



Article

Investigation of *Cannabis sativa* Phytochemicals as Anti-Alzheimer's Agents: An In Silico Study

Nil Patil ^{1,2} , Vaishnavi Chandel ¹, Aarzu Rana ¹, Mukul Jain ^{1,2,*} and Prashant Kaushik ^{3,*} 

¹ Department of Life Sciences, Parul Institute of Applied Sciences, Parul University, Vadodara 391760, Gujarat, India

² Laboratory 209, Cell & Developmental Biology Laboratory, Centre of Research for Development, Parul University, Vadodara 391760, Gujarat, India

³ Instituto de Conservación y Mejora de la Agrodiversidad Valenciana, Universitat Politècnica de València, 46022 Valencia, Spain

* Correspondence: mukul.jain19640@paruluniversity.ac.in (M.J.); prakau@doctor.upv.es (P.K.)

Abstract: *Cannabis sativa* is a medicinal plant that has been known for years and is used as an Ayurvedic medicine. This plant has great potential in treating various types of brain diseases. Phytochemicals present in this plant act as antioxidants by maintaining synaptic plasticity and preventing neuronal loss. Cannabidiol (CBD) and Tetrahydrocannabinol (THC) are both beneficial in treating Alzheimer's disease by increasing the solubility of A β 42 amyloid and Tau aggregation. Apart from these therapeutic effects, there are certain unknown functions of these phytochemicals in Alzheimer's disease that we want to elucidate through this study. In this research, our approach is to analyze the effect of phytochemicals in *Cannabis sativa* on multiple culprit enzymes in Alzheimer's disease, such as AChE (Acetylcholinesterase), BChE (Butyrylcholinesterase), γ -secretase, and BACE-1. In this study, the compounds were selected by Lipinski's rule, ADMET, and ProTox based on toxicity. Molecular docking between the selected compounds (THCV, Cannabinol C2, and Cannabidiol) and enzymes mentioned above was obtained by various software programs including AutoDock Vina 4.2, AutoDock, and iGEMDOCK. In comparison to Donepezil (BA = -8.4 kcal/mol, Ki = 1.46 mM), Rivastigmine (BA = -7.0 kcal/mol, Ki = 0.02 mM), and Galantamine (BA = -7.1, Ki = 2.1 mM), Cannabidiol (BA = -9.4 kcal/mol, Ki = 4.61 mM) shows significant inhibition of AChE. On the other hand, Cannabinol C2 (BA = -9.2 kcal/mol, Ki = 4.32 mM) significantly inhibits Butyrylcholinesterase (BuChE) in comparison to Memantine (BA = -6.8 kcal/mol, Ki = 0.54 mM). This study sheds new light and opens new avenues for elucidating the role of bioactive compounds present in *Cannabis sativa* in treating Alzheimer's disease.

Keywords: Alzheimer's disease; *Cannabis sativa*; AChE; BuChE; AutoDock; drug development



Citation: Patil, N.; Chandel, V.; Rana, A.; Jain, M.; Kaushik, P. Investigation of *Cannabis sativa* Phytochemicals as Anti-Alzheimer's Agents: An In Silico Study. *Plants* **2023**, *12*, 510. <https://doi.org/10.3390/plants12030510>

Academic Editor: Corina Danciu

Received: 4 December 2022

Revised: 17 January 2023

Accepted: 18 January 2023

Published: 22 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Cannabis sativa L. (*C. sativa*) is a dioecious, cross-pollinated, flowering plant that can grow to a height of between one and two meters. It is commonly known as hemp, cannabis, or marijuana [1]. *Cannabis sativa* has long been known to have both therapeutic and psychoactive properties. It is a multipurpose plant that is used for recreational, medicinal, and industrial purposes [2]. Cannabis is currently the focus of a significant amount of research due to its unique phytochemical constituents, i.e., secondary metabolites. The glandular trichomes in the bark, leaves, and especially the leaves of the female plant contain the most potent cannabinoid metabolites, which are known as phytocannabinoids [1,3]. Cannabis consists of a complex mixture of phytochemical compounds made up of flavonoids, terpenoids, cannabinoids [4], alkaloids, glycoproteins [5], and phytosteroids [6]. Over 565 cannabis constituents have been discovered in the cannabis plant [7], and over 150 compounds are considered phytocannabinoids [8–10]. The most investigated cannabinoids are Tetrahydrocannabinol (THC, which includes the two main components 8-THC and 9-THC), a major

psychoactive component of cannabis, Cannabidiol (CBD), and cannabigerol (CBG), which are extracted from the resin formed by the female plants. [11,12].

Seven different classes are used to categorize the additional natural cannabinoids generated from *Cannabis sativa*, including Cannabinol (CBN), Cannabidiol (CBND), Cannabitrinol (CBT), Cannabielsoin (CBE), Cannabichromene (CBC) Cannabicyclol (CBL), and other miscellaneous types [13]. To exert their many beneficial biological effects, Cannabinoids interact with a variety of receptors, including the cannabinoid receptors CB1 and CB2, as well as several additional non-cannabinoid receptors such as G-protein coupled receptors (GPR55, GPR3) and ion channels [14]. It has been suggested that one significant pathway of cannabis is used as an alternative treatment for many diseases, including chronic pain, nausea, multiple sclerosis, schizophrenia, glaucoma, sclerosis multiplex (SM), inflammatory bowel disorders (IBDs), nausea and vomiting, pain, appetite loss, epilepsy, [15] anxiety, Alzheimer's disease, Parkinson's disease, Huntington's disease, and COVID-19. Phytocannabinoids, for example, can bind to these types of GPCRs, as shown in Table 1 [16,17].

Table 1. Major metabolites reported in extracts of *C. sativa* which affects the AD pathway and its medicinal characteristics.

Compound Class & Plant Tissue Type	Name	Effects on AD (In Vivo)	Effects on AD (In Vitro)	Precursor	Medicinal Characteristics [3,18]	References
Neutral cannabinoids (Trichomes, Female flowers, Roots/Apoplast (secretion pathway))	Cannabidiol (CBD)	Male Wistar rats utilise it as a Streptozotocin (STZ)-induced AD model, CBD enhances the brain glucose metabolism. Activation of the PPAR γ via Wnt/ β -catenin pathway	Pretreatment restores the synaptic transmission that was reduced by A β in a C57 mouse hippocampal slice.	CBGA	Anti-fungal and anti-bacterial against methicillin resistant strains, sedative and analgesic potential and anti-epileptic potential	[19–25]
	Tetrahydrocannabinol(THC)	THC reduces the A β burden in 5XFAD/APP mice	Compared to untreated controls, transgenic Tg4-42 mice expressing human A4-42 showed less neuronal death.	CBGA	Psychotropic & psychoactive properties	[20,26–28]
	Cannabichromene (CBC)	CBC (10–75 mgkg ⁻¹ i.p. per day) significantly decreased motor activity in a model of electroshock seizure during the first 10 min interval, but only the maximum dose was beneficial.	in vitro CBC improved the viability of neural stem cells	CBGA	Anti-inflammatory, sedative and analgesic potential	[29–32]
	Cannabidivarin (CBDV)	Inhibits oxytosis and prevents loss of energy in HT22 cells (50% inhibition at 1.1 μ M and 90 nM, respectively), as well as reducing A β toxicity (50% inhibition at 100 nM) and trophic withdrawal (50% inhibition 350 nM);	Prevents oxytosis in HT22 cells (mouse hippocampal cell) MC65 cells (human nerve cell line)	CBGVA	–	[30,33,34]
	Cannabicyclol (CBL)	–	–	CBC	–	[20,33]
	Cannabinol (CBN)	Inhibiting oxytosis and prevent loss of energy in HT22 cells (50% inhibition at 1.1 μ M and 90 nM, respectively), as well as reducing A β toxicity (50% inhibition at 100 nM) and trophic withdrawal (50% inhibition 350 nM);	Along with its capacity to promote the breakdown and clearance of pre-formed A aggregates in MC65 cells at a concentration of 100 nM in HT-22 cells and cortical embryonic E18 neurons	THC	Mild psychoactive potential	[30,33]
	Cannabidiphorol (CBDP)	–	–	–	Antinociceptive,	[35]
	Tetrahydrocannabivarin (THCV)	–	–	CBGVA	Anti-dyskinesia in Parkinson's disease	[35]

Table 1. Cont.

