REVIEW ARTICLE



Nanomaterials in agriculture for plant health and food safety: a comprehensive review on the current state of agro-nanoscience

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Abstract

In the modern epoch, nanotechnology took forward the agriculture and food industry with new tools that promise to increase food production sustainably. It also anticipated that it would become a driving economic force shortly. Nanotechnology has the potential to reduce agricultural inputs, enrich the soil by absorbing nutrients, manage plant diseases, and detect diseases. The aim of the present review is to cover the potential aspects of nanoscience and its trend-setting appliances in modern agricultural production. This review focuses on the impact of various nanomaterials on plant health to improve agricultural production and its cooperative approach to food production. Nanotechnology has great potential compared to conventional approaches. The appealing path of nanotrends in the farming sector raises hopes and illuminates the route of innovative technologies to overcome various diseases in plants with an enhanced yield to meet the growing global population's need for food security.

Keywords Nanomaterials · Nanosensors · Agrochemicals · Nanofertilizers · Nanopesticide · Food security

Introduction

Nanotechnology is a novel scientific approach capable of manipulating physical and chemical properties at molecular levels. On the other hand, biotechnology manages molecular, genetic, and cellular processes to develop products and services. Nanotechnology finds use in diverse fields, from medicine to agriculture, utilising the knowledge and techniques of biology (Fakruddin et al. 2012; Manjunatha et al. 2019). Agriculture, the backbone of developing countries, is the most important and stable sector because it produces and provides the raw materials for food, feed, fibres, and many other products by cultivating plants and raising livestock. Agriculture holds more than 60% of the population depending on it for their livelihood (Brock et al. 2011). The existing natural resources (production land, water, soil, etc.) may not be enough for the burgeoning population that is estimated

to be 8 billion people by 2025 and 9 billion by 2050, and to accomplish the global population requirement, global agricultural production must increase. Agri-food production is one of the primary drivers of the economy. Besides, agriculture offers routes to value-added crops (Ghasemzadeh 2012; Mukhopadhyay 2014; Prasad et al. 2017; Berners-Lee et al. 2018; Mohammad Fakhrul and Karim 2020). On less available land, we have to achieve more food production, which can only be accomplished by intensifying production, in an environmentally safe manner through an ecological increase in yield per unit of area, i.e. escalating production, while meeting acceptable standards of environmental quality (Cassman 1999; Cordell and White 2013). Many believe that the intervention of plant biotechnology holds the key to producing more food. Only genetic advancement cannot fulfil the requirement, so it has to be associated with other materials such as nanofertilizer.

Nanotechnology has an impact due to the alteration of atoms and the development of magnetic power. The nanoparticles (NPs) with a smaller size possess a larger surface area and exhibit more active properties. Nanotechnology is knocking on the doors of perception shortly. In biological processes, nanomaterials play a vital role (Pokropivny et al. 2007; Prasad 2014). Nanotechnology helps in the agricultural and food industries to understand molecular



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management of diseases, rapid disease detection, enhancing the ability of plants to absorb nutrients, among others. Nanobiotechnology of various crops, on the other hand, has the potential to improve yields or nutritional values, as well as develop improved systems for monitoring environmental conditions and enhancing plants' ability to absorb nutrients or pesticides by improving our understanding of biology (Tarafdar et al. 2013). There is a significant demand for fast, reliable, and low-cost systems to detect, monitor, and diagnose biological host molecules in agricultural sectors (Vidotti et al. 2011; Sagadevan and Periasamy 2014). Nanomaterials may include synthesis from plant, bacteria, fungi system; it is considered as a green nanotechnology. Nowadays chemically synthesized nanomaterials are found to be toxic in nature (Prasad 2014). Green nanotechnology has a profound influence on the environment; it is energy efficient. It also reduces waste and greenhouse gas emissions (Prasad 2014: Prasad et al. 2016: Maksimovic and Omanovic-Miklicanin 2017; Verma et al. 2019). In the present decade, green nanotechnology implementation and its functions are still not clear. How will the environmental sustainability of green nanotechnology be achieved in the future? These risks must be mitigated in advancing green nanotechnology solutions (Kandasamy and Sorna Prema 2015). The engineered smart nanotools in modern agricultural systems can make a revolution in agricultural practices and enhance both the quality and quantity of yields (Singh Sekhon 2014; Liu and Lal 2015).

Approaches

Multiple literature searches in Google Scholar, PubMed, and Scopus using the terms nanomaterials, agrochemicals, nanofertilizers, food security, and safety, as well as handsearching recent reviews, were conducted to determine the efficacy of nano-resources in agriculture and food. We included only studies that were published in English and excluded those that were only available in abstract form. Literature search and study selection criteria in order to investigate the efficacy of nanomaterials, multiple literature searches were performed in PubMed, Google Scholar, and Scopus using the terms nanomaterials, agrochemicals; nanofertilizers, food security, and safety, as well as hand searching recent reviews. Only studies published in English were included, and those only available in abstract form were excluded. For observational studies, searches were made for reports of nanomaterials' apparent consequences on crop production and their amicable attitude toward quality food fabrication. What are the requirements for research organisations to use natural solutions to improve the growth and health potential of plants? Aside from minimising the use of hazardous chemicals for short-term benefits, it is



also critical that the ecosystem's strength be improved to bring long-term benefits to agricultural commodities. The critical appraisal of nanomaterials in the path of the review was discussed below, including an analysis of the potential preconceptions of its implications in agriculture and food supplementation.

Insight seascape of nanomaterials for sustainable plant growth and development

Improved agronomic and bioenergy crop production can fulfil the world's rising food and vitality demands through emerging scientific attention and the use of nanomaterials in fertilizers (Boutchuen et al. 2019). Agricultural innovations are crucial for the explosion of the worldwide population through the use of natural and manmade assets. In particular, multiple agriculture-related problems can be solved with the help of nanotechnology (Fig. 1). NPs provide pronounced scientific attention to link the gap between bulk constituents and minute atomic or molecular structures (Prasad et al. 2017; Shang et al. 2019). NPs-based fertilizers are to be used effectively to transport essential nutrients for plants while reducing fertilizer input into the environment (Guo et al. 2018; Boutchuen et al. 2019). The properties of the NPs are important because they influence the morphological and physiological properties of cells during their interaction with plants (Fig. 1; Siddiqui et al. 2015a). Both optimistic and adverse effects on plant growth and development by the use of NPs have been already reviewed by many researchers from their finding (Nowack and Mueller 2008; Handy et al. 2008; Ju-Nam and Lead 2008; Monica and Cremonini 2009; Stampoulis et al. 2009; Kahru and Dubourguier 2010; Ma et al. 2010, 2013; Nair et al. 2010; Menard et al. 2011; Peralta-Videa et al. 2011; Khot et al. 2012; Pan and Xing 2012; Smita et al. 2012; Aslani et al. 2014; Deng et al. 2014; Gardea-Torresdey et al. 2014; Aliofkhazraei 2015; Siddiqui et al. 2015a, b; Chichiriccò and Poma 2015; Huang et al. 2015; Abd-Alla et al. 2016; Le Van et al. 2016; Wen et al. 2016; Gil-Díaz et al. 2016). Engineered NPs have an impact on a plant's system depending on their composition, concentration, size, physical and chemical properties, and also on the plant's species.

Impact of nanoparticles on plant health

Nano-silicon dioxide (SiO₂) in the seed germination

The role of NPs in plant seed germination, growth (shoot and root biomass), and photosynthesis have been illustrated to some extent in many plants. The emblems of plant growth

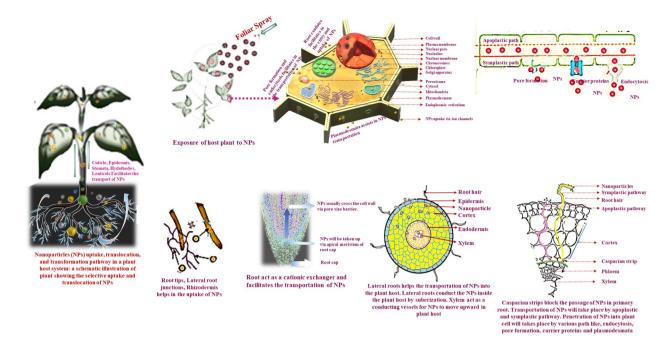


Fig. 1 Transverse section of the plant cell/root absorption zone showing the nanoparticle entry and interaction in the host plant

and development start with seed germination, followed by elongation of root and shoot development. Concerning plant growth and development, reports are available on the effect of the concentration-dependent effect of NPs on seed germination (Fig. 1). Nano-SiO₂, with its lower concentration, improves seed germination parameters in tomatoes (Siddiqui and Al-Whaibi 2014). Suriyaprabha et al. (2012) illustrated the influence of nano-SiO2 with enhanced seed germination in maize seeds by providing restored nutrient availability, pH, and good conductivity to the growth medium. Further, nSiO₂ significantly enhanced the potential of seed germination, average germination time, seed germination and seed vigour index, fresh seedling weight, and dry weight. Nano-SiO₂ application is beneficial for plant growth and enhanced yield. According to Bao-Shan et al. (2004), nano-SiO₂ has a positive effect on *Larix olgensis* seedlings. Nano-SiO₂ improves seedling growth quality parameters, with enriched mean plant height, increased root collar width, increased primary root length, and an amplified number of lateral roots, and also elicits the synthesis of chlorophyll in the host plant. According to Haghighi and Teixeira Da Silva (2014), nano-SiO₂ improves seed germination under abiotic stress. In tomato, Siddiqui and Al-Whaibi (2014) demonstrated that nano-SiO₂ improves seed germination and elicited the antioxidant activity in squash under NaCl stress conditions. Shah and Belozerova (2009) and Frenk et al. (2013) tested gold (Au), silica (Si), palladium (Pd) and copper (Cu) NPs and observed that all these NPs significantly impact seeds and their growth. Also, in soybean, the exogenous application of nano-titanium dioxide (nano-TiO₂) and nano-SiO₂

increases seed germination by improving nitrate reductase (Maity et al. 2018; Acharya et al. 2020; Shinde et al. 2020) and enhances the seeds ability to absorb water and nutrients in spinach, it increases chlorophyll content and seed germination. Under salinity stress, nano-SiO₂ recovers leaf fresh and dry weight, chlorophyll content, and proline accumulation, demonstrating that quantum dots (QDs)-coated silica promotes root growth in rice significantly (Haghighi and Teixeira Da Silva 2014; Karimi and Mohsenzadeh 2015; Rastogi et al. 2019). Nano-SiO₂ promotes plant growth and development by increasing photosynthetic net rate, transpiration frequency, stomatal conductivity, PSII potential action, active photochemical efficacy, profound photochemical proficiency, electron transport rate, and photochemical quench (Siddiqui and Al-Whaibi 2014; Felix Alvarez et al. 2018; Pushpa and Lohani 2018; Zhu et al. 2019; Madany et al. 2020).

Iron nanoparticle

Iron (Fe) is a vital nutrient for all living beings (Zuo and Zhang 2011). Recently, it has been illustrated that iron oxide (Fe_2O_3) NPs have the potency to be an ideal substitution for the traditional Fe fertilizer (Rui et al. 2016). Zero-valent iron NPs (nZVI) elements can penetrate into the peanut seed coats, which improves the uptake of water and fuels the germination of seeds. The growth rate experiments indicated that at higher concentrations, it shows phytotoxicity, while at lower concentrations it stimulates the plant's growth and root development (Li et al. 2015; Li and Lan 2017).



Another study shows that Fe₂O₃ NPs stimulated the growth of peanuts by regulating phytohormone contents, antioxidant enzyme activity, and other parameters also. Accordingly, the likely replacement of the Fe fertilizer with NPs is possible (Rui et al. 2016). The effect of copper (Cu), zinc (Zn), manganese (Mn), and FeO NPs on Lactuca sativa reveals that CuO NPs were slightly more toxic than Cu ions, whereas zinc oxide (ZnO NPs) was similar to that of Zn ions. MnOx NPs and FeOx NPs were less toxic than their ionic forms and also significantly activated the growth of lettuce plant seedlings by 12–54%. It also showed that synthetic NPs were not always more toxic than other chemical types containing the same elements. Mn or Fe NPs can significantly improve the growth of the plant and can be effective nanofertilizers for increasing agronomic productivity (Liu et al. 2016; Zia-ur-Rehman et al. 2018; Rizwan et al. 2019). However, further advancement in this direction is still required to know its possible threat to the environment and to food safety.

