



# Emerging Regulatory Mechanisms of N<sup>6</sup>-Methyladenosine Modification in Cancer Metastasis

Jing Zhao<sup>1,2</sup> · Hao Xu<sup>1,2</sup> · Yinghan Su<sup>1,2</sup> · Junjie Pan<sup>1,2</sup> · Sunzhe Xie<sup>1,2</sup> · Jianfeng Xu<sup>1,2</sup> · Lunxiu Qin<sup>1,2</sup>

Received: 15 August 2021 / Revised: 21 December 2021 / Accepted: 27 December 2021 / Published online: 25 May 2022  
© International Human Phenome Institutes (Shanghai) 2022

## Abstract

Cancer metastasis is the major cause of cancer-related deaths and accounts for poor therapeutic outcomes. A metastatic cascade is a series of complicated biological processes. N<sup>6</sup>-methyladenosine (m<sup>6</sup>A) is the most abundant and conserved epitranscriptomic modification in eukaryotic cells, which has great impacts on RNA production and metabolism, including RNA splicing, processing, degradation and translation. Accumulating evidence demonstrates that m<sup>6</sup>A plays a critical role in regulating cancer metastasis. However, there is a lack of studies that review the recent advances of m<sup>6</sup>A in cancer metastasis. Here, we systematically retrieved the functions and mechanisms of how the m<sup>6</sup>A axis regulates metastasis, and especially summarized the organ-specific liver, lung and brain metastasis mediated by m<sup>6</sup>A in various cancers. Moreover, we discussed the potential application of m<sup>6</sup>A modification in cancer diagnosis and therapy, as well as the present limitations and future perspectives of m<sup>6</sup>A in cancer metastasis. This review provides a comprehensive knowledge on the m<sup>6</sup>A-mediated regulation of gene expression, which is helpful to extensively understand the complexity of cancer metastasis from a new epitranscriptomic point of view and shed light on the developing novel strategies to anti-metastasis based on m<sup>6</sup>A alteration.

**Keywords** Cancer metastasis · m<sup>6</sup>A · Epitranscriptomic modification · RNA metabolism · Organ-specific metastasis

## Abbreviations

m <sup>6</sup> A	N <sup>6</sup> -methyladenosine	WTAP	Wilms tumor 1-associated protein
m <sup>1</sup> A	N <sup>1</sup> -methyladenosine	FTO	Obesity-associated protein
m <sup>3</sup> C	3-Methylcytosine	ALKBH5	AlkB homolog 5
m <sup>5</sup> C	5-Methylcytosine	IGF2BP	Insulinlike growth factor 2 mRNA binding protein family
m <sup>1</sup> G	N <sup>1</sup> -methylguanosine	HNRNPA2B1	Heterogeneous nuclear ribonucleoproteins A2/B1
m <sup>7</sup> G	7-Methylguanosine	PRRC2A	Prolinerich and coiledcoilcontaining protein 2A
ac4C	N <sup>4</sup> -acetylcytidine	CNV	Copy number variation
EMT	Epithelial-mesenchymal transition	OS	Overall survival
VEGF	Vascular endothelial growth factor	DFS	Disease-free survival
HPCs	Hematopoietic progenitor cells	HNSCC	Head and neck squamous cell carcinoma
SDF-1	Stromal cell-derived factor-1	UCEC	Uterine corpus endometrial carcinoma
MMP-9	Matrix metalloproteinase 9	ccRCC	Clear cell renal cell carcinoma
CTCs	Circulating tumor cells	DNMT2	DNA methyltransferase-like 2
METTL3	Methyltransferase-like 3	HCC	Hepatocellular carcinoma
		CRC	Colorectal cancer
		GC	Gastric cancer
		PDAC	Pancreatic ductal adenocarcinoma
		CC	Cervical cancer
		CSC	Cancer stem cell

Jing Zhao and Hao Xu have contributed equally to this work.

✉ Lunxiu Qin  
qinlx@fudan.edu.cn

<sup>1</sup> Department of General Surgery, Huashan Hospital, Fudan University, 12 Urumqi Road (M), Shanghai 200040, China

<sup>2</sup> Cancer Metastasis Institute, Fudan University, Shanghai 200120, China

## Introduction

Cancer has ranked as the second leading cause of death worldwide, which has become a global burden and threat to human health. Cancer metastasis is the principal cause of high mortality rate, and approximately over 90% of cancer patients die of metastasis. Metastasis is a complex biological process including epithelial–mesenchymal transition (EMT), angiogenesis, intravasation and extravasation, and ultimately metastatic outgrowth. Despite great advances in cancer biology, our current knowledge on cancer metastasis is still poor. At present, there is still a lack of available measures to monitor early metastasis and target cancer metastasis for effective therapy. Therefore, it is necessary to enrich our understanding of the progression of cancer metastasis.

Accumulating evidence demonstrates that cancer cells undergo a series of complicated genetic, epigenetic and epitranscriptomic alterations during the process of metastasis. A variety of genomic variations, such as DNA methylation, histone modifications and chromatin remodeling, have been extensively studied in cancer development. With the advances of next-generation sequencing, RNA modifications, such as N<sup>6</sup>-methyladenosine (m<sup>6</sup>A), N<sup>1</sup>-methyladenosine (m<sup>1</sup>A), 3-methylcytosine (m<sup>3</sup>C), 5-methylcytosine (m<sup>5</sup>C), N<sup>1</sup>-methylguanosine (m<sup>1</sup>G), 7-methylguanosine (m<sup>7</sup>G), and N<sup>4</sup>-acetylcytidine (ac4C), have come to prominence and become a hotspot field in cancer (Barbieri and Kouzarides 2020; Roundtree et al. 2017). These highly decorated RNAs with different modifiers are an efficient pathway to regulate gene expression and execute biological functions. Among these chemical RNA modifications, N<sup>6</sup>-methyladenosine (m<sup>6</sup>A) is the most abundant and conserved epitranscriptomic alteration in eukaryotic cells, which plays a critical role in cancer metastasis.

In this review, we first systematically summarized the mechanisms of how m<sup>6</sup>A regulates metastasis in different cancers. Moreover, we also highlighted the limits and perspectives of m<sup>6</sup>A-related researches and discussed the potential application of the m<sup>6</sup>A axis in cancer diagnosis and therapy.

## Cancer Metastasis Is a Complicated Process

Metastasis is the major cause of lethality for cancer patients. Despite the great advances in cancer biology, when and how cancer metastasis occurs remain largely unknown. Cancer metastasis is an extremely complicated process with multiple factors participating in it and

multiple pathways being regulated. First, depolarization is triggered and cancer cells undergo EMT, acquiring invasive properties. Next, the cells successively promote angiogenesis for intravasation to the blood vessels. After successfully surviving the attacks in the circulation, the cells go through extravasation, reside on distant organs, and eventually form metastatic lesions (Gao et al. 2019; Obenauf and Massagué 2015; Suhail et al. 2019).

Cancer metastasis is not a random procedure. Clinically, the organ specificity of metastasis has been recognized early. For example, breast cancers have a propensity to metastasize to bone, small cell lung cancers are prone to brain metastasis, and colon cancers prefer to metastasize in the liver. Along this line, metastasis is highly purposeful and selective (Peinado et al. 2017). In 1889, Paget proposed the “seed-and-soil” hypothesis, which suggested that cancer cells (seeds) can live and grow only when they fall on congenial soil (Paget 1889). Based on this observation, numerous researchers have conducted more in-depth studies on the mechanism of organ-specific metastasis, and proposed that metastasis requires the coordination of cancer cells and the microenvironment. During the process, cancer cells not only gradually adapt to the new microenvironment, but also modify the environment via complex interaction through cytokines, metabolites or certain phenotypical features (Bos et al. 2009; Hoshino et al. 2015; Jin et al. 2020b; Kaplan et al. 2005). Previous analyses have indicated that the subtypes with different histological or molecular characteristics tend to colonize at different locations (Fumagalli et al. 2020; Laughney et al. 2020). The dynamic bidirectional process may be the essence of organ-specific metastasis. In addition to the famous “seed-and-soil theory”, Kaplan et al. first proposed the “pre-metastatic niches” hypothesis in 2005, which suggested that primary tumors induce the formation of pre-determined microenvironments in distant sites to facilitate survival and proliferation of the unreached cancer cells (Kaplan et al. 2005). For instance, the soluble factor, vascular endothelial growth factor (VEGF), is released from the primary tumor and then enters into the circulatory system. After arriving at the remote target organ, VEGF induces the inherent fibroblasts to produce fibronectin, which can mobilize and recruit VEGFR<sup>+</sup>VLA-4<sup>+</sup> hematopoietic progenitor cells (HPCs) to the target organ. In this microenvironment, the HPCs interact with the fibroblasts and increase the expression of stromal cell-derived factor-1 (SDF-1) and matrix metalloproteinase 9 (MMP-9). SDF-1 promotes the adhesion of CXCR4<sup>+</sup> circulating tumor cells (CTCs) at the target site, while MMP-9 is beneficial for remodeling the local microenvironment to make it more suitable for the colonization and growth of CTCs (Kaplan et al. 2005). Subsequently, with the efforts of the researches, some other tumor-derived soluble factors, membrane vesicles, exosomes, and recruited bone marrow-derived cells with functions in line

with the “pre-metastatic niches” hypothesis have been successively identified and further confirmed (Liu et al. 2016b; Murgai et al. 2017; Olmeda et al. 2017; Zeng et al. 2018). The new concept is consistent with the previous “seed-and-soil” hypothesis, but it extends a more dynamic perspective of metastasis that cancer cells have not yet arrived at the target lesions. In this concept, the primary tumor sends special envoys to the specific site, catalyzing the formation of a specific “soil” (pre-metastatic niche), which determines the adhesion, colonization, and growth of CTCs and becomes the critical “speed-limiting” node for the formation of target organ metastases.

Recently, tumor dormancy, highly consistent with the “seed-and-soil” hypothesis, has been emphasized in metastasis by numerous researchers (Phan and Croucher 2020). As early as 1954, Hadfield used the term “dormancy” to describe malignant cancer cells that survive for a long time without significant proliferation (Hadfield 1954). The definition of tumor dormancy covers two scenarios: a solitary cell that enters the G0 phase to undergo cell cycle arrest; or a small cluster of cells having a constant population due to an equal rate of proliferation and apoptosis (Phan and Croucher 2020). Although the two scenarios are more or less the result of the interaction between the tumor and its micro-environment, single-cell dormancy that remains dormant or non-proliferating in unsuitable soil is more consistent with the “seed-and-soil” hypothesis. However, unlike the static perspective proposed by Paget (Paget 1889), dormant cells, after going through the incubation period, can be reactivated and proliferate to metastasize with clinical manifestations (Correia et al. 2021; Vera-Ramirez et al. 2018).

Many studies have indicated that cancer cells undergo a series of complicated genetic, epigenetic and epitranscriptomic alterations during metastasis, such as the well-known genomic variations, DNA methylation, histone modifications and chromatin remodeling (Audia and Campbell 2016; Calabrese et al. 2020; Jones et al. 2016; Klutstein et al. 2016; Koch et al. 2018). Recently, epitranscriptomic alterations, especially the m<sup>6</sup>A modification of RNA, have become a novel scientific hotspot in the cancer metastasis field.

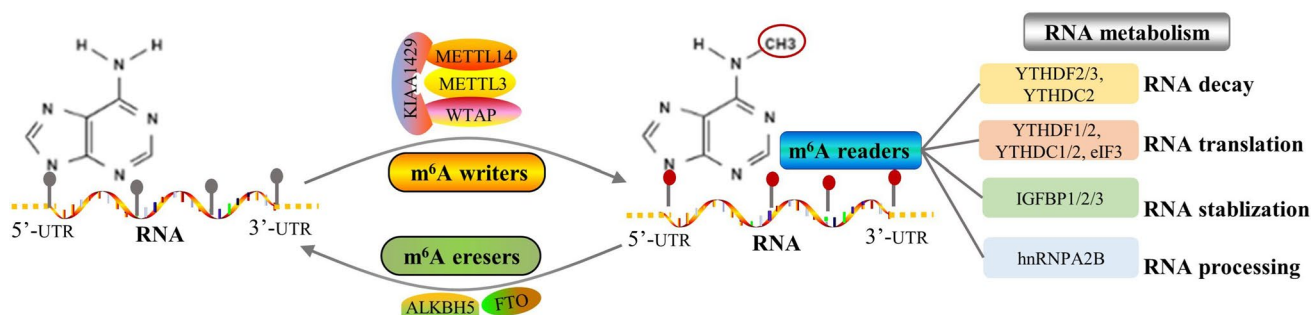
## Critical Components of the m<sup>6</sup>A Modification Machinery

There are hundreds of chemical modifications that have been identified in RNA, such as N<sup>1</sup>-methyladenosine (m<sup>1</sup>A), N<sup>6</sup>-methyladenosine (m<sup>6</sup>A), 5-methylcytosine (m<sup>5</sup>C), 3-methylcytosine (m<sup>3</sup>C), N<sup>1</sup>-methylguanosine (m<sup>1</sup>G), N<sup>7</sup>-methylguanosine (m<sup>7</sup>G), N4-acetylcytidine (ac4C) (Barbieri and Kouzarides 2020). With the improvement of high-throughput sequencing approaches, the m<sup>6</sup>A decoration has

become the best characterized epitranscriptomic alteration at present (Barbieri and Kouzarides 2020).

