



Review

Pediatric Population with Down Syndrome: Obesity and the Risk of Cardiovascular Disease and Their Assessment Using Omics Techniques—Review

Marta Hetman * and Ewa Barg

Department of Basic Medical Sciences, Wrocław Medical University, 50-556 Wrocław, Poland

* Correspondence: martha.hetman@gmail.com

Abstract: People with Down syndrome (PWDS) are more at risk for developing obesity, oxidative stress disorders, metabolic disorders, and lipid and carbohydrate profile disorders than the general population. The presence of an additional copy of genes on chromosome 21 (i.e., the superoxide dismutase 1 gene (SOD1) and gene coding for the cystathionine β -synthase (CBS) enzyme) raises the risk for cardiovascular disease (CVD). As a result of disorders in metabolic processes and biochemical pathways, theoretically protective factors (low homocysteine level, high SOD1 level) do not fulfil their original functions. Overexpression of the CBS gene leads to the accumulation of homocysteine—a CVD risk factor. An excessive amount of protective SOD1, in the case of a lack of compensatory increase in the activity of catalase and peroxidase, leads to intensifying free radical processes. The occurrence of metabolic disorders and the amplified effect of oxidative stress carries higher risk of exposure of people with DS to CVD. At present, classic predispositions are known, but it is necessary to identify early risk factors in order to be able to employ CVD and obesity prophylaxis. Detailed determination of the metabolic and lipid profile may provide insight into the molecular mechanisms underlying CVD.



Citation: Hetman, M.; Barg, E. Pediatric Population with Down Syndrome: Obesity and the Risk of Cardiovascular Disease and Their Assessment Using Omics Techniques—Review. *Biomedicines* **2022**, *10*, 3219. <https://doi.org/10.3390/biomedicines10123219>

Academic Editor: Estefania Nuñez

Received: 21 November 2022

Accepted: 9 December 2022

Published: 12 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: Down syndrome; obesity; metabolomics; lipidomics; cardiovascular disease

1. Introduction

Diagnosing a disease before its first symptoms appear poses a great challenge in medicine. In the past, it was believed that only genes affected the maintenance of homeostasis and health in the body. After many years of research and genome sequencing, we are still unable to diagnose numerous diseases or to design an effective therapy at an early stage. It was assumed that the organism is a complex structure, at the base of which is the genome, and its subsequent levels being transcripts, proteome and metabolome [1]. Metabolism plays a key role in all areas of biology, which is why more and more of those areas are being studied from its perspective. Many adult conditions, such as cardiovascular and metabolic disease, originate during childhood, therefore the knowledge of risk factors for these diseases will allow for the implementation of an early and effective prophylaxis.

Down syndrome (DS, trisomy 21, T21) is the most common chromosome abnormality (caused by trisomy of the whole or a part of chromosome 21) with a worldwide incidence rate of 1:1000–1100 in newborns [2]. The extra chromosome 21, or at least a portion of it, results in a constellation of clinical features (cardiac defects, delayed growth, hematology and endocrine abnormalities, autoimmune diseases, intestinal, stomatognathic disturbances, vision and hearing defects and obstructive sleep apnea, and others) [3,4]. Additionally, people with DS (PWDS) are at increased risk for cardiovascular diseases (CVD) (mitral valve prolapse, endocarditis, atherosclerosis (AS) and congestive heart failure [5]), pulmonary hypoplasia, muscle hypotonia, osteoporosis, arthritis, osteoarthritis, and diabetes mellitus [6,7]. There is no specific DS phenotype. Individuals may differ from each other

both in terms of external features and chronic diseases. Such a complex condition contributes to the demand for profound medical care. Advancements in medicine have led to a marked improvement in life expectancy of PWDS, with the estimated median age of survival approaching 60 years [8]. Obesity-related diseases (CVD, cancer, type II diabetes, and others) have received more attention [9]. The problem of body weight disorders among PWDS is complex and challenging, concerning mainly the rapid transformation between undernutrition in the first period of life and excessive weight gain in later years. For the purposes of this review, however, we will focus on the problems of excess body weight in children and adolescents with DS. The appearance of new disease entities in this population is a challenge for health practitioners. Due to the burden of many conditions, PWDS should be monitored from an early age with the constraints associated with their health status.

2. Omics Techniques: Metabolomics and Lipidomics

Omics techniques are a rapidly evolving field of molecular sciences. Metabolomics, a relatively young branch of omics science, is an interdisciplinary approach that encompasses biology, chemistry, and bioinformatics. The metabolomic examination should enable the detection of abnormalities in the patient's health at an early stage of the development of clinical symptoms or even before their manifestation [10]. The advantage of metabolomics research is its low invasiveness, thanks to the use of mainly readily available body fluids, i.e., blood serum, plasma, saliva, urine, and tissues after prior preparation. It provides a modern bioanalytical tool to define perturbations in metabolic pathways and enables the detection of predictive biomarkers. Thus, metabolomics can contribute to more efficient diagnosis, treatment, and disease prevention. The definition of metabolomics officially appears in the literature in 1999 as “the quantitative measurement of the dynamic multi-parametric metabolic response of living systems to pathophysiological stimuli or genetic modification” [11]. Metabolomics is based on the qualitative and quantitative study of small-molecule (<1.5 kDa) compounds that are intermediates and products of metabolism (lipids, amino acids, short peptides, nucleic acids, sugars, alcohols, or organic acids) and reflects in endogenous metabolism and exogenous sources such as, among others, diet or physical activity. Metabolites are involved in all biochemical reactions (any process that occurs in the body is reflected in the metabolome) and measuring them can potentially evaluate the state of the organism. There are two approaches in metabolomics: untargeted and targeted. The untargeted approach makes it possible to identify metabolic new biomarkers; the targeted approach identifies and quantifies a limited number of known metabolites. Both the presence and absence of specific metabolites can be the source of information about possible disorders in the patient's health and draw attention to a medical problem.

Lipid homeostasis is essential for maintaining full health; therefore its evaluation is of fundamental importance. Any abnormalities in lipid metabolism play an important role in many diseases, including metabolic syndrome, diabetes, CVD, lipodystrophies, neurological/neurodegenerative disorders, and central nervous system damage [12]. Most cases of CVD are difficult to associate with well-known risk factors. Many patients, despite having optimal blood lipid levels, are exposed to CVD [13]. Therefore, it is important to search for new biomarkers that will enable the very early diagnosis and effective prevention of CVD. Single biomarkers in cardiology are very effective in confirming the occurrence of an acute event. However, it is very difficult to precisely estimate the risk of atherosclerotic disease at an early stage. So far, researchers have relied on well-established risk factors, such as smoking, hypertension, dyslipidemia, and diabetes as risk factors of developing CVD [12,14–16]. Detailed determination of the metabolic profile may provide insight into the molecular mechanisms underlying AS [13,17–20].

2.1. Metabolomics

Metabolomics is a very promising tool for investigating human health, however, the analysis of the metabolome is challenging for many reasons, among others, different analytic approaches and the lack of standardization. The techniques used in metabolomics are

magnetic resonance (NMR) spectrometers, mass spectrometers (MS), gas-chromatography (GC), liquid chromatography (LC) systems, ion mobility systems (IMS), capillary electrophoresis (CE) systems, integrated liquid chromatography-mass spectrometry (LC-MS), integrated capillary electrophoresis-mass spectrometry (CE-MS), integrated ion mobility spectrometry-mass spectrometry (IMS-MS) and gas chromatography-mass spectrometry (GC-MS) [21].

As a novel technique, metabolomics can provide insight into obesity and the risk of cardio-metabolic complications, and be used to uncover pathways underlying diet–disease associations. The heart, being a metabolically active organ [22], and its diseases constitute one of the main targets of the omics’ techniques of today. The results of metabolomics studies help to clarify the pathophysiology of many diseases, optimize treatment, and distinguish specific diagnostic biomarkers, which is especially important in the case of asymptomatic diseases [23]. In 1991, the first biomarkers of coronary heart disease using NMR spectroscopy [24] were discovered. The introduction of metabolomics to epidemiological research is of great importance for the understanding of pathophysiology and the discovery of new biomarkers for the early prevention and detection of AS and CVD [25]. In 2017, the American Heart Association published a statement on potential health and CVD effects of metabolomics and its current challenges in clinical practice [26].

Understanding the pathogenesis of childhood obesity with the help of molecular studies is one of the major challenges of current medicine. Metabolomic studies on obesity and comorbidities are conducted in adults on a large scale. Unfortunately, little research has been completed on groups of children and adolescents [27]. Hellmuth et al. combined metabolomics data from four large European cohorts finding a strong positive association of sphingomyelin (SM) 32:2 (molecular species containing myristic acid and sphingadienine) and lyso-phosphatidylcholine (LPC) 14:0 with BMI z-score (this metabolite was found to have a positive association with BMI adults [28]) and no association of non-esterified fatty acid (NEFA) 16:1 with BMI z-score [29]. An LPC 14:0 was considered a predictor of obesity at 6 years of age (study of serum in 6-month-old infants) [30]. The rate of FA 14:0 was also elevated in phospholipids (study among obese 15-year-old children) [31]. The authors [29] additionally concluded that the concentration of lipids with 14:0 (exception of NEFA 14:0) is seemingly higher in children with high BMI and may subsequently be used more often for the synthesis of SM. In addition, the 14:0 synthesis can be enhanced by a high-calorie diet and high glycemic load of food. SM 32:2 may be a potential biochemical marker for the combined effect of genetic predisposition, high dietary intake of total energy, glycemic load, and linoleic acid [29]. Atherosclerosis, the major cause of CVD, is often attributed to lifestyle factors [32]. A high risk of the early development of AS has been proved in people with hyperhomocysteinemia, hypermethioninemia, and homocystinuria [33]. Wurtz et al. identified phenylalanine and various fatty acids as biomarkers for CVD [34]. Biomarkers related to insulin resistance and energy metabolism have also already been identified [35]. A consistent metabolic profile of childhood obesity was observed including amino acids (particularly branched chain and aromatic), carnitines, lipids, and steroids [36,37].

