

Review

Microbiota Effect on Trimethylamine N-Oxide Production: From Cancer to Fitness—A Practical Preventing Recommendation and Therapies

Edoardo Tacconi ^{1,†} , Giuseppe Palma ^{2,*,†} , Davide De Biase ³, Antonio Luciano ², Massimiliano Barbieri ², Filomena de Nigris ⁴ and Francesca Bruzzese ²

¹ Department of Human Science and Quality of Life Promotion, San Raffaele Roma Open University, 00166 Rome, Italy

² S.S.D. Sperimentazione Animale, Istituto Nazionale Tumori-IRCCS-Fondazione G. Pascale, 80131 Naples, Italy

³ Department of Pharmacy, University of Salerno, Via Giovanni Paolo II 132, 84084 Fisciano, Italy

⁴ Department of Precision Medicine, School of Medicine, Università degli Studi della Campania “Luigi Vanvitelli”, Via De Crecchio 7, 80138 Naples, Italy

* Correspondence: giuseppe.palma@istitutotumori.na.it

† These authors contributed equally to this work.

Abstract: Trimethylamine N-oxide (TMAO) is a microbial metabolite derived from nutrients, such as choline, L-carnitine, ergothioneine and betaine. Recently, it has come under the spotlight for its close interactions with gut microbiota and implications for gastrointestinal cancers, cardiovascular disease, and systemic inflammation. The culprits in the origin of these pathologies may be food sources, in particular, high fat meat, offal, egg yolk, whole dairy products, and fatty fish, but intercalated between these food sources and the production of pro-inflammatory TMAO, the composition of gut microbiota plays an important role in modulating this process. The aim of this review is to explain how the gut microbiota interacts with the conversion of specific compounds into TMA and its oxidation to TMAO. We will first cover the correlation between TMAO and various pathologies such as dysbiosis, then focus on cardiovascular disease, with a particular emphasis on pro-atherogenic factors, and then on systemic inflammation and gastrointestinal cancers. Finally, we will discuss primary prevention and therapies that are or may become possible. Possible treatments include modulation of the gut microbiota species with diets, physical activity and supplements, and administration of drugs, such as metformin and aspirin.

Keywords: trimethylamine N-oxide (TMAO); trimethylamine (TMA); gut microbiota; diet; choline; L-carnitine; gastrointestinal cancer; colorectal cancer



Citation: Tacconi, E.; Palma, G.; De Biase, D.; Luciano, A.; Barbieri, M.; de Nigris, F.; Bruzzese, F. Microbiota Effect on Trimethylamine N-Oxide Production: From Cancer to Fitness—A Practical Preventing Recommendation and Therapies. *Nutrients* **2023**, *15*, 563. <https://doi.org/10.3390/nu15030563>

Academic Editor: Emile Levy

Received: 29 December 2022

Revised: 11 January 2023

Accepted: 13 January 2023

Published: 21 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The interest in human microbiota and its modulation of interactions between food sources and some pathologies such as the metabolic syndrome, cardiovascular disease and some types of cancer has been growing in the scientific world. Even if there is a long way to go, our attention has been directed to a molecule involved in systemic inflammation, called trimethylamine N-Oxide (TMAO) [1–3]. TMAO is an amino oxide, produced from the trimethylamine (TMA) through oxidation by some liver enzymes called flavin monooxygenases 1 and 3 (FMO1 and FMO3). Three forms of the enzyme, FMO1 found in fetal liver, FMO3 found in adult liver, and genes clustered in the 1q23-q25 region encode FMO3. Flavin-containing monooxygenases are NADPH-dependent flavoenzymes that catalyze the oxidation of soft nucleophilic heteroatom centers in xenobiotics, such as pesticides and drugs. The human FMO3 enzyme catalyzes several types of reactions, including the N-oxygenation of primary, secondary, and tertiary amines [4]. TMA production is indirectly influenced by some specific compounds such as L-carnitine, choline

and other isoforms, betaine, and lecithin, and directly from gamma-butyrobetaine [2,5,6]. These compounds are metabolized in the gut through interactions with some of microbiota bacteria by different enzymes. First, the major clusters of TMA production start from the mouth with *Streptococcus sanguis* and the genes CutC and CutD, that are necessary for *Desulfovibrio* and *Desulfovibrio desulfuricans* to convert choline in TMA [7]. Other genes such as CntA and CntB contained in *Actinobacter* and *Serratia*, promote the oxidoreductase enzymes from L-carnitine to TMA. Furthermore, YeaW and YeaX are involved in oxygenase and oxidoreductase enzymes for choline, betaine, L-carnitine and gamma-butyrobetaine. Bacteria from gamma-proteobacteria like *Escherichia coli*, *Citrobacter*, *Klebsiella pneumoniae*, and *Shigella*, *Achromobacter* from the strain of *Betaproteobacteria*, *Sporosarcina* from *Firmicutes*, and *Actinobacteria* have orthologue and homologue enzymes such as CntA, CntB, YeaW and YeaX, which encode the gene that can convert all food compounds including choline, betaine, lecithin, gamma-butyrobetaine, ergothioneine and L-carnitine into TMA [7].

Choline, choline esters (e.g., phosphatidylcholine) and lecithin, all important and essential nutrients for the nervous system health [8,9], are converted in TMA by TMA lyase; also, choline can be oxidase in betaine by choline dehydrogenase and choline aldehyde dehydrogenase, which is converted to TMA by betaine oxidoreductase [10]. Furthermore, lecithin, which is a source of choline, can be re-converted to choline by phospholipase D and again converted to TMA [9].

L-carnitine is oxidized directly to TMA by the carnitine oxidoreductase, and indirectly from gamma-butyrobetaine [11]. L-carnitine can be converted by gamma-butyrobetaine hydroxylase in gamma-butyrobetaine, and eventually in TMA by TMA lyase [12]. When the conversion from TMA to TMAO is inhibited, it manifests a particular metabolic syndrome called trimethylaminuria, or “fish odor syndrome”, due to the accumulation of TMA molecules excreted in the urine, sweat and breath which smell like putrid fish [13]. This pathology occurs with a mutation in the gene encoding the liver enzyme FMO3 [14], single nucleotide polymorphism E158K and E308G, which has 10-fold higher specific activity to convert TMA in TMAO than the FMO1 [15]. In some patients, low choline food sources have been recommended [16], but a promising future therapy is to intervene on gut microbiota to modulate the production of TMA and TMAO [17]. Strong evidence correlates high levels of L-carnitine with high levels of TMAO and its potential pro-atherogenic role [18,19] and its role was confirmed by a meta-analysis showing that all causes of mortality increased by 7% per each 10 $\mu\text{mol/L}$ increment of circulating TMAO [20]. Several studies reported a remarkable increment of TMAO achieved by supplementation of L-carnitine, but some of them observed alterations of cardiovascular disease (CVD) markers [21–25]. A diet rich in animal protein, those with high fat content, such as processed and unprocessed meats [26], containing compounds such as choline, carnitine, but even betaine and lecithin in plants usually consumed [27], can produce molecules of TMA by microbiota interaction which can be eventually converted in TMAO [28]. Another molecule in meats, from liver and kidneys, mushrooms, and several type of beans, which is directly involved in TMA production, is ergothioneine, which is converted in TMA through ergothionease enzyme [10]. The TMA molecules are produced directly or indirectly mainly from food compounds containing L-carnitine and choline, but also from betaine and lecithin, ergothioneine and gamma-butyrobetaine. TMAO production is not a clear consequence but can potentially be influenced by dysbiosis and individual polymorphisms in the expression of FMO3 in the liver [14]; as well as the intake of fish and other seafood in the diet [26], and a more systemic disease [29,30]. As well as genetic factors and environmental factors, recent evidence suggests that even the metabolites deriving from the microbiota can play a protective role or promote the onset of tumors. These bacteria produce toxic metabolites, such as secondary bile salt from primary bile salt, hydrogen sulfide, trimethylamine-N-oxide (TMAO) from choline, indoxyl sulfate from amino acid tryptophan, and many more which are likely to promote inflammation, and prolonged inflammation can develop into cancer [5,7,31,32]. The potential role of several gut bacteria metabolites may cause localized

inflammation in normal tissue of colon and promote the genotoxicity of intestinal epithelial cells, determining dysplasia and finally, colorectal cancer (CRC) [1,33].