Compound Class & Plant Tissue Type	Name	Effects on AD (In Vivo)	Effects on AD (In Vitro)	Precursor	Medicinal Characteristics [3,18]	References
Flavonols (leaves, stems, seeds/lypophyl nature suggest cellular retention)	Cannabigerol (CBG)	Retains trophic factors present in cortical neurons of rat (effective concentration 50% = 1.5 μ M) and inhibits the oxytosis in nerve cells (HT22) of mouse	Prevents oxytosis in Ht22 cells (mouse hippocampal cell MC65 cells (human nerve cell line)	OLA, GPP	Analgesic, Anti-inflammatory, Anti-Cancer, Psychotropic, Psychoactive	[30,34,36]
	Canniflavin A	–	Exhibits anti-inflammatory activity	Chrysoeriol	–	[32,36,37]

Phytocannabinoids have gained significant attention due to their neuroactive, strong antioxidant, and anti-inflammatory properties. which may potentially treat different neurodegenerative diseases [38]. Cannabidiol (CBD) and 9-Tetrahydrocannabinol (THC) are two phytocannabinoids that associate with the endocannabinoid system (ECS), have antioxidant, anti-inflammatory, and neuroprotective characteristics, and can improve amyloid- β and NFT-related disorders as well as stimulate neurogenesis [39].

There is a need for a therapeutic drug that can fight against the culprit enzymes of Alzheimer's disease (AD), which is pathologically characterized by the extracellular accumulation of aberrant amyloid-peptide peptides into plaques, the hyperphosphorylation of the microtubule-associated protein Tau resulting in the development of neurofibrillary tangles (NFTs) and neuroinflammation [40]. Preclinical research suggests that some cannabinoids, such as Tetrahydrocannabinol (THC) and Cannabidiol (CBD), may have pharmacological activities on the cholinergic system and amyloid-beta aggregation. These events all contribute to the irreversible and progressive neuronal dysfunction and cell death that cause cerebral atrophy [41]. Furthermore, researchers have been looking for new anti-AD drugs made from secretase enzyme inhibitors that specifically target secretases, such as BACE-1 and γ -secretase.

These are the primary enzymes responsible for breaking down amyloid precursor protein (APP) into neurotoxic A β fragments (A β 42) [42–46]. However, the medications employed in this treatment have limitations and adverse effects, such as nausea and vomiting, and have no impact on the processes that may contribute to AD, such as amyloid genesis, reactive oxygen species, and neuro-inflammation (Figure 1) [47–49]. Plant-based therapeutics for AD are the focus of more research because they have fewer side effects than synthetic medications. In this study, we used in silico methods to examine the effects of a panel of cannabinoids on AChE, BuChE, and BACE-1 activities, including CBD, THC, cannabigerol (CBG), cannabigerolic acid (CBGA), cannabibicitran (CBT), cannabidivarin (CBDV), cannabichromene (CBC), and Cannabinol (CBN).

The effectiveness of psychoactive cannabinoids in the treatment of psychiatric manifestations, particularly agitation and aggression from AD, cannot be conclusively determined from the clinical trials examined. Two factors have significantly hampered the achievement of conclusive results: (1) polypragmasia, which is characterized by the use of established or less established psychotropic drugs (other than cannabinoids) to reduce agitation and aggressive behavior in patients; and (2) a large number of concomitant symptoms, such as pain (which frequently causes anxiety and agitation) [50].

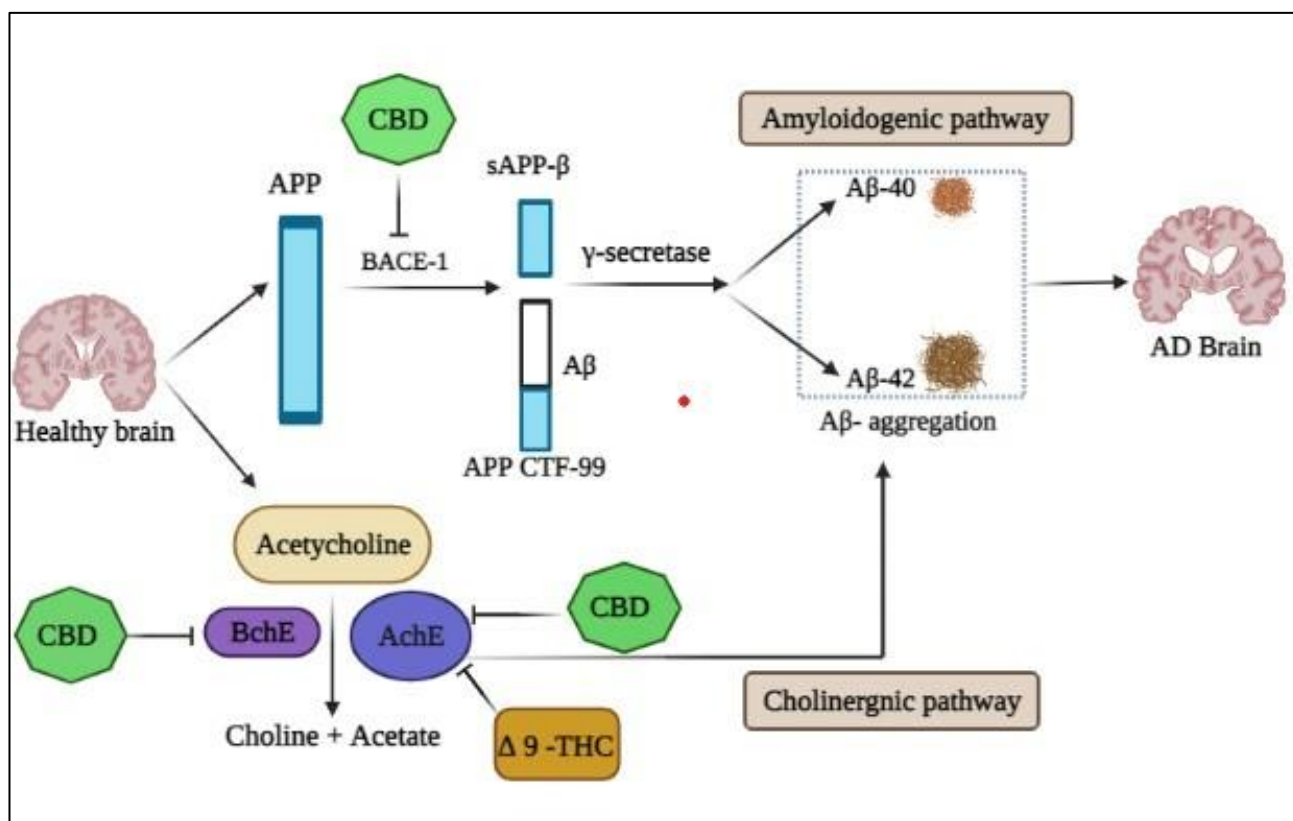


Figure 1. Inhibitory activity of Cannabis constituents on key pathways of Alzheimer's disease. Δ -9-THC and CBD inhibits AChE and BChE. CBD also inhibits amyloid beta aggregate formation by inhibiting BACE-1.

2. Results

2.1. Drug Likelihood Properties Analysis

Drug development is an expensive and drawn-out process. With the aid of computational methods, it is now easier to predict the elements that define a compound's therapeutic potential. Drug-relevant parameters include CLogP, solubility, molecular weight (MW), topological molecular polar surface area (TPSA), etc. The OSIRIS Property Explorer was used to forecast these. OSIRIS accurately predicted the toxicity for 58.66% of compounds that were not harmful and 41.3% of compounds from 75 compounds that were potentially harmful. Lipinski's rule of five is often employed by pharmaceutical chemists in drug design and development to determine the oral bioavailability of possible lead or therapeutic compounds (Supplementary Table S1). The Lipinski rule, Veber rule, Ghose filter, lead likeness, and PAINS (Pan Assay Interference Compounds) were followed by 58.6% of the compounds (44 compounds). Those 44 compounds were further analyzed for pharmacokinetic properties through the SWISS-ADME server for consideration as a drug.

2.2. ADMET Analysis

SWISS-ADME is an online server that lets users create their own ligands or drug molecules and incorporate SMILES data from PubChem. It includes properties such as lipophilicity, water solubility, Log S, and medicinal chemistry. Using five distinct Lipinski, Ghose, Veber, Egan, and Muegge criteria, the SWISS-ADME section provides the physicochemical characteristics of potential oral medication candidates (Table 2). The *in silico* ADMET prediction of selected compounds such as eucalyptol and Cannabinol C2 shows 93.96 and 96.505 absorption with BBB permeability (0.592), respectively, despite the fact that they were not expected to act on p-glycoprotein (Figure 2). Another essential factor in distribution is distribution volume, which describes how medications are distributed

in different tissues *in vivo*. Two of the most crucial enzyme systems in the liver for drug metabolism are the cytochrome P450s (CYP2D6 and CYP3A4). The findings show that none of the substances tested will be broken down by the cytochrome P450 enzymes. In terms of toxicity, CBD derivatives might not have an AMES or hepatotoxicity profile and might not block the hERG channel.

