

REVIEW

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Mechanisms and applications of probiotics in prevention and treatment of swine diseases

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Abstract

Probiotics can improve animal health by regulating intestinal flora balance, improving the structure of the intestinal mucosa, and enhancing intestinal barrier function. At present, the use of probiotics has been a research hotspot in prevention and treatment of different diseases at home and abroad. This review has summarized the researchers and applications of probiotics in prevention and treatment of swine diseases, and elaborated the relevant mechanisms of probiotics, which aims to provide a reference for probiotics better applications to the prevention and treatment of swine diseases.

Keywords Probiotic, Swine diseases, Mechanisms of action, Applications

Introduction

Meat is an essential source of nutrients and energy for modern humans due to its balanced chemical nutrition, culinary potential and excellent digestibility. Swine meat has accounted for a large portion of world-wide meat consumption [1]. Pig farming is often accompanied by diseases. Once the epidemic breaks out, it will not only slow down the growth of pigs, reduce the supply of pork [2], but cause huge economic losses for farmers. The outbreak of Africa swine fever since 2018 has brought a devastating blow to the pig industry [3]. Antibiotics have positive effects in the process of swine disease treatment,

such as inhibiting or killing pathogenic bacteria, reducing the incidence of disease, regulating the balance of intestinal flora, and promoting growth performance [4]. However, the long-term and extensive use of antibiotics has led to the increase of drug resistance, so the massive use of antibiotics in animal husbandry has attracted great attention among farmers and researchers. The complete ban on the use of antibiotics in animal feed among many countries has led to the growth of research on the use of probiotics to combat bacterial infections in humans and animals. [5]. Probiotics have a significant effect in replacing feed antibiotics and are beneficial to host health when ingested in sufficient quantities, so have been developed as alternative feed additives to the prophylactic use of antibiotics [6]. The review provides an overview on the types of probiotics, the application of probiotics in swine disease treatment and the mechanisms, aiming to provide a reference for the better application of probiotics in swine disease treatment.

Characteristics of probiotics in pig industry

Probiotics are a class of live microorganisms which contribute to the health of the host. Probiotics are believed to improve the host health through increasing the

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colonization in host intestine to regulate the balance of the intestinal flora, compete with intestinal pathogenic bacteria, and enhance immunity [7]. The use of probiotics in the swine farming industry must have these following characteristics. Firstly, probiotics survive in the gastrointestinal tract, which means they resist digestion of gastric acid in order to interact with the microorganisms native in the host gut. Secondly, they can improve animal health by stimulating the host's immune response or through indirect mechanisms, such as reducing damage of pathogenic bacteria. Thirdly, meet production requirements, such as suitability for large-scale production, stable shelf storage, good sensory characteristics, etc. Last but not the least, it has no toxic effects on the host and is not pathogenic [8, 9]. The expected characteristics of probiotics is shown in Fig. 1.

Most used probiotics bacteria in agricultural breeding

Lactobacillus

Lactobacillus, a class of lactic acid-producing Gram-positive bacteria. This genus includes many species such as *Lactobacillus acidophilus*, *Lactobacillus rhamnosus*, *Lactobacillus casei*, *Lactobacillus johnsonii* and so on. *Lactobacillus* promote the intestinal health and growth performance of animals by regulating the expression of

cytokines and enhancing intestinal barrier function [10]. Deoxynivalenol (DON) exposure induces liver inflammation and oxidative stress through Keap1-Nrf2 signaling pathway, while *Lactobacillus rhamnosus* oral administration attenuates the liver dysfunction, improve the antioxidant capacity of liver and protect mice from DON injury [11].

Bifidobacterium

Bifidobacterium is a Gram-positive, immobile and strictly anaerobic bacterium. Common probiotic *Bifidobacterium* includes *Bifidobacterium bifidum*, *Bifidobacterium longum*, *Bifidobacterium breve*, *Bifidobacterium adolescentis*, *Bifidobacterium infantis*, etc. There is large number of *Bifidobacterium* in animal guts, and its state is an indicator to measure the health of the host [12]. Studies have shown that *Bifidobacterium* has abilities to inhibit harmful bacteria, modulate the composition of intestinal microbial, improve gastrointestinal barrier function, and promote the growth performance of animals [13–15].

Bacillus

Bacillus is a Gram-positive, aero-anaerobic or facultative aerobic bacterium. Some bacteria of this genus, such as *Bacillus subtilis* and *Bacillus licheniformis*, are often used as supplement to animal feed and play a vital role. Compared with other non-spore-forming bacteria,

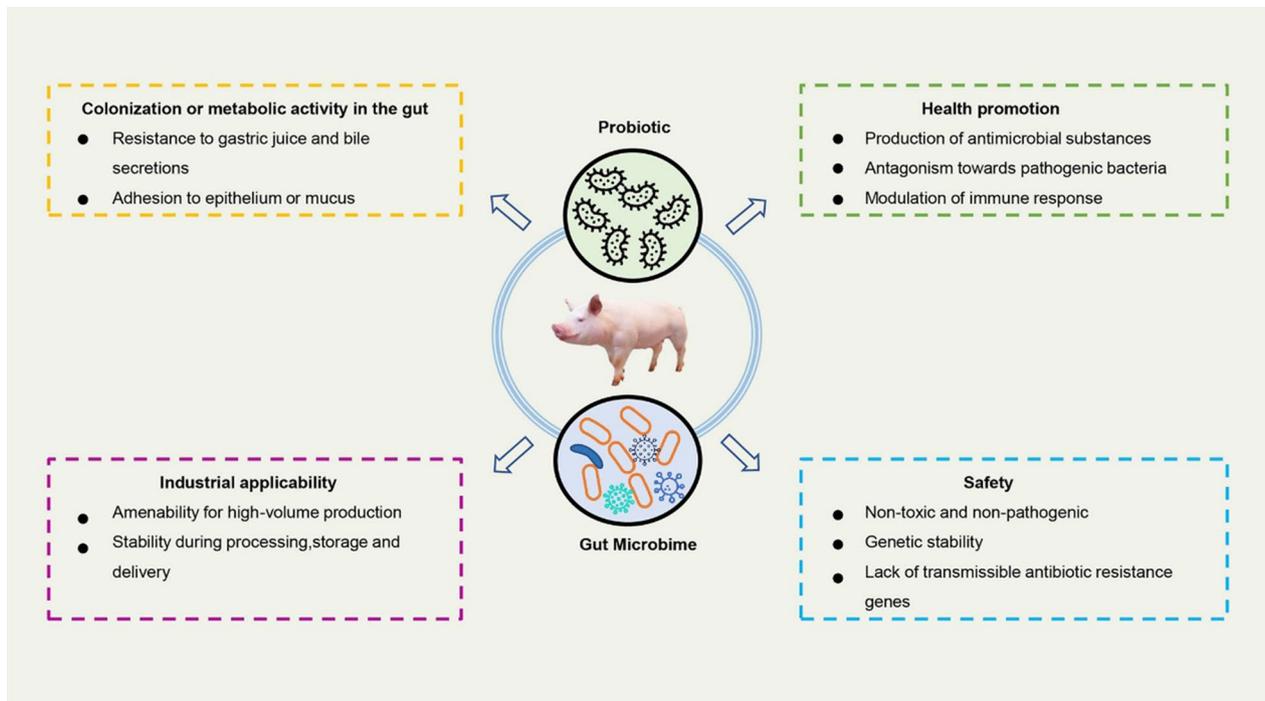


Fig. 1 Expected characteristics of probiotics

Bacillus has many advantages because of its endospores. First of all, it can be stored in dry form at room temperature without any impact on its viability. Secondly, the spores are resistant to the threat of extreme environments, which means that *Bacillus* survives in the low pH environment such as gastric acid [16, 17]. Additionally, *Bacillus* secretes enzymes to improve the digestibility of the feed and promote animal growth. When enters the intestine tract, *Bacillus* inhibits the growth of pathogenic microorganisms, and secrete a variety of antibacterial substances to improve the ability of the intestinal immune system, and reduce the occurrence of diseases [18]. Coccidiosis caused by *Eimeria*, which has been recognized as a parasitic disease in chickens that significantly affects the gastrointestinal tract [19, 20]. Dietary supplementation with *Bacillus* strains resisted the negative impacts caused by coccidiosis through different manners [21, 22]. The mechanisms of action of *Bacillus Subtilis* are shown in Fig. 2.

Lactic acid bacteria

Lactic acid bacteria refer to a class of non-spore, gram positive bacteria, which is the main product of lactic acid and include multiple species, such as *Lactococci* and *Streptococci* [23]. In animal production, *lactic acid bacteria* are commonly found in fermented feeds. *Lactic acid bacteria* further improve the gastrointestinal function by balancing the intestinal flora, and

degrade the macromolecular substances that are not easily absorbed in the body, so they improve the digestive ability of the gastrointestinal tract of animals. The lactic acid, hydrogen peroxide, peptides, etc. produced by *lactic acid bacteria* regulate the immune function and enhance immunity. In addition, *lactic acid bacteria* also reduce the synthesis of cholesterol by reducing the activity of lipid synthetase and assimilating cholesterol. After fermentation, they also synthesize flavor compounds such as acetaldehyde and butanedione to improve the palatability of the feed [24].

