM and NTA indicated that these isolated vesicles (50–150 nm in diameter) have lipid-bilayer membrane structures (Figure 1A and B). Meanwhile, these isolated vesicles were expressed exosome marker proteins CD9, CD63 and CD81 (Figure 1C). Collectively, exosomes were successfully isolated from peripheral blood of healthy participants and patients with keloid.

Hsa_circ_0020792 Level is Increased and miR-193a-5p Level is Reduced in Exosomes Derived from Peripheral Blood of the Patients with Keloid

The data from Starbase (<https://starbase.sysu.edu.cn>) showed that there are two potential binding sites for miR-193a-5p in hsa_circ_0020792 (Supplementary Figure 1), suggesting that hsa_circ_0020792 might be a target of miR-193a-5p. Thus, we investigated whether keloid patient-Exo are enriched with hsa_circ_0020792, and whether keloid patient-Exo could regulate the development of keloids via targeting miR-193a-5p. As indicated in Figure 2A and B, hsa_circ_0020792 level was greatly elevated and miR-193a-5p level was obviously reduced in keloid patient-Exo compared to healthy control-Exo. In addition, hsa_circ_0020792 was increased and miR-193a-5p was decreased in keloid fibroblasts compared with that in NFs (Figure 2C and D). Moreover, the levels of ECM proteins Collagen I, Collagen III and α -SMA were elevated in keloid fibroblasts compared to NFs (Figure 2E). Furthermore, hsa_circ_0020792 level was significantly increased in TGF- β 1-treated NFs (Supplementary Figure 2). To sum up, hsa_circ_0020792 level was increased and miR-193a-5p level was reduced in keloid patient-Exo and keloid fibroblasts.