Zinc oxide nanoparticles

Zinc (Zn) has a foremost role as a biotic micronutrient in plant health systems, and it plays significant roles in metalprotein complexes (Sturikova et al. 2018). Prasad et al. (2012) investigated the pivotal role of ZnO NPs in plant growth and development. Lower concentrations of ZnO NPs were found to have a positive effect on seed germination, seedling vigour, plant growth, flowering, chlorophyll content, root length, and pod yield in Brassica nigra (Sedghi et al. 2013), onion (Raskar and Laware 2014), and wheat (Ramesh et al. 2014). At higher concentrations of ZnO NPs, they significantly inhibit the germination of seeds. The NP influence on seed germination depends on the concentrations of NPs and differs from plant to plant. Studies on the various concentrations of ZnO NPs in alfalfa, cucumber, and tomato plants found that only cucumber plant seed germination was effectively boosted (de la Rosa et al. 2013). Tarafdar et al. (2013) reported that ZnO NPs significantly improve Cyamopsis tetragonoloba plant shoot and root growth, biomass, root area, protein and chlorophyll synthesis, rhizospheric microbial population, alkaline phosphatase, acid phosphatase, and phytase activity in the cluster bean rhizosphere region. It is also apparent by the scanning microscope, correlative light, and inductively coupled plasma/ atomic emission spectroscopy analysis, that ZnO NPs on the roots of Vigna radiata and Cicer arietinum absorption, stimulated the shoot and root lengths and their biomass (Dhoke et al. 2011; Burman et al. 2013). Also in three Solanaceae crops, e.g., Solanum melongena, Lycopersicon escu*lentum*, and *Capsicum annum*, enhanced germination rates and shoot-root biomass were recorded (Younes et al. 2020). Under in vitro conditions, the Nano ZnO supplemented with Murashige and Skoog (MS) media encouraged shooting,



somatic embryogenesis, regeneration of plantlets, elucidated proline synthesis, superoxide dismutase activity, catalase, and peroxidase, thereby inducing resistance to biotic stress (Murashige and Skoog 1962; Elizabath et al. 2017; Ahmad et al. 2020). ZnO NPs show significant influence at varied concentrations on germination, vigour, and premature growth of Capsicum annuum plantlets and also show a better increase of phenolic composites and antioxidant activity in the radicle. ZnO-NPs can elicit the production of desirable secondary metabolites (Garciá-López et al. 2018). ZnO NPs were tested on wheat (Triticum aestivum) grown in hydroponic media. At 50 mg/l, the NP positively affects seed germination rate, the number of roots, plant biomass, and the total growth factors of shoots, roots, and leaves (Awasthi et al. 2017). The development of multiple antioxidant defence biomarkers during the lifecycle of soil-grown soybean (Glycine max cv. Kowsar) and the maximum oxidative stress responses (H₂O₂ synthesis, catalase (CAT), peroxidase (POD), and superoxide dismutase (SOD)) under these NPs treatment was observed. ZnO NPs have the potential to be used as a nanofertilizer for crops cultivated in Zn-deficient soils to recover crop yield, improve food quality, and address malnutrition, globally (Yusefi-Tanha et al. 2020).

Nanoparticles influence in photosynthesis of plant

Plants' ability to change light energy into chemical energy is known as photosynthesis. Although photosynthesis is a vital process in plants, they can only convert 2-4% of the accessible energy in radiation into fresh vegetal growth (Kirschbaum 2011). Advanced research on the point of photosynthesis in plants was done previously through plant breeding and gene manipulation techniques. Recently, studies have demonstrated that incorporation of the two genes of Cyanobacterium, and Synechococcus in tobacco plants leads to enhanced activity of Rubisco's gene in the plant for carbon fixing (Lin et al. 2014; Liang and Lindblad 2017; Liang et al. 2018; Long et al. 2018). Photosynthesis in engineered plants is more efficient than in native plants. The photosynthesis process was faster than expected because the carbon dioxide fixation was working well. Furthermore, in the corral of nanobiotechnology, investigators want to develop bionic plants that can have improved photosynthesis efficacy and biochemical sensing. This study found that the single-wall carbon nanotubes (SWCNTs) in the quarantined chloroplast increased three times greater photosynthetic action than that of controls with significantly improved electron transport rates, and SWCNTs assisted the plants to sense the nitric oxide signalling molecule. The involvement of nanobionics in engineered plants would facilitate innovative and advanced functional properties in the photosynthetic system. Noji et al. (2011) discovered that the complex photosystem II (PSII) absorbed into a nanomesoporous silica compound (SBA) and activated the constant activity of a photosynthetic oxygen-evolving reaction, indicating light-driven electron transport from water to quinone molecules. They suggested that the PSII-SBA conjugate might possess the properties needed to develop artificial photosynthetic systems and photosensors. SiO₂NPs improve the rate of photosynthetic activity by enhancing carbonic anhydrase activity and the synthesis of photosynthetic pigments (Siddiqui et al. 2014). Photosynthesis may improve with the supply of carbonic anhydrase and CO₂ to the Rubisco (Studer et al. 2014; Rudenko et al. 2020). The TiO₂ nanoanatase has a distinctive photocatalyzed feature; it also increases the absorbance of light and the transformation of light energy into electrical and chemical energy and elicits carbon dioxide acclimatization. It is possible that by stimulating Rubisco (a multifaceted form of Rubisco and Rubisco activase), encourage Rubisco carboxylation, thereby increasing plant growth. TiO₂ NPs protect chloroplasts from ageing in long-term illumination (Gao et al. 2006; Studer et al. 2014; Ahmad et al. 2018; Gohari et al. 2020; Suganami et al. 2020). The nano-anatase effect on the molecular mechanism of the carbon reaction proposes that increasing the Rubisco activase protein levels and activities by eliciting the marker gene for Rubisco activase (RCA) mRNA.TiO₂NPs improve the net photosynthetic rate, water conductivity, and transpiration ratio in plants by improving Rubisco carboxylation and increasing the frequency of the photosynthetic carbon reaction (Suganami et al. 2020). As stated by Khatri and Rathore(2018) and Zhang et al. (2020), nano-anatase vigorously promotes the entire electron transport chain, activity of photoreduction in photosystem II, the evolution of O₂, and photophosphorylation activity in chlorophyll under both ultraviolet and visible light. According to Govorov and Carmeli (2007), metal NPs can activate photosynthetic systems by enhancing the efficacy of biochemical energy production. AuNPs and silver (Ag) nanocrystals bind to the centre of the photosynthetic reaction, thereby forming a novel hybrid system that produces exciting electrons around ten times more, due to plasmon resonance and fast electron-hole separation. The improved mechanisms may aid in the plan for artificial light-harvesting schemes. Pradhan et al. (2015) demonstrated that Cu NPs have a significant influence on the elevation of Chl a content in mung bean plants, indicating that it has a significant influence on the photosynthesis process, as Chl a play an important role in the light-dependent photosynthetic pathway. They also observed higher quantities of carotene and xanthophyll in plants treated with Cu NPs as compared to controls; these mechanisms in the photosynthetic process provide a protective shield for plants against external stress environments. Furthermore, they stated that Cu NPs activate

photophosphorylation, the electron transport chain, and the carbon assimilatory pathway in the chloroplast to enhance the process of photosynthetic activity in treated mung bean plants.

Chahardoli et al. (2020) investigated the engineered aluminium and nickel oxide NPs' (Al₂O₃ and NiO NPs) responses at different concentrations on the growth of the plant, oxidative stress, and antioxidant activities in the hydroponically grown tissues of Nigella arvensis L. At lower concentrations, the biomass of the plant was significantly enhanced under Al₂O₃ and NiO NP treatment, but reduced at higher concentrations of NP. Several enzymatic antioxidants assayed in shoots and roots, such as ascorbate peroxidase (APX), CAT, POD, and SOD, specify an overall upsurge of activities after exposure to NPs. The antioxidant activities in the plant were enhanced, as were secondary metabolite formation and other allied physiological factors such as total antioxidant activity, total saponin content, and DPPH scavenging activity (Hong et al. 2005; Zheng et al. 2005; Gao et al. 2006; Yang et al. 2006). These results illustrated that nanosized TiO₂ has a positive influence on the plant growth of spinach, whether it is treated in the seeds or sprayed on the leaves. Nano-TiO₂ significantly increased the SOD, POD, and CAT activities, reduced the increased accumulation of reactive oxygen free radicals and malondialdehyde (MDA) levels, upheld the stability of the membrane, and promoted the adsorption of nitrate and stimulated the conversion of inorganic into organic nitrogen. Also, in the spinach leaves, it shows the activity of Rubisco carboxylase was 2.67 times higher than that of Rubisco in the control, and it can hydrolyze ATP in the same way as Rubisco activase. Rubisco and Rubisco activase were activated in spinach, which increased Rubisco carboxylation and photosynthetic carbon reactions. Normal-sized TiO₂ did not have these effects (Gao et al. 2006). One study reported that NP interaction with green algae, which has a cell wall similar to plants, and a toxicity study of nanosized TiO₂ under illumination indicate that NP is slightly inorganic; oxidic NP can interact with plant cells or green algae by way of a parallel cell wall assembly absorbed at the surface of the root and intake into the cell. The NPs can diffuse into cells via the path of intercellular space, the apoplast, and be adsorbed or merged into membranes (Hund-Rinke and Simon 2006). Because plant surfaces carry negative charge into the apoplast, plant cells easily transport negative charge NPs. The Casparian strip forms a barricade to the apoplastic stream flow, and only symplastic passage is possible into the xylem. Conversely, this barricade is not impeccable, which means compounds can go into the xylem through holes or damaged cells without even crossing a cell membrane and being further transported to the shoots. This mechanism was found to be the prime process for the uptake of metal complexes with chelators like EDTA and their transportation to the shoots



(Nowack et al. 2006; Tandy et al. 2006). Based on these facts, we could theorize that, negatively charged NPs may possibly pass into the apoplast of the root cortex and eventually also into the xylem part.

Promising prospects for nanotechnology for the improved plant health and food production

Massive quantities of fertilizers like urea, ammonium salts, and phosphate or nitrate compounds have a considerable impact on improved food production. Nonetheless, they contribute to a slew of negative effects on beneficial soil microflora. Most of the applied fertilizers are not properly uptaken by plants due to overflow, which causes pollution (Peterson 1975; Prashar and Shah 2016; Berg et al. 2017). Fertilizers covered in nanomaterials can resolve this problem. Nanomaterials have probable influences on the gentle discharge of fertilizers, as NPs hold the material more stably in the plant due to the high surface tension of NPs than the conventional approach. Furthermore, nanocoatings provide surface safeguards for more substantial elements (Giroto et al. 2017; Kottegoda et al. 2017; Niemiec and Komorowska 2018).

Agrochemicals

As the world population increases, the food crisis can be alleviated by utilising plant breeding, fertilizers, and plant protection products. After the green revolution era in India, the use of nitrogen in fertilizer with urea effectively increased production by around 29%. However, rampant use of nitrogen in the production of food increases atmospheric N₂O (a greenhouse gas), which influences increased atmospheric temperature and thus contributes to global warming (Park et al. 2012; Fagodiya et al. 2017; Kumar and Indira 2017; Eliazer Nelson et al. 2019; Manuel 2019; Dimkpa et al. 2020). In agriculture, chemical fertilizers like diammonium phosphate (DAP), urea, and single superphosphate (SSP) are used to meet the deficiency of N, P, and K in the soil. However, most parts of these chemical fertilizers are wasted as vapour or run-off. Among applied fertilizers most of which go to the environment, it is estimated that about 40-70% of the nitrogen, 80–90% of the phosphorus, and 50–70% of the potassium cannot be absorbed by a plant, causing loss to the nation and environmental pollution as well (Trenkel 1997; Ombódi and Saigusa 2000). The recent methodology deceits in the usage of nanocoated urea or other chemical fertilizers. The nanocoating's constancy reduces the degree of dissolution of the compost and permits the slow, constant release of coated fertilizer, which is more efficiently absorbed by the plant roots. It was earlier reported that slow-release fertilizers have become an effective way to save fertilizer consumption while minimizing environmental pollution (Wu et al. 2008). In sulphate-deficient soil, sulphur nanocoating



fertilizers (≤ 100 nm layer) are worthwhile as slow-release fertilizers, since sulphur substances are beneficial (Ronghao et al. 2016; Duhan et al. 2017; Klikocka and Marks 2018). Nanocoated urea and phosphate release will be helpful, and crop demands will be fulfilled for most deficient soil nutrients. Many biodegradable polymeric materials, such as chitosan NPs (~78 nm), have shown promising results for slow NPK fertilizer release (Corradini et al. 2010; Adu-Gyamfi et al. 2019). Polymeric and kaolin biocompatible NPs have potential applications in fertilizer slow discharge actions (Wilson et al. 2008). The nanofertilizers achieve the slow release of fertilizer nitrogen and phosphorus with the plant's absorption, thus preventing the loss of nutrients and avoiding the undesirable nutrients' interactions with microorganisms, air, and water (Emadian et al. 2017). Nutrient absorption by the plants from the soil can be improved using nanofertilizers. The encapsulated form of nanosilica, which forms a dual film on the cell wall of bacteria or fungi after absorption of nutrients and inhibits infections, has a beneficial effect on plant growth under high temperatures and humidity and improves plant resistance to disease infections (Wang et al. 2002; El-shetehy et al. 2020). Silicon-based fertilizers are practiced to activate the resistance of plants, as silicon dioxide NPs can recover the growth of the seedling and root development (Nair et al. 2010) TiO₂, or titanium in its non-toxic form, is used as an additive in fertilizers to enhance food production(Ropers et al. 2017; Lyu et al. 2017). The additives in fertilizers can increase water retention (Emadian et al. 2017). Also, nanoclays play an essential role in the slow release of nitrogen fertilizer (Sharmila et al. 2010). The synthesis of chitosan NPs has been tested with polymethacrylic acid (PMAA) for stuffing NPK fertilizers(Hasaneen et al. 2014). The addition of N, P, and K has a greater anion charge from the calcium phosphate than the potassium chloride and urea charges(Hasaneen et al. 2014; Khalifa and Hasaneen 2018). As a result, it has greater chitosan NPs stability in colloidal polymethacrylic acid (CS-PMAA) suspension (Hasaneen et al. 2014). The dispersals of CS-PMAA pooled with nitrogen are more unstable than those of phosphorus. Positive changes in the colloidal diffusion of CS-PMAA decreased above 500 ppm of nitrogen due to negative groups from the urea molecules (Emadian et al. 2017). For diffusion with potassium, the stability of the solution has been confirmed with the addition of 400 ppm. It revealed that Cl ions (from KCl) didn't disturb the stability of colloidal dispersal with the addition of up to 400 ppm of concentration (de Vasconcelos et al. 2006; Wu et al. 2006).

