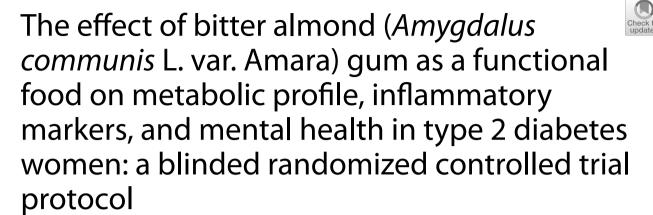
STUDY PROTOCOL Open Access



Saba Saati¹, Parvin Dehghan^{2*}, Fatemeh Azizi-Soleiman³ and Majid Mobasseri⁴

Abstract

Background Using functional foods in the prevention and treatment of type 2 diabetes mellitus (T2DM) has increased across the world owing to their availability, cultural acceptability, and lower side effects. The present study will aim to examine the impact of bitter almond (*Amygdalus communis* L. var. Amara) gum as a functional food on metabolic profile, inflammatory markers, and mental health in women with T2DM.

Methods We will conduct a randomized, triple-blind, placebo-controlled trial. A total of 44 women with T2DM will be randomly allocated into two groups: an intervention group (n = 20) and a placebo group (n = 20). Patients will receive either 5 g/d of bitter melon gum or a placebo for 8 weeks. Clinical and biochemical outcome parameters which include glycemic indices, lipid profile, inflammatory markers, oxidative stress indices, tryptophan (Trp), kynurenine (KYN), cortisol, glucagon-like peptide 1 (GLP-1), leptin, adiponectin, ghrelin, peroxisome proliferator-activated receptor (PPAR) gene expression, brain-derived neurotrophic factor (BDNF), endothelial cell adhesion molecules, plasminogen, cluster deference 4 (CD4), cluster deference 8 (CD8), anthropometric indices, blood pressure, dietary intake, and mental health will be measured at the baseline and end of the study. Statistical analysis will be conducted using the SPSS software (version 24), and P value less than 0.05 will be considered statistically significant.

Discussion The present randomized controlled trial will aim to investigate any beneficial effects of bitter almond gum supplementation on the cardio-metabolic, immune-inflammatory, and oxidative stress biomarkers, as well as mental health in women with T2DM.

Ethics and dissemination The study protocol was approved by the Ethical Committee of the Tabriz University of Medical Sciences (IR.TBZMED.REC.1399.726).

Trial registration Iranian Registry of Clinical Trials (www.irct.ir/IRCT20150205020965N7)

Keywords Type 2 diabetes, Inflammation, Oxidative stress, Bitter almond

*Correspondence: Parvin Dehghan dehghan.nut@gmail.com Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, wist http://creativecommons.org/ficenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Saati et al. Trials (2023) 24:35 Page 2 of 10

Background

Type 2 diabetes mellitus (T2DM), a metabolic disorder characterized by imbalanced blood sugar levels, is thought to be precipitated by a combination of impaired insulin secretion and insulin resistance [1]. Ethnicity, genetic predisposition, obesity, low physical activity, and unhealthy diet are involved in the pathogenesis of T2DM [2]. In 2019, it was estimated that 463 million people are living with T2DM worldwide [3]. This number has almost tripled over the last two decades and is projected to reach 578 million by 2030 and 700 million by 2045 [4]. The prevalence of T2DM has risen dramatically in countries experiencing epidemiologic transitions, including Asia, the Middle East, and North Africa [5]. The economic costs attributable to T2DM are also considerable. T2DM direct health care expenditure was estimated to be US\$760 billion in 2019 [3]. Thus, developing novel adjuvant therapies for the management of T2DM and its associated complications is vital. Recent progress in understanding the development and progression of diabetes has shown a strong association between hyperglycemia, oxidative stress, inflammation, gut microbiota, and T2DM [6].

Intracellular hyperglycemia increases reactive oxygen species (ROS) production, promotes advanced glycation end-product formation and activation of protein kinase C, and enhances polyol pathway flux. ROS stimulates the generation of inflammatory mediators and adhesion molecules, oxidized low-density lipoprotein (LDL) formation, and insulin resistance [7]. As oxidative stress can induce an inflammatory process, inflammation can also induce oxidative stress [8]. Following activation of the immune system, innate immune system cells proinflammatory produce cytokines and chemokines, which stimulate the production of reactive oxygen and/or nitrogen species. Pro-inflammatory cytokines activate macrophages to eliminate pathogens via the generation of ROS [9]. Increased inflammatory markers may be related to mental health conditions [10]. Depressive disorders and impaired mental health-related quality of life are more prevalent among people with T2DM [11]. The relationship between T2DM and depression is thought to be bidirectional because of similar underlying pathological features in these conditions, including inflammation [12]. Higher serum levels and expression of inflammatory and pro-inflammatory markers in depressed patients with T2DM have been reported [13].

The microbiome may play an essential role in the development of T2DM [14]; an increase in the abundance of Firmicutes and Actinobacteria and a decrease in the proportion of Bacteroidetes can result in an inflammatory cascade, insulin resistance, and oxidative stress [15]. Potential mechanisms include modulation of