Yeast

Yeast is a facultative anaerobic eukaryotic single-celled microorganism. In recent years, the interest in using yeasts as probiotics increased, not only for human health but also to improve growth and health of animals and birds, in particular in aquaculture or with respect to industry cattle, pigs, and poultry. *Yeast* cells contains proteins, vitamin B complexes and important trace mineral. And *Yeast* can produce extracellular enzymes, such as amylase, galactosidase, inositol hexaphosphatase, etc., which play an important role in improving the nutritional value and utilization of feed and improving animal production performance [25]. The positive effects of *yeast* on the health and production performance of different animals have been

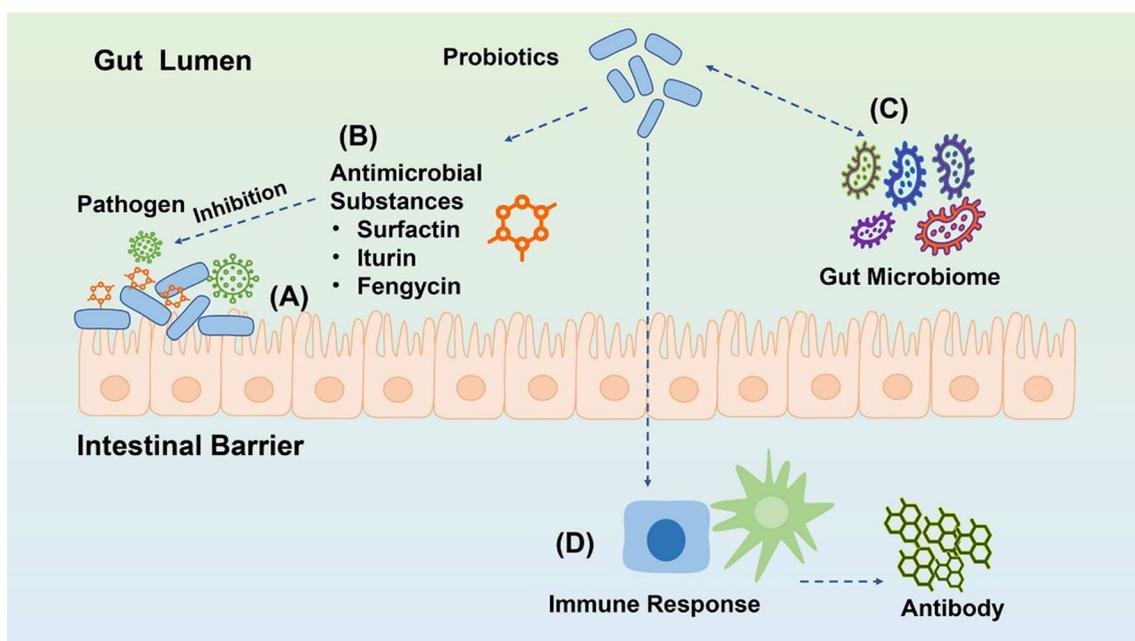


Fig. 2 Mechanisms of action of *Bacillus Subtilis*. **A** Competitive adhesion site of pathogenic microorganisms. **B** Production of antimicrobial substances. **C** Regulation of intestinal flora balance. **D** Stimulation of the immune system

Table 1 The positive influence of yeast supplementation on animal's health, nutrition and performance [25]

Animal species	Effects
Ruminants	Reduces lactate production or accumulation; Balance ruminal pH; Stimulate the growth of cellulolytic and fibrolytic microorganisms, increasing net fiber intake and digestion; Increasing the total ruminal microflora; Improve the health of the ruminant; Reduce methane emissions
Poultry	Modulate of intestinal microflora; Stimulate immune system; Inhibit the growth of pathogenic bacteria; Increase organic phosphorus utilization; Increasing the egg production and improve the internal egg quality of poultry birds; Improve feed intake; Reduce plasma cholesterol; Promote meat quality of poultry
Pigs	Improve the pig's gut microflora balance; Proper maturation of the gut associated tissue; Modulate immune response reducing enteric pathogens; Reduce post-weaning diarrhea (PWD) symptoms in pigs; Exhibit growth promoting properties in weaning and fattening pigs; Improve feed conversion efficiency
Horses	Ameliorates the dysfunction of horse's ecosystem such as lactic acidosis, colitis, laminitis and enterotoxaemia; Increase in digestibility of diet nutrients; Enhances enzyme production in the hindgut
Aquaculture	Improve growth in juvenile and adult fish; Enhance survival rate; Positively modulate cellular innate immune system parameters; Stimulation of enzymatic antioxidative response of farmed fish

Table 2 Some other strains used as probiotics in animal feed

Genus	Species	References
<i>Escherichia</i>	<i>Escherichia coli</i> Nissle (<i>EcN</i>)	[26]
<i>Enterococcus</i>	<i>E. faecalis</i>	[27, 28]
	<i>E. faecium</i>	[29, 30]
	<i>E. gallinarum</i>	[31]
	<i>E. casseliflavus</i>	[32]
<i>Aspergillus</i>	<i>A. oryzae</i>	[33–35]
	<i>A. niger</i>	[36]
<i>Lactococcus</i>	<i>Lc. lactis</i>	[37]
<i>Kluyveromyces</i>	<i>K. fragilis</i>	[38]
	<i>K. marxianus</i>	[39]
<i>Leuconostoc</i>	<i>L. mesenteroides</i>	[40]
<i>Pediococcus</i>	<i>P. acidilactici</i>	[41, 42]
<i>Streptococcus</i>	<i>S. thermophilus</i>	[43]
	<i>S. phocae</i>	[44]

shown in Table 1. Besides improving production performance, *yeast* also regulate balance of intestinal flora, strengthen the immune system, improve animal health, etc.

Other probiotics strains

Some other strains used as probiotics in animal feed are given in Table 2.

Applications of probiotics in the prevention and treatment of swine diseases

Some reports have shown that probiotics not only maintain the microbiome balance of intestinal flora, also enhance the immunity and disease resistance of pigs, therefor reducing the severity and risk of disease in animals. In the process of pig framing, the addition of probiotics efficiently reduce the use of antibiotics, so as

to improve pig production performance and disease response and other multiple challenges [45]. The application effects of probiotics in pigs are shown in Table 3. This review mainly focuses on the research of probiotics in the prevention and treatment of swine diseases.

Applications of probiotics in porcine respiratory diseases

Respiratory diseases in pigs are common in modern pork production. And respiratory diseases are often referred to as Porcine Respiratory Disease Syndrome (PRDC) which is caused by the combination of one or more viruses and bacteria [46]. The main pathogens of pigs include viruses such as porcine reproductive and respiratory syndrome virus (PRRSV), swine influenza virus (SIV), pseudorabies virus (PRV), porcine circovirus type 2 (PCV2), porcine epidemic diarrhea virus (PEDV), as well as bacteria such as *Mycoplasma hyopneumoniae*, *Streptococcus*, *Actinobacillus Pleuropneumoniae*, *Haemophilus Parasuis*, and *Bocherichia bronchialsepticae* [47].

The application effect of probiotics in the prevention and treatment of swine respiratory diseases is mainly manifested in improving the immune capacity of pigs. Probiotics (such as *Bacillus subtilis* and *lactic acid bacteria*) play an important role in disease prevention and treatment as live carriers of bacteria. The genome of PCVs encodes two major proteins, replicase (Rep) and capsid protein (Cap). Rep is involved in viral genomic replication, while Cap is the only structural protein of PCVs, and contains multiple antigen epitopes [48]. For example, a recombinant strain expressing the CAP protein of the PCV2 virus was constructed with *Bacillus subtilis*, and it showed that piglets with oral administration of *Bacillus subtilis* had induced a strong humoral immune response by increasing the levels of PCV2-specific immunoglobulin A (IgA) and IgG [49]. *Mycoplasma hyopneumoniae* (Mhp) is the main pathogen of swine