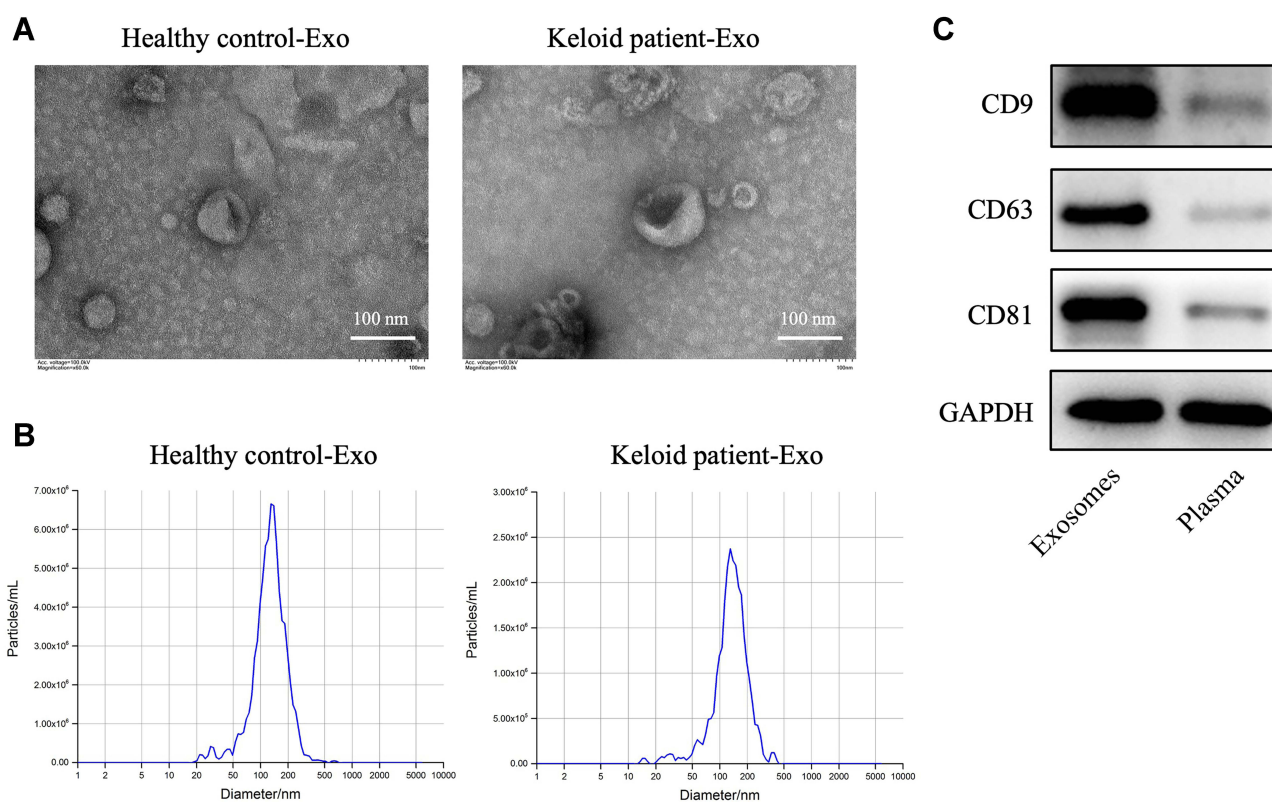


Figure 1 Exosomes are successfully collected from peripheral blood of healthy controls and patients with keloid. (A and B) Exosomes (healthy control-Exo and keloid patient-Exo) were isolated from peripheral blood of healthy controls (n = 3) and patients with keloid (n = 3). The isolated exosomes were identified using TEM and NTA analysis. (C) Western blot assay was used to detect CD9, CD63 and CD81 protein expressions in isolated exosomes and plasma of peripheral blood plasma (n = 3).

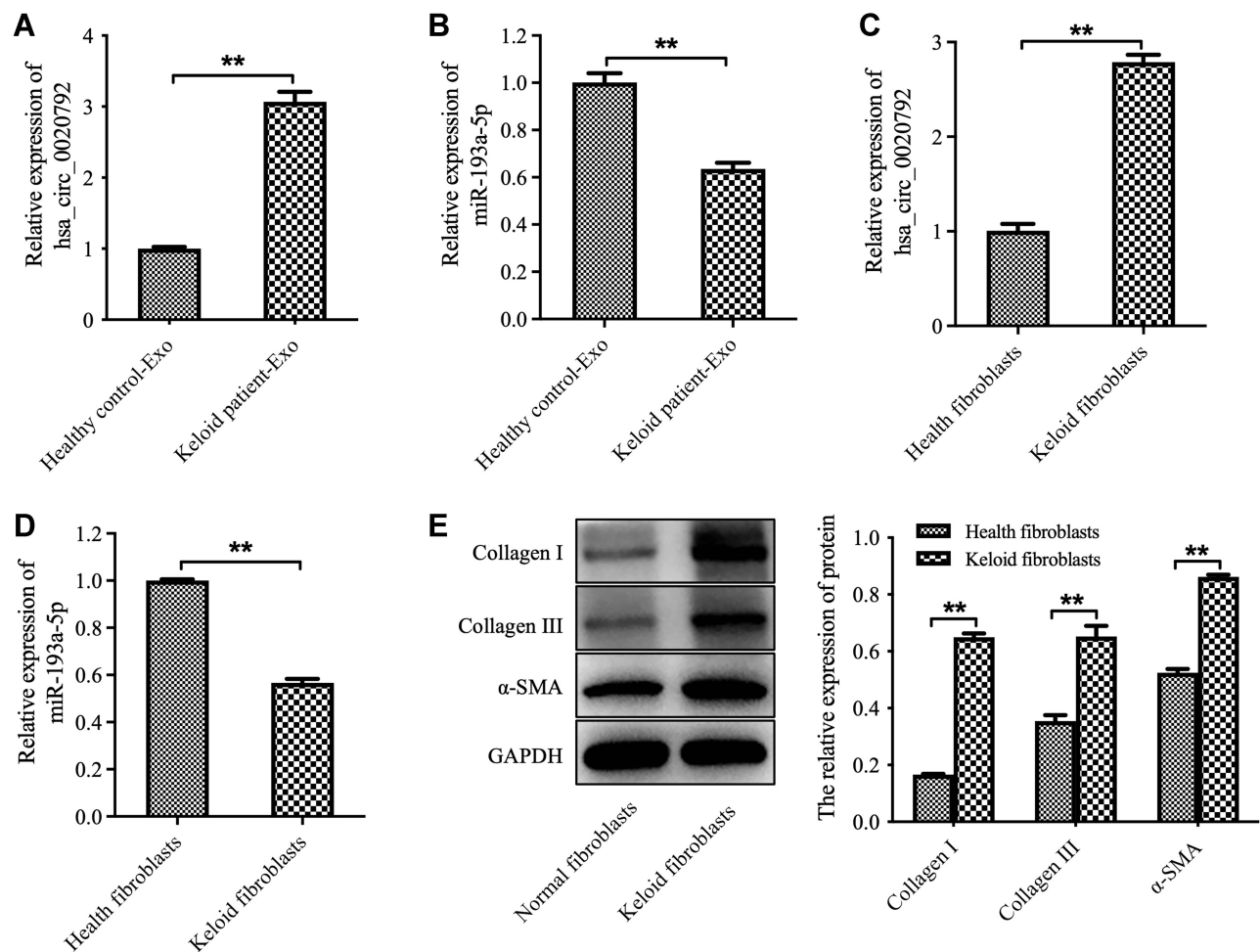


Figure 2 The level of hsa_circ_0020792 is increased and the level of miR-193a-5p is reduced in exosomes derived from patients with keloid. (A and B) The hsa_circ_0020792 and miR-193a-5p level in healthy control-Exo and keloid patient-Exo were detected using RT-qPCR. (C and D) The hsa_circ_0020792 and miR-193a-5p level in normal skin fibroblasts (NFs) and keloid fibroblasts were detected using RT-qPCR. (E) The expressions of Collagen I, Collagen III and α -SMA in NFs and keloid fibroblasts were detected using Western blot assay. **P < 0.01, n = 3.