inflammation, increased intestinal permeability, changed glucose metabolism, altered fatty acid oxidation, synthesis, and energy expenditure [16]. Gut dysbiosis results in an increase in the leakage of bacterial products such as lipopolysaccharides (LPS) into the bloodstream [17]. LPS is recognized by Toll-like receptor 4 (TLR4), which is implicated in inflammation [18]. It has been shown that LPS activate intracellular pathways of c-Jun N-terminal kinase (JNK) and IκB kinase (IKK)-β [19]. Activation of JNK promotes the serine phosphorylation of insulin receptor substrate (IRS)-1, which inhibits normal signal transduction through the insulin receptor/IRS-1 axis, which inhibits its action, and leads to insulin resistance [20]. Activation of the IKKβ pathway induces the activation of nuclear factor (NF)-KB and enhances the expression of numerous pro-inflammatory mediators that are involved in insulin resistance [21]. Therefore, modulation of the gut microbiota can be considered a potential therapeutic strategy. One way would be to improve the gut microbiome by using prebiotics or probiotics. Prebiotics are non-digestible and fermentable fibers and sugars that promote the growth and/or activity of a beneficial gut microbiome [22]. Prebiotics increase the growth of beneficial bacteria such as lactobacilli and bifidobacteria, prevent the proliferation of harmful bacteria, and allow beneficial bacteria to produce short-chain fatty acids (SCFAs), including acetate, butyrate, and propionate [23]. These SCFAs decrease inflammation, improve intestinal membrane integrity, and increase the absorption of nutrients. A systematic review showed that prebiotics might improve metabolic and inflammatory biomarkers in adult women with T2DM [24].

Nowadays, the use of herbal medicine is proposed to be an adjunct to conventional antidiabetic treatments [25]. Medicinal plants contain glycosides, alkaloids, terpenoids, flavonoids, and carotenoids, which can exert antidiabetic effects [26]. Bitter almond gum is a key herb in Iranian medicine which is extracted from Amygdalus scoparia Spach. Remarkable in vitro antioxidant, antimicrobial, and antitumor effects of the water, methanol, and ethanol extracts of bitter almond kernels were demonstrated by Gomaa et al. [27]. Bitter almond gum increased the survival rate of Lactobacillus acidophilus La5 in tomato juice mixtures during fermentation, refrigeration, and exposure to gastric juice [28]. Bitter almond gum is an arabinogalactan polysaccharide that has prebiotic properties [29]. Bitter almond gum may have a favorable immune modulator effect possibly related to its prebiotic content [29]. To the best of our knowledge, there is only one study that evaluated the effect of bitter almond gum on metabolic parameters in overweight hyperlipidemic patients. In this study, bitter almond gum-enriched juice consumption significantly reduced

Saati et al. Trials (2023) 24:35 Page 3 of 10

body weight, body mass index (BMI), serum triglyceride (TG), hyperinsulinemia, and homeostatic model assessment for insulin resistance (HOMA-IR). We hypothesized that bitter almond gum, as adjuvant therapy, is able to improve metabolic, inflammatory, and oxidative stress biomarkers, as well as mental health in T2DM women.

Methods/design

Ethical consideration

The trial protocol is approved by the Tabriz University of Medical Sciences Ethical Committee (IR. TBZMED.REC.1399.726). The clinical trial number is IRCT20150205020965N7, which has been registered on the Iranian Registry of Clinical Trials (IRCT) website (www.irct.ir/IRCT20150205020965N7). This study will be conducted in accordance with the Declaration of Helsinki. Written informed consent will be obtained from all participants prior to study entry and after patient education about the study objective, process, schedule, potential risks, and benefits.

Protocol amendments

Any changes in the study protocol will be reviewed and approved by all authors. If any major change is made to the protocol, additional ethical approval must be obtained from the Ethics Committee.

Dissemination

The results of this experience will be published in a peerreviewed journal.

Safety assessments

The dosage to be administered in this clinical trial is within the recommended range based on the previous studies [30]. Moreover, documentation of all potential adverse events reported by the patients will be recorded by investigators from baseline through the follow-up period. If the reported adverse events are associated with the use of bitter almond gum, patients will be asked to stop taking the supplement, and they will be referred to a physician for therapy. Subjects with adverse events will be followed until the events will be resolved. Serious adverse events will be followed up to determine the final outcome. In addition, any possible negative effects will be notified to the ethics committee.

Study design and participants

The proposed study is a randomized, triple-blind, placebo-controlled, parallel-group clinical trial with a superiority framework to deliver an intention to treat (ITT) to evaluate the efficacy of bitter almond gum on cardiometabolic, immune-inflammatory, and oxidative stress biomarkers, as well as mental health, compared with a

placebo in women with T2DM. The allocation ratio will be 1:1. The study will begin later this year, 2022. Women with T2DM will be recruited from clinics of the Tabriz University of Medical Sciences, Iran, through public announcement systems and doctor referrals. A trained nutritionist will be served to include participants until the target sample size is reached and the project supervisor will assign participants to intervention groups based on the randomization list. The research design is presented in Fig. 1. This clinical trial protocol is based on the Standard Protocol Items: Recommendations for Interventional Trials (SPIRIT) 2013 checklist (Additional file 1: SPIRIT checklist) [31]. The diagram of the study protocol and initial and ongoing participant recruitment is demonstrated in Table 1.

Eligibility of criteria

Women aged between 30 and 65 years and diagnosed with T2DM (fasting blood sugar (FBG) ≥126 mg/dl [32]) that satisfy the inclusion criteria will be recruited. The inclusion criteria will be considered as follows: diagnosed with T2DM at least 6 months, body mass index (BMI) > 25 and < 35 kg/m², no weight changes during the last three months, use of glucose-lowering drugs (including sulfonylureas, glinides, biguanides, thiazolidinediones, dipeptidyl-peptidase 4 inhibitors, sodium-glucose transporter 2 inhibitors, alpha-glucosidase inhibitors, amylin mimetics, and incretin mimetic), dietary fiber intake of < 25 g per day, and propensity to take bitter almond gum supplement during the study. Patients will be excluded if they use insulin, glucocorticoids, laxatives, anti-obesity drugs, nonsteroidal anti-inflammatory drugs (NSAIDs), antidepressants, and antibiotics; take supplements including multivitamins, n-3 fatty acids, Ginkgo biloba, antioxidants, probiotics, and other prebiotics in the previous 3 months before the recruitment; have a history of weight loss or dieting in the last 6 months; have cancers and gastrointestinal, thyroid, heart, kidney, liver, lung, and infectious diseases; consume alcohol and smoke; and are pregnant or lactating.