It is reported that m<sup>6</sup>A is the most abundant and conserved RNA modification in eukaryotic cells. This type of RNA modification is catalyzed by an installed m<sup>6</sup>A machinery composed of multiple methyltransferases, demethylases and m<sup>6</sup>A-binding proteins (He et al. 2019). Methyltransferases are usually called as “writers” that are responsible for methylating the N<sup>6</sup> of adenosine. At present, the well-studied m<sup>6</sup>A writers contain methyltransferase-like 3 (METTL3), METTL14, METTL16, Wilms tumor 1-associated protein (WTAP), RBM15/15B and KIAA1429. The m<sup>6</sup>A modification is a dynamic and reversible process that can be removed by specific RNA demethylases, such as fat mass and obesity-associated protein (FTO) and alkB homolog 5 (ALKBH5). Herein, FTO and ALKBH5 are also termed as “erasers” of m<sup>6</sup>A. The functions and mechanisms of m<sup>6</sup>A modifications are usually recognized and deciphered by various m<sup>6</sup>A RNA-binding proteins called “readers”. Many protein family members have been identified as m<sup>6</sup>A “readers”, such as the members of YTH domain-containing family (YTHDFs and YTHDCs), insulin-like growth factor 2 mRNA-binding protein family (IGF2BP), eukaryotic initiation factor EIF3, heterogeneous nuclear ribonucleoproteins A2/B1 (HNRNPA2B1) and prolinerich and coiledcoilcontaining protein 2A (PRRC2A). YTH domains can directly bind to the m<sup>6</sup>A site of the RNA, and the other readers may bind to the surrounding unfolded RNA. The “writer–eraser–reader” system of m<sup>6</sup>A can determine the fate of the target RNA through regulating its transcription, processing, splicing, RNA stability and translation. The m<sup>6</sup>A regulation is described in Fig. 1.

There are some genomic alterations on m<sup>6</sup>A regulators. In hepatocellular carcinoma, we have reported that m<sup>6</sup>A regulator genes undergo a prevalent alteration of copy number variation (CNV) via analyzing the TCGA database. *YTHDF3*, *CBL1*, *IGF2BP1/3*, *HNRNPA2B1*, *KIAA1429*, and *YTHDF1* are found to display high frequency of CNVs in HCC (Shen et al. 2020). Wang et al. reported that HCC patients with any mutation of the m<sup>6</sup>A regulators may suffer from shorter overall survival (OS) and disease-free survival (DFS) (Wang et al. 2020d), and *METTL16* or *ALKBH5* deletion may predict poor OS and DFS in HCC (Wang et al. 2020d). In some other tumors, such as head and neck squamous cell carcinoma (HNSCC), uterine corpus endometrial carcinoma (UCEC), clear cell renal cell carcinoma (ccRCC) and bladder urothelial carcinoma, m<sup>6</sup>A regulators are characterized by rare somatic mutations. Although most of the writer and eraser genes tend toward loss of copy number in HCC and HNSCC, the reader genes, such as *YTHDC1*, *YTHDC2*, *YTHDF3* and *IGF2BP2*, tend toward gain of copy number in HNSCC, UCEC and ccRCC (Wang et al. 2021b; Wang et al. 2020g; Zhou et al. 2019; Zhou et al. 2020). However, whether these genomic alterations might contribute to



**Fig.1** The regulatory machinery of  $m^6A$  modification. The diagram showed the  $m^6A$  machinery. The  $m^6A$  writers mainly contain METTL3, METTL14, WTAP and KIAA1429, which are responsible for methylating at the  $N^6$  position of adenosine. The  $m^6A$  eraser is composed of RNA demethylases such as FTO and ALKBH5

aberrant expression and dysfunctions of these  $m^6A$  regulators needs to be further illustrated.

In contrast to the  $m^6A$ , the other modifications of RNA have not been extensively studied due to the limitation of sequencing methods and the lack of specific antibodies. For instance, the  $m^1A$  modification, which is mainly found in tRNA and rRNA but few in cytosolic mRNA, usually depends on TRMT10A and TRM61 complex to add methyl group. The YTH protein family is involved in recognizing the  $m^1A$  modification, whereas ALKBH1 and ALKBH3 are responsible for removing the methyl group (Chen et al. 2019; Dai et al. 2018; Liu et al. 2016a; Saikia et al. 2010). The  $m^5C$  is found in a wide range of RNAs, including rRNA, tRNA, mRNA, ncRNA and enhancer RNA. Recent investigation showed that the  $m^5C$ , especially in tRNA, mainly functions as a structural stability regulator to promote translation accuracy (Yang et al. 2017). NSUN family members (NSUN1 to NSUN7) and DNA methyltransferase-like 2 (DNMT2) are involved in  $m^5C$  methylation, and ALYREF accounts for binding and recognizing the modification, but which serves as the eraser of  $m^5C$  remains largely unknown. As for the  $m^3C$  modification, there are four  $m^3C$  methyltransferase-like proteins (METTL2A, METTL2B, METTL6, and METTL8) and two  $m^3C$  demethylases (ALKBH3 demethylating tRNAs and ALKBH1 demethylating mRNA) that have been reported (Cui et al. 2021). The  $m^1G$  alteration is found mainly in eukaryotic tRNAs, and is frequently catalyzed by Trm5 and Trm10 at the position 37 and at the position 9, respectively (Jin et al. 2019b). The  $m^7G$  is identified in mRNA, tRNA and rRNA, and may participate in the cap structure formation and protein translation (Barbieri and Kouzarides 2020). Alexandrov et al. found that  $m^7G$  on tRNA is triggered by the METTL1-WDR4 complex, while  $m^7G$  in rRNA is mediated by WBSR22 protein (Alexandrov et al. 2002; Haag et al. 2015). The readers and erasers of  $m^7G$  need to be further elucidated (Ramanathan et al. 2016). In

addition to the methylation on RNAs, the ac4C modification is the first acetylation event that is found in mRNA, and successively reported in tRNAs and rRNAs. And it is catalyzed by a single enzyme NAT10. A recent study addressed that the ac4C on mRNA can promote mRNA stability and enhance the translation efficiency (Arango et al. 2018).

Accumulating evidence has demonstrated that the  $m^6A$  machinery plays important roles in several physiological and pathological pathways, which have been extensively investigated in embryogenesis, neurogenesis and cancer development (He et al. 2019; Yoon et al. 2017; Zhao et al. 2017). Some of the components of the  $m^6A$  machinery have promising potentials for clinical translation to become ideal diagnostic biomarkers and therapeutic targets. In this review, we mainly focus on the regulatory functions and mechanism of  $m^6A$  in cancer metastasis.

to remove  $m^6A$  modification from target RNAs. The  $m^6A$  reader is essential to recognize and decipher  $m^6A$  modification, which may determine RNA fate to undergo decay, translation, RNA stabilization or RNA processing. The well-documented  $m^6A$  readers include YTHDF1/2/3, YTHDC1/2/3, IGF2BP1/2/3, and hnRNPA2B1

## The Functions and Mechanisms of $m^6A$ Modification in Metastasis of Various Cancers

### $m^6A$ Modification and Hepatocellular Carcinoma Metastasis

Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide with high incidence and mortality. Metastasis accounts for the majority of deaths in HCC. The  $m^6A$  modification plays an essential role in the metastasis of HCC.

The  $m^6A$  writer METTL14 is downregulated in HCC, especially in the metastatic samples. METTL14 can form a complex with the microprocessor protein DGCR8 to regulate  $m^6A$ -dependent *miR-126* processing, which inhibits HCC metastasis (Ma et al. 2017). Another study showed that EGFR is the direct target of METTL14. METTL14 can block EGFR/PI3K/AKT signaling axis to suppress



EMT, metastasis and invasion of HCC (Shi et al. 2020b). KIAA1429 is another critical methyltransferase involved in m<sup>6</sup>A modifications. KIAA1429 promotes tumor growth and metastasis by downregulating GATA3 expression in HCC. Mechanistically, KIAA1429 can disassociate HuR from GATA3 pre-mRNA by inducing m<sup>6</sup>A modification at the 3' UTR, destabilizing GATA3 pre-mRNA and reducing translation. Moreover, *lncRNA GATA3-As*, derived from the anti-sense GATA3 transcript, facilitates the association between KIAA1429 and GATA3 pre-mRNA, which further enhance the pro-metastatic capacity of KIAA1429 (Lan et al. 2019). *CircDLC1* is another target of KIAA1429-mediated m<sup>6</sup>A in HCC, which inhibits metastasis via abolishing the interaction between HuR and *MMP1* mRNA, thus reducing MMP1 expression (Liu et al. 2021).

The m<sup>6</sup>A readers from the YTHDF protein family are widely involved in HCC metastasis. For instance, YTHDF1 binds to m<sup>6</sup>A-modified *ATG2A* and *ATG14* mRNAs to increase their translation, thereby inducing autophagy and metastasis under hypoxia (Li et al. 2021a). A recent study reported that sublethal heat stress can elevate m<sup>6</sup>A binding near the 5'UTR of the *EGFR* mRNA, which facilitates the association of YTHDF1 to improve EGFR protein output. The activation of the m<sup>6</sup>A-YTHDF1-EGFR axis contributes to HCC metastasis after insufficient radiofrequency ablation treatment (Su et al. 2021). The other homolog, YTHDF2, modulates m<sup>6</sup>A binding at 5'UTR of the *OCT4* mRNA to augment OCT4 translation, thus sustaining cancer stem cell properties and promoting metastasis in HCC (Zhang et al. 2020a). Wang et al. demonstrated that YTHDF3 can stabilize *Zeb* mRNA to induce HCC cell migration in an m<sup>6</sup>A-dependent manner (Wang et al. 2020c).

The m<sup>6</sup>A erasers also extensively participate in HCC metastasis via inducing mRNA demethylation. For example, FTO can modulate the m<sup>6</sup>A demethylation of cancer stem cell (CSC)-related genes including *SOX2*, *KLF4* and *NANOG*, which increases the expressions of these genes to maintain stemness and metastasis (Bian et al. 2021). The upregulated oncoprotein AMD1 stabilizes the interaction between *IQGAP1* and FTO, to prevent ubiquitination-mediated degradation of FTO, thereby enhancing HCC metastasis (Bian et al. 2021).

### **M<sup>6</sup>A Methylation and Colorectal Cancer Metastasis**

Colorectal cancer (CRC) is the second leading cause of cancer-related deaths because of the high rate of metastasis and recurrence (Bray et al. 2018). Emerging investigations indicate that m<sup>6</sup>A modification and regulators exert pivotal roles in modulating the metastasis of CRC.

Similar to HCC, the m<sup>6</sup>A writer METTL14 is dramatically downregulated in CRC, which is associated with shorter survival. METTL14 mediates m<sup>6</sup>A methylation of

*SOX4* mRNA and facilitates the degradation of m<sup>6</sup>A-*SOX4* mRNA in a YTHDF2-dependent manner. *SOX4* has an oncogenic capacity to drive EMT and migration by activating the PI3K/AKT signaling pathway. Decreased expression of METTL14 markedly promotes invasion and migration of CRC via increasing *SOX4* expression (Chen et al. 2020a). This study also found that histone demethylase KDM5C-mediated demethylation of H3K4me3 at the METTL14 promoter may account for the low transcription of METTL14 in CRC (Chen et al. 2020a). Another study reported that MeCP2 can associate with METTL14 to decrease m<sup>6</sup>A modification of the tumor suppressor *KLF14* mRNA, thus reducing *KLF14* expression and promoting CRC metastasis (Wang et al. 2021a). METTL14 can mediate m<sup>6</sup>A modification of non-coding RNA, as well as coding mRNAs, in CRC progression. For example, low expression of METTL14 remarkably abolishes m<sup>6</sup>A deposition of *lncRNA XIST* and increases *XIST* expression to promote tumorigenicity and metastasis of CRC cells (Yang et al. 2020a). METTL14 can modulate the processing of *miR-375* via m<sup>6</sup>A methylation, thereby increasing the expression of *miR-375* targets, including YAP1 and SP1 to facilitate the metastasis of CRC (Chen et al. 2020b).

METTL3 serves as an important m<sup>6</sup>A writer that is frequently upregulated in CRC and promotes metastasis via m<sup>6</sup>A methylation (Hou et al. 2021a; Li et al. 2019). *SOX2* and *HMGAI* are the targets of METTL3. The m<sup>6</sup>A reader, IGF2BP2, can recognize m<sup>6</sup>A-*SOX2* RNA and m<sup>6</sup>A-*HMGAI* to increase their protein output, thereby driving CRC metastasis (Hou et al. 2021a; Li et al. 2019). Additionally, METTL3 can induce *circ1662* expression by increasing m<sup>6</sup>A modifications in its flanking region. *Circ1662* can bind to YAP1 and accelerate its nuclear accumulation to increase SMAD3 expression, thereby promoting invasion and migration of CRC (Chen et al. 2021a). Wu et al. reported that m<sup>6</sup>A-induced *lncRNA RP11* mediates the dissemination of CRC cells (Wu et al. 2019). METTL13 is responsible for the m<sup>6</sup>A deposition and the demethylase ALKBH5 can reduce RP11 expression. The m<sup>6</sup>A reader hnRNPA2B1 forms a complex with *lncRNA Rpl11* and other target mRNAs, such as two E3 ligases *Siah-1* and *Fbx45* mRNAs, and subsequently reduces their translation. As a result, ZEB1 is protected from ubiquitin-mediated degradation by E3 ligases *Siah-1* and *Fbx45* to trigger metastasis (Wu et al. 2019).