Keloid Patient-Derived Exosomal hsa_circ_0020792 Enhances the Viability and Reduces the Apoptosis of NFs

Next, to investigate whether hsa_circ_0020792-enriched keloid patient-Exo could affect the proliferation and collagen synthesis in NFs, NFs were transfected with siR-circ-1, siR-circ-2 or siR-circ-3. As shown in Figure 3A, siR-circ-2 remarkably reduced the level of hsa_circ_0020792 in NFs. Additionally, keloid patient-Exo obviously enhanced the viability and reduced the apoptosis of NFs compared to healthy control-Exo, whereas these phenomena were reversed by siR-circ-2 (Figure 3B–D). Meanwhile, keloid patient-Exo notably upregulated the level of Bcl-2 and downregulated the level of Bax and cleaved caspase 3 in NFs, whereas siR-circ-2 was able to reverse these changes (Figure 3E). All in all, keloid patient-derived exosomal hsa_circ_0020792 could promote the viability and inhibit the apoptosis of NFs.

Keloid Patient-Derived Exosomal hsa_circ_0020792 Promotes the Migration and Fibrogenesis of NFs via Activating TGF- β 1/Smad2/3 Signaling

In order to explore the effect of keloid patient-derived exosomal hsa_circ_0020792 on the migration of NFs, transwell assay was performed. The results showed that keloid patient-Exo notably promoted the migration of NFs compared to healthy control-Exo (Figure 4A). However, siR-circ-2 abolished the pro-migratory effect of keloid patient-Exo on NFs (Figure 4A). In addition, compared to healthy control-Exo, keloid patient-Exo remarkably increased the levels of