2.2. Lipidomics

Lipidomics, a discipline belonging to metabolomics, is described as the quantitative characteristic of the complete lipid complex [17]. The subject of research in this subdiscipline is lipids, i.e., a functional unit characterizing the molecular lipid image of a biological sample under study. However, thanks to lipidomics, it is possible to quantify various lipid molecules (acylglycerols, sterols, sphingolipids, and others) [17,19]. Lipidomic evaluation allows for a picture of lipid concentrations, for example, the total plasma lipidomics of the tested total plasma shows a detailed and much more complete picture of lipid metabolism and possible abnormalities of lipid metabolism—as opposed to studies of isolated lipoproteins [12]. Lipidomics had identified ceramides and sphingolipids as potential mediators of cellular dysfunction.

Lipidomics study the structural, signaling and metabolic functions of lipid compounds. Due to the large variety of lipid types in cells/tissues, the lack of homogeneity and their frequent biochemical modifications, detailed characterization may be a difficult task [38]. For these reasons, data on lipidomics rarely appear in the scientific literature when compared with other “omics” technologies. Thus, a sufficient amount of data on the relationship between lipid metabolites with CVD are still lacking [39]. Lipids are presented as CVD risk factors only in major classes and not as individual molecular entities in diabetes [40].

The main challenge of lipidomics is the demonstration of new risk factors and the early detection of the risk of atherogenesis at the clinical level. It seems necessary to identify early risk factors in order to undertake CVD prophylaxis.

3. Down Syndrome

3.1. Cardiovascular Disease

In 1977, Murdoch et al. found a complete absence of atherosclerosis in five posthumously examined PWDS [41]. In addition, Pueschel et al. confirmed the lack of significant differences in the level of total cholesterol (TC), low-density lipoprotein (LDL), apolipoprotein B (apoB), and apoB/apolipoprotein A (apoA) between the examined group of PWDS and the control group [42]. As a result of conducted research in the 1970's, it was thought that people with DS were no more at risk of atherosclerosis than the general population. However, the lifestyle and eating habits differed significantly from those present, and life expectancy was much shorter. Recently, there have been scientific studies that may suggest that significant and known risk factors for CVD and AS have been observed among people with DS: diabetes [43,44], obesity [43,45–47] and hypertension [44], and lipid disorders [48,49]. At the same time, it has been shown that PWDS have a lower incidence of AS [41,44,50–56]. Additionally, lower BPs at rest may have a protective role against the development of atherosclerosis in PWDS [53,57].

Interestingly, Landes et al. showed that PWDS were more likely to die at younger ages from heart diseases compared with the general population [58]. The study of Hill et al., Day et al., and Hermon et al. showed an increased risk of death for PWDS due to CVD in comparison with the general population [59–61]. However, Torr et al. analyzed morbidity and mortality among PWDS and indicated ischemic heart disease to be a minor cause of death [62]. Adelekan et al. found that children with DS have less favorable lipid profiles than their siblings [8]. Sheela et al. showed that youth with DS had more atherogenic lipid and lipoprotein particle profiles, including higher LDL-C levels, compared with those without DS [49]. Buonouomo et al. found high levels of TC, LDL-C, and TG and low HDL-C in individuals aged 2–9 years old with DS [62]. This study group with DS also had a higher prevalence of prediabetes and an increased amount of visceral fat [49]. In general, the increased LDL-C level in youth with DS reveals a greater risk of atherosclerosis. Adults with DS also have a high risk of stroke, driven largely by high cardioembolic risk [44].

Lipoprotein(a) (Lp(a)) seems to be involved in the pathogenesis of CVD [63]. Krześcińska et al. compared lipid parameters, protein composition, antioxidative properties of HDL, and Lp(a) levels in adolescents with DS and healthy individuals [64], and showed unfavorable lipid profiles in conjunction with significantly higher Lp(a) levels and quality changes in HDL particles in adolescents with DS. Serum Lp(a) levels are relatively stable over a lifetime [65], therefore a once-in-a-lifetime Lp(a) measurement could help identify those at increased risk of CVD [66]. Data appearing in the literature seem to be contradictory. Most, however, argue for the need to refute the belief that DS is a disease free from atherosclerosis. In this situation, it is advisable to extend the research on atherosclerosis risk factors and predisposition to related diseases in people with DS with the use of omics techniques.

3.2. Excessive Body Weight and Physical Activity

The literature repeatedly reports that DS children are predisposed to obesity [67–72], abnormal or excessive fat accumulation caused by a positive energy balance, which has

been associated with a negative impact on health [73]. Adults with DS are twice as likely to be obese and nearly four times more likely to be extremely obese in comparison with adults without DS [74]. The literature describes potential causes of obesity tendency among children and adolescents with DS: decreased energy expenditure at rest; increased leptin levels; untreated hypothyroidism; unhealthy diet; and low physical activity [70,75]. Additionally, children and adolescents with DS show less physical activity than their peers without DS [76–78], although the tested level of physical activity in adolescents without DS turned out to be insufficient in 80% of them [79]. What is worse, PWDS tend to become less active as they become older, with higher rates of obesity in girls [80–84]. Although the activity level among children with DS was lower, the caloric intake was higher in this group [75]. The greatest acceleration in obesity occurs between the ages of 2–6 years [84]. In the teenage period, when PWDS gain more independence and the ability to choose the type and amount of food (with the predominance of processed products with excessive amount of salt and sugar), obesity begins to be most visible. Yahia et al. pointed out that prepubertal obese-DS displayed excess body adiposity with pronounced central fat distribution, atherogenic lipid profile, and higher insulin resistance compared with matched obese-control [85,86]. Wernio et al. pointed out that overweight children with DS were characterized by higher levels of triglycerides, atherogenic index of plasma, and apoA2 and apoE levels [87]. Obesity also contributes to the worsening of obstructive sleep apnea symptoms and the burden of congenital heart disease [70,88,89]. With age, it becomes more and more difficult to persuade teenagers to play sports regularly. However, the DONUT STUDY showed that, despite potential difficulties in the pursuit of a correct diet and inadequate approach to physical activity, children with DS could achieve results that are substantially the same as those of non-DS children [90]. Moreover, some children and adolescents with DS are limited by reduced respiratory efficiency and congenital heart diseases [81]. An additional obstacle to increasing physical activity among PWDS is the COVID-19 epidemic that has been present for over 2 years. Amatori et al. showed a negative impact of COVID-19: decreased physical activity and increased sedentary behaviors [91]. It is worth remembering that the patterns of proper nutrition should function throughout a household. Stefanowicz-Bielska et al. proved that in families of overweight and obese children with DS, other members had nutritional disorders more frequently [92]. Caregivers and siblings should be equally involved in shaping healthy habits and lifestyle. Different levels of intellectual disability can also make it difficult to make correct food choices. Hence the repeated emphasis on the importance of the role of the family as a promoter of a healthy lifestyle. Roccatello et al. analyzed meals of choice of the people with DS finding bread, pasta and sweets as their favorite go-to foods [92]. The least-liked food was vegetables. Fruit juices and ready-to-drink tea were the main sources of simple sugars [92], which can contribute to liver steatosis and hypertension (the impact of fructose) [75]. Introducing healthy eating habits may be fundamental to sustaining good health. Jobling et al. conducted an intervention study (education program) [93]. The program was successful in convincing people with DS to reduce their consumption of sweets but the researchers' actions did not change other unhealthy eating habits [93]. However, Naczka et al. enrolled adolescents with DS in a thirty-three weeks swimming program that resulted in decreases in body mass, body fat, and BMI [94]. Because regular physical activity is recommended to reduce the risk of developing health conditions such as heart disease, cancer, type 2 diabetes, high blood pressure, osteoporosis, and obesity [95,96], sports programs of this type play a very important role in acquired heart-disease prevention. As children and adolescents with DS are predisposed to overweight and obesity, and also tend to be physically inactive, they are at a significant risk of mortality and many serious diseases. Me et al. have shown that breastfeeding may be a protective factor for obesity and high body fat in children [97]. In 2022, a systematic review of DS and breastfeeding was conducted: around 50–23.3% of the children with DS were never breastfed and rates of breastfeeding in infants with DS were lower than those in controls in three studies [98].

