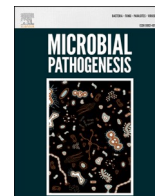




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Benzimidazole compound abrogates SARS-COV-2 receptor-binding domain (RBD)/ACE2 interaction *In vitro*

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ABSTRACT

The development of clinically actionable pharmaceuticals against coronavirus disease (COVID-19); an infectious disease caused by the SARS-CoV-2 virus is very important for ending the pandemic. Coronavirus spike glycoprotein (GP)-Receptor Binding Domain (RBD) and its interaction with host receptor angiotensin converting enzyme 2 (ACE2) is one of the most structurally understood but therapeutically untapped aspect of COVID-19 pathogenesis. Binding interface based on previous x-ray structure of RBD/ACE2 were virtually screened to identify fragments with high-binding score from 12,000 chemical building blocks. The hit compound was subjected to fingerprint-based similarity search to identify compounds within the FDA-approved drug library containing the same core scaffold. Identified compounds were then re-docked into of RBD/ACE2. The best ranked compound was validated for RBD/ACE2 inhibition using commercial kit. Molecular dynamics simulation was conducted to provide further insight into the mechanism of inhibition. From the original 12000 chemical building blocks, benzimidazole (BAZ) scaffold was identified. Fingerprint-based similarity search of the FDA-approved drug library for BAZ-containing compounds identified 12 drugs with the benzimidazole-like substructure. When these compounds were re-docked into GP/ACE2 interface, the consensus docking identified bazedoxifene as the hit. *In vitro* RBD/ACE2 inhibition kinetics showed micromolar IC₅₀ value (1.237 μM) in the presence of bazedoxifene. Molecular dynamics simulation of RBD/ACE2 in the presence BAZ resulted in loss of contact and specific hydrogen-bond interaction required for RBD/ACE2 stability. Taken together, these findings identified benzimidazole scaffold as a building block for developing novel RBD/ACE2 complex inhibitor and provided mechanistic basis for the use of bazedoxifene as a repurposable drug for the treatment of COVID-19 acting at RBD/ACE2 interface.

1. Introduction

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the causative agent of coronavirus disease 2019 (COVID-19); a pneumonia-like illness responsible for the COVID-19 pandemic infecting

approximately 25 million people and causing more than 835,000 deaths (Johns Hopkins, Coronavirus resource center) and counting [1,2]. Two general approaches have been adopted for treating COVID-19 cases; vaccine and therapeutics development.

Small molecules such as remdesivir, hydroxychloroquine, ribavirin,

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and ritonavir were initially repurposed as therapeutics for COVID-19 [3] but have not performed satisfactorily clinically. New drugs specifically developed for COVID-19 by Merck (molnupiravir), Pfizer (ritonavir in combination with PF-07321332) [4] have been adjudged more efficacious [5]. Research efforts at developing plant-based therapeutic agents is also worthy of mention [6].

Although comparatively, vaccines have been more successful in combating COVID-19 than small molecular weight drugs within clinical settings, this success has been threatened by the fast-rising cases of mutations [7]. Thus, making discovery and development of novel anti-COVID-19 agents highly desirable in the long run.

To this end, 12,000 chemical building blocks were computationally screened to identify possible virus entry inhibitor candidates acting at the spike glycoprotein-angiotensin (RBD) converting enzyme II (ACE2) interface [8]. Whilst most of the previous successful efforts have targeted SARS-CoV-2 proteases [9], the current strategy provides a complementary approach such that a successful candidate will potentially reduce virus tissue reservoir as the distribution of ACE2 has been confirmed in most cells with the exception of matured erythrocytes [10]. Indeed, Pfizer/BioNTech's BNT162b2 and Moderna's mRNA-1273 platforms drive bioactivity through SARS-CoV-2 spike glycoprotein interaction with host (human) ACE2 receptor to gain entry into a cell to initiate tissue infection [11]. SARS-CoV-2 spike glycoprotein [12] is indeed a hot spot for mutation which clearly threatens vaccine efficacy. A few other research is also targeting RBD/ACE2 interface, of note is the discovery of bromelain [13], (–)-piperidine, 2-(*p*-hydroxybenzyl)benzofuran-6-ol, 1-(4-hydroxy-3-methoxyphenyl)-2-4-[(E)-3-hydroxy-1-propanyl]-2-methoxyphenoxy-1,3-propanediol, and Rhein [14].