Table 2. Analysis of Pharmacokinetic properties and Protoxicity of three CBD analogues.

Compound	Mw	HA	HD	Absorption	Lipinski's Rule Violation	Solubility	BBB Permeability	CNS Permeability	CYP2D6	LD50 (mg/kg)	Toxicity Class
Cannabidiol	254.3	2	1	92.851	0	−4.599	0.397	−1.368	NO	800	4
THCV	286.4	2	1	91.821	0	−4.403	0.336	−1.99	NO	482	4
Cannabinol C2	268.4	2	1	93.96	0	−4.834	0.5	−1.32	NO	1310	4

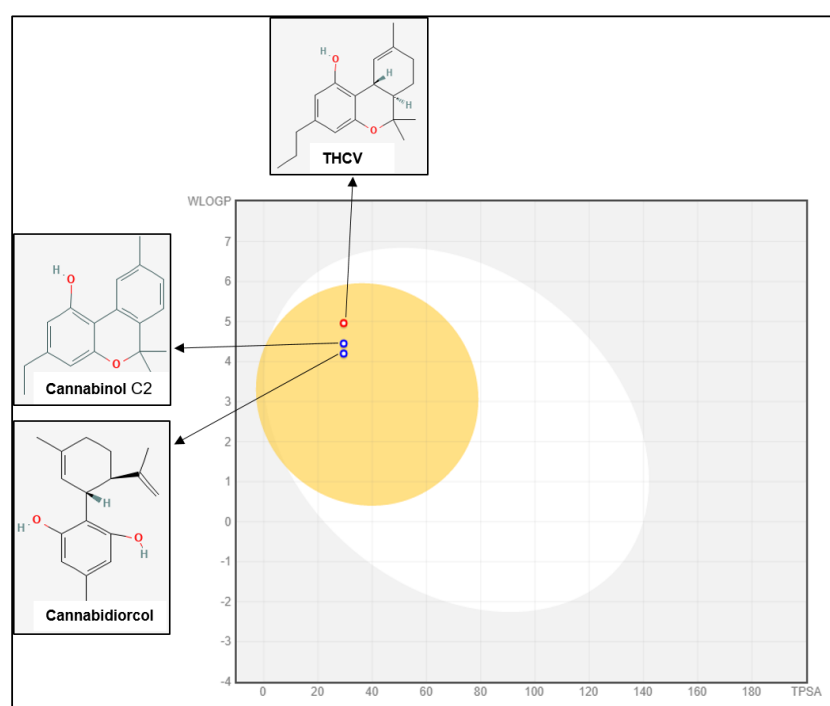


Figure 2. Predicted boiled-egg plot (Swiss ADMET) of THCv, Cannabinol C2 and Cannabidiol. The white region (HIA) predicts the physicochemical space of molecules that are able to be absorbed by the gastrointestinal system, while the yellow region (BBB) predicts the physicochemical space of molecules that are able to penetrate the brain.

2.3. Toxicity Analysis

The projected acute oral toxicity (LD50) of the substances ranged from the least lethal Cannabinol C2 (1310 mg/kg) to THCv (482 mg/kg). The compounds had LD50 values in class IV, indicating that they were not toxic if swallowed. These selected compounds were further compared with the already available AChE and BuChE inhibiting drugs, and of all three analogs, the Cannabinol-C2 ligand could be used in drug development as it possesses an LD50 value that is even lower, thereby making it an efficient candidate.

2.4. Molecular Docking

The binding affinity between the ligand and the enzyme was identified by using AutoDock Vina 4.2, AutoDock, and iGEMDOCK. The K_i of the ligands were found with the help of AutoDock. The ΔG , Van der Waals, and hydrogen bond energies were calculated using iGEMDOCK. The ligands that were used are THCv, Cannabinol C2, and Cannabidiol. The binding interactions and K_i for ligands docked against AChE are summarized

in Table 3. Galanthamine, Donepezil, and Rivastigmine all had binding affinities of -8.4 , -7.0 , and -7.1 kcal/mol, respectively. THCv and Cannabidiol had the highest binding affinity (-9.4 kcal/mol for both) compared to Cannabinol C2 (-9.3 kcal/mol). Donepezil, Rivastigmine, and Galantamine are the drugs taken against AChE. The binding affinity obtained for Memantine is -6.8 kcal/mol, which is taken against butyrylcholinesterase. Among all three ligands, the highest binding affinity obtained was for Cannabinol C2 and Cannabidiol (-9.2 kcal/mol), which is much higher than that of Memantine. The K_i obtained for Donepezil, Rivastigmine, and Galantamine were 1.46, 0.02, and 2.1, respectively. The K_i of standard drugs was less than that of the ligands taken, which signifies that the inhibitor has more potency against any enzyme or protein. The K_i of the ligands, i.e., THCv, Cannabinol C2, and Cannabidiol, were 1.50, 4.42, and 4.61, respectively. Memantine was the standard drug taken against Butyrylcholinesterase, which gave a K_i of 0.54, which is much less compared to the compounds taken i.e., THCv, Cannabinol C2 and Cannabidiol of K_i 3.25, 4.32, and 3.26, respectively (Figure 3). The interactive amino acids were visualized using the BIOVIA Discovery Studio. The 2D and 3D structure of the interactions is shown in Figures 4 and 5. The interactive amino acids of THCv and Cannabidiol were similar, i.e., VAL340, TYR337, TRP439, TYR449, GLY82, THR83, TRP86, VAL132, GLY121, TYR124, and TRP286; whereas the drugs differed in their interactive amino acid residues in the AChE enzyme. For BuChE, the interactive amino acids for Memantine GLU197, TYR440, VAL127, GLY78, SER79, VAL331, TYR332, PHE329, THR327, ALA328, GLY326, LEU286, GLU125, SER198, HIS438, MET437. THCv contained amino acids that are TYR440, VAL127, TRP82, GLY78, ASN83, SER79, PHE329, PRO285, PHE357, TYR332, SER72, PHE73, ASP70; for Cannabinol C2 it was TYR440, VAL127, GLU197, ALA202, ALA229, ASN397, TRP231, ALA199, SER198, GLU325, HIS438, TRP82, GLY78, SER79, PHE329, TYR332, GLY326, LEU286, THR204, ARG242, VAL228 and for Cannabidiol GLY78, GLY435, ASN322, TYR440, VAL127, ARG424, MET437, LEU428, MET434, THR327, LYS339, VAL331, TRP430, ALA328, PHE76, TRP82. The target proteins (AChE and Butyrylcholinesterase) and compounds (THCv, Cannabinol C2, and Cannabidiol) were re-docked with iGEMDOCK (version 2.1). iGEMDOCK finds distinct bond energies that occur between proteins and chemicals, such as hydrogen bonds (H-Bond), Van Der Waals (VDW) interactions, and electrostatic interactions. The docking data showed that Cannabinol C2 had the lowest docking energy (-96.44 kcal/mol) in comparison to AChE, and the highest docking energy (-81.229 kcal/mol) in comparison to Butyrylcholinesterase. As shown in Table 4, Cannabinol C2 had the highest VDW (-81.229 kcal/mol) and the lowest VDW (-93.67 kcal/mol) for AChE and BuChE, respectively. The binding scores obtained using these tools might not be an exact representation of the true binding affinities, but they could be helpful in describing the relative affinities of two or more ligand molecules bound to the same or to different binding sites.

Table 3. Comparison of different docking parameters between three selected CBD analogues with AChE and BChE with reference drugs.

Compounds	AChE				BChE			
	B.A. (kcal/mol)	H bonds	Ki (μ M)	Interactive Amino Acids	B.A. (kcal/mol)	H bonds	Ki (μ M)	Interactive Amino Acids
THCV	-9.4	1	1.50	VAL340, TYR337, TRP439, TYR449, GLY82, THR83, TRP86, VAL132, GLY121, TYR124, TRP286	-8.9	0	3.25	TYR440, VAL127, TRP82, GLY78, ASN83, SER79, PHE329, PRO285, PHE357, TYR332, SER72, PHE73, ASP70
Cannabinol C2	-9.3	1	4.42	VAL340, TYR337, TRP439, TYR449, GLY82, THR83, TRP86, VAL132, GLY121, TYR124, TRP286	-9.2	3	4.32	TYR440, VAL127, GLU197, ALA202, ALA229, ASN397, TRP231, ALA199, SER198, GLU325, HIS438, TRP82, GLY78, SER79, PHE329, TYR332, GLY326, LEU286, THR204, ARG242, VAL228

Table 3. Cont.

