

# Check for updates REVIEW ARTICLE **OPEN** NEDD8-conjugating enzyme E2s: critical targets for c[an](http://crossmark.crossref.org/dialog/?doi=10.1038/s41420-023-01337-w&domain=pdf)cer therapy

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NEDD8-conjugating enzymes, E2s, include the well-studied ubiquitin-conjugating enzyme E2 M (UBE2M) and the poorly characterized ubiquitin-conjugating enzyme E2 F (UBE2F). UBE2M and UBE2F have distinct and prominent roles in catalyzing the neddylation of Cullin or non-Cullin substrates. These enzymes are overexpressed in various malignancies, conferring a worse overall survival. Targeting UBE2M to influence tumor growth by either modulating several biological responses of tumor cells (such as DNA-damage response, apoptosis, or senescence) or regulating the anti-tumor immunity holds strong therapeutic potential. Multiple inhibitors that target the interaction between UBE2M and defective cullin neddylation protein 1 (DCN1), a co-E3 for neddylation, exhibit promising anti-tumor effects. By contrast, the potential benefits of targeting UBE2F are still to be explored. It is currently reported to inhibit apoptosis and then induce cell growth; hence, targeting UBE2F serves as an effective chemo-/ radiosensitizing strategy by triggering apoptosis. This review highlights the most recent advances in the roles of UBE2M and UBE2F in tumor progression, indicating these E2s as two promising anti-tumor targets.

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## FACTS

- More research is being devoted to finding specific inhibitors against neddylation E2s to address the limitations of neddylation E1 inhibitor MLN4924.
- UBE2M and UBE2F are overexpressed in various malignancies, conferring a worse overall survival.
- Targeting UBE2M to influence tumor growth by either modulating several biological responses of tumor cells (such as DNA-damage response, apoptosis, or senescence) or regulating the anti-tumor immunity holds strong therapeutic potential.
- A few inhibitors targeting UBE2M-DCN1 interaction have been identified.
- UBE2F appears to be promising as both an anti-tumor target and a chemo-/radiosensitizing target.

#### OPEN QUESTIONS

- The induction of drug-resistant mutations in UBA3 inhibits the formation of the MLN4924-NEDD8 adduct, and hence there is a need to explore possible alternative targets against the neddylation pathway.
- How do the two E2s regulate anti-tumor immunity? In-depth elucidation of the underlying mechanisms may provide vital cues for targeting UBE2M and UBE2F.
- Deciphering the cellular potency of these UBE2M-DCN1 inhibitors compared to MLN4924?
- Whether these UBE2M-DCN1 inhibitors have any biological function?
- Whether to promote clinical trials of these E2s inhibitors alone or in combination?

#### INTRODUCTION

Protein neddylation involves the conjugation of a ubiquitin-like molecule known as neuronal precursor cell-expressed developmentally down-regulated protein 8 (NEDD8) to the lysine residue of targeted substrate proteins [\[1,](#page-5-0) [2](#page-5-0)]. Like ubiquitin, NEDD8 binds to substrates by forming an isopeptide chain between its C-terminal glycine residue (Gly76) and a lysine residue on targeted proteins. In the initial step of the reaction, NEDD8 is produced as a precursor containing five additional residues downstream from Gly76 cleaved by the C-terminal hydrolases  $[3, 4]$  $[3, 4]$  $[3, 4]$  $[3, 4]$  $[3, 4]$ . The next step involves activation of the mature NEDD8 in an ATP-dependent manner by the NEDD8-activating enzyme (NAE) E1, a heterodimer composed of NAE1 and ubiquitin-like modifier activating enzyme 3 (UBA3) [[5](#page-6-0), [6](#page-6-0)]. After that, a trans-thiolation process occurs during which the NEDD8-loaded NAE is transferred to either UBE2M, also known as UBC12, or UBE2F (two NEDD8-conjugating enzyme E2s) [\[7](#page-6-0)–[9](#page-6-0)]. Finally, covalent attachment drives a substrate-specific E3 ligase, such as defective in cullin neddylation 1 (DCN1) and RINGbox protein 1/2 (RBX1/2), to transfer NEDD8 from the charged E2 to a lysine residue in its target protein (Fig. [1\)](#page-1-0) [\[10](#page-6-0)–[14](#page-6-0)]. The best characterized principal substrates of neddylation are the cullin

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Fig. 1 The process of protein neddylation. Neddylation is a process that conjugates NEDD8 to cullins or non-cullin substrates via a three-step reaction, catalyzed by NEDD8-activating enzyme E1 (a heterodimer of NAE1 and UBA3), NEDD8-conjugating enzyme E2 (UBE2M or UBE2F) and substrate-specific NEDD8-E3 ligases (e.g., RBX1 and RBX2). MLN4924: an inhibitor of UBA3; N8: neuronal precursor cell-expressed developmentally downregulated protein 8.

family members (CUL1, 2, 3, 4A, 4B, and 5), which function as core components of Cullin-RING E3 ubiquitin ligases (CRLs) [\[4\]](#page-5-0). The activation of CRLs requires NEDD8 to attach to a C-terminal lysine residue of cullins, inducing structural changes in the CRLs complex and leading to open conformation to facilitate substrate ubiquitylation [[15](#page-6-0)–[19\]](#page-6-0). As the most prominent family of E3 ubiquitin ligases, CRLs noticeably regulate various essential biological functions, including tumorigenesis, accomplished by enhancing ubiquitylation and consequent degradation of a wide range of critical modulatory proteins [[4](#page-5-0), [20](#page-6-0)]. Besides cullins, several non-cullin proteins have been identified as the substrates of neddylation, such as p53, MDM2, and EGFR [[21](#page-6-0)–[24\]](#page-6-0). However, regulatory mechanisms and physiological functions of non-cullin substrates still await experimental validation.

In multiple cancers, overactivation of the neddylation pathway leads to elevated global neddylation of substrates, such as cullins, and consequent accumulation of tumor suppressors, thereby promoting tumorigenesis and development [[25](#page-6-0)–[28\]](#page-6-0). Targeting the overactivated protein neddylation pathway has been proven to be an effective anti-tumor strategy. MLN4924, commonly referred to as pevonedistat, is an effective and highly selective smallmolecule inhibitor of NAE adopted to inhibit protein neddylation via inactivation of the initial stage of the neddylation cascade [[29](#page-6-0)]. When bound to the active site of UBA3, MLN4924 forms a stable covalent adduct with NEDD8 to block further enzymatic processes [[30\]](#page-6-0). It achieves potent anti-tumor effects by inducing cell cycle arrest, apoptosis, or senescence of tumor cells, or affecting the functions of multiple components of the tumor microenvironment [[28,](#page-6-0) [29,](#page-6-0) [31](#page-6-0)–[37](#page-6-0)]. MLN4924 has been evaluated in Phase I/II clinical trials for treating various solid tumors and hematologic cancers [[38](#page-6-0)-[44\]](#page-6-0). Nevertheless, the induction of drug-resistant mutations in UBA3 inhibits the formation of the MLN4924-NEDD8 adduct, necessitating the need to explore alternative targets against the neddylation pathway [[45,](#page-6-0) [46\]](#page-6-0). To address the limitations of MLN4924, specific inhibitors against neddylation E2s are being investigated [\[47](#page-6-0), [48](#page-6-0)]. This review summarizes the latest progress on validating the neddylation of these enzymes as promising antitumor targets.