Randomization and blinding

After obtaining the informed consent of the study participants, eligible subjects will complete a 2-week run-in period. During this time, patients will be requested to follow dietary and exercise advice according to recommendations for T2DM patients [33]. After the run-in period, the patients will be randomized to one of the two arms (bitter almond gum group or placebo group) in equal numbers, stratified by age and BMI. For stratified block randomization, random allocation software (RAS) will be used with block sizes of 2 and 4. Randomization will be performed by a third party that is not involved in this

Saati et al. Trials (2023) 24:35 Page 4 of 10

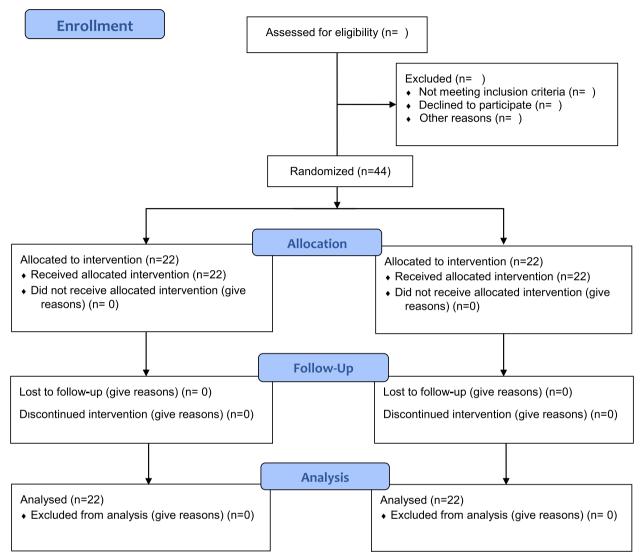


Fig. 1 Consolidated Standards of Reporting Trials (CONSORT) diagram

research. Block size will be concealed to all researchers until code-breaking. To enter a patient into the trial, one of the researchers will open the sequentially numbered sealed opaque envelopes. The study design is blinded to the main investigator, those who collect the data, the statistical consultant, and the participants.

Intervention

Diabetic patients will receive either 5g/d (2.5 g at breakfast and 2.5 g at dinner) of bitter almond gum powder (bitter almond gum, Flavinea Co, Iran) or maltodextrin powder (placebo) (Jiujiang Hurirong Trade Co, China) for 2 months. Participants will be instructed to use supplements with semi-solid food like yogurt or salad. Both bitter almond gum and maltodextrin powders are

odorless and tasteless and will be provided in identical opaque packages. Study participants will be informed on how to take their supplements. Patients will be followed by phone calls twice a week to ensure compliance with the intervention and dietary and physical activity guidelines and receive reports of possible adverse events experienced following bitter almond gum consumption. Compliance will be assessed based on a self-administered checklist and used packages by each person. Also, a checklist will be provided for participants to mark after each consumption of the prescribed supplement to evaluate for cases of non-compliance. Adherence to the regular consumption of powders (bitter almond gum or placebo) will also be determined by counting packages, also at least 10% of the supplements as noncompliant

Saati et al. Trials (2023) 24:35 Page 5 of 10

Table 1 Standard Protocol Items: Recommendations for Interventional Trials (SPIRIT) chart for the study process. The "X" is indicating what is done in the given period

	Enrolment	Allocation	Close-out		
TIMEPOINT**	-t ₁	0	t_1	t_2	t_x
ENROLMENT					
Eligibility screen	X				
Informed consent	X				
General characteristics	X				
Allocation		X			
INTERVENTIONS					
Bitter almond gum group			-	•	
Placebo group			-	•	
ASSESSMENTS					
3-day food record			X	X	
Body composition			X	X	
Blood pressure			X	X	X
 Biochemical assessments: Cardio-metabolic factors Immune-inflammatory factors Oxidative stress biomarkers PPAR gene expression Mental health 	X		Х	X	X
Physical activity status	X		X	X	

PPAR peroxisome proliferator-activated receptor

participants and, consequently, excluded from the study. Participants will be free to withdraw from participation in the study at any time upon request. Withdrawal criteria will be as follows: (1) Any clinical adverse event or medical condition occurs that the patient's continued participation in the study would not be in the best interest of her and (2) the patient meets a newly developed or not previously recognized exclusion criterion that

precludes further study participation. When the study ends, the results will be notified to all participants by post and published on a public website.

Sample size

Given the absence of data in this field, we sought to determine the sample size of our study based on the study of Sheu et al. [34] selected because of its similarity

Saati et al. Trials (2023) 24:35 Page 6 of 10

to our intervention. We calculated the sample size by taking into account the LDL levels. Considering a power of 90%, a 95% confidence interval, and the changes in LDL values as one of the primary outcomes (mean difference of 28.3 mg/dl between the two groups), the required sample size was calculated to be 19 for each study group. To compensate for a drop-out rate of 15% throughout the study, we increased the final sample size to 22 patients in each group.