Compounds	AChE				BChE			
	B.A. (kcal/mol)	H bonds	Ki (μM)	Interactive Amino Acids	B.A. (kcal/mol)	H bonds	Ki (μM)	Interactive Amino Acids
Cannabidiol	−9.4	2	4.61	VAL340, TYR337, TRP439, TYR449, GLY82, THR83, TRP86, VAL132, GLY121, TRP286, TYR124	−9.2	1	3.26	GLY78, GLY435, ASN322, TYR440, VAL127, ARG424, MET437, LEU428, MET434, THR327, LYS339, VAL331, TRP430, ALA328, PHE76, TRP82
Memantine	-	-	-	-	−6.8	1	0.54	GLU197, TYR440, VAL127, TRP82, GLY78, SER79, VAL331, TYR332, PHE329, THR327, ALA328, GLY326, LEU286, GLU125, SER198, HIS438, MET437
Donepezil	−8.4	2	1.46	TYR337, TRP439, TYR449, VAL340, GLY82, THR83, GLY121, TRP86, VAL132, TRP286, TYR124	-	-	-	-
Rivastigmine	−7.0	2	0.02	TYR449, TRP439, TYR337, VAL240, GLY482, THR83, VAL132, TRP86, GLY121, TRP286, TYR124	-	-	-	-
Galantamine	−7.1	0	2.1	TYR337, TYR449, VAL340, TRP439, GLY82, THR83, TRP86, TYR124, GLY121, VAL132	-	-	-	-

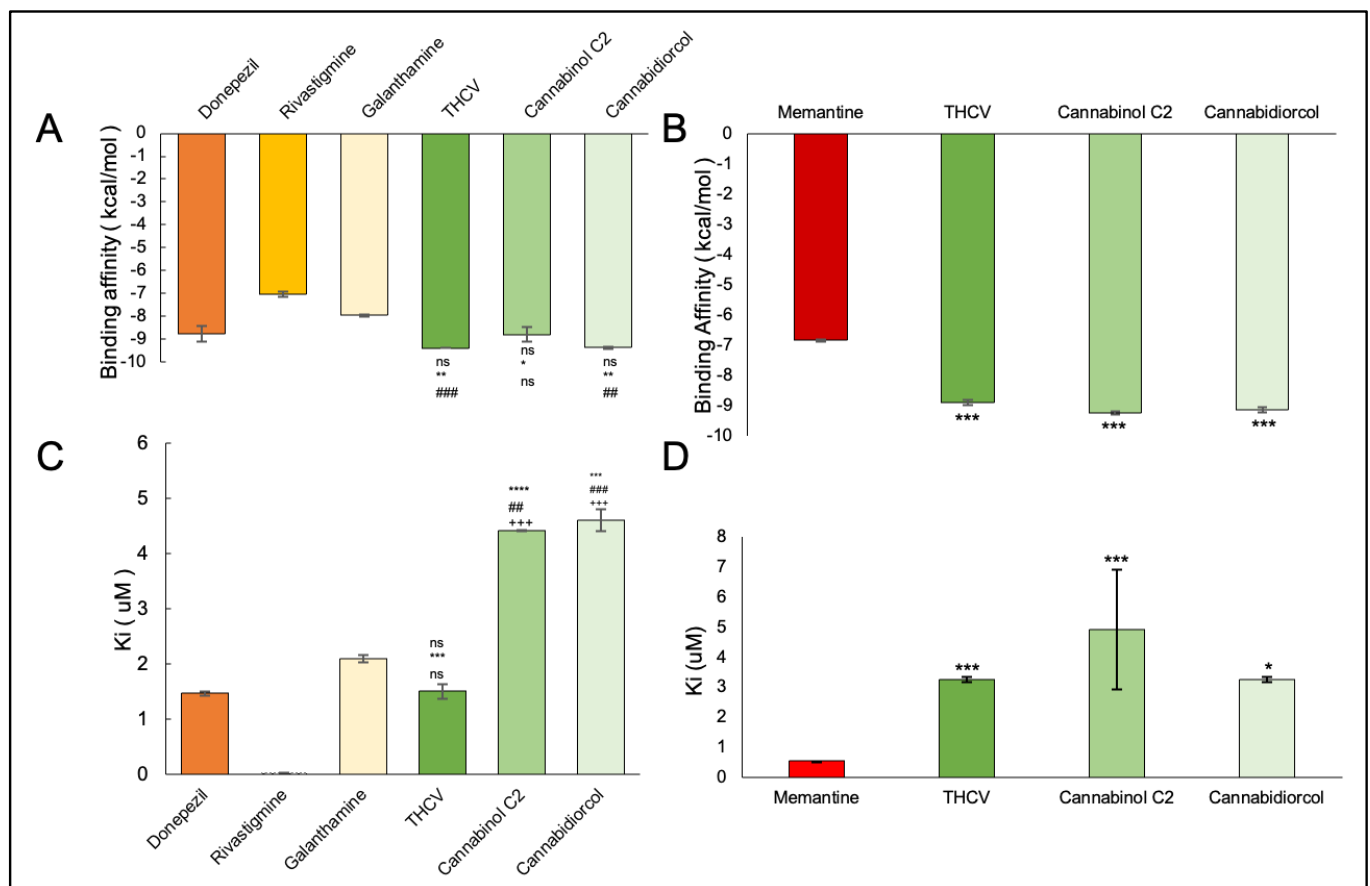


Figure 3. Graphical representation showing comparison of binding affinity and inhibition constant among (A,C) AChE inhibitors (Donepezil, Rivastigmine, Galantamine) and THCv, Cannabinol C2 and Cannabidiol (B,D) Butyrylcholinesterase inhibitors (Memantine) and THCv, Cannabinol C2 and Cannabidiol (* $p < 0.1$, ** $p < 0.01$, *** $p < 0.001$) and values are mean \pm standard deviation where p^* (Donepezil), $p^\#$ (Rivastigmine), p^+ (Galantamine) and p^* (Memantine).

Table 4. Docking energies of compounds with AChE and BChE receptors by iGEMDOCK.

Compounds	AChE			BChE		
	TE	VDW	HB	TE	VDW	HB
THCV	−91.33	−87.579	−4.5	−83.95	−78.95	−5
Cannabinol C2	−96.44	−93.67	5	−81.229	−81.229	0
Cannabidiol	−89.76	−86.579	−4.5	−78.94	−74.3765	−4.5

3. Discussion

Numerous health advantages of cannabinoids have been recognized, including neuroprotective properties against neurodegenerative disorders. Cannabinoids have been linked to a variety of helpful pharmacological benefits on the one hand, and toxic and unpleasant consequences on the other. According to a recent study, cannabinoid dosage and consumer age have a direct impact on health [51]. On the contrary, growing scientific data suggests that CBDs and their changing tone might be viable therapeutic approaches for treating AD. Therefore, the study of AD related pathways and their respective enzyme regulation studies play a significant role. Cannabinoids have been shown to lower oxidative stress and excitotoxicity, as well as the production of amyloid plaques and neurofibrillary tangles [52]. CBD also plays a vital role in AD therapeutics in several manners, such as heat shock proteins and ubiquitin-conjugating enzymes, which are essential regulators of autophagy and are encoded by genes that CBD increases at the mRNA level [53]. The pharmacological effects of CBD on autophagy in AD were validated by a recent study that examined autophagic alterations brought on by prolonged CBD administration in 6-month-old APP/PS1 mice [54]. Additionally, CBD inhibits the hyper phosphorylation of glycogen synthase kinase 3 (GSK-3) brought on by A β and might be a novel treatment option for AD [55]. CBD does not only interact with cannabinoid receptors in the endo cannabinoid system. Instead, it is highly pleiotropic and acts on a variety of other receptors, including adenosine, glycine, non-endocannabinoid G protein-coupled, serotonin (5-HT), opioid, nicotinic acetylcholine, and transient potential Vanilloid receptor type 1 (TRPV1) receptors (i.e., enzymes and ion channels) [56]. CBD has several advantageous features that have been demonstrated in research due to its remarkable diversity of modes of action. These include qualities that are anti-oxidative, anti-inflammatory, analgesic, anticancer, anticonvulsant, anxiolytic, antidepressant, in addition to others [57].