Nanofertilizers

Chemical fertilizer is a major problem and a hindrance to achieving reasonable sustainability in agriculture, with limited nutrient use efficiency and environmental constraints associated with chemical fertilizer use. Also, the resulting cost increase of chemical fertilizer reduces profit margins for growers due to overapplication (Zulfiqar et al. 2019). Agriculture production, crop quality, and about 40%–60% of the total global food fabrication depend on plant nutrition (Roberts 2009). Nanofertilizers are economical for the producer as compared to traditional fertilizers. Also, it increases nutrient efficiency and improves plant nutrition and growth (Table 1). Improvement of nutrient efficiency through nanotechnology helps develop the nanofertilizer. Three classes of nanofertilizers have been proposed:

Nanoscale fertilizers (nutrients loaded with or coated with NPs),

Nanoscale additives (nanoscale additives with traditional fertilizers), and

Nanoscale coating (traditional fertilizers contain NPs).

These nanomaterial coatings (such as nanomembranes) can reduce nutrient discharge or increase the solubility of permeable nanofertilizers. Nanotechnology for fertilizers is still in its infancy; on the other hand, it has been adopted for medical and engineering applications (Mikkelsen 2018).

Nanotechnology makes farming more productive by solving problems that haven't been solved before at the smallest possible scale, which gives researchers more hope. Nanotechnology has the potential to transform agricultural production in plant and animal production by enabling better management and conservation. Nanotechnology can meet the rising population's future requests because NPs have exclusive physicochemical characteristics, i.e. high reactivity, high surface area and tuneable pore size (Abobatta 2018). Sustainable agricultural production and efficiency are not possible without agrochemicals, such as pesticides, fertilizers, etc. Plant growth and development depend on fertilizers. Still, they are essential because most useful fertilizers are inaccessible to plants owing to numerous issues, such as leaching, degradation by photolysis, decomposition, and hydrolysis. So, minimizing the fertilization losses and increasing the crop yield with nanomaterials and nanotechnology. Nanofertilizers or nanoencapsulated nutrients possess the properties that are effective for crops, they release the nutrients on-demand, organize the discharge of chemical fertilizers that control plant growth, and improve objective activity (Monreal et al. 2016; Rai et al. 2015). Agrochemicals are problematic in one way or another, like pollution of water or deposits on food products that threaten human beings and ecological health conditions (Carvalho 2017). Therefore, the development of the modern agricultural system with the use of nanotools is an excellent strategy to make a revolution in agricultural practices; nanotechnology can reduce and eliminate the influence of fertilizer, pesticides, etc. on the atmosphere along with enhancing both the quantity and quality of crop yields (Singh Sekhon 2014; Liu and Lal 2015). Nanoparticles treatment to plants and application of nanofertilizers exhibited profound influence on the secondary metabolites content and antioxidant potential status as reported in various plants (Table 2). Similarly foliar spray and seed priming of silver, zinc, copper and gold nanoparticles is reported to exhibit improved growth and productivity in filed as shown in fenugreek, maize, mustard and foxtail millet (Table 3).

Nanopesticides

In recent years, due to the high toxicity and non-biodegradability of pesticides, environmental problems have brought attention to nanofertilizers. Athanassiou et al. (2018) state that nanoformulations are similar to other regular pesticide formulations in that they help in increasing the apparent solubility of a poorly soluble active element or liberating the active element in a relaxed or directed manner, thus protecting the active component against early degradation. Pesticides developed using the nanoformulation method are called "nanopesticides." The preparation of nanopesticides can be accomplished either by using tiny particles of pesticidal active components or additional nanostructured particles with pesticidal properties. Investigators have advanced diverse nanopesticides, such as nanocapsules, nanogels, nanoemulsions, nanospheres, and metal and metal oxide NP formulations (Hayles et al. 2017; Kah and Hofmann 2014). Nanopesticides are showing viable potential to overcome these limitations. Slow degradation and controlled release of active ingredients in the presence of suitable NMs can offer effective pest control over a long period of time (Chhipa 2017). Therefore, nanopesticides are essential for the effective and sustainable management of different pests and minimize the use of synthetic chemicals and their associated environmental risks(Côa et al. 2019) Nanopesticides behave differently from conventional pesticides to increase their efficacy (Kah et al. 2019). It includes the insertion of nanoscale active ingredients into pesticides. The specific properties of these nanoscale materials, like their aptitude to dissolve in water more efficiently than the existing products or their increased stability, are designed to maximize these pesticides' effectiveness. Various kinds of nanoformulations can be used for agrochemicals like nanoencapsulations, nanoemulsions, nanosuspensions, etc. Some common welfare benefits of NPs based pesticide inventions include the augmented solubility of water-insoluble active elements, improved steadiness of the formulation, the removal of lethal organic solvents in contrast to conventionally used pesticides, the ability to slow the discharge of active elements, recovery of their permanence and avoid their early degradation, improve their flexibility and greater insecticidal



Table 1	Overview	of developed	l nanofertilizer an	d its influence	on plant	t growth parameters
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Nanofertilizer	Crop name	Amount of Nanofertilizer (ppm)	Impact	References
Au	Pearl millet (Pennisetum glaucum)	50	Improved seed germination and growth of seed- lings	Parveen et al. (2016)
Ca	Peanut (<u>Arachis hypogea</u>)		Nutrient content increased in shoot and root	Liu et al. (2004)
CeO ₂	Cucumber (Cucumis sativus)	400	Improved scratch and globulin content	Zhao et al. (2014)
CNTs	Date palm (<i>Phoenix dactylifera</i>)	0.05-0.1	Shoot length and number of leaf increased	Taha et al. (2016)
	Tobacco (Nicotiana tabacum)	5-500	55-64% plant growth improved	Khodakovskaya et al. (2012)
Cu	Lettuce (Lactuca sativa)	130-600	Shoot and root length increased	Shah and Belozerova (2009)
CuO	Maize (Zea mays)	10	51% plant growth	Adhikari et al. (2016)
Fe/SiO ₂	Barley (<i>Hordeum vulgare</i>) and maize (<i>Zea mays</i>)	0–25	Improved mean germination time	Najafi Disfani et al. (2016)
FeO	Soybean (<i>Glycine max</i>)	30- 60	Chlorophyll increased	Ghafariyan et al. (2013)
	Pea (Pisum sativum)	250-500	Seed weight and chlorophyll increased	Delfani et al. (2014)
Mg	Cow peas (Vigna unguiculata)	2.5	Increment in stem Mg content, chlorophyll content and plasma membrane stability	Delfani et al. (2014)
Mn	Mung bean (Vigna radiata)	0.05–1	Shoot length, chlorophyll content and the photo- synthesis rate increased	Pradhan et al. (2013)
	Rice (Oryza sativa)		Improved Zn uptake 5.66 mg/hill	Yuvaraj and Subramanian (2015)
Mo	Chickpea (<i>Cicer arietinum</i>)	8	Plant mass and number of modules increased	Taran et al. (2014)
Р	Soybean (<i>Glycine max</i>)	100	Improved the growth rate and seed yield by 32.6% and 20.4%	Liu and Lal (2014)
TiO2	Spinach (Spinacia oleracea)	0.25–4	Plant dry weight increased by 73%	Zheng et al. (2005)
	Mung bean (Vigna radiata)	10	Improvement in plant growth and nutrient content	
	Cowpea (Vigna unguiculata)	125	Cowpea yield up to 26–51%	Owolade and Ogunleti (2008)
	Spinach (Spinacia oleracea)		N ₂ fixation improvement	
Zn	Ryegrass (<i>Lolium</i>)	01–2000	Root elongation	Lin and Xing (2008)
	Rice (Oryza sativa)	0–150	increase of root and shoot length in rice	Upadhyaya et al. (2015)



Table 1 (continued)

Nanofertilizer	Crop name	Amount of Nanofertilizer (ppm)	Impact	References
ZnO	Mung bean (<i>Vigna radiata</i>) and chickpea (<i>Cicer arietinum</i>)	01–2000	Growth of the plant improved at 20 ppm in chick pea and mung bean at 1 ppm	Mahajan et al. (2011)
	Cucumber (Cucumis sativus)	400-800	Root dry weight and fruit gluten increased	
	Rape seed (Brassica napus)	01–2000	Root elongation	Lin and Xing (2007)
	Peanut (Arachis hypogea)	1000	34% increment in pod yield per plant	Prasad et al. (2012)
	Chickpea (Cicer arietinum)	1.5	Improved shoot dry weight and antioxidant activity	Burman et al. (2013)
	Maize (Zea mays)	10	Improved plant height and dry weight	Adhikari et al. (2015)
	Clusterbean (Cyamopsis tetragonoloba)	10	Improvement in plant growth and nutrient content	Raliya and Tarafdar (2013)

movement due to reduced particle size and higher surface area, and extend their longevity (Usman et al. 2020).

Nanotechnology for food safety and production

Nanotechnology in the farming and food segments is a relatively new trend compared with pharmaceuticals and drug delivery. Agri-food nanotechnology is multidisciplinary in nature. Nanotechnology has a wide-ranging application in agriculture and the food sector, such as protecting vegetation, nursing plant growth, detecting animal and plant diseases, growing worldwide foodstuff fabrication, enhancing food quality features and reducing waste (Omanović-Mikličanin and Maksimović, 2018). The advancement of nanosized food materials and additives as delivery methods for bioactive composites and pioneering food packing were studied (Nile et al. 2020). Nowadays, with the world's population getting higher, fulfilling all the requirements of agriculture and nutrition for food is difficult. Therefore, we have to adopt new technologies to focus on and minimize this problem to get better agricultural production. Nanotechnological applications can help to achieve sustainable agriculture and food nutrition supply (Bajpai et al. 2018). Recently, food and nutrition abundantly implanted in acquaintances. Specific target-oriented goals in agriculture development depend on health, climate change, social inclusion, natural resources, ecosystem progressions, energy, moral supremacy, etc., and must be acknowledged (Meybeck et al. 2018). With sustainable agricultural practices, nanotechnology has the potential to alleviate people's hunger and poverty. In the agriculture

zone, an ecological recital is essential, and simultaneously, the contribution of food chain networks is also a prerequisite for agrarian food fabrication (Prasad et al. 2017; Usman et al. 2020). The global population in 2050 needs about 200 million tons of annual meat. The United Nations of Food and Agriculture Organization has already foreseen fulfilling the food requirements for the increasing population and the pressure and further demands of livestock with land to produce feed (FAO 2017). Also, the need for land fronting competition from the crops for other determinations like, biofuels and pharmaceuticals. So, the production of food faces numerous challenges, including a falling ratio of available land due to an increasing population. Agronomy as a food source is an essential trait in a global world of weakening resources and an increasing human population. Modern technologies like nanotechnology and nanobiotechnology are important for agronomic and food skills to meet the increasing world population. Nanotechnology revolutionizes farming and associated fields, together with fisheries and aquaculture. Agricultural practice now includes nanoagriculture, which uses nanosized particles to increase livestock and crop production (Scott and Chen 2013; Ghidan and Antary 2019; Joshi et al. 2019; Mohammad Fakhrul and Karim 2020).



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Table 2 Table representing the influence of nanoparticles on plant secondary metabolites and antioxidant activity

Plant	Nanoparticles	Effect of Nanoparticles	Reference
<i>Echinacea purpurea</i> (Purple coneflower) callus	ZnO NPs	Secondary metabolite Flavonoids 3.7 fold	Karimi et al. (2018)
Phaseolus vulgaris (Common bean) in vitro and in vivo	Ag and Cu NPs	Physiological Characteristics	Mustafa et al. (2017)
Calendula officinalis (Marigold)	Methyl Jasmonate and AgNPs	MDA, Chlorophyll, Saponin, Carotenoid, Flavonoids	Ghanati and Bakhtiarian (2014)
Eruca sativa (Garden rocket)	Ag, Cu, and Au NPs	seed germination and seedling growth in	Zaka et al. (2015)
<i>Trigonella foenum-graecum</i> (Fenugreek)	AgNPs	Plant growth and diosgenin	Jasim et al. (2017)
Artemisia annua (Sweet wormwood) (Callus)	CoNPs	artemisinin production and gene expression	Ghasemi et al. (2015)
Prunella vulgaris (Common Self-Heal) (Callus)	Ag and Au NPs	Phenolics, Flavonoids, and Anti- oxidant activity	Fazal et al. (2016)
Hyoscyamus reticulatus (Lattice Henbane)	FeO NPs	hyoscyamine and scopolamine	Moharrami et al. (2017)
<i>Capsicum annuum</i> (Cayenne pepper)	CuO NPs	Life cycle	Rawat et al. (2017)
Cicer arietinum (Chickpea) Vigna radiate (Mung bean), Pha- seolus vulgaris (Black bean), and Phaseolus vulgaris (Red bean)	Hematite NPs	Growth of the plant	Boutchuen et al. (2019)
Allium cepa (Bulb onion)	ZnO NPs	Growth, Flowering and Seed Production	Laware and raskar (2014)
Arachis hypogaea (Peanut)	Metal oxide NPs, FeO, CuO, and TiO NPs	physiological response and reduce nutritional quality	Rui et al. (2018)
Stevia rebaudiana (Sweetleaf)	Cu and Au NPs	Secondary metabolite	Ghazal et al. (2018)
Momordica charantia (Bitter gourd)	AgNPs	secondary metabolites	Chung et al. (2018)
Caralluma tuberculate (Pamanghi)	AgNPs	biomass and secondary metabolites	Ali et al. (2019)
Prunella vulgaris (Common self-heal)	Ag and Au NPs	biomass and industrially important secondary metabolites	Fazal et al. (2019)
<i>Capsicum</i> sp. (Chillies or peppers)	AgNPs	capsaicin	Bhat and Bhat (2016)
Triticum aestivum (Common wheat) stress-tolerant Parabola and stress-sensitive Raweta,	Ag NPs and Ag salt	Morphological characteristics, lipid peroxidation and mobilization of defense system	Barbasz et al. (2016)
<i>Gymnema sylvestre</i> (Australian cowplant)	CuO NPs	GA II, total phenolics, and flavo- noids	Chung et al. (2019)
Citrullus lanatu (Watermelon)	Turmeric oil nanoemulsions and Ag NPs	Improves Germination, Growth, Yield, and Quality	Acharya et al. (2020)
<i>Capsicum annuum</i> (Chillies or peppers)	AgNPs	Cytokinin response	Vinković et al. (2017)
Zea mays (Corn)	TiO ₂ NPs	Anthocyanins, Carotenoids, Chlorophyll	Morteza et al. (2013)
Withania somnifera (Ashwagandha)	CuO NPs	polyphenols content and antioxi- dant activity	Singh et al. (2018)
Achillea millefolium (Common yarrow)	Agand Methyl Jasmonate NPs	lipid peroxidation, flavonoid con- tent, and yield of essential oils	Ghanati et al. (2014)