2. TMAO in Physiological Conditions

TMAO molecules are produced in the liver, enter in the blood stream, from where most of them are excreted with urine within 24 h [7], and some can be reconverted in TMA by TMAO reductase [29]. In the intestine, the TMA conversion starts from choline and its isoform with a specific glyceryl-radical-enzyme (GRE), GRE choline TMA-lyase (cutC) and its precursor GRE activase (cutD) [34] and from L-carnitine and gamma-butyrobetaine with Rieske-type oxygenase/reductase (cntA/B) [11]. The production of TMAO mainly depends on its pre-substrate TMA and the expression of liver enzymes FMO1 and mostly FMO3 [28]. TMAO levels also depend on genetic factors such as the presence of the E158K and E308G polymorphism on FMO3 [14]; another gene, founded in mice, called *Slc30a7*, associated with a zinc transporter (*ZNT7*) [35], seems to be correlated with plasma TMAO levels [36]. Although, as some studies report, the genetic factors in human associated with TMAO plasma levels are more complex [37] and correlated with pathological environment, such as CVD and comorbidities [38–40]. Some food sources have free TMA molecules that are absorbed by enterocytes by passive diffusion, oxidized, and expelled with a 3:95 TMA:TMAO ratio through urine (95%), feces (4%) and breathing (1%) [7]. The FMO3 enzyme is expressed in lungs, adrenals, and aorta. There is a gender difference in TMA activity, which is greater in female than male rats [41]. In males, the predominant steroid, testosterone, is responsible for a lower expression of FMO3 in the liver, whereas high levels of estrogen seem to elevate it [41], but a cohort study underlines that males have more TMAO levels than females [42]. However, some studies do not confirm a hormonal influence nor detect any significant sex differences in the levels of circulating TMAO [43–45]. Many other factors potentially mediate the production of TMAO from TMA, e.g., age, pathological status (CVD, low-grade inflammation, diabetes, tumors, genetic polymorphisms, protein-specific transport; sedentary life and inadequate nutritional status that may lead to gut dysbiosis). The production of TMA in physiological condition occurs through the interactions of compounds and some strains of gut microbiota. An in vitro study observed that the production was mainly by *Firmicutes* and *Proteobacteria* phyla, but not *Bacteroidetes* [46]. Yet other studies reported that all the production of TMA from choline and L-carnitine resulted from *Firmicutes*, *Proteobacteria* phyla, and none from *Bacteroidetes* [34,47]. One week with an antibiotic treatment to suppress some strains in the human gut leads to a reduction of TMAO levels, even with an L-carnitine [30] and choline supplementation [48]. From these studies, one could conclude that the amount of TMAO circulating is not only modulated by the ingestion of choline and L-carnitine food sources, but also by gut strains, in particular “high TMAO producers” with a high *Firmicutes*/*Bacteroidetes* ratio (roughly 2:1) [49]. In some studies, it was reported that a healthy gut microbiota in the adult is the one that has a high *Firmicutes* percentage on *Bacteroidetes* or roughly equal relative percentage [50,51]. Sometimes in other studies, it was found that obese people have a higher level of *Bacteroidetes* than thin individuals and a decrease of this strain has been seen after a period of caloric restriction [52]. Other gut microbes who involved in the TMA production are *Deferribacteraceae*, *Anaeroplasmataceae*, *Prevotellaceae* (*Bacteroidetes* phylum abundant in subjects consuming mostly starchy carbohydrates and fiber [53]), and *Enterobacteraceae* [34,54]. Furthermore, an increased TMAO has been found in those who consume a large amount of seafood per day. Similar levels were also found after the consumption of fish rich in omega-3 polyunsaturated fatty acids (e.g., salmon) [55], which may be due to their high content of free TMAO molecules [56]. Half of free TMAO intake from food is absorbed and eliminated in urine, the other half may be converted in TMA by the enzyme TMAO reductase [10]. Whether TMAO levels are correlated with milk and dairy products remains controversial [44]. Another interesting point is the link between the athlete/sportive amateur’s gut microbiota and TMAO production. It is well known that physical activity influences the population of gut microorganisms in a positive way,

but it is quite evident that TMAO levels are also increased after exercise and sports [57]. Maybe this is a consequence of an omnivorous diet or one medium/high in animal protein that most athletes and sportive amateurs follow. The use of some ergogenic types of supplements such as L-carnitine [18] or choline isoforms (e.g., choline bitartrate) [58] may also contribute to high TMAO levels. However, TMAOs levels are not high in individuals on lacto-ovo vegetarian diet (egg, yolks and whole dairy products included) or vegan diet [30]. Nowadays, many professional and amateur athletes are switching to vegetarian and vegan diets, which could be a reason why TMAO plasma levels are low in these subjects [59]. Most of the caloric intake of athletes comes from carbohydrates, mainly starchy foods, such as whole grain cereal and tubers, but even fruits rich in fibers and oligosaccharides [60] that influence the gut microbiota by increasing *Bacteroidetes* phylum such as *Prevotellaceae* family and *Actinobacteria*. These families are partially responsible of TMA derived compounds conversion, but more importantly, they produce short-chain fatty acids (SCFA) [47]. In some studies, *Firmicutes* appears to be high in athletes and it is the main phylum implicated in TMA conversion [34,46,61]. Wolyniec et al. investigated TMAO levels before and after amateur runners' 10 km or 100 Km ultramarathon there was no significant change in the levels, but only an acute increment. The only significant change was a 3.9-fold increase of TMAOs in the fastest runners of the 100 km race. The authors speculated that TMAOs level may impair the runner's performance [62]. Certainly, high intensity physical activity requires a massive turnover of enzymes and substrates, leading to a huge production of metabolites, e.g., many L-carnitine isotypes that may influence TMA and TMAO conversion [63]. The athlete/sportive amateurs' gut microbiota frequently shows some controversial aspects. Some studies report claims that physical activity reduces *Firmicutes* but increases *Bacteroidetes*, possibly due to the influence of the diet rich in whole grain carbohydrates, fruits, and vegetables [53,64]; others report the opposite effects because *Firmicutes* seems to be high in high caloric intake as most of athletes do [65]. It is undeniably controversial that some extreme physical activity shows an increment of TMAO levels, which already know that are strongly associated with CVD, obesity and type II diabetes, low-grade inflammation, gastrointestinal cancer [42], but at the same time reduce all these pathologies [66–71].

3. TMAO in Pathological Conditions

3.1. TMAO in Atherosclerosis and Cardiovascular Disease

Numerous studies indicate that gut microbiota is involved in the pathogenesis and progression of various cardiovascular diseases (CVD), such as heart failure (HF). HF causes changes in the composition of the intestinal microflora, which may affect the circulating levels of TMAO in human body. Researchers suggested intestinal strains, from *Firmicutes* and *Proteobacteria* phyla, which can produce TMA, such as: *Anaerococcus hydrogenalis*, *Clostridium asparagiforme*, *Clostridium hathewayi*, *Clostridium sporogenes*, *Escherichia fergusonii*, *Proteus penneri*, *Providencia rettgeri*, and *Edwardsiella tarda*. The strains of these bacteria show an increased proportion in patients with HF. This indicates that changes in intestinal microbiota may affect TMAO levels by regulating TMA synthesis in the intestines [72]. Direct and indirect active roles of TMAOs in atherosclerosis and CVD are well established [5,30,39,42], also their associations in obesity and diabetes mellitus [19,29,39,73,74] as well as in low-grade inflammation [3,75] that are often are comorbidities. All of these may eventually also develop cancer [1,33,76–78]. The high levels of circulating TMAO in blood are strongly correlated to cardiovascular events such as stroke, myocardial infarctions, peripheral artery disease, acute coronary syndrome, and atherosclerosis [38,48,79,80]. Indeed, TMAO levels are correlated with the size of aortic atherosclerotic plaque, and they play a pivotal role as a pro-atherogenic factor. TMAO participates actively in the early stage of atherosclerotic process by promoting the macrophages migration, contributing to foam cell formation in the arterial intima [30,81]. Furthermore, high TMAO levels lead to an accumulation of ox-LDL particles within the macrophages by upregulating CD36 and scavenger receptor SR-A1 (Scavenger Receptor A1). These scavenger receptors are responsible for the trans-

formation of macrophages into foam cells [82–85]. At the same time, the high levels of TMAO increase the expression of inflammatory cytokines, such as TNF- α and IL-6, that promote the migration of macrophages and their accumulation in arterial intima [82,86]. TMAO molecules seem to be involved even in endothelial dysfunction, a prelude to cardiovascular disease [7,36,81,87]. Vascular endothelial damage was observed in mice fed with a choline-rich diet [88], and high TMAO levels were associated with an increased systemic inflammation, oxidative stress, and fewer circulating endothelial progenitor cells (EPCs) [89–93]. EPCs with a “spindle-shape” morphology, can take up acetylated LDL particles (acLDL) [94], whereas high TMAO levels reduce this. The formation of acLDL in healthy subjects remains controversial, but it is often used as a model of oxidized or glycated LDL taken up by scavenger receptors. The potential accumulation of oxidized LDL particles is a precursor of inflammation and CVD [95]. TMAO can directly and indirectly activate inflammatory signals such as NF- κ B in aortic endothelial cells, and thereby contribute to atherosclerosis [96]. Furthermore, TMAOs reduce the expression of anti-inflammatory cytokines such as IL-10 that can protect the endothelial tissue from damage and inflammation, block the activity of NF- κ B [97], and inhibit the adhesion of monocytes on the endothelial cells by downregulating the expression of CD18 and CD62-L on immune competent cells [98]. Finally, TMAOs can increase oxidative stress and reduce the endothelial nitric oxide synthases (eNOS) [99]. Oxidative stress is one of the most powerful promoters of atherosclerosis [87,96], whereas eNOS is a protective factor of endothelial health [100–102]. TMAO may have direct or indirect effects on (i) promotion macrophages migration and foam cells forming in the arterial intima; (ii) the accumulation of ox-LDL in situ within the macrophages; (iii) the reduction of EPCs, IL-10, eNOS, with the consequent rise of NF- κ B, oxidative stress, and the LDL particles level in blood. All these effects lead to endothelial damages and cardiovascular dysfunction [103].