The immunomodulatory impact of the CB2 receptor, which controls microglial activity, can reduce AD neuroinflammatory processes [58]. Another important effect is on acetylcholine availability and the inhibition of AChE-induced aggregation. BACE-1 and γ -secretase, which cleaves APP and forms amyloid beta oligomers that mediate cholinergic neurotransmission, is a major enzyme responsible for pathological alterations in AD; this dysfunction is directly related to neuroinflammation and cholinergic insufficiency in the CNS. Recent research indicates that *Cannabis sativa* secondary metabolites, CBD, activate PPAR γ via the cell signaling Wnt/ β -catenin pathway [59]. This activation reduces the neurotoxic nature of A β amyloids and oxidative stress in PC12 cells. CBD also increases cell viability and decreases ROS levels with the reduction in the peroxidation of lipids. There is also a decrease in Tau hyper phosphorylation and the inhibition of AChE. According to some in vitro studies, CBD increases the A β amyloid protein's proteolytic pathway on SHY5YAPP+ cells (a neuroblastoma cell line) by inducing APP ubiquitination, which boosts cell viability [60]. Ethyl, which is a functional group in cannabinol C2 and is similar to Galantamine, can effectively enter the active pocket of AChE [61]. Furthermore, THCV consist of Enol as a functional group that might be identified to break up amyloid plaques and enhance the rapid clearance of toxic aggregates in AD which is relatively similar to keto-enol pharmacophores [62].

In a recent study, THCV shows a significant effect at 50 μ M and a decreased epileptiform burst speed in mice brain [63]. In M Sprague–Dawley rats/CB2 knockout mice, THCV also improved motor activities, reduced neuronal loss and reduced microglial acti-

vation [64]. Another analog, Cannabidiol, has shown promising results ($LC_{50} = 0.348 \pm 0.002 \mu\text{g/mL}$), similar to vincristine sulfate in a cytotoxicity assay relating to cancer [65]. In this study, the interactive molecule of compounds (e.g., the Enol and keto groups) were showing significant binding with AChE and BuChE [66]. Our study demonstrates that the previous in vitro analyses were correlated with our in silico, approach and provided an additional step toward its validation as a putative inhibitor for AD.

4. Materials & Methods

The in silico analysis involved screening CBD and its 74 analogs for their binding and interaction with AChE and BuChE receptors. The ligands were visualized for their physicochemical properties, including pharmacokinetic properties and drug-likeness properties, as well as for their consideration as drugs for AChE and BuChE regulation. A detailed description of each part of the methodology is provided below:

4.1. Retrieval of the Ligand Molecule and Protein Structure for ADME Studies

4.1.1. Protein Preparation

The crystal three-dimensional structure of the target receptors AChE (PDB ID: 4PQE), gamma secretase (PDB ID: 6IYC), BACE-1 (PDB ID: 1SZG), and BuChE (PDB ID: 4BDS) was retrieved from PDB <https://www.rcsb.org/> (accessed on 11 November 2022) and used as a rigid receptor. Auto Dock MGLTools was used to prepare the receptor protein. The deposited protein data consisted of water molecules and NAG- and NAM-like molecules to stabilize the crystallographic structure of the proteins. The enzymes were prepared by removing the NAG, NAM, and water molecules from the sequence to be used for further study, assigning bond orders, adding hydrogen atoms, and distributing Kollmann's charge equal to the whole receptor. The energy minimization of protein was done through the Chiron webserver <https://dokhlab.med.psu.edu/chiron/login.php> (accessed on 11 November 2022) [67].

4.1.2. Ligand Preparation

The chemical structures of known inhibitors for AChE, γ secretase, BACE-1, and BuChE were retrieved from the Drug Bank database, and other compounds described in this study were mainly CBD and its 74 derivatives. Those analogs were drawn and retrieved from ChemDraw (<https://chemdrawdirect.perkinelmer.cloud/js/sample/index.html>) (accessed on 13 November 2022) and PubChem <https://pubchem.ncbi.nlm.nih.gov/> (accessed on 13 November 2022). After retrieval, the chemical compounds were energy minimized through Avogadro software 1.2.0 [68].

4.2. Drug Likelihood Properties

The drug-likeness, mutagenic, tumorigenic, reproductive, and irritating impacts of drug-toxicity risk factors were examined using the OSIRIS Data Warrior V5.2.1 program. It was used to study the toxicological characteristics of compounds that are possible orally active therapeutic candidates in clinical applications. An orally active compound should follow the Lipinski rule with zero violations. The "rule of five" model proposed by Lipinski suggested that for a compound to be considered orally bioactive, the violation of two or more of these conditions predicted a molecule to be a non-orally available drug [69].

4.3. ADME and Toxicity Test

4.3.1. ADME Properties

ADME is required to assess the pharmacodynamics of a suggested chemical that might be employed as a medicine. The SWISS-ADME server (<http://www.swissadme.ch/>, accessed on 29 November 2022) was used to analyze the chosen ligands retrieved from PubChem and ChemDraw using canonical SMILES. As an input system, the structural data file and simplified molecular data input format was utilized to calculate the absorption, distribution, metabolism, excretion, and toxicity (ADMET) values. SWISS-ADME allows the

user to study parameters such as lipophilicity, water solubility, Log S, drug likeness rules, and some medicinal chemistry. The observed values of those compounds are presented in Table S1. Through their pharmacokinetic properties, 44 compounds were shown to follow Lipinski's rules [70]. Out of those 44 compounds, seven were able to cross the BBB (Blood-Brain Barrier) [71] and were further evaluated for toxicity determination. Out of those seven compounds, only three (THCV, Cannabinol-C2, and Cannabidiol) were able to cross the CNS and were studied further.

4.3.2. Toxicity Prediction

Toxicology prediction helps predict the tolerability of a chemical before it is employed in a human or animal model. Computer-based methods are now available for obtaining a safety profile of the required substance to quantify toxicity. To analyze the hazardous effects of those three compounds, the Protox-II website <https://tox-new.charite.de/prottoxII/> (accessed on 16 November 2022) was utilized with LD50, Hepatotoxicity, Cytotoxicity, and Immunotoxicity as toxicity parameters of a query chemical molecule that are predicted with this server [72].

4.4. Molecular Docking

The interaction between proteins (AChE and BuChE) and ligands (CBD and its 74 analogs) was determined by conducting molecular docking using AutoDock 4, AutoDock Vina 4.2, and iGEMDOCK in a triplicate manner. For the specific docking, the active site of AChE and BuChE was determined using CASTp 3.0 <http://sts.bioe.uic.edu/castp/> (accessed on 14 November 2022). Kollmann's charges, Gasteiger partial charges, and polar hydrogen atoms were added to the proteins. To achieve the best conformational docking results, the grid box was centered on the binding site of the ligand and the auto grid position was set at (x: -27.195, y: 21.246, z: -10.429) for AChE and at (x: 135.922, y: 140.041, z: 43.131) for BuChE. Gamma secretase and BACE-1 showed non-significant results with Hecogenin and Imatinib, respectively, so they were not used in further research (Supplemental Figure S3). The docking algorithm with 100 runs had the Lamarckian genetic algorithm (LGA) and the empirical free energy function as default parameters. For iGEMDOCK the population size was 200, there were 70 generations, and the number of solutions was two with the standard docking setting. After generating a set of poses, the best fit was selected, which represented the total binding energy in the form of hydrogen bonds (HB), van der Waals forces (VDW), and electrostatic interaction. BIOVIA Discovery Studio v. 21.1.0 was used to display the docked conformations and 3D target-ligand interactions, which assisted in analyzing and predicting the kind of amino acid involved and their interactions. Donepezil, Rivastigmine, and Galantamine, which had binding affinities of -8.77 kcal/mol, -7.0 kcal/mol, and -7.97 kcal/mol, respectively, were chosen as a reference to compare with analogues for the AChE receptor [73]. Memantine had a binding affinity of -6.83 kcal/mol and was chosen as a reference to compare with analogues for the BuChE receptor [74,75].

5. Conclusions

In comparison to known drugs, THCV, Cannabinol C2 and Cannabidiol dominated cannabinoids' inhibitory activities on AChE and BuChE. Computational approaches suggest that THCV, Cannabinol C2, and Cannabidiol are more appropriate for the inhibition of the enzymes AChE and BuChE, which act as the culprits of Alzheimer's disease. Cell and animal studies are needed to improve the efficacy of these cannabinoids and to learn more about the effecting pathways.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants12030510/s1>, Figure S1: 3D & 2D representation of BACE-1 & Gamma Secretase with inhibitors Hecogenin and Imatinib respectively; Table S1: All 75 compounds ADMET properties; Table S2: Compounds passing HHP and Lipinski's rule; Table S3: Compounds passing BBB permeability.

Author Contributions: Conceptualization, M.J.; methodology, M.J., P.K. and N.P.; software, N.P.; validation, M.J., N.P. and P.K.; formal analysis, A.R.; data curation, V.C.; writing—original draft preparation, N.P., A.R. and V.C.; writing—review and editing, M.J. and P.K.; visualization, P.K.; supervision, M.J.; funding acquisition, P.K. All authors have read and agreed to the published version of the manuscript.