Table 3 Application effects of probiotics in pigs

References	Tested animals and adding time	Probiotics and supplemental levels	Effects
Yi et al. [110]	Weaned pigs (21 days of age)	5×10^{10} CFU/kg <i>Lactobacillus reuteri</i> LR1	Improved intestinal morpho-logical structure; Increased intestinal absorption area; Increased in digestibility of diet nutrients; Improved growth performance
He et al. [111]	Weaned pigs (21 days of age)	10 ml 1×10^9 CFU/ml <i>Lactobacillus johnsonii</i> L531	
Sun et al. [55]	Weaned pigs (21 days of age)	<i>L. acidophilus</i> , <i>L. casei</i> , <i>B. thermophilum</i> and <i>E. faecium</i> with the concentration of 0.25×10^8 CFU/g for each strain	Improved immune capacity; Reduced the incidence of intestinal diseases such as diarrhea and intestinal inflammation
Fu et al. [78]	Weaned pig1s (body weight of 6.89 ± 0.15 kg)	2×10^9 CFU/kg <i>Bacillus coagulans</i> , or 2.5×10^{10} CFU/kg yeast hydrolysate	
Inatomi et al. [112]	Pregnant sows	1×10^6 CFU/g <i>B. mesentericus</i> TO-A、 1×10^6 CFU/g <i>C. butyricum</i> TO-A, 1×10^8 CFU/g <i>E. faecalis</i> T-110	Improved immune capacity; Reduced the incidence of intestinal diseases such as diarrhea and intestinal inflammation
Long et al. [71]	Weaned pigs (32 days of age)	1×10^{10} CFU/g yeast	
Li et al. [113]	Weaned pigs (25 ± 2 days of age)	1×10^7 CFU/kg <i>B. subtilis</i>	Modulated intestinal micro-flora; Enhanced epithelial barrier
Li et al. [114]	Weaned pigs (21 days of age)	10ml 5×10^9 CFU/ml <i>Lactobacillus delbrueckii</i>	
Zhao et al. [115]	Weaned pigs (21 ± 2 days of age)	6×10^6 CFU/kg <i>Lactobacillus casei</i>	Modulated intestinal micro-flora; Enhanced epithelial barrier
Zhang et al. [116]	Weaned pigs (1 days of age)	10ml 1×10^9 CFU/ml <i>Lactobacillus rhamnosus</i> GG	
Ding et al. [117]	Weaned pigs (21 days of age)	500 g t^{-1} <i>B. subtilis</i> DSM 32,315	Modulated intestinal micro-flora; Enhanced epithelial barrier
Zhang et al. [118]	Weaned pigs (26 ± 1 days of age)	1×10^9 CFU/kg <i>Clostridium butyrate</i>	
Cao et al. [100]	Finishing pigs	1×10^8 CFU/kg BaSC06	

pneumonia, which leads to symptoms such as coughing and wheezing in pigs. Recombinant *Bacillus subtilis* was constructed by using P97 and P46 of *Mycoplasma pneumoniae* as immunogenic proteins, and piglets were immunized by nasal spray using the recombinant strain. The results showed that the secretory immunoglobulin A (sIgA) in nasal swabs and IgG in pig serum had increased significantly after immunization [50]. The level of IgG and IgA antibodies in serum is an important indicator to evaluate the immune capacity. At the same time, sIgA is considered as an immune barrier on the surface of the intestinal mucosa, which prevents pathogens from adhering to intestinal epithelial cells and inhibit the invasion of pathogens [51]. Probiotics as vectors to express specific immunogenic proteins can induce pigs to produce higher levels of antibodies, thereby increasing the immunity of pigs.

In addition, probiotics inhibit the growth and reproduction of pathogens. Studies have shown that co-infected of weaned piglets with *Salmonella cholerae* in pigs and PRRSV leads to *Salmonella* colonization increasing in the lungs and aggravates the symptoms of pneumonia. The results showed that directly dietary supplementation of *Bacillus subtilis* could reduce the *Salmonella* colonization in the lungs and reduce the existence of PRRSV [52].

Applications of probiotics in porcine digestive tract disease

Digestive tract disease is a common type of pig disease. Although the disease will not cause death of animals, it will affect its growth performance, and then affects the income of farmers. Therefore, many farmers attach great importance to the prevention and treatment of digestive tract diseases. The digestive system is the place where the animal digests and absorbs nutrients. If there are problems with the digestive system of the pig, the digestion and absorption efficiency of pig feed will be reduced, and the weight gain effect of the pig will not be obvious, which will lead to a reduction of farmers' income. If the digestive system diseases are not treated in time, it is very likely that the gastric mucosa of pigs will be damaged, the immunity will be weakened, and the risk of other diseases in pigs will be increased [53].

Studies have shown that probiotics can reduce the severity of diarrhea and improve the health status of animals. PWD in piglets is associated with enterotoxin-producing *Escherichia coli* (ETEC) [54]. Dietary supplementation of a variety of probiotics (including *Lactobacillus acidophilus*, *Lactobacillus casei*, *Bifidobacterium thermophilus*, and *Enterococcus faecalis*) had reduced the severity of diarrhea caused by ETEC F18+ [55]. Oral administration of *Bacillus subtilis* WS-1 at 1.5×10^{10} CFU significantly had inhibited diarrhea and mortality in piglets caused by pathogenic *E. coli*, and the study also found that WS-1 also encodes a variety of functional proteins, such as lipopeptides, which may

confer antibacterial activity to WS-1 [56]. Newborn piglets in the treatment group with 2 ml of sterile skim milk suspended with viable *Lactobacillus rhamnosus* (5×10^8 CFU/mL) had decreased the diarrhea incidence and increased weaning weight and average daily weight gain [57]. Metabolites of probiotics can also inhibit the replication of pathogens. PEDV is an enteropathogenic virus that causes diarrhea in pigs and is also associated with high morbidity and mortality in piglets. One of the main components of *Lactobacillus plantarum* metabolites is *Lactobacillus plantarum* extracellular polysaccharides (LPE), which elevate the transcription and apoptosis levels of tumor necrosis factor- α (TNF- α), inhibiting the replication of PEDV by regulating the apoptosis mechanism and inducing early apoptosis of damaged cells [58].

In addition, probiotics enhance the intestinal defenses by modulating intestinal flora. The addition of a BLS mixture (a mixture of *Bacillus licheniformis* and *Bacillus subtilis*) to piglet diets increased the relative abundance of *Bacteroides* and *Lactococcus* and decreased the abundance of *Brucella* and *Clostridium* [59]. Studies have shown that *Clostridium* is closely associated with the fermentation of proteins and could increase the risk of diarrhea [60]. In addition, the BLS mixture increased cytokine and TLR-4 levels in the ileum and colon. These findings suggest that BLS mixtures may modulate intestinal flora composition and improve intestinal health.

In addition to being added directly to feed, probiotics are used in feed fermentation as well. Through fermentation technology, anti-nutritional factors such as phytate and soybean antigen protein are decomposed, so as to improve feed digestibility. In addition, fermented feed can also affect the intestinal symbiotic flora, activate immune response, and be beneficial to animal health. For example, feeding wheat fermented by *Lactobacillus reuteri* to weaned piglets could increase the content of short-chain fatty acids and improve the intestinal health of weaned piglets, thereby reducing the incidence of diarrhea in pigs [61]. In addition, studies have shown that fermented soybean meal produced with *Lactobacillus* and *Clostridium* butyrate increased the level of immunoglobulin in pig serum and reduced the incidence of diarrhea in weaned piglets [62].

Mechanisms of action of probiotics

Regulation of intestinal flora balance

The microbiota in the gastrointestinal tract plays an important role in host metabolism, immunity, digestion, absorption, and development. The gastrointestinal tract is full of microorganisms that are closely associated with the animal itself and the feed it consumes. A good intestinal flora is essential for animal health by ingesting nutrients from the diet and regulates the development and

function of the digestive and immune systems to benefit the host.

Probiotics can alter the structure of the intestinal flora. Probiotics improve intestinal health by promoting the growth of beneficial bacteria, inhibiting the multiplication of harmful bacteria, and producing metabolites such as short-chain fatty acids. A balanced chicken GIT is predominated by *Firmicutes*, *Tenericutes*, *Bacteroidetes*, and *Proteobacteria* [65]. Once chickens are infected with *Eimeria*, the balance of intestinal flora is destroyed [63–65]. The use of probiotic modifies gut microbiota composition to benefit the growth of chickens [20, 66]. Study has reported that chickens infected with *Eimeria* significantly reduced the abundance of *Firmicutes* and increased abundances of *Proteobacteria* [22, 64]. The addition of *Bacillus subtilis* improved the adverse impacts of *Eimeria* through enhancing the abundance of *Bacillus*, *Weissella*, *Staphylococcus*, *Bacilli* unclassified and *Turicibacter* [22]. The addition of 500 g/t of *Bacillus subtilis* to weaned piglets' diets increased microbial β -diversity and relative abundance of *Bacillus*, *Bifidobacterium* and *Clostridium faecium* in the ileum and colon of weaned piglets [67]. *Phylum Firmicutes* and *Bacteroidetes* are dominant bacteria in the porcine intestine [68], *Bacteroidetes enterica* are involved in the degradation of carbohydrates and proteins and play a vital role in maintaining intestinal health through the production of butyrate [69]; fermentation of carbohydrates by *Clostridium faecalis* produces butyrate and acetate, along with formate or propionate; *Bifidobacterium* produce acetate to prevent intestinal infections and play an important role in immune regulation [70].