### **M<sup>6</sup>A Modification and Gastric Cancer Metastasis**

Gastric cancer (GC) is characterized as one of the most invasive malignancies, ranking as the third most deadly cancer worldwide (Bray et al. 2018). The regulatory roles of m<sup>6</sup>A modification in GC metastasis have attracted increasing attention.

The m<sup>6</sup>A writer METTL3 has been extensively studied in GC metastasis and progression. The oncogene *ZMYM1* was identified as a target of METTL3 for m<sup>6</sup>A modification, and the m<sup>6</sup>A-*ZMYM1* mRNA is recognized by the reader HuR to enhance its stability and argument translation. *ZMYM1* recruits CtBP/LSD1/CoREST transcriptional complex to repress E-cadherin expression, and thus induces EMT and metastasis in GC (Yue et al. 2019). Recently, Wang et al. revealed that METTL3 was involved in tumor growth and liver metastasis of GC (Wang et al. 2020e). Mechanistically, METTL3 modulates m<sup>6</sup>A deposition of *HDGF* mRNA, and the reader protein IGF2BP3 enhances *HDGF* mRNA stability to increase the protein expression. The secreted HDGF facilitates angiogenesis, while the nuclear HDGF transcriptionally activates glycolysis by driving GLUT4 and ENO2 expression (Wang et al. 2020e). In addition to increasing oncogene expression, METTL3 decreases tumor suppressor gene expression levels. The tumor suppressor BATF2 interacts with and improves P53 protein stability to inhibit ERK phosphorylation and activation. METTL3 reduces BATF2 expression in an m<sup>6</sup>A-dependent pathway, thereby activating the ERK pathway and inducing metastasis of GC (Xie et al. 2020b). Of note, some upstream regulators are responsible for METTL3 activation in GC. For instance, P300-mediated H3K27 acetylation within the METTL3 promoter region can trigger its transcription (Wang et al. 2020e). *MiR-338-5p* targets METTL3 reduction to inhibit metastasis, but *miR-338-5p* frequently undergoes methylated silence by EED in GC progression (Zhang et al. 2021).

Besides m<sup>6</sup>A readers HuR and IGF2BP3, high-expressed reader YTHDF1 also participates in metastasis of GC and indicates poor prognosis and shorter survival time (Chen et al. 2021d). YTHDF1 can facilitate *USP14* translation via the m<sup>6</sup>A-modification, thereby inducing tumor growth and metastasis (Chen et al. 2021d).

The m<sup>6</sup>A erasers ALKBH5 and FTO exert vital roles in promoting metastasis of GC. ALKBH5 can bind to and demethylate m<sup>6</sup>A modification of the *lncRNA NEAT1*, which increases the expression of the oncogene *EZH2* to facilitate metastasis in GC (Zhang et al. 2019). A recent study reported that WNT7B reduces FTO expression to elevate *TCF7L2* mRNA expression in an m<sup>6</sup>A-dependent manner, stimulating the Wnt/ $\beta$ -catenin signaling axis to reinforce *WNT7B* expression (Gao et al. 2021). The positive feedback loop WNT7B/m<sup>6</sup>A-TCF7L2/ $\beta$ -catenin pathway firmly promotes GC cancer progression and metastasis (Gao et al. 2021).

### **M<sup>6</sup>A Deposition and Pancreatic Cancer Metastasis**

Pancreatic cancer is one of the most lethal cancers with a 5-year survival rate of no more than 5% (Bray et al. 2018). The m<sup>6</sup>A deposition is markedly increased in 70% of the

pancreatic cancer specimens and exerts critical roles in regulating metastasis (Wang et al. 2020b).

The m<sup>6</sup>A writer METTL14 is upregulated and stimulates tumor growth and metastasis in pancreatic cancer. Further studies indicated that METTL14 mediates the m<sup>6</sup>A modification of *PERP* to accelerate mRNA turnover and decrease PERP protein level, thereby aggravating cancer metastasis and progression (Wang et al. 2020b). The reader YTHDC1 inhibits tumorigenesis and metastasis through the *miR-30d*/RUNX1/SLC2A/HK pathway in pancreatic cancer (Hou et al. 2021b). Mechanistically, the downregulation of YTHDC1 reduces the biogenesis of *miR-30d* in an m<sup>6</sup>A-dependent manner, and thus triggers the RUNX1-induced Warburg effect to induce tumor growth and metastasis in pancreatic cancer (Hou et al. 2021b). The m<sup>6</sup>A eraser ALKBH5 is downregulated in pancreatic cancer, which can inhibit motility via demethylating the *lncRNA KCN15-AS1* (He et al. 2018).

A recent study found that aberrant alternative splicing can regulate m<sup>6</sup>A activation in pancreatic ductal adenocarcinoma (PDAC) (Chen et al. 2021c). CLK1 kinase mediates the phosphorylation of SR-like splicing factors<sup>5250-Ser</sup> (SRSF5<sup>250-Ser</sup>), which suppresses METTL14<sup>exon10</sup> and Cyclin L2<sup>exon6.3</sup> skipping events. The aberrant splicing granted high METTL14 stronger activity to enhance m<sup>6</sup>A modifications and facilitate metastasis. Meanwhile, the aberrant Cyclin L2 splicing promotes proliferation and tumor growth in PDAC (Chen et al. 2021c).

### **M<sup>6</sup>A Methylation and Breast Cancer Metastasis**

Breast cancer has become the second leading cause of cancer-associated mortality among women worldwide (Bray et al. 2018). The 5-year survival rate of metastatic breast cancer patients is only approximately 25%. Breast cancer prefers to metastasize to lung and brain. The roles of m<sup>6</sup>A modification in breast cancer metastasis have been extensively investigated.

In a breast cancer cell model with a high potential of lung metastasis, the m<sup>6</sup>A writer METTL3 displays increased expression, whereas the m<sup>6</sup>A eraser FTO is downregulated (Chen et al. 2021b). KRT7 was identified as the critical effector of m<sup>6</sup>A-mediated lung metastasis of breast cancer. A mechanistic study demonstrated that METTL3 induces m<sup>6</sup>A methylation at the A877 residue of *KRT7-AS*, which stabilizes the *KRT7-AS* and *KRT7* mRNA duplex via IGF2BP1/HuR complexes. Additionally, the downregulation of FTO enhances the methylation of the A950 residue at the exon 6 of *KRT7* to promote translation elongation by recruiting YTHDF1 and eEF1 factors toward *KRT7* mRNA (Chen et al. 2021b). The cancer stem cell regulator SOX2 is the downstream effector of METTL3 in breast cancer as well (Xie et al. 2021). METTL3 induces m<sup>6</sup>A deposition of the *SOX2*

mRNA and increases its protein production via the IGF1R reader, which enhances the cancer stem cell properties and promotes invasion and migration of breast cancer (Xie et al. 2021). In contrast to the aforementioned upregulatory and oncogenic roles of METTL3, Shi et al. found that METTL3 is downregulated in breast cancer, which decreases the m<sup>6</sup>A level of *COL3A1* and increases its expression, leading to metastasis of breast cancer cells (Shi et al. 2020a). These controversial results may be attributable to the highly heterogeneous characterization and different subtypes of breast cancer.

The m<sup>6</sup>A reader YTHDF3 exerts an important role in modulating the interplay of cancer cells with the brain microenvironment and ultimately leads to brain metastasis (Chang et al. 2020). Mechanistically, the upregulation of YTHDF3 promotes the translation of metastasis-related factors, including *ST6GALNAC5*, *GJA1*, and *EGFR*, in an m<sup>6</sup>A-dependent manner. These factors help cancer cells to break the blood–brain barrier, stimulate angiogenesis and grow in brain. The high expression of YTHDF3 in metastatic breast cancer is due to its gene copy number amplification and YTHDF3 self-regulation through m<sup>6</sup>A-dependent translation at its 5'UTR (Chang et al. 2020).

The m<sup>6</sup>A erasers ALKBH5 and FTO have impacts on the metastasis of breast cancer. The hypoxia-inducible factor triggers ALKBH5 transcription and expression. Consequently, overexpression of ALKBH5 decreases *NANOG* mRNA methylation at the 3'-UTR and increases *NANOG* protein levels, which promotes the cancer stem cell properties and aggravates breast cancer progression (Zhang et al. 2016). FTO is upregulated in breast cancer and associated with a poor prognosis. FTO induces the demethylation of tumor suppressor *BNIP3* mRNA in its 3'UTR and triggers its degradation, promoting tumor growth and metastasis (Niu et al. 2019). Additionally, FTO modulates invasion and migration by inhibiting *miR-181b*-targeted silencing of oncogene *ARL5B* (Xu et al. 2020).

### **M<sup>6</sup>A Modification and Lung Cancer Metastasis**

Lung cancer remains the leading cause of cancer-related deaths and has become a seriously global health problem (Bray et al. 2018). The functions of m<sup>6</sup>A modification have attracted more and more attention in the metastasis of lung cancer.

The m<sup>6</sup>A writer METTL3 can promote the biogenesis of precursor *miR-143-3p*, relying on m<sup>6</sup>A methylation, and *miR-143-3p* is upregulated in the brain metastasis samples of lung cancer (Wang et al. 2019a). M<sup>6</sup>A-modified *miR-143-3p* decreases *VASH1* expression and subsequently protects *VEGFA* protein from *VASH1*-mediated proteasome degradation, and thus triggers invasion and angiogenesis, breaking the blood–brain barrier, and promoting brain metastasis of

lung cancer (Wang et al. 2019a). A recent study found that METTL3 induces metastasis and chemotherapeutic resistance by facilitating *YAP* mRNA stability and translation in lung cancer (Jin et al. 2019a). Mechanistically, METTL3 can directly initiate m<sup>6</sup>A modification of *YAP* mRNA and recruit YTHDF1/3 and eIF3b to the translation complex to enhance *YAP* translation. In addition, METTL3 catalyzes m<sup>6</sup>A deposition of the *lncRNA MALAT1* to stabilize *MALAT1* mRNA. *MALAT1* acts as the competing endogenous RNA to sponge *miR-1914-3p*, thereby increasing *YAP* mRNA stability. The m<sup>6</sup>A-mediated dual regulation of *YAP* expression contributes to metastasis and aggravates lung cancer progression (Jin et al. 2019a). Another investigation also confirmed the critical role of m<sup>6</sup>A in regulating *YAP* expression to affect lung cancer metastasis (Jin et al. 2020a). This study indicated that the low expression of *ALKBH5* reduces m<sup>6</sup>A modification levels of *YAP* mRNA and decreases *YAP* translation and activation depending on YTHDF2. Moreover, *ALKBH5* interacts with HuR to augment *LAST2* expression by protecting it from *miR-1914-3p*-mediated degradation, which increases the phosphorylation of *YAP* and inhibits the activity of the *YAP* axis, blocking metastasis of lung cancer (Jin et al. 2020a).

The m<sup>6</sup>A reader YTHDC2 is downregulated in lung cancer and is associated with poor differentiation, lymph node metastasis and advanced TNM stage. YTHDC2 can suppress the proliferation and migration of lung cancer cells (Sun et al. 2020). Li et al. found that the m<sup>6</sup>A reader YTHDF2 is upregulated in lung adenocarcinoma tissues. YTHDF2 promotes tumorigenesis and metastasis by accelerating *AXIN1* decay and consequently activating the Wnt/β-catenin signaling cascade (Li et al. 2021b).

The m<sup>6</sup>A erasers are involved in the metastasis of lung cancer. For instance, FTO can enhance *NELL2* expression to trigger metastasis by declining *E2F1* m<sup>6</sup>A modification levels in lung cancer (Wang et al. 2021c). Guo et al. demonstrated that *ALKBH5*-mediated low m<sup>6</sup>A level facilitates the maintenance of the stability of *UBE2C* mRNA. The oncogenic *UBE2C* induces metastasis by repressing autophagy (Guo et al. 2018).

### **M<sup>6</sup>A Modification and Urological Malignancies**

The m<sup>6</sup>A axis plays critical roles in regulating metastasis of urological cancers such as prostate cancer, bladder cancer and renal cell carcinoma.

Prostate cancer has ranked the most common malignancy among men worldwide. Metastasis is the main risk factor leading to high mortality, and about 80% of metastatic prostate cancer cases may present with bone metastasis. Wen et al. found that a high m<sup>6</sup>A level of the *lncRNA NEAT1* is associated with bone metastasis of prostate cancer (Wen et al. 2020). Investigation of the mechanism indicated that

the *lncRNA NEAT1* may function as a scaffold and bind to CYCLIN1 and CDK19 to phosphorylate Poll at Ser2 in an m<sup>6</sup>A-dependent manner. The *NEAT1/CDK19/CYCLIN1* complex could promote cancer metastasis (Wen et al. 2020). Another *lncRNA PCAT6* was found to be upregulated specifically in prostate cancer specimens with bone metastasis (Lang et al. 2021). METTL3-induced m<sup>6</sup>A modification and IGF2BP3-dependent m<sup>6</sup>A recognition leads to the overexpression of PCAT6. Furthermore, PCAT6 interacts with *IGF2BP3* and *IGF1R* mRNA to enhance IGF1R expression and facilitates bone metastasis (Lang et al. 2021). Li et al. reported that METTL3 modulates ITGB1 expression via the m<sup>6</sup>A-HuR-dependent pathway, which influences the association of ITGB1 with Collagen I to trigger bone metastasis of prostate cancer (Li et al. 2020a).