## BIOLOGICAL CHARACTERISTICS AND CORRELATION OF UBE2M AND UBE2F

UBE2M and UBE2F bind to the ubiquitin-fold domain and the UBA3 hydrophobic groove of E1 via the core domain and the N-terminal motif, respectively [\[9\]](#page-6-0). Acetylation of the N-terminal methionine occurs in both E2s, which facilitates their binding to the PONY domain pocket of neddylation E3 DCN-like (DCNL), thereby increasing the efficiency of the cullin neddylation process [[49](#page-6-0)–[51\]](#page-6-0). In addition to these similarities, protein structure assessment of both UBE2M and UBE2F reveals unique characteristics. UBE2F is very specific to the neddylation of RBX2-related CUL5, while UBE2M can pair with RBX1 to modulate the neddylation of CUL1, 2, 3, 4A, and 4B [\[50](#page-6-0)]. It is noteworthy that glycyl-tRNA synthetase, an enzyme necessary for protein production, binds to the NAE1 subunit of E1 to capture and protect activated UBE2M before it reaches the downstream target [[52\]](#page-6-0).

The enzymes UBE2M and UBE2F can activate distinct cullins by enhancing neddylation modification, and interestingly, there is a cross-talk that leads to UBE2M targeting UBE2F for degradation [[53\]](#page-6-0). Specifically, UBE2M serves as a stress-inducible protein and a dual E2 for neddylation and ubiquitylation to degrade UBE2F. Under physiological conditions, UBE2M acts as a neddylation E2 to promote CUL3 neddylation, which triggers polyubiquitylation and degradation of UBE2F through CUL3-KEAP1 E3 ligase. However, UBE2M is transcriptionally activated under stress by hypoxiainducible factor 1α (HIF-1α) or transcription factor AP-1 (AP-1). UBE2M performs ubiquitylation E2 for Parkin-DJ-1 E3, followed by ubiquitylation and degradation of UBE2F. Ultimately, the degradation of UBE2F is accompanied by the inactivation of CRL5. Collectively, these findings provide evidence that one neddylation E2 (UBE2M) acts as a dual E2 for both neddylation and ubiquitylation to decrease the protein levels of the other (UBE2F), leading to one CRL E3 (CRL3) inactivating the other (CRL5) (Fig. [2](#page-2-0)).

#### UBE2M AND UBE2F AS ATTRACTIVE ANTI-TUMOR TARGETS

Most studies reveal that both two neddylation E2s act as oncogenes, evidenced mainly by their significant upregulation in various human cancers, including esophageal squamous cell carcinoma, osteosarcoma, lung cancer, and hepatocellular carcinoma [[54](#page-6-0)–[59](#page-7-0)]. In addition, the upregulation of the two neddylation E2s correlates closely with illness progression [\[55](#page-6-0)–[58](#page-7-0)]. To attain a greater understanding of the expression profile and prognostic significance of UBE2M and UBE2F in human cancers, we examined their expression with the aid of Tumor Immune Estimation Resource (TIMER) online database, an interactive platform that allows users to conduct in-depth studies of TCGA gene expression profiles [\(http://timer.cistrome.org/\)](http://timer.cistrome.org/). Compared to normal tissues, the mRNA levels of UBE2M are elevated in 17 different kinds of human cancers, while the mRNA levels of UBE2F are elevated in 12 different types of human cancers (Fig. [3A](#page-2-0), B). Kaplan–Meier analysis shows that increased mRNA levels of both enzymes correlated to worse survival for patients with lung adenocarcinoma (LUAD) or with liver hepatocellular carcinoma (LIHC) (Fig. [3](#page-2-0)C, D). They exert oncogenic effects by enhancing the neddylation of certain substrates to mediate a variety of signaling pathways and modulate multiple biological activities, such as apoptosis or senescence (Fig. [4\)](#page-3-0). These findings suggest that the overactivation of these two enzymes could be an oncogenic event throughout the process of tumor occurrence and development.

#### TARGETING UBE2M FOR ANTI-TUMOR THERAPY

In recent decades, increasing studies have shown UBE2M, also known as UBC12, to be an attractive anti-tumor target. Compared with E1 subunits (NAE1 and UBA3), UBE2M is significantly more consistent with the global protein neddylation levels [[56](#page-6-0)]. Targeting UBE2M suppresses the growth of MLN4924-resistant

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Fig. 2 UBE2M acts as a dual E2 for neddylation and ubiquitylation to degrade UBE2F. Under physiological conditions, UBE2M serves as a neddylation E2 to promote CUL3 neddylation, triggering polyubiquitylation and degradation of UBE2F through CUL3-KEAP1 E3 ligase. Under stressed conditions, UBE2M is transcriptionally activated by HIF-1 $\alpha$  or AP-1, which makes it serve as a ubiquitylation E2 to complex with DJ-1/Parkin to promote the ubiquitylation and degradation of UBE2F. Ultimately, the degradation of UBE2F is accompanied by the inactivation of CRL5. TPA: a typical tumor promoter and mitogen stimulator, inducing c-JUN.

cells by inhibiting cullins neddylation and inducing the accumulation of CRLs substrates [\[56](#page-6-0)]. Moreover, UBE2M-mediated protein neddylation is essential for multiple cellular responses, such as DNA-damage response (DDR), apoptosis, senescence, and antitumor immunity. These findings validate UBE2M as an attractive alternative anti-tumor target to efficiently inhibit the neddylation pathway.

# UBE2M PARTICIPATES IN DDR

DNA double-strand break (DSB), contributing significantly to genomic stability [\[60](#page-7-0), [61\]](#page-7-0), is sensed and repaired by DDR, warranting the recruitment and post-translational modification of multiple proteins at the damaged DNA sites. This phenomenon induces checkpoint signaling or essential repair steps [\[62](#page-7-0), [63\]](#page-7-0). Recent studies have shown that UBE2M-mediated neddylation of cullins, or non-cullins, is involved in the DDR pathway (Fig. [4\)](#page-3-0). UBE2M affects DDR and the integrity of the genome by regulating several CRLs substrates, such as CDT1, p21, and claspin, which play complex roles in the increased DNA-damage in UBE2M-silenced cells [\[64\]](#page-7-0). Another study supported the accumulation effects of CRLs substrates CDT1 and ORC1 caused by silenced UBE2M. This phenomenon subsequently induces DSBs, evidenced by the upregulated expression of  $γ$ -H2AX [\[65](#page-7-0)].

Generally, Ku70/Ku80 (Ku) heterodimer is first recruited to the DSB sites, after which, DNA-dependent protein kinase catalytic subunit (DNA-PKcs) is recruited into the process, facilitating nonhomologous end-joining (NHEJ) repair [\[66,](#page-7-0) [67\]](#page-7-0). Upon DNAdamage, UBE2M-mediated cullins neddylation promotes ubiquitylation of Ku, releasing it and its associated proteins from the damaged sites after repair. After NHEJ, ionizing radiation hypersensitivity and decreased cell survival occur in UBE2Mdepleted cells [[68](#page-7-0)]. Moreover, UBE2M also regulates the neddylation of DNA-PKcs, which promotes DNA-PKc autophosphorylation, preferentially activating the NHEJ pathway and facilitating its



Fig. 3 Expression of UBE2M and UBE2F in cancers and their effects on the prognosis of patients with LUAD and LIHC. A, B The expression of UBE2M and UBE2F in different types of cancer was investigated with the TIMER database. The variance was similar between the groups that were being compared. P<0.05 was considered as statistical significance. \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 for the indicated comparison. C, D Kaplan-Meier analysis shows that increased levels of both UBE2M and UBE2F are related to worse survival for patients with lung adenocarcinoma (LUAD) or with liver hepatocellular carcinoma (LIHC).



