REVIEW

Effects of writers, erasers and readers within miRNA-related m6A modification in cancers

Huiru Feng | Xiaofei Yuan | Shuting Wu | Yue Yuan | Linchong Cui | Danfan Lin | Xiaohong Peng | Xiong Liu | Fan Wang

Department of Otolaryngology-Head and Neck Surgery, Nanfang Hospital, Southern Medical University, Baiyun District, Guangzhou, People's Republic of China

Correspondence

Xiong Liu and Fan Wang, Department of Otolaryngology-Head and Neck Surgery, Nanfang Hospital, Southern Medical University, Jingxi Street, Baiyun District, Guangzhou 510515, People's Republic of China.

Email: liux1218@126.com and wf493649053@qq.com

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Abstract

Background: As one of the most abundant post-transcriptional mRNA modifications, N6‐methyladenosine (m6A) has attracted extensive attention from scientists. Emerging evidence indicates that m6A modification plays a significant role in cancer‐related signalling pathways. Existing research demonstrates that m6A modifications were also identified in miRNAs and contribute to cancer‐related signalling pathways.

Methods: A literature retrieval has been performed to collect m6A-miRNA-related original articles published in recent years. Later, a systematic analysis has been conducted to abstract and classify the relationships between m6A modification and miR-NAs, and their contributions to tumorigenesis and cancer development.

Results: Accumulating literature provides important insights into multiple relationships between m6A modifications and miRNAs. Mechanically, m6A writer and eraser alter pri-miRNAs m6A levels, and m6A readers could dually modulate pri-miRNAs processing and pri‐miRNAs degradation. It is also been demonstrated that miRNAs impair m6A regulators' translation to influence m6A medication function in return. Aberrant expressions of m6A regulators and miRNAs could dysregulate proliferative, apoptosis, cell adhesion‐related, and malignant transformation signalling pathways, and contribute to tumour occurrence and development.

Conclusion: This review summarizes the interrelationship between m6A modification and miRNAs; highlights the combined effects of each type of m6A regulator and miR-NAs in cancers. These findings enhance our understanding of m6A‐miRNAs' multiple interactions and significant modulatory role in tumorigenesis and progression.

1 | INTRODUCTION

N6-methyladenosine (m6A) modification is the most prevalent posttranscriptional modification in mammal's mRNAs. It refers to that methylation occurs in the N6-position of adenosine.^{[1,2](#page-13-0)} As the development of high throughput sequencing technology accelerates, m6A is found to be installed at adenosine within RRACH (R corresponds to G or A; H corresponds to A, C or U) motif, which is commonly located at the $3'$ untranslated region (UTR), intron and stop codon of the mRNAs.^{[3,4](#page-13-0)} Mechanically, m6A modification is a reversible process, dynamically regulated by methyltransferases, demethyltransferases and RNA binding proteins, namely writers, erasers and readers,

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respectively.⁵ Writers and erasers are responsible for the installation and dismantlement of methyl, counterbalancing the levels of m6A modification (Figure 1A). As for readers, they could recognize the m6A modification and manipulate the directions of RNA metabolism, including translation, stabilization, degradation, alternative splicing, pri-miRNAs processing and so on (Figure 1B,C). Functionally, m6A modification directly mediates RNA fate to influence proteins, main executors in the life processes, and hence drives a series of subsequent effects on cell biological functions.^{6,7} Extensive studies have elucidated that m6A regulators contribute to variant cell signalling pathways, having profound impacts on disease, stem cell differentiation, cell proliferation, tumorigenesis, cancer metastasis, invasion and clinical prognosis. $8-10$ $8-10$

miRNAs, one of the non-coding RNAs, are a type of bioactive molecules consisting of approximately 22nts single-strand nucleotides. Functionally, miRNAs drive an interfering effect on mRNAs translation and induce mRNAs degradation by targeting mRNA 3′UTR via the complementary base pairing principle. $11-13$ Previous studies have well-validated that miRNAs reflect disturbing effects on cancerassociated mRNAs, exhibiting influence covering cancer stem cell differentiation, proliferation, metastasis, prognosis and therapeutic value in various cancers in the past decades. $14-17$ $14-17$

In addition to mRNAs, miRNAs are proven to be widespread modified with N6-methyl, which are commonly enriched in the consensus sequence of GGAC.^{[18](#page-13-0)} In 2015, Alarcon C. R and colleagues corroborated that primary miRNA (pri-miRNA) processing was associated with an m6A-

dependent manner, raising the curtain on the m6A regulatory mechanism of miRNAs processing and its effect in various cancers.¹⁸ On the other hand, numerous studies have found that miRNAs could alter m6A levels in turn by suppressing the expression of m6A regulators. Emerging investigations have shed light on the molecular basis and effects of m6A and miRNAs interactions. Nevertheless, detailed mechanisms regarding the roles of each type of m6A regulator that contribute to tumour pathological have not yet been completely realized. In this case, it is necessary to comprehensively understand the relationship between m6A and miRNAs from available reports, laying a systematic and theoretical foundation to analyse joint effects on cancer occurrence and development.

In this review, we aim to introduce the basic knowledge of m6A modification, sum up the interaction between m6A regulators and miRNAs, emphasize the contributions of each regulator to cancer physical functions when cooperating with miRNAs and expect to inform future clinical applications.

2 | BIOLOGICAL FUNCTIONS OF M6A REGULATORS

M6A modification is modulated by three types of m6A regulators consisting of methyltransferases, demethyltransferases and RNA-binding proteins three types of proteins, namely m6A writers, erasers and readers (Figure 1A, Table [1\)](#page-2-0).

FIGURE 1 The dynamic process of m6A modification and diverse functions of m6A readers. (A) Methyl was installed at N6 of adenosine by m6A methyltransferase complex which consists of METTL3, METTL14, WTAP, RBM15, VIRMA and ZC3H13. In addition, METTL16 and NSun2 could function individually. M6A modification could be removed by demethyltransferases FTO and ALKBH5. m6A writers and erasers dynamically regulate RNA m6A levels, followed by recognition of m6A binding proteins. Different readers manipulate diverse RNA metabolism processes. (B) In the nucleus, readers could mediate mRNA alternative splicing, maintain mRNA stability and primary miRNAs processing. (C) In the cytoplasm, readers could promote mRNAs translation and the exact opposite function, mRNAs degradation

2.1 | M6A writers

M6A modification is installed by methyltransferase multicomponent involving a catalytic subunit methyltransferase-like enzyme 3 (METTL3), 19 a stabilize subunit methyltransferase-like enzyme 14 (METTL14),^{[20](#page-13-0)} Wilm's tumour-associated protein (WTAP), vir-like m6A methyltransferase-associated (VIRMA, also known as KIAA1429), RNA-binding motif protein 15 (RBM15) and its homologue (RMB15B), zinc finger CCCH-type containing 13 (ZC3H13) and so on (Figure [1A\)](#page-1-0). Somehow, some methyltransferases function by themselves, for instance, METTL16 and NSun2. METTL3, METTL14 and METTL16 all belong to the methyltransferase-like family. METTL14 forms tight conjunction with catalytic METTL3 as a stable heterodimer, providing a platform for METTL3's catalysis. The rest of the components depend on the METTL3-METTL14 core complex, devoting to the activity and localization of the writer complex. WTAP interacts with the METTL3-METTL14 complex and guides it to nuclear speckle.^{[21,22](#page-13-0)} VIRMA recruits METTL3-METTL14-WTAP to 3['] UTR and regions near the stop codon for m6A location-specific installation. 23 23 23 RMB15 and its paralog RMB15B are revealed to connect with the METTL3-WTAP complex, which is necessary for m6Adependent X-inactive-specific transcript (XIST) silencing to mediate X chromosome inactivation. $24,25$ ZC3H13 is required for adenosine

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methylation by bridging RBM15 to the WTAP-VIRMA complex. 26 METTL16 is a novel m6A methyltransferase that modifies the U6 snRNA, other pre-mRNAs and various non-coding RNAs indepen-dently.^{[27](#page-13-0)} NSun2 is one of the tRNA methyltransferases, it is also found to exhibit RNA m6A methylation function in colon-rectal cancer.^{[28](#page-13-0)} Given all that has been mentioned so far, it is demonstrated that m6A methylation is manipulated by complicated machinery, which is orchestrated by various methyltransferases for adenosine methyl installation.

2.2 | M6A erasers

For now, demethyltransferases only account for a few parts of m6A regulators. FTO and ALKBH5, two acknowledged and well-researched demethyltransferases, discharge methyl from adenosine (Figure [1A\)](#page-1-0). FTO, fat mass and obesity-associated protein, was firstly reported to be associated with fatty acid biogenesis and the first demethyl-transferase.^{[29](#page-13-0)} Especially, FTO was mentioned to mediate small RNA demethylation, for example, miRNAs. In addition to m6A demethylation, FTO was shown to modulate m6Am and m1A demethylation. 30 FTO belongs to the AlkB family, thus ALKBH5 was identified by Zheng et al who decided to figure out whether other AlkB homologues are acting as demethyltransferase other than FTO. 31

TABLE 1 Functions of m6A

FIGURE 2 The patterns of interaction between m6A modification and miRNAs. (A) m6A modification mediates pri-miRNAs processing. m6A writer deposits methyl at the m6A motif of pri-miRNA followed by m6A reader recognizing it. Part of readers like HNRNP2B1, NKAP and YTHDC1 recruit microprocessor complex for flanking slicing, subsequently receiving a precursor miRNA, while YTHDF2 could accelerate pre-miRNA degradation. (B) miRNAs target 3's UTR of m6A regulators. miRNAs could induce m6A level alteration or impede m6A reader function implementation through targeting m6A regulators' 3' UTR. (C) miRNAs orchestrate with m6A regulators to exert their intrinsic functions. miRNAs could assist m6A methylation and demethylation by interacting with the writer and eraser respectively. miRNA targets mRNA 3'UTR in an m6Adependent way

Methyltransferase and demethyltransferase counterbalance the m6A modification level in a highly dynamic manner, potentially contributing to disease and cancers associated imbalance.

