

$Review$ **Pharmacological Effects and Clinical Prospects of Cepharanthine Di Liang 1, Qi Li 1,2, Lina Du 1,2,* and Guifang Dou 1,***

Di Liang ¹ [,](https://orcid.org/0000-0002-8993-0436) Qi Li 1,2, Lina Du 1,2,* and Guifang Dou 1,* 1 Depar[tme](https://orcid.org/0000-0001-8103-5974)nt of Pharmaceutical Sciences, Beijing Institute of Radiation Medicine, Beijing 100850, China; China;

- ¹ Department of Pharmaceutical Sciences, Beijing Institute of Radiation Medicine, Beijing 100850, China
- ² School of Pharmacy, Shandong University of Traditional Chinese Medicine, Jinan 250355, China
- ***** Correspondence: dulina@188.com (L.D.); dougf@bmi.ac.cn (G.D.)

Abstract: Cepharanthine is an active ingredient separated and extracted from *Stephania cepharantha* Hayata, a Menispermaceae plant. As a bisbenzylisoquinoline alkaloid, cepharanthine has various pharmacological properties, including antioxidant, anti-inflammatory, immunomodulatory, antitumoral, and antiviral effects. Following the emergence of coronavirus disease 2019 (COVID-19), cepharanthine has been found to have excellent anti-COVID-19 activity. In this review, the important physicochemical properties and pharmacological effects of cepharanthine, particularly the antiviral effect, are systematically described. Additionally, the molecular mechanisms and novel dosage formulations for the efficient, safe, and convenient delivery of cepharanthine are summarized.

Keywords: cepharanthine; coronavirus disease 2019; antiviral; novel formulations **Keywords:** cepharanthine; coronavirus disease 2019; antiviral; novel formulations

1. Introduction 1. Introduction

The Menispermaceae family comprises approximately 70 genera with 420 extant species and is considered a medium-sized family. It mainly consists of dioecious climbers and has a variety of medicinal properties $[1,2]$ $[1,2]$. The main active ingredients of the Menispermaceae family are alkaloids, which can be classified into six types based on their chemical maceae family are alkaloids, which can be classified into six types based on their chemical structure: morphine dienone, lotus alkane, apocynin, proto-apocynin, proto-cotyledonine, and bisbenzylisoquinoline. Cepharanthine is an active ingredient separated and extracted from *Stephania cepharantha* Hayata (Figure [1\)](#page-0-0), which belongs to the bisbenzylisoquinoline type. Cepharanthine has anti-inflammatory, antibacterial, antioxidant, antihemolytic, and immunomodulatory effects [\[3\]](#page-10-2). Clinically, it is used to treat snake bites [\[4\]](#page-10-3), alopecia [\[5\]](#page-10-4), malaria [\[6\]](#page-10-5), and radioactive leucopenia [\[7\]](#page-10-6).

Figure 1. (A) Chemical structure of cepharanthine and (B) plant image of *Stephania cepharantha* Hayata.

Recently, cepharanthine has become a promising therapeutic molecule owing to its remarkable antiviral effect, particularly the inhibitory effect on severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which causes coronavirus disease 2019 (COVID-19). However, its poor water solubility results in low oral bioavailability. Therefore, to improve

Citation: Liang, D.; Li, Q.; Du, L.; Dou, G. Pharmacological Effects and Clinical Prospects of Cepharanthine. *Molecules* **2022**, 27, 8933. [https://](https://doi.org/10.3390/molecules27248933) doi.org/10.3390/molecules27248933

Academic Editor: Olívia R. Pereira

Received: 11 November 2022 $Accepted: 13 December 2022$ Published: 15 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Copyright: \odot 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$). $4.0/$).

its solubility and bioavailability, new dosage forms, such as dry powder inhalers (DPIs), liposomes, and nanoparticles, can be prepared, which can be used to provide a more effective dosing regimen for viral infections via pulmonary, oral, and intravenous administration [\[8\]](#page-11-0).

This review focuses on the important physicochemical and pharmacological properties and molecular mechanisms of cepharanthine and some new dosage forms that can be useful in improving its solubility and bioavailability, providing a basis for its clinical application. Additionally, this review summarizes cepharanthine's future development prospects.

2. Important Physicochemical Properties of Cepharanthine

Cepharanthine is a member of the bisbenzylisoquinoline cyclic alkaloid family [\[7\]](#page-10-6). It is also known as 12-O-methyl cepharanoline and is characterized by the presence of a double 1-benzylisoquinoline portion in its alkyl chain. It is mainly found in plants such as *Stephania epigaea* and *S. cepharantha* [\[9\]](#page-11-1). Its molecular formula is $C_{37}H_{38}N_2O_6$, and it is a generally white or yellow crystalline powder. In the chemical structure of cepharanthine, there are two head-to-head connected coclaurine units, assigning it an elliptical macrocyclic structure. This confers the unique chemical properties of ether solubility, optical activity, and the ability to reduce the mobility of various biofilms [\[5](#page-10-4)[,7\]](#page-10-6). Cepharanthine is easily soluble in acidic aqueous solutions and some organic solvents, such as methanol, ethanol, and DMSO, but is hardly soluble in water. It is cationic and amphiphilic [\[10\]](#page-11-2). These specific physicochemical properties lead to its low bioavailability. Doses of 1–60 mg per day have been safely and effectively used to treat various conditions. For example, the oral dose for alopecia areata treatment is usually 1.5–6 mg of CEP per day. The dose for the treatment of radiation-induced leukopenia is usually 50–60 mg of CEP per day. The dose for the treatment of multiple myeloma is usually 30 mg of CEP per day [\[10](#page-11-2)[,11\]](#page-11-3). CEP has a half-life of 31.3–36.9 h and achieves a steady state after five to six repeated doses of 100 mg/day, with a maximum of 9.6% residual drug after oral administration [\[12\]](#page-11-4). The time to reach the maximum serum concentration after a single oral dose of 10–60 mg in healthy men ranges from 1.1 to 2.5 h. After absorption in the body, CEP is extensively metabolized in the liver and distributed to various tissues [\[5\]](#page-10-4).

3. Pharmacological Effects of Cepharanthine

3.1. Antiviral Effects

A previous study has indicated that cepharanthine has a potent and extensive antiviral activity against viruses such as SARS-CoV, MERS-CoV, and HIV-1 [\[13\]](#page-11-5). It has been demonstrated that in chronically infected U1 cells, cepharanthine inhibits TNF-α or phorbol 12-myristate 13-acetate-induced HIV-1 replication by suppressing NF-κB activation [\[14\]](#page-11-6). The anti-HIV-1 activity of cepharanthine is dependent on its ability to inhibit NF-κB, and it inhibits the entry of HIV-1 by reducing plasma membrane fluidity [\[15\]](#page-11-7). Additionally, cepharanthine can affect the replication of viruses, such as hepatitis B virus [\[16\]](#page-11-8), herpes zoster virus [\[17\]](#page-11-9), and T-lymphotropic virus type 1 [\[18\]](#page-11-10). The extensive antiviral activity of cepharanthine may originate from the inhibition of intracellular inflammatory cytokines and chemokines [\[7\]](#page-10-6).

COVID-19 is caused by a novel coronavirus called severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which has a complex mechanism of action. The virus has been reported to affect several vital organs in the body, including the lungs, heart, brain, kidneys, gastrointestinal tract, blood, and immune system [\[19\]](#page-11-11). Several vaccines have been developed, but their availability is limited, and they can only have a preventive effect. To date, no specific therapeutic agent for coronavirus infection exists. Several drugs approved for COVID-19 by the US Food and Drug Administration (FDA) have previously been used for other diseases, such as raltegravir (Ebola) [\[20\]](#page-11-12), lopinavir–ritonavir (AIDS) [\[21\]](#page-11-13), chloroquine (malaria) [\[22\]](#page-11-14), and favipiravir (influenza) [\[23\]](#page-11-15) (Table [1\)](#page-2-0). In early 2020, Yigang et al. discovered that cepharanthine could be a potent drug for treating COVID-19 infection after screening thousands of monomeric compounds [\[24\]](#page-11-16).

Table 1. Approved drugs with anti-COVID-19 viral activity.

Subsequently, the anti-COVID-19 activity of cepharanthine has been confirmed repeatedly [\[25](#page-11-17)[,26\]](#page-11-18). The combined use of nelfinavir and cepharanthine could effectively inhibit SARS-CoV-2 replication, wherein cepharanthine inhibits the entry of SARS-CoV-2 by blocking the binding of the virus to target cells, and nelfinavir inhibits viral replication by inhibiting proteases [\[27\]](#page-11-19). Moreover, cepharanthine significantly inhibited the purified recombinant SARS-CoV-2 decapping enzyme, Nsp13, via in vitro enzyme activity assays. Nsp13 is essential for viral replication and is the most conserved nonstructural protein in the coronavirus family [\[26\]](#page-11-18). RNA-seq analysis has revealed that cepharanthine may exert antiviral effects through the reversal of heat shock factor-1-mediated heat shock response, endoplasmic reticulum stress/unfolded protein response, and hypoxic pathways of viral interference [\[28\]](#page-11-20). A pseudoviral model of SARS-CoV-2 demonstrated that cepharanthine can inhibit SARS-CoV-2 S protein/angiotensin-converting enzyme 2 (ACE2)-mediated membrane fusion by targeting host calcium ion channels and simultaneously upregulating intracellular cholesterol levels, thereby effectively inhibiting infection by SARS-CoV-2 mutants and different coronaviruses [\[13\]](#page-11-5). The human coronavirus OC43 model was used to screen 1900 clinically used drugs in vitro, which revealed that cepharanthine had the highest in vitro anti-SARS-CoV-2 activity, much higher than that of the already marketed anti-SARS-CoV-2 drug, raltegravir [\[29\]](#page-11-21).