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Table 2 (continued)

Plant	Nanoparticles	Effect of Nanoparticles	Reference
Stevia rebaudiana Bertoni (Honey Leaf)	ZnO and CuO NPs	TPC, TAC, TRP and DPPH free radical scavenging activity	Javed et al. (2017)
Corylus avellana (Common hazel)	AgNPs	phenolics, and flavonoids, antioxi- dants activity, Taxanes content	Jamshidi and Ghanati (2016)
Mentha piperita (Peppermint)	CuSO ₄ and Cu NPs	Morpho-physiological traits and essential oil	Lafmejani et al. (2018)
Punica granatum (Pomegranate)	MgO and CuO NPs	tannins and phenols	Al-oubaidi and Al-khafagi (2018)
Arabidopsis thaliana (Mouseear cress)	FeNPs	Root Elongation, Antioxidant activity	Kim et al. (2014)
Arabidopsis thaliana (Mouseear cress)	Al ₂ O ₃ , SiO2, Magnetite and ZnO NPs	Phytotoxicity (seed germination, root elongation, and number of leaves)	Lee et al. (2010)
Boswellia ovalifoliolata (Salai guggul)	AgNPs	Seed germination and Seedling growth	Savithramma et al. (2012)
<i>Gymnema sylvestre</i> (Australian cowplant)	CuONPs	Gymnemic Acid and Phenolic Compounds	Chung and Thiruvengadam (2019)
Salvia miltiorrhiza (Red sage)	Ag ion,	Tanshinones than Phenolic Acids	Xing et al. (2014)
<i>Nigella arvensis</i> (Wild Fennel)	Al and NiONPs	growth and antioxidant defense	Chahardoli et al. (2020)
Triticum aestivum (Wheat)	TiO ₂ NPs	Antioxidant activity, leaves and root	Silva et al. (2019)
Cucumis sativus (Garden cucumber)	Cu(OH)2 nanopesticide	Antioxidant and detoxification gene expression	Zhao et al. (2017)
Cucumis sativus (Garden cucumber)	Ag ions and AgNPs	Antioxidant defense systems, respiration, inhibited photorespi- ration, altered membrane properties and reduced inorganic nitrogen fixation (down- regulation of glutamine and asparagine)	Zhang et al. (2018)
Capsicum chinense (Habanero peppers)	ZnSO ₄ ion and ZnO NPs	physicochemical quality; phenolic compounds; antioxidant capacity	García-López et al. (2019)
Cucumis sativus (Garden cucumber)	CuNPs	Oxidative stress, genotoxicty, and changes in superoxide dismutase (SOD) Gene expression	Mosa et al. (2018)
<i>Capsicum annuum</i> (Chillies or peppers)	SeNPs	Growth, morphology, anatomy, biochemistry, gene expression, and epigenetic DNA methylation	Sotoodehnia-Korani et al. (2020)
Portulaca oleracea (Duckweed)	AgNPs	Physiological and Molecular	Zare et al. (2019)
Stevia rebaudiana (Sweetleaf)	AgNPs	Glycosides Biosynthetic Pathway	Ramezani et al. (2019)
Capsicum annuum (Chillies or peppers)	Nano-chitosan	growth, morphogenesis, physiol- ogy, and micropropagation	Asgari-Targhi et al. (2018)
<i>Brassica rapa</i> sp. <i>rap</i> a (Napa cabbage)	AgNPs	Physiological, metabolic, and transcriptional effects	Thiruvengadam et al. (2014)



Plant	Type of NPs	Mode	Field relevance	References
Trigonella foenum-graecum (fenugreek)	AgNPs	Foliar application 40 mg/l	AgNPs increased yield compo- nents, growth and biochemical parameters	Sadak, 2019
Zea mays (maize)	Zn encapsulated chitosan NPs	Seed priming and foliar applica- tion (0.01–0.16%)	Increased grain yield from 20.5 to 39.8% and also enriched the grain with zinc micronutrient from 41.27 to 62.21 µg/g dw	Choudhary et al., 2019
Brassica juncea (cv. pusa jaikisan)	AuNPs	Foliar spray	Optimal increase in seed yield was recorded at 10 ppm with 19% increase in seed yield per plant	Arora et al., 2012
Avena sativa (Oat) and Trifolium alexandrinum (Ber- seem)	ZnO, TiO ₂ , CuO and Ag NPs	Seed priming	All the four NPs at lower dose (750 mg) significantly enhanced germination percent- age, seedling vigour and yield traits as compared to control in both the crops	Maity et al. 2018
Zea mays (Maize)	CuNPs	Seed priming	Positive effect on plant produc- tivity with increased total seed number and grain yield under drought stress conditions	Nguyen et al., 2020
Setaria italica (Foxtail Millet)	ZnO NPs	Foliar Spray	Concentrations of 2.6 mgL ⁻¹ ZnO NPs with twice foliar spary in field trial showed the maximum significant changes in millet grain oil and the total nitrogen content nutritional factors	Kolenčík et al. 2019

 Table 3
 Relevance of nanoparticles on crop production under field conditions

Prerequisites of nanotechnology in food industries

Food processing

Food processing is the conversion of raw elements into foods and other forms that are marketable and have a long shelf life. Processing includes pathogen prevention, toxin removal, and improving and preserving food evenness for healthier marketing and distribution. Processed foods are typically less liable to early spoilage than fresh foods and are restored to a state suitable for long-distance transport from the source to the consumer. All of these assets have been enhanced by the current convergence of nanotechnology. Nanocapsule conveyance systems play an essential role in the processing region, and the functional assets are preserved by encapsulating simple solutions like emulsions, colloids, biopolymers, and others into food products (Chellaram et al. 2014; Paredes et al. 2016; Amit et al. 2017). Existing nano-foods include nanoscale ingredients and additives in emerging nanoemulsion-based ice cream with a lower fat content but a fatty consistency and flavour. NPs have improved colour and stability during processing, improved flow properties, and increased shelf life of food products. For example, aluminosilicate is used in granular or powdered processed foods

Table 4 Common ingredients used in food nanoformulations

Components	Examples	References
Functional ingredients	Carotenoids; vitamin E; omega-3, -6, and -9 fatty acids; phytosterols; curcumin; coenzyme Q	Laouini et al. (2012), Qian et al. (2012), Saberi et al. (2013)
Emulsifiers	Sodium dodecyl sulphate; Tween® 20–80; Span® 20–80; phospholipids; casein; whey protein; soy protein	Lam and Nickerson (2013), Adjonu et al. (2014), Guttoff et al. (2015)
Oils	sunflower, soybean, fish, mineral, basil, olive, corn, cin- namon, coconut, castor, and essential oils	Donsì et al. (2011), Ghosh et al. (2013), Lane et al. (2014)
Ripening retardants	Long-chain triglyceride; mineral oil; ester gum	Joe et al. (2012), Ghosh et al. (2014)



as anticaking agents, whereas anatase TiO₂ is a common food whitener and brightener additive found in confectionery, a few sauces, and cheeses (Alfadul and Elneshwy 2010; Masood et al. 2019). Nano-encapsulated bioactives include vitamins, isoflavones, co-enzymes, carotenoids, flavonoids, food colouring substances, preserving agents, and a few other bioactive ingredients. These products can be found in a variety of food additives and beverages, as well as in vitamin E nano-resolution, which is specifically designed for pure beverages such as athletic beverages, flavourings, and enriched waters. The efficiency of these elements depends on conserving and increasing their bioavailability. Nano-encapsulating or nano-sizing active constituents deliver better bioavailability, enhanced solubility and amplified effectiveness than those materials in higher or micro-encapsulated form (Table 4). The higher the force of the NPs additives, the lower the concentration of the prerequisite additives and their chemical reactivity (Mozafari et al. 2006; Coelho et al. 2007; dos Santos et al. 2018). Nanotechnology can be used to manipulate food products for the effective delivery of protein and antioxidants. The encapsulated system developed results in nanospheres or microspheres (Singh et al. 2017; Lengyel et al. 2019). Nanosized self-assembled essential lipids serve as a liquid transferor of healthy components that are insoluble in water and fats, called nanodrops. They are used to constrain cholesterol transportation from the digestive system into the bloodstream (de Azeredo 2009; Arora and Padua 2010; Rhim et al. 2013).

Food safety

Nanomaterials are the essential components of many chemical and biological sensors, which have been explored to improve existing devices' performance or bring a new perspective to food safety. Nanomaterials' functions are implemented for two purposes: to improve the response characteristics of the transducers and as immobilization matrices for the receptors. Nanomaterials used in electrochemical biosensors for food safety mainly include carbon-based nanomaterials, metal and metal oxide NPS, magnetic NPs, and molecularly imprinted polymers (Zeng et al. 2016; Mustafa and Andreescu 2020). Nanotechnology can detect minute amounts of a chemical pollutant, bacteria, or virus in foodstuffs. The combined biosensors of the biological molecules and nanomaterials significantly reduced response time to a prospective problem. It will result in extra care for the food handling scheme. Nano-sensors established by researchers at Purdue and Clemson universities use NPs, which can either be appropriate to colours, flavours, or industrial magnetic resources. These NPs can selectively suppress any food pathogen. These sensors also detect either magnetic materials or infrared light and can signal the occurrence of even a tiny hint of dangerous pathogens. The benefit of such a scheme is that, hypothetically, thousands of NPs can be positioned on a single nanosensor to swiftly and precisely sense any number of diverse bacteria and pathogens. Likewise, these nano-sensors will be able to advance access to the minute clefts where the pathogens habitually hide. The impact of nanotechnologies on the detection of pathogenic microorganisms in food and the development of nanofood safety are also being investigated at Cornell University's biosensors research laboratory and bioanalytical microsystems. These studies focused on developing fast and portable biosensors for detecting pathogenic microorganisms in the environment and food. The scheme places emphasis on the precise and rapid recognition of pathogens in regular drinking water analysis, food examination, environmental analysis, and clinical diagnostics (Shefer and Shefer 2003a, b; 2005). Because of their antagonistic effects on human health and influence on environmental systems, AgNPs could directly improve the world's food supply without being used in food. AgNPs work as antimicrobial agents in foods to grow food-related claims such as microbe-resistant materials or non-biofouling shells. Ag ions are precisely sensitive and may react with the negatively charged cell membrane, providing a supplementary involvement to AgNPs' bactericidal effect (Ghorbanpour et al. 2020; Zorraquín-Peña et al. 2020; Hutapea et al. 2022). The benefits of Ag antimicrobial agents are that they can be easily incorporated into various constituents, such as textiles and plastics. AgNPs are valuable in a broad range of applications, as they preserve their antimicrobial action in situ, whereas traditional antimicrobial agents would be unstable (Franci et al. 2015). According to (Carbone et al. 2016), AgNPs may be assimilated into non-degradable (vinyl alcohol, polyvinyl chloride, and polyethylene) and recyclable (starch, cellulose, agarose, and chitosan) polymers to produce food packages.

The AgNPs may also provide clues to novel methods for killing foodborne pathogens and augmenting food security (Flood 2010). They also illustrated that NPs can provide us with multi-component antimicrobial fusions that would be substantially compatible with other compounds to kill microorganisms and improve food safety (Zorraquín-Peña et al. 2020). Nanotechnology might also be pragmatic in the analysis of food to notice reasonably low-slung quantities of toxins formed by microorganisms. E.g., Staphylococcus aureus can synthesize the thermostable enterotoxins A, B, C, D, and E. Around 100-200 ng of these enterotoxins is enough to cause a lethal infection. Hence, this bacterium in foodstuff might turn into a hazard reaction in health system, if it preserved at a temperature that favours its growth. Nanotechnology has developed a method for detecting S. aureus that employs magnetic beads to detect the maximum number of strains of S. aureus at a low and viable cost. It will lead to improved food safety. Analogous effects were also found for Listeria monocytogenes. Aflatoxin are



teratogenic, carcinogenic, mutagenic, and immunosuppressive materials produced as secondary metabolites on a variety of food products by the fungi *Aspergillus flavus* and *Aspergillus parasiticus*. The highest tolerance level in mice is 2 ng AFB1/g. For AFB1 detection, a competitive ELISA test using a 96-well microplate for immunosensors has been industrialized. This method pools the great selectivity of immune study with the comfort of electrochemical probes and multi-sample analysis (Argudín et al. 2010; Hernández-Cortez et al. 2017; Wang et al. 2017). These applications of nanotechnology in food examination would advance food quality and safety and regulate the vulnerabilities that might disturb human health.