This study therefore identified benzimidazole fragment as particularly interesting, and when compounds bearing benzimidazole scaffold were searched from FDA-approved library, we have identified bazedoxifene as the most likely candidate inhibitor of RBD/ACE2 interaction through molecular docking, atomistic simulation and validation by inhibitory kinetics studies. The previous reports that benzimidazole is a potent entry inhibitor to hepatitis C virus [15] somewhat improves our overall confidence that bazedoxifene may represent a repurposable anti-COVID-19 agent.

2. Materials and methods

2.1. 2D fingerprint search, ligand library, and protein preparation

The database of FDA-approved drugs (open.fda.gov) were retrieved and filtered (Datawarrior Suite) using the molecular weight cutoff $200 \leq 700$ g/mol. The residual compounds were searched (open babel) for benzimidazole substructure using a path-based fingerprint (FP2). Each compound was scored using the Tanimoto coefficient and plotted as a function of the population size using Graphed prism.

Unless otherwise stated, all ligands for molecular docking were incorporated into Maestro and prepared using the Schrodinger suite version 2018–1. Employing Ligprep [16], Epik [17] with OPLS3 force field for protonation, stereoisomerization, tautomers generation, and to achieve biological conformers. Energy minimization was achieved for all tautomeric states at a pH of 7 ± 2 . 2019-nCoV RBD/ACE2 Complex (2.9 Å) was retrieved from PDB [18] with the PDB ID 6M17 [19]. The complex was prepared using the protein preparation wizard module in maestro 11.5 by default settings. The complex was optimized and then minimized using the OPLS3 force field by converging heavy atoms to RMSD of 0.3 Å. The complex interaction interface for docking was generated with the receptor grid generation tool in maestro 11.5. The interaction interface between RBD/ACE2 complex was defined by amino acid residues: K417/Q409/R403/D405/R408/Y505 (RBD) and D30/N33/T324/F356/(ACE2), with van der Waals scaling factor 1.00 and charge cutoff of 0.25 around the complex interaction interface residues.

2.2. Molecular docking

Molecular docking was conducted using Autodock Vina [20], and Glide module on maestro 11.5 [21]. The library of compounds was docked into the interface of the target using the standard precision (SP) and Extra precision (XP) algorithm [22], applying a scaling factor of 0.8 and partial charge cutoff of 0.15, the ligand was handled as flexible. 30 Å in all directions of the interface between RBD/ACE2 complex defined by amino acid residues: K417/Q409/R403/D405/R408/Y505 (RBD) and D30/N33/T324/F356/(ACE2) was used as dock grid dimension. Lastly, the binding affinity of the receptor-ligand complex was ranked according to Glide score (SP and XP) while Vina score was utilized for the Autodock vina program.

2.3. Molecular dynamics simulation

2.3.1. Biosystems setup for atomistic simulation

The input files for MD simulation for each of the biosystems (apo-RBD/ACE2, BAZ-RBD/ACE2; ACE2 = aa21-559; RBD = aa336-518) was generated using CHARMM-GUI webserver (www.charmm-gui.org). BAZ parametrization was performed using ParamChem service (<https://cgenff.paramchem.org>) as implemented on CHARMM-GUI webserver. During biosystem build-up, protein and ligand atoms were defined by CHARMM36 all-atom additive force field parameters, solvated in TIP3P explicit water model and neutralized with Na^+/Cl^- at 0.15 M [23]. Details of biosystem setup using CHARMM-GUI have been previously published [24–26].