Funding: Not funded.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Not Applicable.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

Abbreviations

CBD	Cannabidiol
THC	Tetrahydrocannabinol
AChE	Acetylcholinesterase
BuChE	Butyrylcholinesterase
AD	Alzheimer’s disease
γ-secretase	Gamma-secretase
BACE-1	Beta secretase-1
ADMET	Absorption, Distribution, Metabolism, Excretion and Toxicity
THCV	Tetrahydrocannabivarin
iGEMDOCK	A Graphical Environment for Recognizing Pharmacological Interactions and Virtual Screening
GPR55	G protein-coupled receptors 55
GPR3	G protein-coupled receptors 3
NF-κB	nuclear factor κ B
PPARγ	peroxisome proliferator-activated receptor γ
GPCRs	G protein-coupled receptors
BBB	Blood brain barrier
APP	Amyloid precursor protein
GPP	geranyl diphosphate
OLA	olivetolic acid
NFT	neurofibrillary tangles
Aβ	beta-amyloid
sAPP-β	soluble amyloid precursor protein β
CYP2D6	Cytochrome P450 2D6
CYP3A4	Cytochrome P450 3A4
P450s	cytochrome P450 enzymes
hERG	human ether-a-go-go-related gene
LD50	lethal dose 50
VDW	van der Waals interactions
HB	hydrogen bonds
SHY5YAPP⁺ cells	SH-SY5Y cells transfected with the amyloid precursor protein
CASTp	Computed Atlas of Surface Topography of proteins
K_i	inhibition constant of enzymes
ΔG	Gibbs free energy
TE	electrostatic forces
Kcal/mol	kilo-calorie per mole
PAINS	Pan Assay Interference Compounds
TPSA	topological molecular polar surface area
ClogP	$\log(C_{\text{octanol}}/C_{\text{water}})$
HIA	human gastrointestinal absorption

References

1. Bonini, S.A.; Premoli, M.; Tambaro, S.; Kumar, A.; Maccarinelli, G.; Memo, M.; Mastinu, A. *Cannabis sativa*: A comprehensive ethnopharmacological review of a medicinal plant with a long history. *J. Ethnopharmacol.* **2018**, *227*, 300–315. [[CrossRef](#)] [[PubMed](#)]
2. Dowling, C.A.; Melzer, R.; Schilling, S. Timing is everything: The genetics of flowering time in *Cannabis sativa*. *Biochemist* **2021**, *43*, 34–38. [[CrossRef](#)]
3. Andre, C.M.; Hausman, J.F.; Guerriero, G. *Cannabis sativa*: The plant of the thousand and one molecules. *Front. Plant Sci.* **2016**, *7*, 19. [[CrossRef](#)] [[PubMed](#)]
4. Banister, S.D.; Arnold, J.C.; Connor, M.; Glass, M.; McGregor, I.S. Dark classics in chemical neuroscience: Δ^9 -tetrahydrocannabinol. *ACS Chem. Neurosci.* **2019**, *10*, 2160–2175. [[CrossRef](#)]
5. Gill, E.W.; Paton, W.D.M.; Pertwee, R.G. Preliminary Experiments on the Chemistry and Pharmacology of Cannabis. *Nature* **1970**, *228*, 134–136. [[CrossRef](#)]
6. McPartland, J.M.; Russo, E.B. Cannabis and Cannabis Extracts: Greater Than the Sum of Their Parts? *J. Cannabis Ther.* **2001**, *1*, 103–132. [[CrossRef](#)]
7. Chandra, S.; Lata, H.; ElSohly, M.A. *Cannabis sativa L.-Botany and Biotechnology*; Springer: Berlin/Heidelberg, Germany, 2017; ISBN 3-319-54564-7.
8. Hanuš, L.O.; Meyer, S.M.; Muñoz, E.; Tagliatalata-Scafati, O.; Appendino, G. Phytocannabinoids: A Unified Critical Inventory. *Nat. Prod. Rep.* **2016**, *33*, 1357–1392. [[CrossRef](#)]
9. Citti, C.; Linciano, P.; Panseri, S.; Vezzalini, F.; Forni, F.; Vandelli, M.A.; Cannazza, G. Cannabinoid Profiling of Hemp Seed Oil by Liquid Chromatography Coupled to High-Resolution Mass Spectrometry. *Front. Plant Sci.* **2019**, *10*, 120. [[CrossRef](#)]
10. Pavlovic, R.; Panseri, S.; Giupponi, L.; Leoni, V.; Citti, C.; Cattaneo, C.; Cavaletto, M.; Giorgi, A. Phytochemical and Ecological-Analysis of Two Varieties of Hemp (*Cannabis sativa* L.) Grown in a Mountain Environment of Italian Alps. *Front. Plant Sci.* **2019**, *10*, 1265. [[CrossRef](#)]
11. Pertwee, R.G. Cannabinoid pharmacology: The first 66 years. *Br. J. Pharmacol.* **2006**, *147*, 163–171. [[CrossRef](#)]
12. Curran, H.V.; Freeman, T.P.; Mokrysz, C.; Lewis, D.A.; Morgan, C.J.A.; Parsons, L.H. Keep off the grass? Cannabis, cognition and addiction. *Nat. Rev. Neurosci.* **2016**, *17*, 293–306. [[CrossRef](#)] [[PubMed](#)]
13. ElSohly, M.A.; Radwan, M.M.; Gul, W.; Chandra, S.; Galal, A. Phytochemistry of *Cannabis sativa* L. In *Progress in the Chemistry of Organic Natural Products*; Kinghorn, A., Falk, H., Gibbons, S., Kobayashi, J., Eds.; Springer: Cham, Switzerland, 2017; Volume 103, pp. 1–36.
14. Lewis, M.M.; Yang, Y.; Wasilewski, E.; Clarke, H.A.; Kotra, L.P. Chemical profiling of medical cannabis extracts. *ACS Omega* **2017**, *2*, 6091–6103. [[CrossRef](#)] [[PubMed](#)]
15. Kumar, P.; Mahato, D.K.; Kamle, M.; Borah, R.; Sharma, B.; Pandhi, S.; Tripathi, V.; Yadav, H.S.; Devi, S.; Patil, U.; et al. Pharmacological properties, therapeutic potential, and legal status of *Cannabis sativa* L.: An overview. *Phytother. Res.* **2021**, *35*, 6010–6029. [[CrossRef](#)]
16. Abuhasira, R.; Schleider, L.B.L.; Mechoulam, R.; Novack, V. Epidemiological characteristics, safety and efficacy of medical cannabis in the elderly. *Eur. J. Intern. Med.* **2018**, *49*, 44–50. [[CrossRef](#)] [[PubMed](#)]
17. Rosenberg, E.C.; Tsien, R.W.; Whalley, B.J.; Devinsky, O. Cannabinoids and epilepsy. *Neurotherapeutics* **2015**, *12*, 747–768. [[CrossRef](#)]
18. Nuutinen, T. European journal of medicinal chemistry medicinal properties of terpenes found in cannabis sativa and *Humulus lupulus*. *Eur. J. Med. Chem.* **2018**, *157*, 198–228. [[CrossRef](#)] [[PubMed](#)]
19. Aizpurua-Olaizola, O.; Soydaner, U.; Oztürk, E.; Schibano, D.; Simsir, Y.; Navarro, P.; Etxebarria, N.; Usobiaga, A. Evolution of the cannabinoid and terpene content during the growth of cannabis sativa plants from different chemotypes. *J. Nat. Prod.* **2016**, *79*, 324–331. [[CrossRef](#)] [[PubMed](#)]
20. Lal, S.; Shekher, A.; Puneet Narula, A.S.; Abrahamse, H.; Gupta, S.C. Cannabis and its constituents for cancer: History, biogenesis, chemistry and pharmacological activities. *Pharmacol. Res.* **2021**, *163*, 105302. [[CrossRef](#)] [[PubMed](#)]
21. Iuvone, T.; Esposito, G.; Esposito, R.; Santamaria, R.; Di Rosa, M.; Izzo, A.A. Neuroprotective effect of cannabidiol, a non-psychoactive component from Cannabis sativa, on β -amyloid-induced toxicity in PC12 cells. *J. Neurochem.* **2004**, *89*, 134–141. [[CrossRef](#)]
22. Esposito, G.; De Filippis, D.; Carnuccio, R.; Izzo, A.A.; Iuvone, T. The marijuana component cannabidiol inhibits beta-amyloid-induced tau protein hyperphosphorylation through Wnt/beta-catenin pathway rescue in PC12 cells. *J. Mol. Med.* **2006**, *84*, 253–258. [[CrossRef](#)]
23. Eubanks, L.M.; Rogers, C.J.; Beuscher, A.; Koob, G.F.; Olson, A.J.; Dickerson, T.J.; Janda, K.D. A molecular link between the active component of marijuana and Alzheimer’s disease pathology. *Mol. Pharm.* **2006**, *3*, 773–777. [[CrossRef](#)] [[PubMed](#)]
24. Cassano, T.; Villani, R.; Pace, L.; Carbone, A.; Bukke, V.N.; Orkisz, S.; Avolio, C.; Serviddio, G. From Cannabis sativa to cannabidiol: Promising therapeutic candidate for the treatment of neurodegenerative diseases. *Front. Pharmacol.* **2020**, *6*, 124. [[CrossRef](#)] [[PubMed](#)]
25. Nuñez-Borque, E.; González-Naranjo, P.; Bartolomé, F.; Alquézar, C.; Reinares-Sebastián, A.; Pérez, C.; Ceballos, M.L.; Páez, J.A.; Campillo, N.E.; Martín-Requero, Á. Targeting cannabinoid receptor activation and BACE-1 activity counteracts TgAPP mice memory impairment and Alzheimer’s disease lymphoblast alterations. *Mol. Neurobiol.* **2020**, *57*, 1938–1951. [[CrossRef](#)] [[PubMed](#)]