2.3 | M6A readers

M6A readers are a type of RNA-binding protein that can decipher the m6A mark and specifically manipulate RNA metabolism, dominating the downstream biological functions. In general, there are mainly two types of m6A readers, YT521-B homology (YTH) domain and heterogeneous nuclear ribonucleoproteins (HNRNPs). YTH domain consists of YTH domain family 1–3 (YTHDF1-3) and YTH domain containing 1–2 (YTHDC1-2) in humans.^{[32](#page-13-0)} In terms of YTHDF1-3, they could dominate transcripts into exactly opposite destinies. YTHDF1 accelerates translation or promotes stability and storage of transcripts. 33 On the contrary, YTHDF2 expedites mRNA decay and impairs its enrichment.^{[34](#page-14-0)} As for YTHDF3, it coordinates with YTHDF1 or YTHDF2 to exert their effects on corresponding m6A-containing mRNAs.³⁵ YTHDC1 assists mRNA precursor export to the cytoplasm for further maturation and translation.^{36,37} YTHDC2 exerts a critical effect in mammalian spermatogenesis by enhancing translation efficiency and decreasing mRNA abundance.^{[38](#page-14-0)} In contrast to the former finding, YTHDC2 is found to facilitate mRNA degradation and hence decrease translation efficiency.³⁹

Another group of HNRNPs, including HNRNPC, HNRNPG and HNRNPA2B1, is characterized by alternative splicing, transcript processing, local RNA structure remodelling and RNA-protein interaction modulation[.40](#page-14-0) Among these HNRNPs, HNRNPA2B1 is found to directly bind to m6A and mediate primary miRNA processing with miRNA microprocessor complex.[41](#page-14-0) Apart from these two main family m6A readers, insulin-like growth factor-2 mRNA-binding proteins (IGF2BPs) are found to be a new group of m6A readers. Compelling evidence revealed that IGF2BPs facilitate mRNAs translation and stability. 42 Some known molecules are also identified as m6A readers. Eukaryotic initiation factor 3 (eIF3) is elucidated to initiate translation through recruiting 43S ribosomal complex via binding m6A containing 5'UTR mRNAs in a cap-independent manner.^{[43](#page-14-0)} Nuclear factor κB -related protein (NKAP) could exhibit miRNAs biogenesis assistance effect in a similar pattern of HNRNPA2B1.⁴⁴ Beyond all that, m6A readers' functions are much more than available evidence, deeper investigation is recommended for general understanding.

3 | INTERACTION PATTERNS BETWEEN M6A MODIFICATION AND MIRNAS

From the available literature, we found multiple interactions between m6A modification and miRNAs. We develop the following interaction patterns for better understanding (Figure 2).

3.1 | M6A modification mediates pri-miRNAs processing (Figure [2A](#page-3-0))

miRNA maturation undergoes a series of stages. Primary miRNAs, transcribed from the genome, include 100s to 1000s of nucleotides and basically consist of at least one loop-stem-flanking region. Primary miRNAs require flanking slicing to form precursor miRNAs, which were manipulated by microprocessor complex DGCR8 and DROSHA. Next, precursor miRNAs that contain the stem structure of the mature part of miRNA/miRNA* duplex at the opposite side and the loop structure are exported outward cytoplasm by exportin5-RAN-GTP complex. In the cytoplasm, DICER removes the loop structure of precursor miRNAs, followed by AGO cleaving the leaving miRNA/miRNA* duplex into single-stranded mature miRNAs.^{[45,46](#page-14-0)} In 2015, Alarcon et al firstly inves-tigated the role of METTL3 toward primary miRNA processing.^{[18](#page-13-0)} It is confirmed that METTL3 enhanced pri-miRNAs m6A abundance and miRNAs expressions. Researchers proposed that m6A modification initiated the interaction between DGCR8 and m6Acontaining pri-miRNAs for slicing during the processing. On the contrary, demethyltransferase ALKBH5 has been reported to sup-press pri-miR-193a processing.^{[47](#page-14-0)} Recently, Zhang and colleagues revealed that overexpressed-METTL3 elevated the pri-miR-25 m6A level, and subsequently reader NKAP bound with m6A and recruited DGCR8 for further processing.^{[44](#page-14-0)} These findings illustrated a pri-miRNA processing pattern that is featured by the m6A modification and reader-dependent foundation for microprocessor complex recruitment, providing a more complete theory frame for understanding the effect of m6A modification on miRNAs synthesis modulation.

3.2 | miRNAs target 3's UTR of m6A regulators (Figure [2B](#page-3-0))

It is well-validated that miRNAs as vital bioactive molecules, aim at disturbing the translation of targeted mRNAs. In this way, miRNAs could base pair with N6-methyladenosine regulators' mRNAs followed by downstream m6A modification alteration. MiR-483p has been found to target METTL3 3'UTR to reduce the m6A level of p21, driving an antiproliferation effect in breast cancer.^{[48](#page-14-0)} Similarly, miR-193a-3p could repress ALKBH5 expression, thus elevating AKT2 m6A abundance and inhibiting apoptosis in glioma.^{[49](#page-14-0)} Though miRNA could not change m6A amounts when repressing readers' expression, it does influence readers' function implementation. It is uncovered that MALAT1-sponged-miR-204 failed to bind to reader IGF2BP2. Hence, elevated-IGF2BP2 could recognize m6Acontaining MYC and promote translation, exhibiting an oncogenic effect in thyroid cancer.^{[50](#page-14-0)} Therefore, the disturbing effect of miR-NAs could exhibit global impacts on cell m6A modification as well as downstream biological functions, providing an overall insight into the bidirectional relationship between m6A modification and miRNAs.

3.3 | miRNAs orchestrate with m6A regulators to exert their intrinsic functions (Figure [2C](#page-3-0))

Recently, miRNAs are somehow identified as possible readers of m6A motifs, assisting writers and erasers to better exert functions of methylation and demethylation respectively. miRNAs binding sites are discovered to be enriched at m6A motifs of mRNAs 3'UTRs, which is a fundamental basis that miRNAs mediate m6A regulators to be located at the m6A motifs. A report carried out by Chen et al suggested that miRNAs orchestrated METTL3's recognition of the m6A motif for methyl installation. Further examination confirmed that overexpression of miRNAs mediated METTL3 to bind with mRNAs m6A motifs for methylation. 51 And not coincidentally, John P. Zepecki and colleagues revealed that miR-145 elevated the demethylation activity of FTO by forming FTO/AGO1/ILF3/miR-145 complex. Further investigation demonstrated that miR-145 induced FTO's binding to mRNAs and reduced the m6A level of target mRNAs during glioma stem cells (GSCs) state transition to differential glioma cells (DGCs).⁵² For both reports of miRNAs assisting catalysis of METTL3 or FTO, overexpression or downregulation of miRNAs do not cause expression alteration of these m6A writers and erasers. It is miRNAs that modulate localization and binding of METTL3 and FTO to more perfectly perform their functions. Another novel study conducted by Junmei Cheng et al indicated that m6A modification in MYCN is indispensable for miR-98's binding to MYCN 3'UTR. Mutations of the m6A motif of MYCY impeded the interaction between miR-98 and MYCN. Hence, overexpressed pseudo reader miR-98 disturbed MYCN expression in a novel m6A-dependent way.⁵³ These findings supported the phenomenon that miRNAs could be viewed as a kind of feasible readers of m6A motifs, showing more possibility of regulative patterns between miRNAs and m6A modification.

4 | COMBINED ROLES OF M6A WRITERS AND MIRNAS IN CANCERS

4.1 | Methyltransferases regulate pri-miRNAs processing

After Alarcon's research concerning METTL3 and m6A set a research foundation and reference for m6A-dependent miRNAs processing, accumulating studies have been conducted and showed that writermediated miRNAs maturation exhibited generality of aggressive phenotypes in various cancers. Next, we will illustrate that m6A writers' functions via mediating miRNAs expression from two diverse biological aspects, cancer proliferation and metastasis (Table [2](#page-5-0)).

The limitless replicative potential is one of the most prominent features of tumours. In gallbladder cancer, it is elucidated that reduced-deoxycholic acid (DAC) failed to disrupt METTL3 assemble with methyltransferase complex, assisting METTL3-m6A-dependent pri-miR-92b processing. In this situation, miR-92b-3p silenced tumour suppressor PTEN and subsequently stimulated PI3K/AKT pathway, fostering cell growth and exerting an oncogenic effect in gallbladder

TABLE 2 M6A writers regulate miRNAs processing

Abbreviations: CRC, colorectal cancer; EMT, epithelial-mesenchymal transition; HCC, hepatocellular cancer; MeRIP-qPCR, methylated RNA immunoprecipitation quantitative polymerase chain reaction; NSCLC, non-small cell lung cancer; 1, promote pri-miRNA processing; 1, inhibit pri-miRNA processing.

cancer[.54](#page-14-0) In pancreatic duct adenocarcinoma (PDAC), cigarette smoke condensate (CSC)-induced METTL3 promoted proliferation, driving an oncogenic function. METTL3 accelerated m6A-containing pri-miR-25 processing by recruiting a microprocessor DGCR8 with recognition of m6A reader NKAP. Consequently, miR-25-3p inhibited PHLPP2 expression followed by an evocation of AKT-p70S6K signalling.^{[44](#page-14-0)} In ovarian cancer, elevated METTL3 was found to contribute to miR-126-5p maturation and exert a tumour-promoting effect. Excessive miR-126-5p stimulated PTEN/PI3K/Akt/mTOR pathway, promoting cancer cell proliferation as well as suppressing apoptosis.⁵⁵ In bladder cancer, upregulated METTL3 was involved in facilitating pri-miR-221/222 processing to encourage cell proliferation. Enhanced mature miR-221/222 directly bound to PTEN mRNA, exerting an oncogenic effect.^{[56](#page-14-0)} So far, these observations confirmed that the wellresearched molecule METTL3, seemed to have a close connection with PTEN/PI3K/AKT/mTOR pathway. METTL3 utilized different miRNAs as a bridge to drive cell proliferation and tumour-promoting

effect in an m6A-based pattern. Furthermore, it is indicated that METTL3 has a prevalent modification function toward diverse primiRNAs, suggesting the ubiquitous role of m6A modification in miRNA metabolism.