In addition, some substances in probiotics inhibit the growth and reproduction of harmful bacteria to reduce the occurrence of intestinal diseases. For example, the incidence of diarrhea in piglets could be reduced by adding *Saccharomyces cerevisiae* to the diet, which was mainly attributed to the cell wall polysaccharides of *Saccharomyces cerevisiae* [71]. The main active components of yeast cell wall polysaccharides are β -glucan and mannan, which play an important role in improving the immune system and protecting the health of animals [72]. Oligosaccharides promote the formation of beneficial bacteria in the intestine, inhibit the proliferation of pathogenic bacteria and regulate the intestinal flora as a way to maintain intestinal health. Study found that adding 0.3% and 0.5% *Saccharomyces boulardii* yeast wall polysaccharides to the diet could significantly suppressed the growth of *Salmonella* and *Clostridium perfringens* in early weaned lambs [73]. The above findings suggest that probiotics inhibit the growth of harmful bacteria in the intestine tract while increasing the number of dominant

bacteria, maintaining the balance of the intestinal flora and thus improving animal health.

Improvement intestinal mucosal structure and enhancement intestinal barrier function

The structure of the small intestine and the integrity of the intestinal barrier play a pivotal role in the digestion, absorption and transport of nutrients. In general, the digestion and absorption of nutrients in animals is positively correlated with the intestinal villi height and the ratio of intestinal villi height to crypt depth [74]. Studies have suggested that reduced villus height and increased crypt depth may be responsible for nutrient malabsorption, increased gastric secretion, and diarrhea [75]. Yang et al. [76] found that the addition of *Lactobacillus plantarum* during piglet feeding alleviated the decrease in jejunal villi height that occurs after enterotoxigenic *E. coli* attack. *Bacillus subtilis* increase the villi height and the ratio of villi height to crypt depth in the ileum, expanding the absorption area of nutrients and improving digestion and absorption in weaned piglets [67]. In addition, there is a certain relationship between the morphological structure, development and related genes of the intestine. *IGF1*, *IGF-1R*, *GLP-2* and *TGF- β 2* are the main markers of intestinal development and cell differentiation [77]. *Bacillus subtilis* upregulated the expression of *IGF1* and *GLP-2* in the jejunal mucosa and yeast hydrolysates upregulated the expression of mRNA of *GLP-2* and *TGF- β 2* [78]. Probiotics improve intestinal structure by promoting the expression of genes related to intestinal morphology and intestinal development, helping to increase the total surface area of the intestine, which is an effective guarantee for the absorption of nutrients such as nucleotides [79].

The tight junctions (TJs) between intestinal epithelial cells play an important role in the intestinal mucosal barrier. Damage to TJs leads to increased cell permeability, and bacteria and pathogens in the intestinal lumen penetrate the intestinal mucosa and enter other tissues, organ or circulatory system, thus leading to the occurrence of diseases. Probiotics enhance intestinal TJs and improve the defense of intestinal epithelial cells against pathogenic invasion. Studies have shown that *Lactobacillus plantarum* prevents the adhesion of enterotoxin-producing *E. coli* to intestinal epithelial cells and maintains the integrity of the intestinal barrier [80]. *Bacillus amyloliquefaciens* SC06 (BaSC06) protects the integrity of TJs and villi [81]. The TJs consists of tight junction proteins (claudins) and occludins, and pathogenic bacteria invade the intestines by attacking TJs through various virulence factors. *E. coli* destroys TJs by transferring the occlusion proteins from TJs to the cytoplasm, the absence of which leads to the infection of the organism with pathogenic bacteria

[82]. Therefore, tight junction proteins and occlusion proteins play an important role in TJs and intestinal barrier function. *E. coli* Nissle 1917 upregulated the expression of tight junction protein-1 (ZO-1) in intestinal epithelial cells [83]. *Lactobacillus rhamnosus* GG could prevent a decrease in ZO-1 expression induced by *E. coli* O157:H7 [84]. Probiotics increase the trans-epithelial resistance (TER) of intestinal epithelial cells by increasing the expression of occlusion and tight junction proteins, thereby repairing epithelial cell damage caused by pathogenic bacteria.

Production of antibacterial substances and inhibition the growth of pathogenic microorganisms

Antimicrobial substances, produced by probiotics, such as bacteriocins, hydrogen peroxide, organic acids and biosurfactants, inhibit the growth of pathogenic microorganisms and maintain intestinal health.

Bacteriocins, antimicrobial peptides produced by ribosomes, may inhibit or directly kill pathogenic microorganisms, thereby limiting the colonization ability of pathogenic microorganisms in the intestine. Bacteriocins can induce increased cytoplasmic membrane permeability in bacteria, which leads to cell leakage and inhibition of DNA, RNA synthesis and/or cell wall protein synthesis [85]. For example, nisin from *Streptococcus lactis* acts by forming complexes with cell membrane lipid II precursors, followed by aggregation of polypeptides bound to form discrete pores in the bacterial cell membrane [86].

Another mechanism of action of probiotics in the intestine is to make the intestinal environment unsuitable for the growth of pathogenic microorganisms by lowering pH. *Lactic acid bacteria* and commensal microbiota ferment carbohydrates in the gastrointestinal tract, leading to the production of metabolites such as acetic acid, formic acid, succinic acid and lactic acid, making the intestinal environment acidic and inhibiting the growth of pathogenic microorganisms. Organic acids (especially lactic and acetic acids) inhibit the growth of many pathogenic bacteria in the gastrointestinal tract [87]. The undissociated form of lactic acid acts as a permeabilizing agent for the outer cell membrane of Gram-negative bacteria, followed by dissociation into the bacterial cytoplasm, exerting a bactericidal effect by accumulating ionized forms of organic acids and other antibacterial compounds within the cytoplasm. Studies have shown that the strong inhibitory effect of *Lactobacillus rhamnosus* GG on *Salmonella typhimurium* is due to the production of lactic acid [88]. Lactic acid affects the expression of *HilA* and *InvF* virulence factors in *Salmonella* [89].

In addition to the production of organic acids and bacteriocins, the production of hydrogen peroxide by symbiotic microbiota and *lactic acid bacteria* may be an

important antimicrobial mechanism. Hydrogen peroxide reduces the virulence of pathogenic microorganisms, decreases the invasion of epithelial cells by pathogenic microorganisms, or causes the death of intestinal pathogenic microorganisms by diffusing within epithelial cells to alter gene transcription and signal transduction [90]. Biosurfactants are a class of compounds with surface and emulsifying activity that cause increased permeability of cells by disrupting or dissolving cell membranes [91]. For example, bacteriocins and biosurfactants produced by *Lactobacillus casei* MRTL3 could inhibit *Listeria monocytogenes*, *Staphylococcus aureus*, *Shigella* and *Pseudomonas aeruginosa* [92].

Enhancement of immune capacity

Probiotics stimulate immune system of the organism by increasing the production of antibodies and activating immune cells. Vibriosis is a common bacterial disease, which has a negative impact on economically farmed shrimp, marine fish and some freshwater fish [93, 94]. The dietary probiotic can significantly modulate the immune responses to reduce the mortality caused by several *Vibrio* species [95–97]. Serum immunoglobulin is one of the indicators of the immune status of the organism, and the addition of *Bacillus subtilis* and *Bacillus licheniformis* to the diet of weaned piglets could promote the production of serum IgG [98]. Fermented soybean meal produced with *Lactobacillus* and *Clostridium butyrate* enhanced immunity in weaned piglets by increasing serum IgG and IgA levels [62]. sIgA is the main immunoglobulin in the mucosal system and essential to protect the mucosal surface from toxins, viruses and enteric intestinal pathogens. *Lactobacillus* BS15 could increase intestinal sIgA levels and delay the decline in sIgA levels, which contributed to the maintenance of intestinal health [99].

Toll-like receptors (TLRs) are one of the key recognition receptors in the innate immune system and are expressed in various intestinal mucosal cells such as mucosal epithelial cells, macrophages, and dendritic cells. Increased TLR expression results in the release of cytokines including tumor necrosis factor (TNF), interleukin-4 (IL-4), and interferon- γ (IFN- γ) when probiotics are used to stimulate the innate immune system. *Bacillus amyloliquefaciens* SC06 (BaSC06) alleviated intestinal inflammation in fattening pigs by regulating the expression of pro-inflammatory cytokines IL-6, IL-8 and MCP1 in the intestinal mucosa [100]. *Lactobacillus fermentum* and *Pediococcus acidilactici* reduced the concentration of IL-6, IL-1 β , IFN- γ in the serum of weaned piglets, which helped to reduce the damage caused by inflammation [101].