Primary and secondary outcomes

Primary outcomes of the study will be FPG, glycosylated hemoglobin (HbA1c), and insulin. Secondary outcomes will include the lipid profile (total cholesterol (TC), highdensity lipoproteins (HDL), low-density lipoproteins (LDL), TG), high-sensitivity C-reactive protein (hs-CRP), tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1), interleukin-6 (IL-6), interleukin-10 (IL-10), interleukin-17 (IL-17), lipopolysaccharides (LPS), total antioxidant capacity (TAC), malondialdehyde (MDA), oxidative stress index (OSI), total oxidant status (TOS), superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), catalase (CAT), soluble receptor for AGEs (sRAGE), carboxymethyl lysine (CML), pentosidine, 8-iso-prostaglandin $F2\alpha$ (8-iso-PGF2 α), nitric oxide (NO), tryptophan (Trp), kynurenine (KYN), cortisol, glucagon-like peptide 1 (GLP-1), receptor for advanced glycation end products (RAGE), leptin, adiponectin, ghrelin, peroxisome proliferator-activated receptor (PPAR) gene expression, brain-derived neurotrophic factor (BDNF), endothelial cell adhesion molecules, plasminogen, cluster deference 4 (CD4), cluster deference 8 (CD8), anthropometric indices, blood pressure, dietary intake, and mental health. The following variables will be considered covariates: energy intake, weight changes, and the baseline values of glycemic indices, lipid profile, LPS, inflammatory and oxidative stress biomarkers, adipokines, and parameters related to mental health and the immune system. All variables will be measured at baseline and at the end of the study.

Clinical, para-clinical assessment

Demographic details and current use of medications will be recorded at baseline. Evaluation of patients' physical activity level (PAL) will be done at the onset and end of the study using the International Physical Activity Questionnaire Short-Form (IPAQ-SF) [35]. The Depression, Anxiety, and Stress Scale (DASS) and a general health questionnaire (GHQ) will be used for assessing patient mental health [36, 37]. The DASS questionnaire consists of 14 items that are divided into three subscales to measure depression, anxiety, and stress [38]. Additionally, four other subscales (somatic symptoms, anxiety, insomnia,

social dysfunction, and severe depression) will be assessed using the 28-item GHQ [39]. A 3-day food diary (one for a weekend day and two for 2 non-consecutive weekdays) [40] will be completed by patients at baseline and at the end of the study and applied for dietary assessment. Collected dietary records will be analyzed using the "Nutritionist 4" software (First Databank Inc., Hearst Corp., San Bruno, CA, USA).

At the start and end of the study, weight and height will be measured to the nearest 0.1 kg and 0.1 cm employing a reliable digital scale (Seca, Hamburg, Germany) and a meter mounted on the wall, respectively. A non-elastic tape will be placed on the midpoint between the lowest rib and the upper iliac crest to measure the waist circumference (WC) [41]. The hip circumference (HC) will be recorded at the maximum circumference over the hips without pressing the skin. Neck circumference (NC) will be measured at the level of the mid-cervical spine below the cricoid cartilage, and a minimum circumference will be calculated to the nearest 0.1 cm [42]. The BMI will be derived from weight (kg) divided by the square of the height (m²). All the anthropometric variables will be measured by the same person to minimize measurement errors. Blood pressure will be measured three times by a DinaMap Compact after 30 min of resting, and the mean of the three will be reported.

A venous blood sample (10 mL) will be drawn from all participants following 10 to 12 h overnight fasting and centrifuged for serum isolation at baseline and at week 8 of the trial. FPG, lipid profile, and hs-CRP will be immediately determined after collecting samples. FPG and lipid profiles will be evaluated using the enzymatic colorimetric assay and commercial kits (Pars Azmoon, Tehran, Iran). The LDL will be derived from the Fried Ewald calculation [43]. The remainder of the serum samples will be stored at -70° C until the end of the study. Serum hs-CRP, TNF-α, IL-1, IL-6, IL-10, IL-17, LPS, CML, 8-iso-PGF2a, NO, Trp, KYN, cortisol, GLP-1, RAGE, leptin, adiponectin, ghrelin, BDNF, endothelial cell adhesion molecules, and plasminogen levels will be measured using an enzyme-linked immunosorbent assay (ELISA) kit. Flow cytometry and dual-color reagents will be applied to measure CD4 and CD8 (Dako Company, Denmark). HbA1c A will be determined using a highpressure liquid chromatography D-10 system. Plasma insulin concentrations will be assessed using a chemiluminescent immunoassay method. TAC will be determined through the colorimetric method. Thiobarbituric acid reactive substances (TBARS) assay will be utilized to measure MDA levels using a spectrofluorometer [44]. The reverse transcription polymerase chain reaction (RT-PCR) will be used to determine the altered expression of the PPAR gene. The subsequent formulas will be used to

Saati et al. Trials (2023) 24:35 Page 7 of 10

calculate the oxidative stress index (OSI), the HOMA-IR, and the quantitative insulin sensitivity check index (QUICKI):

OSI =100 \times (TOS/TAC) [45]. Total oxidant status HOMA-IR = [fasting insulin (μ U/mL) \times FPG (mM/L)] / 22.5 QUICKI = 1/ (log (fasting insulin, μ U/ml) + log (FPG, mg/dl)) [46].

Data management

The study progress, protocol, validity, and integrity of the info, also as ethical requirements, will occasionally be supervised by a clinical trial monitor. All participants will be encouraged to answer the questions honestly. To analyze the data of the participants who withdraw from this trial for any reason, the ITT analysis will be used. All participants will be followed up for 8 weeks after the treatment assignment.

Confidentiality

Each participant will be assigned a unique code number. All data will be stored on the drive and available only to the study team. Questionnaire data and consent forms will be stored on paper, remain separate from study data, and be coded with a particular code number.