Apart from cell proliferation-related signalling, m6A writers could mediate cell cycle protein and proliferation-associated protein to facilitate cancer cell growth via utilizing m6A-related miRNAs synthesis. In cervical cancer, it is elucidated that METTL3 could promote pri-miR-193b m6A level and processing. However, METTL3 was reduced in cervical cancer, hence failed to elevate mature miR-193b and under-mine the silencing effect of miR-193b on cell cycle protein CCND1.^{[57](#page-14-0)} In contrast to the findings described above, METTL3 exerted an infrequent tumour-suppressing effect in this article by negatively regulating CCND1. Though only METTL3 exhibits catalysis, METTL14 was validated to be crucial for methylation. METTL14 was also confirmed to be reduced and associated with a tumour-suppressing effect in colorectal cancer (CRC). METTL14 was found to be responsible for

<code>TABLE 3 $\,$ miRNAs</code> target m6A writers 3 $^{\prime}$ UTR $\,$

Abbreviations: EMT, epithelial-mesenchymal transition; HCC, hepatocellular cancer; LUAC, lung adenocarcinoma; NSCLC, non-small cell lung cancer.

facilitating miR-375 processing. Since miR-375 targeted YAP1, a proliferation-associated protein, METTL14 eventually repressed can-cer cell proliferation in CRC.^{[58](#page-14-0)}

Tissue invasion and metastasis are other predominant features of aggressive cancer. It has been widely uncovered that m6A writermodulated miRNAs could regulate EMT (epithelial-to-mesenchymal transition), angiogenesis, metastasis-associated molecules, and signal transduction pathways, contributing to tumour distant metastasis and undesirable prognosis. In lung cancer, overexpressed-METTL3 was involved in vasohibin-1(VASH1)-induced brain metastasis by facilitating pri-miR-143 processing. Functionally, VASH1 mediated angiogenesis and tubulin depolymerization through regulating vascular endothelial growth factor-A (VEGFA) degradation and tubulin detyrosination. So METTL3/miR-143 aggravated hematogenous metastasis and migration to offer a proper opportunity for brain metastasis.^{[59](#page-14-0)} SPRED2 was elucidated to function as a tumour-suppress regulator and repress metastasis-relevant MAPK pathway in various cancers. In CRC, it is reported that METTL3 modulated miR-1246 maturation and promoted aggressive phenotype. miR-1246 suppressed SPRED2 expression, exerting metastasis-facilitating function via MAPK path-way.^{[60](#page-14-0)} In breast cancer, aberrant elevated METTL14 promoted

migration and invasion through EMT via reshaping the miRNAs profile. Lately, hsa-miR-146a-5p was identified as a downstream regulative objective of METTL14 and was enriched in the cell adhesion aspect through bioinformatics analysis.⁶¹

The evidence presented in this section depicted a general phenomenon that writers increase pri-miRNAs m6A levels followed by facilitating pri-miRNAs processing. Writers are capable of mediating tumorigenesis, cancer cell growth and distant metastasis by enhancing miRNA levels. However, in proteinase-activated receptor 2 (PAR2) correlated CRC, one unanticipated finding was that another writer NSun2 inhibited miR-125 processing while elevating the pri-miR-125 m6A level. Eventually, NSun2 intensified PAR2-correlated CRC invasion capacity via activating the miR-125/Gab2/PI3K/AKT axis, exert-ing an oncogenic effect.^{[28](#page-13-0)} Unlike the prevalent writer characteristic of facilitating miRNAs processing, this article is worth thinking over the radical roles of m6A writers in pri-miRNAs processing. It is required more evidence to support writers' mutual modulations of miRNAs maturation. These not only indicated the close connection between m6A writers and cancer aggressive phenotypes but also provided more comprehensive insights into the way that we view writers' effect on pri-miRNAs processing.

4.2 | miRNAs target methyltransferases

Plenty of attempts have been made to demonstrate that miRNAs participate in reversing m6A modification amount, which is theoretically able to reflect on downstream pathways and biological functions via directly binding with m6A regulators' mRNAs in various cancers (Table [3](#page-6-0)).

Current studies have uncovered that as reduced-miRNAs failed to target METTL3 in diverse cancers, augmented METTL3 elevated proliferative-correlative proteins and signalling pathways in a miRNAs-METTL3-related way. In NSCLC, miR-33a was validated to be decreased and attenuated aggressive tumour phenotype via disturbing METTL3 expression. Further exploration revealed that METTL3 stimulated the epithelial growth factor receptor (EGFR) pathway, a type of self-sufficiency growth signalling. Hence reduced miR-33a substantially restored METTL3 oncogenic effect through retriev-ing EGFR pathway.^{[62](#page-14-0)} Similarly, it is also revealed that METTL3 referred to the miR-338-5p candidate target in NSCLC. Dramatically decreased miR-338 rescued METTL3 expression. Differently for this time, METTL3 enhanced a proliferative-associated protein c-MYC expression, facilitating proliferation and migration function in lung cancer cells.⁶³ In hepatoblastoma, bioinformatic software discovered miR-186 as a regulatory molecule of METTL3. METTL3 is revealed to be particularly capable of triggering the multiplication-associated Wnt/β-catenin pathway. miR-186 and METTL3 jointly contributed to cell proliferation as well as distant migration in hepatoblastoma. 64 In breast cancer, metformin (a traditional diabetes drug) induced miR-483-3p expression and miR-483-3p was validated to target METTL3. Reduced-METTL3 downregulated m6A-p21level but promoted p21 expression, a cycle inhibition-associated molecule. Therefore, metformin/miR-483-3p alleviated METTL3's oncogenic effect, promoting anti-proliferation activity in breast cancer. 48 As the articles mentioned above, various miRNAs could be available to bind to METTL3 3'UTR. A possible explanation for this might be that multiple miRNAs share similar sequences with METTL3 3′UTR, building a mutual and complicated communication in the m6A-miRNAs research field.

Similar to cell interminate growth, resisting cell death is another distinct way leading to tumorigenesis and growth. The miRNAs-METTL3 regulatory mechanism also participates in cancer cell death control. Cell apoptosis is the most prominent death form, mainly consisting of endoplasmic reticulum (ER), mitochondrial and death receptor pathways. SEC62 is a negative key transport molecule within ER apoptosis pathway. In gastric cancer, decreased-miR-4429 weakened silencing effect of oncogenic METTL3. Elevated-METTL3 enhanced m6A-SEC62 followed by the recognition of translation-promoting reader IGF2BP1. Reduced miR-4429 accelerated SEC62 expression and suppressed ER apoptosis via the METTL3/m6A/IGF2BP1 axis.^{[65](#page-14-0)} And anti-apoptosis Bcl2 is a crucial initiator of mitochondrial apoptosis. In lung adenocarcinoma (LUAD), miR-600 could suppress METTL3 expression, and simultaneously METTL3 was capable of stimulating PI3K/AKT/Bcl2 pathway. Therefore, repressed-miR-600 successfully restored METTL3 level, eventually alleviating mitochondrial apoptosis and leading to LUAD progression.^{[66](#page-14-0)}

Compared with apoptosis, ferroptosis is a pathological, ferdependent and more intense necrosis death. Since apoptosis deficiency in many cancers, ferroptosis might be a potential tumour therapeutic target. It is illustrated that miR-4443 was enriched in exosomes of cisplatin-resistant NSCLC and was also found to target METTL3 3'UTR. Decreased-METTL3 repressed m6A-dependent ferroptosis suppressor protein 1(FSP1) expression. Hence, miR-4443 suppressed NSCLC cancer ferroptosis and impaired cisplatin efficacy in a METTL3-FSP1-silencing way. 67 Contrary to the former literature, it is the first time to be observed that METTL3 functioned as a tumour suppressor in a miRNA-targeted way in this study.

Though WTAP does not possess a catalytic effect as well, it is critical to preserve catalytic function. WTAP was also implied to drive a significant role in cancer progression. In HCC, miR-139 functioned as a negative regulator of WTAP, which was revealed to be associated with stimulation of the EMT pathway. Later, it is further confirmed that alleviated miR-139 restored WTAP expression, facilitating HCC EMT proceeding.^{[68](#page-14-0)} In accordance with the available combination of findings concerning METTL14 and WTAP, the significance of the integrality of the m6A methyltransferase complex is further supported.

From cancer cell proliferation, cell death, invasion and metastasis, writers and various miRNAs infiltrated into cancer diverse aspects, driving significant impacts on tumour characteristics. Notably, research about the roles of methyltransferase in cancers is like a dualedge sword, presenting both oncogenic and tumour-suppressing effects with different writer-miRNA combinations. Based on available limited data, though METTL3 has a preference to exert an oncogenic effect, it is worthwhile mentioning that METTL14 exhibits dual effects but tends to present a protective, tumour-suppressing role in cancers. However, until now scientists are incapable of providing a reasonable explanation. Understandably, they did not possess decisive effects since m6A modification and miRNAs both function as posttranscriptional modifications. Even so, scientists are still looking forward to discovering some regularity or inclination for future applications.