The only clinical study to date of the proposed use of cepharanthine in the treatment of COVID-19 is the patent for an enteric formulation of cepharanthine by the Canadian pharmaceutical company, Pharmadrug Inc. This patent prepared cepharanthine as an enteric formulation for oral administration (PD-001), and its bioavailability in animal models was significantly improved. The company has already established GMP manufacturing for PD-001 and FDA-compliant preclinical safety/efficacy studies. Pharmadrug has submitted a Pre-Investigational New Drug Application to the FDA for the treatment of mild-tomoderate COVID-19 and will be working with the regulator to bring this medication for a complete evaluation.

3.2. Prevention of Leukopenia

Cepharanthine was first used to increase the number of leukocytes in the peripheral blood of patients undergoing radiotherapy or chemotherapy [\[7\]](#page-10-6). The major mechanisms involved are now widely recognized as the stimulation of the reticuloendothelial system, the activation of hematopoietic tissue, and the promotion of bone marrow proliferation, which ultimately increases the white blood cell count [\[30\]](#page-11-22).

Leukopenia is clinically defined as a reduction in the number of neutrophils and

can be caused by many factors, including chemotherapy and radiotherapy for tumors and chemicals such as benzene. In the 1930s and 1940s, cepharanthine was used to treat pulmonary tuberculosis. It was found that patients with tuberculosis had an increased number of leukocytes in the peripheral blood after cepharanthine administration, which indicated the prospect of cepharanthine in preventing leukopenia. In several clinical studies involving more than 350 patients, radiotherapy was used along with cepharanthine to treat head and neck tumors and ovarian, lung [\[10\]](#page-11-2), and breast cancers [\[31\]](#page-11-23). All studies showed that cepharanthine helped prevent leukopenia in patients receiving anticancer treatment.

3.3. Antitumor Effects

Cepharanthine exhibits antitumor effects by enhancing immunity [\[32\]](#page-11-24) and inhibiting tumor cell proliferation [\[33\]](#page-12-0), as well as increasing the sensitivity of tumor cells to radiotherapy while reducing the adverse effects caused by the radiotherapy [\[34\]](#page-12-1). A more promising approach is the use of cepharanthine in combination with other chemotherapeutic drugs to reverse multidrug resistance (MDR) of tumor cells [\[35\]](#page-12-2). CEP effectively inhibits the drug transporter protein ABCC10 (also known as MRP7) on cancer cell membranes, thereby reversing anticancer drug resistance in cells expressing ABCC10. This allows for the increased accumulation of anticancer drugs in cancer cells and inhibits their efflux [\[36\]](#page-12-3), thus improving the efficacy of anticancer drugs. CEP also interferes with other transporter proteins, such as ABCB1 (also known as MDR1 or P-glycoprotein) [\[37\]](#page-12-4), possibly by inhibiting the PI3K/Akt signaling pathway, leading to the downregulation of ABCB1 expression in cancer cells, which in turn reverses MDR [\[38\]](#page-12-5). CEP potentially interacts directly with specific drug transporter proteins present on the plasma membrane. For example, neferine, another bisbenzylisoquinoline alkaloid with a similar structure to CEP, binds strongly to and inhibits ABCB1, thereby reversing MDR. Therefore, it is speculated that CEP has a similar biological effect [\[39\]](#page-12-6).

Cepharanthine can be involved in tumor prevention and treatment through various mechanisms, including increasing the white blood cell count and exerting immunomodulatory effects on macrophages, T cells, and natural killer cells to increase the immune capacity [\[32](#page-11-24)[,40\]](#page-12-7). Additionally, it can play a role in inducing apoptosis, inhibiting tumor cell infiltration and metastasis.

3.4. Anti-Inflammatory Effects

Cepharanthine is also effective for inflammation in vivo, especially in the ears, nose, throat, and mouth. It can significantly decrease the expression of certain inflammatory cytokines, such as IL-6, IL-1 β , and TNF- α [\[41\]](#page-12-8). In a mouse model of lipopolysaccharide (LPS)-induced mastitis, cepharanthine significantly reduced neutrophil infiltration and decreased TNF- α , IL-1 β , and IL-6 levels [\[41\]](#page-12-8). Presently, cepharanthine is used to treat inflammatory diseases such as rheumatism, arthritis, lumbago, nephritis, edema, and dysentery [\[7\]](#page-10-6).

3.5. Immunomodulation

Cepharanthine can be used as an immunomodulator, and it has excellent potential in the treatment of various autoimmune diseases and allergies [\[42\]](#page-12-9). At a low dose, cepharanthine could effectively prevent progressive thrombocytopenia and was used to successfully treat a patient with multiple myeloma combined with immune thrombocytopenic purpura [\[43\]](#page-12-10). Additionally, cepharanthine specifically inhibits abnormally activated T cells or steroid-resistant human leukemia T cells [\[44,](#page-12-11)[45\]](#page-12-12). Furthermore, cepharanthine regulates several signaling pathways in abnormally activated T cells, such as NF-κB, caspase cascade, cell cycle, mitogen-activated protein kinase (MAPK), and PI3K/Akt/mTOR pathways [\[46\]](#page-12-13).

4. Molecular Mechanisms of Cepharanthine *4.1. NF‐κB Pathway* $T_{\text{F}-k}$ R pathway plays a central role in induction in induction T

4.1. NF-κB Pathway

The NF-κB pathway plays a central role in inducing pro-inflammatory gene expression and is a potent regulator of inflammatory molecules. In resting cells, NF- κ B exists in the upique is the upique of the IRB protein via the URB. cytoplasm as a latent, inactive IκB-binding complex. Upon relevant stimulation, IκB
history (IKK) in human handscription of the LP anattic right the nucleus and conven kinase (IKK) induces phosphorylation of the I_{KB} protein via the ubiquitin–proteasome
and activating specific target general decoration allowing ME and disconsistent in and produce and pathway, resulting in its rapid degradation, allowing NF-κB dimers to enter the nucleus and ranthing, resulting in the rapid degradation, allowing τις που allied to enter the Indeceds and
activating specific target gene expression [\[46\]](#page-12-13). In an LPS-stimulated model of RAW264.7 cells, cepharanthine inhibited NF-κB activation by blocking the IKK pathway [\[47\]](#page-12-14) (Figure [2\)](#page-4-0). In this study, cepharanthine (2.5, 5, and 10 μ g/mL) inhibited the release of pro-inflammatory in this study, cepharanthine (2.5, 5, and 10 μ g/mL) inhibited the release of pro-inflammatory $\frac{1}{2}$ factors such as TNF α , IL-6, and IL-1 β in a dose-dependent manner with potentially no cytotoxicity. In addition, 10 mg/kg/day of cepharanthine could significantly inhibit the expression of NF-κB and reduce the levels of the pro-inflammatory cytokines, IL-1β and TNF- α , in diabetic rats [\[48\]](#page-12-15). Thus, it seems that cepharanthine can reduce the expression of inflammatory factors in related disease models through the NF-KB pathway and exert anti-inflammatory effects as a potential drug for the treatment of certain related diseases. the tyto-kb pattiway plays a central fole in including pro-imagination y gene expression.

Figure 2. Possible molecular mechanisms of cepharanthine in cells. (1) NF-KB pathway, (2) apoptosis, (3) cell cycle control, (4) MAPK pathway, (5) PI3K/Akt/mTOR signaling pathway, and (6) P-glycoprotein expression.

4.2. Apoptosis 4.2. Apoptosis

Apoptosis is a regulated cellular suicide mechanism characterized by nuclear conden-sation, cell shrinkage, membrane leakage, and DNA fragmentation [\[49\]](#page-12-16). The cascade of caspases, which belong to the cysteine protease family, is a key apoptosis regulator. To date, only two types of caspases have been defined, including initiator and effector caspases. Initiator caspases, such as caspase-2, -4, -8, -9, -10, and -12, are closely associated with proapoptotic signaling. Upon stimulation by apoptosis, these initiator caspases divide and activate downstream effector caspases, such as caspase-3, -6, and -7. These effector caspases hydrolyze a range of substrates, generating signals that lead to cell death [\[50](#page-12-17)[,51\]](#page-12-18). The cell growth inhibition of primary effusion lymphoma (PEL) cell lines was enhanced in a dose-dependent manner as the cepharanthine dose was increased from 1 to 10 μ g/mL. Moreover, cepharanthine-treated PEL cells induced the activation of caspase-3, which is an apoptosis marker [\[52\]](#page-12-19). In addition, cepharanthine at high concentrations $(10-20 \mu g/mL)$ decreased the mitochondrial membrane potential of dendritic cells, and high levels of cellular stress caused the release of cytochrome c from mitochondria. This activates caspase-9, which then activates the effector caspase-3/7, and finally triggers apoptosis [\[53\]](#page-12-20).