26. Citti, C.; Linciano, P.; Russo, F.; Luongo, L.; Iannotta, M.; Maione, S.; Laganà, A.; Capriotti, A.L.; Forni, F.; Vandelli, M.A.; et al. A novel phytocannabinoid isolated from *Cannabis sativa* L. with an in vivo cannabimimetic activity higher than Δ^9 -tetrahydrocannabinol: Δ^9 -Tetrahydrocannabiphorol. *Sci. Rep.* **2019**, *9*, 20335. [[CrossRef](#)]
27. Chen, R.; Zhang, J.; Fan, N.; Teng, Z.; Wu, Y.; Yang, H.; Tang, Y.; Sun, H.; Song, Y.; Chen, C. Δ^9 -THC-Caused Synaptic and Memory Impairments Are Mediated through COX-2 Signaling. *Cell* **2013**, *155*, 1154–1165. [[CrossRef](#)]
28. Franke, T.N.; Irwin, C.; Beindorff, N.; Bouter, Y.; Bouter, C. Effects of tetrahydrocannabinol treatment on brain metabolism and neuron loss in a mouse model of sporadic Alzheimer's disease. *Nuklearmedizin-Nucl.* **2019**, *58*, P94.
29. Wiles, D.; Shanbhag, B.K.; O'Brien, M.; Doblin, M.S.; Bacic, A.; Beddoe, T. Heterologous production of Cannabis sativa-derived specialised metabolites of medicinal significance—Insights into engineering strategies. *Phytochemistry* **2022**, *203*, 113380. [[CrossRef](#)]
30. Schubert, D.; Kepchia, D.; Liang, Z.; Dargusch, R.; Goldberg, J.; Maher, P. Efficacy of cannabinoids in a pre-clinical drug-screening platform for Alzheimer's disease. *Mol. Neurobiol.* **2019**, *56*, 7719–7730. [[CrossRef](#)]
31. Shinjyo, N.; Di Marzo, V. The effect of cannabichromene on adult neural stem/progenitor cells. *Neurochem. Int.* **2013**, *63*, 432–437. [[CrossRef](#)]
32. Werz, O.; Seegers, J.; Schaible, A.M.; Weinigel, C.; Barz, D.; Koeberle, A.; Appendino, G. Cannflavins from hemp sprouts, a novel cannabinoid-free hemp food product, target microsomal prostaglandin E2 synthase-1 and 5-lipoxygenase. *PharmaNutrition* **2014**, *2*, 53–60. [[CrossRef](#)]
33. Colović, M.B.; Krstić, D.Z.; Lazarević-Pašti, T.D.; Bondžić, A.M.; Vasić, V.M. Acetylcholinesterase inhibitors: Pharmacology and toxicology. *Curr. Neuropharmacol.* **2013**, *11*, 315–335. [[CrossRef](#)] [[PubMed](#)]
34. Djurfeldt, M.; Hjorth, J.; Eppler, J.M.; Dudani, N.; Helias, M.; Potjans, T.C.; Ekeberg, Ö. Run-time interoperability between neuronal network simulators based on the MUSIC framework. *Neuroinformatics* **2010**, *8*, 43–60. [[CrossRef](#)] [[PubMed](#)]
35. Espadas, I.; Keifman, E.; Palomo-Garo, C.; Burgaz, S.; García, C.; Fern'andez-Ruiz, J.; Moratalla, R. Beneficial effects of the phytocannabinoid Δ^9 -THCV in L-DOPA-induced dyskinesia in Parkinson's disease. *Neurobiol. Dis.* **2020**, *141*, 104892. [[CrossRef](#)]
36. Rea, K.A.; Casaretto, J.A.; Al-Abdul-Wahid, M.S.; Sukumaran, A.; Geddes-McAlister, J.; Rothstein, S.J.; Akhtar, T.A. Biosynthesis of cannflavins A and B from *Cannabis sativa* L. *Phytochemistry* **2019**, *164*, 162–171. [[CrossRef](#)] [[PubMed](#)]
37. Barrett, M.L.; Gordon, D.; Evans, F.J. Isolation from *Cannabis sativa* L. of cannflavin—A novel inhibitor of prostaglandin production. *Biochem. Pharmacol.* **1985**, *34*, 2019–2024. [[CrossRef](#)] [[PubMed](#)]
38. Bhunia, S.; Kolishetti, N.; Arias, A.Y.; Vashist, A.; Nair, M. Cannabidiol for neurodegenerative disorders: A comprehensive review. *Front. Pharmacol.* **2022**, *13*, 989717. [[CrossRef](#)] [[PubMed](#)]
39. Coles, M.; Steiner-Lim, G.Z.; Karl, T. Therapeutic properties of multi-cannabinoid treatment strategies for Alzheimer's disease. *Front. Neurosci.* **2022**, *16*, 962922. [[CrossRef](#)]
40. Tiwari, S.; Atluri, V.; Kaushik, A.; Yndart, A.; Nair, M. Alzheimer's disease: Pathogenesis, diagnostics, and therapeutics. *Int. J. Nanomed.* **2019**, *14*, 5541. [[CrossRef](#)]
41. Alzheimer's Association. 2021 Alzheimer's disease facts and figures. *Alzheimer's Dement.* **2021**, *17*, 327–406. [[CrossRef](#)]
42. Bortolato, B.; Miskowiak, K.W.; Köhler, C.A.; Maes, M.; Fernandes, B.S.; Berk, M.; Carvalho, A.F. Cognitive remission: A novel objective for the treatment of major depression? *BMC Med.* **2016**, *14*, 1–8. [[CrossRef](#)]
43. Menting, K.W.; Claassen, J.A. β -secretase inhibitor; a promising novel therapeutic drug in Alzheimer's disease. *Front. Aging Neurosci.* **2014**, *6*, 165. [[CrossRef](#)] [[PubMed](#)]
44. Yan, X.; Tang, J.; dos Santos Passos, C.; Nurisso, A.; SimõesPires, C.A.; Ji, M.; Lou, H.; Fan, P. Characterization of Lignanamides from hemp (*Cannabis sativa* L.) seed and their antioxidant and acetylcholinesterase inhibitory activities. *J. Agric. Food Chem.* **2015**, *63*, 10611–10619. [[CrossRef](#)] [[PubMed](#)]
45. Panda, S.S.; Jhanji, N. Natural products as potential anti-Alzheimer agents. *Curr. Med. Chem.* **2020**, *27*, 5887–5917. [[CrossRef](#)]
46. Hanseeuw, B.J.; Betensky, R.A.; Jacobs, H.I.L.; Schultz, A.P.; Sepulcre, J.; Becker, J.A.; Cosio, D.M.O.; Farrell, M.; Quiroz, Y.T.; Mormino, E.C.; et al. Association of amyloid and tau with cognition in preclinical Alzheimer disease: A longitudinal study. *JAMA Neurol.* **2019**, *76*, 915–924. [[CrossRef](#)] [[PubMed](#)]
47. Martinez, A.; Castro, A. Novel cholinesterase inhibitors as future effective drugs for the treatment of Alzheimer's disease. *Expert Opin. Investig. Drugs* **2006**, *15*, 1–12. [[CrossRef](#)]
48. Alzheimer's Association Report. Alzheimer's disease facts and figures. *Alzheimer's Dement.* **2020**, *11*, 1–88.
49. Masondo, N.A.; Stafford, G.I.; Aremu, A.O.; Makunga, N.P. Acetylcholinesterase inhibitors from southern African plants: An overview of ethnobotanical, pharmacological potential and phytochemical research including and beyond Alzheimer's disease treatment. *S. Afr. J. Bot.* **2019**, *120*, 39–64. [[CrossRef](#)]
50. Paunescu, H.; Dima, L.; Ghita, I.; Coman, L.; Ifteni, P.I.; Fulga, I.; Coman, O.A. A systematic review of clinical studies on the effect of psychoactive cannabinoids in psychiatric conditions in Alzheimer Dementia. *Am. J. Ther.* **2020**, *27*, e249–e269. [[CrossRef](#)]
51. Holst, B.; Williamson, G. Nutrients and phytochemicals: From bioavailability to bioefficacy beyond antioxidants. *Curr. Opin. Biotech.* **2008**, *19*, 73–82. [[CrossRef](#)]
52. Campbell, V.A.; Gowran, A. Alzheimer's disease; taking the edge off with cannabinoids? *Brit. J. Pharmacol.* **2007**, *152*, 655–662. [[CrossRef](#)]
53. Xiong, Y.; Lim, C.S. Understanding the Modulatory Effects of Cannabidiol on Alzheimer's Disease. *Brain Sci.* **2021**, *11*, 1211. [[CrossRef](#)] [[PubMed](#)]