5 | COMBINED ROLES OF M6A ERASERS AND MIRNAS IN CANCERS

Demethyltransferases are responsible for removing N6-methyl from pri-miRNA, reversing the effect of methyltransferase. However, demethyltransferases do not always drive the opposite function against methyltransferase's common pri-miRNAs processingpromoting effect. So far, the discovery of demethyltransferase is at a standstill, consistent with the low production of relative research articles. Literature regarding FTO and AKLBH5 constitutes the whole available evidence but is virtually the tip of the iceberg (Table [4](#page-8-0)).

Though FTO has always been a high-profile topic for scientists, literature about FTO and miRNAs remains few. Since FTO's multiple functions, the interrelationship between FTO and miRNAs catalysis are not limited in an m6A-dependent manner. In HER-2 positive breast cancer, highly expressed FTO was observed to reduce miR-

TABLE 4 The interaction between m6A erasers and miRNAs

M ₆ A regulators	Cancer type	miRNA	Erasers' role	Mechanism	Validation approach/model	Function of erasers	Ref
FTO	Breast cancer	$miR-$ 181-3p	Oncogene	Processing	Not in m6A- dependent way	Promote metastasis- related adhesion	69
FTO	NSCLC	miR-607	Oncogene	miRNA target erasers 3'UTR	Dual luciferase report assay, miRNA mimic/ inhibitor (cell)	Promote tumorigenesis and invasion	70
ALKBH5	Osteosarcoma	$miR-$ 181b- 3p	Suppress	Processing↑	MeRIP-qPCR (cell)	Inhibit YAP expression	71
ALKBH5	Oesophageal cancer	miR- 193a- 3p	Oncogene	Processing↑ miRNA target erasers 3'UTR	MeRIP-qPCR, dual luciferase report assay (cell)	Promote cancer cell proliferation and metastasis	47
ALKBH5	Glioma	$miR-$ 193a- 3p	Suppress	miRNA target erasers 3'UTR	Dual luciferase report assay, miRNA mimic/ inhibitor (cell)	Stimulate AKT2/Bcl2/ survivin, inhibit intrinsic apoptosis	49
ALKBH5	Ovarian cancer	$miR-7$	Oncogene	miRNA target erasers 3'UTR	HuR-dependent (cell)	Stimulate EGFR-PI3K/ AKT/mTOR pathway	72

Abbreviations: MeRIP-qPCR, methylated RNA immunoprecipitation quantitative polymerase chain reaction; NSCLC, non-small cell lung cancer; processing \uparrow , promote pri-miRNAs processing; processing., inhibit pri-miRNAs processing.

181b-3p and drove migration and invasion effects through miR-181b-3p/ADP ribosylation factor-like 5B(ARL5B) axis. Nonetheless, it is not demonstrated that FTO regulated miR-181b-3p level in an m6Adependent way since no observation of FTO-mediated miR-181b-3p m6A alteration based on MeRIP-seq result.⁶⁹ It was also found that FTO mRNA was identified as a downstream target of miR-607 in LUAD. miR-607 was suppressed by Circ-0072309 through miRNA response elements (MREs), which restored FTO expression and con-sequently facilitated LUAD tumorigenesis and invasion.^{[70](#page-15-0)}

Research concerning another demethyltransferase ALKBH5 faced poor production as well. Available literature elucidated that ALKBH5 was involved in cancer cell growth and apoptosis in an m6Adependent way. In osteosarcoma, ALKBH5 was revealed to remove m6A from pre-miR-181b-3p. Therefore, YTHDF2, a degradationfacilitating m6A reader, failed to degrade pre-miR-181b-3p without m6A modifications. Eventually, this combination of m6A regulators upregulated miR-181–3p to repress YAP, promoting apoptosis and attenuating malignancy phenotype.^{[71](#page-15-0)} In an addition, if ignoring the YTHDF2's degradation impact, this article provided a novel sight of ALKBH5's promoting effect on miRNA maturation while decreasing m6A levels. On the other hand, it is implied that ALKBH5 cooperated with miR-193a-3p to aggravate oesophageal carcinoma (ESCC) progression. Notably, further evidence indicated that miR-193a-3p could silence ALKBH5, and simultaneously ALKBH5 impeded miR-193a-3p processing in an m6A-dependent way in turn. In another word, ALKBH5 and miR-193a created a positive feedback loop, verifying the existence of m6A-miRNAs mutual regulation and intensifying oncogenic effects.⁴⁷ The same combination but in glioma, while ALKBH5 was only screened as a target of miR-193a-3p. ALKBH5 positively

regulated AKT2, a key molecule that inhibits the intrinsic apoptosis signalling pathway. As miR-193a-3p alleviated, restored-ALKBH5 facilitated the AKT2 pathway to exert an apoptosis-promoting effect.^{[49](#page-14-0)} In epithelial ovarian carcinoma, it is elucidated that ALKBH5 suppressed miR-7 processing and drove a tumour-promoting effect. However, in this study, ALKBH5 reduced miR-7 expression in a HuRrelative way, instead of the m6A-dependent one. 72

Resembling methyltransferases, demethyltransferases also drive dual effects in diverse cancers, especially when they link to multiple readers and downstream pathways. Surprisingly, ALKBH5 exhibited both effects of promoting miRNAs processing and degradation. Researchers provide a reasonable explanation that ALKBH5 accelerated miRNAs processing when cooperating with decay-inducing reader YTHDF2. Hence, in terms of m6A-associated miRNA metabolism, it could not be ignored that m6A readers are the ones who dominate m6A functions beyond writers and erasers. There is thereby an urgent necessity to comprehensively consider m6A regulators' combined effects on miRNAs processing.

6 | COMBINED ROLES OF M6A READERS AND MIRNAS IN CANCERS

RNA-binding proteins possess multiple types and functions, taking charge of recognizing m6A modification and guiding corresponding metabolic processes. Similarly, the phenomenon of readers' dual functions toward mRNAs also exists when it comes to miRNAs, including processing-promoting and degradation-promoting. Even though methyltransferase and demethyltransferase are in charge of altering

TABLE 5 The interaction between m6A readers and miRNAs

Abbreviations: HCC, hepatocellular cancer; HITS-CLIP, high through sequence-crosslinking immunoprecipatation; NSCLC, non-small cell lung cancer; PDAC, pancreatic ductal adenocarcinoma; processing†, promote pri-miRNAs processing; processing, inhibit pri-miRNAs processing; RIP-qPCR, RNA immunoprecipitation quantitative polymerase chain reaction.

m6A levels, readers are the decisive factors that devote to real functional implementation of m6A modification. It is obvious that m6A readers' dual functions and writer/eraser m6A regulatory effects complicate the conditions of miRNAs biosynthesis and cancer-related physical behaviours (Table [5](#page-9-0)).

After reporting miRNAs processing in a METTL3 and m6A dependent way, Alarcon et al continued subsequent research of m6A reader HNRNPA2B1. Alarcon and colleagues firstly confirmed that HNRNPA2B1 recognized m6A of pri-miRNAs and recruited microprocessor complex to facilitate the processing of primary miRNAs.⁴¹ m6A was commonly identified in the GGAC motif which is usually located near a junction of flanking and stem region of primary miRNAs. This excellent site allows localization of microprocessor complex followed by flanking region slicing. Since HNRNPA2B1's general processingpromoting effect on miRNAs and miRNAs' universal tumoursuppressing silencing function, HNRNPA2B1 seemed to act as an oncogenic factor in various cancers. To date, it is uncovered that HNRPRA2B1 took part in cancer cell proliferation and drug resistance aspects. In NSCLC, HNRNPA2B1, cooperating with lncRNA LINC01234, promoted miR-106b-5p maturation and drove an aggressive effect. Cryptochrome 2(CRY2) was identified to be a tumour suppressor and upstream inhibitor of oncogenic c-MYC. Enhanced miR-106b-5p attacked CRY2 and hence stabilized c-MYC, accelerat-ing cell proliferation and resulting in poor clinical outcomes.^{[73](#page-15-0)} In addition, breast cancer tamoxifen-resistant LCC9 cells exhibited highly expressed HNRNPA2B1. To investigate whether HNRNPA2B1 induced endocrine resistance in breast cancer cells, researchers constructed HNRNPA2B1-overexpressed MCF-7 cells (formerly tamoxifen sensitive). It is confirmed that excessive HNRNPA2B1 eliminated the ability of endocrine blockers like tamoxifen, supporting its important role in endocrine-resistance breast cancer. Subsequently, global miRome analysis and qPCR had been performed to confirm the downregulation of selected miR-29a-3p, miR-29b-3p and miR-222, and upregulation of selected miR-1266-5p, miR-1268a and miR-671–3p in the HNRNPA2B1-overexpressed MCF-7 cell. 74 It is somewhat astonishing that HNRNPA2B1 was also capable to downregulate miR-NAs. It is possible to hypothesize that this condition is likely to occur when there were more intermediate links or different patterns involved.