3.3. Oxidative Stress

PWDS have been identified as having high oxidative stress (the imbalance between free radical production and the prooxidative state within the cell determines a biological state [99]), which is connected with the risk of the development of AS, neurodegeneration, cell ageing, cancer, and immunological disorders [100,101]. Oxidative stress, which damages blood vessel tissues, also plays a role in the pathogenesis of AS. In oxidatively damaged tissues, the development of AS is facilitated [102]. Endothelial cell function may be impaired in PWDS despite their protection against AS [103]. Furthermore, high oxidative stress has been related to elevated insulin resistance, poor insulin sensitivity, and hypertension [104]. It has been shown that T21 is associated with pro-oxidant status and increased susceptibility to oxidative damage [105–107]. T21 of the chromosome increases the representation and expression of Cu/Zn superoxide dismutase (SOD1), the gene which is located on the distal segment of chromosome 21 (21q22.1) [108]. It has been shown that PWDS have an increased SOD1 activity by as much as 150% compared with people without DS [109]. SOD-1 is the main enzyme in the antioxidant defense system. Under physiological conditions SOD-1, together with catalase and peroxidases, protects the body against the harmful effects of very reactive free oxygen radicals which are a potential threat to cellular structures. Radicals and reactive oxygen species (ROS) are formed during normal cellular metabolism, however, in the conditions of their increased formation or disturbances in the activity of antioxidant enzymes, free radical damage occurs. It is believed that excessive SOD-1 activity is responsible for the increased formation of hydrogen peroxide and it heightens the risk of oxidative stress (prolonged higher SOD activity may lead to glutathione depletion, deficiencies in catalase and peroxidases' activity). SOD-1 catalyzes the conversion of the peroxide anion to hydrogen peroxide, which leads to the continuous production of two major reactive oxygen species in oxygen cells in the mitochondria [109]. In the pathogenesis of AS, ROS are responsible for the formation of oxidatively modified LDLs (oxLDL), which are pro-atherogenic substances [110–112]. The biological effects of ROS are controlled by a wide spectrum of antioxidant defense mechanisms such as the action of vitamins E and C, uric acid, glutathione, and antioxidant enzymes. The reduced concentration of glutathione in the blood along with the overexpression of the SOD-1 gene in PWDS additionally contributes to elevated exposure to the negative effects of oxidative stress. The increased activity of SOD-1 as an antioxidant enzyme could explain the protective role in preventing atherosclerotic lesions. In the situation of disturbed antioxidant balance, as is the case in DS, due to the lack of compensatory higher activity of catalase and peroxidase, free radical processes are intensified. It is known that high SOD-1 activity means a disturbed balance of the antioxidant system: the peroxidation processes of lipid peroxides, participating in the formation of atherosclerotic plaques, dominate. In people who experience increased oxidative stress, biologically important molecules such as lipids, proteins, or nucleic acids are oxidized, which significantly affects the incorrect function of both individual organs and the entire body. Therefore, it seems that PWDS will be additionally exposed [111]. Lipid peroxidation (LPO) in free radical reactions is the process of oxidation of unsaturated fatty acids or lipids in which peroxides of these compounds are formed. They are an important link in the atherosclerotic process [112]. They modify physical properties of cell membranes and inhibit the activity of membrane enzymes and transport proteins. There is also a link between an aerobic modification of LDL-C and inflammatory activity of macrophages through the induction of macrophage cyclooxygenase 2 expression by LPO products [110]. Chronic oxidative stress leads to the intensification of degenerative processes and premature aging of tissues. With age, numerous hormonal and metabolic disorders appear and worsen, which is the leading problem in older children and adolescents with DS who require constant and targeted medical care. An earlier description of DS as a “non-atherosclerotic model” could be justified, *inter alia*, by increased activity of the defense enzyme SOD-1 and the altered metabolism of homocysteine. The results of the current research seem to contradict this assumption. In this situation, it is advisable to extend the research workshop on atherosclerosis risk factors and predisposition to other

diseases in people with DS to include metabolomics research. Wernio et al. pointed out that fat mass, fat mass/height² index, and visceral fat mass in children with DS correlated with advanced oxidative protein product level [87]. Figure 1 shows the processes described above.

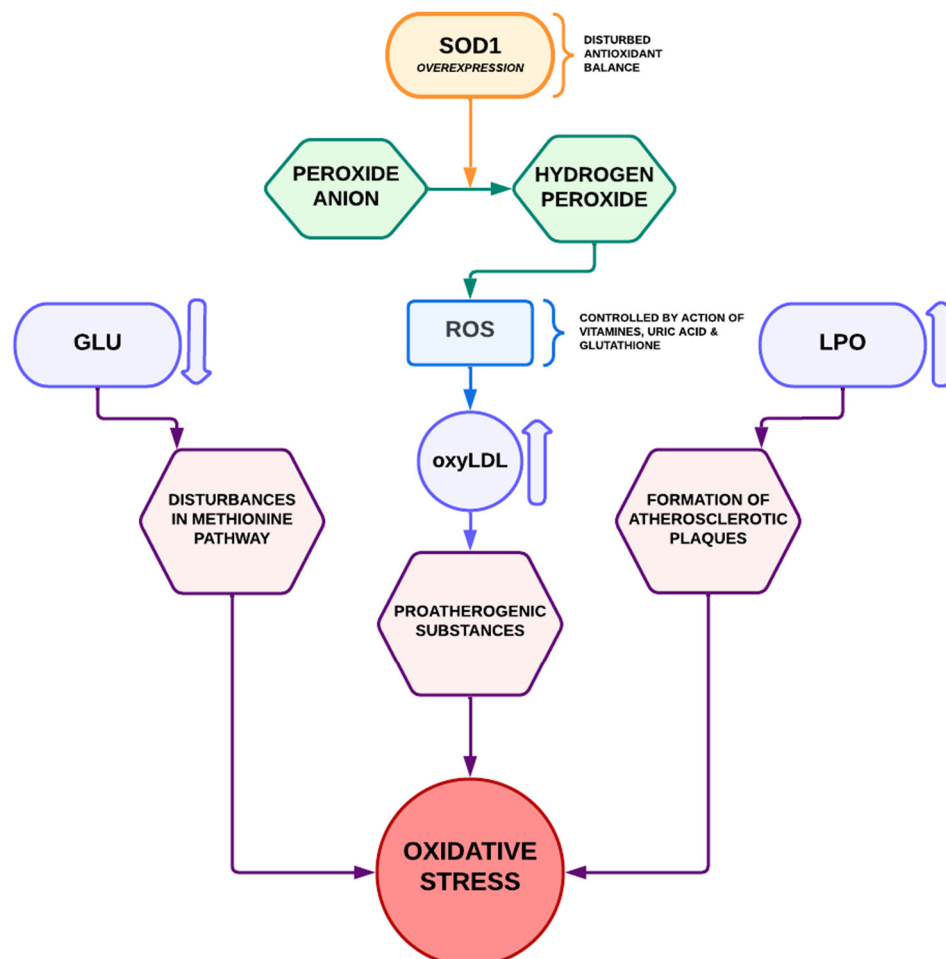


Figure 1. Oxidative stress processes in people with Down syndrome. SOD1-Superoxide dismutase; GLU-glutathione; ROS-reactive oxygen species; LPO-lipid peroxidation; oxyLDL-oxidatively modified LDLs.

3.4. Metabolic and Endocrinological Disorders

The gene coding for the enzyme β -cystathionine synthase (CBS) is located on chromosome 21. This enzyme is responsible for converting homocysteine (Hcy) and serine into cystathionine in the methionine metabolic pathway. Hcy, being a by-product of methionine metabolism, must be converted back to methionine (by re-methylation or by conversion to cysteine). In this process, the important part is played by folic acid and vitamins B (B6 and B12). Three copies of the CBS enzyme genes cause its overexpression; thus, people with DS have a reduced level of homocysteine, which should result in a reduced risk of AS. The reduced concentration of homocysteine also means a lower concentration of methionine, deficiency of tetrahydro folic acid (THF) (the so-called THF trap), and the participation of B vitamins in the methionine pathways. Additionally, the low concentration of homocysteine results in DNA hypermethylation. The disruption of the methionine metabolism pathways is caused by a number of unfavorable metabolic disorders that can be detected using metabolomics studies. Low availability of vitamin B (B6, B12, and folic acid) leads to impaired re-methylation of homocysteine to methionine and thus accumulation of homocysteine [113]. Recent data indicate that homocysteine accumulates in states of increased

oxidative stress associated with immune activation [113]. To better understand the above processes, simplified diagrams have been prepared: Figure 2 shows the correct metabolic pathways; Figure 3 shows the disorders occurring in DS.

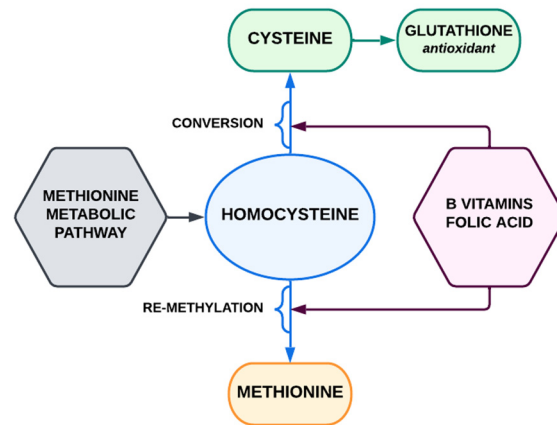


Figure 2. Methionine metabolic pathway (simplified).