Food packaging

Nanomaterials useful in food packaging are categorized into two classes: inorganic and organic materials. For the prior materials, metals, metal oxides, and clay NPs merged into bionanocomposite films and nanofibers can be considered (Huang et al. 2018; Chaudhary et al. 2020; Jafarzadeh et al. 2020). Besides conventional bacteriostatic AgNPs, some inorganic agents, like oxidized NPs, including ZnO, CuO, MgO, TiO₂, and Fe₃O₄, have attracted considerable attention due to their conflict with rough processing conditions and improvements in potent inhibition against foodborne pathogens. As far, the resources like several clays, they may possibly bid resistance to water vapor and gases and recover biopolymers' mechanical strength (Attaran et al. 2017). The second group is organic materials including, halogenated compounds, phenols, plastic polymers, quaternary ammonium salts, plus natural polysaccharide or protein materials such as chitin, chitosan, zein, and whey protein isolates, which have been currently greatly assessed (Perinelli et al. 2018).

Recent nanotechnology applications in food science have ensured a significant influence on the food industry with the molecular combination of novel food products or fixings that increase the security and shelf-life of the food. The growth of nanostructured materials comprises physical, chemical, and biological properties that are significantly different from those of their unpackaged parallels in food structures. Nanotechnology has impending applications in the food zone, including food packaging and safety, leading to enhanced eminence and conservation of foodstuffs. Some foods, like milk and milk derivatives, contain nanoscale constituents such as casein and other milk proteins (Pathakoti et al. 2017; Nile et al. 2020). The foremost tenders of nanostructured supplies in food are liposomes, nanocomposites, nanoemulsions, NPs, proteins, biopolymeric nanostructured films, and cellulose-based NPs. These materials enable precise proclamation, improve solubility, advance bioavailability, and



defend bioactive modules during production and packing (Pathakoti et al. 2017).

Nanotoxicity

The toxicity of nanomaterials and nanocomposites on plant systems was well illustrated (Ibrahim et al. 2019). Studies in this regard prove that it has full applications in agriculture. For example, the effect of nanohexaconazole on soil microbes behaves similarly to the commercial formulation hexaconazole (Contaf Plus) with no side effect on soil microbiota (Kumar et al. 2015). In the case of ZnO NPs with thiram, it has antifungal properties and degrades the extra thiram after the antifungal process (Xue et al. 2014). CeO₂ NPs and ZnO NPs' effects on the life cycle of corn (Zea mays) plants were found to be that CeO₂ NPs altered the position of calcium in kernels, and ZnO NPs reduced photosynthesis by 12%, stomatal conductivity by 15%, and chlorophyll content by 10%. The yield was reduced by 38 and 49% in the cases of CeO2 and ZnO, respectively (Zhao et al. 2015).

Nanomaterials' mobility, bioavailability, and toxicity have required attention for a better understanding of risk factors. The use of nanomaterials and their exposure may be a hazard, which still needs to be known in-depth. NPs and their materials will most likely be released into the environment from construction services, landfills, wastewater treatment plants, or nonpoint sources such as clothing made from NP-containing materials. Unintentional discharge of NPs during creation or transportation is also possible. There are various NPs that directly get injected into groundwater, leading to pollution, especially with chlorinated solvents. The different ways to dump the NPs, whether in soil/ water, or the atmosphere, all result in the interaction of NPs with plants, either directly or indirectly, via sewage treatment plants (Nowack and Bucheli 2007). The AgNPs affect the growth, physiology, and morphology of the aquatic plant duckweed (Spirodela polyrhiza). The harmfulness of AgNPs and AgNO₃ indicated that Ag content in plant tissue increased considerably with higher concentrations of AgNO₃ and AgNPs. AgNPs and AgNO₃ expressively reduced the biomass of the plant, produced colonies of S. polyrhiza to crumble, and also resulted in root abscission. The physiological study revealed that AgNPs and AgNO₃ pointedly reduced nitrate-nitrogen content in plant tissue, chlorophyll a/b (Chl a/b) content, and chlorophyll fluorescence (Fv/Fm). Variations in soluble carbohydrate and proline content were also identified after AgNO3 and AgNPs treatment (Jiang et al. 2017). These acts of NP exposure will have a harmful effect on the environment and human health. There are some studies regarding the application of NPs to plants. There is a crucial point in the view that metal NPs cause phytotoxicity

via the closure and discharge of greater concentrations of essential ions (Ma and Mumper 2013; Anjum et al. 2015).

Overlying challenges and future perspectives

Nature has both disease-inducing and disease-preventive incentives. Science is the link between nature and disease, examining biological problems in nature and developing solutions to them. Any particle has the power to bring about changes in the environment. When we go with the disease protection concept, we always look forward to an agent or an elicitor with significant activity and minor concentration. In this context, nanoformulation is one of the most significant approaches. Nanotechnology includes diverse fields of science involving the manipulation of materials at the atomic and molecular level. Agricultural nanotechnology was developed in the 1990s, and it is now functionalized globally in various sectors of farming. Agro-nanotechnology enables sustainable food production by developing healthy potential plant systems that can improve plant germination, growth, yield, and quality. The scientific venture of nanotechnology provides us considerable benefits in all areas, and it is also essential to move toward a responsibility to make wise use of nanotechnology. The vision of nanotechnology was initiated in 1959 by Richard Feynman in his visionary talk "There is Plenty of Room at the Bottom," wherein he illustrated the possible manipulation of atoms. Presently, a branch of that vision has been recognized, but it remains indistinct as to the possible extent of manipulations at the atomic level. The concept of nanotechnology in the farming sector helps develop good health in society's economic status by transforming a pathogenic form into a non-pathogenic, environmentally friendly, and sustainable form. Also, minimize the use of hazardous chemicals for short-term benefits. Nanotechnology has created an era by grabbing significant funds globally. There is no doubt, that it will further channel and revolutionize the mainstream of the science vision to overcome all the constraints in agriculture by improving the green economy. Further, research bodies have to engage in the utilization of naturally available solutions to improve the plants' growth and health potential. Besides, it is also essential to minimize the use of hazardous chemicals for short-term benefits and step forward to improve the ecosystem's strength in bringing out long-term benefits to agricultural commodities.

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References

- Abd-Alla MH, Nafady NA, Khalaf DM (2016) Assessment of silver nanoparticles contamination on faba bean-*Rhizobium leguminosarum* bv. viciae-*Glomus aggregatum* symbiosis: Implications for induction of autophagy process in root nodule. Agric Ecosyst Environ 218:163–177. https://doi.org/10.1016/j.agee.2015. 11.022
- Abobatta WF (2018) Nanotechnology application in agriculture. Acta Sci Agric 2. https://doi.org/10.1007/BF01568729
- Acharya P, Jayaprakasha GK, Crosby KM, Jifon JL, Patil BS (2020) Nanoparticle-mediated seed priming improves germination, growth, yield, and quality of watermelons (*Citrullus lanatus*) at multi-locations in Texas. Sci Rep 10:1–16. https://doi.org/10. 1038/s41598-020-61696-7
- Adu-Gyamfi R, Agyin-Birikorang S, Tindjina I, Manu Y, Singh U (2019) Minimizing nutrient leaching from maize production systems in northern ghana with one-time application of multinutrient fertilizer briquettes. Sci Total Environ 694:133667 https://doi.org/10.1016/j.scitotenv.2019.133667
- Ahmad P, Alyemeni MN, Al-Huqail AA, Alqahtani MA, Wijaya L, Ashraf M, Kaya C, Bajguz A (2020) Zinc oxide nanoparticles application alleviates arsenic (As) toxicity in soybean plants by restricting the uptake of as and modulating key biochemical attributes, antioxidant enzymes, ascorbate-glutathione cycle and glyoxalase system. Plants 9:1–18. https://doi.org/10.3390/plant s9070825
- Ahmad B, Shabbir A, Jaleel H, Khan MMA, Sadiq Y (2018) Efficacy of titanium dioxide nanoparticles in modulating photosynthesis, peltate glandular trichomes and essential oil production and quality in *Mentha piperita* L. Curr Plant Biol 13:6–15. https://doi.org/ 10.1016/j.cpb.2018.04.002
- Alfadul SM, Elneshwy AA (2010) Use of nanotechnology in food processing, packaging and safety—review. Afr J Food Agri Nut Develop 10:2719–2739. https://doi.org/10.4314/ajfand.v10i6. 58068
- Aliofkhazraei M (2015) Handbook of Nanoparticles. Springer Reference, Iran. https://doi.org/10.1007/978-3-319-15338-4
- Amit SK, Uddin MM, Rahman R, Islam SMR, Khan MS (2017) A review on mechanisms and commercial aspects of food preservation and processing. Agric Food Secur 6:1–22. https://doi.org/10. 1186/s40066-017-0130-8
- Anjum NA, Adam V, Kizek R, Duarte AC, Pereira E, Iqbal M, Lukatkin AS, Ahmad I (2015) Nanoscale copper in the soil-plant



system—toxicity and underlying potential mechanisms. Environ Res 138:306–325. https://doi.org/10.1016/j.envres.2015.02.019

- Argudín MÁ, Mendoza MC, Rodicio MR (2010) Food poisoning and Staphylococcus aureus enterotoxins. Toxins (basel) 2:1751– 1773. https://doi.org/10.3390/toxins2071751
- Aslani F, Bagheri S, Muhd Julkapli N, Juraimi AS, Hashemi FSG, Baghdadi A (2014) Effects of engineered nanomaterials on plants growth: An overview. Sci World J 2014:641759. https://doi.org/ 10.1155/2014/641759
- Athanassiou CG, Kavallieratos NG, Benelli G, Losic D, Usha Rani P, Desneux N (2018) Nanoparticles for pest control: current status and future perspectives. J Pest Sci 91:1–15. https://doi.org/10. 1007/s10340-017-0898-0
- Attaran SA, Hassan A, Wahit MU (2017) Materials for food packaging applications based on bio-based polymer nanocomposites. J Thermoplast Compos Mater 30:143–173. https://doi.org/10. 1177/0892705715588801
- Awasthi A, Bansal S, Jangir LK, Awasthi G, Awasthi KK, Awasthi K (2017) Effect of ZnO Nanoparticles on germination of *Triticum* aestivum seeds. Macromol Symp 376:1–5. https://doi.org/10. 1002/masy.201700043
- de Azeredo HMC (2009) Nanocomposites for food packaging applications. Food Res Int 42:1240–1253. https://doi.org/10.1016/j. foodres.2009.03.019
- Bajpai VK, Kamle M, Shukla S, Mahato DK, Chandra P, Hwang SK, Kumar P, Huh YS, Han YK (2018) Prospects of using nanotechnology for food preservation, safety, and security. J Food Drug Anal 26:1201–1214. https://doi.org/10.1016/j.jfda. 2018.06.011
- Bao-shan LIN, Chun-hui LI, Li-jun F, Shu-chun Q, Min YU (2004) Effect of TMS (nanostructured silicon dioxide) on growth of *Changbai larch* seedlings. J for Res 15:138–140. https://doi.org/ 10.1007/bf02856749
- Berg M, Meehan M, Scherer T (2017) Environmental implications of excess fertilizer and manure on water quality. Environ Implic Excess Fertil Manure Water Qual — Publ. 1281: 1.
- Berners-Lee M, Kennelly C, Watson R, Hewitt CN (2018). Current global food production is sufficient to meet human nutritional needs in 2050 provided there is radical societal adaptation. Elementa 6. https://doi.org/10.1525/elementa.310
- Boutchuen A, Zimmerman D, Aich N, Masud AM, Arabshahi A, Palchoudhury S (2019) Increased plant growth with hematite nanoparticle fertilizer drop and determining nanoparticle uptake in plants using multimodal approach. J Nanomater 2019:7–9. https://doi.org/10.1155/2019/6890572
- Brock DA, Douglas TE, Queller DC, Strassmann JE (2011) Primitive agriculture in a social amoeba. Nature 469:393–396. https://doi. org/10.1038/nature09668
- Burman U, Saini M, Kumar P (2013) Effect of zinc oxide nanoparticles on growth and antioxidant system of chickpea seedlings. Toxicol Environ Chem 95:605–612. https://doi.org/10.1080/02772248. 2013.803796
- Carbone M, Donia DT, Sabbatella G, Antiochia R (2016) Silver nanoparticles in polymeric matrices for fresh food packaging. J King Saud Univ - Sci 28:273–279. https://doi.org/10.1016/j.jksus. 2016.05.004
- Carvalho FP (2017) Pesticides, environment, and food safety. Food Energy Secur 6:48–60. https://doi.org/10.1002/fes3.108
- Cassman KG (1999) Ecological intensification of cereal production systems: Yield potential, soil quality, and precision agriculture. Proc Natl Acad Sci U. S A 96:5952–5959. https://doi.org/10. 1073/pnas.96.11.5952
- Chahardoli A, Karimi N, Ma X, Qalekhani F (2020) Effects of engineered aluminum and nickel oxide nanoparticles on the growth and antioxidant defense systems of *Nigella arvensis* L. Sci Rep 10:1–11. https://doi.org/10.1038/s41598-020-60841-6