Comparable to HNRNPA2B1, YTHDC1 and NKAP could decode m6A and exhibit pri-miRNAs processing effect as well. In consideration that NKAP has been illustrated in the former part, 44 here is no more detailed description. YTHDC1 was also suggested to expedite miR-30d processing in PDAC and was involved in cancerassociated metabolism reprogramming Warburg effect. Mechanically, miR-30d bound to transcriptional factor RUNX, which failed to evoke glucose transporter-encoding SLC2A and hexokinase (HK1) transcription. Therefore, YTHDC1 attenuated PADC aerobic glycolysis, repressing PDAC occurrence and progression through pri-miR-30d processing.^{[75](#page-15-0)}

As mentioned before, another known m6A reader YTHDF2 could destabilize m6A-containing RNAs, exerting a negative effect on RNA stability and subsequent procedure. As for miRNAs, YTHDF2 impeded

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their processing through degradation as well. According to the aforementioned research, YTHDF2 failed to decay pre-miR-181 when ALKBH5 removed m6A, while YTHDF2 degrade pre-miR-181 when it received m6A again.^{[71](#page-15-0)} In addition to that YTHDF2 could degrade miR-NAs, YTHDF2 was also confirmed to be the target of diverse miRNAs. In ovarian cancer and HCC, YTHDF2 was both screened as a miR-145 target object and functioned as an oncogenic factor. Since miR-145 was repressed, enhanced-YTHDF2 destabilized m6A-carrying RNAs, contributing to cancer cell proliferation, apoptosis inhibition and migration. $76,77$ In prostate cancer, YTHDF2 was also suppressed by miR-495 and exerted oncogenic influence as well. As reduced miR-495 failed to silence YTHDF2, elevated-YTHDF2 degraded MOB family kinase activator 3B (MOB3B), expediting cell proliferation and distant metastasis. These results consistently suggest that YTHDF2 seemed to tend to drive a tumour-promoting effect.⁷⁸

From existing literature, no evidence suggests that YTHDF1 and IGF2BP2 exhibited function regarding miRNA slicing processing. On the contrary, anti-tumour miR-376c targeted oncogenic YTHDF1 in NSCLC. As miR-376c silenced YTHDF1, it disrupted the Wnt/βcatenin pathway, inhibiting NSCLC cell aggressive phenotype and poor progression.⁷⁹ In glioma, YTHDF1 was identified as a miR-346 downstream target as well. As miR-346 decreased, accumulating YTHDF1 promoted tumour growth and indicated a poor prognosis.^{[80](#page-15-0)} Alternatively, IGF2BP2 was revealed to be targeted by miR-204 in thyroid cancer. However, LNR MALAT could sponge with miR-204, impeding miR-204 to attack IGF2BP2. Restored-IGF2BP2 promoted m6A-c-MYC translation efficiency, facilitating c-MYC-mediated migration, invasion and apoptosis suppression.⁵⁰ Though there is no available source concerning YTHDF1 and IGF2BP2's pri-miRNAs processing-promoting function, they contributed to cancer development via miRNAs-m6A way.

To date, it is found that three readers, including HNRNPA2B1, NKAP and YTHDC1, exhibited miRNAs processing function. There is only YTHDF2 was revealed as a negative reader of miRNAs processing. As for readers' roles in cancers, it is demonstrated that m6A readers and miRNAs' multiple relationships contribute to the occurrence and development of various cancers, relatively showing an oncogenic tendency. Being convinced of readers' dual effects on miRNA processing, it is understandable that the writer could drive both influences on miRNAs like NSun2 and METTL3. On the other hand, demethyltransferase ALKBH5 is somehow able to exhibit both processing-promoting and degradation effects. However, for now, a majority of current reports separated m6A readers from research content, and depicted writers or erasers unilaterally. Exploring their individual superficial devotions narrows our horizon against the detailed miRNAs biosynthesis regulatory mechanism in an m6A-dependent way. These results had an important implication for the developing conceptual premise that the m6A modification function concerning pri-miRNAs processing depends on m6A levels alteration and readers' decisive recognizing effects. The exertion of the m6A modification function lies in the relative expression of these three types of regulators, commanding a rigorous and scientific attitude toward experiment and analysis.

FIGURE 3 M6A modification and miRNAs are involved in various cancers. Articles concerning m6A and miRNAs cover the digestive system, respiratory system, urogenital system, neural system, skeletal system and endocrine system

7 | THE EXPECTATION FOR THERAPEUTIC VALUE OF M6A AND MIRNAS

As we investigate closely on literature concerning m6A and miRNAs, we find out that more attention has been focused on clinical personality treatment. A comprehensive analysis of the combined effects of m6A regulators and miRNAs supports that certain combination indeed facilitates tumour malignancy or suppress tumour progression. Molecule-specific mediated therapy is one of the foundations of precision medicine. In this case, it is suggested that m6A has inestimable clinical value via utilizing blocking agents to provide precise attacks and applying analogues for specific deficiency compensation (Table [6\)](#page-11-0). In gallbladder cancer, DAC exhibited a possible therapeutic value for those with METTL3 high expression through disturbing METTL3/miR-92b-3p related cancer progression. 54 In breast cancer, metformin exerted an anti-growth effect via reinforcing miR-483-3p level, which could silence oncogenic METTL3 and restore apoptosis-related p21. Consequently, metformin might be clinically potential to apply to METTL3-overexpressed breast cancer patients.⁴⁸

What is more, according to available research findings, aberrant m6A regulators and miRNAs amount may imply certain drug sensitivity or drug resistance phenomena. It is well-known that drug resistance is the main challenge of anti-tumour treatment all the way long. Under this circumstance, m6A regulators could enhance therapeutic response and alleviate drug resistance, laying a foundation for combination therapy. m6A-dependent miR-17-92 cluster biogenesis activated the mTOR pathway, accelerating mTOR inhibitor everolimus sensitivity in gastric cancer. 81 mTOR inhibitor everolimus is now applied in advanced renal cancer patients. If the METTL3/AKT/mTOR signalling exists in renal cancer as well, the patients with overexpressed METTL3 might benefit more from everolimus. Carolyn M. Klinge and colleagues reported that overexpressed HNPNPA2B1 is involved in endocrine resistance, which suggested that HNPNPA2B1 inhibitors might be applied to alleviate endocrine resistance to improve clinical efficacy.⁷⁴ In breast cancer, excessive METTL3 resisted cell apoptosis as well as exhibited Adriamycin resistance. METTL3-induced mature miR-221-3p bound to HIPK2 3'UTR, attenuating to repress drug resistance-related Che1 expression and finally driving Adriamycin resistance. METTL3 inhibitor might pro-vide a reference for treatment.^{[82](#page-15-0)}

Serious research has provided evidence about targeted-based therapy at a cellular level or in animal models. However, a majority problem of m6A-miRNAs-based therapy with these studies is how to construct an exchanging platform of basic research and clinical applications. As for miRNA therapeutics, firstly, multiple miRNA targets indicated a major uncertain miRNA-based therapeutic effect. A contradiction might occur while a particular miRNA targets both oncogenes and tumour suppressors. In this situation, apart from attacking expected targets, the off-target effect might cause toxicity in other normal tissue or organs.⁸³ Furthermore, miRNA delivery vehicle construction is another difficulty in targeting function implementation. An excellent delivery system helps to avoid miRNAs degradation in serum and enables tissue-specific delivery to alleviate off-target toxicity.^{[84](#page-15-0)} Finally, the modulatory degree of miRNAs is somehow within a small

scope of downstream gene expression, the miRNA-based therapy effect might not be promising. So, it is a key point to explore how to magnify miRNA-based drug efficiency before clinical application. And with regard to m6A regulator analogue or antagonist, its clinical research still lags far behind basic research. Similarly, with wideranging implications and dual effects on cancers, m6A-miRNA-related targeting therapy still faces the problem of off-target toxicity. As mentioned above, there is a long way to go before m6A-miRNAs-based targeting treatment is available in clinical application.

8 | CONCLUSIONS

N6-methyladenosine modification is one of the most prevalent posttranscriptional epigenetics in mammalian RNAs. This review shares the mutual interactions between m6A regulators and miRNAs; focuses on the combined effect of various m6A regulators and miRNAs in different cancers. m6A regulators could regulate miRNAs processing, meanwhile, miRNAs could attack m6A regulator mRNAs to alter the m6A level in return. Particularly, miRNAs are capable of assisting m6A regulators to better exert their functions through binding to the m6A motif. Notably, m6A readers possess the fateful effects of miRNAs biosynthesis, transcending writers and eraser's m6A abundance alteration functions. Future research regard to m6A-related miRNAs processing should concentrate on the combined effect of three types of m6A regulators.

From the digestive system, respiratory system, productive system, neural system, skeletal system and endocrine system (Figure [3](#page-11-0)), m6A and miRNAs contribute to cancer involving aspects converting from selfsufficient growth signals, resisting cell death, tissue metastasis, and invasion as well as deregulating cell energy. Theoretically, different combinations of m6A regulators and miRNAs and downstream pathways create infinite possibilities. Indeed, the role of m6A modification in miRNArelative cancer depends on its function and location within the whole signalling, affected by miRNAs regulative effects as well as subsequent pathways. Though it is reasonable that m6A regulators are like a double-edged sword in cancers, existing published literature indicated certain trends.

For the reason that aberrant N6-methyladenosine level contributes to cancer malignancy and clinical deterioration, m6A modification exhibited potential therapeutic value. However, subject to the existing limited acquaintance and complexity of m6A regulators and miRNAs, it is a huge challenge to apply to clinical medicine from available reports. Consequently, further and overall studies are indispensable for paving a way for clinical applications.

AUTHOR CONTRIBUTIONS

Huiru Feng and Fan Wang contributed to the conception and design of this work. Xiong Liu supervised the study. All authors contributed to the collection and interpretation of literature and writing, review and revision of the manuscript. All authors read and approved the final manuscript.