Statistical analysis

SPSS version 24.0 (SPSS Inc., Chicago, IL, USA) will be applied to analyze the data. Mean \pm standard deviation (SD) or median (25th-75th percentile) and frequency (percentage) will be used to present quantitative and qualitative variables, respectively. The normality of our data will be assessed with the Kolmogorov-Smirnov test. Log transformation in the cases of non-normally distributed data will be done. In order to examine differences in qualitative and quantitative baseline variables between groups, we will apply the chi-square test and the unpaired Student T-test, respectively. To compare quantitative variables between groups, an analysis of covariance (ANCOVA) will be performed post-intervention. For within-group comparison, we will conduct the paired sample Student's T-test or its nonparametric equivalent, the Wilcoxon test. To find the percentage difference, the absolute value of the change will be divided by the average of the values and multiplied by 100 [100 \times (intervention values – placebo values)/placebo values]. P value < 0.05 will be considered statistically significant.

Patient and public involvement

Involving patients in designing, conducting, reporting, or disseminating will not be necessary for this study.

Discussion

This study is a randomized controlled trial to evaluate the usefulness of bitter almond gum as a functional food on the cardio-metabolic, immune-inflammatory, and oxidative stress biomarkers, as well as mental health in women with T2DM. Documented studies suggest the beneficial effects of consuming functional food on macro- and micro-vascular complications of T2DM [47-49]. It is well established that prebiotics exerts wide-ranging impacts on human health and disease. There are numerous studies on the positive effects of prebiotics on cancer, vascular diseases, obesity, and mental disorders [50]. Bitter almond gum, as a possible source of polyphenols and prebiotic fiber, might have anticancer, anti-inflammatory, antioxidant, antilipidemic, antimicrobial, antiviral, and immunomodulatory properties [51, 52]. A randomized study found that 6-week bitter almond gum consumption significantly decreased anthropometric measures and insulin resistance in hyperlipidemic patients [30]. Flavonoids and phenolic acids activate the PPAR gene [53]. PPAR activates the expression of genes encoding lipoprotein lipase (LPL) and apo C-II oxidation [54]. These mechanisms might play a role in the improvement of the lipid profile after bitter almond gum supplementation. On the other hand, bitter almond gum, as a prebiotic, improves glucose tolerance via an increase in GLP-1 secretion [55]. Studies on the effect of bitter almond gum on health are rare. Prebiotic compounds detected in bitter almond gum resemble gum Arabic in being composed of the arabinogalactan polysaccharide [29]. Babiker et al. reported the glucose and lipid-lowering effect of Arabic gum in patients with T2DM [56]. Ahmed et al. revealed that Arabic gum supplementation could downregulate adipose triglyceride lipase (ATGL) and super conserved receptor expressed in brain2 (SREB2) and upregulate hormone-sensitive lipase (HSL) gene expression in the liver of mice fed a high-fat diet [57]. The positive effects of Arabic gum in reducing oxidative stress and inflammation have also been shown [58, 59]. An animal study evaluated the effect of Arabic gum on inflammatory and oxidative stress biomarkers in the gastrointestinal tract of the experimental model of chronic kidney disease (CKD), lower levels of TNF-α, IL-6, transforming growth factor β 1 (TGF- β 1), lipid peroxidation, nitrite, and higher concentrations of IL-10, catalase, glutathione reductase, TAC, SOD, and nuclear factor erythroid 2-related factor 2 reported in the duodenal mucosa [60]. Other antiinflammatory effects attributed to Arabic gum include blockage of the liver macrophage function, modulation of nuclear factor-kB, the maturation of dendritic cells (DCs), and consequent improvement of CD4+ T cell proliferation [61]. Arabic gum may directly scavenge free radicals or ROSs through the presence of various antioxidant Saati et al. Trials (2023) 24:35 Page 8 of 10

compounds or by elevating the synthesis of antioxidant biomolecules [62, 63]. It is possible that Arabic gum can improve the antioxidant capacity due to the presence of amino acid residues such as lysine, tyrosine, and histidine, which show antioxidant properties [64, 65]. This trial is the first randomized, triple-blind controlled study investigating the effects of bitter almond gum consumption on the cardio-metabolic risk factors, oxidative stress, inflammatory biomarkers, LPS, CML, 8-iso-PGF2 α , NO, Trp, KYN, cortisol, GLP-1, RAGE, leptin, adiponectin, ghrelin, BDNF, and mental health in women with T2DM. We hypothesize that bitter almond gum supplementation would improve cardio-metabolic, inflammatory, and oxidative stress indices and mental health through gut microbiota modulation in T2DM women.

Strengths and limitations of the study

The strengths of our trial include using a triple-blind, placebo-controlled, randomized clinical design and also exploring the effects of bitter almond gum supplementation on metabolic parameters, inflammatory markers, and mental health in women with T2DM for the first time. Also, as there is a need to find promising therapeutic agents for managing diabetes and its complications, the application of bitter almond gum might be a natural product to achieve this goal. However, there are some limitations to this research. First, self-reported dietary and physical activity habits may impact the results. Second, there is no compliance biomarker for measuring bitter almond gum intake. Finally, a longer duration of bitter almond gum supplementation might be advisable to find changes in trial biomarkers.

Conclusion

We hope bitter almond gum supplementation will improve metabolic parameters, inflammation, oxidative stress, and mental health in women with T2DM. We expect that the results of this trial will provide scientific evidence in support of bitter almond gum intake for the management of T2DM and its comorbidities.

Trial status

The present protocol is version 1, dated Aug 25, 2022. Recruiting of participants is underway, but the trial has not yet started.