In bladder cancer, the m<sup>6</sup>A machinery, including METTL3 and YTHDF2, degrades the m<sup>6</sup>A-modified mRNAs of tumor suppressors *SETD* and *KLF4*, thereby promoting tumorigenesis and metastasis (Xie et al. 2020a). Gu et al. found that the m<sup>6</sup>A writer METTL14 is downregulated in bladder cancer tissues and tumor-initiating cells (Gu et al. 2019). Low expression of METTL14 facilitates self-renewal capacity, malignant proliferation, and metastasis through decreasing m<sup>6</sup>A modification levels of *Notch1* mRNA and increasing its translation (Gu et al. 2019). The m<sup>6</sup>A eraser FTO is upregulated and modulates tumor growth and metastasis in bladder cancer (Tao et al. 2021). FTO demethylates m<sup>6</sup>A of the *lncRNA MALAT1* and increases its mRNA stability via the YTHDF2 reader. MALAT1 elevates the expression level of MAL2 by sponging *miR-384* to aggravate bladder cancer progression (Tao et al. 2021).

In renal cell carcinoma, the *lncRNA DMDRMR* binds to m<sup>6</sup>A reader IGF2BP3 to stabilize m<sup>6</sup>A-modified mRNAs including *CDK4*, *COL6A1*, *LAMA5* and *FN1*, and increase their protein production. Therefore, *DMDRMR* accelerates tumor growth by increasing CDK expression and coordinates cell invasion and metastasis partially by elevating *FN1* translation in an m<sup>6</sup>A-dependent manner (Gu et al. 2021). Gu et al. reported that low expression of METTL14 can decrease the m<sup>6</sup>A modification level of *P2RX6* to augment *P2RX6* expression and thus activate the p-ERK1/2/MMP9 signaling axis, thereby promoting renal cancer development (Gu et al. 2021).

### **M<sup>6</sup>A Deposition and the Metastasis of Gynecological Tumors**

Cervical, ovarian and endometrial cancers are three common lethal gynecological malignancies. Metastasis is the foremost cause for the poor prognosis and mortality of patients with these cancers. The regulatory roles of m<sup>6</sup>A modification have been deeply investigated in these gynecological cancers.

Cervical cancer (CC) is the second most prevalent cancer in women worldwide. Yang et al. found that METTL3 mediates the *lncRNA ZAFS1* to sponge *miR-647* in an m<sup>6</sup>A-dependent manner, and this RNA–RNA interaction modulates tumor growth and metastasis in CC (Yang et al. 2020b). Another *lncRNA GAS5-AS1* is markedly downregulated, which is associated with lymphatic and distant metastasis in CC (Wang et al. 2019b). Further studies showed that *lncRNA GAS5-AS1* can form a ternary complex with GAS5 and the m<sup>6</sup>A eraser ALKBH5, which decreases the m<sup>6</sup>A modification levels of the tumor suppressor *GAS5* and blocks *GAS5* RNA decay in a YTHDF2-dependent way (Wang et al. 2019b). These m<sup>6</sup>A-induced effects confer the anti-metastatic function of *lncRNA GAS5-AS1* in CC (Wang et al. 2019b). A recent study demonstrated that the overexpression of the m<sup>6</sup>A writer METTL3 is closely linked to lymphatic metastasis in CC (Wang et al. 2020f). Mechanistically, METTL3 can enrich m<sup>6</sup>A deposition of 3'UTR of *HK2* and recruit the reader YTHDF1 to enhance *HK2* expression. The m<sup>6</sup>A-mediated *HK2* elevation promotes the Warburg effect and aggravates cancer progression (Wang et al. 2020f). The m<sup>6</sup>A eraser FTO is closely involved in the proliferation and migration of CC cells (Zou et al. 2019). FTO interacts with *E2F1* and *Myc* mRNAs to accelerate the translation of these oncogenic transcripts, thereby promoting CC development (Zou et al. 2019).

Ovarian cancer has the highest mortality among gynecological tumors and has become an enormous threat to women's health. The m<sup>6</sup>A writer METTL3 is upregulated in ovarian cancer and associated with lymph node metastasis and an advanced pathological grade (Liang et al. 2020). METTL3 activates the AKT pathway and promotes cyclin D1 expression (Liang et al. 2020). The m<sup>6</sup>A reader YTHDF1 is overexpressed in ovarian cancer, which is closely correlated with poor prognosis (Liu et al. 2020b). YTHDF1 recognizes m<sup>6</sup>A-modified *EIF3C* mRNA and facilitates *EIF3C* protein output, so as to induce tumorigenesis and metastasis (Liu et al. 2020b).

In endometrial cancer, the m<sup>6</sup>A eraser FTO mediates demethylation at the 3'-UTR of *HOXB13* mRNA to enhance *HOXB13* protein translation, thereby activating the Wnt signaling axis to stimulate invasion and metastasis (Zhang et al. 2020b).

### **M<sup>6</sup>A Modification and Metastasis in Other Cancer Types**

In addition to the aforementioned cancers, the regulatory functions of the m<sup>6</sup>A axis have been found in other cancers, such as nasopharyngeal carcinoma, oral squamous cell carcinoma, osteosarcoma and thyroid cancer.

In nasopharyngeal carcinoma, m<sup>6</sup>A modification is enriched on *lncRNA FAM225A* to increase its RNA stability.



The upregulation of *FAM225A* sequesters *miR-590-3p* and *miR-1275* to elevate *ITGB3* expression and activate the FAK/PI3K/AKT signaling pathway, which leads to malignant proliferation and invasion of nasopharyngeal carcinoma (Zheng et al. 2019).

In oral squamous cell carcinoma, high expression of *METTL3* is closely correlated with poor prognosis (Liu et al. 2020a). *METTL3* mediates the m<sup>6</sup>A modification at the 3' UTR of *BM11* mRNA and facilitates *BM11* translation via *IGF2BP1*, thereby promoting proliferation, self-renewal and metastasis (Liu et al. 2020a).

The m<sup>6</sup>A writer WTAP was found to promote cancer progression in osteosarcoma (Lian et al. 2018). WTAP induces m<sup>6</sup>A enrichment at the 3'UTR of *HMBOX1* mRNA and enhances its expression, which activates the PI3K/AKT pathway to stimulate osteosarcoma growth and metastasis (Lian et al. 2018).

M<sup>6</sup>A modification plays a critical role in regulating thyroid cancer development. Ye et al. reported that the *lncRNA MALAT1* increases the expression levels of the m<sup>6</sup>A reader *IGF2BP2* and *Myc* by sponging *miR-204* in an m<sup>6</sup>A-dependent manner, to stimulate migration and invasion of thyroid cancer (Ye et al. 2021). A recent study found that the m<sup>6</sup>A writer *METTL3* is downregulated and markedly associates with poor prognosis in papillary thyroid carcinoma (He et al. 2021). Low expression of *METTL3* activates the NF- $\kappa$ B pathway via abolishing the m<sup>6</sup>A modification of *C-Rel* and *Rel A* and inducing IL-18 secretion to recruit tumor-associated neutrophils, thereby aggravating cancer progression of papillary thyroid cancer (He et al. 2021).

Collectively, the versatile mechanisms of m<sup>6</sup>A modification in regulating cancer metastasis are summarized in Table 1. Additionally, it is well known that cancer stem cell (CSC) maintenance and EMT are two critical events in driving cancer metastasis. The m<sup>6</sup>A modification plays an important role in regulating the CSC-like properties and EMT process in various types of cancer as mentioned above. Herein, we also summarized the shared common functions of m<sup>6</sup>A regulators in sustaining CSC-like features and inducing EMT in Fig. 2.

### M<sup>6</sup>A-Mediated Cancer Metastasis Organotropism

It is noteworthy that most cancers are prone to metastasize to a specific organ, known as “organotropism”. For instance, colorectal cancer has a high propensity to metastasize to the liver. Breast cancer preferably metastasizes to the lungs, bones and brain. Hepatocellular carcinoma is prone to lung metastasis, whereas prostate cancer frequently relapses in the bone. The specificity of the metastatic process is determined by numerous factors, including tumor-intrinsic properties, organ-specific niches, and the complicated interplay between

tumor and the surrounding microenvironment. Increasing evidence demonstrates that m<sup>6</sup>A modification plays an important role in regulating metastasis organotropism in various cancer.

The liver is one of the most frequently distant metastatic organs in multiple cancers such as colorectal cancer, gastric cancer and pancreatic cancer. In colorectal cancer development, m<sup>6</sup>A writers, readers and erasers are extensively involved in liver-specific metastasis through the following mechanisms, including *METTL14/XIST*, *METTL13/RP11*, *METTL3/SOX2* and *HMGAI*, *METTL14/miR-375/ YAP1* and *SP1*, *IGF2BP2/SOX2* and *HMGAI*, *hnRNPA2B1-Rp11-Siah-1-Fbx45/ZEB1*, and the *ALKBH5/RP11* cascade (Chen et al. 2020b; Wu et al. 2019; Xie et al. 2021; Yang et al. 2020a). In advanced gastric cancer, several m<sup>6</sup>A pathways greatly contribute to the liver metastasis of gastric cancer including *METTL3/BATF2/ERK*, *METTL3/ZMYM1/CtBP/LSD1/CoREST*, *YTHDF1/USP14*, *METTL3-IGF2BP3/HDGF*, *ALKBH5/NEAT1/EZH2*, and *FTO/TCF7L2/Wnt/β-catenin* feedback (Chen et al. 2021d; Gao et al. 2021; Wang et al. 2020e; Xie et al. 2020b; Yue et al. 2019; Zhang et al. 2019). In pancreatic cancer, *METTL14*-mediated *PERP* m<sup>6</sup>A and *YTHDC1*-mediated Warburg effect promoted liver metastasis (Hou et al. 2021b; Wang et al. 2020b).

The lung is another favored metastatic site for solid tumors such as HCC, breast cancer, bladder cancer and renal cell carcinoma. There are various m<sup>6</sup>A factors to trigger lung metastasis of HCC, such as m<sup>6</sup>A writers *METTL14-DGCR8/miR-126*, *METTL14/EGFR/PI3K/AKT*, *KIAA1429/GATA3-HuR*, and *KIAA1429/CircDLC1/MMP1-HuR*, m<sup>6</sup>A readers *YTHDF1/ATG2A* and *ATG14*, *YTHDF1/EGFR*, *YTHDF2/OCT4*, and *YTHDF3/Zeb*, and m<sup>6</sup>A erasers *FTO/SOX2*, *KLF4* and *NANOG* (Bian et al. 2021; Lan et al. 2019; Li et al. 2021a; Ma et al. 2017; Shi et al. 2020b; Su et al. 2021; Wang et al. 2020c; Zhang et al. 2020a). In bladder cancer, these m<sup>6</sup>A-related mechanisms might account for increased liver metastasis including *METTL14/Notch1*, *METTL3-YTHDF2/SETD* and *KLF4*, *YTHDF2/AXIN1/Wnt/β-catenin* and *FTO-YTHDF2/MALAT1/MAL2* (Gu et al. 2019; Li et al. 2021b; Tao et al. 2021; Xie et al. 2020a). In breast cancer, *METTL3/KRT7*, *METTL3/COL3A1*, *FTO/BNIP3*, and *FTO-YTHDF1-eEF1/KRT7* axis are responsible for liver metastasis (Chen et al. 2021b; Niu et al. 2019; Shi et al. 2020a). Gu et al. found that *IGF2BP3*-induced *FNI* m<sup>6</sup>A modification contributes to liver metastasis of renal cell carcinoma (Gu et al. 2021).