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CONFLICT OF INTEREST

The authors declare no competing interests.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Fan Wang <https://orcid.org/0000-0003-0914-947X>

REFERENCES

- 1. Wei CM, Gershowitz A, Moss B. 5'-Terminal and internal methylated nucleotide sequences in HeLa cell mRNA. Biochemistry. 1976;15:397- 401. doi:[10.1021/bi00647a024](info:doi/10.1021/bi00647a024)
- 2. Wei CM, Moss B. Nucleotide sequences at the N6-methyladenosine sites of HeLa cell messenger ribonucleic acid. Biochemistry. 1977;16: 1672-1676. doi:[10.1021/bi00627a023](info:doi/10.1021/bi00627a023)
- 3. Dominissini D, Moshitch-Moshkovitz S, Schwartz S, et al. Topology of the human and mouse m(6)A RNA methylomes revealed by m(6)Aseq. Nature. 2012;485:201-U84. doi:[10.1038/nature11112](info:doi/10.1038/nature11112)
- 4. Meyer KD, Saletore Y, Zumbo P, Elemento O, Mason CE, Jaffrey SR. Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. Cell. 2012;149:1635-1646. doi[:10.](info:doi/10.1016/j.cell.2012.05.003) [1016/j.cell.2012.05.003](info:doi/10.1016/j.cell.2012.05.003)
- 5. Yang Y, Hsu PJ, Chen YS, Yang YG. Dynamic transcriptomic m(6)A decoration: writers, erasers, readers and functions in RNA metabolism. Cell Res. 2018;28:616-624. doi[:10.1038/s41422-018-0040-8](info:doi/10.1038/s41422-018-0040-8)
- 6. Shi HL, Wei JB, He C. Where, when, and how: context-dependent functions of RNA methylation writers, readers, and erasers. Mol Cell. 2019;74:640-650. doi:[10.1016/j.molcel.2019.04.025](info:doi/10.1016/j.molcel.2019.04.025)
- 7. Liu Q, Gregory RI. RNAmod: an integrated system for the annotation of mRNA modifications. Nucleic Acids Res. 2019;47:W548-W555. doi: [10.1093/nar/gkz479](info:doi/10.1093/nar/gkz479)
- 8. Dai DJ, Wang HY, Zhu LY, Jin HC, Wang X. N6-methyladenosine links RNA metabolism to cancer progression. Cell Death Dis. 2018;9:124. doi:[10.1038/s41419-017-0129-x](info:doi/10.1038/s41419-017-0129-x)
- 9. Pan YT, Ma P, Liu Y, Li W, Shu YQ. Multiple functions of m(6)A RNA methylation in cancer. J Hematol Oncol. 2018;11:48. doi:[10.1186/](info:doi/10.1186/s13045-018-0590-8) [s13045-018-0590-8](info:doi/10.1186/s13045-018-0590-8)
- 10. Zhou ZJ, Lv JC, Yu H, et al. Mechanism of RNA modification N6-methyladenosine in human cancer. Mol Cancer. 2020;19:104. doi: [10.1186/s12943-020-01216-3](info:doi/10.1186/s12943-020-01216-3)
- 11. Lagos-Quintana M, Rauhut R, Lendeckel W, Tuschl T. Identification of novel genes coding for small expressed RNAs. Science. 2001;294: 853-858. doi:[10.1126/science.1064921](info:doi/10.1126/science.1064921)
- 12. Reinhart BJ, Slack FJ, Basson M, et al. The 21-nucleotide let-7 RNA regulates developmental timing in Caenorhabditis elegans. Nature. 2000;403:901-906. doi:[10.1038/35002607](info:doi/10.1038/35002607)
- 13. Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE, Mello CC. Potent and specific genetic interference by double-stranded RNA in Caenorhabditis elegans. Nature. 1998;391:806-811. doi:[10.1038/](info:doi/10.1038/35888) [35888](info:doi/10.1038/35888)
- 14. Lee YS, Dutta A. MicroRNAs in cancer. Annu Rev Pathol-Mech Dis. 2009;4:199-227. doi[:10.1146/annurev.pathol.4.110807.092222](info:doi/10.1146/annurev.pathol.4.110807.092222)
- 15. Iorio MV, Croce CM. MicroRNAs in cancer: small molecules with a huge impact. J Clin Oncol. 2009;27:5848-5856. doi[:10.1200/jco.](info:doi/10.1200/jco.2009.24.0317) [2009.24.0317](info:doi/10.1200/jco.2009.24.0317)
- 16. Garzon R, Calin GA, Croce CM. MicroRNAs in cancer. Annu Rev Med. 2009;60:167-179. doi:[10.1146/annurev.med.59.053006.104707](info:doi/10.1146/annurev.med.59.053006.104707)
- 17. Croce CM. Causes and consequences of microRNA dysregulation in cancer. Nat Rev Genet. 2009;10:704-714. doi[:10.1038/nrg2634](info:doi/10.1038/nrg2634)
- 18. Alarcon CR, Lee H, Goodarzi H, Halberg N, Tavazoie SF. N-6-methyladenosine marks primary microRNAs for processing. Nature. 2015;519:482-485. doi:[10.1038/nature14281](info:doi/10.1038/nature14281)
- 19. Bokar JA, Shambaugh ME, Polayes D, Matera AG, Rottman FM. Purification and cDNA cloning of the AdoMet-binding subunit of the human mRNA (N6-adenosine)-methyltransferase. RNA (New York, NY). 1997;3:1233-1247.
- 20. Liu JZ, Yue YN, Han DL, et al. A METTL3-METTL14 complex mediates mammalian nuclear RNA N-6-adenosine methylation. Nat Chem Biol. 2014;10:93-95. doi[:10.1038/nchembio.1432](info:doi/10.1038/nchembio.1432)
- 21. Scholler E, Weichmann F, Treiber T, et al. Interactions, localization, and phosphorylation of the m(6)A generating METTL3-METTL14-WTAP complex. RNA. 2018;24:499-512. doi[:10.1261/rna.064063.117](info:doi/10.1261/rna.064063.117)
- 22. Ping XL, Sun BF, Wang L, et al. Mammalian WTAP is a regulatory subunit of the RNA N6-methyladenosine methyltransferase. Cell Res. 2014;24:177-189. doi:[10.1038/cr.2014.3](info:doi/10.1038/cr.2014.3)
- 23. Yue YA, Liu J, Cui XL, et al. VIRMA mediates preferential m(6)A $mRNA$ methylation in $3'$ UTR and near stop codon and associates with alternative polyadenylation. Cell Discov. 2018;4:10. doi:[10.1038/](info:doi/10.1038/s41421-018-0019-0) [s41421-018-0019-0](info:doi/10.1038/s41421-018-0019-0)
- 24. Patil DP, Chen CK, Pickering BF, et al. m(6)a RNA methylation promotes XIST-mediated transcriptional repression. Nature. 2016;537: 369-373. doi:[10.1038/nature19342](info:doi/10.1038/nature19342)
- 25. Moindrot B, Cerase A, Coker H, et al. A pooled shRNA screen identifies Rbm15, Spen, and Wtap as factors required for Xist RNAmediated silencing. Cell Rep. 2015;12:562-572. doi:[10.1016/j.celrep.](info:doi/10.1016/j.celrep.2015.06.053) [2015.06.053](info:doi/10.1016/j.celrep.2015.06.053)
- 26. Knuckles P, Lence T, Haussmann IU, et al. Zc3h13/Flacc is required for adenosine methylation by bridging the mRNA-binding factor Rbm15/Spenito to the m(6)A machinery component Wtap/Fl(2)d. Genes Dev. 2018;32:415-429. doi[:10.1101/gad.309146.117](info:doi/10.1101/gad.309146.117)
- 27. Warda AS, Kretschmer J, Hackert P, et al. Human METTL16 is a N-6-methyladenosine (m(6)A) methyltransferase that targets premRNAs and various non-coding RNAs. EMBO Rep. 2017;18:2004- 2014. doi:[10.15252/embr.201744940](info:doi/10.15252/embr.201744940)
- 28. Yuan S, Tang H, Xing JY, et al. Methylation by NSun2 represses the levels and function of MicroRNA 125b. Mol Cell Biol. 2014;34:3630- 3641. doi:[10.1128/mcb.00243-14](info:doi/10.1128/mcb.00243-14)
- 29. Jia GF, Fu Y, Zhao X, et al. N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. Nat Chem Biol. 2011; 7:885-887. doi[:10.1038/nchembio.687](info:doi/10.1038/nchembio.687)
- 30. Wei JB, Liu FG, Lu ZK, et al. Differential m(6)A, m(6)A(m), and m (1)A demethylation mediated by FTO in the cell nucleus and cytoplasm. Mol Cell. 2018;71:973–985.e5. doi[:10.1016/j.molcel.](info:doi/10.1016/j.molcel.2018.08.011) [2018.08.011](info:doi/10.1016/j.molcel.2018.08.011)
- 31. Zheng GQ, Dahl JA, Niu YM, et al. ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. Mol Cell. 2013;49:18-29. doi[:10.1016/j.molcel.2012.10.015](info:doi/10.1016/j.molcel.2012.10.015)
- 32. Xu C, Liu K, Ahmed H, Loppnau P, Schapira M, Min JR. Structural basis for the discriminative recognition of N-6-methyladenosine RNA by the human YT521-B homology domain family of proteins. J Biol Chem. 2015;290:24902-24913. doi:[10.1074/jbc.M115.](info:doi/10.1074/jbc.M115.680389) [680389](info:doi/10.1074/jbc.M115.680389)
- 33. Wang X, Zhao BS, Roundtree IA, et al. N-6-methyladenosine modulates messenger RNA translation efficiency. Cell. 2015;161:1388- 1399. doi:[10.1016/j.cell.2015.05.014](info:doi/10.1016/j.cell.2015.05.014)
- 34. Du H, Zhao Y, He JQ, et al. YTHDF2 destabilizes m(6)A-containing RNA through direct recruitment of the CCR4-NOT deadenylase complex. Nat Commun. 2016;7:12626. doi:[10.1038/ncomms12626](info:doi/10.1038/ncomms12626)
- 35. Shi HL, Wang X, Lu ZK, et al. YTHDF3 facilitates translation and decay of N-6-methyladenosine-modified RNA. Cell Res. 2017;27:315- 328. doi[:10.1038/cr.2017.15](info:doi/10.1038/cr.2017.15)
- 36. Roundtree IA, Luo GZ, Zhang ZJ, et al. YTHDC1 mediates nuclear export of N-6 - methyladenosine methylated mRNAs. Elife. 2017;6: e31311. doi:[10.7554/eLife.31311](info:doi/10.7554/eLife.31311)
- 37. Xiao W, Adhikari S, Dahal U, et al. Nuclear m(6)A reader YTHDC1 regulates mRNA splicing. Mol Cell. 2016;61:507-519. doi:[10.1016/j.](info:doi/10.1016/j.molcel.2016.01.012) [molcel.2016.01.012](info:doi/10.1016/j.molcel.2016.01.012)