Abbreviations

8-iso-PGF2 α 8-iso-prostaglandin F2 α ATGL Adipose triglyceride lipase

ANCOVA Analysis of covariance

BMI Body mass index

BDNF Brain-derived neurotrophic factor

CML Carboxymethyl lysine CKD Chronic kidney disease CD4 Cluster deference 4
CD8 Cluster deference 8
GLP-1 Glucagon-like peptide 1
DCs Dendritic cells

DASS Depression, Anxiety, and Stress Scale ELISA Enzyme-linked immunosorbent assay

FPG Fasting plasma glucose
GHQ General health questionnaire
HbA1c Glycosylated hemoglobin
HDL High-density lipoproteins
hs-CRP High-sensitivity C-reactive protein

HC Hip circumference

HOMA-IR Homeostasis model assessment insulin resistance

HSL Hormone-sensitive lipase

IKK IkB kinase

IRS Insulin receptor substrate
ITT Intention to treat
IL-1 Interleukin-1
IL-10 Interleukin-10
IL-17 Interleukin-17

IL-6 Interleukin-6

IDF International Diabetes Federation

IPAQ-SF International Physical Activity Questionnaire Short-Form

KYN Kynurenine LPS Lipopolysaccharides LPL Lipoprotein lipase LDI Low-density lipoprotein MDA Malondialdehyde NC Neck circumference NF Nuclear factor NO Nitric oxide

PPAR Peroxisome proliferator-activated receptor

PAL Physical activity level ROS Reactive oxygen species

RT-PCR Reverse transcription polymerase chain reaction AGE Receptor for advanced glycation end products

SPIRIT Standard Protocol Items: Recommendations for Interventional

Trials
TG Triglyceride
TLR4 Toll-like receptor 4
SCFAs Short-chain fatty acids
SD Standard deviation

SREB2 Super conserved receptor expressed in brain2
TBARS Thiobarbituric acid reactive substances

TAC Total antioxidant capacity TC Total cholesterol

TGF- β 1 Transforming growth factor β 1

Trp Tryptophan

TNF-a Tumor necrosis factor-a T2DM Type 2 diabetes mellitus WC Waist circumference

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13063-023-07085-7.

Additional file 1. SPIRIT checklist.

Acknowledgements

The authors would like to acknowledge the patients, their dedication and commitment, and the staff and personnel at the hospital clinic.

Authors' contributions

Each author contributed significantly to the conception and design of the study. PD, MM, and SS conceived and developed the research idea. PD, FAS, and MM reviewed the manuscript, and SS is conducting the research as part of her Master of Science (MS.C) dissertation. The author(s) read and approved the final manuscript.

Saati et al. Trials (2023) 24:35 Page 9 of 10

Funding

This work was supported by the Tabriz University of Medical Science (grant number: 70646).

Availability of data and materials

Related articles will be published with all data generated or analyzed during this study.

Declarations

Ethics approval and consent to participate

The Tabriz University of Medical Sciences ethical committee approved the trial (Ethical Code: IR.TBZMED.REC.1399.726). Participants will be asked to complete an informed consent form (in Persian).

Consent for publication

Not applicable. All the authors take public responsibility for the content of the protocol.

Competing interests

The authors declare that they have no competing interests.

Author details

¹ Student Research Committee, Faculty of Nutrition and Food Science, Tabriz University of Medical Sciences, Tabriz, Iran. ² Nutrition Research Center, Department of Biochemistry and Diet Therapy Faculty of Nutrition and Food Science, Tabriz University of Medical Sciences, Tabriz 5166614711, Iran. ³ Department of Nutrition, School of Health, Arak University of Medical Sciences, Arak, Iran. ⁴ Department of Internal Medicine, School of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran.

Received: 19 November 2022 Accepted: 9 January 2023 Published online: 17 January 2023

References

- Stumvoll M, Goldstein BJ, Van Haeften TW. Type 2 diabetes: principles of pathogenesis and therapy. Lancet. 2005;365(9467):1333–46.
- Galicia-Garcia U, Benito-Vicente A, Jebari S, Larrea-Sebal A, Siddiqi H, Uribe KB, et al. Pathophysiology of type 2 diabetes mellitus. Int J Mol Sci. 2020;21(17):6275.
- Atlas D. International diabetes federation. IDF Diabetes Atlas. 7th ed. Brussels: International Diabetes Federation; 2015. p. 33.
- Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: results from the International Diabetes Federation Diabetes Atlas. Diabetes Res Clin Pract. 2019;157:107843.
- 5. Tinajero MG, Malik VS. An update on the epidemiology of type 2 diabetes: a global perspective. Endocrinol Metab Clin. 2021;50(3):337–55.
- Oguntibeju OO. Type 2 diabetes mellitus, oxidative stress and inflammation: examining the links. Int J Physiol Pathophysiol Pharmacol. 2019;11(3):45.
- Yuan T, Yang T, Chen H, Fu D, Hu Y, Wang J, et al. New insights into oxidative stress and inflammation during diabetes mellitus-accelerated atherosclerosis. Redox Biol. 2019;20:247–60.
- 8. Emmendoerffer A, Hecht M, Boeker T, Mueller M, Heinrich U. Role of inflammation in chemical-induced lung cancer. Toxicol Lett. 2000;112:185–91.
- Costa AD, Garlid KD. Intramitochondrial signaling: interactions among mitoKATP, PKCε, ROS, and MPT. Am J Phys Heart Circ Phys. 2008;295(2):H874–H82.
- Kim J-R, Kim H-N, Song S-W. Associations among inflammation, mental health, and quality of life in adults with metabolic syndrome. Diabetol Metab Syndr. 2018;10(1):1–8.
- 11. Panagi L, Poole L, Steptoe A, Hackett RA. Inflammatory stress responses and future mental health outcomes in people with type 2 diabetes. Brain Behav Immun Health. 2022;23:100472.
- 12. Dona AC, DeLouize AM, Eick G, Thiele E, Salinas Rodriguez A, Manrique Espinoza BS, et al. Inflammation and central adiposity as mediators of