3.2. TMAO in Chronic Kidney Disease

High TMAO levels were found in patients with a chronic kidney disease (CKD). In patients with CKD treated with hemodialysis, pre-dialysis TMAO plasma levels were $77 \pm 26 \mu\text{M/dL}$, whereas in control group $2 \pm 1 \mu\text{M/dL}$, and other studies confirm that TMAO plasma levels of CKD patients are up to 40 times greater than normal [7,73,104]. CKD is also associated with CVD or a low-grade inflammation. All these show high levels of circulating TMAOs [20]. Missailidis et al. suggest that after a renal transplantation, the levels of TMAO were normalized [105], indicating not only a crucial role of the kidney in the excretion of these molecules, but also in their production. Indeed, some authors report a strong link between TMAO circulating levels, events of CVD and an effect on kidney health [105–107], with a direct connection with a low renal function [42], because there is a transporter called organic cation transporter 2 (OCT2) in basolateral kidney membrane that is responsible for the uptake of TMAOs [108]. A recent revision of Dongsheng et al. [109] confirms that the mechanism by which TMAO may enhance renal damage and aggravate nephropathy has not been well established. High TMAO plasma and urine levels may have a negative impact on CKD, due to the activation genes expression in the kidney tissue, but the mechanism is not known yet.

3.3. TMAO and Type II Diabetes

Several studies associate high plasma levels of TMAO and T2D or prediabetes [19, 29,39,73,74,110–113], but none has proven a direct cause–effect. In mice feed with high fat diets plus 0.2% of TMAO molecules, it seems to impair glucose tolerance by affecting gene expression on insulin pathways in the liver and by increasing mRNA levels of pro-inflammatory cytokines in adipose tissue [114]. A bidirectional Mendelian assessment reports that high TMAO levels in T2D patients is more like a consequence [115]. For now, we only conclude that the overall microbiota disproportion found in obese patients with pre-diabetes or T2D can lead to high plasma levels of TMAO [74,110–113].

3.4. TMAO, Microbiota Homeostasis and Cancer

The latest literature proved how the gut microbiota plays an important role in gastrointestinal cancer (GIC) [116]. Nowadays, the human microbiota is considered an organ that communicates in synergy with the other apparatus and with many physiological influences. Therefore, its homeostasis, called *eubiosis*, must be protected. When the microbiota is compromised, or when the proportions of microorganisms in it are modified, a syndrome called dysbiosis occurs [116]. Approximately 90% of these microorganisms constituting the microbiota belong to *Firmicutes*, *Bacteroidetes*, *Proteobacteria* and *Actinobacteria*. Viruses, eukarya, fungi, blastocystis, amoebzoa and archea [117–121] represent the other components. A good and healthy microbiota is considered necessary to regulate the immune function, intestinal mucosal protection, vitamins production, correct digestion, and nutrients absorption [122]. The most important phyla correlating with a healthy gut and, consequently, overall health, are the *Firmicutes* and *Cytophaga-Flavobacterium-Bacteroides* (CFB) that are indeed from *Bacteroidetes* phyla [118]. Dysbiosis is not only a difference of the microorganisms' proportion, but it is correlated with various pathologies with different nature such as being overweight and severe obesity, which may carry over even to cardiovascular events, insulin resistance and type II diabetes [74]. A microbiota disproportion was founded and confirmed in some mental pathologies [123,124], such as depression, that it is important to include in the context of other pathologies which affect the digestive apparatus, such as inflammatory bowel disease (IBD) [125], liver disease [126], leaky gut and the intestinal mucosal function [127,128], and gastrointestinal cancer [116,129,130]. Gastrointestinal cancer is one of the most common neoplasia all over the world [131], and lot of data show how the microbiota interacts with this pathology [116,132–136]. Obesity and low-grade inflammation are two main factors which can lead to cancer development [75], and there is a strong correlation within obesity, low-grade inflammation, dysbiosis and colorectal cancer (CRC) [129,132,136,137].

In patients with gastrointestinal cancers, the most abundant family of microorganisms are the *Enterobacteriaceae* that are situated in the small intestine. A low presence of *Lactobacillaceae* and *Acidoaminococcaceae* is typical of colon cancers and *Bifidobacteriaceae* in those of the rectum [138]. At the species level, *Bacteroides fragilis* seem to account for many colorectal cancers and *Helicobacter pylori* for gastric cancers [116,129,139,140]. Even *Escherichia coli* [141] and *Enterococcus faecalis* [142] may be involved in cancer development by DNA mutation genes. Some studies have shown that chemotherapy in patients with GIC leads to a modification/restoration of microbiota dysbiosis [143], a much greater richness in *Lactobacillaceae* has been found after therapy, compared to untreated controls [138]. Another genus correlated with CLC, *Fusobacterium*, showed a significative reduction after debulking surgery, but not chemotherapy [144]. At last, in a very recent study that compared the microbiota composition in obese patients with CLC, non-obese patients with CLC and a healthy control, shows enormous differences in diversity and richness of the gut species [145]. A reduction in richness was found in both CLC groups in comparison with the healthy group, but much more incisive was the decrease in diversity. The major phyla detected in healthy group was *Bacteroidetes*, with more than 50%, whereas it was below 30% in the CLC groups. *Firmicutes* was 39–43% in CLC groups and about 21% in the healthy one, and *Fusobacterium* was 9.1% and 1.2%, respectively. In conclusion, there is strong evidence for a reduction of species which produce SCFA (*Butyricimonas*, *Roseburia*, *Blautia*, *Faecalibacterium*, and *Ruminococcus*), and an increase of pathogenic and induced-cancer ones (*Fusobacterium*, *Clostridium*, *Prevotella*, *Desulfovibrio*, and *Enterococcus*) [145]. In this context, the TMAOs molecules may be implicated too. An increased level in TMAO concentration may be caused by diet, changes in the composition of intestinal microflora, gut dysbiosis or impairment of the gut–blood barrier. Studies on mice have shown that intestinal bacteria are essential to convert dietary compounds to TMA [146]. The production of TMA and TMAO can be almost completely suppressed using broad spectrum antibiotics, and after one month of withdrawal of antibiotics, the TMAO concentration returns to normal [147]. Indeed, some studies have associated the high levels of TMAO with high TNF-alpha, IL-6,

C-reactive protein [44], pro-inflammatory cytokines, IL-1beta [96] and even as a coadjuvant of *Helicobacter pylori* to promote infection in the gastric epithelial cells, increasing the activity of IL-6 and chemokine ligands. This suggests a potential link between TMAO and gastric cancer via the inflammatory process. Consistent with this, Yue et al. also demonstrated that TMAO can trigger the activation of the nod-like receptor family pyrin domain containing 3 (NLRP3) inflammasome [148,149], which was suggested to be implicated in the growth and/or metastasis of a variety of cancers including head and neck cancer, oral cancer, lung cancer, prostate cancer, and colorectal cancer [150,151]. Association studies provided further support for a link between TMAO and inflammation. The serum level of TMAO was shown to be positively correlated with the level of certain pro-inflammatory mediators including tumor necrosis factor-alpha (TNF- α) and IL-6. Moreover, studies have also displayed that an enhanced level of TMAO prompted the initiation of NF-Kappa-B route and improved the expression of pro-inflammatory genes involving chemokines, adhesion molecules and inflammatory cytokines [152]. Several data show, indeed, how the TMAO levels are quite high in patients with colorectal cancer and even higher in obese subjects with the same type of cancer [145,153,154]. Moreover, the presence of high quantity of *Firmicutes phylum*, *Prevotellaceae*, *Enterobacteraceae* and *Desulfovibrio* that can increase the conversion of choline to TMA by the expression of the cutC gene [34], and low levels of *Bacteroidetes phylum* [145] in relation to healthy controls with neither cancer nor obesity, are very illuminating and confirm the role of TMAO. This would indicate reducing all foods containing choline, betaine, lecithin, gamma-butyrobetaine, ergothioneine and L-carnitine, to avoid a massive conversion to TMAO [1]. Conversion of these compounds in TMA not only depends on the food sources [4,24,29], but also on microbiota composition [42] which is compromised by pathological status [129,137,155]. It is now clear that many gastrointestinal pathologies are related to bacteria families of *Firmicutes phylum* [49], which even includes healthy strains of gut microbiota [50,51]. TMAO is frequently used as a risk marker of diseases, but its usefulness as a standalone marker is limited due to the high intra individual variability that may reflect dietary changes from day to day, in particular the intake of meat or fish [156]. Oxidative stress might also be one of the factors linking TMAO and cancer. Recent studies showed that TMAO could be implicated in oxidative stress and increased circulating TMAO was shown to induce superoxide production, a reactive oxygen species (ROS) linked to oxidative stress. In an in vitro assay, TMAO was also shown to stimulate the production of ROS in cells [157–159].

4. Practical Recommendations for Prevention and Treatments Reducing TMAO Production

4.1. Diet

To modulate the TMA and TMAO levels, the manipulation of food intake and diet is probably the best/most promising initial treatment. The targets are primarily choline, betaine, lecithin, gamma-butyrobetaine, ergothioneine and L-carnitine [3], naturally found in animal protein, processed and unprocessed meat, egg yolk, dairy products, and even fatty fish [26]. However, some of these compounds are essential nutrients, which eventually may necessitate supplementation, if their intake is inadequate. High fat, high protein or Western diets contain/are associated with high levels of TMAO [3], even when supplemented with fish oil [160]. In contrast, inclusion of some nuts such as pistachios [161] or some indigestible fiber [162], or vegetarian diets [30], tended to attenuate TMAO production.