- 38. Hsu PJ, Zhu YF, Ma HH, et al. Ythdc2 is an N-6-methyladenosine binding protein that regulates mammalian spermatogenesis. Cell Res. 2017;27:1115-1127. doi:[10.1038/cr.2017.99](info:doi/10.1038/cr.2017.99)
- 39. Ma LF, Chen TX, Zhang X, et al. The m(6)A reader YTHDC2 inhibits lung adenocarcinoma tumorigenesis by suppressing SLC7A11-dependent antioxidant function. Redox Biol. 2021;38:101801. doi[:10.1016/j.redox.](info:doi/10.1016/j.redox.2020.101801) [2020.101801](info:doi/10.1016/j.redox.2020.101801)
- 40. Liu N, Dai Q, Zheng GQ, He C, Parisien M, Pan T. N-6-methyladenosine-dependent RNA structural switches regulate RNA-protein interactions. Nature. 2015;518:560-564. doi:[10.1038/nature14234](info:doi/10.1038/nature14234)
- 41. Alarcon CR, Goodarzi H, Lee H, Liu XH, Tavazoie S, Tavazoie SF. HNRNPA2B1 is a mediator of m(6)A-dependent nuclear RNA processing events. Cell. 2015;162:1299-1308. doi:[10.1016/j.cell.2015.](info:doi/10.1016/j.cell.2015.08.011) [08.011](info:doi/10.1016/j.cell.2015.08.011)
- 42. Huang HL, Weng HY, Sun WJ, et al. Recognition of RNA N-6-methyladenosine by IGF2BP proteins enhances mRNA stability and translation. Nat Cell Biol. 2018;20:285-295. doi[:10.1038/s41556-](info:doi/10.1038/s41556-018-0045-z) [018-0045-z](info:doi/10.1038/s41556-018-0045-z)
- 43. Meyer KD, Patil DP, Zhou J, et al. 5' UTR m(6)A promotes capindependent translation. Cell. 2015;163:999-1010. doi[:10.1016/j.cell.](info:doi/10.1016/j.cell.2015.10.012) [2015.10.012](info:doi/10.1016/j.cell.2015.10.012)
- 44. Zhang JL, Bai RH, Li M, et al. Excessive miR-25-3p maturation via N-6-methyladenosine stimulated by cigarette smoke promotes pancreatic cancer progression. Nat Commun. 2019;10:1858. doi[:10.](info:doi/10.1038/s41467-019-09712-x) [1038/s41467-019-09712-x](info:doi/10.1038/s41467-019-09712-x)
- 45. Gregory RI, Yan KP, Amuthan G, et al. The microprocessor complex mediates the genesis of microRNAs. Nature. 2004;432:235-240. doi: [10.1038/nature03120](info:doi/10.1038/nature03120)
- 46. Bartel DP. Metazoan microRNAs. Cell. 2018;173:20-51. doi[:10.1016/](info:doi/10.1016/j.cell.2018.03.006) [j.cell.2018.03.006](info:doi/10.1016/j.cell.2018.03.006)
- 47. Xue JL, Xiao P, Yu XD, Zhang X. A positive feedback loop between AlkB homolog 5 and miR-193a-3p promotes growth and metastasis in esophageal squamous cell carcinoma. Hum Cell. 2021;34:502-514. doi:[10.1007/s13577-020-00458-z](info:doi/10.1007/s13577-020-00458-z)
- 48. Cheng L, Zhang X, Huang YZ, et al. Metformin exhibits antiproliferation activity in breast cancer via miR-483-3p/METTL3/m(6) A/p21 pathway. Oncogenesis. 2021;10:7. doi[:10.1038/s41389-](info:doi/10.1038/s41389-020-00290-y) [020-00290-y](info:doi/10.1038/s41389-020-00290-y)
- 49. Cui Y, Wang Q, Lin J, et al. miRNA-193a-3p regulates the AKT2 pathway to inhibit the growth and promote the apoptosis of glioma cells by targeting ALKBH5. Front Oncol. 2021;11:600451. doi[:10.3389/](info:doi/10.3389/fonc.2021.600451) [fonc.2021.600451](info:doi/10.3389/fonc.2021.600451)
- 50. Ye M, Dong S, Hou HT, Zhang T, Shen MH. Oncogenic role of long noncoding RNAMALAT1 in thyroid cancer progression through regulation of the miR-204/IGF2BP2/m6A-MYC signaling. Mol Ther Nucl Acids. 2021;23:1-12. doi:[10.1016/j.omtn.2020.09.023](info:doi/10.1016/j.omtn.2020.09.023)
- 51. Chen T, Hao YJ, Zhang Y, et al. m(6)A RNA methylation is regulated by microRNAs and promotes reprogramming to pluripotency. Cell Stem Cell. 2015;16:289-301. doi[:10.1016/j.stem.2015.01.016](info:doi/10.1016/j.stem.2015.01.016)
- 52. Zepecki JP, Karambizi D, Fajardo JE, et al. miRNA-mediated loss of m (6)A increases nascent translation in glioblastoma. PLoS Genet. 2021; 17:e1009086. doi:[10.1371/journal.pgen.1009086](info:doi/10.1371/journal.pgen.1009086)
- 53. Cheng JM, Xu LL, Deng LQ, et al. RNA N-6-methyladenosine modification is required for miR-98/MYCN axis-mediated inhibition of neuroblastoma progression. Sci Rep. 2020;10:13624. doi:[10.1038/](info:doi/10.1038/s41598-020-64682-1) [s41598-020-64682-1](info:doi/10.1038/s41598-020-64682-1)
- 54. Lin RR, Zhan M, Yang LH, et al. Deoxycholic acid modulates the progression of gallbladder cancer through N-6-methyladenosinedependent microRNA maturation. Oncogene. 2020;39:4983-5000. doi:[10.1038/s41388-020-1349-6](info:doi/10.1038/s41388-020-1349-6)
- 55. Bi XH, Lv X, Liu DJ, et al. METTL3-mediated maturation of miR-126-5p promotes ovarian cancer progression via PTEN-mediated PI3K/Akt/mTOR pathway. Cancer Gene Ther. 2021;28:335-349. doi: [10.1038/s41417-020-00222-3](info:doi/10.1038/s41417-020-00222-3)
- 56. Han J, Wang JZ, Yang X, et al. METTL3 promote tumor proliferation of bladder cancer by accelerating pri-miR221/222 maturation in m6A-dependent manner. Mol Cancer. 2019;18:110. doi:[10.1186/](info:doi/10.1186/s12943-019-1036-9) [s12943-019-1036-9](info:doi/10.1186/s12943-019-1036-9)
- 57. Huang CX, Liang JX, Lin SD, et al. N-6-methyladenosine associated silencing of miR-193b promotes cervical cancer aggressiveness by targeting CCND1. Front Oncol. 2021;11:666597. doi:[10.3389/fonc.](info:doi/10.3389/fonc.2021.666597) [2021.666597](info:doi/10.3389/fonc.2021.666597)
- 58. Chen XX, Xu M, Xu XN, et al. METTL14 suppresses CRC progression via regulating N6-methyladenosine-dependent primary miR-375 processing. Mol Ther. 2020;28:599-612. doi:[10.1016/j.ymthe.2019.](info:doi/10.1016/j.ymthe.2019.11.016) [11.016](info:doi/10.1016/j.ymthe.2019.11.016)
- 59. Wang HS, Deng QQ, Lv ZY, et al. N6-methyladenosine induced miR-143-3p promotes the brain metastasis of lung cancer via regulation of VASH1. Mol Cancer. 2019;18:181. doi:[10.1186/s12943-019-1108-x](info:doi/10.1186/s12943-019-1108-x)
- 60. Peng W, Li J, Chen RR, et al. Upregulated METTL3 promotes metastasis of colorectal cancer via miR-1246/SPRED2/MAPK signaling pathway. J Exp Clin Cancer Res. 2019;38:393. doi[:10.1186/s13046-](info:doi/10.1186/s13046-019-1408-4) [019-1408-4](info:doi/10.1186/s13046-019-1408-4)
- 61. Yi DD, Wang R, Shi XB, Xu L, Yilihamu Y, Sang JF. METTL14 promotes the migration and invasion of breast cancer cells by modulating N6-methyladenosine and hsa-miR-146a-5p expression. Oncol Rep. 2020;43:1375-1386. doi[:10.3892/or.2020.7515](info:doi/10.3892/or.2020.7515)
- 62. Du MJ, Zhang YJ, Mao YS, et al. MiR-33a suppresses proliferation of NSCLC cells via targeting METTL3 mRNA. Biochem Biophys Res Commun. 2017;482:582-589. doi[:10.1016/j.bbrc.2016.11.077](info:doi/10.1016/j.bbrc.2016.11.077)
- 63. Wu HY, Li FJ, Zhu R. miR-338-5p inhibits cell growth and migration via inhibition of the METTL3/m6A/c-Myc pathway in lung cancer. Acta Biochim Biophys Sin. 2021;53:304-316. doi[:10.1093/abbs/](info:doi/10.1093/abbs/gmaa170) [gmaa170](info:doi/10.1093/abbs/gmaa170)
- 64. Cui XC, Wang ZF, Li JH, et al. Cross talk between RNA N6-methyladenosine methyltransferase-like 3 and miR-186 regulates hepatoblastoma progression through Wnt/beta-catenin signalling pathway. Cell Prolif. 2020;53:e12768. doi[:10.1111/cpr.12768](info:doi/10.1111/cpr.12768)
- 65. He HB, Wu W, Sun ZE, Chai LX. MiR-4429 prevented gastric cancer progression through targeting METTL3 to inhibit m(6)A-caused stabilization of SEC62. Biochem Biophys Res Commun. 2019;517:581-587. doi:[10.1016/j.bbrc.2019.07.058](info:doi/10.1016/j.bbrc.2019.07.058)
- 66. Wei WW, Huo BS, Shi XL. miR-600 inhibits lung cancer via downregulating the expression of METTL3. Cancer Manag Res. 2019;11: 1177-1187. doi:[10.2147/cmar.S181058](info:doi/10.2147/cmar.S181058)
- 67. Song ZY, Jia G, Ma PZ, Cang SD. Exosomal miR-4443 promotes cisplatin resistance in non-small cell lung carcinoma by regulating FSP1 m6A modification-mediated ferroptosis. Life Sci. 2021;276:119399. doi:[10.1016/j.lfs.2021.119399](info:doi/10.1016/j.lfs.2021.119399)
- 68. Liu WL, Gao XW, Chen XL, et al. miR-139-5p loss-mediated WTAP activation contributes to hepatocellular carcinoma progression by promoting the epithelial to mesenchymal transition. Front Oncol. 2021;11:611544. doi:[10.3389/fonc.2021.611544](info:doi/10.3389/fonc.2021.611544)
- 69. Xu YY, Ye S, Zhang N, et al. The FTO/miR-181b-3p/ARL5B signaling pathway regulates cell migration and invasion in breast cancer. Cancer Commun. 2020;40:484-500. doi[:10.1002/cac2.12075](info:doi/10.1002/cac2.12075)