- Chaudhary P, Fatima F, Kumar A (2020) Relevance of nanomaterials in food packaging and its advanced future prospects. J Inorg Organomet Polym Mater 30:5180–5192. https://doi.org/10.1007/ s10904-020-01674-8
- Chellaram C, Murugaboopathi G, John AA, Sivakumar R, Ganesan S, Krithika S, Priya G (2014) Significance of nanotechnology in food Industry. APCBEE Proc 8:109–113. https://doi.org/10. 1016/j.apcbee.2014.03.010
- Chhipa H (2017) Nanofertilizers and nanopesticides for agriculture. Environ Chem Lett 15:15–22. https://doi.org/10.1007/ s10311-016-0600-4
- Chichiriccò G, Poma A (2015) Penetration and toxicity of nanomaterials in higher plants. Nanomaterials 5:851–873. https://doi.org/ 10.3390/nano5020851
- Coelho CMM, de Bellato C, Santos JCP, Ortega EMM, Tsai SM, (2007) Effect of phytate and storage conditions on the development of the 'hard-to-cook.' J Sci Food Agric 1243:1237–1243. https://doi.org/10.1002/jsfa
- Cordell D, White S (2013) Sustainable phosphorus measures: strategies and technologies for achieving phosphorus security. Agronomy 3:86–116. https://doi.org/10.3390/agronomy3010086
- Corradini E, de Moura MR, Mattoso LHC (2010) A preliminary study of the incorparation of NPK fertilizer into chitosan nanoparticles. Express Polym Lett 4:509–515. https://doi.org/10.3144/expre sspolymlett.2010.64
- Côa F, Bortolozzo LS, Petry R, Silva GH, Da Martins CHZ, Medeiros AMZ, de Sabino CMS, Costa RS, Khan LU, Delite FS, Martinez DST (2019) Nanopesticides from research and development to mechanisms of action and sustainable use in agriculture, in: Fraceto LF, Lima R, Ávila D, Oliveira HC, Castro VLSS, de Grillo R (Eds.), Nanopesticides. Springer, Brazil, pp. 227–280 10.1007%2F978-3-030-44873-8
- Deng YQ, White JC, Xing BS (2014) Interactions between engineered nanomaterials and agricultural crops: implications for food safety. J Zhejiang Univ Sci A 15:552–572. https://doi.org/10. 1631/jzus.A1400165
- Dhoke SK, Mahajan P, Khanna AS (2011) Effect of nano-ZnO particle suspension on growth of mung (Vigna radiata) and gram (*Cicer* arietinum) seedlings using plant agar method. J Nanotechnol 2011. https://doi.org/10.1155/2011/696535
- Dimkpa CO, Fugice J, Singh U, Lewis TD (2020) Development of fertilizers for enhanced nitrogen use efficiency—Trends and perspectives. Sci Total Environ 731: 139113. https://doi.org/10. 1016/j.scitotenv.2020.139113
- Duhan JS, Kumar R, Kumar N, Kaur P, Nehra K, Duhan S (2017) Nanotechnology: the new perspective in precision agriculture. Biotechnology Reports 15:11–23. https://doi.org/10.1016/j.btre. 2017.03.002
- El-shetehy M, Moradi A, Maceroni M, Reinhardt D, Petri-fink A, Rothen-rutishauser B Mauch, F, Schwab F (2020). Silica nanoparticles enhance disease resistance in Arabidopsis plants. Nat Nanotechnol https://doi.org/10.1038/s41565-020-00812-0
- Eliazer Nelson ARL, Ravichandran K, Antony U (2019) The impact of the green revolution on indigenous crops of India. J Ethn Foods 6:1–10. https://doi.org/10.1186/s42779-019-0011-9
- Elizabath A, Bahadur V, Misra P, Mashi Prasad V, Thomas T, Ambily Elizabath C (2017) Effect of different concentrations of iron oxide and zinc oxide nanoparticles on growth and yield of carrot (*Daucus carota* L.). J Pharmacogn Phytochem JPP 6:1266–1269
- Emadian SM, Onay TT, Demirel B (2017) Biodegradation of bioplastics in natural environments. Waste Manag 59:526–536. https:// doi.org/10.1016/j.wasman.2016.10.006
- Fagodiya RK, Pathak H, Kumar A, Bhatia A, Jain N (2017) Global temperature change potential of nitrogen use in agriculture: A

50-year assessment. Sci Rep 7:1–8. https://doi.org/10.1038/srep4 4928

- Fakruddin M, Hossain Z, Afroz H (2012) Prospects and applications of nanobiotechnology: a medical perspective. J Nanobiotechnology 10:1–8. https://doi.org/10.1186/1477-3155-10-31
- Felix Alvarez R de C Prado R de M, Felisberto G, Fernandes Deus AC, Lima De Oliveira RL (2018) Effects of soluble silicate and nanosilica application on rice nutrition in an oxisol. Pedosphere 28:597–606. https://doi.org/10.1016/S1002-0160(18)60035-9
- Flood J (2010) The importance of plant health to food security. Food Secur 2:215–231. https://doi.org/10.1007/s12571-010-0072-5
- Frenk S, Ben-Moshe T, Dror I, Berkowitz B, Minz D (2013) Effect of metal oxide nanoparticles on microbial community structure and function in two different soil types. PLoS ONE 8:1–12. https:// doi.org/10.1371/journal.pone.0084441
- Gao F, Hong F, Liu C, Zheng L, Su M, Wu X, Yang F, Wu C, Yang P (2006) Mechanism of nano-anatase TiO₂ on promoting photosynthetic carbon reaction of spinach: inducing complex of Rubisco-Rubisco activase. Biol Trace Elem Res 111:239–253. https://doi.org/10.1385/BTER:111:1:239
- Garciá-López JI, Zavala-Garcia F, Olivares-Saénz E, Lira-Saldivar RH, Barriga-Castro ED, Ruiz-Torres NA, Ramos-Cortez E, Vázquez-Alvarado R, Ninõ-Medina G (2018) Zinc Oxide nanoparticles boosts phenolic compounds and antioxidant activity of *Capsicum annuum* L. during germination. Agronomy 8. https://doi.org/10. 3390/agronomy8100215
- Gardea-Torresdey JL, Rico CM, White JC (2014) Trophic transfer, transformation, and impact of engineered nanomaterials in terrestrial environments. Environ Sci Technol 48:2526–2540. https:// doi.org/10.1021/es4050665
- Ghasemzadeh A (2012) Global Issues of Food Production Agrotechnology 01:1–2. https://doi.org/10.4172/2168-9881.1000e102
- Ghidan AY, Antary TM Al (2019) Applications of nanotechnology in agriculture, methods in microbiology. Intechopen, Jordan. https:// doi.org/10.5772/intechopen.88390
- Ghorbanpour M, Bhargava P, Varma A, Choudhary DK (2020). Biogenic Nano-Particles and Their Use in Agro-Ecosystems. https:// doi.org/10.1007/978-981-15-2985-6
- Gil-Díaz M, González A, Alonso J, Lobo MC (2016) Evaluation of the stability of a nanoremediation strategy using barley plants. J Environ Manage 165:150–158. https://doi.org/10.1016/j.jenvm an.2015.09.032
- Giroto AS, Guimarães GGF, Foschin M, Ribeiro C (2017) Role of slow-release nanocomposite fertilizers on nitrogen and phosphate availability in soil. Sci Rep 7. https://doi.org/10.1038/srep46032
- Gohari G, Mohammadi A, Akbari A, Panahirad S, Dadpour MR, Fotopoulos V, Kimura S, (2020) Titanium dioxide nanoparticles (TiO₂ NPs) promote growth and ameliorate salinity stress effects on essential oil profile and biochemical attributes of *Dracocephalum moldavica*. Sci Rep 10:1–14. https://doi.org/10.1038/ s41598-020-57794-1
- Govorov AO, Carmeli I (2007) Hybrid structures composed of photosynthetic system and metal nanoparticles: Plasmon enhancement effect. Nano Lett 7:620–625. https://doi.org/10.1021/n1062528t
- Guo H, White JC, Wang Z, Xing B (2018) Nano-enabled fertilizers to control the release and use efficiency of nutrients. Curr Opin Environ Sci Heal 6:77–83. https://doi.org/10.1016/j.coesh.2018. 07.009
- Haghighi M, Teixeira Da Silva JA (2014) The effect of N-TiO₂ on tomato, onion, and radish seed germination. J Crop Sci Biotechnol 17:221–227. https://doi.org/10.1007/s12892-014-0056-7
- Handy RD, Von Der Kammer F, Lead JR, Hassellöv M, Owen R, Crane M (2008) The ecotoxicology and chemistry of manufactured

nanoparticles. Ecotoxicology 17:287–314. https://doi.org/10. 1007/s10646-008-0199-8

- Hasaneen MNA, Abdel-Aziz HMM, El-Bialy DMA, Omer AM (2014) Preparation of chitosan nanoparticles for loading with NPK fertilizer. Afr J Biotech 13:3158–3164. https://doi.org/10.5897/ajb20 14.13699
- Hayles J, Johnson L, Worthley C, Losic D (2017) Nanopesticides: a review of current research and perspectives. Elsevier Inc., New Pesticides and Soil Sensors. https://doi.org/10.1016/b978-0-12-804299-1.00006-0
- Hernández-Cortez C, Palma-Martínez I, Gonzalez-Avila LU, Guerrero-Mandujano A, Solís RC, Castro-Escarpulli G (2017) Food poisoning caused by bacteria (Food Toxins). poisoning - From Specif. Toxic Agents to Nov. Rapid Simpl. Tech. Anal. https:// doi.org/10.5772/intechopen.69953
- Hong F, Zhou J, Liu C, Yang F, Wu C, Zheng L, Yang P (2005) Effect of Nano-TiO₂ on photochemical reaction of chloroplasts of spinach. Biol Trace Elem Res 105:269–279. https://doi.org/10.1385/ BTER:105:1-3:269
- Huang S, Wang L, Liu L, Hou Y, Li L (2015) Nanotechnology in agriculture, livestock, and aquaculture in China. A Review, Agronomy for Sustainable Development. https://doi.org/10.1007/ s13593-014-0274-x
- Hund-Rinke K, Simon M (2006) Ecotoxic effect of photocatalytic active nanoparticles (TiO₂) on algae and daphnids. Environ Sci Pollut Res 13:225–232. https://doi.org/10.1065/espr2006.06.311
- Ibrahim K, Saeed K, Idrees K (2019) Nanoparticles: Properties, applications and toxicities. Arab J Chem 12:908–931. https://doi.org/ 10.1016/j.arabjc.2017.05.011
- Jafarzadeh S, Salehabadi A, Jafari SM (2020) Metal nanoparticles as antimicrobial agents in food packaging, Handbook of Food Nanotechnology. INC. https://doi.org/10.1016/b978-0-12-815866-1.00010-8
- Jiang HS, Yin LY, Ren NN, Zhao ST, Li Z, Zhi Y, Shao H, Li W, Gontero B (2017) Silver nanoparticles induced reactive oxygen species via photosynthetic energy transport imbalance in an aquatic plant. Nanotoxicology 11:157–167. https://doi.org/10. 1080/17435390.2017.1278802
- Joshi R, Bhati R, Kandpal J, Professor A (2019) Nutrient and pest management through nanotechnology. Int Res J Eng Technol 06:1887
- Ju-Nam Y, Lead JR (2008) Manufactured nanoparticles: an overview of their chemistry, interactions and potential environmental implications. Sci Total Environ 400:396–414. https://doi.org/10.1016/j. scitotenv.2008.06.042
- Kah M, Hofmann T (2014) Nanopesticide research: Current trends and future priorities. Environ Int 63:224–235. https://doi.org/10. 1016/j.envint.2013.11.015
- Kah M, Tufenkji N, White JC (2019) Nano-enabled strategies to enhance crop nutrition and protection. Nat Nanotechnol 14:532– 540. https://doi.org/10.1038/s41565-019-0439-5
- Kahru A, Dubourguier HC (2010) From ecotoxicology to nanoecotoxicology. Toxicology 269:105–119. https://doi.org/10.1016/j. tox.2009.08.016
- Kandasamy S, Sorna Prema R (2015) Methods of synthesis of nanoparticles and its applications. Available online www.jocpr.com J Chem Pharm Res 7: 278–285.
- Karimi J, Mohsenzadeh S (2015) Effects of silicon oxide nanoparticles on growth and physiology of wheat Seedlings. Физиология Растений 63:126–130. https://doi.org/10.7868/s001533031 6010103
- Khalifa NS, Hasaneen MN (2018) The effect of chitosan–PMAA–NPK nanofertilizer on *Pisum sativum* plants. 3 Biotech 8. https://doi. org/10.1007/s13205-018-1221-3



- Khatri K, Rathore M.S (2018) Plant nanobionics and its applications for developing plants with improved photosynthetic capacity, In: photosynthesis - from its evolution to future improvements in photosynthetic efficiency using nanomaterials. Intechopen pp 95–112 https://doi.org/10.5772/intechopen.76815
- Khot LR, Sankaran S, Maja JM, Ehsani R, Schuster EW (2012) Applications of nanomaterials in agricultural production and crop protection: a review. Crop Prot 35:64–70. https://doi.org/10.1016/j. cropro.2012.01.007
- Kirschbaum MUF (2011) Does enhanced photosynthesis enhance growth? Lessons learned from CO2 enrichment studies. Plant Physiol 155:117–124. https://doi.org/10.1104/pp.110.166819
- Klikocka H, Marks M (2018) Sulphur and nitrogen fertilization as a potential means of agronomic biofortification to improve the content and uptake of microelements in spring wheat grain DM. J Chem 2018. https://doi.org/10.1155/2018/9326820
- Kolenčík M, Ernst D, Komár M, Urík M, Šebesta M, Dobročka E, Černý I, Illa R, Kanike R, Qian Y, Feng H, Orlová D, Kratošová G (2019) Effect of foliar spray application of zinc oxide nanoparticles on quantitative, nutritional, and physiological parameters of foxtail millet (*Setaria italica* L.) under field conditions. Nanomaterials 9. https://doi.org/10.3390/nano9111559
- Kottegoda N, Sandaruwan C, Priyadarshana G, Siriwardhana A, Rathnayake UA, Berugoda Arachchige DM, Kumarasinghe AR, Dahanayake D, Karunaratne V, Amaratunga GAJ (2017) Urea-Hydroxyapatite nanohybrids for slow release of nitrogen. ACS Nano 11:1214–1221. https://doi.org/10.1021/acsnano.6b07781
- Kumar R, Nair KK, Alam MI, Gogoi R, Singh PK, Srivastava C, Gopal M, Goswami A (2015) Development and quality control of nanohexaconazole as an effective fungicide and its biosafety studies on soil nitifiers. J Nanosci Nanotechnol 15:1350–1356. https://doi.org/10.1166/jnn.2015.9088
- Kumar LMP, M Indira (2017) Trends in fertilizer consumption and food grain production in India: A co-integration analysis. SDMIMD J Manag 8: 45–50. https://doi.org/10.18311/ sdmimd/2017/18025
- Lengyel M, Kállai-Szabó N, Antal V, Laki AJ, Antal I (2019) Microparticles, microspheres, and microcapsules for advanced drug delivery. Sci Pharm 87. https://doi.org/10.3390/scipharm87 030020
- Li W, Lan P (2017) The understanding of the plant iron deficiency responses in strategy I plants and the role of ethylene in this process by omic approaches. Front Plant Sci 8:1–15. https://doi. org/10.3389/fpls.2017.00040
- Li X, Yang Y, Gao B, Zhang M (2015) Stimulation of peanut seedling development and growth by zero-valent iron nanoparticles at low concentrations. PLoS ONE 10:1–12. https://doi.org/10. 1371/journal.pone.0122884
- Liang F, Lindberg P, Lindblad P (2018) Engineering photoautotrophic carbon fixation for enhanced growth and productivity. Sustain Energy Fuels 2:2583–2600. https://doi.org/10.1039/c8se00281a
- Liang F, Lindblad P (2017) Synechocystis PCC 6803 overexpressing RuBisCO grow faster with increased photosynthesis. Metab Eng Commun 4:29–36. https://doi.org/10.1016/j.meteno.2017.02.002
- Lin MT, Occhialini A, Andralojc PJ, Parry MAJ, Hanson MR (2014) A faster Rubisco with potential to increase photosynthesis in crops. Nature 513:547–550. https://doi.org/10.1038/nature13776
- Liu R, Lal R (2015) Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions. Sci Total Environ 514:131–139. https://doi.org/10.1016/j.scitotenv.2015.01.104
- Liu, Ruiqiang, Zhang H, Lal R (2016) Effects of Stabilized Nanoparticles of Copper, Zinc, Manganese, and iron oxides in low concentrations on Lettuce (*Lactuca sativa*) Seed Germination:

Nanotoxicants or Nanonutrients? Water Air Soil Pollut 227. https://doi.org/10.1007/s11270-015-2738-2

- Long BM, Hee WY, Sharwood RE, Rae BD, Kaines S, Lim YL, Nguyen ND, Massey B, Bala S, von Caemmerer S, Badger MR, Price GD (2018) Carboxysome encapsulation of the CO₂-fixing enzyme Rubisco in tobacco chloroplasts. Nat Commun 9. https:// doi.org/10.1038/s41467-018-06044-0
- Lyu S, Wei X, Chen J, Wang C, Wang X, Pan) D, (2017) Titanium as a beneficial element for crop production. Front Plant Sci 8:1–19. https://doi.org/10.3389/fpls.2017.00597
- Ma X, Geiser-Lee J, Deng Y, Kolmakov A (2010) Interactions between engineered nanoparticles (ENPs) and plants: Phytotoxicity, uptake and accumulation. Sci Total Environ 408:3053–3061. https://doi.org/10.1016/j.scitotenv.2010.03.031
- Ma H, Williams PL, Diamond SA (2013) Ecotoxicity of manufactured ZnO nanoparticles - A review. Environ Pollut 172:76–85. https:// doi.org/10.1016/j.envpol.2012.08.011
- Madany MMY, Saleh AM, Habeeb TH, Hozzein WN, AbdElgawad H (2020) Silicon dioxide nanoparticles alleviate the threats of broomrape infection in tomato by inducing cell wall fortification and modulating ROS homeostasis. Environ Sci Nano 7:1415–1430. https://doi.org/10.1039/c9en01255a
- Maity A, Natarajan N, Pastor M, Vijay D, Gupta CK, Wasnik VK (2018) Nanoparticles influence seed germination traits and seed pathogen infection rate in forage sorghum (*Sorghum bicolour*) and cowpea (*Vigna unguiculata*). Indian J Exp Biol 56:363–372
- Maksimovic M, Omanovic-Miklicanin E (2017) Towards green nanotechnology: maximizing benefits and minimizing harm Mirjana. IFMBE Proc 62:164–165. https://doi.org/10.1007/ 978-981-10-4166-2
- Manjunatha R, Naik D, Usharani K (2019) Nanotechnology application in agriculture: A review. J Pharmacogn Phytochem 8:1073–1083
- Manuel T (2019) The challenge of feeding the poor. Sustainability 11:5816. https://doi.org/10.3390/su11205816
- Masood F, Siddiqui Z, Ahmad S, Malik A (2019) Health and safety aspects of food processing technologies, Health and safety aspects of food processing technologies. Springer, Turkey. https://doi.org/10.1007/978-3-030-24903-8
- Menard A, Drobne D, Jemec A (2011) Ecotoxicity of nanosized TiO₂. Review of in vivo data. Environ Pollut 159:677–684. https://doi.org/10.1016/j.envpol.2010.11.027
- Meybeck A, Laval E, Levesque R, Parent G (2018) Food security and nutrition in the age of climate change, proceedings of the international symposium organized by the government of Québec in collaboration with FAO.
- Mikkelsen R (2018) Nanofertilizer and nanotechnology: a quick look. Better Crop with Plant Food 102: 18–19. https://doi.org/ 10.24047/bc102318
- Mohammad Fakhrul Islam S, Karim Z (2020) World's demand for food and water: the consequences of climate change, In: Desalination - challenges and opportunities. Intechopen, bangladesh, pp. 1–27. https://doi.org/10.5772/intechopen.85919
- Monica RC, Cremonini R (2009) Nanoparticles and higher plants. Caryologia 62:161–165. https://doi.org/10.1080/00087114. 2004.10589681
- Monreal CM, Derosa M, Mallubhotla SC, Bindraban PS, Dimkpa C (2016) Nanotechnologies for increasing the crop use efficiency of fertilizer-micronutrients. Biol Fertil Soils 52:423–437. https://doi.org/10.1007/s00374-015-1073-5
- Mozafari MR (2006). Nanocarrier Technologies: Frontiers of Nanotherapy. https://doi.org/10.1007/978-1-4020-5041-1
- Mukhopadhyay SS (2014) Nanotechnology in agriculture: Prospects and constraints. Nanotechnol Sci Appl 7:63–71. https://doi.org/ 10.2147/NSA.S39409



- Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assay with Tobacco tissur culture. Physiol Plant 15:474–497
- Mustafa F, Andreescu S (2020) Nanotechnology-based approaches for food sensing and packaging applications. RSC Adv 10:19309–19336. https://doi.org/10.1039/d0ra01084g
- Mustafa HS, Oraibi AG, Ibrahim KM, Ibrahim NK (2017) Influence of Silver and Copper Nanoparticles on Physiological Characteristics of Phaseolus vulgaris L. in vitro and in vivo. Int J Curr Microbiol App Sci 6(1):834–843. https://doi.org/10. 20546/ijcmas.2017
- Nair R, Varghese SH, Nair BG, Maekawa T, Yoshida Y, Kumar DS (2010) Nanoparticulate material delivery to plants. Plant Sci 179:154–163. https://doi.org/10.1016/j.plantsci.2010.04.012
- Niemiec M, Komorowska M (2018) The use of slow-release fertilizers as a part of optimization of celeriac production technology. Agric Eng 22:59–68. https://doi.org/10.1515/agriceng-2018-0016
- Nile SH, Baskar V, Selvaraj D, Nile A, Xiao J, Kai G (2020) Nanotechnologies in food science: applications, recent trends, and future perspectives, Nano-Micro Letters. Springer Singapore. https:// doi.org/10.1007/s40820-020-0383-9
- Noji T, Kamidaki C, Kawakami K, Shen JR, Kajino T, Fukushima Y, Sekitoh T, Itoh S (2011) Photosynthetic oxygen evolution in mesoporous silica material: adsorption of photosystem II reaction center complex into 23 nm nanopores in SBA. Langmuir 27:705–713. https://doi.org/10.1021/la1032916
- Nowack B, Bucheli TD (2007) Occurrence, behavior and effects of nanoparticles in the environment. Environ Pollut 150:5–22. https://doi.org/10.1016/j.envpol.2007.06.006
- Nowack B, Mueller NC (2008) Exposure modeling of engineered nanoparticles in the environment. Environ Sci Technol 42:4447–4453
- Nowack B, Schulin R, Robinson BH (2006) Critical assessment of chelant-enhanced metal phytoextraction. Environ Sci Technol 40:5225–5232. https://doi.org/10.1021/es0604919
- Omanović-Mikličanin E, Maksimović M (2018) Application of nanotechnology in agriculture and food production, nanofood and nanoagriculture, in: IcETRAN. Palic, Serbia.
- Ombódi A, Saigusa M (2000) Broadcast application versus band application of polyolefln-coated fertilizer on green peppers grown on andisol. J Plant Nutr 23:1485–1493. https://doi.org/ 10.1080/01904160009382116
- Padua GW (2010) Nanocomposites in food packaging. Food Eng Ingredients 35:35–37. https://doi.org/10.1002/9781119179 108.ch15
- Pan B, Xing B (2012) Applications and implications of manufactured nanoparticles in soils: a review. Eur J Soil Sci 63:437–456. https://doi.org/10.1111/j.1365-2389.2012.01475.x
- Paredes AJ, Asencio CM, Manuel LJ, Alberto D (2016) Nanoencapsulation in the food industry: manufacture, applications and characterization. J Food Bioeng Nanoprocessing 1:56–79
- Park S, Croteau P, Boering KA, Etheridge DM, Ferretti D, Fraser PJ, Kim KR, Krummel PB, Langenfelds RL, Van Ommen TD, Steele LP, Trudinger CM (2012) Trends and seasonal cycles in the isotopic composition of nitrous oxide since 1940. Nat Geosci 5:261–265. https://doi.org/10.1038/ngeo1421
- Pathakoti K, Manubolu M, Hwang HM (2017) Nanostructures: current uses and future applications in food science. J Food Drug Anal 25:245–253. https://doi.org/10.1016/j.jfda.2017.02.004
- Peralta-Videa JR, Zhao L, Lopez-Moreno ML, de la Rosa G, Hong J, Gardea-Torresdey JL (2011) Nanomaterials and the environment: a review for the biennium 2008–2010. J Hazard Mater 186:1–15. https://doi.org/10.1016/j.jhazmat.2010.11.020
- Perinelli DR, Fagioli L, Campana R, Lam JKW, Baffone W, Palmieri GF, Casettari L, Bonacucina G (2018) Chitosan-based

nanosystems and their exploited antimicrobial activity. Eur J Pharm Sci 117:8–20. https://doi.org/10.1016/j.ejps.2018.01.046

- Peterson HL (1975) Effects of high concentrations of several fertilizer salts on microbial biomass, numbers, and activity in several Iowa soils.
- Pokropivny V, Lohmus R, Hussainova I, Pokropivny A, Vlassov S (2007) Introduction to nanomaterials and nanotechnology. Tartu University Press, Ukraine. https://doi.org/10.1201/9781315153 285-1
- Pradhan S, Patra P, Mitra S, Dey KK, Basu S, Chandra S, Palit P, Goswami A (2015) Copper nanoparticle (CuNP) nanochain arrays with a reduced toxicity response: a biophysical and biochemical outlook on *Vigna radiata*. J Agric Food Chem 63:2606–2617. https://doi.org/10.1021/jf504614w
- Prasad R (2014) Synthesis of silver nanoparticles in photosynthetic plants. J Nanoparticles 2014:1–8. https://doi.org/10.1155/2014/ 963961
- Prasad R, Bhattacharyya A, Nguyen QD (2017) Nanotechnology in sustainable agriculture: Recent developments, challenges, and perspectives. Front Microbiol 8:1–13. https://doi.org/10.3389/ fmicb.2017.01014
- Prasad R, Pandey R, Barman I (2016) Engineering tailored nanoparticles with microbes: Quo vadis? Wiley Interdiscip. Rev Nanomedicine Nanobiotechnology 8:316–330. https://doi.org/10. 1002/wnan.1363
- Prasad TNVKV, Sudhakar P, Sreenivasulu Y, Latha P, Munaswamy V, Raja Reddy K, Sreeprasad TS, Sajanlal PR, Pradeep T (2012) Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. J Plant Nutr 35:905–927. https:// doi.org/10.1080/01904167.2012.663443
- Prashar P, Shah S (2016) Impact of fertilizers and pesticides on soil microflora in agriculture. https://doi.org/10.1007/ 978-3-319-26777-7_8
- Pushpa SS, Lohani (2018) Silica nanoparticles : Its green synthesis and importance in agriculture. J Pharmacogn Phytochem 7: 3383–3393
- Rai M, Ribeiro C, Mattoso L, Duran N (2015). Nanotechnologies in Food and Agriculture. https://doi.org/10.1007/ 978-3-319-14024-7
- Ramesh A, Sharma SK, Sharma MP, Yadav N, Joshi OP (2014) Inoculation of zinc solubilizing Bacillus aryabhattai strains for improved growth, mobilization and biofortification of zinc in soybean and wheat cultivated in Vertisols of central India. Appl Soil Ecol 73:87–96. https://doi.org/10.1016/j.apsoil.2013.08.009
- Raskar SV, Laware SL (2014) Effect of zinc oxide nanoparticles on cytology and seed germination in onion. Int J Curr Microbiol Appl Sci 3:467–473
- Rastogi A, Tripathi DK, Yadav S, Chauhan DK, Živčák M, Ghorbanpour M, El-Sheery NI, Brestic M (2019) Application of silicon nanoparticles in agriculture. 3 Biotech 9: 1–11. https://doi.org/ 10.1007/s13205-019-1626-7
- Rhim JW, Park HM, Ha CS (2013) Bio-nanocomposites for food packaging applications. Prog Polym Sci 38:1629–1652. https://doi. org/10.1016/j.progpolymsci.2013.05.008
- Rizwan M, Ali S, Ali B, Adrees M, Arshad M, Hussain A, Zia ur Rehman M, Waris AA, (2019) Zinc and iron oxide nanoparticles improved the plant growth and reduced the oxidative stress and cadmium concentration in wheat. Chemosphere 214:269–277. https://doi.org/10.1016/j.chemosphere.2018.09.120
- Roberts TL (2009) The role of fertilizer in growing the world's food. Better Crop 93:12–15
- Ronghao L, Kang Y, Pei L, Wan S, Shiping L, Liu S (2016) Use of a new controlled-loss-fertilizer to reduce nitrogen losses during winter wheat cultivation in the Danjiangkou Reservoir Area of