Molecular dynamics simulation was run on GROMACS (ver. 5) [27] software. During equilibration, the biosystems were subjected to constant pressure and temperature (NPT; 310K, 1 bar) conditions using Berendsen temperature and pressure coupling algorithms as implemented in GROMACS. van der Waals interactions were estimated at 10 Å, long-range electrostatic interactions were computed using particle mesh Ewald (PME) summation scheme while equation of atomic motion was integrated using the leap-frog algorithm at 2 fs time step for a total time of 50 ns with positional restraints imposed on the heavy atoms in all directions. Production simulations were performed at 100 ns with the removal of atomic restraints. All calculations were performed on Super-Micro workstations (32-E2600 Intel Xeon CPUs, M6000 GPUs Accelerator PCI-E $\times 16$ Card/node) housed at Bio-Computing Research Unit (B-cRU), Mols and Sims, Ado Ekiti, Ekiti State, Nigeria.

2.3.2. Post-simulation trajectory analysis and MMPBSA calculation

Unless otherwise stated, 3D atomic representations were drawn using Visual Molecular Dynamics (VMD) software [28]. Interatomic distance was calculated using *g_dist* tool inbuilt in GROMACS software. Population plots from distance values were plotted using GraphPad-Prism software. Binding free energy was calculated GROMACS Tool for high-throughput MM-PBSA calculations (*g_mmpbsa*) algorithm as described [29]. All line graphs were plotted as mean of 2 independent runs using GraphPad prism (ver 6.0e, 2014).

2.4. RBD/ACE2 complex inhibition kinetics

Assessment of SARS-CoV2 RBD and hACE2 binding inhibition by Bazedoxifene (Cat: HY-A0031/CS-0932, Lot: 58414) was performed using COVID-19 Spike-ACE2 binding assay kit (RayBiotech, Inc, Cat #: CoV-SACE2-1, <https://www.raybiotech.com/covid-19-spike-ace2-binding-assay-kit/>) following the manufacturer's protocol) Bazedoxifene was dissolved in water and aliquoted at concentrations 0.01, 0.03, 0.1, 0.3, 1.0, 3.0 mM in triplicates for the study. The SARS-CoV2 RBD and hACE2 binding inhibitory capacity of each concentration was assessed in triplicate. Compounds at noted concentration were mixed with recombinant hACE2 protein or PBS (Control), added to ELISA plate pre-coated with recombinant SARS-Cov2 S-protein RBD and incubated overnight at 4 °C with shaking. Unbound ACE2 was removed by