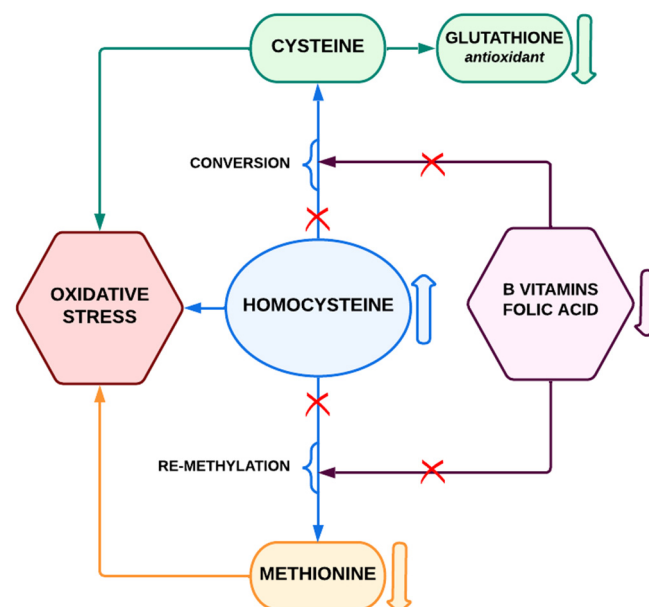


Figure 3. Methionine metabolic pathway (simplified)—pathway disorders in people with Down syndrome.

Children with DS have a higher likelihood of developing endocrine and metabolic disorders such as thyroid dysfunction, diabetes mellitus, short stature, vitamin D deficiency, and obesity than the general population [44,89,114–118]. Thyroid dysfunction is the most common endocrine abnormality in DS children: it is about 38 times more common in individuals with DS than in other people [119,120]. Thyroid hormones are involved in the regulation of carbo–lipid metabolism. They are related to oxidative stress by stimulating cellular metabolism and influencing antioxidant mechanisms as well as regulating oxygen consumption and producing free radicals [111,121]. It is estimated that the incidence of thyroid gland disorders in people with DS increases with age [122]. Aslam et al. demonstrated that at younger ages the incidence of diabetes in patients with DS is four times higher than that of control patients. Peak mean BMI is higher and established earlier in DS, contributing to T2DM risk [123]. The prevalence of type 2 diabetes mellitus in children with DS ranged between 0% and 3.6% [117]. Wernio et al. pointed out that in children with

DS fat mass, fat mass/height² index, and visceral fat mass correlated with thiobarbituric acid reactive-substances and advanced oxidative protein product-levels [87].

3.5. Metabolomics in Down Syndrome

To date, numerous disturbances in the concentration of metabolites in DS have been described, such as: increased levels of phenylalanine and tyrosine in blood serum [124]; lower plasma levels of free histidine, lysine, tyrosine, phenylalanine, leucine, isoleucine, and tryptophan [125]; increased plasma concentrations of leucine, isoleucine, cysteine, and phenylalanine [126]; decreased plasma concentration of serine [127]; increased plasma lysine concentration [127]; elevated concentrations of metabolites related to the methylation cycle such as cysteine, cystathionine, choline, and dimethylglycine [125]; and increased concentrations of S-adenosylhomocysteine and S-adenosylmethionine [125]. Little data exist on the use of metabolomics among PWZD [125,126]. At the same time, there are no data on the use of lipidomics in DS. Orozco et al. analyzed metabolomics of 31 PWZD and observed alterations to methylation metabolism, carnitine/O-acetyl carnitine, dimethyl sulfone, and myo-inositol in children with DS [128]. Obeid et al. reported similar findings in methylation pathway metabolites and found elevated blood cystathionine, cysteine, betaine, choline, and N,N-dimethylglycine in children and young adults with DS [125]. Caracausi et al. analyzed plasma and urine of children with DS and revealed DS/normal ratio in plasma being 1.23 (pyruvate), 1.47 (succinate), 1.39 (fumarate), 1.33 (lactate), and 1.4 (formate) [126]. As most of the altered concentrations were consistent with the 3:2 gene dosage model, there is a possibility that the mentioned changes are caused by the presence of three copies of chromosome 21 [126]. As a result of the use of different methods of omics techniques, as well as the differences in metabolites among children and adults, it is very difficult at the present stage to compare the results obtained in the studies mentioned in the review. However, it is very important to perform metabolomic and lipidomic tests in children with DS in order to be able to compare and analyze the data.

4. Summary and Conclusions

Trisomy of 21 chromosome affects the cardiovascular system in anatomical and physiological ways. Numerous hormonal and metabolic disorders are a leading problem in children and adolescents with DS. Those disorders aggravate with age and require constant targeted medical care. As a result of disorders in metabolic processes and biochemical pathways, theoretically protective factors (low homocysteine level, high SOD1 level) do not fulfil their original functions. The results of the current research seem to contradict the assumption that PWDS are not at risk of developing cardiovascular disease. At present, some classic predispositions are known but CVD prophylaxis requires identifying early risk factors. In such case, it is advisable to extend the research of omics techniques on atherosclerosis risk factors and predisposition to include related diseases in people with DS.

Author Contributions: Conceptualization, E.B. and M.H.; methodology, M.H.; software, M.H.; validation, E.B. and M.H.; formal analysis, E.B. and M.H.; investigation, M.H.; resources, E.B. and M.H.; data curation, M.H.; writing—original draft preparation, M.H.; writing—review and editing, E.B. and M.H.; visualization, M.H.; supervision, E.B.; project administration, E.B. and M.H.; funding acquisition, E.B. and M.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding. However, the publication will be financed by the Wroclaw Medical University, including subsidy funds (Wroclaw Medical University; SUBK.D130.22.055) for the project “Children and young adults with Down Syndrome-metabolomics”.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. van der Greef, J.; Smilde, A.K. Symbiosis of Chemometrics and Metabolomics: Past, Present, and Future. *J. Chemom.* **2005**, *19*, 376–386. [[CrossRef](#)]
2. World Health Organization (WHO). *International Classification of Functioning, Disability and Health*; World Health Organization: Geneva, Switzerland, 2018.
3. Bull, M.J. Down Syndrome. *N. Engl. J. Med.* **2020**, *382*, 2344–2352. [[CrossRef](#)] [[PubMed](#)]
4. Chicoine, B.; Rivelli, A.; Fitzpatrick, V.; Chicoine, L.; Jia, G.; Rzhetsky, A. Prevalence of Common Disease Conditions in a Large Cohort of Individuals with down Syndrome in the United States. *J. Patient-Cent. Res. Rev.* **2021**, *8*, 86–97. [[CrossRef](#)] [[PubMed](#)]
5. Vis, J.C.; Duffels, M.G.J.; Winter, M.M.; Weijerman, M.E.; Cobben, J.M.; Huisman, S.A.; Mulder, B.J.M. Down Syndrome: A Cardiovascular Perspective. *J. Intellect. Disabil. Res.* **2009**, *53*, 419–425. [[CrossRef](#)]
6. Heller, T.; Hsieh, K.; Rimmer, J. Barriers and Supports for Exercise Participation among Adults with down Syndrome. *J. Gerontol. Soc. Work.* **2003**, *38*, 161–178. [[CrossRef](#)]
7. Castro-Piñero, J.; Carbonell-Baeza, A.; Martinez-Gomez, D.; Gómez-Martínez, S.; Cabanas-Sánchez, V.; Santiago, C.; Veses, A.M.; Bandrés, F.; Gonzalez-Galo, A.; Gomez-Gallego, F.; et al. Follow-up in Healthy Schoolchildren and in Adolescents with down Syndrome: Psycho-Environmental and Genetic Determinants of Physical Activity and Its Impact on Fitness, Cardiovascular Diseases, Inflammatory Biomarkers and Mental Health; the Up & Down Study. *BMC Public Health* **2014**, *14*, 400. [[CrossRef](#)]
8. Adelekan, T.; Magge, S.; Shults, J.; Stallings, V.; Stettler, N. Lipid Profiles of Children with down Syndrome Compared with Their Siblings. *Pediatrics* **2012**, *129*, e1382–e1387. [[CrossRef](#)]
9. Rowland, M.; Peterson-Besse, J.; Dobbertin, K.; Walsh, E.S.; Horner-Johnson, W.; Expert Panel on Disability and Health Disparities. Health Outcome Disparities among Subgroups of People with Disabilities: A Scoping Review. *Disabil. Health J.* **2014**, *7*, 136–150. [[CrossRef](#)]
10. Klupczyńska, A.; Dereziński, Z.; Kokot, J. Metabolomics in medical sciences and trends, challenges and perspectives. *Acta Pol. Pharm. Drug Res.* **2015**, *72*, 629–641.
11. Nicholson, J.K.; Lindon, J.C.; Holmes, E. “Metabonomics”: Understanding the Metabolic Responses of Living Systems to Pathophysiological Stimuli via Multivariate Statistical Analysis of Biological NMR Spectroscopic Data. *Xenobiotica* **1999**, *29*, 1181–1189. [[CrossRef](#)]
12. Perk, J.; De Backer, G.; Gohlke, H.; Graham, I.; Reiner, Ž.; Verschuren, M.; Albus, C.; Benlian, P.; Boysen, G.; Cifkova, R.; et al. European Guidelines on Cardiovascular Disease Prevention in Clinical Practice (Version 2012) the Fifth Joint Task Force of the European Society of Cardiology and Other Societies on Cardiovascular Disease Prevention in Clinical Practice (Constituted by Representatives of Nine Societies and by Invited Experts) Developed with the Special Contribution of the European Association for Cardiovascular Prevention & Rehabilitation (EACPR). *Eur. Heart J.* **2012**, *33*, 1635–1701. [[CrossRef](#)]
13. Inouye, M.; Ripatti, S.; Kettunen, J.; Lyytikäinen, L.-P.; Oksala, N.; Laurila, P.-P.; Kangas, A.J.; Soininen, P.; Savolainen, M.J.; Viikari, J.; et al. Novel Loci for Metabolic Networks and Multi-Tissue Expression Studies Reveal Genes for Atherosclerosis. *PLoS Genet.* **2012**, *8*, e1002907. [[CrossRef](#)] [[PubMed](#)]
14. Reiner, Z.; Catapano, A.L.; de Backer, G.; Graham, I.; Taskinen, M.-R.; Wiklund, O.; Agewall, S.; Alegria, E. ESC/EAS Guidelines for the Management of Dyslipidaemias: The Task Force for the Management of Dyslipidaemias of the European Society of Cardiology (ESC) and the European Atherosclerosis Society (EAS). *Eur. Heart J.* **2011**, *32*, 1769–1818. [[CrossRef](#)] [[PubMed](#)]
15. Wang, T.J. Assessing the Role of Circulating, Genetic, and Imaging Biomarkers in Cardiovascular Risk Prediction. *Circulation* **2011**, *123*, 551–565. [[CrossRef](#)] [[PubMed](#)]
16. Yla-Herttuala, S.; Bentzon, J.F.; Daemen, M.; Falk, E.; Garcia-Garcia, H.M.; Herrmann, J.; Hofer, I.; Jauhiainen, S.; Jukema, J.W.; Krams, R.; et al. Stabilization of Atherosclerotic Plaques: An Update. *Eur. Heart J.* **2013**, *34*, 3251–3258. [[CrossRef](#)]
17. Inouye, M.; Kettunen, J.; Soininen, P.; Silander, K.; Ripatti, S.; Kumpula, L.S.; Hämäläinen, E.; Jousilahti, P.; Kangas, A.J.; Männistö, S.; et al. Metabonomic, Transcriptomic, and Genomic Variation of a Population Cohort. *Mol. Syst. Biol.* **2010**, *6*, 441. [[CrossRef](#)]
18. Shah, S.H.; Kraus, W.E.; Newgard, C.B. Metabolomic Profiling for the Identification of Novel Biomarkers and Mechanisms Related to Common Cardiovascular Diseases. *Circulation* **2012**, *126*, 1110–1120. [[CrossRef](#)]
19. Wurtz, P.; Raiko, J.R.; Magnussen, C.G.; Soininen, P.; Kangas, A.J.; Tynkkynen, T.; Thomson, R.; Laatikainen, R.; Savolainen, M.J.; Laurikka, J.; et al. High-Throughput Quantification of Circulating Metabolites Improves Prediction of Subclinical Atherosclerosis. *Eur. Heart J.* **2012**, *33*, 2307–2316. [[CrossRef](#)]
20. Tang, W.H.; Wang, Z.; Levison, B.S. Intestinal Microbial Metabolism of Phosphatidylcholine and Cardiovascular Risk. *J. Vasc. Surg.* **2013**, *58*, 549. [[CrossRef](#)]
21. Dunn, W.B.; Bailey, N.J.C.; Johnson, H.E. Measuring the Metabolome: Current Analytical Technologies. *Analyst* **2005**, *130*, 606. [[CrossRef](#)]
22. Kolwicz, S.C.; Purohit, S.; Tian, R. Cardiac Metabolism and Its Interactions with Contraction, Growth, and Survival of Cardiomyocytes. *Circ. Res.* **2013**, *113*, 603–616. [[CrossRef](#)]