Lung cancer and breast cancer exhibit great propensity for brain-specific metastasis, and m<sup>6</sup>A modification is extensively involved in the malignant behaviors. As for the brain metastasis of lung cancer, multiple m<sup>6</sup>A pathways regulate the process, such as

**Table 1** The mechanisms of m<sup>6</sup>A regulators in cancer metastasis

m <sup>6</sup> A regulators	Functions in Cancer	Target genes	Cancer type	References
Writer METTL14↓	Invasion and migration	<i>miR-126</i>	HCC	Ma et al. (2017)
METTL14↓	Migration, invasion and EMT	<i>EGFR</i>	HCC	Shi et al. (2020b)
METTL14↓	EMT, invasion and migration	<i>SOX4</i>	CRC	Chen et al. (2020a)
METTL14↓	Tumorigenicity and metastasis	<i>lncRNA XIST</i>	CRC	Yang et al. (2020a)
METTL14↓	Metastasis	<i>miR-375</i>	CRC	Chen et al. (2020b)
METTL14↑	Metastasis and progression	<i>PERP</i>	Pancreatic cancer	Wang et al. (2020b)
METTL14↓	Self-renewal capacity, malignant proliferation and metastasis	<i>Notch1</i>	Bladder cancer	Gu et al. (2019)
METTL14	Proliferation	<i>P2RX6</i>	Renal cancer	Gu et al. (2021)
Writer METTL3↑	Metastasis	<i>SOX2</i> and <i>HMGAI</i>	CRC	Hou et al. (2021a); Li et al. (2019)
METTL3↑	Metastasis	<i>KRT7</i>	Breast cancer	Chen et al. (2021b)
METTL3↑	Stemness, invasion and migratory	<i>SOX2</i>	Breast cancer	Xie et al. (2021)
METTL3↑	Metastasis	<i>COL3A1</i>	Breast cancer	Shi et al. (2020a)
METTL3	Invasion and migration	<i>circ1662</i>	CRC	Chen et al. (2021a)
METTL3↑	EMT, progression and metastasis	<i>ZMYM1</i>	GC	Yue et al. (2019)
METTL3↑	Metastasis	<i>BATF2</i>	GC	Xie et al. (2020b)
METTL3	Metastasis	A877 residue of <i>KRT7-AS</i>	Breast cancer	Chen et al. (2021b)
METTL3↑	Invasion, angiogenesis and metastasis	<i>miR143-3p</i>	Lung cancer	Wang et al. (2019a)
METTL3↑	Metastasis and chemotherapeutic resistance	<i>YAP</i>	Lung cancer	Jin et al. (2019a, b)
METTL3↑	Progression and metastasis	<i>MALAT1</i>	Lung cancer	Jin et al. (2019a, b)
METTL3	Metastasis	<i>PCAT6</i>	Prostate cancer	Lang et al. (2021)
METTL3	Metastasis	<i>ITGB1</i>	Prostate cancer	Li et al. (2020a)
METTL3	Tumorigenesis and metastasis	<i>SETD</i> and <i>KLF4</i>	Bladder cancer	Xie et al. (2020a)
METTL3↑	Proliferation and metastasis	<i>lncRNA ZAFS1</i>	Cervical cancer	Yang et al. (2020b)
METTL3↑	Progression and metastasis	<i>HK2</i>	Cervical cancer	Wang et al. (2020e)
METTL3	Metastasis	<i>Cyclin D1</i>	Ovarian cancer	Liang et al. (2020)
METTL3	Proliferation, self-renewal and metastasis	<i>BMI1</i>	Oral squamous cell carcinoma	Liu et al. (2020a)
METTL3↓	Progression	<i>C-Rel</i> and <i>Rel A</i>	Papillary thyroid cancer	He et al. (2021)
Writer METTL13	Glycolysis and angiogenesis	<i>HDGF</i>	GC	Wang et al. (2020d)
METTL13	Metastasis	<i>Rp11</i>	CRC	Wu et al. (2019)
Writer KIAA1429	Proliferation, apoptosis, invasion and migration	<i>GATA3</i>	HCC	Lan et al. (2019)
KIAA1429	Proliferation, invasion and metastasis	<i>circDLC1-HuR-MMP1</i> axis	HCC	Liu et al. (2021)
Writer WTAP	Proliferation and metastasis	<i>HMBOX1</i>	Osteosarcoma	Lian et al. (2018)
Reader hnRNPA2B1	Metastasis	<i>lncRNA Rp11, Siah-1</i> and <i>Fbx45</i>	CRC	Wu et al. (2019)
Reader HuR	EMT and metastasis	<i>ZMYM1</i>	GC	Yue et al. (2019)
Reader YTHDF1↑	Autophagy and metastasis	<i>ATG2A</i> and <i>ATG14</i>	HCC	Li et al. (2021a)
YTHDF1↑	Metastasis	<i>EGFR</i>	HCC	Su et al. (2021)
YTHDF1↑	Proliferation and metastasis	<i>USP14</i>	GC	Chen et al. (2021d)
YTHDF1↑	Tumorigenesis and metastasis	<i>EIF3C</i>	Ovarian cancer	Liu et al. (2020b)

**Table 1** (continued)

m <sup>6</sup> A regulators	Functions in Cancer	Target genes	Cancer type	References
YTHDF1	Progression and metastasis	<i>HK2</i>	Cervical cancer	Wang et al. (2020e)
Reader YTHDC1↑	Inhibition of tumorigenesis and metastasis	<i>miR-30d/RUNX1/SLC2A1/HK pathway</i>	Pancreatic cancer	Hou et al. (2021b)
Reader YTHDF2↑	Stemness and metastasis	<i>OCT4</i>	HCC	Zhang et al. (2020a)
YTHDF2	Tumorigenesis and metastasis	<i>AXIN1</i>	Lung cancer	Li et al. (2021b)
YTHDF2	Tumorigenesis and metastasis	<i>SETD</i> and <i>KLF4</i>	Bladder cancer	Xie et al. (2020a)
YTHDF2↑	Proliferation and metastasis	<i>MALAT1</i>	Bladder cancer	Tao et al. (2021)
Reader YTHDF3↑	Migration	<i>Zeb</i>	HCC	Wang et al. (2020c)
YTHDF3↑	Angiogenesis and outgrowth	<i>ST6GALNAC5</i> , <i>GJA1</i> , and <i>EGFR</i>	Breast cancer	Chang et al. (2020)
Reader IGF2BP2	Metastasis	<i>SOX2</i> , <i>HMGAI</i>	CRC	Hou et al. (2021a); Li et al. (2019)
IGF2BP3	Glycolysis and angiogenesis	<i>HDGF</i>	GC	Wang et al. (2020d)
IGF2BP3	Metastasis	<i>PCAT6</i>	Prostate cancer	Lang et al. (2021)
IGF2BP3↑	Proliferation and metastasis	<i>lncRNA DMDRMR</i>	Renal cell carcinoma	Gu et al. (2021)
Eraser FTO↑	stemness and metastasis	<i>SOX2</i> , <i>KLF4</i> and <i>NANOG</i>	HCC	Bian et al. (2021)
FTO	Progression and metastasis	<i>TCF7L2</i>	GC	Gao et al. (2021)
FTO↓	Metastasis	A950 in <i>KRT7</i> exon 6	Breast cancer	Chen et al. (2021b)
FTO↑	Proliferation and metastasis	<i>BNIP3</i>	Breast cancer	Niu et al. (2019)
FTO	Invasion and migration	<i>miR-181b</i>	Breast cancer	Xu et al. (2020)
FTO	Metastasis	<i>E2F1</i>	Lung cancer	Wang et al. (2021b)
FTO↑	Proliferation and metastasis	<i>MALAT1</i>	Bladder cancer	Tao et al. (2021)
FTO	Proliferation and migration	<i>E2F1</i> and <i>Myc</i> mRNAs	Cervical cancer	Zou et al. (2019)
FTO	Invasion and metastasis	<i>HOXB13</i>	Endometrial cancer	Zhang et al. (2020b)
Eraser ALKBH5	Metastasis	<i>lncRNA NEAT1</i>	GC	Zhang et al. (2019)
ALKBH5↓	Motility	<i>lncRNA KCNK15-AS1</i>	Pancreatic cancer	He et al. (2018)
ALKBH5↑	Stemness and progression	<i>NANOG</i>	Breast cancer	Zhang et al. (2016)
ALKBH5↓	Metastasis	<i>YAP</i> axis	Lung cancer	Jin et al. (2020a)
ALKBH5↓	Metastasis	<i>miR-1914-3p</i>	Lung cancer	Jin et al. (2020a)
ALKBH5↓	Repression of autophagy, metastasis	<i>UBE2C</i>	Lung cancer	Guo et al. (2018)
ALKBH5	Anti-metastatic	<i>lncRNA GAS5-AS1</i>	Cervical cancer	Wang et al. (2019b)
ALKBH5	Metastasis	<i>Rp11</i>	CRC	Wu et al. (2019)

↓Indicates downregulated expression and ↑indicates upregulated expression in cancer. HCC is hepatocellular carcinoma, CRC is colorectal carcinoma and GC is gastric cancer

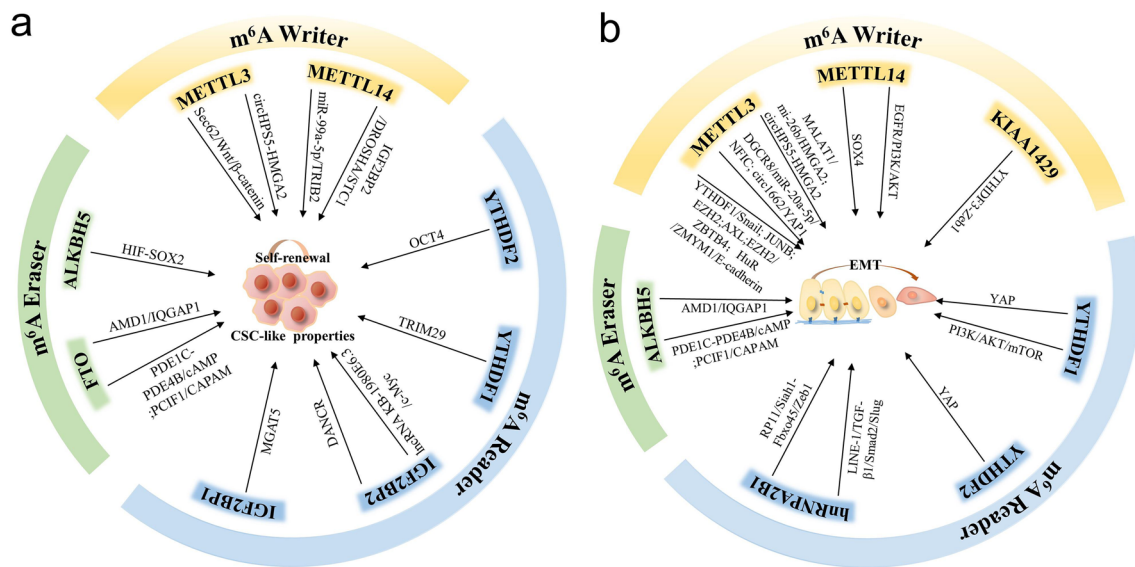
METTL3/*miR-143-3p/VASH1/VEGFA*, METTL3-YTHDF1/3-eIF3b/*YAP*, METTL3/*MALAT1/miR-1914-3p/YAP*, YTHDF2/*AXIN1/Wnt/β-catenin*, ALKBH5-YTHDF2/*YAP*, ALKBH5-HuR/*miR-1914-3p/LAST2/YAP*, FTO-E2F1/*NELL2*, and ALKBH5/*UBE2C* (Guo et al. 2018; Jin et al. 2019a; Jin et al. 2020a; Li et al. 2021b; Wang et al. 2019a; Wang et al. 2021c). In breast cancer, YTHDF3-induced translation of m<sup>6</sup>A-enriched transcripts for *ST6GALNAC5*, *GJA1* and *EGFR* promotes brain metastasis (Chang et al. 2020).

The above-mentioned m<sup>6</sup>A-mediated liver-, lung- and brain-specific metastasis in various cancers are summarized in Fig. 3. Whether m<sup>6</sup>A modification and related

regulators could serve as biomarkers and targets for cancer metastasis organotropism still warrants extensive studies.

## The Clinical Translational Potential of the m<sup>6</sup>A Axis in Cancer Diagnosis and Treatment

Given the essential functions of the m<sup>6</sup>A axis in cancer development, the clinical translation potential of m<sup>6</sup>A has attracted increasing attention. The m<sup>6</sup>A axis has become a promising target for diagnosis, prognostic prediction and therapy.



**Fig. 2** The functions of m<sup>6</sup>A regulators in maintaining CSC-like properties and inducing EMT. The schematic diagrams showed the shared common functions of m<sup>6</sup>A-mediated CSC-like features and EMT in **a** and **b**, respectively. The m<sup>6</sup>A regulators, including writers, read-

ers and erasers, have been extensively studied how to affect CSC and EMT, which are two critical biological events to drive cancer metastasis

Ge et al. claimed that m<sup>6</sup>A modification has the potential to serve as an ideal biomarker in GC progression (Ge et al. 2020). It was reported that the m<sup>6</sup>A levels are significantly increased in the peripheral blood RNA in GC, compared to benign gastric disease (BGD) and healthy controls (HCs). Furthermore, m<sup>6</sup>A levels exhibit an elevated trend with the progression and metastasis of GC (Ge et al. 2020).

Our group comprehensively assessed the clinically predictive potential of m<sup>6</sup>A modification, HCC progression and therapeutic responses. According to the established m<sup>6</sup>A score system, three distinct m<sup>6</sup>A patterns were identified in HCC. The HCC cluster with a lower m<sup>6</sup>A score frequently showed metabolic hyperactivity, better prognosis and lower response rate to sorafenib treatment. In contrast, the HCC cluster with a higher m<sup>6</sup>A score usually exhibited hypoactive metabolism, poorer prognosis, and favorable response to sorafenib therapy (Shen et al. 2020). In future, these findings still need to be confirmed in a large cohort, and further efforts are required to explore the clinical translation potential of the m<sup>6</sup>A machinery in HCC metastasis.

Recently, targeting the critical enzymes of m<sup>6</sup>A modification for cancer therapy has made some inspiring progressions. For example, METTL3 contains a Rossmann fold, which binds the S-adenosyl methionine (SAM) methyl donor. STM2457 is a bioavailable inhibitor of METTL3 catalytic activity through a SAM-competitive mode, which has

been developed to apply for the treatment of acute myeloid leukemia (AML) (Yankova et al. 2021). For m<sup>6</sup>A demethylases, two targeted FTO inhibitors, FB23 and FB23-2, have also been designed. FB23-2 significantly inhibits the progression of AML in cell lines and xeno-transplanted mice (Huang et al. 2019). Moreover, the two potent inhibitors of FTO showed promising anti-tumor effects in multiple types of cancers (Su et al. 2020). Another FTO-targeted molecule, Saikosaponin D (SsD), increases the global m<sup>6</sup>A RNA methylation, effectively overcoming FTO/m<sup>6</sup>A-mediated resistance to tyrosine kinase inhibitors in leukemia (Sun et al. 2021).