4.3. Cell Cycle Control

The cell cycle is regulated by different cellular proteins, such as the cell cycle protein A/B/D [\[54\]](#page-12-21). It has been shown that cepharanthine affects the cell cycle, usually arresting cells in the G1 and S phases. Jurkat T cells treated with 5, 10, and 15 μ M CEP showed a dose-dependent inhibition of cell cycle progression in the S phase, significantly reducing the number of cells in the G0/G1 phase [\[45\]](#page-12-12). Further research has revealed that cepharanthine upregulates the expression of cell cycle proteins A2 and B1 but downregulates that of the cell cycle protein D1 in Jurkat T cells, possibly relating to cell cycle arrest [\[46\]](#page-12-13). Using $0, 1, 5, 10$, and 20μ M cepharanthine also effectively reverses MDR-mediated resistance to cisplatin in esophageal squamous cell carcinoma (ESCC) cells, inhibiting the proliferation of ESCC cell lines and inducing G2/M phase cell cycle arrest. Dose-dependent upregulation of p53 and downstream p21 induced by cepharanthine explains its mechanism of causing cell cycle arrest [\[55\]](#page-12-22).

4.4. MAPK Pathway

Activated MAPK has been reported to play an important role in promoting and maintaining T lymphocyte populations [\[56\]](#page-12-23). Cepharanthine regulates multiple signaling pathways that aberrantly activate T cells at low toxicity, and the antiproliferative effect on human peripheral blood T cells occurs partly through the blockade of MAPKs, such as JNK, p38, and ERK activity [\[57\]](#page-12-24). It was reported that another class of bisbenzylisoquinoline alkaloids, tetrandrine, in combination with methylprednisolone (a glucocorticoid), significantly inhibited the phosphorylation of the mitogen-activated protein kinase family. Powdered tetrandrine (3 μ M) in combination with 0.5 ng/mL methylprednisolone showed synergistic inhibition of both ERK1/2 and P38. The powdered antifungal base significantly reduced the IC50 value of methylprednisolone but had no significant toxic effect on normal cells [\[46\]](#page-12-13). These evaluations suggest that CEP, a member of the bisbenzylisoquinoline alkaloid family, may have similar efficacy and could be used as a lead compound for the development of new drugs for the treatment of T-cell-related diseases or to address glucocorticoid resistance.

4.5. PI3K/Akt/mTOR Signaling Pathway

The PI3K/Akt/mTOR signaling pathway is involved in regulating various cellular responses, such as metabolic regulation [\[58\]](#page-13-0), cell proliferation, transcription, translation, survival, and growth [\[58](#page-13-0)[,59\]](#page-13-1). This pathway is essential for the pathological and physiological conditioning of humans, and changes in the regulation of this pathway may lead to the development of various cancers [\[60\]](#page-13-2). Thus, the PI3K/Akt/mTOR signaling pathway is a potential target for antitumor therapy. In a study on breast cancer cells, treatment with 5 and 10 μ M cepharanthine was found to reduce the levels of both phosphorylated AKT and mTOR. The downstream targets of mTOR (p70S6K, p-S6, and p-4E-BP1) were significantly reduced, whereas GSK3β, a downstream effector of AKT [\[61\]](#page-13-3), showed similarly reduced levels in cells after CEP treatment. These results suggest that cepharanthine induces apoptosis and autophagy in breast cancer cells by inhibiting the AKT/mTOR signaling pathway[\[62\]](#page-13-4). Cepharanthine can inhibit the expression of p-PI3K and mTOR in Jurkat T cells; however, this resulted in high expression of p-Akt1 [\[45\]](#page-12-12). This result may appear to be incongruous, as Akt is considered a major downstream effector of PI3K in physiological processes [\[63\]](#page-13-5). However, several studies have demonstrated that different groups of tyrosine (Ack1/TNK2, Src, and PTK6) and serine/threonine (TBK1, IKBKE, and DNAPKcs) kinases can directly activate Akt [\[64\]](#page-13-6). Thus, CEP may regulate p-PI3K and mTOR expression independently of Akt.

4.6. P-glycoprotein Expression

P-glycoprotein (also known as ABCB1) belongs to the ABC superfamily of transporter proteins [\[65\]](#page-13-7) and is encoded by multidrug resistance gene 1 (*MDR-1*), which is an ATPdependent membrane transporter protein [\[66\]](#page-13-8). The overexpression of drug transporter

proteins present in cancer cell membranes is a major cause of MDR to cancer chemotherapy. Cepharanthine can reverse the resistance of tumor cells to many chemotherapy drugs by interfering with P-glycoprotein [\[37\]](#page-12-4). Cepharanthine may act as a potent P-glycoprotein inhibitor, dose-dependently restoring antitumor activity [\[67\]](#page-13-9). In a study on glucocorticoid resistance, the human T-lymphoblast leukemia cell line MOLT-4 with low P-glycoprotein expression, and its MDR subline MOLT-4/DNR, were used as the model. Tetrandrine (Figure [3\)](#page-6-0), being likewise a bisbenzylisoquinoline-like alkaloid and highly resembling cepharanthine, indirectly modulated the translocation of glucocorticoid receptors by inhibiting the efflux function of P-glycoprotein in MOLT-4/DNR cells and downregulated its protein expression, thereby enhancing the pharmacodynamic effects of glucocorticoids. The expression of P-glycoprotein was downregulated in MOLT-4/DNR cells in a concentrationdependent manner under treatment with 0.03, 0.3, and 1 μ M tetrandrine. Although 1 μ M tetrandrine significantly inhibited P-glycoprotein expression, MOL T-4/DNR 296 cells exhibited cytotoxic effects at this dose. In contrast, glucocorticoid receptor translocation was almost completely restored at 0.3 μ M with no notable side effects [\[68\]](#page-13-10). However, studies on the resistance of CEP to glucocorticoids remain unclear, and the actual effective dose needs to be explored further. However, it can be deduced that this bisbenzylisoquinoline alkaloid of natural plant origin is beneficial for glucocorticoid-resistant diseases caused by P-glycoprotein overexpression.

porter proteins present in cancer cell membranes is a major cause of MDR to cancer chem‐

Figure 3. Chemical structure of tetrandrine.

5. New Dosage Forms of Cepharanthine

Presently, the commercial formulations of cepharanthine are ordinary tablets. As cepharanthine is insoluble in water, it is distributed in tablets in the form of larger granules. It is administered at larger doses, and the rate of dissolution and diffusion after oral administration is slow, significantly affecting its absorption and utilization in vivo. Therefore, it is important to explore new dosage forms of cepharanthine to widen its clinical application.

5.1. Oral Formulation

5.1.1. Oral Disintegrating Tablets

Oral administration is the most widely used and convenient route of administration, with a high degree of stability and portable packaging [\[69\]](#page-13-11). Orally disintegrating tablets, as a delivery system, are those that disintegrate or dissolve rapidly in the mouth without needing water and are absorbed through the antral mucosa [\[70\]](#page-13-12). Compared with ordinary tablets, orally disintegrating tablets have a faster onset of action, avoid the liver first-pass effect, have higher bioavailability, are less irritating to the esophagus and gastrointestinal tract, and have higher compliance. Therefore, they are ideal for children, elderly patients, and patients with swallowing difficulties [\[71,](#page-13-13)[72\]](#page-13-14). The preparation methods include conventional techniques such as freeze drying [\[73\]](#page-13-15), spray drying [\[74\]](#page-13-16), direct compression [\[75\]](#page-13-17), and molding [\[76\]](#page-13-18) and new techniques such as marshmallow technology [\[77\]](#page-13-19), microwaveassisted irradiation [\[78\]](#page-13-20), sublimation [\[79\]](#page-13-21), and 3D printing [\[80\]](#page-13-22). It is possible to consider formulating cepharanthine as an orally disintegrating tablet to improve its bioavailability and reduce its dose.

5.1.2. Dropping Pills

As a form of solid dispersion, dropping pills are the preparations made by heating or mixing drugs with a matrix, then dropping them into an immiscible condensate, causing the droplets to shrink and condense due to surface tension [\[81\]](#page-13-23). Unlike conventional tablets, dropping pills have the advantages of small dosage, fewer adverse effects, fast dissolution, high bioavailability, good efficacy, easy administration, excellent drug stability, simple preparation, and easy quality control [\[82\]](#page-13-24). They are good candidates to improve drug dissolution and increase oral bioavailability. In the preparation of dropping pills, a solid dispersion is formed, and drugs can be present in the form of separated molecules or amorphous particles, which substantially improves their solubility [\[83\]](#page-13-25). Cepharanthine dropping pills have been prepared with PEG 4000 and PEG 6000 serving as the excipients using the solid dispersion technology to improve their solubility, thereby achieving rapid and high efficiency [\[84\]](#page-13-26).

5.2. Injections

Although oral administration is the preferred route of administration due to its convenience and noninvasiveness, it is impossible to deliver all drugs orally due to low bioavailability and patients' tolerance to oral intake. Therefore, injections can be an effective alternative [\[85\]](#page-13-27). The advantages of injections include rapid efficacy, direct entry into the bloodstream when administered without passing through the gastrointestinal tract, improved bioavailability and reduced drug interactions, targeted efficacy, and suitability for drugs that are unsuitable in oral dosage forms or for patients who cannot receive oral administration [\[86\]](#page-13-28).

After the COVID-19 outbreak, several Chinese medicine injections, including the Reduning injection [\[87\]](#page-14-0), Xiyanping injection [\[88\]](#page-14-1), and Xuebijing injection [\[89\]](#page-14-2), were used as specific therapeutical agents. The Chinese pharmaceutical company BUCHANG has applied for a patent on the preparation and testing method of cepharanthine hydrochloride injection. The preparation method is simple and controllable, with reliable quality and high bioavailability, which can overcome many disadvantages of oral administration of cepharanthine and has good application prospects.