Choline is a component of choline phospholipids, which are essential components of cell membranes [163,164]; it is also a precursor of acetylcholine, which acts as neurotransmitter [165] and plays a pivotal role in the correct development of brain cells such as astrocytes [166]. Furthermore, its presence is required in other tissues to interact with hormones, growth factors and neural cells [167]. Although it can be produced endogenously, dietary supplementation is necessary in certain conditions, such as vegan diets, pregnancy, diets very low in protein sources, or parenteral nutrition [163,164,168–170]. The recommended choline intake is 7 mg/kg/day for adults, 450 mg/day for women throughout

pregnancy, and up to 550 mg/day to support breastfeeding [171]. An inadequate intake of choline and betaine is linked to pro-atherogenic changes [172,173]. For example, betaine is required to homocysteine methylation to methionine, and its insufficiency leads to a high circulating level of homocysteine [174], which is correlated, with CVD [175], cancers [176], neurodegenerative disease [177] and osteoporosis [178]. There is an interesting hypothesis of choline/1-carbon (betaine) crosstalk metabolism, in which low intake of choline and betaine may interact with several mitochondrial pathways having an impact on the systemic insulin sensitivity of muscle and adipose tissues, impairing the body composition, energy homeostasis and thus the health status [179]. A “sweet pot” daily dose of choline and betaine seems to be associated with better body composition [180]. The choline food sources are egg yolk, whole milk and whole dairy products, beef, pork, liver, seafood and fatty fish [9], whereas for betaine whole grains, shellfish beets and spinach are the common sources [181]. Apart from the hypothesized crosstalk with choline, betaine itself is another essential nutrient that can be obtained from veggies, shellfish, sugar beet and cereal [27] but is also produced from oxidation of choline in betaine aldehyde by choline dehydrogenase, and betaine aldehyde is converted in betaine by the betaine aldehyde dehydrogenase [182]. The molecule is also known as trimethyl-glycine for its 3 methyl groups; its function is to protect cells from oxidative stress and to add a methyl group, as aforementioned, during the conversion of homocysteine in methionine [181], an important process to low levels of homocysteine in the blood that is a supposed CVD marker [175]. It is established that choline and betaine are precursors of TMA and eventually TMAO, but it is also certain that they are essential nutrients, so it is suggested to cover, but not to exceed the Recommended Dietary Allowance (RDA) of choline. For the betaine, association studies in some populations suggest that its intake is usually half that of choline [180]. The choline and betaine content in food is shown in Table 1. The risks of conversion of choline in TMA by the bacteria *Desulfovibrio* cutC gene expression [34] is tangible, but only choline bitartrate and not phosphatidylcholine seems to raise the TMAO levels and excretion with urine [183] in those who are considered “high TMAO producers” with abundance in microbiota of *Firmicutes* phylum, *Clostridia* class such as *Clostridium*, *Ruminococcaceae* and *Lachnospiraceae* [184]. Since egg yolks contain mostly phosphatidylcholine, 2–3 whole eggs/day (roughly 400 mg choline) result in high levels of choline but not TMAOs in the bloodstream [185,186].

L-carnitine is an amino acid that is essential for the organism at mitochondrial level to transport long chain fatty acids into the matrix to produce energy via beta-oxidation [187]. Supplementation is not required to maintain a physiological level, because the body can produce the right amount endogenously from lysine and methionine [187] and the kidneys reduce or increase the excretion based on current blood levels [187,188]. L-carnitine is easily converted into TMA in gut microbiota by the 2-component Rieske-type L-carnitine oxygenated CntA/B in the presence of oxygen molecule [11] and is strongly correlated with an increase of TMAO levels [12,30]. L-carnitine is contained mostly in red meat and for a very small part in white meat (poultry) [189]. A diet rich in red meat results in a high level of L-carnitine and gamma-butyrobetaine and consequently in high TMAO levels [1,11,18,21,23–25,30,44,189,190]. Vegetarians, who include in their diet whole dairy products and whole eggs are less predisposed to convert L-carnitine into TMA, even with a supplementations protocol [30]. This may be due to their microbiota composition [30] or their low levels of L-carnitine from the meatless diet. Since L-carnitine supplementation shows no benefits at all, with an exception for those who have a specific deficiency [18], the most recent international guidelines against cancers [191] recommend the entire population, for patients with CVD, type II diabetes, CKD, and cancer, to avoid supplements of any form of carnitine and to limit all foods containing it [1]. At last, a diet very rich in fat, a normal Western diet or a diet mainly composed of protein and fat with low or no sources of carbohydrates and fibers (e.g., ketogenic diet, very low carb diet) leads to an acute postprandial and chronic production of TMAOs, by changing the ratio between *Firmicutes* and *Bacteroidetes* in favor of the former [3]. Poor quality diets also create dysbiosis [122].

The classical beneficial Mediterranean diet does not lower TMAOs levels after six months of intervention, maybe because of the fish intake that directly brings free TMAOs molecules into the organism. Indeed, as Griffin et al. suggest, it would probably be useful to investigate whether the Mediterranean or another adequate healthy diet along the anti-cancer international guidelines, could affect TMAOs serum levels with or without fish [192].

Table 1. Betaine, phosphocholine and free choline raw food contents in g for 100 g [1].

	Betaine	Free Choline	Phosphocholine
1-4			
Egg yolk	0.0	2.38	5.84
<i>Milk</i>			
Whole 3.25% Fat	0.6	3.7	1.8
2% Fat	0.9	2.8	1.6
<i>Butter</i>	0.3	0.5	0.7
<i>Cheese</i>			
Cheddar	0.7	1.6	0.6
Cottage 2%	0.6	2.9	1.3
Mozzarella	0.7	2.3	0.9
Swiss	0.6	4.5	0.0
<i>Chicken</i>			
Meat and skin	7.8	6.0	3.6
Liver	16.9	49.2	4.1
<i>Pork</i>			
Sausage	3.4	8.0	0.5
Bacon	0.9	4.4	1.4
Loin, lean only	2.4	1.6	2.2
<i>Beef</i>			
Ground 80% lean	8.2	2.6	0.4
Liver	4.4	56.2	11.8
<i>Fish and seafoods (cooked)</i>			
Cod (Atlantic)	9.7	17.7	1.6
Shrimp	33.0	1.5	0.8
Salmon (Sockeye)	2.1	8.6	1.1
Tilapia	25.3	21.4	2.5
Tuna (canned in water)	2.7	2.1	0.0
<i>Vegetables</i>			
Beets	128.7	4.1	0.9
Broccoli	0.1	18.1	0.4
Cabbage	0.4	6.1	2.3
Carrots	0.4	6.8	1.1
Lettuce iceberg	0.1	4.8	1.5
Mushrooms	10.7	5.9	1.3
Potato, white, flesh and skin	0.2	7.9	0.3
Spinach boiled, drained	726	1.7	1.1
Tomato paste	0.4	26.2	4.3
Tomato	0.1	4.4	1.8
<i>Nuts (roasted and dried)</i>			
Almonds	0.5	9.4	1.9
Brazilnuts	0.4	16.1	0.3
Cashews	11.2	19.6	0.9
Hazelnuts	0.4	15.2	0.9
Macadamia	0.3	11.3	1.0
Pecans	0.7	9.7	1.3
Pine	0.4	8.4	2.1
Pistachio	0.8	10.7	8.5
walnuts	0.5	8.3	0.5
<i>Legumes</i>			
Beans, kidney, canned	0.1	19.7	0.5
Peanut butter, smooth	0.4	25.8	0.7
Soy milk	0.8	13.1	3.4
Soy sauce (shoyu)	39.6	31.0	0.0
<i>Cereal grains and others</i>			
Oat bran	35.7	4.4	0.7
Rice, brown	0.5	4.7	0.0
Pasta, dry	460	9.7	0.0
Wheat flour, white	124.4	5.7	0.1
Bread, wheat	85.2	11.5	0.3
Kellogg's all-bran	360.0	25.5	1.7