In view of the widespread clinical application of chemotherapy and targeted therapies in metastasis treatment, researchers have also explored the possibility of co-administration of m<sup>6</sup>A-related molecules. Depletion of methyltransferases, METTL3 and METTL14, was found to enhance the response to anti-PD-1 treatment in CRC and melanoma, by increasing cytotoxic tumor-infiltrating CD8+ T cells and elevating the secretion of IFN- $\gamma$ , CXCL9, and CXCL10 in the tumor microenvironment (Wang et al. 2020a; Yang et al. 2019). Similarly, the deletion of the m<sup>6</sup>A demethylase ALKBH5 modulates the composition of tumor-infiltrating Treg and myeloid-derived suppressor cells, thereby sensitizing tumors to cancer immunotherapy (Li et al. 2020b). Despite the promising perspective of m<sup>6</sup>A in clinical translation, we should be aware that most of the results were achieved in vitro or



in mice models. Large-scale clinical trials are urgently required to evaluate the effectiveness of targeting m<sup>6</sup>A for cancer therapy.

Nowadays, with the development of m<sup>6</sup>A mapping technologies, such as Mazter-Seq, it is possible to determine the precise quantity of m<sup>6</sup>A modification at specific sites of specific RNAs (Garcia-Campos et al. 2019). These new methods will greatly promote the clinical application of m<sup>6</sup>A as a novel biomarker. Additionally, the combination of m<sup>6</sup>A and target therapy or immune therapy may be a promising strategy for anti-metastasis treatment.

### Limitations and Future Perspectives of m<sup>6</sup>A Modifications

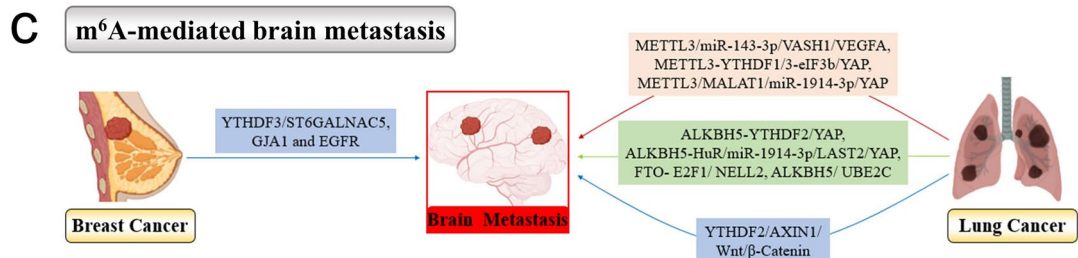
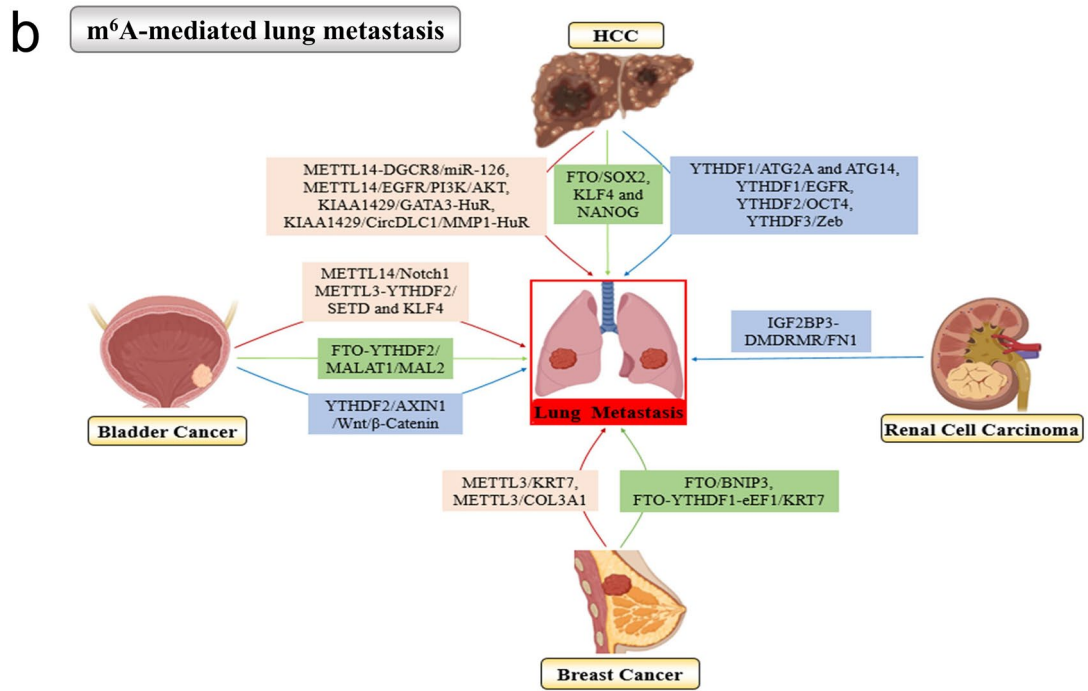
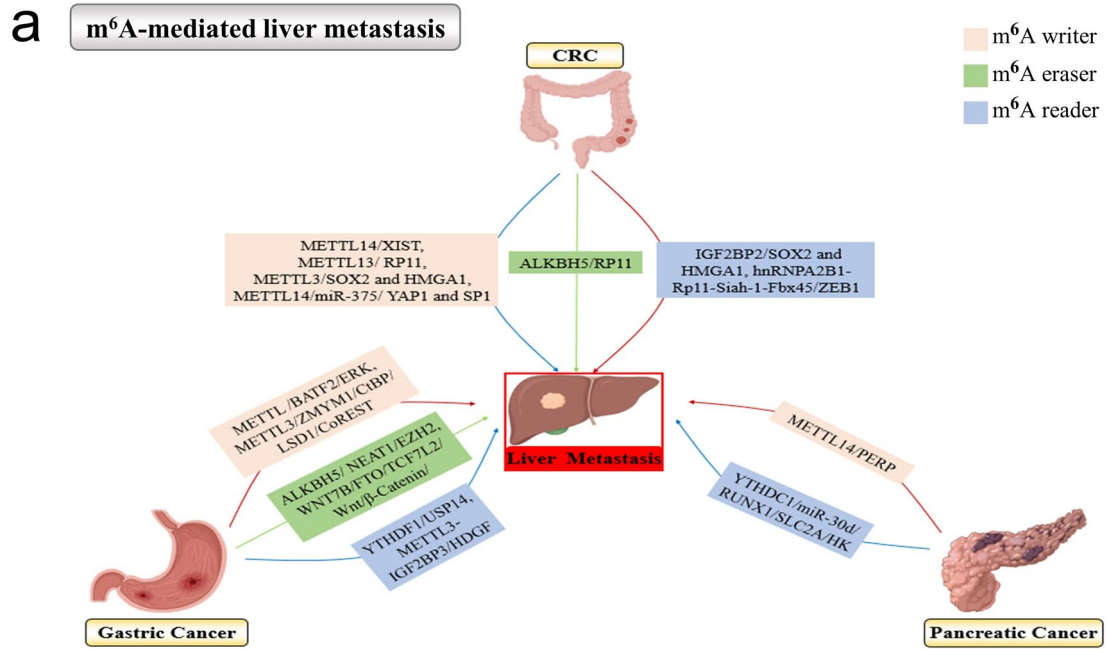
Despite the great progression of m<sup>6</sup>A-related research in cancer development, there are still some limitations to be noted. First, the writers, readers and erasers of m<sup>6</sup>A are not specific to a single target, and m<sup>6</sup>A regulators have multiple substrates, including oncogenes and tumor suppressor genes. These may explain the controversial roles of a single m<sup>6</sup>A regulator that may exert opposite functions. Meanwhile, the uncertainty of m<sup>6</sup>A-targeting also increases the difficulty to develop a therapeutic strategy. Second, m<sup>6</sup>A modification may induce completely different effects on RNA metabolism. For some RNAs, m<sup>6</sup>A enrichment may facilitate RNA decay, such as *EGFR*, *Oct4* and *Sox2*, but in some cases, m<sup>6</sup>A deposition may increase the stability of the RNA, such as in the case of tumor suppressor *KLF14*. The different fates of m<sup>6</sup>A-RNAs may develop in a contextspecific manner for different targets and different types of cancers, and the underlying mechanisms need to be clarified. Additionally, a single m<sup>6</sup>A regulator may display different expression patterns in different cancers, as well as in the same type of cancer. Taking METTL14 as an

example, it is downregulated in multiple types of cancers and can inhibit cancer metastasis, but in pancreatic cancer, METTL14 is upregulated, promoting metastasis (Wang et al. 2020b). Another example is METTL3, several studies have demonstrated that METTL3 is upregulated and acts as an oncogene in breast cancer (Chen et al. 2021b; Xie et al. 2021). However, Shi et al. reported that METTL3 is downregulated and can inhibit the development of breast cancer (Shi et al. 2020a). These inconsistencies may result from different pathogenesis pathways and backgrounds of different cancer types. Moreover, the heterogeneity of cancer may lead to a discrepancy.

The m<sup>6</sup>A modification is a new frontier in the cancer field, and most of the present studies focus on it in cancer cells. As the microenvironment is essential for cancer metastasis, further study of m<sup>6</sup>A alterations and functions in the tumor microenvironment are warranted. Additionally, it is important to identify the determinative m<sup>6</sup>A site with a regulatory function, as some m<sup>6</sup>A sites may present just a constitutive event lacking of function. A particular m<sup>6</sup>A signature in some specific transcript loci may be more informative than the global m<sup>6</sup>A profiles, which would serve as better biomarkers for clinical translation in cancer. These promising perspectives of m<sup>6</sup>A deserve to be further probed in future investigations.

### Conclusion

Cancer metastasis is a complicated biological event and accounts for a high mortality rate. The m<sup>6</sup>A modifications may exert versatile roles to determine the fate of RNAs, which play essential roles in regulating cancer development. Through systematically reviewing the latest progression of m<sup>6</sup>A modification in cancer metastasis, we hope to comprehensively enrich our knowledge on m<sup>6</sup>A regulation



**Fig. 3** The m<sup>6</sup>A-mediated organ-specific metastasis in various cancers. The schematic diagram indicated the m<sup>6</sup>A-mediated liver metastasis (a), lung metastasis (b), and brain metastasis (c) in different cancers, and the m<sup>6</sup>A writers, readers and erasers all play essential roles in the organ-specific metastatic process. The diagram of various cancer types were created with the help of BioRender.com

and provide some new clues to develop effective strategies to monitor and treat cancer metastasis based on m<sup>6</sup>A dysregulation.

**Acknowledgements** This study was supported by the Key Program of the National Natural Science Foundation of China (81930074, 2020–2024), the Major Program of National Natural Science Foundation of China (91959203, 2020–2023) and the Natural Science Foundation of China (81672820, 2017–2020; 81672378, 2017–2020; 82173093)

**Authors' Contributions** LXQ designed the outline and directed the writing of the paper. JZP, SZX and JFX collected the related papers. JZ and HX retrieved the related studies, drafted manuscript and prepared the figures and tables. YHS revised the language and provided comments. All authors critically revised the manuscript.