5.3. Pulmonary Drug Delivery Systems—DPIs

A pulmonary drug delivery system is a system that delivers drugs to the lungs to produce local or systemic therapeutic effects [\[90\]](#page-14-3). Compared with conventional drug delivery, pulmonary drug delivery acts faster, avoids hepatic first-pass effects, reduces the dose required and adverse effects associated with systemic drug delivery, and substantially improves patient compliance [\[91\]](#page-14-4). Currently, there are three types of pulmonary delivery systems: metered-dose inhalers, DPIs, and nebulizers. DPI sales account for approximately 50% of all global sales of pulmonary drug delivery systems, far exceeding the sales of the other two dosage forms. It is safe and environmentally friendly as no projectile agent or preservative is used in the DPI. Furthermore, the stability of the drugs existing as a dry powder is significantly improved [\[92\]](#page-14-5). Common processes for preparing DPIs include spray drying, freeze drying, airflow pulverization, supercritical fluid pulverization, grinding and pulverization, etc. [\[93\]](#page-14-6). The ideal DPI should meet the following criteria: high dispersibility, drug stability, narrow aerodynamic particle size distribution, low particle surface energy and potential, good dose reproducibility, and a sustained release or specific targeting mechanism when required [\[94\]](#page-14-7).

Cepharanthine is a water-insoluble drug that is appropriate as a DPI for pulmonary inhalation to treat COVID-19 associated with its sequelae and complications because the vital infection organ of SARS-CoV-2 is the lung.

5.4. Nanoformulations

5.4.1. Liposomes

Liposomes are miniature vesicles formed by encapsulating a drug within a lipid-like bilayer. It is considered a highly promising drug delivery system because of its biofilm-like structure, ability to encapsulate both water and fat-soluble drugs, ability to reduce drug dose, reduce toxicity, mitigate metabolic and immune reactions, delay release, reduce the rate of elimination in the body, alter drug distribution in the body, and its ability to target drug release [\[95\]](#page-14-8). The liposome encapsulation technology can solve the challenge of drug dissolution. By encapsulating unstable, oxidizable drugs in liposomes, these drugs are protected by lipid membranes, which can reduce the probability of events such as enzymatic degradation, chemical and immunological inactivation, and rapid plasma clearance, helping improve and prolong their action [\[96\]](#page-14-9). Among the hundreds of nanoformulations approved by the FDA, liposomes account for the highest percentage (33%) [\[97\]](#page-14-10). Furthermore, various administration routes, such as injection, oral, transdermal, transnasal, transpulmonary, and ophthalmic, have been developed.

As a water-insoluble drug, cepharanthine can be designed as a formulation encapsulated in liposomes for transpulmonary or oral administration for treating COVID-19. This solves the problem of the low solubility of cepharanthine in water and improves its bioavailability in vivo.

5.4.2. Nanoparticles

Nanoparticles are sphere-like particles formed by dissolving or encapsulating a drug in a polymeric material, usually between 10 nm and 100 nm in size, which are more effectively absorbed by cells than larger molecules [\[98\]](#page-14-11). In recent years, nanoparticles have been widely used for drug delivery and have proven to be effective in drug delivery and diagnostics [\[92\]](#page-14-5). Compared with other types of drug-carrying particles, nanoparticles have a higher drugcarrying capacity [\[99\]](#page-14-12), permeate permeability barriers, prolong circulation times [\[100\]](#page-14-13), and increase cellular uptake [\[101\]](#page-14-14), thus improving safety and efficacy. Nanoparticles typically have a large surface-area-to-volume ratio, and this feature improves their dissolution properties, thereby increasing solubility and intracellular drug delivery potential. However, engineered nanoparticles are foreign to the organism, and once they enter the body, they immediately activate the innate immune system and generate a specific immune response based on their properties [\[102\]](#page-14-15). Currently, it is possible to evade the immune response by designing nanoparticles that mimic host cells [\[103\]](#page-14-16), adjusting their physicochemical properties such as preparing them in different sizes and shapes with a surface charge [\[104\]](#page-14-17), or modifying the surface of the particles with a bionic coating [\[105\]](#page-14-18).

Bionic anti-inflammatory nanoformulations of cepharanthine for the treatment of acute lung injury (ALI) have been reported, which are based on the "homing" of macrophages to sites of inflammation and cell membrane coating nanotechnology. Researchers used macrophage membranes isolated from the RAW264.7 monocyte/macrophage cell line to encapsulate nanostructured lipid carriers loaded with cepharanthine. The nanoparticles were then injected into the lung inflammation sites of ALI mice via tail vein injection. Long-term circulation was achieved in vivo and showed the ability to target inflamed lungs [\[106\]](#page-14-19).

5.5. Summary

Based on the above-listed several new dosage forms of cepharanthine, we have summarized their advantages and disadvantages as well as those that need attention during clinical development. Orally disintegrating tablets are convenient to administer and highly stable, while avoiding the first-pass effect in the liver and improving the low bioavailability of cepharanthine. However, orally disintegrating tablets have the disadvantages of fragility and moisture absorption; therefore, there is still a need to improve the formulation and packaging materials to enhance their application. Another oral formulation, drops, in the form of a solid dispersion formed during its preparation can effectively improve the

solubility of cepharanthine while enhancing its bioavailability. However, quality control of this new dosage form is difficult, and quality standards are lacking; therefore, there are no new dosage forms of drops approved for marketing. Injections are another drug delivery method that have rapid efficacy and are suitable for patients who cannot receive oral administration. However, injectable drug delivery faces the challenge of poor water solubility of active ingredients, and suitable solubilization strategies need to be adopted according to the chemical structure and physicochemical properties of insoluble drugs. In addition, pulmonary drug delivery systems are superior to traditional drug delivery methods for their advantages, such as fast-acting effects and avoidance of the hepatic first-pass effect. Cepharanthine is a water-insoluble drug suitable for use as a DPI for pulmonary inhalation. What requires attention when developing dosage forms is designing the formulation so that it meets the ideal DPI criteria. Nanoformulations, including liposomes and nanoparticles, have been widely used for drug delivery in recent years as a new dosage form. They can encapsulate water-insoluble drugs in microcapsules or particles to achieve targeted and delayed drug release. However, these substances are foreign to the organism and can stimulate an immune response when entering the body. Therefore, the most important factor when designing a nanoformulation is addressing the immune rejection reaction.

6. Conclusions

Cepharanthine has become a promising molecule due to its excellent antiviral activity against SARS-CoV-2. This review summarizes the important physicochemical properties and pharmacological effects of cepharanthine (Table [2\)](#page-9-0), including its anti-COVID-19 effect. Additionally, the molecular mechanisms (Table [3\)](#page-10-7) and novel dosage formulations for the efficient, safe, and convenient delivery of cepharanthine are summarized, including oral, injectable, DPIs, and nanoformulations. The number of studies on cepharanthine has shown exponential growth following the discovery of the remarkable potential of cepharanthine, an "old" drug derived from a natural plant, in the fight against COVID-19. However, cepharanthine is mainly used clinically for treating leukopenia caused by radiotherapy and chemotherapy for tumors, while its excellent anti-COVID-19 activity has only been demonstrated in vitro and at the cellular level. Therefore, it is important to determine methods to improve the bioavailability of cepharanthine in vivo so that it could exert a marked anti-SARS-CoV-2 effect in vivo, making it truly applicable in COVID-19 treatment.

Table 2. Pharmacological effects of cepharanthine.

Table 3. Detailed molecular mechanisms of cepharanthine in cells.