Fig. 4 The function of UBE2M and UBE2F in tumor growth. UBE2M influences tumor growth by modulating several biological responses of tumor cells, such as DNA-damage response, cell cycle arrest, apoptosis, or senescence. UBE2F promotes the degradation of NOXA and then inhibits apoptosis and induces tumor growth.

release from DNA-damage sites after Ku [[69\]](#page-7-0). Consistently, the knockdown of UBE2M significantly enhances the sensitivity of hormone-resistant prostate cancer cells to radiation-induced DNA damage [\[70\]](#page-7-0).

Furthermore, in response to various stimuli, NEDD8 may accumulate at DNA-damage sites by relying on UBE2M rather than UBE2F. UBE2M, together with an E3 ubiquitin ligase RNF111, promotes ionizing radiation-induced histone H4 neddylation and links another E3 ligase RNF168 to DNA-damage sites. This occurrence leads to the recruitment of BRCA1 and other downstream DDR factors to repair the damaged DNA [[71\]](#page-7-0). Notably, another group has reported that UBE2M/RNF111-mediated neddylation inhibits BRCA1 and CtIP-regulated DNA end resection, an essential mechanism aiding the selection of an appropriate repair pathway [[72\]](#page-7-0).

# UBE2M INHIBITION INDUCES CELL CYCLE ARREST

The aberrant functioning of cell cycle regulators leads to unregulated cell proliferation, making them promising therapeutic targets for cancer treatment [[73,](#page-7-0) [74](#page-7-0)]. UBE2M has been shown to crucially regulate the tumor-suppressive cell cycle inhibitors (Fig. 4). In lung cancer and esophageal squamous cell carcinoma cells, UBE2M knockdown disturbs cell cycle progression by triggering G2 phase cell cycle arrest, specifically by inhibiting cullins neddylation and upregulating CRL substrates (p21, p27, and Wee1) [[55,](#page-6-0) [56\]](#page-6-0). In hepatocellular carcinoma cells, UBE2M-mediated stabilization of β-catenin, leading to the upregulation of its downstream effectors, known as cyclin D1, promotes the G1/S transition of cells [[75\]](#page-7-0). These data suggest that suppression of UBE2M can potentially trigger cell cycle arrest at distinct stages in a cell type-dependent manner via multiple mechanisms.

# UBE2M INHIBITION INDUCES APOPTOSIS OR SENESCENCE

UBE2M plays an essential role in regulating apoptosis and senescence (Fig. 4). In intrahepatic cholangiocarcinoma cells, UBE2M knockdown induces apoptosis, demonstrated by shrinkage of cellular morphology and the upregulation of cleaved PARP and caspase-3/-9 [[65\]](#page-7-0). In hepatocellular cells, UBE2M knockdown promotes apoptosis by inducing the accumulation of cleaved PARP and caspase-3 and increasing the mRNA levels of apoptosis-associated proteins, including p53, PUMA, and Bax [\[76](#page-7-0)]. Interestingly, UBE2M knockdown leads to p53-mediated apoptosis by activating and stabilizing the protein [[76\]](#page-7-0). In esophageal squamous cell carcinoma cells, UBE2M knockdown triggers apoptosis or senescence in a cell line-dependent manner. EC1 cells with UBE2M knockdown exhibit typical senescent morphology, wellcharacterized by an enlarged and flattened cellular shape and positive staining for senescence-related β-galactosidase [\[55](#page-6-0)]. In contrast, KYSE450 cells with UBE2M knockdown show prominent apoptotic features, shrunk morphology, and a substantial increase in cell numbers positive for annexin V [[55\]](#page-6-0). Moreover, UBE2M knockdown increases the accumulation of CRL substrate activating transcription factor 4 (ATF4), activating death receptor 5 (DR5) mediated extrinsic apoptosis and proapoptotic protein NOXAmediated intrinsic apoptosis [\[55](#page-6-0), [77](#page-7-0), [78\]](#page-7-0). Thus, induction of apoptosis or senescence establishes UBE2M as a promising antitumor target.

# UBE2M IS INVOLVED IN ANTI-TUMOR IMMUNITY

Immune evasion is a hallmark of cancer. Anti-tumorigenic immune cells are often exhausted or repressed by immune suppressive cell populations, such as tumor-associated macrophages (TAMs), regulatory T (Treg), and myeloid-derived suppressor cells (MDSCs),

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Fig. 5 The correlation of UBE2M and immune suppressive cell populations. A Graphs generated from TIMER database show the correlations between UBE2M and the abundance of macrophage, Treg, and MDSCs. B, C The expression of UBE2M was significantly correlated with multiple genes that are related to macrophage M2, MDSCs, or Treg in liver hepatocellular carcinoma. Spearman correlation analysis was used to assess the correlation.  $P < 0.05$  was considered as statistical significance.

leading to tumor immune evasion [\[79](#page-7-0)–[81\]](#page-7-0). Hence, targeting these immune suppressive cells is a promising anti-tumor immunotherapy. It is known that UBE2M knockdown inhibits the expression of proinflammatory cytokines triggered by exposure to lipopolysaccharides (LPS) (e.g., IL-6 and TNF-α) in macrophages. UBE2M knockdown suppresses CUL1 neddylation, inactivates CRL1, and induces the accumulation of phosphorylated IκBα and the subsequent suppression of NF-κB nuclear translocation. This phenomenon transcriptionally inhibits macrophage-associated cytokines [[82\]](#page-7-0). Recent reports suggest that the Ube2m/Rbx1 axis, rather than the Ube2f/Rbx2 one, is crucial to the homeostasis and survival of Treg cells [\[83\]](#page-7-0). We then investigated the association between UBE2M expression and the abundance of immune cell populations in tumors using the TIMER database to assess further the effects of UBE2M on these immune suppressive cells (Fig. 5A). This analysis revealed that in hepatocellular carcinoma, the expression of UBE2M was significantly positively correlated with the abundance of Treg, MDSCs, and macrophage M2 (Fig. 5A). Moreover, the expression of UBE2M was also significantly correlated with multiple genes related to macrophage M2, MDSCs, or Treg (Fig. 5B, C). These results collectively suggest that UBE2M may drive immunosuppression in hepatocellular carcinoma. However, future investigation of detailed mechanisms is warranted.