**Data and Material Availability** Not applicable.

**Code Availability** Not applicable.

## Declarations

**Conflicts of Interest** No conflicts of interest to disclose.

**Ethical Approval** Not applicable.

**Consent to Participate** Not applicable.

**Consent to Publish** Not applicable.

## References

- Alexandrov A, Martzen MR, Phizicky EM (2002) Two proteins that form a complex are required for 7-methylguanosine modification of yeast tRNA. *RNA* 8:1253–1266. <https://doi.org/10.1017/s1355838202024019>
- Arango D et al (2018) Acetylation of cytidine in mRNA promotes translation efficiency. *Cell* 175:1872–1886.e1824. <https://doi.org/10.1016/j.cell.2018.10.030>
- Audia JE, Campbell RM (2016) Histone Modifications and cancer cold spring. *Harb Perspect Biol* 8:a019521. <https://doi.org/10.1101/cshperspect.a019521>
- Barbieri I, Kouzarides T (2020) Role of RNA modifications in cancer. *Nat Rev Cancer* 20:303–322. <https://doi.org/10.1038/s41568-020-0253-2>
- Bian X, Shi D, Xing K, Zhou H, Lu L, Yu D, Wu W (2021) AMD1 upregulates hepatocellular carcinoma cells stemness by FTO mediated mRNA demethylation. *Clin Transl Med* 11:e352. <https://doi.org/10.1002/ctm2.352>
- Bos PD et al (2009) Genes that mediate breast cancer metastasis to the brain. *Nature* 459:1005–1009. <https://doi.org/10.1038/nature08021>
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer. J Clin* 68:394–424. <https://doi.org/10.3322/caac.21492>
- Calabrese C et al (2020) Genomic basis for RNA alterations in cancer. *Nature* 578:129–136. <https://doi.org/10.1038/s41586-020-1970-0>
- Chang G et al (2020) YTHDF3 induces the translation of m(6) A-enriched gene transcripts to promote breast cancer brain metastasis. *Cancer Cell* 38:857–871.e857. <https://doi.org/10.1016/j.ccell.2020.10.004>
- Chen Z et al (2019) Transfer RNA demethylase ALKBH3 promotes cancer progression via induction of tRNA-derived small RNAs. *Nucleic Acids Res* 47:2533–2545. <https://doi.org/10.1093/nar/gky1250>
- Chen X et al (2020a) METTL14-mediated N6-methyladenosine modification of SOX4 mRNA inhibits tumor metastasis in colorectal cancer. *Mol Cancer* 19:106. <https://doi.org/10.1186/s12943-020-01220-7>
- Chen X et al (2020b) METTL14 suppresses CRC progression via regulating N6-methyladenosine-dependent primary miR-375 processing. *Mol Ther* 28:599–612. <https://doi.org/10.1016/j.ymthe.2019.11.016>
- Chen XY, Liang R, Yi YC, Fan HN, Chen M, Zhang J, Zhu JS (2021) The m(6)A reader YTHDF1 facilitates the tumorigenesis and metastasis of gastric cancer via USP14 translation in an m(6) A-dependent manner. *Front Cell Dev Biol* 9:647702. <https://doi.org/10.3389/fcell.2021.647702>
- Chen C et al (2021a) N6-methyladenosine-induced circ1662 promotes metastasis of colorectal cancer by accelerating YAP1 nuclear localization. *Theranostics* 11:4298–4315. <https://doi.org/10.7150/thno.51342>
- Chen F et al (2021b) N(6)-methyladenosine regulates mRNA stability and translation efficiency of KRT7 to promote breast cancer lung metastasis. *Cancer Res* 81:2847–2860. <https://doi.org/10.1158/0008-5472.Can-20-3779>
- Chen S et al (2021c) CLK1/SRSF5 pathway induces aberrant exon skipping of METTL14 and Cyclin L2 and promotes growth and metastasis of pancreatic cancer. *J Hematol Oncol* 14:60. <https://doi.org/10.1186/s13045-021-01072-8>
- Correia AL et al (2021) Hepatic stellate cells suppress NK cell-sustained breast cancer dormancy. *Nature*. <https://doi.org/10.1038/s41586-021-03614-z>
- Cui J, Liu Q, Sendinc E, Shi Y, Gregory RI (2021) Nucleotide resolution profiling of m3C RNA modification by HAC-seq. *Nucleic Acids Res* 49:e27. <https://doi.org/10.1093/nar/gkaa1186>
- Dai X, Wang T, Gonzalez G, Wang Y (2018) Identification of YTH domain-containing proteins as the readers for N1-methyladenosine in RNA. *Anal Chem* 90:6380–6384. <https://doi.org/10.1021/acs.analchem.8b01703>
- Fumagalli A et al (2020) Plasticity of Lgr5-negative cancer cells drives metastasis in colorectal cancer. *Cell Stem Cell* 26:569–578.e567. <https://doi.org/10.1016/j.stem.2020.02.008>
- Gao Y, Bado I, Wang H, Zhang W, Rosen JM, Zhang XH (2019) Metastasis organotropism: redefining the congenial soil. *Dev Cell* 49:375–391. <https://doi.org/10.1016/j.devcel.2019.04.012>
- Gao Q et al (2021) A WNT7B-m(6)A-TCF7L2 positive feedback loop promotes gastric cancer progression and metastasis. *Signal Transduct Target Ther* 6:43. <https://doi.org/10.1038/s41392-020-00397-z>
- Garcia-Campos MA et al (2019) Deciphering the “m(6)A Code” via antibody-independent quantitative profiling. *Cell* 178:731–747. e716. <https://doi.org/10.1016/j.cell.2019.06.013>
- Ge L et al (2020) Level of N6-methyladenosine in peripheral blood RNA: a novel predictive biomarker for gastric cancer. *Clin Chem* 66:342–351. <https://doi.org/10.1093/clinchem/hvz004>

- Gu C et al (2019) Mettl14 inhibits bladder TIC self-renewal and bladder tumorigenesis through N(6)-methyladenosine of Notch1. *Mol Cancer* 18:168. <https://doi.org/10.1186/s12943-019-1084-1>
- Gu Y et al (2021) DMDRMR-mediated regulation of m(6)A-modified CDK4 by m(6)A reader IGF2BP3 drives ccRCC progression. *Cancer Res* 81:923–934. <https://doi.org/10.1158/0008-5472.Can-20-1619>
- Guo J et al (2018) Deregulation of UBE2C-mediated autophagy repression aggravates NSCLC progression. *Oncogenesis* 7:49. <https://doi.org/10.1038/s41389-018-0054-6>
- Haag S, Kretschmer J, Bohnsack MT (2015) WBSR22/Merm1 is required for late nuclear pre-ribosomal RNA processing and mediates N7-methylation of G1639 in human 18S rRNA. *RNA* 21:180–187. <https://doi.org/10.1261/rna.047910.114>
- Hadfield G (1954) The dormant cancer cell. *Br Med J* 2:607–610. <https://doi.org/10.1136/bmj.2.4888.607>
- He Y et al (2018) ALKBH5 inhibits pancreatic cancer motility by decreasing long non-coding RNA KCNK15-AS1 methylation. *Cell Physiol Biochem* 48:838–846. <https://doi.org/10.1159/000491915>
- He L, Li H, Wu A, Peng Y, Shu G, Yin G (2019) Functions of N6-methyladenosine and its role in cancer. *Mol Cancer* 18:176. <https://doi.org/10.1186/s12943-019-1109-9>
- He J et al (2021) METTL3 restrains papillary thyroid cancer progression via m(6)A/c-Rel/IL-8-mediated neutrophil infiltration. *Mol Ther* 29:1821–1837. <https://doi.org/10.1016/j.ymthe.2021.01.019>
- Hoshino A et al (2015) Tumour exosome integrins determine organotropic metastasis. *Nature* 527:329–335. <https://doi.org/10.1038/nature15756>
- Hou P et al (2021a) LINC00460/DHX9/IGF2BP2 complex promotes colorectal cancer proliferation and metastasis by mediating HMGA1 mRNA stability depending on m6A modification. *J Exp Clin Cancer Res* 40:52. <https://doi.org/10.1186/s13046-021-01857-2>
- Hou Y et al (2021b) YTHDC1-mediated augmentation of miR-30d in repressing pancreatic tumorigenesis via attenuation of RUNX1-induced transcriptional activation of Warburg effect. *Cell Death Differ*. <https://doi.org/10.1038/s41418-021-00804-0>
- Huang Y et al (2019) Small-molecule targeting of oncogenic FTO demethylase in acute myeloid leukemia. *Cancer Cell* 35:677–691. <https://doi.org/10.1016/j.ccell.2019.03.006>
- Jin D et al (2019a) m(6)A mRNA methylation initiated by METTL3 directly promotes YAP translation and increases YAP activity by regulating the MALAT1-miR-1914-3p-YAP axis to induce NSCLC drug resistance and metastasis. *J Hematol Oncol* 12:135. <https://doi.org/10.1186/s13045-019-0830-6>
- Jin X et al (2019b) AtTrm5a catalyses 1-methylguanosine and 1-methylinosine formation on tRNAs and is important for vegetative and reproductive growth in *Arabidopsis thaliana*. *Nucleic Acids Res* 47:883–898. <https://doi.org/10.1093/nar/gky1205>
- Jin D et al (2020a) m(6)A demethylase ALKBH5 inhibits tumor growth and metastasis by reducing YTHDFs-mediated YAP expression and inhibiting miR-107/LATS2-mediated YAP activity in NSCLC. *Mol Cancer* 19:40. <https://doi.org/10.1186/s12943-020-01161-1>
- Jin X et al (2020b) A metastasis map of human cancer cell lines. *Nature* 588:331–336. <https://doi.org/10.1038/s41586-020-2969-2>
- Jones PA, Issa JP, Baylin S (2016) Targeting the cancer epigenome for therapy. *Nat Rev Genet* 17:630–641. <https://doi.org/10.1038/nrg.2016.93>
- Kaplan RN et al (2005) VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature* 438:820–827. <https://doi.org/10.1038/nature04186>
- Klutstein M, Nejman D, Greenfield R, Cedar H (2016) DNA methylation in cancer and aging. *Cancer Res* 76:3446–3450. <https://doi.org/10.1158/0008-5472.Can-15-3278>
- Koch A et al (2018) Analysis of DNA methylation in cancer: location revisited. *Nat Rev Clin Oncol* 15:459–466. <https://doi.org/10.1038/s41571-018-0004-4>
- Lan T et al (2019) KIAA1429 contributes to liver cancer progression through N6-methyladenosine-dependent post-transcriptional modification of GATA3. *Mol Cancer* 18:186. <https://doi.org/10.1186/s12943-019-1106-z>
- Lang C et al (2021) m(6) A modification of lncRNA PCAT6 promotes bone metastasis in prostate cancer through IGF2BP2-mediated IGF1R mRNA stabilization. *Clin Transl Med* 11:e426. <https://doi.org/10.1002/ctm2.426>
- Laughney AM et al (2020) Regenerative lineages and immune-mediated pruning in lung cancer metastasis. *Nat Med* 26:259–269. <https://doi.org/10.1038/s41591-019-0750-6>
- Li T et al (2019) METTL3 facilitates tumor progression via an m(6) A-IGF2BP2-dependent mechanism in colorectal carcinoma. *Mol Cancer* 18:112. <https://doi.org/10.1186/s12943-019-1038-7>
- Li E, Wei B, Wang X, Kang R (2020a) METTL3 enhances cell adhesion through stabilizing integrin  $\beta$ 1 mRNA via an m6A-HuR-dependent mechanism in prostatic carcinoma. *Am J Cancer Res* 10:1012–1025
- Li N et al (2020b) ALKBH5 regulates anti-PD-1 therapy response by modulating lactate and suppressive immune cell accumulation in tumor microenvironment. *Proc Natl Acad Sci USA* 117:20159–20170. <https://doi.org/10.1073/pnas.1918986117>
- Li Q et al (2021a) HIF-1 $\alpha$ -induced expression of m6A reader YTHDF1 drives hypoxia-induced autophagy and malignancy of hepatocellular carcinoma by promoting ATG2A and ATG14 translation. *Signal Transduct Target Ther* 6:76. <https://doi.org/10.1038/s41392-020-00453-8>
- Li Y et al (2021b) RNA m(6)A reader YTHDF2 facilitates lung adenocarcinoma cell proliferation and metastasis by targeting the AXIN1/Wnt/ $\beta$ -catenin signaling. *Cell Death Dis* 12:479. <https://doi.org/10.1038/s41419-021-03763-z>
- Lian H, Wang QH, Zhu CB, Ma J, Jin WL (2018) Deciphering the epitranscriptome in cancer. *Trends Cancer* 4:207–221. <https://doi.org/10.1016/j.trecan.2018.01.006>
- Liang S, Guan H, Lin X, Li N, Geng F, Li J (2020) METTL3 serves an oncogenic role in human ovarian cancer cells partially via the AKT signaling pathway. *Oncol Lett* 19:3197–3204. <https://doi.org/10.3892/ol.2020.11425>
- Liu F et al (2016a) ALKBH1-mediated tRNA demethylation regulates translation. *Cell* 167:816–828.e816. <https://doi.org/10.1016/j.cell.2016.09.038>
- Liu Y et al (2016b) Tumor exosomal RNAs promote lung pre-metastatic niche formation by activating alveolar epithelial TLR3 to recruit neutrophils. *Cancer Cell* 30:243–256. <https://doi.org/10.1016/j.ccell.2016.06.021>
- Liu L et al (2020a) METTL3 promotes tumorigenesis and metastasis through BMI1 m(6)A methylation in oral squamous cell carcinoma. *Mol Ther* 28:2177–2190. <https://doi.org/10.1016/j.ymthe.2020.06.024>
- Liu T et al (2020b) The m6A reader YTHDF1 promotes ovarian cancer progression via augmenting EIF3C translation. *Nucleic Acids Res* 48:3816–3831. <https://doi.org/10.1093/nar/gkaa048>
- Liu H et al (2021) Circular RNA circDLC1 inhibits MMP1-mediated liver cancer progression via interaction with HuR. *Theranostics* 11:1396–1411. <https://doi.org/10.7150/thno.53227>
- Ma JZ et al (2017) METTL14 suppresses the metastatic potential of hepatocellular carcinoma by modulating N(6)-methyladenosine-dependent primary MicroRNA processing. *Hepatology* 65:529–543. <https://doi.org/10.1002/hep.28885>