54. Hao, F.; Feng, Y. Cannabidiol (CBD) enhanced the hippocampal immune response and autophagy of APP/PS1 Alzheimer's mice uncovered by RNA-seq. *Life Sci.* **2021**, *264*, 118624. [[CrossRef](#)] [[PubMed](#)]
55. Sirbu, C.A.; Manole, A.M.; Vasile, T.M.; Toma, G.S.; Dobrican, L.R.; Virvara, D.G.; Vasiliu, O. Cannabinoids—A new therapeutic strategy in neurology. *Rom. J. Mil. Med.* **2022**, *125*, 349. [[CrossRef](#)]
56. Ibeas Bih, C.; Chen, T.; Nunn, A.V.; Bazelot, M.; Dallas, M.; Whalley, B.J. Molecular targets of cannabidiol in neurological disorders. *Neurotherapeutics* **2015**, *12*, 699–730. [[CrossRef](#)] [[PubMed](#)]
57. Liang, J.; Zhang, Y.; Chi, P.; Liu, H.; Jing, Z.; Cao, H.; Du, Y.; Zhao, Y.; Qin, X.; Zhang, W.; et al. Essential Oils: Chemical Constituents, Potential Neuropharmacological Effects and Aromatherapy—A Review. *Pharmacol. Res. Mod. Chin. Med.* **2022**, *6*, 100210. [[CrossRef](#)]
58. Komorowska-Müller, J.A.; Schmöle, A.C. CB2 receptor in microglia: The guardian of self-control. *Int. J. Mol. Sci.* **2020**, *22*, 19. [[CrossRef](#)]
59. Vallée, A.; Lecarpentier, Y.; Guillevin, R.; Vallée, J.N. Effects of cannabidiol interactions with Wnt/ β -catenin pathway and PPAR γ on oxidative stress and neuroinflammation in Alzheimer's disease. *Acta Biochim. Biophys. Sin.* **2017**, *49*, 853–866. [[CrossRef](#)]
60. Kim, S.H.; Yang, J.W.; Kim, K.H.; Kim, J.U.; Yook, T.H. A Review on Studies of Marijuana for Alzheimer's Disease—Focusing on CBD, THC. *J. Pharmacopunct.* **2019**, *22*, 225. [[CrossRef](#)]
61. Jiang, X.; Zhang, Z.; Zuo, J.; Wu, C.; Zha, L.; Xu, Y.; Wang, S.; Shi, J.; Liu, X.H.; Zhang, J.; et al. Novel cannabidiol—Carbamate hybrids as selective BuChE inhibitors: Docking-based fragment reassembly for the development of potential therapeutic agents against Alzheimer's disease. *Eur. J. Med. Chem.* **2021**, *223*, 113735. [[CrossRef](#)]
62. Makhouri, F.R.; Ghasemi, J.B. In silico studies in drug research against neurodegenerative diseases. *Curr. Neuropharmacol.* **2018**, *16*, 664–725. [[CrossRef](#)]
63. Hill, A.J.; Weston, S.E.; Jones, N.A.; Smith, I.; Bevan, S.A.; Williamson, E.M.; Stephens, G.J.; Williams, C.M.; Whalley, B.J. Δ^9 -Tetrahydrocannabivarin suppresses in vitro epileptiform and in vivo seizure activity in adult rats. *Epilepsia* **2010**, *51*, 1522–1532. [[CrossRef](#)] [[PubMed](#)]
64. García, C.; Palomo-Garo, C.; García-Arencibia, M.; Ramos, J.A.; Pertwee, R.G.; Fernández-Ruiz, J. Symptom-relieving and neuroprotective effects of the phytocannabinoid Δ^9 -THCV in animal models of Parkinson's disease. *Br. J. Pharmacol.* **2011**, *163*, 1495–1506. [[CrossRef](#)] [[PubMed](#)]
65. Baroi, S.; Saha, A.; Bachar, R.; Bachar, S.C. Cannabinoid as potential aromatase inhibitor through molecular modeling and screening for anti-cancer activity. *Dhaka Univ. J. Pharm. Sci.* **2020**, *26*, 47–58. [[CrossRef](#)]
66. Zoltán, O.; László, P.; Éva, K.; Béla, V. A “keto-enol” plaque buster mechanism to diminish Alzheimer's β -Amyloid burden. *Biochem. Biophys. Res. Comm.* **2020**, *532*, 82–87. [[CrossRef](#)] [[PubMed](#)]
67. Scuderi, C.; Steardo, L.; Esposito, G. Cannabidiol promotes amyloid precursor protein ubiquitination and reduction of beta amyloid expression in SHSY5YAPP+ cells through PPAR γ involvement. *Phytother. Res.* **2014**, *28*, 1007–1013. [[CrossRef](#)]
68. Ramachandran, S.; Kota, P.; Ding, F.; Dokholyan, N.V. Homology modeling: Generating structural models to understand protein function and mechanism. In *Computational modeling of biological systems. PROTEINS Struct. Funct. Bioinform.* **2011**, *79*, 261–270. [[CrossRef](#)]
69. Patel, N.B.; Patel, L.N.; Patel, K.D.; Patel, M.V.; Kalasariya, H.S. Admet & Cytotoxicity Prediction of Red Seaweed Gracillaria Dura: An in Silico Approach. Available online: https://www.researchgate.net/profile/Nikunj-Patel-22/publication/339973699_ADMET_CYTOTOXICITY_PREDICTION_OF_RED_SEAWEED_GRACILLARIA_DURA_AN_IN_SILICO_APPROACH/links/5e708cdd92851c1a689a7c90/ADMET-CYTOTOXICITY-PREDICTION-OF-RED-SEAWEED-GRACILLARIA-DURA-AN-IN-SILICO-APPROACH.pdf (accessed on 29 November 2022).
70. Hussain, H.; Ahmad, S.; Shah, S.W.; Ghias, M.; Ullah, A.; Rahman, S.U.; Kamal, Z.; Khan, F.A.; Khan, N.M.; Muhammad, J.; et al. Neuroprotective Potential of Synthetic Mono-Carbonyl Curcumin Analogs Assessed by Molecular Docking Studies. *Molecules* **2021**, *26*, 7168. [[CrossRef](#)]
71. Van De Waterbeemd, H.; Gifford, E. ADMET in silico modelling: Towards prediction paradise? *Nat. Rev. Drug Discov.* **2003**, *2*, 192–204. [[CrossRef](#)]
72. Daina, A.; Zoete, V. A boiled-egg to predict gastrointestinal absorption and brain penetration of small molecules. *ChemMedChem* **2016**, *11*, 1117–1121. [[CrossRef](#)]
73. Banerjee, P.; Eckert, A.O.; Schrey, A.K.; Preissner, R. ProTox-II: A webserver for the prediction of toxicity of chemicals. *Nucl. Acids Res.* **2018**, *46*, W257–W263. [[CrossRef](#)]
74. Santos, T.C.; Gomes, T.M.; Pinto, B.A.; Camara, A.L.; Paes, A.M. Naturally occurring acetylcholinesterase inhibitors and their potential use for Alzheimer's disease therapy. *Front. Pharmacol.* **2018**, *18*, 1192. [[CrossRef](#)] [[PubMed](#)]
75. Folch, J.; Busquets, O.; Ettcheto, M.; Sánchez-López, E.; Castro-Torres, R.D.; Verdaguer, E.; Garcia, M.L.; Olloquequi, J.; Casadesús, G.; Beas-Zarate, C.; et al. Memantine for the treatment of dementia: A review on its current and future applications. *J. Alz. Dis.* **2018**, *62*, 1223–1240. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.