Author Contributions: Conceptualization, D.L., L.D. and G.D.; writing—original draft preparation, D.L.; drawing figures, Q.L.; writing—review and editing, L.D.; project administration, L.D. and G.D.; funding acquisition, L.D. and G.D. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Beijing Municipal Natural Science Foundation (No. 20SWAQK22).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Rdel, C.O.; Kellogg, E.A.; Werff, H.V. Molecular phylogeny of the moonseed family (Menispermaceae): Implications for morphological diversification. *Am. J. Bot.* **2007**, *94*, 1425–1438.
- 2. Thavamani, B.S.; Mathew, M.; Dhanabal, S.P. In vitro cytotoxic activity of menispermaceae plants against HeLa cell line. *Anc. Sci. Life* **2013**, *33*, 81–84. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25284939)
- 3. Kao, M.-C.; Yang, C.-H.; Chou, W.-C.; Sheu, J.-R.; Huang, C.-J. Cepharanthine mitigates lung injury in lower limb ischemia– reperfusion. *J. Surg. Res.* **2015**, *199*, 647–656. [\[CrossRef\]](http://doi.org/10.1016/j.jss.2015.06.041)
- 4. Hifumi, T.; Yamamoto, A.; Morokuma, K.; Okada, I.; Kiriu, N.; Ogasawara, T.; Hasegawa, E.; Kato, H.; Inoue, J.; Koido, Y.; et al. Clinical Efficacy of Antivenom and Cepharanthine for the Treatment of Mamushi (*Gloydius blomhoffii*) Bites in Tertiary Care Centers in Japan. *Jpn. J. Infect. Dis.* **2013**, *66*, 26–31. [\[CrossRef\]](http://doi.org/10.7883/yoken.66.26) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23429081)
- 5. Rogosnitzky, M.; Okediji, P.; Koman, I. Cepharanthine: A review of the antiviral potential of a Japanese-approved alopecia drug in COVID-19. *Pharmacol. Rep.* **2020**, *72*, 1509–1516. [\[CrossRef\]](http://doi.org/10.1007/s43440-020-00132-z) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32700247)
- 6. Desgrouas, C.; Dormoi, J.; Chapus, C.; Ollivier, E.; Parzy, D.; Taudon, N. In vitro and in vivo combination of cepharanthine with anti-malarial drugs. *Malar. J.* **2014**, *13*, 90. [\[CrossRef\]](http://doi.org/10.1186/1475-2875-13-90)
- 7. Bailly, C. Cepharanthine: An update of its mode of action, pharmacological properties and medical applications. *Phytomedicine* **2019**, *62*, 152956. [\[CrossRef\]](http://doi.org/10.1016/j.phymed.2019.152956)
- 8. Fan, H.; He, S.T.; Han, P.; Hong, B.; Liu, K.; Li, M.; Wang, S.; Tong, Y. Cepharanthine: A Promising Old Drug against SARS-CoV-2. *Adv. Biol. (Weinh.)* **2022**, e2200148. [\[CrossRef\]](http://doi.org/10.1002/adbi.202200148)
- 9. Hong, L.; Guo, Z.; Huang, K.; Wei, S.; Liu, B.; Meng, S.; Long, C. Ethnobotanical study on medicinal plants used by Maonan people in China. *J. Ethnobiol. Ethnomed.* **2015**, *11*, 32. [\[CrossRef\]](http://doi.org/10.1186/s13002-015-0019-1)
- 10. Rogosnitzky, M.; Danks, R. Therapeutic potential of the biscoclaurine alkaloid, cepharanthine, for a range of clinical conditions. *Pharmacol. Rep.* **2011**, *63*, 337–347. [\[CrossRef\]](http://doi.org/10.1016/S1734-1140(11)70500-X)
- 11. Kikukawa, Y.; Okuno, Y.; Tatetsu, H.; Nakamura, M.; Harada, N.; Ueno, S.; Kamizaki, Y.; Mitsuya, H.; Hata, H. Induction of cell cycle arrest and apoptosis in myeloma cells by cepharanthine, a biscoclaurine alkaloid. *Int. J. Oncol.* **2008**, *33*, 807–814.
- 12. Yasuda, K.; Moro, M.; Akasu, M.; Ohnishi, A. Pharmacokinetic disposition of Cepharanthin following single and multiple intravenous doses in healthy subjects. *Rinsho Yakuri/Jpn. J. Clin. Pharmacol. Ther.* **1989**, *20*, 741–749. [\[CrossRef\]](http://doi.org/10.3999/jscpt.20.741)
- 13. He, C.L.; Huang, L.Y.; Wang, K.; Gu, C.J.; Hu, J.; Zhang, G.J.; Xu, W.; Xie, Y.H.; Tang, N.; Huang, A.L. Identification of bisbenzylisoquinoline alkaloids as SARS-CoV-2 entry inhibitors from a library of natural products. *Signal Transduct. Target* **2021**, *6*, 131. [\[CrossRef\]](http://doi.org/10.1038/s41392-021-00531-5)
- 14. Okamoto, M.; Ono, M.; Baba, M. Potent Inhibition of HIV Type 1 Replication by an Antiinflammatory Alkaloid, Cepharanthine, in Chronically Infected Monocytic Cells. *AIDS Res. Hum. Retrovir.* **1998**, *14*, 1239–1245. [\[CrossRef\]](http://doi.org/10.1089/aid.1998.14.1239)
- 15. Matsuda, K.; Hattori, S.; Komizu, Y.; Kariya, R.; Ueoka, R.; Okada, S. Cepharanthine inhibited HIV-1 cell–cell transmission and cell-free infection via modification of cell membrane fluidity. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 2115–2117. [\[CrossRef\]](http://doi.org/10.1016/j.bmcl.2014.03.041) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24704028)
- 16. Zhou, Y.-B.; Wang, Y.-F.; Zhang, Y.; Zheng, L.-Y.; Yang, X.-A.; Wang, N.; Jiang, J.-H.; Ma, F.; Yin, D.-T.; Sun, C.-Y.; et al. In vitro activity of cepharanthine hydrochloride against clinical wild-type and lamivudine-resistant hepatitis B virus isolates. *Eur. J. Pharmacol.* **2012**, *683*, 10–15. [\[CrossRef\]](http://doi.org/10.1016/j.ejphar.2012.02.030) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22387093)
- 17. Liu, Y.; Tang, Q.; Rao, Z.; Fang, Y.; Jiang, X.; Liu, W.; Luan, F.; Zeng, N. Inhibition of herpes simplex virus 1 by cepharanthine via promoting cellular autophagy through up-regulation of STING/TBK1/P62 pathway. *Antivir. Res.* **2021**, *193*, 105143. [\[CrossRef\]](http://doi.org/10.1016/j.antiviral.2021.105143) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34303748)
- 18. Toyama, M.; Hamasaki, T.; Uto, T.; Aoyama, H.; Okamoto, M.; Hashmoto, Y.; Baba, M. Synergistic inhibition of HTLV-1-infected cell proliferation by combination of cepharanthine and a tetramethylnaphthalene derivative. *Anticancer Res.* **2012**, *32*, 2639–2645.
- 19. Guo, G.; Ye, L.; Pan, K.; Chen, Y.; Xing, D.; Yan, K.; Chen, Z.; Ding, N.; Li, W.; Huang, H.; et al. New Insights of Emerging SARS-CoV-2: Epidemiology, Etiology, Clinical Features, Clinical Treatment, and Prevention. *Front. Cell Dev. Biol.* **2020**, *8*, 410. [\[CrossRef\]](http://doi.org/10.3389/fcell.2020.00410)
- 20. Beigel, J.H.; Tomashek, K.M.; Dodd, L.E.; Mehta, A.K.; Zingman, B.S.; Kalil, A.C.; Hohmann, E.; Chu, H.Y.; Luetkemeyer, A.; Kline, S.; et al. Remdesivir for the Treatment of COVID-19—Preliminary report. *N. Engl. J. Med.* **2020**, *383*, 1813–1826. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa2007764)
- 21. Cao, B.; Wang, Y.; Wen, D.; Liu, W.; Wang, J.; Fan, G.; Ruan, L.; Song, B.; Cai, Y.; Wei, M.; et al. A Trial of Lopinavir–Ritonavir in Adults Hospitalized with Severe COVID-19. *N. Engl. J. Med.* **2020**, *382*, 1787–1799. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa2001282) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32187464)
- 22. Boulware, D.R.; Pullen, M.F.; Bangdiwala, A.S.; Pastick, K.A.; Lofgren, S.M.; Okafor, E.C.; Skipper, C.P.; Nascene, A.A.; Nicol, M.R.; Abassi, M.; et al. A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. *N. Engl. J. Med.* **2020**, *383*, 517–525. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa2016638) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32492293)
- 23. Dong, L.; Hu, S.; Gao, J. Discovering drugs to treat coronavirus disease 2019 (COVID-19). *Drug Discov. Ther.* **2020**, *14*, 58–60. [\[CrossRef\]](http://doi.org/10.5582/ddt.2020.01012)
- 24. Fan, H.; Liu, K.; Hong, B.; He, S.; Han, P.; Li, M.; Wang, S.; Tong, Y. Progress in the study of antiviral activity of cepharanthine against SARS-CoV-2. *Nan Fang Yi Ke Da Xue Xue Bao* **2022**, *42*, 955–956. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35790449)
- 25. Ohashi, H.; Watashi, K.; Saso, W.; Shionoya, K.; Iwanami, S.; Hirokawa, T.; Shirai, T.; Kanaya, S.; Ito, Y.; Kim, K.S.; et al. Multidrug treatment with nelfinavir and cepharanthine against COVID-19. *bioRxiv* **2020**, 039925. [\[CrossRef\]](http://doi.org/10.1101/2020.04.14.039925)