Changes in T cell subsets in the peripheral blood are an important indicator of overall immunity levels, and mature T cells can be divided into two main subpopulations (CD4+ and CD8+) based on the cell surface proteins they express. CD4+ T cells are associated with major histocompatibility complex (MHC) class II molecules and act as helper or inflammatory cells in response to exogenous antigens. CD8+ T cells are associated with MHC class I molecules and play a key role in resistance to endogenous antigens [102, 103]. The ratio of CD4+ and CD8+ T lymphocytes is closely related to the immune function. For example, oral administration of *Bacillus subtilis* and total inactivation of PEDV resulted in an upregulated ratio of CD4+CD8+ and enhanced proliferation of memory T cells in the intestinal mucosa-associated lymphocytes of pigs [104].

Inhibition for the colonization of pathogenic microorganisms

Pathogenic microorganisms have binding molecules on their surfaces that can interact with host cell membranes in a similar manner to antigens. Pathogenic microorganisms have binding molecules on their surfaces that can interact with host cell membranes in a similar manner to antigens. For example, the *Lactobacillus plantarum* CCMA 0743 strain was able to reduce the adhesion of *Salmonella* to two types of cells (Caco-2 and HT-29) [105]. *Lactobacillus plantarum* ZLP001 has the ability to inhibit the adhesion of ETEC to IPEC-J2 cells, and the probiotic prevents the adhesion of pathogens via a competitive mechanism at the colonization site [106]. *Lactobacillus reuteri* improves intestinal health by producing extracellular polysaccharides to increase probiotic colonization in the intestine and inhibits the proliferation of enterotoxin-producing *E. coli* [107]. To gain this competitive advantage, probiotics can alter the intestinal environment by creating inhibitory compounds, lowering pH, and competing for nutrients [108].

In addition, probiotics may weaken the flagellar motility of intestinal pathogenic microorganisms, thereby preventing their colonization of the intestinal tract. The flagellum is the motility organ of bacteria and foodborne pathogens (e.g., *Salmonella typhi*) need to actively move the flagellum in order to cross the intestinal epithelial cells. Following the use of the flagellar inhibitor Sal4, *Salmonella typhimurium* was found to remain non-invasive in infected mice [109].

Prospect

Due to the increase in antibiotic resistance and the potential harm to human health, many countries have banned the addition of antibiotics to feeds. In the context of the ban on antibiotics in feed, probiotics have

become an alternative to antibiotics and are widely used in pig farming industry. Probiotics have the effect of regulating intestinal flora, improving immunity and intestinal mucosal barrier function, inhibiting the growth of pathogenic microorganisms, etc., Therefore, they can improve the immunity of pigs and are beneficial to the prevention and treatment of swine diseases. However, there are some issues regarding the application of probiotics that need to be addressed. Firstly, the effect of probiotics depends on the specific strain, dose and environment, as well as the host specificity, so the results of different studies may vary when evaluating probiotics. Secondly, although the good adhesion of probiotics is beneficial to its action on the intestinal mucosa, it is also possible to increase the movement of bacteria and make animals sick. Thirdly, the possibility of probiotics used in animal feed entering the human food chain cannot be completely excluded. Although probiotics have been used in the pig farming industry for many years, the mechanism of probiotics in the prevention and treatment of swine diseases and the problems existing in the application of probiotics still need to be further investigated in order to play a greater role in the pig farming industry.

Abbreviations

DON	Deoxynivalenol
PWD	Post-weaning diarrhea
PRRSV	Porcine reproductive and respiratory syndrome virus
SIV	Swine influenza virus
PCV2	Porcine circovirus type 2
PEDV	Porcine epidemic diarrhea virus
IgG	Immunoglobulin G
IgA	Immunoglobulin A
sIgA	Secretory immunoglobulin A
ETEC	Enterotoxin-producing <i>Escherichia coli</i>
TJs	Tight junctions

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Author contributions

ZY, Z-YY, and YJ contributed to the conception and overall idea of the study and completed the manuscript. ZH, R-SF, G-LH, CZ, and NH helped in searching for related articles. LF, M-YW, Z-YM, and W-JQ revised the manuscript. All authors contributed to the article and finally the submitted version is approved by YJ. All authors read and approved the final version of the manuscript.

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References

- Soare E, Chirciu I-A. Study on the pork market worldwide. *Sci Papers Ser Manag Econ Eng Agric Rural Dev*. 2017;17(4):321–6.
- Halasa T, Bøtner A, Mortensen S, Christensen H, Toft N, Boklund A. Simulating the epidemiological and economic effects of an African swine fever epidemic in industrialized swine populations. *Vet Microbiol*. 2016;193:7–16.
- Zhao D, Liu R, Zhang X, Li F, Wang J, Zhang J, et al. Replication and virulence in pigs of the first African swine fever virus isolated in China. *Emerg Microbes Infect*. 2019;8(1):438–47.
- Allen HK, Levine UY, Looft T, Bandrick M, Casey TA. Treatment, promotion, commotion: antibiotic alternatives in food-producing animals. *Trends Microbiol*. 2013;21(3):114–9.
- Patil A, Kumar S, Verma A, Baghel R. Probiotics as feed additives in weaned pigs: a review. *Livest Res Int*. 2015;3(2):31–9.
- Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol*. 2014;11(8):506–14.
- Sánchez B, Delgado S, Blanco-Míguez A, Lourenço A, Gueimonde M, Margolles A. Probiotics, gut microbiota, and their influence on host health and disease. *Mol Nutr Food Res*. 2017;61(1):1600240.
- Gaggia F, Mattarelli P, Biavati B. Probiotics and prebiotics in animal feeding for safe food production. *Int J Food Microbiol*. 2010;141(Suppl 1):15–28.
- Musa HH, Wu S, Zhu C, Seri H, Zhu G. The potential benefits of probiotics in animal production and health. *J Anim Vet Adv*. 2009;8(2):313–21.
- Wu Z, Yang K, Zhang A, Chang W, Zheng A, Chen Z, et al. Effects of *Lactobacillus acidophilus* on the growth performance, immune response, and intestinal barrier function of broiler chickens challenged with *Escherichia coli* O157. *Poult Sci*. 2021;100(9):101323.
- Bai Y, Ma K, Li J, Li J, Bi C, Shan A. Deoxynivalenol exposure induces liver damage in mice: inflammation and immune responses, oxidative stress, and protective effects of *Lactobacillus rhamnosus* GG. *Food Chem Toxicol*. 2021;156:112514.
- Gaggia F, Mattarelli P, Biavati B. Probiotics and prebiotics in animal feeding for safe food production. *Int J Food Microbiol*. 2010;141:15–28.
- Afonso ER, Parazzi LJ, Marino CT, Martins SMMK, Araújo LF, Araújo CSdS, et al. Probiotics association in the suckling and nursery in piglets challenged with *Salmonella typhimurium*. *Braz Arch Biol Technol*. 2013;56:249–58.
- Jungersen M, Wind A, Johansen E, Christensen JE, Stuer-Lauridsen B, Eskesen D. The science behind the probiotic strain *Bifidobacterium animalis* subsp. *Lactis* BB-12[®]. *Microorganisms*. 2014;2(2):92–110.
- Barba-Vidal E, Castillejos L, Roll VF, Cifuentes-Orjuela G, Moreno Munoz JA, Martín-Orúe SM. The probiotic combination of *Bifidobacterium longum* subsp. *Infantis* CECT 7210 and *Bifidobacterium animalis* subsp. *Lactis* BPL6 reduces pathogen loads and improves gut health of weaned piglets orally challenged with *Salmonella Typhimurium*. *Front Microbiol*. 2017;8:1570.