China. Commun Soil Sci Plant Anal 47:1137–1147. https://doi. org/10.1080/00103624.2016.1166245

- Ropers MH, Terrisse H, Mercier-Bonin M, Humbert B (2017) Titanium Dioxide as Food Additive. Appl Titan Dioxide 3–22. https://doi. org/10.5772/intechopen.68883
- de la Rosa G, López-Moreno ML, de Haro D, Botez CE, Peralta-Videa JR, Gardea-Torresdey JL (2013) Effects of ZnO nanoparticles in alfalfa, tomato, and cucumber at the germination stage: root development and X-ray absorption spectroscopy studies. Pure Appl Chem 85:2161–2174. https://doi.org/10.1351/ PAC-CON-12-09-05
- Rudenko NN, Mubarakshina MMB, Ignatova LK, Fedorchuk TP, Ivanov EMNZBN (2020) Role of plant carbonic anhydrases under stress conditions, In: Plant Stress Physiology. https://doi. org/10.5772/intechopen.91971
- Rui M, Ma C, Hao Y, Guo J, Rui Y, Tang X, Zhao Q, Fan X, Zhang Z, Hou T, Zhu S (2016) Iron oxide nanoparticles as a potential iron fertilizer for peanut (*Arachis hypogaea*). Front Plant Sci 7:1–10. https://doi.org/10.3389/fpls.2016.00815
- Sagadevan S, Periasamy M (2014) Recent trends in nanobiosensors and their applications -a review. Rev Adv Mater Sci 36:62–69
- dos Santos PP, Andrade LDA, Flôres SH, Rios ADO, (2018) Nanoencapsulation of carotenoids: a focus on different delivery systems and evaluation parameters. J Food Sci Technol 55:3851–3860. https://doi.org/10.1007/s13197-018-3316-6
- Scott N, Chen H (2013) Nanoscale science and engineering for agriculture and food systems. Ind Biotechnol 9:17–18. https://doi. org/10.1089/ind.2013.1555
- Sedghi M, Hadi M, Toluie S (2013) Effect of nano zinc oxide on the germination parameters of soybean seeds under drought stress. Ann West Univ Timişoara Ser Biol 16:73–78
- Shah V, Belozerova I (2009) Influence of metal nanoparticles on the soil microbial community and germination of lettuce seeds. Water Air Soil Pollut 197:143–148. https://doi.org/10.1007/ s11270-008-9797-6
- Shang Y, Hasan K, Ahammed GJ, Li M, Yin H (2019) Applications of nanotechnology in plant growth and crop protection: A review. Molecules 24:2558. https://doi.org/10.3390/molecules24142558
- Shefer A, Shefer S D (2003a) Multicomponent biodegradable bioadhesive controlled release system for oral care products. US 6,589,562 B1.
- Shefer A, Shefer SD (2003b) Biodegradable bioadhesive controlled release system of nano-particles for oral care products. US 2003b/0147956A1.
- Shefer A, Shefer SD (2005) Multi component controlled release system for oral care, food products, nutraceutical and beverages. US 2005/0112235 A1.
- Shinde S, Paralikar P, Ingle AP, Rai M (2020) Promotion of seed germination and seedling growth of *Zea mays* by magnesium hydroxide nanoparticles synthesized by the filtrate from *Aspergillus niger*. Arab J Chem 13:3172–3182. https://doi.org/10. 1016/j.arabjc.2018.10.001
- Siddiqui MH, Al-Khaishany MY, Al-Qutami MA, Al-Whaibi MH, Grover A, Ali HM, Al-Wahibi MS (2015a) Morphological and physiological characterization of different genotypes of faba bean under heat stress. Saudi J Biol Sci 22:656–663. https://doi.org/ 10.1016/j.sjbs.2015.06.002
- Siddiqui MH, Al-Whaibi MH (2014) Role of nano-SiO₂ in germination of tomato (*Lycopersicum esculentum* seeds Mill.). Saudi J Biol Sci 21:13–17. https://doi.org/10.1016/j.sjbs.2013.04.005
- Siddiqui MH, Al-Whaibi MH, Faisal M, Al Sahli AA (2014) Nanosilicon dioxide mitigates the adverse effects of salt stress on *Cucurbita pepo* L. Environ Toxicol Chem 33:2429–2437. https:// doi.org/10.1002/etc.2697

- Siddiqui MH, Al-Whaibi MH, Mohammad F (2015b) Nanotechnology and plant sciences: Nanoparticles and their impact on plants. 1–303. https://doi.org/10.1007/978-3-319-14502-0
- Singh Sekhon B (2014) Nanotechnology in agri-food production: an overview. Nanotechnol Sci Appl 7:31–53. https://doi.org/10. 2147/NSA.S39406
- Singh T, Shukla S, Kumar P, Wahla V, Bajpai VK (2017) Application of nanotechnology in food science: perception and overview. Front Microbiol 8:1–7. https://doi.org/10.3389/fmicb.2017. 01501
- Smita S, Gupta SK, Bartonova A, Dusinska M, Gutleb AC, Rahman Q (2012) Nanoparticles in the environment: assessment using the causal diagram approach. Environ Heal A Glob Access Sci Source 11:1–11. https://doi.org/10.1186/1476-069X-11-S1-S13
- Stampoulis D, Sinha SK, White JC (2009) Assay-dependent phytotoxicity of nanoparticles to plants. Environ Sci Technol 43:9473– 9479. https://doi.org/10.1021/es901695c
- Studer AJ, Gandin A, Kolbe AR, Wang L, Cousins AB, Brutnell TP (2014) A limited role for carbonic anhydrase in C₄ photosynthesis as revealed by a ca1ca₂ double mutant in maize. Plant Physiol 165:608–617. https://doi.org/10.1104/pp.114.237602
- Sturikova H, Krystofova O, Huska D, Adam V (2018) Zinc, zinc nanoparticles and plants. J Hazard Mater 349:101–110. https://doi. org/10.1016/j.jhazmat.2018.01.040
- Suganami M, Suzuki Y, Kondo E, Nishida S, Konno S, Makino A (2020) Effects of overproduction of rubisco activase on rubisco content in transgenic rice grown at different N levels. Int J Mol Sci 21. https://doi.org/10.3390/ijms21051626
- Suriyaprabha R, Karunakaran G, Yuvakkumar R, Rajendran V, Kannan N (2012) Silica nanoparticles for increased silica availability in maize (*Zea mays. L*) seeds under hydroponic conditions. Curr Nanosci 8:902–908. https://doi.org/10.2174/157341312803989 033
- Tandy S, Schulin R, Nowack B (2006) The influence of EDDS on the uptake of heavy metals in hydroponically grown sunflowers. Chemosphere 62:1454–1463. https://doi.org/10.1016/j.chemo sphere.2005.06.005
- Tarafdar JC, Sharma S, Raliya R (2013) Nanotechnology: interdisciplinary science of applications. African J Biotechnol 12:219–226. https://doi.org/10.5897/AJB12
- Tarafder C, Daizy M, Alam MM, Ali MR, Islam MJ, Islam R, Ahommed MS, Aly Saad Aly M, Khan MZH (2020) Formulation of a hybrid nanofertilizer for slow and sustainable release of micronutrients. ACS Omega 5:23960–23966. https://doi.org/10. 1021/acsomega.0c03233
- Usman M, Farooq M, Wakeel A, Nawaz A, Cheema SA, Rehman Hur, Ashraf I, Sanaullah M (2020) Nanotechnology in agriculture: current status, challenges and future opportunities. Sci Total Environ 721: 137778. https://doi.org/10.1016/j.scitotenv.2020. 137778
- Le Van N, Ma C, Shang J, Rui Y, Liu S, Xing B (2016) Effects of CuO nanoparticles on insecticidal activity and phytotoxicity in conventional and transgenic cotton. Chemosphere 144:661–670. https://doi.org/10.1016/j.chemosphere.2015.09.028
- Verma A, Gautam S, Bansal K, Prabhakar N, Rosenholm J (2019) Green Nanotechnology: advancement in phytoformulation research. Medicines 6:39. https://doi.org/10.3390/medicines6 010039
- Vidotti M, Carvalhal RF, Mendes RK, Ferreira DCM, Kubota LT (2011) Biosensors based on gold nanostructures. J Braz Chem Soc 22:3–20. https://doi.org/10.1590/S0103-505320110001000 02
- Wang L, Hu C, Shao L (2017) The-antimicrobial-activity-of-nanoparticles-present-situati. Int J Nanomedicine 12:1227–1249



- Wang LJ, Wang YH, Li M, Fan MS, Zhang FS, Wu XM, Yang WS, Li TJ (2002) Synthesis of ordered biosilica materials. Chinese J Chem 20:107–110. https://doi.org/10.1002/cjoc.20020200121
- Wen Y, Zhang L, Chen Z, Sheng X, Qiu J, Xu D (2016) Co-exposure of silver nanoparticles and chiral herbicide imazethapyr to *Arabidopsis thaliana*: Enantioselective effects. Chemosphere 145:207– 214. https://doi.org/10.1016/j.chemosphere.2015.11.035
- Wilson MA, Tran NH, Milev AS, Kannangara GK, Volk H, Lu GM (2008) Nanomaterials in soils. Geoderma 146(1-2):291-302. https://doi.org/10.1016/j.geoderma.2008.06.004
- Wu S, Mickley LJ, Jacob DJ, Rind D, Streets DG (2008) Effects of 2000–2050 changes in climate and emissions on global tropospheric ozone and the policy-relevant background surface ozone in the United States, J. Geophys Res 113, D18312, https://doi. org/10.1029/2007JD009639
- Xue J, Luo Z, Li P, Ding Y, Cui Y, Wu Q (2014) A residue-free green synergistic antifungal nanotechnology for pesticide thiram by ZnO nanoparticles. Sci Rep 4:1–9. https://doi.org/10.1038/srep0 5408
- Yang F, Hong F, You W, Liu C, Gao F, Wu C, Yang P (2006) Influences of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. Biol Trace Elem Res 110:179–190. https://doi.org/10. 1385/BTER:110:2:179
- Younes NA, Hassan HS, Elkady MF, Hamed AM, Dawood MFA (2020) Impact of synthesized metal oxide nanomaterials on seedlings production of three Solanaceae crops. Heliyon 6: e03188. https://doi.org/10.1016/j.heliyon.2020.e03188
- Yusefi-Tanha E, Fallah S, Rostamnejadi A, Pokhrel LR (2020) Zinc oxide nanoparticles (ZnONPs) as a novel nanofertilizer: Influence on seed yield and antioxidant defense system in soil grown soybean (*Glycine max* cv. *Kowsar*). Sci Total Environ 738. https://doi.org/10.1016/j.scitotenv.2020.140240
- Zeng W, Wang H, Li Z (2016) Nanomaterials for Sensing Applications J Nanotechnol 2016:2–4. https://doi.org/10.1155/2016/2083948
- Zhang Y, Liu N, Wang W, Sun J, Zhu L (2020) Photosynthesis and related metabolic mechanism of promoted rice (*Oryza sativa* L.)

growth by TiO₂ nanoparticles. Front Environ Sci Eng 14. https:// doi.org/10.1007/s11783-020-1282-5

- Zhao L, Sun Y, Hernandez-Viezcas JA, Hong J, Majumdar S, Niu G, Duarte-Gardea M, Peralta-Videa JR, Gardea-Torresdey JL (2015) Monitoring the environmental effects of CeO₂ and ZnO nanoparticles through the life cycle of corn (*Zea mays*) plants and in situ μ-XRF mapping of nutrients in kernels. Environ Sci Technol 49:2921–2928. https://doi.org/10.1021/es5060226
- Zheng L, Hong F, Lu S, Liu C (2005) Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. Biol Trace Elem Res 104:83–91. https://doi.org/10.1385/BTER:104:1:083
- Zhu YX, Gong HJ, Yin JL (2019) Role of silicon in mediating salt tolerance in plants: a review. Plants 8:6–10. https://doi.org/10. 3390/plants8060147
- Zia-ur-Rehman M, Naeem A, Khalid H, Rizwan M, Ali S, Azhar M (2018) Responses of plants to iron oxide nanoparticles, nanomaterials in plants, algae, and microorganisms. Elsevier Inc. https:// doi.org/10.1016/B978-0-12-811487-2.00010-4
- Zorraquín-Peña I, Cueva C, Bartolomé B, Moreno-Arribas MV (2020) Silver nanoparticles against foodborne bacteria. Effects at intestinal level and health limitations. Microorganisms 8. https://doi. org/10.3390/microorganisms8010132
- Zulfiqar F, Navarro M, Ashraf M, Akram NA, Munné-Bosch S (2019 Nanofertilizer use for sustainable agriculture: Advantages and limitations. Plant Sci 289. https://doi.org/10.1016/j.plantsci. 2019.110270
- Zuo Y, Zhang F (2011) Soil and crop management strategies to prevent iron deficiency in crops. Plant Soil 339:83–95. https://doi.org/10. 1007/s11104-010-0566

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