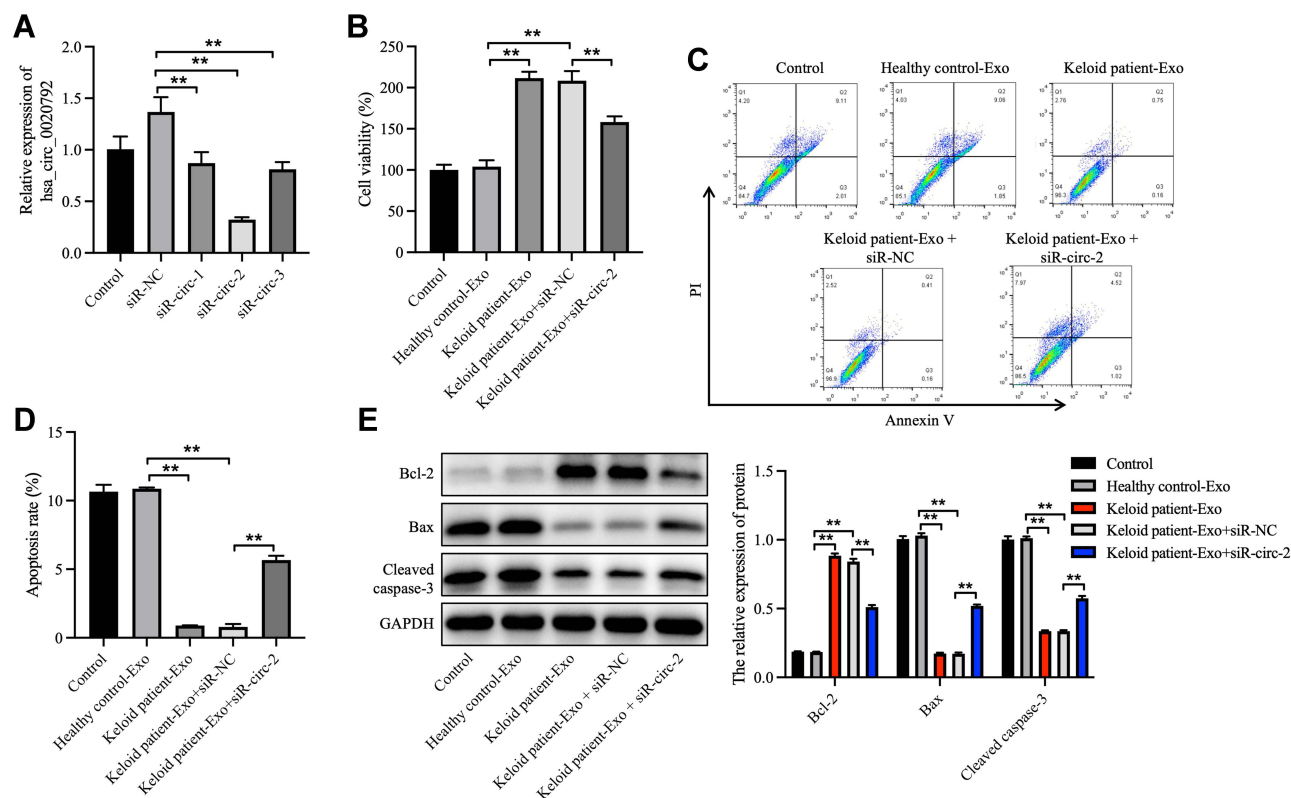


Figure 3 Keloid patient-derived exosomal hsa_circ_0020792 enhances the viability and reduces the apoptosis in normal skin fibroblasts. **(A)** Normal skin fibroblasts (NFs) were transfected with siR-NC, siR-circ-1, siR-circ-2 or siR-circ-3. The level of hsa_circ_0020792 in NFs was detected using RT-qPCR. **(B)** NFs were treated with healthy control-Exo, keloid patient-Exo, keloid patient-Exo + siR-NC and keloid patient-Exo + siR-circ-2. The viability of NFs was detected using CCK-8 assay. **(C and D)** The apoptosis of NFs was assessed using flow cytometry assay. **(E)** The expressions of Bcl-2, Bax and cleaved caspase 3 in NFs were detected using Western blot assay. ** $P < 0.01$, $n = 3$.

Collagen I, Collagen III, α -SMA, TGF- β 1, p-Smad2 and p-Smad3 in NFs, whereas these increases were reduced by siR-circ-2 (Figure 4B and C). These data indicated that keloid patient-derived exosomal hsa_circ_0020792 could promote the migration and fibrogenesis of NFs via activating TGF- β 1/Smad2/3 signaling.

Knockdown of hsa_circ_0020792 Inhibits the Viability, Migration, and Fibrogenesis of NFs via Upregulating miR-193a-5p

To clarify the effect of hsa_circ_0020792 and miR-193a-5p on the cell growth, apoptosis, migration and collagen synthesis on NFs, siR-NC or siR-circ-2 and miR-inhibitor or miR-NC were transfected into NFs. As illustrated in Figure 5A and B, siR-circ-2 significantly increased miR-193a-5p level in NFs, whereas miR-inhibitor obviously declined miR-193a-5p level in NFs. Besides, downregulation of hsa_circ_0020792 by siR-circ-2 notably reduced the viability and triggered the apoptosis of NFs, whereas these phenomena were abolished by miR-193a-5p inhibition (Figure 5C–E). Moreover, knockdown of hsa_circ_0020792 markedly downregulated the expressions of Bcl-2 and upregulated the expressions of Bax and cleaved caspase 3 in NFs, whereas these phenomena were reversed in the presence of miR-inhibitor (Figure 5F). Meanwhile, hsa_circ_0020792 knockdown notably suppressed the migration of NFs, whereas miR-inhibitor was able to reverse this effect (Figure 6A). Moreover, downregulation of hsa_circ_0020792 significantly reduced the level of Collagen I, Collagen III, α -SMA, TGF- β 1, p-Smad2 and p-Smad3 in NFs, whereas these phenomena were reversed by miR-193a-5p inhibition (Figure 6B and C). Collectively, knockdown of hsa_circ_0020792 could reduce the viability, migration and fibrogenesis and induce the apoptosis of NFs via upregulating miR-193a-5p.