16. Barbosa TM, Serra CR, La Ragione RM, Woodward MJ, Henriques AO. Screening for *Bacillus* isolates in the broiler gastrointestinal tract. *Appl Environ Microbiol.* 2005;71(2):968–78.
17. Spinosa MR, Braccini T, Ricca E, De Felice M, Morelli L, Pozzi G, et al. On the fate of ingested *Bacillus spores*. *Res Microbiol.* 2000;151(5):361–8.
18. Mingmongkolchai S, Panbangred W. *Bacillus probiotics*: an alternative to antibiotics for livestock production. *J Appl Microbiol.* 2018;124(6):1334–46.
19. Behnamifar A, Rahimi S, Kiaei M, Fayazi H. Comparison of the effect of probiotic, prebiotic, salinomycin and vaccine in control of coccidiosis in broiler chickens. *Iran J Vet Res.* 2019;20(1):51.
20. Tsukahara T, Inoue R, Nakayama K, Inatomi T. Inclusion of *Bacillus amyloliquefaciens* strain TOA 5001 in the diet of broilers suppresses the symptoms of coccidiosis by modulating intestinal microbiota. *Anim Sci J.* 2018;89(4):679–87.
21. Chaudhari AA, Lee Y, Lillehoj HS. Beneficial effects of dietary supplementation of *Bacillus strains* on growth performance and gut health in chickens with mixed coccidiosis infection. *Vet Parasitol.* 2020;277:109009.
22. Memon FU, Yang Y, Zhang G, Leghari IH, Lv F, Wang Y, et al. Chicken gut microbiota responses to dietary *Bacillus subtilis* probiotic in the presence and absence of eimeria infection. *Microorganisms.* 2022;10(8):1548.
23. König H, Fröhlich J. Lactic acid bacteria. In: König H, Uden G, Fröhlich J, editors. *Biology of Microorganisms on Grapes, in Must and in Wine*. London: Springer; 2017. p. 3–41.
24. Missotten JA, Michiels J, Degroote J, De Smet S. Fermented liquid feed for pigs: an ancient technique for the future. *J Anim Sci Biotechnol.* 2015;6(1):1–9.
25. Vohra A, Syal P, Madan A. Probiotic yeasts in livestock sector. *Anim Feed Sci Technol.* 2016;219:31–47.
26. Secher T, Kassem S, Benamar M, Bernard I, Boury M, Barreau F, et al. Oral administration of the probiotic strain *Escherichia coli* Nissle 1917 reduces susceptibility to neuroinflammation and repairs experimental autoimmune encephalomyelitis-induced intestinal barrier dysfunction. *Front Immunol.* 2017;8:1096.
27. Hu Y, Dun Y, Li S, Zhang D, Peng N, Zhao S, et al. Dietary *Enterococcus faecalis* LAB31 improves growth performance, reduces diarrhea, and increases fecal *Lactobacillus* number of weaned piglets. *PLoS ONE.* 2015;10(1):e0116635.
28. Allameh SK, Ringø E, Yusoff F, Daud H, Ideris A. Dietary supplement of *Enterococcus faecalis* on digestive enzyme activities, short-chain fatty acid production, immune system response and disease resistance of Javanese carp (*Puntius gonionotus*, Bleeker 1850). *Aquacult Nutr.* 2017;23(2):331–8.
29. Huang L, Luo L, Zhang Y, Wang Z, Xia Z. Effects of the dietary probiotic, *Enterococcus faecium* NCIMB11181, on the intestinal barrier and system immune status in *Escherichia coli* O78-challenged broiler chickens. *Probiotics Antimicrob Proteins.* 2019;11(3):946–56.
30. Wu Y, Zhen W, Geng Y, Wang Z, Guo Y. Effects of dietary *Enterococcus faecium* NCIMB 11181 supplementation on growth performance and cellular and humoral immune responses in broiler chickens. *Poult Sci.* 2019;98(1):150–63.
31. Sorroza L, Real F, Acosta F, Acosta B, Déniz S, Román L, et al. A probiotic potential of *Enterococcus gallinarum* against *Vibrio anguillarum* infection. *Fish Pathol.* 2013;48(1):9–12.
32. Safari R, Adel M, Lazado CC, Caiyang CMA, Dadar M. Host-derived probiotics *Enterococcus casseliflavus* improves resistance against *Streptococcus iniae* infection in rainbow trout (*Oncorhynchus mykiss*) via immunomodulation. *Fish Shellfish Immunol.* 2016;52:198–205.
33. Feng J, Liu X, Xu Z, Lu Y, Liu Y. The effect of *Aspergillus oryzae* fermented soybean meal on growth performance, digestibility of dietary components and activities of intestinal enzymes in weaned piglets. *Anim Feed Sci Technol.* 2007;134(3–4):295–303.
34. Zahirian M, Seidavi A, Solka M, Nosrati M, Corazzin M. Dietary supplementation of *Aspergillus oryzae* meal and its effect on performance, carcass characteristics, blood variables, and immunity of broiler chickens. *Trop Anim Health Prod.* 2019;51(8):2263–8.
35. Dawood MA, Eweedah NM, Moustafa Moustafa E, Shahin MG. Effects of feeding regimen of dietary *Aspergillus oryzae* on the growth performance, intestinal morphometry and blood profile of *Nile tilapia* (*Oreochromis niloticus*). *Aquacult Nutr.* 2019;25(5):1063–72.
36. Dang DX, Liu Y, Chen N, Kim IH. Dietary supplementation of *Aspergillus niger*-expressed glucose oxidase ameliorates weaning stress and improves growth performance in weaning pigs. *J Anim Physiol Anim Nutr.* 2022;106(2):258–65.
37. Balcázar JL, Vendrell D, De Blas I, Ruiz-Zarzuela I, Múzquiz JL. Effect of *Lactococcus lactis* CLFP 100 and *Leuconostoc mesenteroides* CLFP 196 on *Aeromonas salmonicida* infection in brown trout (*Salmo trutta*). *Microb Physiol.* 2009;17(3):153–7.
38. Keimer B, Pieper R, Simon A, Zentek J. Effect of time and dietary supplementation with processed yeasts (*Kluyveromyces fragilis*) on immunological parameters in weaned piglets. *Anim Feed Sci Technol.* 2018;245:136–46.
39. Wang W, Li Z, Gan L, Fan H, Guo Y. Dietary supplemental *Kluyveromyces marxianus* alters the serum metabolite profile in broiler chickens. *Food Funct.* 2018;9(7):3776–87.
40. Hossain M, Begum M, Kim I. Effect of *Leuconostoc mesenteroides* KCCM35046 fermented aged garlic extract on egg production, egg quality, odour gas emissions, targeted *E. coli* colony, haematological characteristics and fatty acids composition of egg yolk in laying hens. *J Appl Anim Res.* 2016;44(1):458–65.
41. Al-Hisnawi A, Rodiles A, Rawling MD, Castex M, Waines P, Gioacchini G, et al. Dietary probiotic *Pediococcus acidilactici* MA18/5 M modulates the intestinal microbiota and stimulates intestinal immunity in rainbow trout (*Oncorhynchus mykiss*). *J World Aquaculture Soc.* 2019;50(6):1133–51.
42. Ahmadi E, Dawood MA, Moghadam MS, Shahrestanaki AH, Van Doan H, Saad AH, et al. The effect of *Pediococcus acidilactici* MA 18/5 M on immune responses and mRNA levels of growth, antioxidant and immune-related genes in zebrafish (*Danio rerio*). *Aquaculture Rep.* 2020;17:100374.
43. Additives EPo F, PoSuiA. Scientific opinion on the safety and efficacy of Probiotic LACTINA® (*Lactobacillus acidophilus*, *Lactobacillus helveticus*, *Lactobacillus bulgaricus*, *Lactobacillus lactis*, *Streptococcus thermophilus* and *Enterococcus faecium*) for chickens for fattening and piglets. *EFSA J.* 2013;11(4):3170.
44. Swain SM, Singh C, Arul V. Inhibitory activity of probiotics *Streptococcus pneumoniae* P180 and *Enterococcus faecium* MC13 against vibriosis in shrimp *Penaeus monodon*. *World J Microbiol Biotechnol.* 2009;25(4):697–703.
45. Pereira WA, Franco SM, Reis IL, Mendonça CM, Piazzentin AC, Azevedo PO, et al. Beneficial effects of probiotics on the pig production cycle: an overview of clinical impacts and performance. *Vet Microbiol.* 2022;269:109431.
46. Opriessnig T, Gimenez-Lirola LG, Halbur PG. Polymicrobial respiratory disease in pigs. *Anim Health Res Rev.* 2011;12(90):133–48.
47. Brockmeier SL, Halbur PG, Thacker EL. Porcine respiratory disease complex. In: Brogden KA, Guthmiller JM, editors. *Polymicrobial diseases*. Washington, DC: ASM Press; 2002. p. 231–58.
48. Ji W, Zhang X, Niu G, Chen S, Li X, Yang L, et al. Expression and immunogenicity analysis of the capsid proteins of porcine circovirus types 2 to 4. *Int J Biol Macromol.* 2022;218:828–38.
49. Zhang S, Mou C, Cao Y, Zhang E, Yang Q. Immune response in piglets orally immunized with recombinant *Bacillus subtilis* expressing the capsid protein of porcine circovirus type 2. *Cell Commun Signal.* 2020;18(47):23.
50. Wang Y, Wang J, Zhou M, Liu P, Zhang E, Li Y, et al. Mucosal and systemic immune responses induced by intranasal immunization of recombinant *Bacillus subtilis* expressing the P97R1, P46 antigens of *Mycoplasma hyopneumoniae*. *Biosci Rep.* 2019;39(10):BSR20191126.
51. Pietrzak B, Tomela K, Olejnik-Schmidt A, Mackiewicz A, Schmidt M. Secretory IgA in intestinal mucosal secretions as an adaptive barrier against microbial cells. *Int J Mol Sci.* 2020;21(23):9254.
52. Zuckermann FA, Husmann R, Chen W, Roady P, Pfeiff J, Leistikow KR, et al. Bacillus-based direct-fed microbial reduces the pathogenic synergy of a coinfection with *Salmonella enterica* Serovar Choleraesuis and porcine reproductive and respiratory syndrome virus. *Infect Immun.* 2022;90(4):e00574–21.
53. Pluske JR, Turpin DL, Kim J-C. Gastrointestinal tract (gut) health in the young pig. *Anim Nutr.* 2018;4(2):187–96.