Targeting immune checkpoints mediated by programmed cell death 1 (PD-1) and its ligand PD-L1 is a practical approach to enhance anti-tumor immunity. This line of therapy has been approved for treating various human cancers with durable clinical benefits [\[84](#page-7-0), [85\]](#page-7-0). Notably, inhibition of UBE2M-mediated neddylation significantly upregulates the expression of PD-L1 by inactivating CUL1 and CUL3 in glioblastoma cancer cells. This phenomenon is mainly achieved by the transcriptional activation of PD-L1 by dysregulating the CUL1-FBXW7/c-MYC axis and stabilizing PD-L1 protein by inhibiting CUL3 E3 ligase activity, leading to T-cell exhaustion [[86\]](#page-7-0). Moreover, inhibition of CUL3SPOP E3 ligase impairs ubiquitination-mediated PD-L1 degradation, increasing PD-L1 protein levels and reducing the numbers of tumor-infiltrating lymphocytes in mouse tumors and primary human prostate cancer [\[87](#page-7-0)]. A recent study also revealed that inhibition of the neddylation pathway by MLN4924 activates ERK and JNK signals, leading to AP-1 activation. Activated AP-1 transactivates PD-L1 expression, inducing tumor immune evasion to fight the anti-tumor activity of MLN4924 [\[88](#page-7-0)]. These findings offer novel insights for further clinical experiments on tumor patients using a combined method of UBE2M targeting and anti-PD-L1/PD-1 therapy. Altogether, the identification of the role of UBE2M in modulating anti-tumor immunity deserves further research.

## TARGETING UBE2M-DCN1 INTERACTION FOR REGULATION OF NEDDYLATION PATHWAY

Efforts have been directed to develop more specific smallmolecule inhibitors that preferentially target neddylation E2s to address the shortcomings of MLN4924. The co-crystal structure assessment of DCN1, which has no RING finger domain and acts as a co-NEDD8-E3 with RBX1, and its binding partners UBE2M, harbors the potential to develop strong small-molecule inhibitors [\[47,](#page-6-0) [51](#page-6-0), [89](#page-7-0)-[91](#page-7-0)]. Over the past few years, several different research facilities have developed small-molecule inhibitors for suppressing interactions between UBE2M and DCN1. Zhou et al. created potent peptidomimetics, such as DI-591 and DI-404, by significantly modifying the N-terminal 12-residue peptide of UBE2M to inhibit UBE2M-DCN1 interaction [\[92](#page-7-0), [93](#page-7-0)]. Subsequently, Guy et al. identified non-peptidic and potent small-molecule UBE2M-DCN1 inhibitors, such as NAcM-HIT, by high-throughput screening of over 600,000 compounds. The chemical optimization of NAcM-HIT led to designing two more potent inhibitors, NAcM-OPT and NAcM-COV [\[94](#page-7-0)–[96](#page-7-0)]. Liu et al. discovered the triazolo[1,5-α] pyrimidine-based inhibitor WS-383, which targets the UBE2M-

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DCN1 interaction [[97](#page-7-0)]. In addition, Zhao et al. discovered smallmolecular DCN1 inhibitors DC-1 and DC-2 based on pyrimidines [[98\]](#page-7-0).

Such studies have successfully discovered potent smallmolecule inhibitors that are either covalent or non-covalent with powerful affinities to DCN1. These UBE2M-DCN1 inhibitors have been shown to efficiently block the neddylation of CUL1 and/or CUL3 while exerting no or minimal impact on the other members of the cullin family. These UBE2M-DCN1 inhibitors, as predicted, cause an accumulation of the CUL1 or CUL3 substrates, such as p21, p27, and NRF2 [\[92](#page-7-0), [95](#page-7-0)–[98](#page-7-0)]. However, compared to MLN4924, the UBE2M-DCN1 inhibitors have moderate cellular potency, suggesting the need for further potency improvement.

# TARGETING UBE2F FOR ANTI-TUMOR THERAPY

In contrast with many studies done on UBE2M, UBE2F receives little attention. Recently, UBE2F has been shown to inhibit apoptosis and induce cell growth. UBE2F can be efficiently targeted as a chemo-/radiosensitizing strategy by triggering apoptosis.

#### UBE2F INHIBITION INDUCES APOPTOSIS

Zhou et al. reported that by coupling to RBX2, UBE2F neddylates CUL5, which activates CRL5, eventually leading to the ubiquitylation and degradation of NOXA through the K11-linkage [\[57](#page-7-0)]. In lung cancer cells, overexpression of UBE2F activates CRL5 and promotes NOXA degradation, leading to inhibition of apoptosis and improvement of cell survival [\[57](#page-7-0)]. HA-9104 is recently discovered as a novel small-molecule inhibitor targeting UBE2F-CRL5 axis. HA-9104 interacts with UBE2F to reduce its protein levels (via a yet-to-defined mechanism), thereby inhibiting CUL5 neddylation. Blockage of CUL5 neddylation results in CRL5 inhibition and NOXA accumulation to trigger apoptosis [[48\]](#page-6-0). Since UBE2F inhibition promotes apoptosis and suppresses cancer cell growth (Fig. [4\)](#page-3-0), it is a promising target for anti-tumor therapy.

Moreover, peroxiredoxin PRDX1 binds to UBE2F and CUL5 to form a triple-molecule complex, PRDX1-UBE2F-CUL5, essential for CUL5 neddylation. Silencing PRDX1 or blocking PRDX1 oligomerization significantly inhibits CUL5 neddylation, suppressing NOXA ubiquitination and degradation. Etoposide, an anti-cancer chemotherapeutic DNA damaging agent, increases NOXA transcription, leading to apoptosis. Colorectal cancer cells increase CUL5 neddylation to accelerate NOXA degradation, which prevents etoposide-induced apoptosis. At the same time, the knockdown of PRDX1 eliminates etoposide-induced CUL5 neddylation and increases the sensitivity of colorectal cancer cells to etoposide therapy [\[99](#page-7-0)].

# UBE2F SERVES AS A TARGET FOR CHEMO-/ RADIOSENSITIZATION

UBE2F, an apoptotic regulatory protein, is a probable viable target for chemosensitization [[100,](#page-7-0) [101](#page-7-0)]. In lung cancer cells, depletion of UBE2F renders the cells more sensitive to multiple anti-tumor agents (e.g., an inhibitor of anti-apoptotic protein MCL1) by accumulating NOXA [\[101\]](#page-7-0). UBE2F upregulation allows lung cancer cells to evade apoptosis caused by platinum exposure. Mechanistically, platinum prevents the generation of the complex required for proteasome-mediated UBE2F degradation, which ultimately results in UBE2F accumulation. This phenomenon demonstrates an increase in the CUL5 neddylation level, consistent with the decreased protein levels of NOXA. UBE2F knockdown dramatically increases cell sensitivity to platinum therapy by increasing NOXA protein levels and consequently promoting apoptosis [\[100\]](#page-7-0). Notably, these effects manifest in other cancer cells, such as breast and ovarian cancer cells, indicating the role of UBE2F, a universal drug target of platinumsensitization [\[100\]](#page-7-0). Nevertheless, the expressions of other

neddylation factors, such as UBE2M, are not affected following platinum treatment [[100](#page-7-0)].

Besides chemosensitization, targeting UBE2F also displays a sensitizing effect on radiotherapy [[102](#page-7-0)]. Elevated UBE2F levels can be attributed to oxidative stress induced by irradiation or other stimuli, causing the degradation of ROS-induced NOXA, consequently inducing apoptotic resistance to radiotherapy. Moreover, silencing UBE2F suppresses NOXA degradation and increases cancer cells' susceptibility to irradiation-mediated apoptosis [\[102](#page-7-0)]. Taken together, UBE2F-mediated activation of CRL5 and subsequent ubiquitylation and degradation of NOXA potentially hold great promise as both an anti-tumor target and a chemo-/ radiosensitizing target.