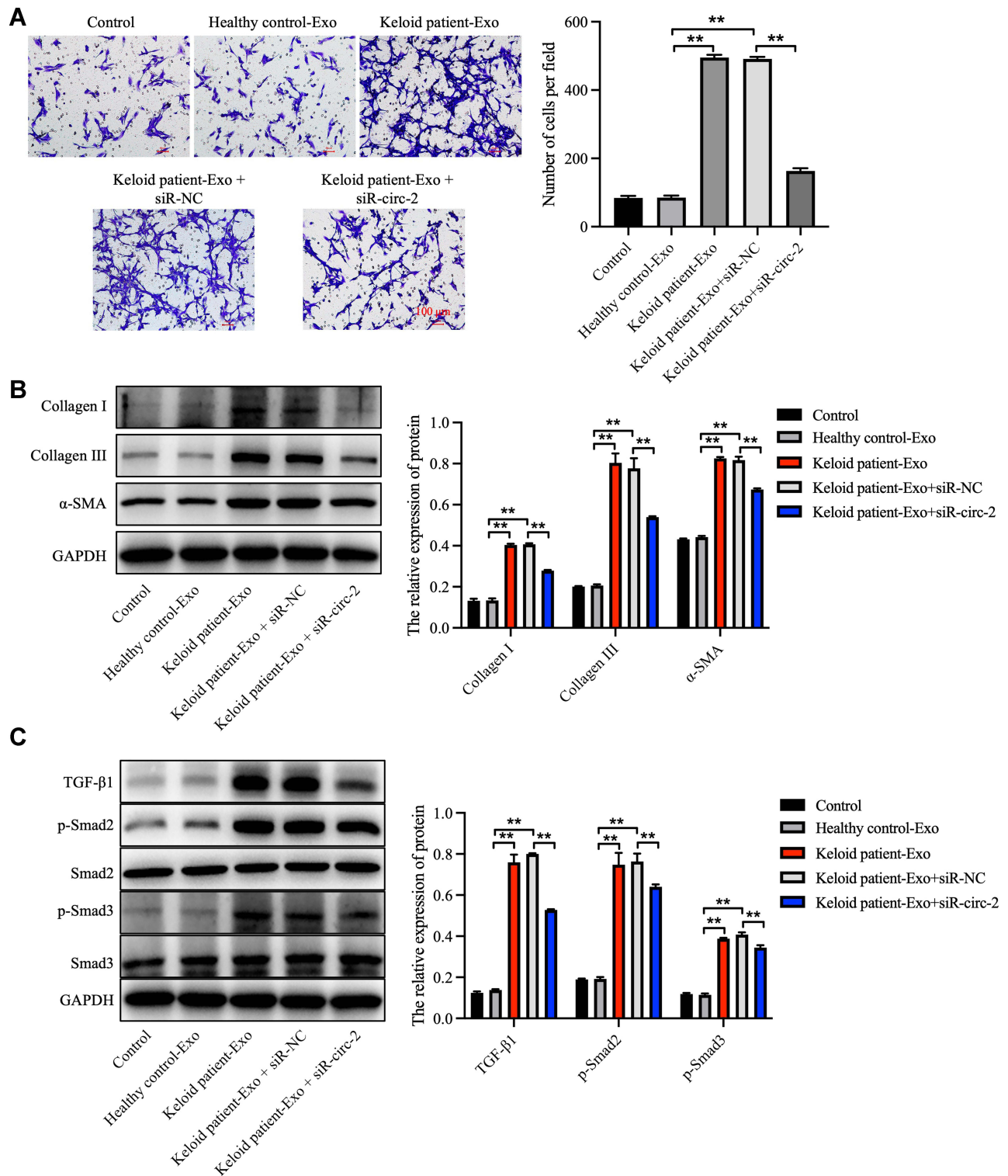


Figure 4 Keloid patient-derived exosomal hsa_circ_0020792 promotes the migration and ECM synthesis of normal skin fibroblasts via activating TGF-β1/Smad2/3 signaling. Normal skin fibroblasts (NFs) were treated with healthy control-Exo, keloid patient-Exo, keloid patient-Exo + siR-NC and keloid patient-Exo + siR-circ-2. **(A)** The migratory ability of NFs was detected using transwell assay. **(B, C)** The expressions of Collagen I, Collagen III, α-SMA, TGF-β1, p-Smad2, Smad2, Smad3 and p-Smad3 in NFs were detected using Western blot assay. **P < 0.01, n = 3.