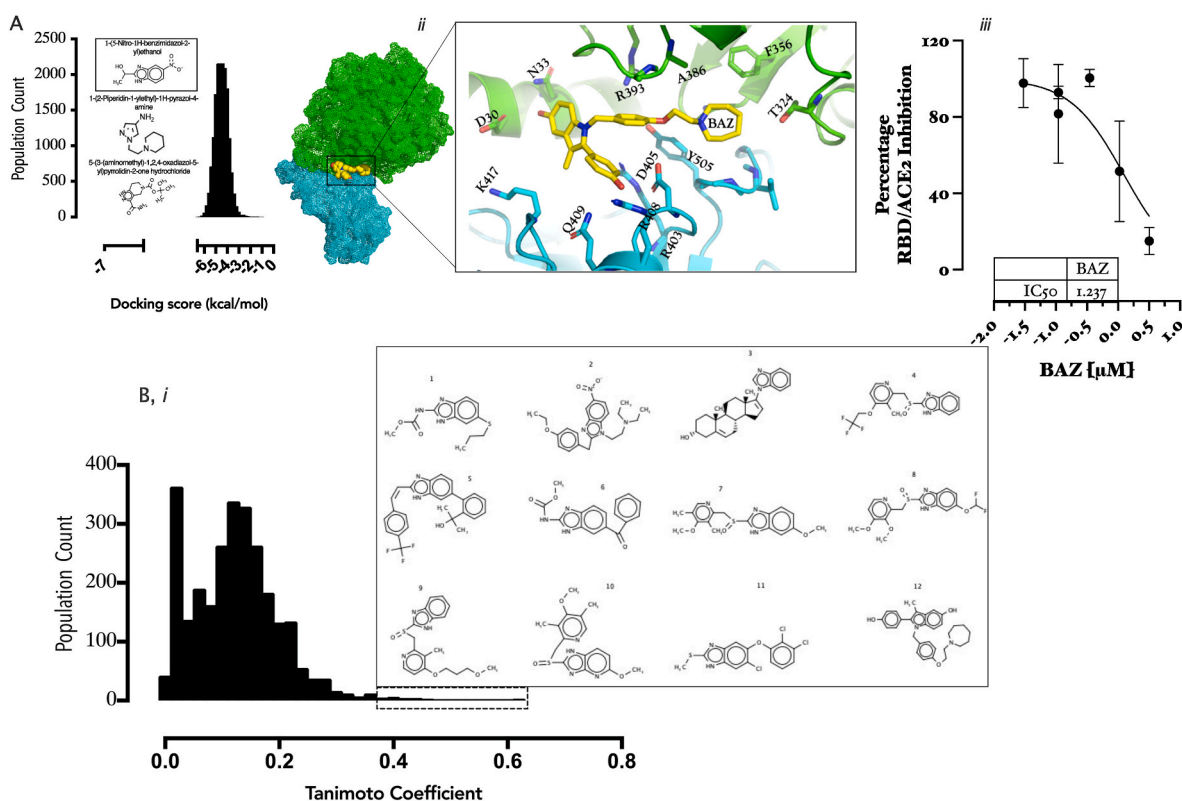


Fig. 1. (A) Population count of docking score distribution of building block into RBD/ACE-2 interface; the inset show the best 3 ranked compounds 2D structures. **Fig. 1B:** (i) Population count of FDA-approved drugs with benzimidazole scaffold based on Tanimoto coefficient scores (inset shows 12 compounds with benzimidazole substructure). (ii) A snapshot of the placement of bazedoxifene (12) within the RBD/ACE2 interface and a detailed projection of the amino acids contributing to the binding interaction. **Fig. 1B.** (iii) Dose-response curve fitting of RBD/ACE2 binding in the presence of graded concentration of bazedoxifene (BAZ).

washing, and binding was assessed based on anti-ACE2 antibody- HRP-conjugated anti-goat IgG reaction with 3,3',5,5'-tetramethylbenzidine (TMB). Absorbance at 450 nm was measured with a ELISA microplate reader. Dose-response graph was plotted using GraphPad Prism and IC50 value was calculated.

3. Results

3.1. Virtual screening identifies benzimidazole fragment as high-affinity binder at RBD/ACE2 interface

Fragment-based lead discovery (FBLD) represents an efficient approach to drug discovery, taking advantage of the highly diverse fragment libraries (12,000 compounds) available at the Chem Bridge Corporation (<https://www.chembridge.com>), a virtual screening performed to identify potential hits at the RBD/ACE2 interface. Whilst most of the fragments had docking scores between -1.0 and -6.0 kcal/mol, three compounds namely: 1-(5-Nitro-1H-benzimidazol-2-yl)ethanol, 1-(2-piperidin-1-ylethyl)-1H-pyrazol-4-amine and 5-(3-(aminoethyl)-1,2,4-oxadiazol-5-yl)pyrrolidine-2-one have docking scores of -7.0 kcal/mol (Fig. 1A). Although, -7.0 kcal/mol docking score is weak, instead of subsequent growing and/or combining the lead fragments to produce other leads with a higher affinity and improved physicochemical properties, we rather screened the FDA-library for chemical compounds containing benzimidazole fragment as 1-(5-Nitro-1H-benzimidazol-2-yl)ethanol had the best docking score overall.

2D fingerprint search was conducted into the database of FDA-approved drugs to identify and rank (Tanimoto coefficient) compounds with benzimidazole scaffold which can be rapidly repurposed as entry inhibitors in the current pandemic. The best ranked compounds were twelve (12) including albendazole(1), etonitazene(2), galeterone (3), lansoprazole(4), mavatrep(5), mebendazole(6), omeprazole(7),

Table 1.0

Compounds with high Tanimoto coefficient with benzimidazole.