23. Kordalewska, M.; Markuszewski, M.J. Metabolomics in Cardiovascular Diseases. *J. Pharm. Biomed. Anal.* **2015**, *113*, 121–136. [[CrossRef](#)] [[PubMed](#)]
24. Otvos, J.D.; Jeyarajah, E.J.; Bennett, D.W. Quantification of Plasma Lipoproteins by Proton Nuclear Magnetic Resonance Spectroscopy. *Clin. Chem.* **1991**, *37*, 377–386. [[CrossRef](#)] [[PubMed](#)]
25. Griffin, J.L.; Atherton, H.; Shockcor, J.; Atzori, L. Metabolomics as a Tool for Cardiac Research. *Nat. Rev. Cardiol.* **2011**, *8*, 630–643. [[CrossRef](#)] [[PubMed](#)]
26. Cheng, S.; Shah, S.H.; Corwin, E.J.; Fiehn, O.; Fitzgerald, R.L.; Gerszten, R.E.; Illig, T.; Rhee, E.P.; Srinivas, P.R.; Wang, T.J.; et al. Potential Impact and Study Considerations of Metabolomics in Cardiovascular Health and Disease: A Scientific Statement from the American Heart Association. *Circ. Cardiovasc. Genet.* **2017**, *10*, e000032. [[CrossRef](#)]
27. Klein, M.S.; Shearer, J. Metabolomics and Type 2 Diabetes: Translating Basic Research into Clinical Application. *J. Diabetes Res.* **2016**, *2016*, 3898502. [[CrossRef](#)]
28. Rauschert, S.; Uhl, O.; Koletzko, B.; Kirchberg, F.; Mori, T.A.; Huang, R.-C.; Beilin, L.J.; Hellmuth, C.; Oddy, W.H. Lipidomics Reveals Associations of Phospholipids with Obesity and Insulin Resistance in Young Adults. *J. Clin. Endocrinol. Metab.* **2016**, *101*, 871–879. [[CrossRef](#)] [[PubMed](#)]
29. Hellmuth, C.; Kirchberg, F.F.; Brandt, S.; Moß, A.; Walter, V.; Rothenbacher, D.; Brenner, H.; Grote, V.; Gruszfeld, D.; Socha, P.; et al. An Individual Participant Data Meta-Analysis on Metabolomics Profiles for Obesity and Insulin Resistance in European Children. *Sci. Rep.* **2019**, *9*, 5053. [[CrossRef](#)] [[PubMed](#)]
30. Rzehak, P.; Hellmuth, C.; Uhl, O.; Kirchberg, F.F.; Peissner, W.; Harder, U.; Grote, V.; Weber, M.; Xhonneux, A.; Langhendries, J.-P.; et al. Rapid Growth and Childhood Obesity Are Strongly Associated with LysoPC (14:0). *Ann. Nutr. Metab.* **2014**, *64*, 294–303. [[CrossRef](#)]
31. Steffen, L.M.; Vessby, B.; Jacobs, D.R.; Steinberger, J.; Moran, A.; Hong, C.-P.; Sinaiko, A.R. Serum Phospholipid and Cholesteryl Ester Fatty Acids and Estimated Desaturase Activities Are Related to Overweight and Cardiovascular Risk Factors in Adolescents. *Int. J. Obes.* **2008**, *32*, 1297–1304. [[CrossRef](#)]
32. Rafieian-kopaei, M.; Setorki, M.; Douidi, M.; Baradaran, A.; Nasri, H.R. Atherosclerosis: Process, Indicators, Risk Factors and New Hopes. *Int. J. Prev. Med.* **2014**, *5*, 927–946.
33. Cattaneo, M. Hyperhomocysteinemia, Atherosclerosis and Thrombosis. *Thromb. Haemost.* **1999**, *81*, 165–176. [[CrossRef](#)] [[PubMed](#)]
34. Würtz, P.; Havulinna, A.S.; Soininen, P.; Tynkkynen, T.; Prieto-Merino, D.; Tillin, T.; Ghorbani, A.; Artati, A.; Wang, Q.; Tiainen, M.; et al. Metabolite Profiling and Cardiovascular Event Risk. *Circulation* **2015**, *131*, 774–785. [[CrossRef](#)] [[PubMed](#)]
35. Bhattacharya, S.; Granger, C.B.; Craig, D.; Haynes, C.; Bain, J.; Stevens, R.D.; Hauser, E.R.; Newgard, C.B.; Kraus, W.E.; Newby, L.K.; et al. Validation of the Association between a Branched Chain Amino Acid Metabolite Profile and Extremes of Coronary Artery Disease in Patients Referred for Cardiac Catheterization. *Atherosclerosis* **2014**, *232*, 191–196. [[CrossRef](#)]
36. Handakas, E.; Lau, C.H.; Alfano, R.; Chatzi, V.L.; Plusquin, M.; Vineis, P.; Robinson, O. A Systematic Review of Metabolomic Studies of Childhood Obesity: State of the Evidence for Metabolic Determinants and Consequences. *Obes. Rev.* **2021**, *23*, e13384. [[CrossRef](#)] [[PubMed](#)]
37. Polidori, N.; Grasso, E.A.; Chiarelli, F.; Giannini, C. Amino Acid-Related Metabolic Signature in Obese Children and Adolescents. *Nutrients* **2022**, *14*, 1454. [[CrossRef](#)]
38. Gofman, J.W.; Lindgren, F.; Elliott, H.; Mantz, W.; Hewitt, J.; Strisower, B.; Herring, V.; Lyon, T.P. The Role of Lipids and Lipoproteins in Atherosclerosis. *Science* **1950**, *111*, 166–186. [[CrossRef](#)]
39. Zhao, Y.-Y.; Cheng, X.; Lin, R.-C. Lipidomics Applications for Discovering Biomarkers of Diseases in Clinical Chemistry. *Int. Rev. Cell Mol. Biol.* **2014**, *313*, 1–26. [[CrossRef](#)]
40. Kim, E.J.; Ramachandran, R.; Wierzbicki, A.S. Lipidomics in Diabetes. *Curr. Opin. Endocrinol. Diabetes Obes.* **2021**, *29*, 124–130. [[CrossRef](#)]
41. Murdoch, J.C.; Rodger, J.C.; Rao, S.S.; Fletcher, C.D.; Dunnigan, M.G. Down’s Syndrome: An Atheroma-Free Model? *BMJ* **1977**, *2*, 226–228. [[CrossRef](#)]
42. Pueschel, S.M.; Craig, W.Y.; Hadow, J.E. Lipids and Lipoproteins in Persons with Down’s Syndrome. *J. Intellect. Disabil. Res.* **2008**, *36*, 365–369. [[CrossRef](#)] [[PubMed](#)]
43. Alexander, M.; Petri, H.; Ding, Y.; Wandel, C.; Khwaja, O.; Foskett, N. Morbidity and Medication in a Large Population of Individuals with down Syndrome Compared to the General Population. *Dev. Med. Child Neurol.* **2015**, *58*, 246–254. [[CrossRef](#)]
44. Sobey, C.G.; Judkins, C.P.; Sundararajan, V.; Phan, T.G.; Drummond, G.R.; Srikanth, V.K. Risk of Major Cardiovascular Events in People with down Syndrome. *PLoS ONE* **2015**, *10*, e0137093. [[CrossRef](#)] [[PubMed](#)]
45. Bell, A.J.; Bhat, M.S. Prevalence of Overweight and Obesity in Down’s Syndrome and Other Mentally Handicapped Adults Living in the Community. *J. Intellect. Disabil. Res.* **2008**, *36*, 359–364. [[CrossRef](#)] [[PubMed](#)]
46. Prasher, V.P. Overweight and Obesity amongst Down’s Syndrome Adults. *J. Intellect. Disabil. Res.* **1995**, *39*, 437–441. [[CrossRef](#)]
47. Melville, C.A.; Cooper, S.-A.; McGrother, C.W.; Thorp, C.F.; Collacott, R. Obesity in Adults with down Syndrome: A Case-Control Study. *J. Intellect. Disabil. Res.* **2005**, *49*, 125–133. [[CrossRef](#)]
48. Dörner, K.; Gaethke, A.-S.; Tolksdorf, M.; Schumann, K.P.; Gustmann, H. Cholesterol Fractions and Triglycerides in Children and Adults with Down’s Syndrome. *Clin. Chim. Acta* **1984**, *142*, 307–311. [[CrossRef](#)] [[PubMed](#)]
49. Magge, S.N.; Zemel, B.S.; Pipan, M.E.; Gidding, S.S.; Kelly, A. Cardiometabolic Risk and Body Composition in Youth with down Syndrome. *Pediatrics* **2019**, *144*, e20190137. [[CrossRef](#)]