4.2. Drugs, Supplements and Physical Activity

An adequate diet rich in whole grains, natural starchy foods, fruits, and vegetables, containing moderated choline sources and low to no L-carnitine would seem to be the first step to reduce circulating TMAOs by directly decreasing those compounds that convert to TMA. Probiotics and prebiotics could be used to modify microbiota targeting TMAOs. Indeed, the administration of probiotic *Lactocaseibacillus paracasei* [193], but not *Lactocaseibacillus casei* [194] or *Lactiplantibacillus plantarum* [195] have demonstrated to reduce circulating TMA and TMAO levels; *Enterobacter aerogenes* ZDY01 increased *Bacteroidales* (*Bacteroidetes* phylum) and decreased *Prevotellaceae* and *Helicobacteraceae* families [196], and supplementing with *Archeobacteria* phylum, which subtract methyl compounds required to form TMA and decrease in TMAO [17,197]. In contrast, prebiotic supplementing with *Arabinoxylan* oligosaccharide plus vitamins B and D showed only a little reduction in serum TMAO [198]. Using a prebiotic such as resveratrol, *Bacteroidetes* phylum is increased, while *Firmicutes*, aside *Lactobacillus* and *Bifidobacterium* genus, decreased along with TMAO plasma levels [199,200]. Therefore, a prudent approach would be to evaluate probiotics and prebiotics as adjuvant therapy with diet on TMAOs modulation. Physical activity also has an important effect on the composition of gut microbiota [201,202]. Endurance training, for example, lowers the *Proteobacteria* and enhances *Akkermansia muciniphilia* [203], decreases *Clostridium difficile*, increases *Oscillospira* [204] and augments beneficial short-chain fatty acids in lean people [205]. As the intensity and volume of physical activity increases, such as in professional athletes or the military, the positive effects on gut microbiota becomes inversely proportional [206,207], with an increment of potentially pathogenic *Staphylococcus*, *Peptostreptococcus*, *Peptoniphilus*, *Acidaminococcus* and *Fusobacterium*, and a reduction of potentially beneficial strains [208]. However, prebiotics and probiotics are/seem indicated in athletes with compromised immune function, upper respiratory tract illnesses (URTIs), or gastrointestinal disorders such as diarrhea, bloating, abdominal pain or gastroesophageal reflux, all of which strongly correlate with dysbiosis [206,209]. Even though physical activity and sports in general does not seem to affect the gut microbiota and TMA conversion, they remain correlated to an overall beneficial impact on various conditions that favor gastro-intestinal cancer development [66,68,69,71,206]. A highlight could be put on the *Akkermansia muciniphilia*, which is increased by physical activity [202,203], administration of metformin [210,211] and berberine [212–216]. *Akkermansia muciniphilia* appears to be a promising strain that can be beneficial in gastro-intestinal cancer and in the management of glucose in obese patients [50,132,217,218].

Antibiotics have a great impact on the gut microbiota. They are the most incisive drug that can block TMAOs production, but at the same time, they kill other beneficial microorganisms.

Ciprofloxacin and metronidazole are the most effective suppressors of TMAOs, but after just one month of use, TMAO levels rise again [48]. Even a mix of various broad-spectrum antibiotics such as vancomycin, neomycin-sulphate, metronidazole and ampicillin block the conversion of choline into TMA, and therefore reduce TMAO, but result in relatively fast development of antibiotic-resistance and extinction of other beneficial phyla [6,48].

The compound 3, 3-Dimethyldimethyl-1-butanol (DMB), a natural compound derived from vinegar, olive, and grapeseed oil, could be used to limit the conversion of choline, betaine and L-carnitine to TMA with imbibition of TMA lyase but unfortunately not of gamma-butyrobetaine (GBB) to TMA and neither the FMO3 conversion to TMAO [219]. Other second generation of choline analogues are Flouro-methylcholine and Iodo-methylcholine, which irreversibly block TMA lyase, showed a decrease of TMAOs and thrombotic events too without any toxic effects [220].

Meldonium, another molecule used in ischemic and atherosclerosis events, decreases TMAO levels by blocking the conversion from L-carnitine to TMAO and GBB to L-carnitine [221]. In fact, it leads to an accumulation of GBBs and does not have any effects on choline conversion to TMA [222].

Enalapril is an ACE inhibitor drug that lowers TMAO levels. It increases the excretion of TMAO, possibly by a common sodium mechanism, but does not reduce TMA levels [223].

A study of 2 g/day metformin without diet control did not show an effect on TMAO levels, even though metformin affects the microbiota [224]. In contrast, a very recent study reported that 250 mg/kg metformin for 4–8 weeks lowered TMA and TMAO levels in db/db mice that was treated only metformin and with metformin plus a choline bolus to mimic a typical Western diet [225]. This study also evaluated the effects of metformin on the choline-TMA lyase genes expression (CutD, CutC, cmcA) and FMO3 liver enzymes, and found that metformin had an impact on gut microbiota species that decrease the conversion of substrates to TMA resulting in a decrease of TMAOs [225].

Finally, one study reported that 81 mg aspirin plus choline diet (two whole eggs a day) significantly reduced TMAO plasma levels over 12 months [107,226,227].

5. Conclusions

Different studies have highlighted how the microbiota can be a direct or indirect cause of the onset or progression of various types of pathologies. Foods, or nutrient molecules extracted from the diet, can be powerful modulators of the microbiota. One of these, TMAO, is formed from the precursor of TMA (trimethylamine) through the combined action of the intestinal microbiota and the liver, as we can see in the Figure 1. TMAO directly interferes with hepatic gluconeogenesis and glucose transport, increasing the susceptibility to insulin resistance. A high level of TMAO has been associated with an increased risk of adverse events in cardiovascular disease. Furthermore, TMAO can cause epigenetic changes to DNA and, through the formation of N-nitrous, compounds damage DNA, which can lead to malignant transformations in exposed cells. Before blaming some natural foods, that contain important and useful compounds for our health, it would be right and proper to understand how the intestinal microbiota, individual polymorphism and lifestyle of a subject can modulate the expression of TMAO. Several therapeutic strategies are being studied to reduce TMAO levels, including the use of broad-spectrum oral antibiotics, promoting the growth of bacteria that use TMAO as a substrate and the development of target-specific molecules. Despite the accumulated evidence, one wonders whether TMAO is a spectator's mediator in the disease process. Therefore, it is important to undertake studies to establish the role of TMAO in human health and disease. Recent investigations into the cross-dialogue between GM and human health have opened new approaches for diet-based interventions. Manipulation of gut microbiota through diet is a visionary approach to improve human health. On the bases of this knowledge, we propose that the administration of specific foods could be a new support in daily practice, as shown in Table 2. Based on the current range of carnitine intake (i.e., from 2 to 12 mol/kg/day or from 22.7 to 136.1 mg/day for a 70 kg human being), a standard diet provides enough carnitine, 3.4 times the lower level or 96% of the average recommendation. However, a detailed characterization of both the extent and the mechanisms by which these interactions occur will be necessary. The principle of primary prevention of oncological and cardiovascular diseases is certainly based on the dietary modifications and healthy behaviors. The intake of a specific food category, and the combination with the genetic, epigenetic and characteristics of the microbiota of a single subject has to be considered more frequently in future precision medicine.

Table 2. L-carnitine raw food contents in mg for 100 g [2].

<i>Beef</i>	
Steak	65.0
Ground	87.5
Tenderloin	78.6
T-Bone	84.2
Loin	64.6
<i>Chicken</i>	
Liver	94.0
Meat	10.4
Wing Meat	10.0
<i>Turkey meat</i>	21.2
<i>Lamb Chop</i>	40.5
<i>Pork</i>	
Shoulder	21.1
Ham	53.5
White Ham	33.5
Sausage	7.1
<i>Veal</i>	
Shoulder	78.2
Sirloin	132.8
<i>Milk</i>	
2% fat	2.9
4% fat	2.3
<i>Butter</i>	1.3
<i>Cheese</i>	
Camembert	14.4
Gruyere	6.5
Feta	1.8
Goat cheese	15.3
Mozzarella	0.3
Parmesan	0.7
<i>Yogurt</i>	
Regular	12.2
0% fat	12.5
<i>Egg</i>	
White	0.3
Yolk	0.8
<i>Fish and seafood</i>	
Anchovy	1.8
Shrimp	0.7
Cod (Atlantic)	1.8
Hake (boiled)	2.9
Mussels (cooked)	2.6
Salmon (cooked)	5.8
Smoked salmon	1.0
Tuna	1.5

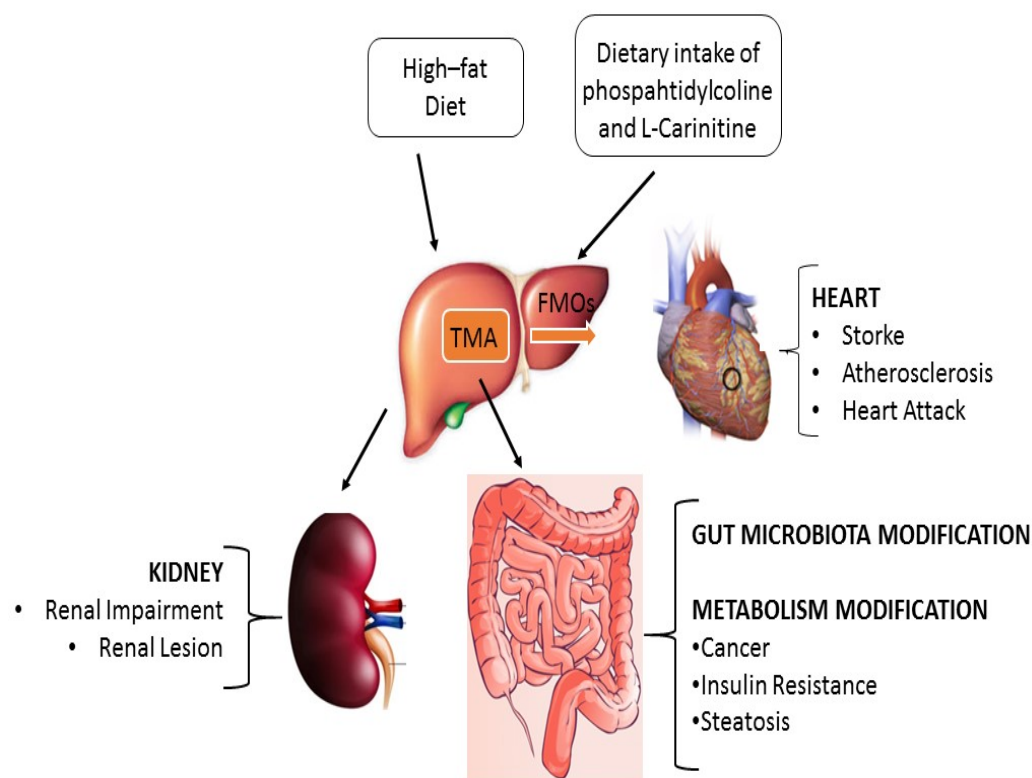


Figure 1. The action of TMA in pathological condition. The dietary intake of choline isoforms, carnitine, gamma-butyrobetaine contribute to increment the plasma level of TMA. TMA are converted in TMAO in the liver. TMAO are involved and correlated with kidney and liver disease, gastrointestinal cancers, diabetes type II, atherosclerosis, and cardiac muscle damage.