Compounds	Vina Dock	Glide XP	Glide SP	Mean Score
Albendazole (1)	-5.20	-2.84	-4.27	-4.10
etonitazene (2)	-6.00	-2.48	-4.78	-4.42
galeterone (3)	-6.80	-2.52	-4.03	-4.45
lansoprazole (4)	-6.30	-5.23	-5.83	-5.79
mavatrep (5)	-8.00	-3.81	-5.55	-5.79
mebendazole (6)	-6.20	-3.41	-3.85	-4.49
omeprazole (7)	-5.40	-5.48	-5.27	-5.38
pantoprazole (8)	-6.50	-3.94	-5.81	-5.42
rabeprazole (9)	-5.70	-5.74	-5.05	-5.50
tenatoprazole (10)	-6.00	-4.03	-3.91	-4.65
triclabendazole (11)	-5.70	-4.18	-4.69	-4.86
Bazedoxifene* (12)	-7.40	-5.13	-6.20	-6.24

pantoprazole(8), rabeprazole(9), tenatoprazole(10), triclabendazole (11), and bazedoxifene(12) (Fig. 1B, i, Table 1). When these compounds were docked into RBD/ACE2 interface using consensus scoring method [30], bazedoxifene (BAZ) was identified as exhibiting the lowest binding score (-6.24 kcal/mol) (Supplementary Table 1.0). An overview of the binding position within the RBD/ACE2 intercalates between a network of aromatic and charged amino acids at the RBD/ACE2 interface; some of the key residues proximal to BAZ binding pocket include: D30/N33/T324/F356/(ACE2) and K417/Q409/R403/D405/R408/Y505 (RBD) (Fig. 1B, ii). These amino acids have identified as key to successful initiation of cellular invasion [31]. When the finding was validated in SARS-COV-2 spike-ACE2 binding assay, a mean IC50 value of 1.237 μ M was obtained (Fig. 1B, iii) thus, strongly suggesting that BAZ is a micromolar level inhibitor of RBD/ACE2 interaction and a potential drug in the current pandemic.