- 26. White, M.A.; Lin, W.; Cheng, X. Discovery of COVID-19 Inhibitors Targeting the SARS-CoV-2 Nsp13 Helicase. *bioRxiv* **2020**, 243246. [\[CrossRef\]](http://doi.org/10.1021/acs.jpclett.0c02421) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33052685)
- 27. Ohashi, H.; Watashi, K.; Saso, W.; Shionoya, K.; Iwanami, S.; Hirokawa, T.; Shirai, T.; Kanaya, S.; Ito, Y.; Kim, K.S.; et al. Potential anti-COVID-19 agents, cepharanthine and nelfinavir, and their usage for combination treatment. *Iscience* **2021**, *24*, 102367. [\[CrossRef\]](http://doi.org/10.1016/j.isci.2021.102367)
- 28. Li, S.; Liu, W.; Chen, Y.; Wang, L.; An, W.; An, X.; Song, L.; Tong, Y.; Fan, H.; Lu, C. Transcriptome analysis of cepharanthine against a SARS-CoV-2-related coronavirus. *Brief Bioinf.* **2021**, *22*, 1378–1386. [\[CrossRef\]](http://doi.org/10.1093/bib/bbaa387)
- 29. Drayman, N.; De Marco, J.K.; Jones, K.A.; Azizi, S.A.; Froggatt, H.M.; Tan, K.; Maltseva, N.I.; Chen, S.; Nicolaescu, V.; Dvorkin, S.; et al. Masitinib is a broad coronavirus 3CL inhibitor that blocks replication of SARS-CoV-2. *Science* **2021**, *373*, 931–936. [\[CrossRef\]](http://doi.org/10.1126/science.abg5827)
- 30. Oyaizu, H.; Adachi, Y.; Yasumizu, R.; Ono, M.; Ikebukuro, K.; Fukuhara, S.; Ikehara, S. Protection of T cells from radiation-induced apoptosis by Cepharanthin®. *Int. Immunopharmacol.* **2001**, *1*, 2091–2099. [\[CrossRef\]](http://doi.org/10.1016/S1567-5769(01)00127-8)
- 31. Terasaki, M.; Abe, T.; Miyagi, N.; Ogo, E.; Shigemori, M. Feasibility and response to 1-(4-amino-2-methyl-5-pyrimidynyl) methyl-3-(2-chloroethyl)-3-nitrosourea hydrochloride chemotherapy with pre-treated procarbazine for elderly patients with newly diagnosed glioblastoma. *J. Neuro-Oncol.* **2007**, *81*, 265–269. [\[CrossRef\]](http://doi.org/10.1007/s11060-006-9223-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16937011)
- 32. Ito, H.; Ito, H.; Amano, H.; Noda, H. Inhibitory effect of a biscoclaurine alkaloid, cepharanthin, on lung metastasis of Lewis lung carcinoma. *Jpn. J. Pharmacol.* **1991**, *56*, 195–202. [\[CrossRef\]](http://doi.org/10.1016/S0021-5198(19)39880-4) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/1880998)
- 33. Wu, J.; Suzuki, H.; Zhou, Y.-W.; Liu, W.; Yoshihara, M.; Kato, M.; Akhand, A.A.; Hayakawa, A.; Takeuchi, K.; Hossain, K.; et al. Cepharanthine activates caspases and induces apoptosis in Jurkat and K562 human leukemia cell lines. *J. Cell. Biochem.* **2001**, *82*, 200–214. [\[CrossRef\]](http://doi.org/10.1002/jcb.1155) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11527146)
- 34. Tamatani, A.M.; Motegi, K.; Takamaru, N.; Kawashima, Y.; Bando, T. Cepharanthin-enhanced radiosensitivity through the inhibition of radiation-induced nuclear factor-kappaB activity in human oral squamous cell carcinoma cells. *Int. J. Oncol.* **2007**, *31*, 761–768.
- 35. Paradowska-Gorycka, A.; Grzybowska-Kowalczyk, A.; Wojtecka-Lukasik, E.; Maslinski, S. IL-23 in the Pathogenesis of Rheumatoid Arthritis. *Scand. J. Immunol.* **2010**, *71*, 134–145. [\[CrossRef\]](http://doi.org/10.1111/j.1365-3083.2009.02361.x)
- 36. Zhou, Y.; Hopper-Borge, E.; Shen, T.; Huang, X.-C.; Shi, Z.; Kuang, Y.-H.; Furukawa, T.; Akiyama, S.-I.; Peng, X.-X.; Ashby, C.R.; et al. Cepharanthine is a potent reversal agent for MRP7(ABCC10)-mediated multidrug resistance. *Biochem. Pharmacol.* **2009**, *77*, 993–1001. [\[CrossRef\]](http://doi.org/10.1016/j.bcp.2008.12.005)
- 37. Kathawala, R.J.; Wang, Y.-J.; Ashby, C.R., Jr.; Chen, Z.-S. Recent advances regarding the role of ABC subfamily C member 10 (ABCC10) in the efflux of antitumor drugs. *Chin. J. Cancer* **2014**, *33*, 223–230. [\[CrossRef\]](http://doi.org/10.5732/cjc.013.10122)
- 38. Huang, C.Z.; Wang, Y.F.; Zhang, Y.; Peng, Y.M.; Liu, Y.X.; Ma, F.; Jiang, J.H.; Wang, Q.D. Cepharanthine hydrochloride reverses P-glycoprotein-mediated multidrug resistance in human ovarian carcinoma A2780/Taxol cells by inhibiting the PI3K/Akt signaling pathway. *Oncol. Rep.* **2017**, *38*, 2558–2564. [\[CrossRef\]](http://doi.org/10.3892/or.2017.5879)
- 39. Kadioglu, O.; Law, B.Y.K.; Mok, S.W.F.; Xu, S.-W.; Efferth, T.; Wong, V.K.W. Mode of Action Analyses of Neferine, a Bisbenzylisoquinoline Alkaloid of Lotus (*Nelumbo nucifera*) against Multidrug-Resistant Tumor Cells. *Front. Pharmacol.* **2017**, *8*, 238. [\[CrossRef\]](http://doi.org/10.3389/fphar.2017.00238)
- 40. Okada, K.; Sakusabe, N.; Kobayashi, A.; Hoshi, N.; Sato, K. Prevention of Lung Metastasis by Intra-tumoral Injection of Cepharanthin and Staphylococcal Enterotoxin B in Transplantable Rat Osteosarcoma. *Jpn. J. Cancer Res.* **1999**, *90*, 928–934. [\[CrossRef\]](http://doi.org/10.1111/j.1349-7006.1999.tb00837.x)
- 41. Ershun, Z.; Yunhe, F.; Zhengkai, W.; Yongguo, C.; Naisheng, Z.; Zhengtao, Y. Cepharanthine attenuates lipopolysaccharideinduced mice mastitis by suppressing the NF-kappaB signaling pathway. *Inflammation* **2014**, *37*, 331–337. [\[CrossRef\]](http://doi.org/10.1007/s10753-013-9744-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24062060)
- 42. Morikawa, K.; Oseko, F.; Morikawa, S. Inhibition of proliferation and differentiation of human B-lymphocytes by a biscoclaurine alkaloid. *Int. J. Immunopharmacol.* **1992**, *14*, 941–949. [\[CrossRef\]](http://doi.org/10.1016/0192-0561(92)90094-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/1387392)
- 43. Tabata, R.; Tabata, C.; Tazoh, A.; Nagai, T. Low dose cepharanthine ameliorates immune thrombocytopenic purpura associated with multiple myeloma. *Int. Immunopharmacol.* **2012**, *13*, 242–244. [\[CrossRef\]](http://doi.org/10.1016/j.intimp.2012.04.015) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22561120)
- 44. Liu, Z.-L.; Hirano, T.; Tanaka, S.; Onda, K.; Oka, K. Persistent reversal of P-glycoprotein-mediated daunorubicin resistance by tetrandrine in multidrug-resistant human T lymphoblastoid leukemia MOLT-4 cells. *J. Pharm. Pharmacol.* **2003**, *55*, 1531–1537. [\[CrossRef\]](http://doi.org/10.1211/0022357022115) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/14713364)
- 45. Xu, W.; Wang, X.; Tu, Y.; Masaki, H.; Tanaka, S.; Onda, K.; Sugiyama, K.; Yamada, H.; Hirano, T. Tetrandrine and cepharanthine induce apoptosis through caspase cascade regulation, cell cycle arrest, MAPK activation and PI3K/Akt/mTOR signal modification in glucocorticoid resistant human leukemia Jurkat T cells. *Chem. Interact.* **2019**, *310*, 108726. [\[CrossRef\]](http://doi.org/10.1016/j.cbi.2019.108726) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31255635)
- 46. Xu, W.; Chen, S.; Wang, X.; Tanaka, S.; Onda, K.; Sugiyama, K.; Yamada, H.; Hirano, T. Molecular mechanisms and therapeutic implications of tetrandrine and cepharanthine in T cell acute lymphoblastic leukemia and autoimmune diseases. *Pharmacol. Ther.* **2020**, *217*, 107659. [\[CrossRef\]](http://doi.org/10.1016/j.pharmthera.2020.107659)
- 47. Huang, H.; Hu, G.; Wang, C.; Xu, H.; Chen, X.; Qian, A. Cepharanthine, an Alkaloid from Stephania cepharantha Hayata, Inhibits the Inflammatory Response in the RAW264.7 Cell and Mouse Models. *Inflammation* **2014**, *37*, 235–246. [\[CrossRef\]](http://doi.org/10.1007/s10753-013-9734-8)
- 48. Samra, Y.A.; Said, H.S.; Elsherbiny, N.M.; Liou, G.I.; El-Shishtawy, M.M.; Eissa, L.A. Cepharanthine and Piperine ameliorate diabetic nephropathy in rats: Role of NF-kappaB and NLRP3 inflammasome. *Life Sci.* **2016**, *157*, 187–199. [\[CrossRef\]](http://doi.org/10.1016/j.lfs.2016.06.002)
- 49. Golan-Goldhirsh, A.; Gopas, J. Plant derived inhibitors of NF-κB. *Phytochem. Rev.* **2013**, *13*, 107–121. [\[CrossRef\]](http://doi.org/10.1007/s11101-013-9293-5)
- 50. Gerschenson, L.E.; Rotello, R.J. Apoptosis: A different type of cell death. *FASEB J.* **1992**, *6*, 2450–2455. [\[CrossRef\]](http://doi.org/10.1096/fasebj.6.7.1563596)