- depression and uncontrolled diabetes in the study on global AGEing and adult health (SAGE). Am J Hum Biol. 2020;32(6):e23413.
- Hajebrahimi B, Kiamanesh A, Farid AA, Asadikaram G. Type 2 diabetes and mental disorders; a plausible link with inflammation. Cell Mol Biol. 2016;62(13):71–7.
- 14. He C, Shan Y, Song W. Targeting gut microbiota as a possible therapy for diabetes. Nutr Res. 2015;35(5):361–7.
- Zhang Y, Zhang H. Microbiota associated with type 2 diabetes and its related complications. Food Sci Human Wellness. 2013;2(3-4):167–72.
- Gurung M, Li Z, You H, Rodrigues R, Jump DB, Morgun A, et al. Role of gut microbiota in type 2 diabetes pathophysiology. EBioMedicine. 2020;51:102590.
- 17. Violi F, Cammisotto V, Bartimoccia S, Pignatelli P, Carnevale R, Nocella C. Gut-derived low-grade endotoxaemia, atherothrombosis and cardiovascular disease. Nat Rev Cardiol. 2022:1–14.
- 18. Zou Y, Song X, Liu N, Sun W, Liu B. Intestinal flora: a potential new regulator of cardiovascular disease. Aging Dis. 2022;13(3):753.
- Han J-L, Lin H-L. Intestinal microbiota and type 2 diabetes: from mechanism insights to therapeutic perspective. World J Gastroenterol: WJG. 2014;20(47):17737.
- Khalid M, Alkaabi J, Khan MA, Adem A. Insulin signal transduction perturbations in insulin resistance. Int J Mol Sci. 2021;22(16):8590.
- Rehman K, Akash MSH. Mechanisms of inflammatory responses and development of insulin resistance: how are they interlinked? J Biomed Sci. 2016;23(1):1–18.
- Gibson GR, Probert HM, Van Loo J, Rastall RA, Roberfroid MB. Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. Nutr Res Rev. 2004;17(2):259–75.
- 23. Gibson GR, Hutkins RW, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, et al. The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. 2017.
- Colantonio AG, Werner SL, Brown M. The effects of prebiotics and substances with prebiotic properties on metabolic and inflammatory biomarkers in individuals with type 2 diabetes mellitus: a systematic review. J Acad Nutr Diet. 2020;120(4):587–607 e2.
- Willcox ML, Elugbaju C, Al-Anbaki M, Lown M, Graz B. Effectiveness of medicinal plants for glycaemic control in type 2 diabetes: an overview of meta-analyses of clinical trials. Front Pharmacol. 2021;12:777561.
- Kooti W, Farokhipour M, Asadzadeh Z, Ashtary-Larky D, Asadi-Samani M.
 The role of medicinal plants in the treatment of diabetes: a systematic review. Electron Physician. 2016;8(1):1832.
- 27. Gomaa EZ. In vitro antioxidant, antimicrobial, and antitumor activities of bitter almond and sweet apricot (Prunus armeniaca L.) kernels. Food Sci Biotechnol. 2013;22(2):455–63.
- 28. Rasekhi Kazeruni A, Hosseini E. Effect of bitter almond gum (Amygdalus scoparia Spach) on the survival of Lactobacillus acidophilus La5 in tomato juice during refrigeration storage and exposure to simulated gastric juice. J Maz Univ Med Sci. 2017;27(147):75–86.
- Fadavi G, Mohammadifar MA, Zargarran A, Mortazavian AM, Komeili R. Composition and physicochemical properties of Zedo gum exudates from Amygdalus scoparia. Carbohydr Polym. 2014;101:1074–80.
- Chahibakhsh N, Hosseini E, Rahbar AR. Bitter almond gum reduces body mass index, serum triglyceride, hyperinsulinemia and insulin resistance in overweight subjects with hyperlipidemia. J Funct Foods. 2019;55:343–51.
- Chan A-W, Tetzlaff JM, Gøtzsche PC, Altman DG, Mann H, Berlin JA, et al. SPIRIT 2013 explanation and elaboration: guidance for protocols of clinical trials. BMJ. 2013;346.
- 32. Kerner W, Brückel J. Definition, classification and diagnosis of diabetes mellitus. Exp Clin Endocrinol Diabetes. 2014;122(07):384–6.
- Evert AB, Boucher JL, Cypress M, Dunbar SA, Franz MJ, Mayer-Davis EJ, et al. Nutrition therapy recommendations for the management of adults with diabetes. Diabetes Care. 2014;37(Supplement_1):S120–S43.
- 34. Sheu WH-H, Lee I-T, Chen W, Chan Y-C. Effects of xylooligosaccharides in type 2 diabetes mellitus. J Nutr Sci Vitaminol. 2008;54(5):396–401.
- Craig C, Marshall A, Sjostrom M, Bauman A, Lee P, Macfarlane D, et al. International physical activity questionnaire-short form. J Am Coll Heal. 2017;65(7):492–501.
- Parkitny L, McAuley J. The depression anxiety stress scale (DASS). J Physiother. 2010;56(3):204.