50. Brattström, L.; Englund, E.; Brun, A. Does down Syndrome Support Homocysteine Theory of Arteriosclerosis? *Lancet* **1987**, *329*, 391–392. [[CrossRef](#)] [[PubMed](#)]
51. Ylä-Herttua, S.; Luoma, J.; Nikkari, T.; Kivimäki, T. Down's Syndrome and Atherosclerosis. *Atherosclerosis* **1989**, *76*, 269–272. [[CrossRef](#)]
52. Draheim, C.C.; Geijer, J.R.; Dengel, D.R. Comparison of Intima-Media Thickness of the Carotid Artery and Cardiovascular Disease Risk Factors in Adults with versus without the down Syndrome. *Am. J. Cardiol.* **2010**, *106*, 1512–1516. [[CrossRef](#)]
53. Parra, P.; Costa, R.; de Asúa, D.R.; Moldenhauer, F.; Suárez, C. Atherosclerotic Surrogate Markers in Adults with down Syndrome: A Case-Control Study. *J. Clin. Hypertens.* **2016**, *19*, 205–211. [[CrossRef](#)] [[PubMed](#)]
54. Corsi, M.M.; Dogliotti, G.; Pedroni, F.; Galliera, E.; Malavazos, A.E.; Villa, R.; Chiappelli, M.; Licastro, F. Adipocytokines in Down's Syndrome, an Atheroma-Free Model: Role of Adiponectin. *Arch. Gerontol. Geriatr.* **2009**, *48*, 106–109. [[CrossRef](#)] [[PubMed](#)]
55. Head, E.; Phelan, M.J.; Doran, E.; Kim, R.C.; Poon, W.W.; Schmitt, F.A.; Lott, I.T. Cerebrovascular Pathology in down Syndrome and Alzheimer Disease. *Acta Neuropathol. Commun.* **2017**, *5*, 93. [[CrossRef](#)] [[PubMed](#)]
56. de Almeida, E.W.; Greguol, M. Lipid Profile in People with down Syndrome: A Literature Review. *J. Hum. Growth Dev.* **2020**, *30*, 197–208. [[CrossRef](#)]
57. Rodrigues, A.N.; Coelho, L.C.; Goncalves, W.L.S.; Gouvea, S.A.; Vasconcellos, M.J.R.; Cunha, R.S.; Abreu, G.R. Stiffness of the Large Arteries in Individuals with and without down Syndrome. *Vasc. Health Risk Manag.* **2011**, *7*, 375–381. [[CrossRef](#)] [[PubMed](#)]
58. Landes, S.D.; Stevens, J.D.; Turk, M.A. Cause of Death in Adults with down Syndrome in the United States. *Disabil. Health J.* **2020**, *13*, 100947. [[CrossRef](#)]
59. Hill, D.A.; Gridley, G.; Cnattingius, S.; Mellekjaer, L.; Linet, M.; Adami, H.-O.; Olsen, J.H.; Nyren, O.; Fraumeni, J.F. Mortality and Cancer Incidence among Individuals with down Syndrome. *Arch. Intern. Med.* **2003**, *163*, 705. [[CrossRef](#)]
60. Day, S.M.; Strauss, D.J.; Shavelle, R.M.; Reynolds, R.J. Mortality and Causes of Death in Persons with down Syndrome in California. *Dev. Med. Child Neurol.* **2007**, *47*, 171–176. [[CrossRef](#)]
61. Hermon, C.; Alberman, E.; Beral, V.; Swerdlow, A.J. Mortality and Cancer Incidence in Persons with Down's Syndrome, Their Parents and Siblings. *Ann. Hum. Genet.* **2001**, *65*, 167–176. [[CrossRef](#)]
62. Buonomo, P.S.; Bartuli, A.; Mastrogiorgio, G.; Vittucci, A.; Di Camillo, C.; Bianchi, S.; Pires Marafon, D.; Villani, A.; Valentini, D. Lipid Profiles in a Large Cohort of Italian Children with down Syndrome. *Eur. J. Med. Genet.* **2016**, *59*, 392–395. [[CrossRef](#)]
63. Jang, A.Y.; Han, S.H.; Sohn, I.S.; Oh, P.C.; Koh, K.K. Lipoprotein(A) and Cardiovascular Diseases Revisited. *Circ. J.* **2020**, *84*, 867–874. [[CrossRef](#)] [[PubMed](#)]
64. Krzesińska, A.; Klosowska, A.; Sałaga-Zaleska, K.; Ćwiklińska, A.; Mickiewicz, A.; Chyła, G.; Wierzbza, J.; Jankowski, M.; Kuchta, A. Lipid Profile, Lp(A) Levels, and HDL Quality in Adolescents with down Syndrome. *J. Clin. Med.* **2022**, *11*, 4356. [[CrossRef](#)] [[PubMed](#)]
65. Kamstrup, P.R. Lipoprotein(A): The Common, Likely Causal, yet Elusive Risk Factor for Cardiovascular Disease. *J. Lipid Res.* **2017**, *58*, 1731–1732. [[CrossRef](#)] [[PubMed](#)]
66. Trinder, M.; Uddin, M.M.; Finneran, P.; Aragam, K.G.; Natarajan, P. Clinical Utility of Lipoprotein(A) and LPA Genetic Risk Score in Risk Prediction of Incident Atherosclerotic Cardiovascular Disease. *JAMA Cardiol.* **2021**, *6*, 287. [[CrossRef](#)]
67. Roizen, N.J.; Patterson, D. Down's Syndrome. *Lancet* **2003**, *361*, 1281–1289. [[CrossRef](#)]
68. Harris, N.; Rosenberg, A.; Jangda, S.; O'Brien, K.; Gallagher, M.L. Prevalence of Obesity in International Special Olympic Athletes as Determined by Body Mass Index. *J. Am. Diet. Assoc.* **2003**, *103*, 235–237. [[CrossRef](#)]
69. Nordstrøm, M.; Retterstøl, K.; Hope, S.; Kolset, S.O. Nutritional Challenges in Children and Adolescents with down Syndrome. *Lancet Child Adolesc. Health* **2020**, *4*, 455–464. [[CrossRef](#)]
70. Bertapelli, F.; Pitetti, K.; Agiovlasitis, S.; Guerra-Junior, G. Overweight and Obesity in Children and Adolescents with down Syndrome—Prevalence, Determinants, Consequences, and Interventions: A Literature Review. *Res. Dev. Disabil.* **2016**, *57*, 181–192. [[CrossRef](#)]
71. Luke, A.; Sutton, M.; Schoeller, D.A.; Roizen, N.J.M. Nutrient Intake and Obesity in Prepubescent Children with down Syndrome. *J. Am. Diet. Assoc.* **1996**, *96*, 1262–1267. [[CrossRef](#)]
72. Bull, M.J. Health Supervision for Children with down Syndrome. *Pediatrics* **2011**, *128*, 393–406. [[CrossRef](#)]
73. Apovian, C.M. Obesity: Definition, comorbidities, causes, and burden. *Am. J. Manag. Care* **2016**, *22* (Suppl. 7), s176–s185. [[PubMed](#)]
74. Rimmer, J.H.; Wang, E. Obesity Prevalence among a Group of Chicago Residents with Disabilities. *Arch. Phys. Med. Rehabil.* **2005**, *86*, 1461–1464. [[CrossRef](#)] [[PubMed](#)]
75. Nordstrøm, M.; Hansen, B.H.; Paus, B.; Kolset, S.O. Accelerometer-Determined Physical Activity and Walking Capacity in Persons with down Syndrome, Williams Syndrome and Prader-Willi Syndrome. *Res. Dev. Disabil.* **2013**, *34*, 4395–4403. [[CrossRef](#)] [[PubMed](#)]
76. Shields, N.; Plant, S.; Warren, C.; Wollersheim, D.; Peiris, C. Do Adults with down Syndrome Do the Same Amount of Physical Activity as Adults without Disability? A Proof of Principle Study. *J. Appl. Res. Intellect. Disabil.* **2017**, *31*, 459–465. [[CrossRef](#)]
77. Fox, B.; Moffett, G.E.; Kinnison, C.; Brooks, G.; Case, L.E. Physical Activity Levels of Children with down Syndrome. *Pediatr. Phys. Ther.* **2019**, *31*, 33–41. [[CrossRef](#)]
78. Hsieh, K.; Hilgenkamp, T.; Murthy, S.; Heller, T.; Rimmer, J. Low Levels of Physical Activity and Sedentary Behavior in Adults with Intellectual Disabilities. *Int. J. Environ. Res. Public Health* **2017**, *14*, 1503. [[CrossRef](#)]