## **CONCLUSIONS**

Targeting the overactivated neddylation pathway has been demonstrated as a promising anti-tumor strategy, supported by the development of MLN4924, a potent inhibitor of the neddylation E1 subunit UBA3. However, the emergence of drugresistant mutations in UBA3 warrants the identification of alternative targets against the neddylation pathway. Recent and ongoing research has revealed that UBE2M and UBE2F perform an integral function in the biology of tumors. Overexpression of both UBE2M and UBE2F in cancer cells is associated with increased cell proliferation and poor survival. UBE2M regulates tumor growth by modulating several cellular responses, such as DDR, senescence, or apoptosis. The functional role of UBE2F, on the other hand, remains poorly characterized. Recent studies have shown that UBE2F inhibits apoptosis, induces cell growth, and serves as an effective chemo-/radiosensitizing target. However, the effects of both E2s on anti-tumor immunity undoubtedly demand more experimental investigations. In-depth elucidation of the mechanisms of these E2s may provide more in-depth knowledge for targeting UBE2M and UBE2F as attractive anti-tumor therapy.

Many inhibitors targeting UBE2M-DCN1 interaction have been discovered to overcome the limitations of MLN4924 and tackle the crucial role of UBE2M on the neddylation pathway and tumor growth. However, compared to MLN4924, the tumor cell-killing potency of these UBE2M-DCN1 inhibitors is moderate, suggesting the need for further potency improvement. Promoting anti-tumor clinical trials of these E2 inhibitors alone or in combination may direct further research. Recent studies reported that, in addition to effective tumor treatment, MLN4924 also plays a potential role in the treatment of obesity [[103\]](#page-7-0), insulin resistance [\[103,](#page-7-0) [104](#page-7-0)], nonalcoholic fatty liver [\[105\]](#page-7-0), and ischemia-reperfusion injury [[106](#page-7-0)–[108\]](#page-7-0). Hence, it would be interesting to identify the function of these UBE2M-DCN1 inhibitors in these non-tumor diseases.

In summary, the current research offers profound insights into the role of UBE2M and UBE2F in tumor progression, which is highly conducive to aiding the development of targeted inhibitors with higher potency and selectivity.

#### DATA AVAILABILITY

The corresponding author will provide the data and materials upon reasonable request.

# **REFERENCES**

- 1. Kamitani T, Kito K, Nguyen HP, Yeh ET. Characterization of NEDD8, a developmentally down-regulated ubiquitin-like protein. J Biol Chem. 1997;272:28557–62.
- 2. Xirodimas DP. Novel substrates and functions for the ubiquitin-like molecule NEDD8. Biochem Soc Trans. 2008;36:802–6.
- 3. Zhao Y, Morgan MA, Sun Y. Targeting Neddylation pathways to inactivate cullin-RING ligases for anticancer therapy. Antioxid Redox Signal. 2014;21:2383–400.
- 4. Enchev RI, Schulman BA, Peter M. Protein neddylation: beyond cullin-RING ligases. Nat Rev Mol Cell Biol. 2015;16:30–44.
- <span id="page-6-0"></span>5. Walden H, Podgorski MS, Huang DT, Miller DW, Howard RJ, Minor DL Jr, et al. The structure of the APPBP1-UBA3-NEDD8-ATP complex reveals the basis for selective ubiquitin-like protein activation by an E1. Mol Cell. 2003;12:1427–37.
- 6. Bohnsack RN, Haas AL. Conservation in the mechanism of Nedd8 activation by the human AppBp1-Uba3 heterodimer. J Biol Chem. 2003;278:26823–30.
- 7. Gong L, Yeh ET. Identification of the activating and conjugating enzymes of the NEDD8 conjugation pathway. J Biol Chem. 1999;274:12036–42.
- 8. Huang DT, Paydar A, Zhuang M, Waddell MB, Holton JM, Schulman BA. Structural basis for recruitment of Ubc12 by an E2 binding domain in NEDD8's E1. Mol Cell. 2005;17:341–50.
- 9. Huang DT, Miller DW, Mathew R, Cassell R, Holton JM, Roussel MF, et al. A unique E1-E2 interaction required for optimal conjugation of the ubiquitin-like protein NEDD8. Nat Struct Mol Biol. 2004;11:927–35.
- 10. Zhou L, Zhang W, Sun Y, Jia L. Protein neddylation and its alterations in human cancers for targeted therapy. Cell Signal. 2018;44:92–102.
- 11. Duan H, Wang Y, Aviram M, Swaroop M, Loo JA, Bian J, et al. SAG, a novel zinc RING finger protein that protects cells from apoptosis induced by redox agents. Mol Cell Biol. 1999;19:3145–55.
- 12. Kamura T, Conrad MN, Yan Q, Conaway RC, Conaway JW. The Rbx1 subunit of SCF and VHL E3 ubiquitin ligase activates Rub1 modification of cullins Cdc53 and Cul2. Genes Dev. 1999;13:2928–33.
- 13. Kurz T, Chou YC, Willems AR, Meyer-Schaller N, Hecht ML, Tyers M, et al. Dcn1 functions as a scaffold-type E3 ligase for cullin neddylation. Mol Cell. 2008;29:23–35.
- 14. Meyer-Schaller N, Chou YC, Sumara I, Martin DD, Kurz T, Katheder N, et al. The human Dcn1-like protein DCNL3 promotes Cul3 neddylation at membranes. Proc Natl Acad Sci USA. 2009;106:12365–70.
- 15. Merlet J, Burger J, Gomes JE, Pintard L. Regulation of cullin-RING E3 ubiquitinligases by neddylation and dimerization. Cell Mol Life Sci. 2009;66:1924–38.
- 16. Sakata E, Yamaguchi Y, Miyauchi Y, Iwai K, Chiba T, Saeki Y, et al. Direct interactions between NEDD8 and ubiquitin E2 conjugating enzymes upregulate cullin-based E3 ligase activity. Nat Struct Mol Biol. 2007;14:167–8.
- 17. Saha A, Deshaies RJ. Multimodal activation of the ubiquitin ligase SCF by Nedd8 conjugation. Mol Cell. 2008;32:21–31.
- 18. Duda DM, Borg LA, Scott DC, Hunt HW, Hammel M, Schulman BA. Structural insights into NEDD8 activation of cullin-RING ligases: conformational control of conjugation. Cell 2008;134:995–1006.
- 19. Baek K, Krist DT, Prabu JR, Hill S, Klugel M, Neumaier LM, et al. NEDD8 nucleates a multivalent cullin-RING-UBE2D ubiquitin ligation assembly. Nature 2020;578:461–6.
- 20. Petroski MD, Deshaies RJ. Function and regulation of cullin-RING ubiquitin ligases. Nat Rev Mol Cell Biol. 2005;6:9–20.
- 21. Dohmesen C, Koeppel M, Dobbelstein M. Specific inhibition of Mdm2-mediated neddylation by Tip60. Cell cycle. 2008;7:222–31.
- 22. Xirodimas DP, Saville MK, Bourdon JC, Hay RT, Lane DP. Mdm2-mediated NEDD8 conjugation of p53 inhibits its transcriptional activity. Cell 2004;118:83–97.
- 23. Oved S, Mosesson Y, Zwang Y, Santonico E, Shtiegman K, Marmor MD, et al. Conjugation to Nedd8 instigates ubiquitylation and down-regulation of activated receptor tyrosine kinases. J Biol Chem. 2006;281:21640–51.
- 24. Zhou L, Jia L. Targeting protein neddylation for cancer therapy. Adv Exp Med Biol. 2020;1217:297–315.
- 25. Li L, Wang M, Yu G, Chen P, Li H, Wei D, et al. Overactivated neddylation pathway as a therapeutic target in lung cancer. J Natl Cancer Inst. 2014;106:dju083.
- 26. Jia X, Li C, Li L, Liu X, Zhou L, Zhang W, et al. Neddylation inactivation facilitates FOXO3a nuclear export to suppress estrogen receptor transcription and improve fulvestrant sensitivity. Clin Cancer Res. 2019;25:3658–72.
- 27. Hua W, Li C, Yang Z, Li L, Jiang Y, Yu G, et al. Suppression of glioblastoma by targeting the overactivated protein neddylation pathway. Neuro Oncol. 2015;17:1333–43.
- 28. Olaizola P, Lee-Law PY, Fernandez-Barrena MG, Alvarez L, Cadamuro M, Azkargorta M, et al. Targeting NAE1-mediated protein hyper-NEDDylation halts cholangiocarcinogenesis and impacts on tumor-stroma crosstalk in experimental models. J Hepatol. 2022;77:177–90.
- 29. Soucy TA, Smith PG, Milhollen MA, Berger AJ, Gavin JM, Adhikari S, et al. An inhibitor of NEDD8-activating enzyme as a new approach to treat cancer. Nature 2009;458:732–6.
- 30. Brownell JE, Sintchak MD, Gavin JM, Liao H, Bruzzese FJ, Bump NJ, et al. Substrate-assisted inhibition of ubiquitin-like protein-activating enzymes: the NEDD8 E1 inhibitor MLN4924 forms a NEDD8-AMP mimetic in situ. Mol Cell. 2010;37:102–11.
- 31. Jia L, Li H, Sun Y. Induction of p21-dependent senescence by an NAE inhibitor, MLN4924, as a mechanism of growth suppression. Neoplasia 2011;13:561–9.
- 32. Milhollen MA, Traore T, Adams-Duffy J, Thomas MP, Berger AJ, Dang L, et al. MLN4924, a NEDD8-activating enzyme inhibitor, is active in diffuse large B-cell