54. Luppi A, Gibellini M, Gin T, Vangroenweghe F, Vandenbroucke V, Bauerfeind R, et al. Prevalence of virulence factors in enterotoxigenic *Escherichia coli* isolated from pigs with post-weaning diarrhoea in Europe. *Porcine Health Manag*. 2016;2:20.
55. Sun Y, Duarte ME, Kim SW. Dietary inclusion of multispecies probiotics to reduce the severity of post-weaning diarrhea caused by *Escherichia coli* F18(+) in pigs. *Anim Nutr*. 2021;7(67):326–33.
56. Du Y, Xu Z, Yu G, Liu W, Zhou Q, Yang D, et al. A newly isolated *Bacillus subtilis* strain named WS-1 inhibited diarrhea and death caused by pathogenic *Escherichia coli* in newborn piglets. *Front Microbiol*. 2019;10(93):1248.
57. Wang Y, Gong L, Wu Y-p, Cui Z-w, Wang Y-q, Huang Y, et al. Oral administration of *Lactobacillus rhamnosus* GG to newborn piglets augments gut barrier function in pre-weaning piglets. *J Zhejiang Univ Sci B*. 2019;20(2):180–92.
58. Huang S, Yu Q, Xie L, Ran L, Wang K, Yang Y, et al. Inhibitory effects of *Lactobacillus plantarum* metabolites on porcine epidemic diarrhea virus replication. *Res Vet Sci*. 2021;139(68):32–42.
59. Wang X, Tian Z, Azad MAK, Zhang W, Blachier F, Wang Z, et al. Dietary supplementation with *Bacillus* mixture modifies the intestinal ecosystem of weaned piglets in an overall beneficial way. *J Appl Microbiol*. 2021;130(59):233–46.
60. Rist VT, Weiss E, Sauer N, Mosenthin R, Eklund M. Effect of dietary protein supply originating from soybean meal or casein on the intestinal microbiota of piglets. *Anaerobe*. 2014;25:72–9.
61. Le M, Galle S, Yang Y, Landero J, Beltranena E, Gänzle M, et al. Effects of feeding fermented wheat with *Lactobacillus reuteri* on gut morphology, intestinal fermentation, nutrient digestibility, and growth performance in weaned pigs. *J Anim Sci*. 2016;94(11):4677–87.
62. Cheng Y-H, Su L-W, Horng Y-B, Yu Y-H. Effects of soybean meal fermented by *Lactobacillus* species and *Clostridium butyricum* on growth performance, diarrhea incidence, and fecal bacteria in weaning piglets. *Ann Anim Sci*. 2019;19(99):1051–62.
63. Awais MM, Jamal MA, Akhtar M, Hameed MR, Anwar MI, Ullah MI. Immunomodulatory and ameliorative effects of *Lactobacillus* and *Saccharomyces* based probiotics on pathological effects of eimeriasis in broilers. *Microb Pathog*. 2019;126:101–8.
64. Chen H-L, Zhao X-Y, Zhao G-X, Huang H-B, Li H-R, Shi C-W, et al. Dissection of the cecal microbial community in chickens after *Eimeria tenella* infection. *Parasites Vectors*. 2020;13(1):1–15.
65. Madlala T, Okpeku M, Adeleke MA. Understanding the interactions between *Eimeria* infection and gut microbiota, towards the control of chicken coccidiosis: a review. *Parasite*. 2021;28:48.
66. Wang X, Farnell YZ, Kiess AS, Peebles ED, Wamsley KG, Zhai W. Effects of *Bacillus subtilis* and coccidial vaccination on cecal microbial diversity and composition of *Eimeria*-challenged male broilers. *Poult Sci*. 2019;98(9):3839–49.
67. Ding H, Zhao X, Azad MAK, Ma C, Gao Q, He J, et al. Dietary supplementation with *Bacillus subtilis* and xylo-oligosaccharides improves growth performance and intestinal morphology and alters intestinal microbiota and metabolites in weaned piglets. *Food Funct*. 2021;12(58):5837–49.
68. Xu J, Li Y, Yang Z, Li C, Liang H, Wu Z, et al. Yeast probiotics shape the gut microbiome and improve the health of early-weaned piglets. *Front Microbiol*. 2018;9:2011.
69. Thomas F, Hehemann J-H, Rebuffet E, Czjzek M, Michel G. Environmental and gut bacteroidetes: the food connection. *Front Microbiol*. 2011;2:93.
70. Fukuda S, Toh H, Hase K, Oshima K, Nakanishi Y, Yoshimura K, et al. Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature*. 2011;469(7331):543–7.
71. Long S, He T, Kim SW, Shang Q, Kiros T, Mahfuz SU, et al. Live yeast or live yeast combined with zinc oxide enhanced growth performance, antioxidative capacity, immunoglobulins and gut health in nursery pigs. *Anim (Basel)*. 2021;11:62.
72. Kogan G, Kocher A. Role of yeast cell wall polysaccharides in pig nutrition and health protection. *Livest Sci*. 2007;109(1–3):161–5.
73. Liu M, Liu W, Zhang W, Yao J, Mo X. Ultrasound-assisted extraction of boulardii yeast cell wall polysaccharides: characterization and its biological functions on early-weaned lambs. *Food Sci Nutr*. 2021;9(7):3617–30.
74. Li A, Jiang X, Wang Y, Zhang L, Zhang H, Mehmood K, et al. The impact of *Bacillus subtilis* 18 isolated from Tibetan yaks on growth performance and gut microbial community in mice. *Microb Pathog*. 2019;128:153–61.
75. Xu Z, Hu C, Xia M, Zhan X, Wang M. Effects of dietary fructooligosaccharide on digestive enzyme activities, intestinal microflora and morphology of male broilers. *Poult Sci*. 2003;82(6):1030–6.
76. Yang K, Jiang Z, Zheng C, Wang L, Yang X. Effect of *Lactobacillus plantarum* on diarrhea and intestinal barrier function of young piglets challenged with enterotoxigenic *Escherichia coli* K88. *J Anim Sci*. 2014;92(4):1496–503.
77. Liu P, Che L, Yang Z, Feng B, Che L, Xu S, et al. A maternal high-energy diet promotes intestinal development and intrauterine growth of offspring. *Nutrients*. 2016;8(5):258.
78. Fu R, Liang C, Chen D, Yan H, Tian G, Zheng P, et al. Effects of dietary *Bacillus coagulans* and yeast hydrolysate supplementation on growth performance, immune response and intestinal barrier function in weaned piglets. *J Anim Physiol Anim Nutr (Berl)*. 2021;105(87):898–907.
79. Che L, Hu L, Liu Y, Yan C, Peng X, Xu Q, et al. Dietary nucleotides supplementation improves the intestinal development and immune function of neonates with intra-uterine growth restriction in a pig model. *PLoS ONE*. 2016;11(6):e0157314.
80. Liu Z, Shen T, Zhang P, Ma Y, Qin H. *Lactobacillus plantarum* surface layer adhesive protein protects intestinal epithelial cells against tight junction injury induced by enteropathogenic *Escherichia coli*. *Mol Biol Rep*. 2011;38(5):3471–80.
81. Du W, Xu H, Mei X, Cao X, Gong L, Wu Y, et al. Probiotic *Bacillus* enhance the intestinal epithelial cell barrier and immune function of piglets. *Benef Microb*. 2018;9(5):743–54.
82. Fasano A. Zonulin and its regulation of intestinal barrier function: the biological door to inflammation, autoimmunity, and cancer. *Physiol Rev*. 2011;91:151–75.
83. Ukena SN, Singh A, Dringenberg U, Engelhardt R, Seidler U, Hansen W, et al. Probiotic *Escherichia coli* Nissle 1917 inhibits leaky gut by enhancing mucosal integrity. *PLoS ONE*. 2007;2(12):e1308.
84. Johnson-Henry K, Donato K, Shen-Tu G, Gordanpour M, Sherman P. *Lactobacillus rhamnosus* strain GG prevents enterohemorrhagic *Escherichia coli* O157:H7-induced changes in epithelial barrier function. *Infect Immun*. 2008;76(4):1340–8.
85. Cotter PD, Ross RP, Hill C. Bacteriocins—A viable alternative to antibiotics? *Nat Rev Microbiol*. 2013;11(2):95–105.
86. Bierbaum G, Sahl H-G. Lantibiotics: mode of action, biosynthesis and bioengineering. *Curr Pharm Biotechnol*. 2009;10(1):2–18.
87. Servin AL. Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *FEMS Microbiol Rev*. 2004;28(4):405–40.
88. De Keersmaecker SC, Verhoeven TL, Desair J, Marchal K, Vanderleyden J, Nagy I. Strong antimicrobial activity of *Lactobacillus rhamnosus* GG against *Salmonella typhimurium* is due to accumulation of lactic acid. *FEMS Microbiol Lett*. 2006;259(1):89–96.
89. Durant JA, Corrier DE, Ricke SC. Short-chain volatile fatty acids modulate the expression of the hilA and invF genes of *Salmonella typhimurium*. *J Food Prot*. 2000;63(5):573–8.
90. Knaus UG, Hertzberger R, Pircalabioru GG, Yousefi SPM, Branco dos Santos F. Pathogen control at the intestinal mucosa—H2O2 to the rescue. *Gut Microb*. 2017;8(1):67–74.
91. Duarte C, Gudiña EJ, Lima CF, Rodrigues LR. Effects of biosurfactants on the viability and proliferation of human breast cancer cells. *AMB Exp*. 2014;4(1):1–12.
92. Sharma D, Singh Saharan B. Simultaneous production of biosurfactants and bacteriocins by probiotic *Lactobacillus casei* MRTL3. *Int J Microbiol*. 2014;2014:698713.
93. Mohamad N, Amal MNA, Yasin ISM, Saad MZ, Nasruddin NS, Al-saari N, et al. Vibriosis in cultured marine fishes: a review. *Aquaculture*. 2019;512:734289.
94. Ina-Salwany M, Al-saari N, Mohamad A, Mursidi FA, Mohd-Aris A, Amal M, et al. Vibriosis in fish: a review on disease development and prevention. *J Aquat Anim Health*. 2019;31(1):3–22.
95. Ghanei-Motlagh R, Gharibi D, Mohammadian T, Khosravi M, Mahmoudi E, Zarea M, et al. Feed supplementation with quorum quenching probiotics with anti-virulence potential improved innate immune responses,