79. Guthold, R.; Stevens, G.A.; Riley, L.M.; Bull, F.C. Global Trends in Insufficient Physical Activity among Adolescents: A Pooled Analysis of 298 Population-Based Surveys with 1.6 Million Participants. *Yearb. Paediatr. Endocrinol.* **2020**, *4*, 23–35. [[CrossRef](#)]
80. Barr, M.; Shields, N. Identifying the Barriers and Facilitators to Participation in Physical Activity for Children with down Syndrome. *J. Intellect. Disabil. Res.* **2011**, *55*, 1020–1033. [[CrossRef](#)]
81. De Lausnay, M.; Ides, K.; Wojciechowski, M.; Boudewyns, A.; Verhulst, S.; Van Hoorenbeek, K. Pulmonary Complications in Children with down Syndrome: A Scoping Review. *Paediatr. Respir. Rev.* **2021**, *40*, 65–72. [[CrossRef](#)]
82. McGarty, A.M.; Melville, C.A. Parental Perceptions of Facilitators and Barriers to Physical Activity for Children with Intellectual Disabilities: A Mixed Methods Systematic Review. *Res. Dev. Disabil.* **2018**, *73*, 40–57. [[CrossRef](#)]
83. Hatch-Stein, J.A.; Zemel, B.S.; Prasad, D.; Kalkwarf, H.J.; Pipan, M.; Magge, S.N.; Kelly, A. Body Composition and BMI Growth Charts in Children with down Syndrome. *Pediatrics* **2016**, *138*, e20160541. [[CrossRef](#)] [[PubMed](#)]
84. Pierce, M.; Ramsey, K.; Pinter, J. Trends in Obesity and Overweight in Oregon Children with down Syndrome. *Glob. Pediatr. Health* **2019**, *6*, 2333794X1983564. [[CrossRef](#)]
85. Yahia, S.; El-Farahaty, R.; EL-Gilany, A.-H.; Shoaib, R.; Ramadan, R.; Salem, N. Serum Adiponectin, Body Adiposity and Metabolic Parameters in Obese Egyptian Children with down Syndrome. *J. Pediatr. Endocrinol. Metab.* **2021**, *34*, 1401–1410. [[CrossRef](#)] [[PubMed](#)]
86. Hetman, M.; Moreira, H.; Barg, E. The Best Tool for the Assessment of Developmental Disorders in Children with down Syndrome: Comparison of Standard and Specialized Growth Charts-Cross Sectional Study. *Front. Endocrinol.* **2022**, *13*, 928151. [[CrossRef](#)]
87. Wernio, E.; Kłosowska, A.; Kuchta, A.; Ćwiklińska, A.; Sałaga-Zaleska, K.; Jankowski, M.; Kłosowski, P.; Wiśniewski, P.; Wierzba, J.; Małgorzewicz, S. Analysis of Dietary Habits and Nutritional Status of Children with down Syndrome in the Context of Lipid and Oxidative Stress Parameters. *Nutrients* **2022**, *14*, 2390. [[CrossRef](#)] [[PubMed](#)]
88. Haligheri, G.; Johnson, T.; Kathol, M.; Kuzava, L.; Goth, N.; Staggs, V.S.; Donnelly, J.E.; Ptomey, L.T.; Forsha, D. Early Cardiac Dysfunction in Obese Adolescents with down Syndrome or Autism. *Cardiol. Young* **2022**, 1–8. [[CrossRef](#)]
89. Basil, J.S.; Santoro, S.L.; Martin, L.J.; Healy, K.W.; Chini, B.A.; Saal, H.M. Retrospective Study of Obesity in Children with down Syndrome. *J. Pediatr.* **2016**, *173*, 143–148. [[CrossRef](#)]
90. Pecoraro, L.; Solfa, M.; Ferron, E.; Mirandola, M.; Lauriola, S.; Piacentini, G.; Pietrobelli, A. Mediterranean Diet and Physical Activity in down Syndrome Pediatric Subjects: The DONUT STUDY. *Int. J. Food Sci. Nutr.* **2022**, *73*, 973–980. [[CrossRef](#)]
91. Amatori, S.; Sisti, D.; Perroni, F.; Brandi, G.; Rocchi, M.B.L.; Gobbi, E. Physical Activity, Sedentary Behaviour and Screen Time among Youths with down Syndrome during the COVID-19 Pandemic. *J. Intellect. Disabil. Res.* **2022**, *66*, 903–912. [[CrossRef](#)]
92. Roccatello, G.; Cocchi, G.; Dimastromatteo, R.T.; Cavallo, A.; Biserni, G.B.; Selicati, M.; Forchielli, M.L. Eating and Lifestyle Habits in Youth with down Syndrome Attending a Care Program: An Exploratory Lesson for Future Improvements. *Front. Nutr.* **2021**, *8*, 641112. [[CrossRef](#)]
93. Jobling, A.; Cuskelly, M. Young People with down Syndrome: A Preliminary Investigation of Health Knowledge and Associated Behaviours. *J. Intellect. Dev. Disabil.* **2006**, *31*, 210–218. [[CrossRef](#)] [[PubMed](#)]
94. Naczka, A.; Gajewska, E.; Naczka, M. Effectiveness of Swimming Program in Adolescents with down Syndrome. *Int. J. Environ. Res. Public Health* **2021**, *18*, 7441. [[CrossRef](#)] [[PubMed](#)]
95. Eckel, R.H.; Jakicic, J.M.; Ard, J.D.; de Jesus, J.M.; Miller, N.H.; Hubbard, V.S.; Lee, I.-M.; Lichtenstein, A.H.; Loria, C.M.; Millen, B.E.; et al. 2013 AHA/ACC Guideline on Lifestyle Management to Reduce Cardiovascular Risk. *Circulation* **2013**, *129* (Suppl. 2), S76–S99. [[CrossRef](#)]
96. Loprinzi, P.D.; Lee, I.-M.; Andersen, R.E.; Crespo, C.J.; Smit, E. Association of Concurrent Healthy Eating and Regular Physical Activity with Cardiovascular Disease Risk Factors in U.S. Youth. *Am. J. Health Promot. AJHP* **2015**, *30*, 2–8. [[CrossRef](#)] [[PubMed](#)]
97. Ma, J.; Qiao, Y.; Zhao, P.; Li, W.; Katzmarzyk, P.T.; Chaput, J.; Fogelholm, M.; Kuriyan, R.; Lambert, E.V.; Maher, C.; et al. Breastfeeding and Childhood Obesity: A 12-Country Study. *Matern. Child Nutr.* **2020**, *16*, e12984. [[CrossRef](#)]
98. Magenis, M.L.; de Faveri, W.; Castro, K.; Forte, G.C.; Grande, A.J.; Perry, I.S. Down Syndrome and Breastfeeding: A Systematic Review. *J. Intellect. Disabil.* **2020**, *26*, 244–263. [[CrossRef](#)]
99. Sies, H.; Berndt, C.; Jones, D.P. Oxidative Stress. *Annu. Rev. Biochem.* **2017**, *86*, 715–748. [[CrossRef](#)]
100. Ordonez, F.J.; Rosety, I.; Rosety, M.A.; Camacho-Molina, A.; Fornieles, G.; Rosety, M.; Rosety-Rodriguez, M. Aerobic Training at Moderate Intensity Reduced Protein Oxidation in Adolescents with down Syndrome. *Scand. J. Med. Sci. Sport.* **2010**, *22*, 91–94. [[CrossRef](#)]
101. Shields, N.; Downs, J.; de Haan, J.B.; Taylor, N.F.; Torr, J.; Fernhall, B.; Kingsley, M.; Mnatzaganian, G.; Leonard, H. What Effect Does Regular Exercise Have on Oxidative Stress in People with down Syndrome? A Systematic Review with Meta-Analyses. *J. Sci. Med. Sport* **2018**, *21*, 596–603. [[CrossRef](#)]
102. Tangvarasittichai, S. Oxidative Stress, Insulin Resistance, Dyslipidemia and Type 2 Diabetes Mellitus. *World J. Diabetes* **2015**, *6*, 456. [[CrossRef](#)]
103. Cappelli-Bigazzi, M.; Santoro, G.; Battaglia, C.; Palladino, M.T.; Carrozza, M.; Russo, M.G.; Pacileo, G.; Calabrò, R. Endothelial Cell Function in Patients with Down's Syndrome. *Am. J. Cardiol.* **2004**, *94*, 392–395. [[CrossRef](#)] [[PubMed](#)]
104. Flore, P.; Bricout, V.-A.; van Biesen, D.; Guinot, M.; Laporte, F.; Pépin, J.-L.; Eberhard, Y.; Favre-Juvin, A.; Wuyam, B.; de Vliet, P.; et al. Oxidative Stress and Metabolism at Rest and during Exercise in Persons with down Syndrome. *Eur. J. Cardiovasc. Prev. Rehabil.* **2008**, *15*, 35–42. [[CrossRef](#)] [[PubMed](#)]