- 51. Degterev, A.; Yuan, J. Expansion and evolution of cell death programmes. *Nat. Rev. Mol. Cell Biol.* **2008**, *9*, 378–390. [\[CrossRef\]](http://doi.org/10.1038/nrm2393) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18414491)
- 52. Kurokawa, M.; Kornbluth, S. Caspases and Kinases in a Death Grip. *Cell* **2009**, *138*, 838–854. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2009.08.021) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19737514)
- 53. Uto, T.; Toyama, M.; Yoshinaga, K.; Baba, M. Cepharanthine induces apoptosis through the mitochondria/caspase pathway in murine dendritic cells. *Immunopharmacol. Immunotoxicol.* **2016**, *38*, 238–243. [\[CrossRef\]](http://doi.org/10.3109/08923973.2016.1173059) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27121492)
- 54. Vermeulen, K.; Van Bockstaele, D.R.; Berneman, Z.N. The cell cycle: A review of regulation, deregulation and therapeutic targets in cancer. *Cell Prolif.* **2003**, *36*, 131–149. [\[CrossRef\]](http://doi.org/10.1046/j.1365-2184.2003.00266.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12814430)
- 55. Zhou, P.; Zhang, R.; Wang, Y.; Xu, D.; Zhang, L.; Qin, J.; Su, G.; Feng, Y.; Chen, H.; You, S.; et al. Correction: Cepharanthine hydrochloride reverses the mdr1 (P-glycoprotein)-mediated esophageal squamous cell carcinoma cell cisplatin resistance through JNK and p53 signals. *Oncotarget* **2021**, *12*, 61–62. [\[CrossRef\]](http://doi.org/10.18632/oncotarget.27638) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33456714)
- 56. Chen, L.; Flies, D.B. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat. Rev. Immunol.* **2013**, *13*, 227–242. [\[CrossRef\]](http://doi.org/10.1038/nri3405)
- 57. Xu, W.; Meng, K.; Tu, Y.; Tanaka, S.; Onda, K.; Sugiyama, K.; Hirano, T.; Yamada, H. Tetrandrine potentiates the glucocorticoid pharmacodynamics via inhibiting P-glycoprotein and mitogen-activated protein kinase in mitogen-activated human peripheral blood mononuclear cells. *Eur. J. Pharmacol.* **2017**, *807*, 102–108. [\[CrossRef\]](http://doi.org/10.1016/j.ejphar.2017.04.007)
- 58. Durinck, K.; Goossens, S.; Peirs, S.; Wallaert, A.; Van Loocke, W.; Matthijssens, F.; Pieters, T.; Milani, G.; Lammens, T.; Rondou, P.; et al. Novel biological insights in T-cell acute lymphoblastic leukemia. *Exp. Hematol.* **2015**, *43*, 625–639. [\[CrossRef\]](http://doi.org/10.1016/j.exphem.2015.05.017)
- 59. Tewari, D.; Patni, P.; Bishayee, A.; Sah, A.N.; Bishayee, A. Natural products targeting the PI3K-Akt-mTOR signaling pathway in cancer: A novel therapeutic strategy. *Semin. Cancer Biol.* **2022**, *80*, 1–17. [\[CrossRef\]](http://doi.org/10.1016/j.semcancer.2019.12.008)
- 60. Polivka, J., Jr.; Janku, F. Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. *Pharmacol. Ther.* **2014**, *142*, 164–175. [\[CrossRef\]](http://doi.org/10.1016/j.pharmthera.2013.12.004)
- 61. Takahashi-Yanaga, F.; Sasaguri, T. GSK-3beta regulates cyclin D1 expression: A new target for chemotherapy. *Cell Signal* **2008**, *20*, 581–589. [\[CrossRef\]](http://doi.org/10.1016/j.cellsig.2007.10.018) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18023328)
- 62. Gao, S.; Li, X.; Ding, X.; Qi, W.; Yang, Q. Cepharanthine Induces Autophagy, Apoptosis and Cell Cycle Arrest in Breast Cancer Cells. *Cell. Physiol. Biochem.* **2017**, *41*, 1633–1648. [\[CrossRef\]](http://doi.org/10.1159/000471234) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28359054)
- 63. Faes, S.; Dormond, O. PI3K and AKT: Unfaithful Partners in Cancer. *Int. J. Mol. Sci.* **2015**, *16*, 21138. [\[CrossRef\]](http://doi.org/10.3390/ijms160921138) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26404259)
- 64. Mahajan, K.; Mahajan, N.P. PI3K-independent AKT activation in cancers: A treasure trove for novel therapeutics. *J. Cell. Physiol.* **2012**, *227*, 3178–3184. [\[CrossRef\]](http://doi.org/10.1002/jcp.24065) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22307544)
- 65. Levatić, J.; Ćurak, J.; Kralj, M.; Šmuc, T.; Osmak, M.; Supek, F. Accurate Models for P-gp Drug Recognition Induced from a Cancer Cell Line Cytotoxicity Screen. *J. Med. Chem.* **2013**, *56*, 5691–5708. [\[CrossRef\]](http://doi.org/10.1021/jm400328s)
- 66. Genovese, I.; Ilari, A.; Assaraf, Y.G.; Fazi, F.; Colotti, G. Not only P-glycoprotein: Amplification of the ABCB1-containing chromosome region 7q21 confers multidrug resistance upon cancer cells by coordinated overexpression of an assortment of resistance-related proteins. *Drug Resist. Updat.* **2017**, *32*, 23–46. [\[CrossRef\]](http://doi.org/10.1016/j.drup.2017.10.003)
- 67. Li, H.; Yan, Z.; Ning, W.; Xiao-Juan, G.; Cai-Hong, Z.; Jin-Hua, J.; Fang, M.; Qing-Duan, W. Using Rhodamine 123 Accumulation in CD8+ Cells as a Surrogate Indicator to Study the P-Glycoprotein Modulating Effect of Cepharanthine Hydrochloride In Vivo. *J. Biomed. Biotechnol.* **2011**, *2011*, 281651. [\[CrossRef\]](http://doi.org/10.1155/2011/281651)
- 68. Xu, W.; Wang, X.; Chen, S.; Wu, H.; Tanaka, S.; Onda, K.; Sugiyama, K.; Yamada, H.; Hirano, T. Tetrandrine enhances glucocorticoid receptor translocation possibly via inhibition of P-glycoprotein in daunorubicin-resistant human T lymphoblastoid leukemia cells. *Eur. J. Pharmacol.* **2020**, *881*, 173232. [\[CrossRef\]](http://doi.org/10.1016/j.ejphar.2020.173232)
- 69. Vanbillemont, B.; De Beer, T. Application of polyvinyl acetate in an innovative formulation strategy for lyophilized orally disintegrating tablets. *Int. J. Pharm.* **2020**, *588*, 119717. [\[CrossRef\]](http://doi.org/10.1016/j.ijpharm.2020.119717)
- 70. Cantor, S.L.; Khan, M.A.; Gupta, A. Development and optimization of taste-masked orally disintegrating tablets (ODTs) of clindamycin hydrochloride. *Drug Dev. Ind. Pharm.* **2015**, *41*, 1156–1164. [\[CrossRef\]](http://doi.org/10.3109/03639045.2014.935392)
- 71. Yıldız, S.; Aytekin, E.; Yavuz, B.; Pehlivan, S.B.; Vural, I.; Ünlü, N. Development and evaluation of orally disintegrating tablets comprising taste-masked mirtazapine granules. *Pharm. Dev. Technol.* **2018**, *23*, 488–495. [\[CrossRef\]](http://doi.org/10.1080/10837450.2017.1315670)
- 72. Comoglu, T.; Ozyilmaz, E.D. Orally disintegrating tablets and orally disintegrating mini tablets-novel dosage forms for pediatric use. *Pharm. Dev. Technol.* **2019**, *24*, 902–914. [\[CrossRef\]](http://doi.org/10.1080/10837450.2019.1615090) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31215850)
- 73. Mahmoud, A.A.; Salah, S. Fast relief from migraine attacks using fast-disintegrating sublingual zolmitriptan tablets. *Drug Dev. Ind. Pharm.* **2012**, *38*, 762–769. [\[CrossRef\]](http://doi.org/10.3109/03639045.2011.625949)
- 74. Song, H.; Moon, C.; Lee, B.-J.; Oh, E. Mesoporous Pravastatin Solid Dispersion Granules Incorporable into Orally Disintegrating Tablets. *J. Pharm. Sci.* **2018**, *107*, 1886–1895. [\[CrossRef\]](http://doi.org/10.1016/j.xphs.2018.03.003) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29530714)
- 75. Shanmugam, S. Granulation techniques and technologies: Recent progresses. *BioImpacts* **2015**, *5*, 55–63. [\[CrossRef\]](http://doi.org/10.15171/bi.2015.04) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25901297)
- 76. Gryczke, A.; Schminke, S.; Maniruzzaman, M.; Beck, J.; Douroumis, D. Development and evaluation of orally disintegrating tablets (ODTs) containing Ibuprofen granules prepared by hot melt extrusion. *Colloids Surfaces B Biointerfaces* **2011**, *86*, 275–284. [\[CrossRef\]](http://doi.org/10.1016/j.colsurfb.2011.04.007) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21592751)
- 77. Badgujar, B.; Mundada, A. The technologies used for developing orally disintegrating tablets: A review. *Acta Pharm.* **2011**, *61*, 117–139. [\[CrossRef\]](http://doi.org/10.2478/v10007-011-0020-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21684842)