Author Contributions: E.T. and G.P. conceptualization and writing original draft, A.L., M.B.—resources, D.D.B. and F.d.N.—investigation and validation, F.B.—supervision and funding acquisition. All authors have read and agreed to the published version of the manuscript.

Funding: Ricerca Corrente Funds to Istituto Nazionale Tumori G. Pascale project Linea 3/12 to G.P.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Edoardo Tacconi is grateful to his mentor Giuseppe Palma, his family and Giulia.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Oellgaard, J.; Winther, S.A.; Hansen, T.S.; Rossing, P.; von Scholten, B.J. Trimethylamine N-oxide (TMAO) as a New Potential Therapeutic Target for Insulin Resistance and Cancer. *Curr. Pharm. Des.* **2017**, *23*, 3699–3712. [[CrossRef](#)] [[PubMed](#)]
- Tang, W.H.W.; Hazen, S.L. Microbiome, trimethylamine N-oxide, and cardiometabolic disease. *Transl. Res.* **2017**, *179*, 108–115. [[CrossRef](#)] [[PubMed](#)]
- Janeiro, M.H.; Ramírez, M.J.; Milagro, F.I.; Martínez, J.A.; Solas, M. Implication of trimethylamine n-oxide (TMAO) in disease: Potential biomarker or new therapeutic target. *Nutrients* **2018**, *10*, 1398. [[CrossRef](#)] [[PubMed](#)]
- Bain, M.; Fornasini, G.; Evans, A. Trimethylamine: Metabolic, Pharmacokinetic and Safety Aspects. *Curr. Drug Metab.* **2005**, *6*, 227–240. [[CrossRef](#)]
- Ahmadmehrabi, S.; Tang, W.H.W. Gut microbiome and its role in cardiovascular diseases. *Curr. Opin. Cardiol.* **2017**, *32*, 761–766. [[CrossRef](#)]
- Jie, Z.; Xia, H.; Zhong, S.L.; Feng, Q.; Li, S.; Liang, S.; Zhong, H.; Liu, Z.; Gao, Y.; Zhao, H.; et al. The gut microbiome in atherosclerotic cardiovascular disease. *Nat. Commun.* **2017**, *8*, 845. [[CrossRef](#)]

7. Zeisel, S.H.; Warriar, M. Trimethylamine N-Oxide, the Microbiome, and Heart and Kidney Disease. *Annu. Rev. Nutr.* **2017**, *37*, 157–181. [[CrossRef](#)]
8. Wiedeman, A.M.; Barr, S.I.; Green, T.J.; Xu, Z.; Innis, S.M.; Kitts, D.D. Dietary choline intake: Current state of knowledge across the life cycle. *Nutrients* **2018**, *10*, 1513. [[CrossRef](#)]
9. Zeisel, S.H.; Da Costa, K.A. Choline: An essential nutrient for public health. *Nutr. Rev.* **2009**, *67*, 615–623. [[CrossRef](#)]
10. Fennema, D.; Phillips, I.R.; Shephard, E.A. Trimethylamine and trimethylamine N-oxide, a Flavin-Containing Monooxygenase 3 (FMO3)-mediated host-microbiome metabolic axis implicated in health and disease. *Drug Metab. Dispos.* **2016**, *44*, 1839–1850. [[CrossRef](#)]
11. Zhu, Y.; Jameson, E.; Crosatti, M.; Schäfer, H.; Rajakumar, K.; Bugg, T.D.H.; Chen, Y. Carnitine metabolism to trimethylamine by an unusual Rieske-type oxygenase from human microbiota. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 4268–4273. [[CrossRef](#)]
12. Koeth, R.A.; Levison, B.S.; Culley, M.K.; Buffa, J.A.; Wang, Z.; Gregory, J.C.; Org, E.; Wu, Y.; Li, L.; Smith, J.D.; et al. γ -butyrobetaine is a proatherogenic intermediate in gut microbial metabolism of L-carnitine to TMAO. *Cell Metab.* **2014**, *20*, 799–812. [[CrossRef](#)]
13. Akerman, B.R.; Lemass, H.; Chow, L.M.L.; Lambert, D.M.; Greenberg, C.; Bibeau, C.; Mamer, O.A.; Treacy, E.P. Trimethylaminuria is caused by mutations of the FMO3 gene in a North American cohort. *Mol. Genet. Metab.* **1999**, *68*, 24–31. [[CrossRef](#)]
14. Treacy, E.P.; Akerman, B.R.; Chow, L.M.L.; Youil, R.; Bibeau, C.; Lin, J.; Bruce, A.G.; Knight, M.; Danks, D.M.; Cashman, J.R.; et al. Mutations of the flavin-containing monooxygenase gene (FMO3) cause trimethylaminuria, a defect in detoxication. *Hum. Mol. Genet.* **1998**, *7*, 839–845. [[CrossRef](#)]
15. Bennett, B.J.; Vallim, T.Q.D.A.; Wang, Z.; Shih, D.M.; Meng, Y.; Gregory, J.; Allayee, H.; Lee, R.; Graham, M.; Croke, R.; et al. Trimethylamine-N-Oxide, a metabolite associated with atherosclerosis, exhibits complex genetic and dietary regulation. *Cell Metab.* **2013**, *17*, 49–60. [[CrossRef](#)]
16. Busby, M.G.; Fischer, L.; Da Costa, K.A.; Thompson, D.; Mar, M.H.; Zeisel, S.H. Choline- and betaine-defined diets for use in clinical research and for the management of trimethylaminuria. *J. Am. Diet. Assoc.* **2004**, *104*, 1836–1845. [[CrossRef](#)]
17. Brugère, J.F.; Borrel, G.; Gaci, N.; Tottey, W.; O’Toole, P.W.; Malpuech-Brugère, C. Archaeobiotics: Proposed therapeutic use of archaea to prevent trimethylaminuria and cardiovascular disease. *Gut Microbes* **2013**, *5*, 5–10. [[CrossRef](#)]
18. Sawicka, A.K.; Renzi, G.; Olek, R.A. The bright and the dark sides of L-carnitine supplementation: A systematic review. *J. Int. Soc. Sports Nutr.* **2020**, *17*, 1–10. [[CrossRef](#)]
19. Heianza, Y.; Ma, W.; Manson, J.A.E.; Rexrode, K.M.; Qi, L. Gut microbiota metabolites and risk of major adverse cardiovascular disease events and death: A systematic review and meta-analysis of prospective studies. *J. Am. Heart Assoc.* **2017**, *6*, 4947. [[CrossRef](#)]
20. Schiattarella, G.G.; Sannino, A.; Toscano, E.; Giugliano, G.; Gargiulo, G.; Franzone, A.; Trimarco, B.; Esposito, G.; Perrino, C. Gut microbe-generated metabolite trimethylamine-N-oxide as cardiovascular risk biomarker: A systematic review and dose-response meta-analysis. *Eur. Heart J.* **2017**, *38*, 2948–2956. [[CrossRef](#)]
21. Samulak, J.J.; Sawicka, A.K.; Hartmane, D.; Grinberga, S.; Pugovics, O.; Lysiak-Szydłowska, W.; Olek, R.A. L-Carnitine supplementation increases trimethylamine-N-oxide but not markers of atherosclerosis in healthy aged women. *Ann. Nutr. Metab.* **2019**, *74*, 11–17. [[CrossRef](#)] [[PubMed](#)]
22. Olek, R.A.; Samulak, J.J.; Sawicka, A.K.; Hartmane, D.; Grinberga, S.; Pugovics, O.; Lysiak-Szydłowska, W. Increased Trimethylamine N-Oxide Is Not Associated with Oxidative Stress Markers in Healthy Aged Women. *Oxid. Med. Cell. Longev.* **2019**, *2019*, 6247169. [[CrossRef](#)] [[PubMed](#)]
23. Bordoni, L.; Sawicka, A.K.; Szarmach, A.; Winklewski, P.J.; Olek, R.A.; Gabbianelli, R. A pilot study on the effects of L-carnitine and trimethylamine-N-oxide on platelet mitochondrial dna methylation and CVD biomarkers in aged women. *Int. J. Mol. Sci.* **2020**, *21*, 1047. [[CrossRef](#)] [[PubMed](#)]
24. Fukami, K.; Yamagishi, S.I.; Sakai, K.; Kaida, Y.; Yokoro, M.; Ueda, S.; Wada, Y.; Takeuchi, M.; Shimizu, M.; Yamazaki, H.; et al. Oral L-carnitine supplementation increases trimethylamine-N-oxide but reduces markers of vascular injury in hemodialysis patients. *J. Cardiovasc. Pharmacol.* **2015**, *65*, 289–295. [[CrossRef](#)] [[PubMed](#)]
25. Vallance, H.D.; Koochin, A.; Branov, J.; Rosen-Heath, A.; Bosdet, T.; Wang, Z.; Hazen, S.L.; Horvath, G. Marked elevation in plasma trimethylamine-N-oxide (TMAO) in patients with mitochondrial disorders treated with oral L-carnitine. *Mol. Genet. Metab. Rep.* **2018**, *15*, 130–133. [[CrossRef](#)]
26. Wallace, T.C.; Blusztajn, J.K.; Caudill, M.A.; Klatt, K.C.; Natker, E.; Zeisel, S.H.; Zelman, K.M. The underconsumed and underappreciated essential nutrient. *Nutr. Today* **2018**, *53*, 240–253. [[CrossRef](#)]
27. Zeisel, S.H.; Mar, M.H.; Howe, J.C.; Holden, J.M. Concentrations of choline-containing compounds and betaine in common foods. *J. Nutr.* **2003**, *133*, 1302–1307. [[CrossRef](#)]
28. Schugar, R.C.; Brown, J.M. Emerging roles of flavin monooxygenase 3 in cholesterol metabolism and atherosclerosis. *Curr. Opin. Lipidol.* **2015**, *26*, 426–431. [[CrossRef](#)]
29. Chhibber-Goel, J.; Gaur, A.; Singhal, V.; Parakh, N.; Bhargava, B.; Sharma, A. The complex metabolism of trimethylamine in humans: Endogenous and exogenous sources. *Expert Rev. Mol. Med.* **2016**, *18*, e8. [[CrossRef](#)]
30. Koeth, R.A.; Wang, Z.; Levison, B.S.; Buffa, J.A.; Org, E.; Sheehy, B.T.; Britt, E.B.; Fu, X.; Wu, Y.; Li, L.; et al. Intestinal microbiota metabolism of l-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat. Med.* **2013**, *19*, 576–585. [[CrossRef](#)]
31. Tremaroli, V.; Bäckhed, F. Functional interactions between the gut microbiota and host metabolism. *Nature* **2012**, *489*, 242–249. [[CrossRef](#)]