105. Brooksbank, B.W.L.; Balazs, R. Superoxide Dismutase, Glutathione Peroxidase and Lipoperoxidation in Oown's Syndrome Fetal Brain. *Dev. Brain Res.* **1984**, *16*, 37–44. [[CrossRef](#)] [[PubMed](#)]
106. Epstein, C.J.; Avraham, K.B.; Lovett, M.; Smith, S.; Elroy-Stein, O.; Rotman, G.; Bry, C.; Groner, Y. Transgenic Mice with Increased Cu/Zn-Superoxide Dismutase Activity: Animal Model of Dosage Effects in down Syndrome. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 8044–8048. [[CrossRef](#)] [[PubMed](#)]
107. Pallardó, F.V.; Degan, P.; d'Ischia, M.; Kelly, F.J.; Zatterale, A.; Calzone, R.; Castello, G.; Fernandez-Delgado, R.; Dunster, C.; Lloret, A.; et al. Multiple Evidence for an Early Age Pro-Oxidant State in down Syndrome Patients. *Biogerontology* **2006**, *7*, 211–220. [[CrossRef](#)]
108. Tan, Y.H.; Tischfield, J.; Ruddle, F.H. The Linkage of Genes for the Human Interferon-Induced Antiviral Protein and Indophenol Oxidase-B Traits to Chromosome G-21. *J. Exp. Med.* **1973**, *137*, 317–330. [[CrossRef](#)]
109. Halliwell, B.; Gutteridge, J.M.C. Free Radicals in Biology and Medicine. *J. Free Radic. Biol. Med.* **1985**, *1*, 331–332. [[CrossRef](#)]
110. Kumagai, T.; Matsukawa, N.; Kaneko, Y.; Kusumi, Y.; Mitsumata, M.; Uchida, K. A Lipid Peroxidation-Derived Inflammatory Mediator. *J. Biol. Chem.* **2004**, *279*, 48389–48396. [[CrossRef](#)]
111. Muchova, J.; Žitnanova, I.; Ďurackova, Z. Oxidative Stress and down Syndrome. Do Antioxidants Play a Role in Therapy? *Physiol. Res.* **2014**, *63*, 535–542. [[CrossRef](#)]
112. Moor, A.C.E. Signaling Pathways in Cell Death and Survival after Photodynamic Therapy. *J. Photochem. Photobiol. B Biol.* **2000**, *57*, 1–13. [[CrossRef](#)]
113. Copus, A.W.; Fekkes, D.; Verhoeven, W.M.A.; Tuinier, S.; Egger, J.I.M.; van Duijn, C.M. Plasma Amino Acids and Neopterin in Healthy Persons with Down's Syndrome. *J. Neural Transm.* **2007**, *114*, 1041–1045. [[CrossRef](#)] [[PubMed](#)]
114. Vockley, J.; Andersson, H.C.; Antshel, K.M.; Braverman, N.E.; Burton, B.K.; Frazier, D.M.; Mitchell, J.; Smith, W.E.; Thompson, B.H.; Berry, S.A.; et al. Phenylalanine Hydroxylase Deficiency: Diagnosis and Management Guideline. *Genet. Med.* **2013**, *16*, 188–200. [[CrossRef](#)] [[PubMed](#)]
115. Hinckson, E.A.; Dickinson, A.; Water, T.; Sands, M.; Penman, L. Physical Activity, Dietary Habits and Overall Health in Overweight and Obese Children and Youth with Intellectual Disability or Autism. *Res. Dev. Disabil.* **2013**, *34*, 1170–1178. [[CrossRef](#)] [[PubMed](#)]
116. Mulu, B.; Fantahun, B. Thyroid Abnormalities in Children with down Syndrome at St. Paul's Hospital Millennium Medical College, Ethiopia. *Endocrinol. Diabetes Metab.* **2022**, *5*, e00337. [[CrossRef](#)]
117. Butler, A.E.; Sacks, W.; Rizza, R.A.; Butler, P.C. Down Syndrome-Associated Diabetes Is Not due to a Congenital Deficiency in β Cells. *J. Endocr. Soc.* **2017**, *1*, 39–45. [[CrossRef](#)]
118. Moreau, M.; Benhaddou, S.; Dard, R.; Tolu, S.; Hamzé, R.; Vialard, F.; Movassat, J.; Janel, N. Metabolic Diseases and down Syndrome: How Are They Linked Together? *Biomedicines* **2021**, *9*, 221. [[CrossRef](#)]
119. Metwalley, K.A.; Farghaly, H.S. Endocrinal Dysfunction in Children with down Syndrome. *Ann. Pediatr. Endocrinol. Metab.* **2022**, *27*, 15–21. [[CrossRef](#)]
120. Rivelli, A.; Fitzpatrick, V.; Wales, D.; Chicoine, L.; Jia, G.; Rzhetsky, A.; Chicoine, B. Prevalence of Endocrine Disorders among 6078 Individuals with down Syndrome in the United States. *J. Patient-Cent. Res. Rev.* **2022**, *9*, 70–74. [[CrossRef](#)]
121. Mullur, R.; Liu, Y.-Y.; Brent, G.A. Thyroid Hormone Regulation of Metabolism. *Physiol. Rev.* **2014**, *94*, 355–382. [[CrossRef](#)]
122. Karlsson, B.; Gustafsson, J.; Hedov, G.; Ivarsson, S.-A.; Anneren, G. Thyroid Dysfunction in Down's Syndrome: Relation to Age and Thyroid Autoimmunity. *Arch. Dis. Child.* **1998**, *79*, 242–245. [[CrossRef](#)]
123. Aslam, A.A.; Baksh, R.A.; Pape, S.E.; Strydom, A.; Gulliford, M.C.; Chan, L.F.; Herault, Y.; Strydom, A.; Chan, L.; Potier, M.-C.; et al. Diabetes and Obesity in down Syndrome across the Lifespan: A Retrospective Cohort Study Using U.K. Electronic Health Records. *Diabetes Care* **2022**, *45*, 2892–2899. [[CrossRef](#)] [[PubMed](#)]
124. Heggarty, H.J.; Ball, R.; Smith, M.; Henderson, M.J. Amino Acid Profile in Down's Syndrome. *Arch. Dis. Child.* **1996**, *74*, 347–349. [[CrossRef](#)] [[PubMed](#)]
125. Obeid, R.; Hartmuth, K.; Herrmann, W.; Gortner, L.; Rohrer, T.R.; Geisel, J.; Reed, M.C.; Nijhout, H.F. Blood Biomarkers of Methylation in down Syndrome and Metabolic Simulations Using a Mathematical Model. *Mol. Nutr. Food Res.* **2012**, *56*, 1582–1589. [[CrossRef](#)] [[PubMed](#)]
126. Caracausi, M.; Ghini, V.; Locatelli, C.; Mericio, M.; Piovesan, A.; Antonaros, F.; Pelleri, M.C.; Vitale, L.; Vacca, R.A.; Bedetti, F.; et al. Plasma and Urinary Metabolomic Profiles of down Syndrome Correlate with Alteration of Mitochondrial Metabolism. *Sci. Rep.* **2018**, *8*, 2977. [[CrossRef](#)] [[PubMed](#)]
127. Mircher, C.; Salabelle, A.; Peeters, M.; Rabier, D.; Parvy, P.; Kamoun, P.; Lejeune, J. Variation Des Acides Aminés En Fonction de l'Âge Chez Des Sujets Trisomiques 21. *Arch. Pédiatrie* **1997**, *4*, 1093–1099. [[CrossRef](#)]
128. Orozco, J.S.; Hertz-Picciotto, I.; Abbeduto, L.; Slupsky, C.M. Metabolomics Analysis of Children with Autism, Idiopathic-Developmental Delays, and down Syndrome. *Transl. Psychiatry* **2019**, *9*, 243. [[CrossRef](#)]