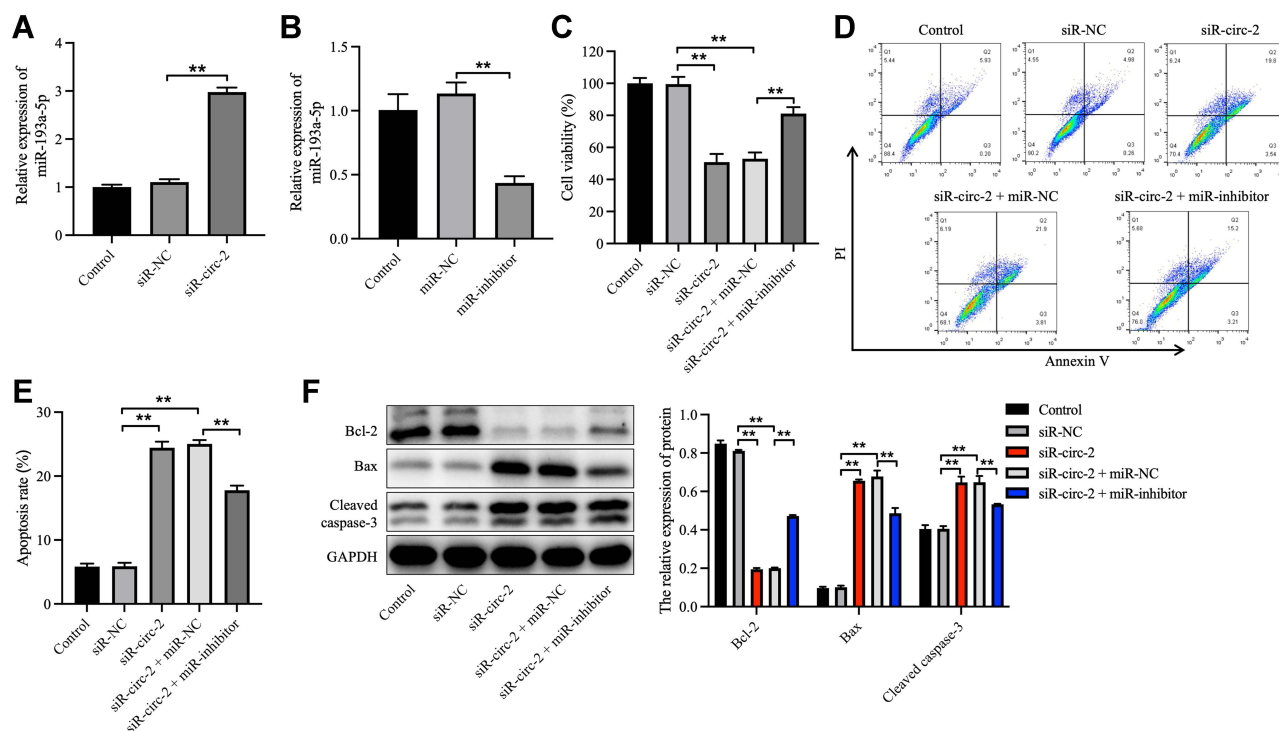


Figure 5 Knockdown of *hsa_circ_0020792* reduces the viability and increases the apoptosis of normal skin fibroblasts via upregulating miR-193a-5p. **(A)** The miR-193a-5p level in normal skin fibroblasts (NFs) transfected with siR-NC or siR-circ-2 was detected using RT-qPCR. **(B)** The miR-193a-5p level in NFs transfected with miR-NC or miR-inhibitor was detected using RT-qPCR. **(C)** NFs were transfected with siR-NC, siR-circ-2, or siR-circ-2 + miR-NC or siR-circ-2 + miR-inhibitor. The viability of NFs was detected using CCK-8 assay. **(D and E)** The apoptosis of NFs was measured using flow cytometry assay. **(F)** The expressions of Bcl-2, Bax and cleaved caspase 3 in NFs were determined using Western blot assay. ***P* < 0.01, *n* = 3.

Knockdown of *hsa_circ_0020792* Inhibits the Viability and Induces the Apoptosis of Keloid Fibroblasts via Upregulating miR-193a-5p

To further confirm the effect of *hsa_circ_0020792* and miR-193a-5p on the growth of keloid fibroblasts, siR-circ-1, siR-circ-2 or siR-circ-3 was transfected into keloid fibroblasts. As revealed in [Supplementary Figure 3A](#), siR-circ-2 sharply reduced *hsa_circ_0020792* level in keloid fibroblasts. In addition, downregulation of *hsa_circ_0020792* notably reduced the viability and induced the apoptosis of keloid fibroblasts, whereas these phenomena were reversed by miR-193a-5p inhibitor ([Supplementary Figure 3B and C](#)). To sum up, knockdown of *hsa_circ_0020792* could inhibit the viability and induce the apoptosis of keloid fibroblasts via upregulating miR-193a-5p.

Discussion

It has been shown that peripheral circulating blood exosomes are involved in the pathogenesis of various diseases including skin diseases.^{26–28} CircRNAs have been found in exosomes derived from human blood samples with clinical implications.²⁹ In addition, circRNAs exert key roles in the occurrence and development of keloid.³⁰ Jiao et al reported that circCOL5A1 was able to suppress the proliferation, migration and ECM production of keloid fibroblasts through targeting miR-877-5p.³¹ Gao et al found that *hsa_circ_0057452* could facilitate keloid progression by targeting miR-1225-3p.³² In addition, Shi et al found that *hsa_circ_0020792* level was abnormally upregulated in the keloid tissues compared to healthy skin tissues.¹⁷ In this study, we found that *hsa_circ_0020792* level was increased in exosomes derived from keloid patients compared to exosomes from healthy controls, which was consistent with the previous study. However, the role of exosomal *hsa_circ_0020792* in the formation and development of keloids is not well understood.