32. Kamada, N.; Seo, S.U.; Chen, G.Y.; Núñez, G. Role of the gut microbiota in immunity and inflammatory disease. *Nat. Rev. Immunol.* **2013**, *13*, 321–335. [[CrossRef](#)]
33. Yang, S.; Dai, H.; Lu, Y.; Li, R.; Gao, C.; Pan, S. Trimethylamine N-Oxide Promotes Cell Proliferation and Angiogenesis in Colorectal Cancer. *J. Immunol. Res.* **2022**, *2022*, 7043856. [[CrossRef](#)]
34. Craciun, S.; Balskus, E.P. Microbial conversion of choline to trimethylamine requires a glycy radical enzyme. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 21307–21312. [[CrossRef](#)] [[PubMed](#)]
35. Hartiala, J.; Bennett, B.J.; Tang, W.H.W.; Wang, Z.; Stewart, A.F.R.; Roberts, R.; McPherson, R.; Luskis, A.J.; Hazen, S.L.; Allayee, H. Comparative genome-wide association studies in mice and humans for trimethylamine N-Oxide, a proatherogenic metabolite of choline and L-carnitine. *Arterioscler. Thromb. Vasc. Biol.* **2014**, *34*, 1307–1313. [[CrossRef](#)]
36. Obeid, R.; Awwad, H.M.; Rabagny, Y.; Graeber, S.; Herrmann, W.; Geisel, J. Plasma trimethylamine N-oxide concentration is associated with choline, phospholipids, and methyl metabolism. *Am. J. Clin. Nutr.* **2016**, *103*, 703–711. [[CrossRef](#)]
37. Aslibekyan, S.; Irvin, M.R.; Hidalgo, B.A.; Perry, R.T.; Jeyarajah, E.J.; Garcia, E.; Shalurova, I.; Hopkins, P.N.; Province, M.A.; Tiwari, H.K.; et al. Genome- and CD4+ T-cell methylome-wide association study of circulating trimethylamine-N-oxide in the Genetics of Lipid Lowering Drugs and Diet Network (GOLDN). *J. Nutr. Intermed. Metab.* **2017**, *8*, 1–7. [[CrossRef](#)]
38. Senthong, V.; Wang, Z.; Fan, Y.; Wu, Y.; Hazen, S.L.; Tang, W.H.W. Trimethylamine N-oxide and mortality risk in patients with peripheral artery disease. *J. Am. Heart Assoc.* **2016**, *5*, e4237. [[CrossRef](#)]
39. Randrianarisoa, E.; Lehn-Stefan, A.; Wang, X.; Hoene, M.; Peter, A.; Heinzmann, S.S.; Zhao, X.; Königsrainer, I.; Königsrainer, A.; Balletshofer, B.; et al. Relationship of serum trimethylamine N-oxide (TMAO) levels with early atherosclerosis in humans. *Sci. Rep.* **2016**, *6*, 26745. [[CrossRef](#)]
40. Nie, J.; Xie, L.; Zhao, B.X.; Li, Y.; Qiu, B.; Zhu, F.; Li, G.F.; He, M.; Wang, Y.; Wang, B.; et al. Serum trimethylamine N-oxide concentration is positively associated with first stroke in hypertensive patients. *Stroke* **2018**, *49*, 2021–2028. [[CrossRef](#)]
41. Esposito, T.; Varriale, B.; D’Angelo, R.; Amato, A.; Sidoti, A. Regulation of flavin-containing mono-oxygenase (Fmo3) gene expression by steroids in mice and humans. *Horm. Mol. Biol. Clin. Investig.* **2014**, *20*, 99–109. [[CrossRef](#)] [[PubMed](#)]
42. Manor, O.; Zubair, N.; Conomos, M.P.; Xu, X.; Rohwer, J.E.; Krafft, C.E.; Lovejoy, J.C.; Magis, A.T. A Multi-omic Association Study of Trimethylamine N-Oxide. *Cell Rep.* **2018**, *24*, 935–946. [[CrossRef](#)] [[PubMed](#)]
43. Kühn, T.; Rohrmann, S.; Sookthai, D.; Johnson, T.; Katzke, V.; Kaaks, R.; Von Eckardstein, A.; Müller, D. Intra-individual variation of plasma trimethylamine-N-oxide (TMAO), betaine and choline over 1 year. *Clin. Chem. Lab. Med.* **2017**, *55*, 261–268. [[CrossRef](#)] [[PubMed](#)]
44. Rohrmann, S.; Linseisen, J.; Allenspach, M.; Von Eckardstein, A.; Müller, D. Plasma concentrations of trimethylamine- n-oxide are directly associated with dairy food consumption and low-grade inflammation in a german adult population. *J. Nutr.* **2016**, *146*, 283–289. [[CrossRef](#)] [[PubMed](#)]
45. Krüger, R.; Merz, B.; Rist, M.J.; Ferrario, P.G.; Bub, A.; Kulling, S.E.; Watzl, B. Associations of current diet with plasma and urine TMAO in the KarMeN study: Direct and indirect contributions. *Mol. Nutr. Food Res.* **2017**, *61*, 1700363. [[CrossRef](#)]
46. Romano, K.A.; Vivas, E.I.; Amador-Noguez, D.; Rey, F.E. Intestinal microbiota composition modulates choline bioavailability from diet and accumulation of the proatherogenic metabolite trimethylamine-N-oxide. *MBio* **2015**, *6*, e02481-14. [[CrossRef](#)]
47. Falony, G.; Vieira-Silva, S.; Raes, J. Microbiology Meets Big Data: The Case of Gut Microbiota-Derived Trimethylamine. *Annu. Rev. Microbiol.* **2015**, *69*, 305–321. [[CrossRef](#)]
48. Tang, W.H.W.; Wang, Z.; Levison, B.S.; Koeth, R.A.; Britt, E.B.; Fu, X.; Wu, Y.; Hazen, S.L. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N. Engl. J. Med.* **2013**, *368*, 1575–1584. [[CrossRef](#)]
49. Cho, C.E.; Taesuwan, S.; Malysheva, O.V.; Bender, E.; Tulchinsky, N.F.; Yan, J.; Sutter, J.L.; Caudill, M.A. Trimethylamine-N-oxide (TMAO) response to animal source foods varies among healthy young men and is influenced by their gut microbiota composition: A randomized controlled trial. *Mol. Nutr. Food Res.* **2017**, *61*, 324. [[CrossRef](#)]
50. Biagi, E.; Franceschi, C.; Rampelli, S.; Severgnini, M.; Ostan, R.; Turroni, S.; Consolandi, C.; Quercia, S.; Scurti, M.; Monti, D.; et al. Gut Microbiota and Extreme Longevity. *Curr. Biol.* **2016**, *26*, 1480–1485. [[CrossRef](#)]
51. Santoro, A.; Ostan, R.; Candela, M.; Biagi, E.; Brigidi, P.; Capri, M.; Franceschi, C. Gut microbiota changes in the extreme decades of human life: A focus on centenarians. *Cell. Mol. Life Sci.* **2018**, *75*, 129–148. [[CrossRef](#)]
52. Ley, R.E.; Turnbaugh, P.J.; Klein, S.; Gordon, J.I. Microbial ecology: Human gut microbes associated with obesity. *Nature* **2006**, *444*, 1022–1023. [[CrossRef](#)]
53. Ley, R.E. Gut microbiota in 2015: Prevotella in the gut: Choose carefully. *Nat. Rev. Gastroenterol. Hepatol.* **2016**, *13*, 69–70. [[CrossRef](#)]
54. Velasquez, M.T.; Ramezani, A.; Manal, A.; Raj, D.S. Trimethylamine N-oxide: The good, the bad and the unknown. *Toxins* **2016**, *8*, 326. [[CrossRef](#)]
55. Landfald, B.; Valeur, J.; Berstad, A.; Raa, J. Microbial trimethylamine- N -oxide as a disease marker: Something fishy? *Microb. Ecol. Health Dis.* **2017**, *28*, 1327309. [[CrossRef](#)]
56. Cheung, W.; Keski-Rahkonen, P.; Assi, N.; Ferrari, P.; Freisling, H.; Rinaldi, S.; Slimani, N.; Zamora-Ros, R.; Rundle, M.; Frost, G.; et al. A metabolomic study of biomarkers of meat and fish intake. *Am. J. Clin. Nutr.* **2017**, *105*, 600–608. [[CrossRef](#)]
57. Barton, W.; Penney, N.C.; Cronin, O.; Garcia-Perez, I.; Molloy, M.G.; Holmes, E.; Shanahan, F.; Cotter, P.D.; O’Sullivan, O. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut* **2018**, *67*, 625–633. [[CrossRef](#)]