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- 70. Mo WL, Deng LJ, Cheng Y, Yu WJ, Yang YH, Gu WD. Circular RNA hsa_circ_0072309 promotes tumorigenesis and invasion by regulating the miR-607/FTO axis in non-small cell lung carcinoma. Aging-Us. 2021;13:11629-11645.
- 71. Yuan Y, Yan GG, He MY, et al. ALKBH5 suppresses tumor progression via an m(6)A-dependent epigenetic silencing of pre-miR-181b-1/YAP signaling axis in osteosarcoma. Cell Death Dis. 2021;12:60. doi: [10.1038/s41419-020-03315-x](info:doi/10.1038/s41419-020-03315-x)
- 72. Zhu HT, Gan XL, Jiang XW, Diao S, Wu H, Hu JG. ALKBH5 inhibited autophagy of epithelial ovarian cancer through miR-7 and BCL-2. J Exp Clin Cancer Res. 2019;38:163. doi:[10.1186/s13046-019-](info:doi/10.1186/s13046-019-1159-2) [1159-2](info:doi/10.1186/s13046-019-1159-2)
- 73. Chen ZY, Chen X, Lei TY, et al. Integrative analysis of NSCLC identifies LINC01234 as an oncogenic lncRNA that interacts with HNRNPA2B1 and regulates miR-106b biogenesis. Mol Ther. 2020;28: 1479-1493. doi[:10.1016/j.ymthe.2020.03.010](info:doi/10.1016/j.ymthe.2020.03.010)
- 74. Klinge CM, Piell KM, Tooley CS, Rouchka EC. HNRNPA2/B1 is upregulated in endocrine-resistant LCC9 breast cancer cells and alters the miRNA transcriptome when overexpressed in MCF-7 cells. Sci Rep. 2019;9:9430. doi:[10.1038/s41598-019-45636-8](info:doi/10.1038/s41598-019-45636-8)
- 75. Hou YC, Zhang QW, Pang WJ, et al. YTHDC1-mediated augmentation of miR-30d in repressing pancreatic tumorigenesis via attenuation of RUNX1-induced transcriptional activation of Warburg effect. Cell Death Differ. 2021;28:3105-3124. doi[:10.1038/s41418-021-](info:doi/10.1038/s41418-021-00804-0) [00804-0](info:doi/10.1038/s41418-021-00804-0)
- 76. Li J, Wu L, Pei ML, Zhang Y. YTHDF2, a protein repressed by miR-145, regulates proliferation, apoptosis, and migration in ovarian cancer cells. J Ovarian Res. 2020;13:111. doi[:10.1186/s13048-020-](info:doi/10.1186/s13048-020-00717-5) [00717-5](info:doi/10.1186/s13048-020-00717-5)
- 77. Yang Z, Li J, Feng GX, et al. MicroRNA-145 modulates N-6-methyladenosine levels by targeting the 3'-untranslated mRNA region of the N-6-methyladenosine binding YTH domain family 2 protein. J Biol Chem. 2017;292:3614-3623. doi:[10.1074/jbc.M116.](info:doi/10.1074/jbc.M116.749689) [749689](info:doi/10.1074/jbc.M116.749689)
- 78. Du C, Lv CH, Feng Y, Yu SW. Activation of the KDM5A/miRNA-495/YTHDF2/m6A-MOB3B axis facilitates prostate cancer progression. J Exp Clin Cancer Res. 2020;39:223. doi[:10.1186/s13046-020-](info:doi/10.1186/s13046-020-01735-3) [01735-3](info:doi/10.1186/s13046-020-01735-3)
- 79. Zhou JY, Xiao D, Qiu TT, Li J, Liu ZT. Loading microRNA-376c in extracellular vesicles inhibits properties of non-small cell lung cancer cells by targeting YTHDF1. Technol Cancer Res Treat. 2020;19: 1533033820977525. doi:[10.1177/1533033820977525](info:doi/10.1177/1533033820977525)
- 80. Xu C, Yuan B, He T, Ding B, Li S. Prognostic values of YTHDF1 regulated negatively by mir-3436 in glioma. J Cell Mol Med. 2020;24: 7538-7549. doi[:10.1111/jcmm.15382](info:doi/10.1111/jcmm.15382)
- 81. Sun YT, Li S, Yu WB, et al. N-6-methyladenosine-dependent pri-miR-17-92 maturation suppresses PTEN/TMEM127 and promotes sensitivity to everolimus in gastric cancer. Cell Death Dis. 2020;11:836. doi:[10.1038/s41419-020-03049-w](info:doi/10.1038/s41419-020-03049-w)
- 82. Pan XP, Hong XL, Li SM, Meng P, Xiao F. METTL3 promotes adriamycin resistance in MCF-7 breast cancer cells by accelerating pri-micro-RNA-221-3p maturation in a m6A-dependent manner. Exp Mol Med. 2021;53:91-102. doi[:10.1038/s12276-020-00510-w](info:doi/10.1038/s12276-020-00510-w)
- 83. Rupaimoole R, Slack FJ. MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. Nat Rev Drug Discov. 2017;16:203-221. doi:[10.1038/nrd.2016.246](info:doi/10.1038/nrd.2016.246)
- 84. Cheng CJ, Bahal R, Babar IA, et al. MicroRNA silencing for cancer therapy targeted to the tumour microenvironment. Nature. 2015;518: 107-110. doi:[10.1038/nature13905](info:doi/10.1038/nature13905)
- 85. Ma JZ, Yang F, Zhou CC, et al. METTL14 suppresses the metastatic potential of hepatocellular carcinoma by modulating N-6-methyladenosine-dependent primary microRNA processing. Hepatology. 2017; 65:529-543. doi[:10.1002/hep.28885](info:doi/10.1002/hep.28885)
- 86. Cai XL, Wang X, Cao C, et al. HBXIP-elevated methyltransferase METTL3 promotes the progression of breast cancer via inhibiting tumor suppressor let-7g. Cancer Lett. 2018;415:11-19. doi[:10.1016/j.](info:doi/10.1016/j.canlet.2017.11.018) [canlet.2017.11.018](info:doi/10.1016/j.canlet.2017.11.018)
- 87. Hu H, Kong Q, Huang XX, et al. Longnon-coding RNA BLACAT2 promotes gastric cancer progression via the miR-193b-5p/METTL3 pathway. J Cancer. 2021;12:3209-3221. doi:[10.7150/jca.50403](info:doi/10.7150/jca.50403)
- 88. Li LL, Xie RR, Wei QC. Network analysis of miRNA targeting m6Arelated genes in patients with esophageal cancer. PeerJ. 2021;9: e11893. doi:[10.7717/peerj.11893](info:doi/10.7717/peerj.11893)
- 89. Li KX, Chen JY, Lou XY, et al. HNRNPA2B1 affects the prognosis of esophageal cancer by regulating the miR-17-92 cluster. Front Cell Dev Biol. 2021;9:658642. doi:[10.3389/fcell.2021.658642](info:doi/10.3389/fcell.2021.658642)
- 90. Li JF, Meng S, Xu MJ, et al. Downregulation of N-6-methyladenosine binding YTHDF2 protein mediated by miR-493-3p suppresses prostate cancer by elevating N-6-methyladenosine levels. Oncotarget. 2018;9:3752-3764. doi:[10.18632/oncotarget.23365](info:doi/10.18632/oncotarget.23365)

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