The excessive ECM production and abnormal increase in fibroblast proliferation and migration are risk factors in the development of keloid.^{31,33} In addition, Xie et al found that keloid-derived fibroblasts expressed high levels of ECM proteins collagen I and collagen III,³⁴ which was consistent with our results. Furthermore, we found that keloid patient-

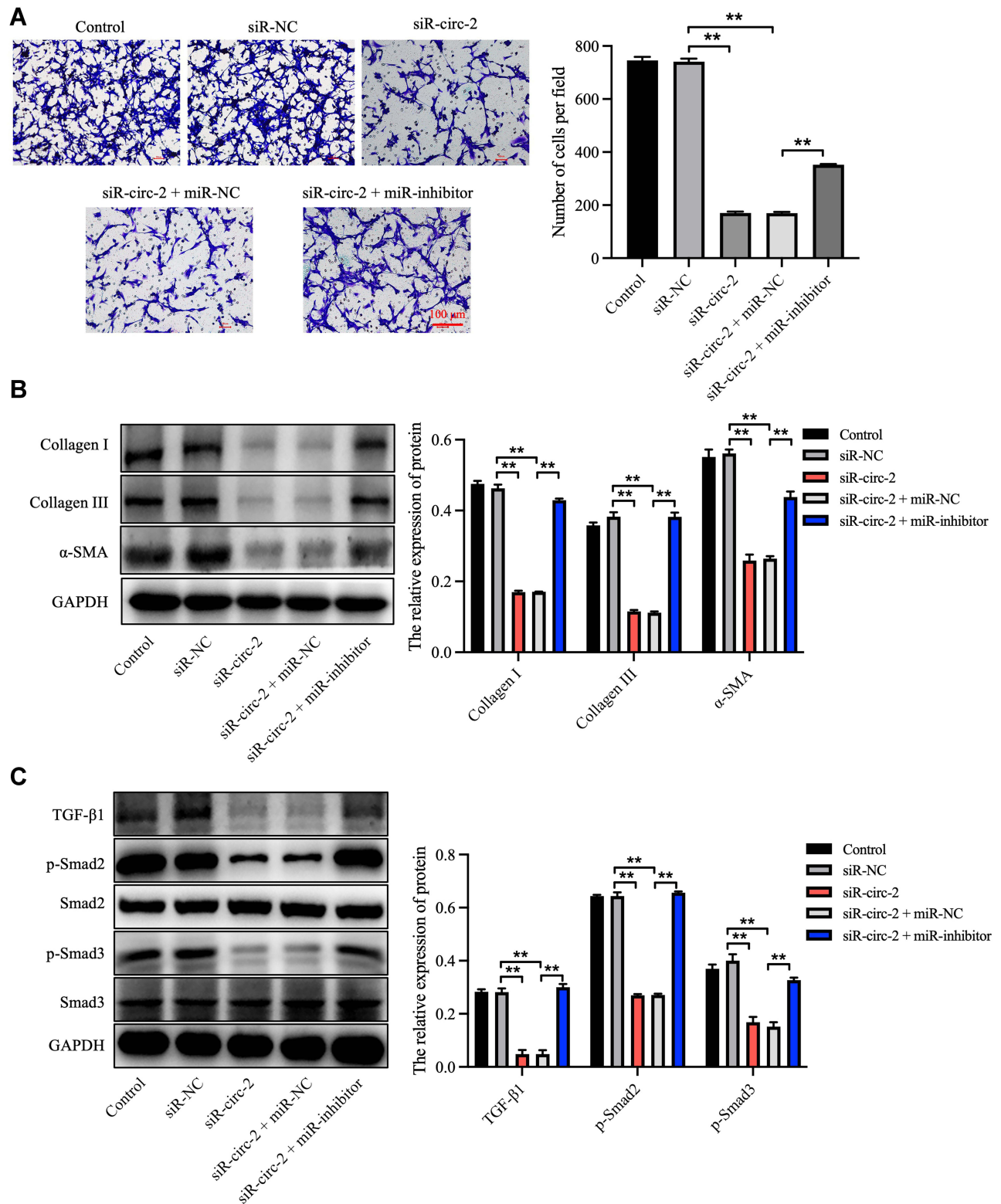


Figure 6 Knockdown of hsa_circ_0020792 inhibits the migration and the fibrogenesis of normal skin fibroblasts by upregulating miR-193a-5p. Normal skin fibroblasts (NFs) were transfected with siR-NC, siR-circ-2, or siR-circ-2 + miR-NC or siR-circ-2 + miR-inhibitor. (A) The migratory ability of NFs was assessed using transwell assay. (B and C) The level of Collagen I, Collagen III, α -SMA, TGF- β 1, p-Smad2, Smad2, Smad3 and p-Smad3 in NFs were detected using Western blot assay. **P < 0.01, n = 3.