lymphoma models: rationale for treatment of NF-{kappa}B-dependent lymphoma. Blood 2010;116:1515–23.

- 33. Milhollen MA, Narayanan U, Soucy TA, Veiby PO, Smith PG, Amidon B. Inhibition of NEDD8-activating enzyme induces rereplication and apoptosis in human tumor cells consistent with deregulating CDT1 turnover. Cancer Res. 2011;71:3042–51.
- 34. Knorr KL, Schneider PA, Meng XW, Dai H, Smith BD, Hess AD, et al. MLN4924 induces Noxa upregulation in acute myelogenous leukemia and synergizes with Bcl-2 inhibitors. Cell Death Differ. 2015;22:2133–42.
- 35. Zhou L, Jiang Y, Liu X, Li L, Yang X, Dong C, et al. Promotion of tumor-associated macrophages infiltration by elevated neddylation pathway via NF-kappaB-CCL2 signaling in lung cancer. Oncogene 2019;38:5792–804.
- 36. Best S, Lam V, Liu T, Bruss N, Kittai A, Danilova OV, et al. Immunomodulatory effects of pevonedistat, a NEDD8-activating enzyme inhibitor, in chronic lymphocytic leukemia-derived T cells. Leukemia 2021;35:156–68.
- 37. Zhou L, Zhang L, Chen S, Sun D, Qu J. Elevated neddylation pathway promotes Th2 cells infiltration by transactivating STAT5A in hepatocellular carcinoma. Front Oncol. 2021;11:709170.
- 38. Zhou L, Jiang Y, Luo Q, Li L, Jia L. Neddylation: a novel modulator of the tumor microenvironment. Mol Cancer. 2019;18:77.
- 39. Sarantopoulos J, Shapiro GI, Cohen RB, Clark JW, Kauh JS, Weiss GJ, et al. Phase I study of the investigational NEDD8-activating enzyme inhibitor pevonedistat (TAK-924/MLN4924) in patients with advanced solid tumors. Clin Cancer Res. 2016;22:847–57.
- 40. Shah JJ, Jakubowiak AJ, O'Connor OA, Orlowski RZ, Harvey RD, Smith MR, et al. Phase I study of the novel investigational NEDD8-activating enzyme inhibitor pevonedistat (MLN4924) in patients with relapsed/refractory multiple myeloma or lymphoma. Clin Cancer Res. 2016;22:34–43.
- 41. Sekeres MA, Watts J, Radinoff A, Sangerman MA, Cerrano M, Lopez PF, et al. Randomized phase 2 trial of pevonedistat plus azacitidine versus azacitidine for higher-risk MDS/CMML or low-blast AML. Leukemia 2021;35:2119–24.
- 42. Swords RT, Coutre S, Maris MB, Zeidner JF, Foran JM, Cruz J, et al. Pevonedistat, a first-in-class NEDD8-activating enzyme inhibitor, combined with azacitidine in patients with AML. Blood 2018;131:1415–24.
- 43. Handa H, Cheong JW, Onishi Y, Iida H, Kobayashi Y, Kim HJ, et al. Pevonedistat in East Asian patients with acute myeloid leukemia or myelodysplastic syndromes: a phase 1/1b study to evaluate safety, pharmacokinetics and activity as a single agent and in combination with azacitidine. J Hematol Oncol. 2022;15:56.
- 44. Swords RT, Watts J, Erba HP, Altman JK, Maris M, Anwer F, et al. Expanded safety analysis of pevonedistat, a first-in-class NEDD8-activating enzyme inhibitor, in patients with acute myeloid leukemia and myelodysplastic syndromes. Blood Cancer J. 2017;7:e520.
- 45. Toth JI, Yang L, Dahl R, Petroski MD. A gatekeeper residue for NEDD8-activating enzyme inhibition by MLN4924. Cell Rep. 2012;1:309–16.
- 46. Milhollen MA, Thomas MP, Narayanan U, Traore T, Riceberg J, Amidon BS, et al. Treatment-emergent mutations in NAEbeta confer resistance to the NEDD8 activating enzyme inhibitor MLN4924. Cancer Cell. 2012;21:388–401.
- 47. Zhou H, Lu J, Yang CY, Sun Y, Wang S. Targeting DCN1-UBC12 protein-protein interaction for regulation of neddylation pathway. Adv Exp Med Biol. 2020;1217:349–62.
- 48. Xu T, Ma Q, Li Y, Yu Q, Pan P, Zheng Y, et al. A small molecule inhibitor of the UBE2F-CRL5 axis induces apoptosis and radiosensitization in lung cancer. Signal Transduct Target Ther. 2022;7:354.
- 49. Monda JK, Scott DC, Miller DJ, Lydeard J, King D, Harper JW, et al. Structural conservation of distinctive N-terminal acetylation-dependent interactions across a family of mammalian NEDD8 ligation enzymes. Structure 2013;21:42–53.
- 50. Huang DT, Ayrault O, Hunt HW, Taherbhoy AM, Duda DM, Scott DC, et al. E2- RING expansion of the NEDD8 cascade confers specificity to cullin modification. Mol Cell. 2009;33:483–95.
- 51. Scott DC, Monda JK, Bennett EJ, Harper JW, Schulman BA. N-terminal acetylation acts as an avidity enhancer within an interconnected multiprotein complex. Science 2011;334:674–8.
- 52. Mo Z, Zhang Q, Liu Z, Lauer J, Shi Y, Sun L, et al. Neddylation requires glycyltRNA synthetase to protect activated E2. Nat Struct Mol Biol. 2016;23:730–7.
- 53. Zhou W, Xu J, Tan M, Li H, Li H, Wei W, et al. UBE2M is a stress-inducible dual E2 for neddylation and ubiquitylation that promotes targeted degradation of UBE2F. Mol Cell. 2018;70:1008–24 e6.
- 54. Zhang Y, Shi CC, Zhang HP, Li GQ, Li SS. MLN4924 suppresses neddylation and induces cell cycle arrest, senescence, and apoptosis in human osteosarcoma. Oncotarget 2016;7:45263–74.
- 55. Wang S, Xian J, Li L, Jiang Y, Liu Y, Cai L, et al. NEDD8-conjugating enzyme UBC12 as a novel therapeutic target in esophageal squamous cell carcinoma. Signal Transduct Target Ther. 2020:5:123.
- 56. Li L, Kang J, Zhang W, Cai L, Wang S, Liang Y, et al. Validation of NEDD8 conjugating enzyme UBC12 as a new therapeutic target in lung cancer. EBio-Medicine 2019;45:81–91.