- antioxidant capacity and disease resistance in asian seabass (*Lates calcarifer*). *Aquaculture*. 2021;535:736345.
96. Zhang Q, Yu H, Tong T, Tong W, Dong L, Xu M, et al. Dietary supplementation of *Bacillus subtilis* and fructooligosaccharide enhance the growth, non-specific immunity of juvenile ovate pompano, *Trachinotus ovatus* and its disease resistance against *Vibrio vulnificus*. *Fish Shellfish Immunol*. 2014;38(1):7–14.
 97. Lee S, Katya K, Park Y, Won S, Seong M, Bai SC. Comparative evaluation of dietary probiotics *Bacillus subtilis* WB60 and *Lactobacillus plantarum* KCTC3928 on the growth performance, immunological parameters, gut morphology and disease resistance in Japanese eel, *Anguilla japonica*. *Fish Shellfish Immunol*. 2017;61:201–10.
 98. Ahmed ST, Hoon J, Mun H-S, Yang C-J. Evaluation of *Lactobacillus* and *Bacillus*-based probiotics as alternatives to antibiotics in enteric microbial challenged weaned piglets. *Afr J Microbiol Res*. 2014;8(1):96–104.
 99. Xin J, Zeng D, Wang H, Sun N, Zhao Y, Dan Y, et al. Probiotic *Lactobacillus johnsonii* BS15 promotes growth performance, intestinal immunity, and gut microbiota in piglets. *Probiotics Antimicrob Proteins*. 2020;12(1):184–93.
 100. Cao X, Tang L, Zeng Z, Wang B, Zhou Y, Wang Q, et al. Effects of probiotics BaSC06 on intestinal digestion and absorption, antioxidant capacity, microbiota composition, and macrophage polarization in pigs for fattening. *Front Vet Sci*. 2020;7:570593.
 101. Wang S, Yao B, Gao H, Zang J, Tao S, Zhang S, et al. Combined supplementation of *Lactobacillus fermentum* and *Pediococcus acidilactici* promoted growth performance, alleviated inflammation, and modulated intestinal microbiota in weaned pigs. *BMC Vet Res*. 2019;15(1):1–11.
 102. Wilson A, Stokes C, Bourne F. Morphology and functional characteristics of isolated porcine intraepithelial lymphocytes. *Immunology*. 1986;59(1):109.
 103. Brisbin JT, Zhou H, Gong J, Sabour P, Akbari MR, Haghghi HR, et al. Gene expression profiling of chicken lymphoid cells after treatment with *Lactobacillus acidophilus* cellular components. *Dev Comp Immunol*. 2008;32(5):563–74.
 104. Huang L, Wang J, Wang Y, Zhang E, Li Y, Yu Q, et al. Upregulation of CD4+ CD8+ memory cells in the piglet intestine following oral administration of *Bacillus subtilis* spores combined with PEDV whole inactivated virus. *Vet Microbiol*. 2019;235:1–9.
 105. Fonseca HC, de Sousa Melo D, Ramos CL, Dias DR, Schwan RF. Probiotic properties of lactobacilli and their ability to inhibit the adhesion of enteropathogenic bacteria to Caco-2 and HT-29 cells. *Probiotics Antimicrob Proteins*. 2021;13(1):102–12.
 106. Wang J, Zeng Y, Wang S, Liu H, Zhang D, Zhang W, et al. Swine-derived probiotic *Lactobacillus plantarum* inhibits growth and adhesion of enterotoxigenic *Escherichia coli* and mediates host defense. *Front Microbiol*. 2018;9(92):1364.
 107. Hou C, Zeng X, Yang F, Liu H, Qiao S. Study and use of the probiotic *Lactobacillus reuteri* in pigs: a review. *J Anim Sci Biotechnol*. 2015;6(1):1–8.
 108. Dicks L, Botes M. Probiotic lactic acid bacteria in the gastro-intestinal tract: health benefits, safety and mode of action. *Benef Microb*. 2010;1(1):11–29.
 109. Forbes SJ, Eschmann M, Mantis NJ. Inhibition of *Salmonella enterica* serovar typhimurium motility and entry into epithelial cells by a protective antilipopolysaccharide monoclonal immunoglobulin a antibody. *Infect Immun*. 2008;76(9):4137–44.
 110. Yi H, Wang L, Xiong Y, Wen X, Wang Z, Yang X, et al. Effects of *Lactobacillus reuteri* LR1 on the growth performance, intestinal morphology, and intestinal barrier function in weaned pigs. *J Anim Sci*. 2018;96(6):2342–51.
 111. He T, Zhu Y-H, Yu J, Xia B, Liu X, Yang G-Y, et al. *Lactobacillus johnsonii* L531 reduces pathogen load and helps maintain short-chain fatty acid levels in the intestines of pigs challenged with *Salmonella enterica* infantis. *Vet Microbiol*. 2019;230:187–94.
 112. Inatomi T, Amatatsu M, Romero-Perez GA, Inoue R, Tsukahara T. Dietary probiotic compound improves reproductive performance of porcine epidemic diarrhea virus-infected sows reared in a Japanese commercial swine farm under vaccine control condition. *Front Immunol*. 2017;8:1877.
 113. Li H-H, Jiang X-R, Qiao J-Y. Effect of dietary *Bacillus subtilis* on growth performance and serum biochemical and immune indexes in weaned piglets. *J Appl Anim Res*. 2021;49(1):83–8.
 114. Li Y, Hou S, Peng W, Lin Q, Chen F, Yang L, et al. Oral administration of *Lactobacillus delbrueckii* during the suckling phase improves antioxidant activities and immune responses after the weaning event in a piglet model. *Oxid Med Cell Longev*. 2019;2019:6919803.
 115. Zhao D, Wu T, Yi D, Wang L, Li P, Zhang J, et al. Dietary supplementation with *Lactobacillus casei* alleviates lipopolysaccharide-induced liver injury in a porcine model. *Int J Mol Sci*. 2017;18(12):2535.
 116. Zhang W, Wu Q, Zhu Y, Yang G, Yu J, Wang J, et al. Probiotic *Lactobacillus rhamnosus* GG induces alterations in ileal microbiota with associated CD3-CD19-T-bet + IFN γ +/-Cell subset homeostasis in pigs challenged with *Salmonella enterica* Serovar 4,[5], 12: i. *Front Microbiol*. 2019;10:977.
 117. Ding H, Zhao X, Ma C, Gao Q, Yin Y, Kong X, et al. Dietary supplementation with *Bacillus subtilis* DSM 32315 alters the intestinal microbiota and metabolites in weaned piglets. *J Appl Microbiol*. 2021;130(86):217–32.
 118. Zhang J, Chen X, Liu P, Zhao J, Sun J, Guan W, et al. Dietary *Clostridium butyricum* induces a phased shift in fecal microbiota structure and increases the acetic acid-producing bacteria in a weaned piglet model. *J Agric Food Chem*. 2018;66(20):5157–66.

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