derived exosomal hsa_circ_0020792 obviously promoted the viability, migration and inhibited the apoptosis of NFs. Moreover, keloid patient-derived exosomal hsa_circ_0020792 notably accelerated the synthesis of ECM in NFs, as characterized by the increased levels of collagen I and collagen III. Meanwhile, α -SMA, a myofibroblast marker, exerts an important role in fibrogenesis.³⁵ Our results showed that keloid patient-derived exosomal hsa_circ_0020792 obviously elevated α -SMA expression in NFs, suggesting that exosomal hsa_circ_0020792 promoted fibrogenesis in NFs. These results showed that hsa_circ_0020792-enriched exosomes derived from keloid patients were able to endow normal skin fibroblast cells with the biological properties of keloid fibroblasts.

Evidence has shown that TGF- β /Smads signaling plays vital roles in promoting the proliferation, collagen synthesis and fibrogenesis in keloid fibroblasts.^{36,37} Activation of TGF- β /Smads pathway could promote the keloid development.³⁸ Chen et al showed that circ_0008450 knockdown could suppress the proliferation of human keratinized epithelial cells via inactivating the TGF- β /Smad2/3 signaling.³⁹ In this study, we found that keloid patient-derived exosomal hsa_circ_0020792 remarkably upregulated the expressions of TGF- β 1, p-Smad2 and p-Smad3 in NFs, whereas these changes were reduced by hsa_circ_0020792 knockdown. Collectively, hsa_circ_0020792-enriched exosomes derived from patients with keloids could promote the proliferation, migration, collagen synthesis and fibrogenesis of NFs via activating TGF- β 1/Smad2/3 signaling.

It has been shown that miR-193a is associated with skin-related diseases.^{40,41} Polini et al showed that miR-193a-5p could exert a tumor suppressor role on cutaneous melanoma.⁴² However, the role of miR-193a-5p in keloid development remains unclear. In this study, our results showed that miR-193a-5p level was reduced in keloid patient-derived exosomes and keloid fibroblasts. Inhibition of miR-193a-5p was able to reverse the effect of hsa_circ_0020792 knockdown on the proliferation and migration of NFs, suggesting that downregulation of hsa_circ_0020792 could suppress the viability and migration of NFs via upregulating miR-193a-5p. Additionally, the inhibitory effects of hsa_circ_0020792 knockdown on the ECM synthesis and fibrogenesis of NFs were reversed by miR-193a-5p inhibitor, as shown by the increased levels of collagen I, collagen III, TGF- β 1, p-Smad2 and p-Smad3, suggesting that hsa_circ_0020792 knockdown could suppress the ECM synthesis and fibrogenesis of NFs via upregulating miR-193a-5p. In pancreatic cancer, miR-193a was able to regulate tumor metastasis via targeting TGF- β 2/SMAD2/3 signaling.⁴³ Our results showed that key factors participated in TGF- β 1/Smad2/3 pathway could be affected by hsa_circ_0020792 and miR-193a-5p. Furthermore, we further confirmed that hsa_circ_0020792 knockdown could inhibit the viability and induce the apoptosis of keloid fibroblasts via upregulating miR-193a-5p. These data showed that hsa_circ_0020792 could affect the progression of keloid via targeting miR-193a-5p.

Conclusion

In conclusion, keloid patient-derived exosomal hsa_circ_0020792 could promote normal skin fibroblasts proliferation, migration, and fibrogenesis via modulating miR-193a-5p and activating TGF- β 1/Smad2/3 signaling. These results showed that hsa_circ_0020792 could affect keloid progression by miR-193a-5p/TGF- β 1/Smad2/3 axis, suggesting a novel hsa_circ_0020792/miR-193a-5p/TGF- β 1/Smad2/3 pathway underlying keloid treatment.

Data Sharing Statement

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics Approval and Consent to Participate

The study complies with the declaration of Helsinki and was proved by Ethics Committee of Shanghai Sixth People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine (S2017-0023). The written informed consents have been obtained from patients and healthy donors.

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Disclosure

The authors declare that there are no conflicts of interest.

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