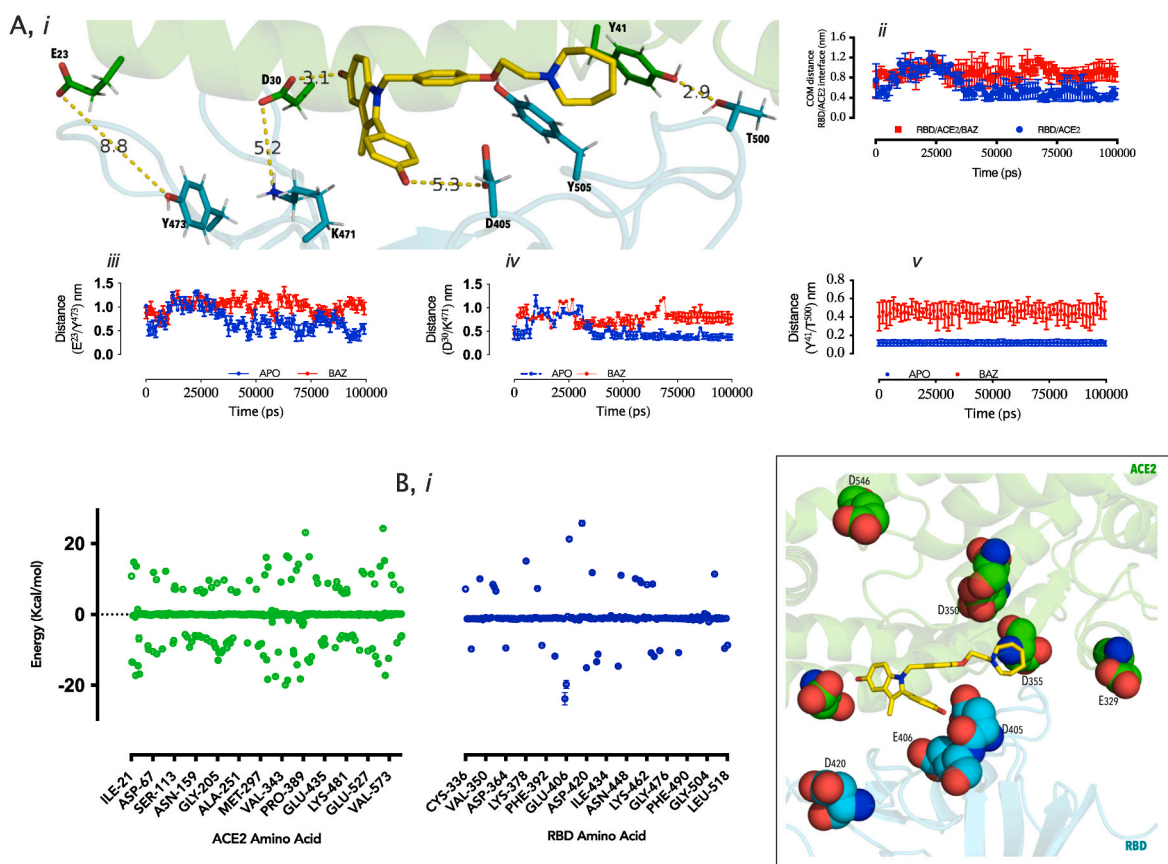


Fig. 2. (I) 3D representation of selected hydrogen-bond interaction at RBD (blue) and ACE2 (green) interface proximal to bazedoxifene (yellow stick) binding site. (ii-v) Time-dependent distance plots of RBD/ACE2, and selected residue pairs (see Y-axes). (D). (i) Energy contribution of amino acids (ACE2 = green points, RBD = blue points) to bazedoxifene binding based on MMPBSA energy calculations; a 3D projection of the most contributing amino acids based on 14.0 kJ/mol energy cutoff.

3.2. Molecular dynamics simulation identified the atomistic basis for RBD/ACE2 inhibition

Molecular dynamics (MD) simulation has become a routine *in silico* tools for drug discovery due to its key advantage of incorporating structural flexibility and entropic effects into target-drug interaction, which is limited in molecular docking. Thus, MD simulation is more accurate when monitoring atomistic motions and kinetics associated with drug–target recognition and binding [32]. Since our group has previous experience in deploying MD simulation for delineating mechanism of drug–target interaction in complexes such as lysophosphatidate-LPA1 [26], nefiracetam-NMDA receptors [33], we therefore deployed MD simulation for the current studies in order to provide better insight into the mechanism of inhibition.

First, investigation was conducted into the center of mass distance between residues constituting the interface (Fig. 2A, i) showed that at approximately 40 ns, the distance reduced to 0.4 nm in apo state, indicating contact establishment; the presence of BAZ resulted in loss of contact with mean distance 0.8–1.2 nm starting at 40 ns till the end of the simulation. (Fig. 2A, ii). Three amino acid pairs were further studied providing insight into loss of contact. Glu23/Tyr473 (Fig. 2A, iii) and Asp30/Lys471 (Fig. 2A, iv) distance evolution with time strongly

suggest loss of contact around 40 ns but Tyr41/T500 had already contact post equilibration in the presence of BAZ (Fig. 2A, v). Clearly, the presence of BAZ within the RBD/ACE2 interface promotes loss of contact and possibly dissociation as suggested by the *in vitro* data. Lastly, the details of the amino acid network required for BAZ binding and the energy of binding were worked out using MMPBSA method [29]. Several amino acids at the RBD/ACE2 interface participate in BAZ binding (Fig. 2B, i) but in terms of contribution, Glu329/Asp355/Asp350/D546 (ACE2) and Asp405/Glu406 and Asp420 (ACE2) have the most contribution using the energy -14.0 kJ/mol energy cutoff, while the binding energy was estimated at -409.178 ± 21.114 kJ/mol (Table 2) These data strongly suggest high affinity of RBD/ACE2 interface for BAZ.

4. Discussion

In addition to the challenges of developing and distributing safe and effective SARS-CoV-2 (COVID-19) vaccines [34], evidence for the presence of variants of concern (VOC) now mounts [35]. These VOCs in most cases accumulate mutations around the epitopes; causing viral escape or highly diminished vaccine efficacy [36].