- Murgai M et al (2017) KLF4-dependent perivascular cell plasticity mediates pre-metastatic niche formation and metastasis. *Nat Med* 23:1176–1190. <https://doi.org/10.1038/nm.4400>
- Niu Y et al (2019) RNA N<sup>6</sup>-methyladenosine demethylase FTO promotes breast tumor progression through inhibiting BNIP3. *Mol Cancer* 18:46. <https://doi.org/10.1186/s12943-019-1004-4>
- Obenauf AC, Massagué J (2015) Surviving at a distance: organ-specific metastasis trends. *Cancer* 1:76–91. <https://doi.org/10.1016/j.trecan.2015.07.009>
- Olmeda D et al (2017) Whole-body imaging of lymphovascular niches identifies pre-metastatic roles of midkine. *Nature* 546:676–680. <https://doi.org/10.1038/nature22977>
- Paget S (1889) The distribution of secondary growths in cancer of the breast. *Lancet* 133:571–573. [https://doi.org/10.1016/S0140-6736\(00\)49915-0](https://doi.org/10.1016/S0140-6736(00)49915-0)
- Peinado H et al (2017) Pre-metastatic niches: organ-specific homes for metastases. *Nat Rev Cancer* 17:302–317. <https://doi.org/10.1038/nrc.2017.6>
- Phan TG, Croucher PI (2020) The dormant cancer cell life cycle. *Nat Rev Cancer* 20:398–411. <https://doi.org/10.1038/s41568-020-0263-0>
- Ramanathan A, Robb GB, Chan SH (2016) mRNA capping: biological functions and applications. *Nucleic Acids Res* 44:7511–7526. <https://doi.org/10.1093/nar/gkw551>
- Roundtree IA, Evans ME, Pan T, He C (2017) Dynamic RNA modifications in gene expression regulation. *Cell* 169:1187–1200. <https://doi.org/10.1016/j.cell.2017.05.045>
- Saikia M, Fu Y, Pavon-Eternod M, He C, Pan T (2010) Genome-wide analysis of N<sup>1</sup>-methyl-adenosine modification in human tRNAs. *RNA* 16:1317–1327. <https://doi.org/10.1261/rna.2057810>
- Shen X et al (2020) The m<sup>6</sup>A methylation landscape stratifies hepatocellular carcinoma into 3 subtypes with distinct metabolic characteristics. *Cancer Biol Med* 17:937–952
- Shi Y et al (2020a) Reduced expression of METTL3 promotes metastasis of triple-negative breast cancer by m<sup>6</sup>A methylation-mediated COL3A1 up-regulation. *Front Oncol* 10:1126. <https://doi.org/10.3389/fonc.2020.01126>
- Shi Y, Zhuang Y, Zhang J, Chen M, Wu S (2020b) METTL14 inhibits hepatocellular carcinoma metastasis through regulating EGFR/PI3K/AKT signaling pathway in an m<sup>6</sup>A-dependent manner. *Cancer Manag Res* 12:13173–13184. <https://doi.org/10.2147/cmar.S286275>
- Su R et al (2020) Targeting FTO suppresses cancer stem cell maintenance and immune evasion. *Cancer Cell* 38:79–96.e11. <https://doi.org/10.1016/j.ccell.2020.04.017>
- Su T et al (2021) Insufficient radiofrequency ablation promotes hepatocellular carcinoma metastasis through m<sup>6</sup>A mRNA methylation dependent mechanism. *Hepatology*. <https://doi.org/10.1002/hep.31766>
- Suhail Y, Cain MP, Vanaja K, Kurywachak PA, Levchenko A, Kalhuri R (2019) Systems biology of cancer metastasis. *Cell Syst* 9:109–127. <https://doi.org/10.1016/j.cels.2019.07.003>
- Sun S, Han Q, Liang M, Zhang Q, Zhang J, Cao J (2020) Downregulation of m<sup>6</sup>A reader YTHDC2 promotes tumor progression and predicts poor prognosis in non-small cell lung cancer. *Thorac Cancer* 11:3269–3279. <https://doi.org/10.1111/1759-7714.13667>
- Sun K et al (2021) Saikosaponin D exhibits anti-leukemic activity by targeting FTO/m<sup>6</sup>A signaling. *Theranostics* 11:5831–5846. <https://doi.org/10.7150/thno.55574>
- Tao L et al (2021) FTO modifies the m<sup>6</sup>A level of MALAT1 and promotes bladder cancer progression. *Clin Transl Med* 11:e310. <https://doi.org/10.1002/ctm2.310>
- Vera-Ramirez L, Vodnala SK, Nini R, Hunter KW, Green JE (2018) Autophagy promotes the survival of dormant breast cancer cells and metastatic tumour recurrence. *Nat Commun* 9:1944. <https://doi.org/10.1038/s41467-018-04070-6>
- Wang H et al (2019a) N<sup>6</sup>-methyladenosine induced miR-143–3p promotes the brain metastasis of lung cancer via regulation of VASH1. *Mol Cancer* 18:181. <https://doi.org/10.1186/s12943-019-1108-x>
- Wang X, Zhang J, Wang Y (2019b) Long noncoding RNA GAS5-AS1 suppresses growth and metastasis of cervical cancer by increasing GAS5 stability. *Am J Transl Res* 11:4909–4921
- Wang L et al (2020a) m<sup>6</sup>A RNA methyltransferases METTL3/14 regulate immune responses to anti-PD-1 therapy. *EMBO J* 39(20):e104514. <https://doi.org/10.15252/embo.2020104514>
- Wang M et al (2020b) Upregulation of METTL14 mediates the elevation of PERP mRNA N<sup>6</sup> adenosine methylation promoting the growth and metastasis of pancreatic cancer. *Mol Cancer* 19:130. <https://doi.org/10.1186/s12943-020-01249-8>
- Wang M, Yang Y, Yang J, Yang J, Han S (2020c) circ\_KIAA1429 accelerates hepatocellular carcinoma advancement through the mechanism of m<sup>6</sup>A-YTHDF3-Zeb1. *Life Sci* 257:118082. <https://doi.org/10.1016/j.lfs.2020.118082>
- Wang P, Wang X, Zheng L, Zhuang C (2020d) Gene signatures and prognostic values of m<sup>6</sup>A regulators in hepatocellular carcinoma. *Front Genet* 11:540186. <https://doi.org/10.3389/fgene.2020.540186>
- Wang Q et al (2020e) METTL3-mediated m<sup>6</sup>A modification of HDGF mRNA promotes gastric cancer progression and has prognostic significance. *Gut* 69:1193–1205. <https://doi.org/10.1136/gutjnl-2019-319639>
- Wang Q, Guo X, Li L, Gao Z, Su X, Ji M, Liu J (2020f) N<sup>6</sup>-methyladenosine METTL3 promotes cervical cancer tumorigenesis and Warburg effect through YTHDF1/HK2 modification. *Cell Death Dis* 11:911. <https://doi.org/10.1038/s41419-020-03071-y>
- Wang Y, Ren F, Song Z, Wang X, Ma X (2020g) Multiomics profile and prognostic gene signature of m<sup>6</sup>A regulators in uterine corpus endometrial carcinoma. *J Cancer* 11:6390–6401. <https://doi.org/10.7150/jca.46386>
- Wang S, Gan M, Chen C, Zhang Y, Kong J, Zhang H, Lai M (2021a) Methyl CpG binding protein 2 promotes colorectal cancer metastasis by regulating N<sup>6</sup>-methyladenosine methylation through methyltransferase-like 14. *Cancer Sci*. <https://doi.org/10.1111/cas.15011>
- Wang X et al (2021b) Copy number variation analysis of m<sup>6</sup>A regulators identified METTL3 as a prognostic and immune-related biomarker in bladder cancer. *Cancer Med* 10:7804–7815. <https://doi.org/10.1002/cam4.3981>
- Wang Y, Li M, Zhang L, Chen Y, Zhang S (2021c) m<sup>6</sup>A demethylase FTO induces NELL2 expression by inhibiting E2F1 m<sup>6</sup>A modification leading to metastasis of non-small cell lung cancer. *Mol Ther Oncolytics* 21:367–376. <https://doi.org/10.1016/j.omto.2021.04.011>
- Wen S, Wei Y, Zen C, Xiong W, Niu Y, Zhao Y (2020) Long non-coding RNA NEAT1 promotes bone metastasis of prostate cancer through N<sup>6</sup>-methyladenosine. *Mol Cancer* 19:171. <https://doi.org/10.1186/s12943-020-01293-4>
- Wu Y et al (2019) m<sup>6</sup>A-induced lncRNA RP11 triggers the dissemination of colorectal cancer cells via upregulation of Zeb1. *Mol Cancer* 18:87. <https://doi.org/10.1186/s12943-019-1014-2>
- Xie H et al (2020a) METTL3/YTHDF2 m<sup>6</sup>A axis promotes tumorigenesis by degrading SETD7 and KLF4 mRNAs in bladder cancer. *J Cell Mol Med* 24:4092–4104. <https://doi.org/10.1111/jcmm.15063>
- Xie JW et al (2020b) m<sup>6</sup>A modification-mediated BATF2 acts as a tumor suppressor in gastric cancer through inhibition of ERK signaling. *Mol Cancer* 19:114. <https://doi.org/10.1186/s12943-020-01223-4>

- Xie J, Ba J, Zhang M, Wan Y, Jin Z, Yao Y (2021) The m6A methyltransferase METTL3 promotes the stemness and malignant progression of breast cancer by mediating m6A modification on SOX2. *J Buon* 26:444–449
- Xu Y et al (2020) The FTO/miR-181b-3p/ARL5B signaling pathway regulates cell migration and invasion in breast cancer. *Cancer Commun (lond)* 40:484–500. <https://doi.org/10.1002/cac2.12075>
- Yang X et al (2017) 5-methylcytosine promotes mRNA export - NSUN2 as the methyltransferase and ALYREF as an m(5)C reader. *Cell Res* 27:606–625. <https://doi.org/10.1038/cr.2017.55>
- Yang S et al (2019) m(6)A mRNA demethylase FTO regulates melanoma tumorigenicity and response to anti-PD-1 blockade. *Nat Commun* 10:2782. <https://doi.org/10.1038/s41467-019-10669-0>
- Yang X et al (2020a) METTL14 suppresses proliferation and metastasis of colorectal cancer by down-regulating oncogenic long non-coding RNA XIST. *Mol Cancer* 19:46. <https://doi.org/10.1186/s12943-020-1146-4>
- Yang Z, Ma J, Han S, Li X, Guo H, Liu D (2020b) ZFAS1 exerts an oncogenic role via suppressing miR-647 in an m(6)A-dependent manner in cervical cancer. *Onco Targets Ther* 13:11795–11806. <https://doi.org/10.2147/ott.S274492>
- Yankova E et al (2021) Small-molecule inhibition of METTL3 as a strategy against myeloid leukaemia. *Nature* 593:597–601. <https://doi.org/10.1038/s41586-021-03536-w>
- Ye M, Dong S, Hou H, Zhang T, Shen M (2021) Oncogenic role of long noncoding RNAMALAT1 in thyroid cancer progression through regulation of the miR-204/IGF2BP2/m6A-MYC signaling. *Mol Ther Nucleic Acids* 23:1–12. <https://doi.org/10.1016/j.omtn.2020.09.023>
- Yoon KJ et al (2017) Temporal control of mammalian cortical neurogenesis by m(6)A methylation. *Cell* 171:877–889.e817. <https://doi.org/10.1016/j.cell.2017.09.003>
- Yue B, Song C, Yang L, Cui R, Cheng X, Zhang Z, Zhao G (2019) METTL3-mediated N6-methyladenosine modification is critical for epithelial-mesenchymal transition and metastasis of gastric cancer. *Mol Cancer* 18:142. <https://doi.org/10.1186/s12943-019-1065-4>
- Zeng Z et al (2018) Cancer-derived exosomal miR-25–3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. *Nat Commun* 9:5395. <https://doi.org/10.1038/s41467-018-07810-w>
- Zhang C et al (2016) Hypoxia induces the breast cancer stem cell phenotype by HIF-dependent and ALKBH5-mediated m<sup>6</sup>A-demethylation of NANOG mRNA. *Proc Natl Acad Sci USA* 113:E2047–2056. <https://doi.org/10.1073/pnas.1602883113>
- Zhang J et al (2019) ALKBH5 promotes invasion and metastasis of gastric cancer by decreasing methylation of the lncRNA NEAT1. *J Physiol Biochem* 75:379–389. <https://doi.org/10.1007/s13105-019-00690-8>
- Zhang L, Wan Y, Zhang Z, Jiang Y, Lang J, Cheng W (2020) Zhu L (2020b) FTO demethylates m6A modifications in HOXB13 mRNA and promotes endometrial cancer metastasis by activating the WNT signalling pathway. *RNA Biol*. <https://doi.org/10.1080/154762861841458>
- Zhang C et al (2020a) YTHDF2 promotes the liver cancer stem cell phenotype and cancer metastasis by regulating OCT4 expression via m6A RNA methylation. *Oncogene* 39:4507–4518. <https://doi.org/10.1038/s41388-020-1303-7>
- Zhang F, Yan Y, Cao X, Zhang J, Li Y, Guo C (2021) Methylation of microRNA-338–5p by EED promotes METTL3-mediated translation of oncogene CDCP1 in gastric cancer. *Aging* 13:12224–12238. <https://doi.org/10.18632/aging.103822>
- Zhao BS, Roundtree IA, He C (2017) Post-transcriptional gene regulation by mRNA modifications. *Nat Rev Mol Cell Biol* 18:31–42. <https://doi.org/10.1038/nrm.2016.132>
- Zheng ZQ et al (2019) Long noncoding RNA FAM225A promotes nasopharyngeal carcinoma tumorigenesis and metastasis by acting as ceRNA to sponge miR-590–3p/miR-1275 and upregulate ITGB3. *Cancer Res* 79:4612–4626. <https://doi.org/10.1158/0008-5472.Can-19-0799>
- Zhou J et al (2019) Gene signatures and prognostic values of m6A regulators in clear cell renal cell carcinoma: a retrospective study using TCGA database. *Aging* 11:1633–1647. <https://doi.org/10.18632/aging.101856>
- Zhou X et al (2020) Analysis of genetic alteration signatures and prognostic values of m6A regulatory genes in head and neck squamous cell carcinoma. *Front Oncol* 10:718. <https://doi.org/10.3389/fonc.2020.00718>
- Zou D, Dong L, Li C, Yin Z, Rao S, Zhou Q (2019) The m(6)A eraser FTO facilitates proliferation and migration of human cervical cancer cells. *Cancer Cell Int* 19:321. <https://doi.org/10.1186/s12935-019-1045-1>