- 78. Kande, K.V.; Kotak, D.J.; Degani, M.S.; Kirsanov, D.; Legin, A.; Devarajan, P.V. Microwave-Assisted Development of Orally Disintegrating Tablets by Direct Compression. *AAPS PharmSciTech* **2017**, *18*, 2055–2066. [\[CrossRef\]](http://doi.org/10.1208/s12249-016-0683-z)
- 79. Hannan, P.; Khan, J.; Safiullah, S. Oral dispersible system: A new approach in drug delivery system. *Indian J. Pharm. Sci.* **2016**, *78*, 2–7. [\[CrossRef\]](http://doi.org/10.4103/0250-474X.180244)
- 80. Khaled, S.A.; Alexander, M.R.; Wildman, R.D.; Wallace, M.J.; Sharpe, S.; Yoo, J.; Roberts, C.J. 3D extrusion printing of high drug loading immediate release paracetamol tablets. *Int. J. Pharm.* **2018**, *538*, 223–230. [\[CrossRef\]](http://doi.org/10.1016/j.ijpharm.2018.01.024)
- 81. Li, G.F. Research progress on dosage form and clinical application of drop pills. *West China Pharm. J.* **2020**, *35*, 579–583.
- 82. Hu, L.; Shi, Y.; Li, J.H.; Gao, N.; Ji, J.; Niu, F.; Chen, Q.; Yang, X.; Wang, S. Enhancement of Oral Bioavailability of Curcumin by a Novel Solid Dispersion System. *AAPS PharmSciTech* **2015**, *16*, 1327–1334. [\[CrossRef\]](http://doi.org/10.1208/s12249-014-0254-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25804949)
- 83. Vo, C.L.-N.; Park, C.; Lee, B.-J. Current trends and future perspectives of solid dispersions containing poorly water-soluble drugs. *Eur. J. Pharm. Biopharm.* **2013**, *85*, 799–813. [\[CrossRef\]](http://doi.org/10.1016/j.ejpb.2013.09.007) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24056053)
- 84. Wang, L.; Zhao, J. Comparison of the dissolution rate of cepharanthine tablets and Qianjintiansu dropping pills. *J. Jinggangshan Univ.* **2009**, *30*, 83–84, 88.
- 85. Goldberg, M.; Gomez-Orellana, I. Challenges for the oral delivery of macromolecules. *Nat. Rev. Drug Discov.* **2003**, *2*, 289–295. [\[CrossRef\]](http://doi.org/10.1038/nrd1067)
- 86. Chaudhary, K.; Patel, M.M.; Mehta, P.J. Long-Acting Injectables: Current Perspectives and Future Promise. *Crit. Rev. Ther. Drug Carr. Syst.* **2019**, *36*, 137–181. [\[CrossRef\]](http://doi.org/10.1615/CritRevTherDrugCarrierSyst.2018025649)
- 87. Ma, Q.; Xie, Y.; Wang, Z.; Lei, B.; Chen, R.; Liu, B.; Jiang, H.; Wang, Y.; Liu, Q.; Yang, Z. Efficacy and safety of ReDuNing injection as a treatment for COVID-19 and its inhibitory effect against SARS-CoV-2. *J. Ethnopharmacol.* **2021**, *279*, 114367. [\[CrossRef\]](http://doi.org/10.1016/j.jep.2021.114367)
- 88. Zhang, X.Y.; Lv, L.; Zhou, Y.L.; Xie, L.D.; Xu, Q.; Zou, X.F.; Ding, Y.; Tian, J.; Fan, J.L.; Fan, H.W.; et al. Efficacy and safety of Xiyanping injection in the treatment of COVID-19: A multicenter, prospective, open-label and randomized controlled trial. *Phytother. Res.* **2021**, *35*, 4401–4410. [\[CrossRef\]](http://doi.org/10.1002/ptr.7141)
- 89. Xing, Y.; Hua, Y.-R.; Shang, J.; Ge, W.-H.; Liao, J. Traditional Chinese medicine network pharmacology study on exploring the mechanism of Xuebijing Injection in the treatment of coronavirus disease 2019. *Chin. J. Nat. Med.* **2020**, *18*, 941–951. [\[CrossRef\]](http://doi.org/10.1016/S1875-5364(20)60038-3)
- 90. Kim, Y.-K.; Xing, L.; Chen, B.-A.; Xu, F.; Jiang, H.-L.; Zhang, C. Aerosol delivery of programmed cell death protein 4 using polysorbitol-based gene delivery system for lung cancer therapy. *J. Drug Target.* **2014**, *22*, 829–838. [\[CrossRef\]](http://doi.org/10.3109/1061186X.2014.932796)
- 91. Kaur, S.S. Pulmonary drug delivery system: Newer patents. *Pharm. Pat. Anal.* **2017**, *6*, 225–244. [\[CrossRef\]](http://doi.org/10.4155/ppa-2017-0019) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28818001)
- 92. Muralidharan, P.; Malapit, M.; Mallory, E.; Hayes, D.; Mansour, H.M. Inhalable nanoparticulate powders for respiratory delivery. *Nanomed. Nanotechnol. Biol. Med.* **2015**, *11*, 1189–1199. [\[CrossRef\]](http://doi.org/10.1016/j.nano.2015.01.007) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25659645)
- 93. Bharti, N.; Kumar, S.L.H.; Budhiraja, A. Pulmonary drug delivery as a vital route for delivering nanoparticles e a review. *World J. Pharm. Pharm. Sci.* **2012**, *2*, 4037–4060. [\[CrossRef\]](http://doi.org/10.4103/0970-2113.92361)
- 94. Buttini, F.; Colombo, P.; Rossi, A.; Sonvico, F.; Colombo, G. Particles and powders: Tools of innovation for non-invasive drug administration. *J. Control. Release* **2012**, *161*, 693–702. [\[CrossRef\]](http://doi.org/10.1016/j.jconrel.2012.02.028)
- 95. Nisini, R.; Poerio, N.; Mariotti, S.; De Santis, F.; Fraziano, M. The Multirole of Liposomes in Therapy and Prevention of Infectious Diseases. *Front. Immunol.* **2018**, *9*, 155. [\[CrossRef\]](http://doi.org/10.3389/fimmu.2018.00155)
- 96. Guimarães, D.; Cavaco-Paulo, A.; Nogueira, E. Design of liposomes as drug delivery system for therapeutic applications. *Int. J. Pharm.* **2021**, *601*, 120571. [\[CrossRef\]](http://doi.org/10.1016/j.ijpharm.2021.120571)
- 97. D'Mello, S.R.; Cruz, C.N.; Chen, M.-L.; Kapoor, M.; Lee, S.L.; Tyner, K.M. The evolving landscape of drug products containing nanomaterials in the United States. *Nat. Nanotechnol.* **2017**, *12*, 523–529. [\[CrossRef\]](http://doi.org/10.1038/nnano.2017.67)
- 98. Hoshyar, N.; Gray, S.; Han, H.; Bao, G. The effect of nanoparticle size on in vivo pharmacokinetics and cellular interaction. *Nanomedicine* **2016**, *11*, 673–692. [\[CrossRef\]](http://doi.org/10.2217/nnm.16.5)
- 99. Pandey, R.; Khuller, G.K. Antitubercular inhaled therapy: Opportunities, progress and challenges. *J. Antimicrob. Chemother.* **2005**, *55*, 430–435. [\[CrossRef\]](http://doi.org/10.1093/jac/dki027)
- 100. Mitchell, M.J.; Billingsley, M.M.; Haley, R.M.; Wechsler, M.E.; Peppas, N.A.; Langer, R. Engineering precision nanoparticles for drug delivery. *Nat. Rev. Drug Discov.* **2021**, *20*, 101–124. [\[CrossRef\]](http://doi.org/10.1038/s41573-020-0090-8)
- 101. Al-Hallak, M.K.; Sarfraz, M.K.; Azarmi, S.; Roa, W.H.; Finlay, W.H.; Löbenberg, R. Pulmonary delivery of inhalable nanoparticles: Dry powder inhalers. *Ther. Deliv.* **2011**, *2*, 1313–1324. [\[CrossRef\]](http://doi.org/10.4155/tde.11.100) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22826885)
- 102. Liu, Y.; Hardie, J.; Zhang, X.; Rotello, V.M. Effects of engineered nanoparticles on the innate immune system. *Semin. Immunol.* **2017**, *34*, 25–32. [\[CrossRef\]](http://doi.org/10.1016/j.smim.2017.09.011) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28985993)
- 103. de Braganca, L.; Ferguson, G.J.; Santos, J.L.; Derrick, J.P. Adverse immunological responses against non-viral nanoparticle (NP) delivery systems in the lung. *J. Immunotoxicol.* **2021**, *18*, 61–73. [\[CrossRef\]](http://doi.org/10.1080/1547691X.2021.1902432) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33956565)
- 104. Thorp, E.B.; Boada, C.; Jarbath, C.; Luo, X. Nanoparticle Platforms for Antigen-Specific Immune Tolerance. *Front. Immunol.* **2020**, *11*, 945. [\[CrossRef\]](http://doi.org/10.3389/fimmu.2020.00945)
- 105. Liu, C.; Zhang, W.; Li, Y.; Chang, J.; Tian, F.; Zhao, F.; Ma, Y.; Sun, J. Microfluidic Sonication to Assemble Exosome Membrane-Coated Nanoparticles for Immune Evasion-Mediated Targeting. *Nano Lett.* **2019**, *19*, 7836–7844. [\[CrossRef\]](http://doi.org/10.1021/acs.nanolett.9b02841) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31597431)
- 106. Lu, C.; Zheng, J.; Ding, Y.; Meng, Y.; Tan, F.; Gong, W.; Chu, X.; Kong, X.; Gao, C. Cepharanthine loaded nanoparticles coated with macrophage membranes for lung inflammation therapy. *Drug Deliv.* **2021**, *28*, 2582–2593. [\[CrossRef\]](http://doi.org/10.1080/10717544.2021.2009936) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34866533)