It is therefore not unimaginable that cheaper, more effective therapeutic options must be investigated. One of the key successes here is the

Table 2. MMPBSA Energy Estimation of Bazedoxifene at RBD/ACE2 interface.

Compound	Energy Terms (Kj/mol)				
BAZ	van der Waal energy	Electrostatic energy	Polar solvation energy	SASA energy	Binding energy
	-128.110 ± 20.285	-431.479 ± 18.550	167.395 ± 39.644	-16.984 ± 2.664	-409.178 ± 21.114

Pfizer-developed PF-07321332⁴. PF-07321332 is an orally bioavailable SARS-CoV-2 main protease inhibitor with excellent off-target selectivity and *in vivo* safety profile [5].

In current study, Fragment-based lead discovery-aided fingerprint-based similarity search [37] of the FDA-approved chemical library identified benzimidazole scaffold-containing compounds as potential inhibitor of RBD/ACE2 interaction. Indeed, some previous reports have identified benzimidazole scaffold-containing compounds as potent anti-viral agents [38,39]. For instance, linear (dialkylamino)alkyl-derivatized 2-[(benzotriazol-1/2-yl)methyl]benzimidazoles exhibited nanomolar EC(50) activity on respiratory syncytial virus (RSV) [40].

The FDA-approved drug with the best docking result in this study is bazedoxifene. Bazedoxifene is a selective estrogen receptor modulator indicated for endometriosis [41]. It has also been indicated for postmenopausal osteoporosis [42]. The anti-SARS-CoV-2 activity of bazedoxifene has been mentioned in some prior reports. For instance, in a study, it was demonstrated that bazedoxifene inhibits IL-6 signaling at therapeutic doses, leading to blockage of cytokine storm, ARDS and mortality in severe COVID-19 patients [43]. In another report, anti-SARS-CoV-2 activity with IC50 value of 3.44 μ M, and CC50 value of 14.97 had been reported [44]. Here, we provide a mechanistic insight that bazedoxifene inhibits RBD/ACE2 complex. It is worthy of note that the computational data obtained by Yele et al. [45] and Mohapatra et al. [46] lend credence to our findings. Mudi et al. [47] also reported that benzimidazole preferentially inhibits SARS-CoV-2-plagued VeroE6 cells while proposing main protease and non-structural as potential targets based on molecular docking and MD simulation studies. A further study by this group also confirmed that SARS-CoV-2 is susceptible to five-membered heterocycle-derivatives of benzimidazoles [48]. Indeed, *in vitro* conformation of bazedoxifene will encourage validation of gonadorelin, fondaparinux and atorvastatin which have been recently identified as RBD/ACE2 inhibitors through similar computational methods [13].

In conclusion, we have provided further evidence in support of bazedoxifene as a repurposable drug for the treatment of COVID-19 acting at RBD/ACE2 interface. These findings are of immediate importance as vaccine production, distribution and efficacy present a challenge in the global flight against COVID-19.

Ethics approval and consent to participate

The current study does not involve the use of human or animal subjects, therefore, ethical approval or consent to participate is not applicable.

Consent for publication

All authors have consented to publish this manuscript.

Authors' contributions

Authors OO, and SF and ON conceived and designed the study. Authors OMO and BO performed molecular docking and compiled the manuscript. OS and AI performed the kinetic assay studies. All authors prepared the final manuscript, and proofreading.

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Availability of data and material

All materials including primary data relating to this study will be made available upon request.

CRedit authorship contribution statement

Olaposi Omotuyi: Writing – original draft, Methodology, Funding acquisition, Conceptualization. **Olusina M. Olatunji:** Validation. **Oyekanmi Nash:** Conceptualization. **Babatunji Oyinloye:** Formal analysis. **Opeyemi Soremekun:** Writing – review & editing, Writing – original draft, Investigation. **Ayodeji Ijagbuji:** Visualization, Validation. **Segun Fatumo:** Supervision, Investigation, Conceptualization.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.micpath.2023.105994>.

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