- <span id="page-7-0"></span> $\overline{8}$
- 57. Zhou W, Xu J, Li H, Xu M, Chen ZJ, Wei W, et al. Neddylation E2 UBE2F promotes the survival of lung cancer cells by activating CRL5 to degrade NOXA via the K11 linkage. Clin Cancer Res. 2017;23:1104–16.
- 58. Gao Q, Yu GY, Shi JY, Li LH, Zhang WJ, Wang ZC, et al. Neddylation pathway is up-regulated in human intrahepatic cholangiocarcinoma and serves as a potential therapeutic target. Oncotarget 2014;5:7820–32.
- 59. Heo MJ, Kang SH, Kim YS, Lee JM, Yu J, Kim HR, et al. UBC12-mediated SREBP-1 neddylation worsens metastatic tumor prognosis. Int J Cancer. 2020;147:2550–63.
- 60. Jackson SP, Bartek J. The DNA-damage response in human biology and disease. Nature 2009;461:1071–8.
- 61. Ciccia A, Elledge SJ. The DNA damage response: making it safe to play with knives. Mol Cell. 2010;40:179–204.
- 62. Hauer MH, Gasser SM. Chromatin and nucleosome dynamics in DNA damage and repair. Genes Dev. 2017;31:2204–21.
- 63. Polo SE, Jackson SP. Dynamics of DNA damage response proteins at DNA breaks: a focus on protein modifications. Genes Dev. 2011;25:409–33.
- 64. Cukras S, Morffy N, Ohn T, Kee Y. Inactivating UBE2M impacts the DNA damage response and genome integrity involving multiple cullin ligases. PLoS One. 2014;9:e101844.
- 65. Zhao B, Gao C, Shi D, Mao J, Zhao J, Guo L, et al. Knockdown of Nedd8conjugating enzyme UBE2M suppresses the proliferation and induces the apoptosis of intrahepatic cholangiocarcinoma cells. Oncol Rep. 2019;42:2670–9.
- 66. Hudson JJ, Hsu DW, Guo K, Zhukovskaya N, Liu PH, Williams JG, et al. DNA-PKcsdependent signaling of DNA damage in Dictyostelium discoideum. Curr Biol. 2005;15:1880–5.
- 67. Spagnolo L, Rivera-Calzada A, Pearl LH, Llorca O. Three-dimensional structure of the human DNA-PKcs/Ku70/Ku80 complex assembled on DNA and its implications for DNA DSB repair. Mol Cell. 2006;22:511–9.
- 68. Brown JS, Lukashchuk N, Sczaniecka-Clift M, Britton S, le Sage C, Calsou P, et al. Neddylation promotes ubiquitylation and release of Ku from DNA-damage sites. Cell Rep. 2015;11:704–14.
- 69. Guo Z, Wang S, Xie Y, Han Y, Hu S, Guan H, et al. HUWE1-dependent DNA-PKcs neddylation modulates its autophosphorylation in DNA damage response. Cell Death Dis. 2020;11:400.
- 70. Wang X, Zhang W, Yan Z, Liang Y, Li L, Yu X, et al. Radiosensitization by the investigational NEDD8-activating enzyme inhibitor MLN4924 (pevonedistat) in hormone-resistant prostate cancer cells. Oncotarget 2016;7:38380–91.
- 71. Ma T, Chen Y, Zhang F, Yang CY, Wang S, Yu X. RNF111-dependent neddylation activates DNA damage-induced ubiquitination. Mol Cell. 2013;49:897–907.
- 72. Jimeno S, Fernandez-Avila MJ, Cruz-Garcia A, Cepeda-Garcia C, Gomez-Cabello D, Huertas P. Neddylation inhibits CtIP-mediated resection and regulates DNA double strand break repair pathway choice. Nucleic Acids Res. 2015;43:987–99.
- 73. Otto T, Sicinski P. Cell cycle proteins as promising targets in cancer therapy. Nat Rev Cancer. 2017;17:93–115.
- 74. Suski JM, Braun M, Strmiska V, Sicinski P. Targeting cell-cycle machinery in cancer. Cancer cell. 2021;39:759–78.
- 75. Zhang GC, Yu XN, Sun JL, Xiong J, Yang YJ, Jiang XM, et al. UBE2M promotes cell proliferation via the beta-catenin/cyclin D1 signaling in hepatocellular carcinoma. Aging 2020;12:2373–92.
- 76. Kim JH, Jung JH, Lee HJ, Sim DY, Im E, Park J, et al. UBE2M drives hepatocellular cancer progression as a p53 negative regulator by binding to MDM2 and ribosomal protein L11. Cancers. 2021;13:4901.
- 77. Chen P, Hu T, Liang Y, Li P, Chen X, Zhang J, et al. Neddylation inhibition activates the extrinsic apoptosis pathway through ATF4-CHOP-DR5 axis in human esophageal cancer cells. Clin Cancer Res. 2016;22:4145–57.
- 78. Liu X, Jiang Y, Wu J, Zhang W, Liang Y, Jia L, et al. NEDD8-activating enzyme inhibitor, MLN4924 (Pevonedistat) induces NOXA-dependent apoptosis through up-regulation of ATF-4. Biochem Biophys Res Commun. 2017;488:1–5.
- 79. Nakamura K, Smyth MJ. Myeloid immunosuppression and immune checkpoints in the tumor microenvironment. Cell Mol Immunol. 2020;17:1–12.
- 80. Gonzalez H, Hagerling C, Werb Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. Genes Dev. 2018;32:1267–84.
- 81. Chiu DK, Tse AP, Xu IM, Di Cui J, Lai RK, Li LL, et al. Hypoxia inducible factor HIF-1 promotes myeloid-derived suppressor cells accumulation through ENTPD2/ CD39L1 in hepatocellular carcinoma. Nat Commun. 2017;8:517.
- 82. Chang FM, Reyna SM, Granados JC, Wei SJ, Innis-Whitehouse W, Maffi SK, et al. Inhibition of neddylation represses lipopolysaccharide-induced proinflammatory cytokine production in macrophage cells. J Biol Chem. 2012;287:35756–67.
- 83. Wu D, Li H, Liu M, Qin J, Sun Y. The Ube2m-Rbx1 neddylation-Cullin-RING-Ligase proteins are essential for the maintenance of Regulatory T cell fitness. Nat Commun. 2022;13:3021.
- 84. Topalian SL, Taube JM, Pardoll DM. Neoadjuvant checkpoint blockade for cancer immunotherapy. Science. 2020; 367:eaax0182.
- 85. Yi M, Zheng X, Niu M, Zhu S, Ge H, Wu K. Combination strategies with PD-1/PD-L1 blockade: current advances and future directions. Mol Cancer. 2022;21:28.