Saati et al. Trials (2023) 24:35 Page 10 of 10

- Montazeri A, Harirchi AM, Shariati M, Garmaroudi G, Ebadi M, Fateh A. The 12-item General Health Questionnaire (GHQ-12): translation and validation study of the Iranian version. Health Qual Life Outcomes. 2003;1(1):1–4.
- 38. Sahebi A, Asghari MJ, Salari RS. Validation of depression anxiety and stress scale (DASS-21) for an Iranian population; 2005.
- Yaghubi H, Karimi M, Omidi A. Validity and factor structure of the General Health Questionnaire (GHQ-12) in university students. Int J Behav Sci. 2012;6(2).
- 40. Yang YJ, Kim MK, Hwang SH, Ahn Y, Shim JE, Kim DH. Relative validities of 3-day food records and the food frequency questionnaire. Nutr Res Pract. 2010;4(2):142–8.
- Ness-Abramof R, Apovian CM. Waist circumference measurement in clinical practice. Nutr Clin Pract. 2008;23(4):397–404.
- 42. Joshipura K, Muñoz-Torres F, Vergara J, Palacios C, Pérez CM. Neck circumference may be a better alternative to standard anthropometric measures. J Diabetes Res. 2016;2016:6058916.
- 43. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem. 1972;18(6):499–502.
- Papastergiadis A, Mubiru E, Van Langenhove H, De Meulenaer B. Malondialdehyde measurement in oxidized foods: evaluation of the spectrophotometric thiobarbituric acid reactive substances (TBARS) test in various foods. J Agric Food Chem. 2012;60(38):9589–94.
- Abuelo A, Hernández J, Benedito JL, Castillo C. Oxidative stress index (OSi) as a new tool to assess redox status in dairy cattle during the transition period. Animal. 2013;7(8):1374–8.
- Holzinger U, Kitzberger R, Fuhrmann V, Funk G-C, Madl C, Ratheiser K. Correlation of calculated indices of insulin resistance (QUICKI and HOMA) with the euglycaemic hyperinsulinaemic clamp technique for evaluating insulin resistance in critically ill patients. Eur J Anaesthesiol. 2007;24(11):966–70.
- Mirmiran P, Bahadoran Z, Azizi F. Functional foods-based diet as a novel dietary approach for management of type 2 diabetes and its complications: a review. World J Diabetes. 2014;5(3):267.
- 48. Alkhatib A, Tsang C, Tiss A, Bahorun T, Arefanian H, Barake R, et al. Functional foods and lifestyle approaches for diabetes prevention and management. Nutrients. 2017;9(12):1310.
- 49. Monjiote DP, Leo EEM, Campos MRS. Functional and biological potential of bioactive compounds in foods for the dietary treatment of type 2 diabetes mellitus. In: Functional food—Improve health through adequate food; 2017. p. 143–63.
- Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi SJ, et al. Prebiotics: definition, types, sources, mechanisms, and clinical applications. Foods. 2019;8(3):92.
- 51. Patel AK, Singhania RR, Awasthi MK, Varjani S, Bhatia SK, Tsai M-L, et al. Emerging prospects of macro-and microalgae as prebiotic. Microb Cell Factories. 2021;20(1):1–16.
- 52. Cory H, Passarelli S, Szeto J, Tamez M, Mattei J. The role of polyphenols in human health and food systems: a mini-review. Front Nutr. 2018;5:87.
- Liu L, Shan S, Zhang K, Ning ZQ, Lu XP, Cheng YY. Naringenin and hesperetin, two flavonoids derived from Citrus aurantium up-regulate transcription of adiponectin. Phytother Res. 2008;22(10):1400–3.
- Auwerx J, Schoonjans K, Fruchart J-C, Staels B. Regulation of triglyceride metabolism by PPARs: fibrates and thiazolidinediones have distinct effects. J Atheroscler Thromb. 1996;3(2):81–9.
- Kellow NJ, Coughlan MT, Reid CM. Metabolic benefits of dietary prebiotics in human subjects: a systematic review of randomised controlled trials. Br J Nutr. 2014;111(7):1147–61.
- Babiker R, Elmusharaf K, Keogh MB, Banaga AS, Saeed AM. Metabolic effect of gum Arabic (Acacia Senegal) in patients with type 2 diabetes mellitus (T2DM): randomized, placebo controlled double blind trial. Funct Foods Health Dis. 2017;7(3):222–34.
- Ahmed AA, Musa HH, Fedail JS, Sifaldin AZ, Musa TH. Gum arabic suppressed diet-induced obesity by alteration the expression of mRNA levels of genes involved in lipid metabolism in mouse liver. Bioact Carbohydr Diet Fibre. 2016;7(1):15–20.
- Ali NE, Kaddam LA, Alkarib SY, Kaballo BG, Khalid SA, Higawee A, et al. Gum arabic (Acacia Senegal) augmented total antioxidant capacity and reduced C-reactive protein among haemodialysis patients in Phase II trial. Int J Nephrol. 2020;2020.

- Kamal E, Kaddam LA, Dahawi M, Osman M, Salih MA, Alagib A, et al. Gum arabic fibers decreased inflammatory markers and disease severity score among rheumatoid arthritis patients, phase II trial. Int J Rheumatol. 2018;2018:4197537.
- Ali BH, Za'abi A, Al Suleimani Y, Manoj P, Ali H, Ribeiro DA, et al. Gum arabic reduces inflammation, oxidative, and nitrosative stress in the gastrointestinal tract of mice with chronic kidney disease. Naunyn Schmiedeberg's Arch Pharmacol. 2020;393(8):1427–36.
- 61. Kaddam L, Fdl-Elmula I, Saeed A. Gum Arabic beneficial effects, clinical applications, and future prospective. Gum Arabic: Elsevier; 2018. p. 211–20.
- 62. Ali BH, Ziada A, Blunden G. Biological effects of gum arabic: a review of some recent research. Food Chem Toxicol. 2009;47(1):1–8.
- Kong H, Yang J, Zhang Y, Fang Y, Nishinari K, Phillips GO. Synthesis and antioxidant properties of gum arabic-stabilized selenium nanoparticles. Int J Biol Macromol. 2014;65:155–62.
- 64. Marcuse R. Antioxidative effect of amino-acids. Nature. 1960;186(4728):886–7.
- Park EY, Murakami H, Matsumura Y. Effects of the addition of amino acids and peptides on lipid oxidation in a powdery model system. J Agric Food Chem. 2005;53(21):8334–41.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