- 86. Zhou S, Zhao X, Yang Z, Yang R, Chen C, Zhao K, et al. Neddylation inhibition upregulates PD-L1 expression and enhances the efficacy of immune checkpoint blockade in glioblastoma. Int J Cancer. 2019;145:763–74.
- 87. Zhang J, Bu X, Wang H, Zhu Y, Geng Y, Nihira NT, et al. Cyclin D-CDK4 kinase destabilizes PD-L1 via cullin 3-SPOP to control cancer immune surveillance. Nature 2018;553:91–5.
- 88. Zhang S, You X, Xu T, Chen Q, Li H, Dou L, et al. PD-L1 induction via the MEK-JNK-AP1 axis by a neddylation inhibitor promotes cancer-associated immunosuppression. Cell Death Dis. 2022;13:844.
- 89. Scott DC, Sviderskiy VO, Monda JK, Lydeard JR, Cho SE, Harper JW, et al. Structure of a RING E3 trapped in action reveals ligation mechanism for the ubiquitin-like protein NEDD8. Cell. 2014;157:1671–84.
- 90. Kurz T, Ozlu N, Rudolf F, O'Rourke SM, Luke B, Hofmann K, et al. The conserved protein DCN-1/Dcn1p is required for cullin neddylation in C. elegans and S. cerevisiae. Nature 2005;435:1257–61.
- 91. Scott DC, Monda JK, Grace CR, Duda DM, Kriwacki RW, Kurz T, et al. A dual E3 mechanism for Rub1 ligation to Cdc53. Mol Cell. 2010:39:784-96.
- 92. Zhou H, Lu J, Liu L, Bernard D, Yang CY, Fernandez-Salas E, et al. A potent smallmolecule inhibitor of the DCN1-UBC12 interaction that selectively blocks cullin 3 neddylation. Nat Commun. 2017;8:1150.
- 93. Zhou H, Zhou W, Zhou B, Liu L, Chern TR, Chinnaswamy K, et al. High-affinity peptidomimetic inhibitors of the DCN1-UBC12 protein-protein interaction. J Med Chem. 2018;61:1934–50.
- 94. Scott DC, Hammill JT, Min J, Rhee DY, Connelly M, Sviderskiy VO, et al. Blocking an N-terminal acetylation-dependent protein interaction inhibits an E3 ligase. Nat Chem Biol. 2017;13:850–7.
- 95. Hammill JT, Scott DC, Min J, Connelly MC, Holbrook G, Zhu F, et al. Piperidinyl ureas chemically control defective in cullin neddylation 1 (DCN1)-mediated cullin neddylation. J Med Chem. 2018;61:2680–93.
- 96. Hammill JT, Bhasin D, Scott DC, Min J, Chen Y, Lu Y, et al. Discovery of an orally bioavailable inhibitor of defective in cullin neddylation 1 (DCN1)-mediated cullin neddylation. J Med Chem. 2018;61:2694–706.
- 97. Wang S, Zhao L, Shi XJ, Ding L, Yang L, Wang ZZ.et al. Development of highly potent, selective, and cellular active triazolo[1,5- a]pyrimidine-based inhibitors targeting the DCN1-UBC12 protein-protein interaction. J Med Chem. 2019;62:2772–97.
- 98. Zhou W, Ma L, Ding L, Guo Q, He Z, Yang J, et al. Potent 5-cyano-6-phenylpyrimidin-based derivatives targeting DCN1-UBE2M interaction. J Med Chem. 2019;62:5382–403.
- 99. Xu S, Ma Y, Tong Q, Yang J, Liu J, Wang Y, et al. Cullin-5 neddylation-mediated NOXA degradation is enhanced by PRDX1 oligomers in colorectal cancer. Cell Death Dis. 2021;12:265.
- 100. Zhou L, Zhu J, Chen W, Jiang Y, Hu T, Wang Y, et al. Induction of NEDD8 conjugating enzyme E2 UBE2F by platinum protects lung cancer cells from apoptosis and confers to platinum-insensitivity. Cell Death Dis. 2020;11:975.
- 101. Kabir S, Cidado J, Andersen C, Dick C, Lin PC, Mitros T, et al. The CUL5 ubiquitin ligase complex mediates resistance to CDK9 and MCL1 inhibitors in lung cancer cells. eLife. 2019;8:e44288.
- 102. Zhou L, Dong C, Xu Z, Wang X, Zhang L, Chen S, et al. NEDD8-conjugating enzyme E2 UBE2F confers radiation resistance by protecting lung cancer cells from apoptosis. J Zhejiang Univ Sci B 2021;22:959–65.
- 103. Wang Q, Li H, Tajima K, Verkerke ARP, Taxin ZH, Hou Z, et al. Post-translational control of beige fat biogenesis by PRDM16 stabilization. Nature. 2022;609:151–158.
- 104. Chen C, Gu L, Matye DJ, Clayton YD, Hasan MN, Wang Y, et al. Cullin neddylation inhibitor attenuates hyperglycemia by enhancing hepatic insulin signaling through insulin receptor substrate stabilization. Proc Natl Acad Sci USA. 2022;119:e2111737119.
- 105. Ju UI, Jeong DW, Seo J, Park JB, Park JW, Suh KS, et al. Neddylation of sterol regulatory element-binding protein 1c is a potential therapeutic target for nonalcoholic fatty liver treatment. Cell Death Dis. 2020;11:283.
- 106. Zhang J, Cui J, Zhao F, Yang L, Xu X, Shi Y, et al. Cardioprotective effect of MLN4924 on ameliorating autophagic flux impairment in myocardial ischemiareperfusion injury by Sirt1. Redox Biol. 2021;46:102114.
- 107. Yu H, Luo H, Chang L, Wang S, Geng X, Kang L, et al. The NEDD8-activating enzyme inhibitor MLN4924 reduces ischemic brain injury in mice. Proc Natl Acad Sci USA. 2022;119:e2111896119.
- 108. Yu S, Xie L, Liu Z, Li C, Liang Y. MLN4924 exerts a neuroprotective effect against oxidative stress via sirt1 in spinal cord ischemia-reperfusion injury. Oxid Med Cell Longev. 2019;2019:7283639.

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# AUTHOR CONTRIBUTIONS

LSZ, XZL, and JZ collected the related paper and drafted the manuscript. LYZ and SYC searched the databases and analyzed the data. LSZ, HY, LJJ, and BFC revised and finalized the manuscript. All authors read and approved the final manuscript.

## COMPETING INTERESTS

The authors declare no competing interests.

# ADDITIONAL INFORMATION

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