58. Penry, J.T.; Manore, M.M. Choline: An important micronutrient for maximal endurance-exercise performance? *Int. J. Sport Nutr. Exerc. Metab.* **2008**, *18*, 191–203. [[CrossRef](#)]
59. Rogerson, D. Vegan diets: Practical advice for athletes and exercisers. *J. Int. Soc. Sports Nutr.* **2017**, *14*, 1–15. [[CrossRef](#)]
60. Hamaker, B.R.; Tuncil, Y.E. A perspective on the complexity of dietary fiber structures and their potential effect on the gut microbiota. *J. Mol. Biol.* **2014**, *426*, 3838–3850. [[CrossRef](#)]
61. Sharma, P.; Bhandari, C.; Kumar, S.; Sharma, B.; Bhadwal, P.; Agnihotri, N. Dietary fibers: A way to a healthy microbiome. In *Diet, Microbiome and Health*; Elsevier: Amsterdam, The Netherlands, 2018; ISBN 9780128114407.
62. Wołyniec, W.; Kasprowicz, K.; Giebułtowicz, J.; Korytowska, N.; Zorena, K.; Bartoszewicz, M.; Rita-Tkachenko, P.; Renke, M.; Ratkowski, W. Changes in water soluble uremic toxins and urinary acute kidney injury biomarkers after 10-and 100-km runs. *Int. J. Environ. Res. Public Health* **2019**, *16*, 4153. [[CrossRef](#)] [[PubMed](#)]
63. Schraner, D.; Kastenmüller, G.; Schönfelder, M.; Römisch-Margl, W.; Wackerhage, H. Metabolite Concentration Changes in Humans After a Bout of Exercise: A Systematic Review of Exercise Metabolomics Studies. *Sport. Med.-Open* **2020**, *6*, 1–17. [[CrossRef](#)] [[PubMed](#)]
64. Kerkick, C.M.; Wilborn, C.D.; Roberts, M.D.; Smith-Ryan, A.; Kleiner, S.M.; Jäger, R.; Collins, R.; Cooke, M.; Davis, J.N.; Galvan, E.; et al. ISSN exercise & sports nutrition review update: Research & recommendations. *J. Int. Soc. Sports Nutr.* **2018**, *5*, 38.
65. Hughes, R.L. A Review of the Role of the Gut Microbiome in Personalized Sports Nutrition. *Front. Nutr.* **2020**, *6*, 191. [[CrossRef](#)]
66. Ekelund, U.; Ward, H.A.; Norat, T.; Luan, J.; May, A.M.; Weiderpass, E.; Sharp, S.J.; Overvad, K.; Østergaard, J.N.; Tjønneland, A.; et al. Physical activity and all-cause mortality across levels of overall and abdominal adiposity in European men and women: The European prospective investigation into cancer and nutrition study (EPIC). *Am. J. Clin. Nutr.* **2015**, *101*, 613–621. [[CrossRef](#)]
67. Kyu, H.H.; Bachman, V.F.; Alexander, L.T.; Mumford, J.E.; Afshin, A.; Estep, K.; Veerman, J.L.; Delwiche, K.; Iannarone, M.L.; Moyer, M.L.; et al. Physical activity and risk of breast cancer, colon cancer, diabetes, ischemic heart disease, and ischemic stroke events: Systematic review and dose-response meta-analysis for the Global Burden of Disease Study 2013. *BMJ* **2016**, *354*, i3857. [[CrossRef](#)]
68. Wolin, K.Y.; Yan, Y.; Colditz, G.A.; Lee, I.M. Physical activity and colon cancer prevention: A meta-analysis. *Br. J. Cancer* **2009**, *100*, 611–616. [[CrossRef](#)]
69. World Cancer Research Fund/American Institute for Cancer Research. *Diet, Nutrition, Physical Activity and Cancer: A Global Perspective*; World Cancer Research Fund/American Institute for Cancer Research: Washington, DC, USA, 2018; ISBN 9781912259465.
70. Fuchs, R. Physical Activity and Health. In *International Encyclopedia of the Social & Behavioral Sciences*, 2nd ed.; Elsevier: Amsterdam, The Netherlands, 2015; ISBN 9780080970875.
71. Saint-Maurice, P.F.; Coughlan, D.; Kelly, S.P.; Keadle, S.K.; Cook, M.B.; Carlson, S.A.; Fulton, J.E.; Matthews, C.E. Association of Leisure-Time Physical Activity Across the Adult Life Course with All-Cause and Cause-Specific Mortality. *JAMA Netw. Open* **2019**, *2*, e190355. [[CrossRef](#)]
72. Zhang, Y.; Wang, Y.; Ke, B.; Du, J. TMAO: How gut microbiota contributes to heart failure. *Transl. Res.* **2021**, *228*, 109–125. [[CrossRef](#)]
73. Mueller, D.M.; Allenspach, M.; Othman, A.; Saely, C.H.; Muendlein, A.; Vonbank, A.; Drexel, H.; von Eckardstein, A. Plasma levels of trimethylamine-N-oxide are confounded by impaired kidney function and poor metabolic control. *Atherosclerosis* **2015**, *243*, 638–644. [[CrossRef](#)]
74. Hansen, T.H.; Gøbel, R.J.; Hansen, T.; Pedersen, O. The gut microbiome in cardio-metabolic health. *Genome Med.* **2015**, *7*, 33. [[CrossRef](#)]
75. Wu, S.; Rhee, K.J.; Albesiano, E.; Rabizadeh, S.; Wu, X.; Yen, H.R.; Huso, D.L.; Brancati, F.L.; Wick, E.; McAllister, F.; et al. A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. *Nat. Med.* **2009**, *15*, 1016–1022. [[CrossRef](#)] [[PubMed](#)]
76. Bae, S.; Ulrich, C.M.; Neuhouser, M.L.; Malysheva, O.; Bailey, L.B.; Xiao, L.; Brown, E.C.; Cushing-Haugen, K.L.; Zheng, Y.; Cheng, T.Y.D.; et al. Plasma choline metabolites and colorectal cancer risk in the women’s health initiative observational study. *Cancer Res.* **2014**, *74*, 7442–7452. [[CrossRef](#)]
77. Xu, R.; Wang, Q.Q.; Li, L. A genome-wide systems analysis reveals strong link between colorectal cancer and trimethylamine N-oxide (TMAO), a gut microbial metabolite of dietary meat and fat. *BMC Genom.* **2015**, *16*, S4. [[CrossRef](#)]
78. Chan, C.W.H.; Law, B.M.H.; Waye, M.M.Y.; Chan, J.Y.W.; Wei So, W.K.; Chow, K.M. Trimethylamine-N-oxide as one hypothetical link for the relationship between intestinal microbiota and cancer-Where we are and where shall we go? *J. Cancer* **2019**, *10*, 5874. [[CrossRef](#)]
79. Li, X.S.; Obeid, S.; Klingenberg, R.; Gencer, B.; Mach, F.; Räber, L.; Windecker, S.; Rodondi, N.; Nanchen, D.; Muller, O.; et al. Gutmicrobiota-dependent trimethylamine N-oxide in acute coronary syndromes: A prognostic marker for incident cardiovascular events beyond traditional risk factors. *Eur. Heart J.* **2017**, *38*, 814–824. [[CrossRef](#)]
80. Senthong, V.; Wang, Z.; Li, X.S.; Fan, Y.; Wu, Y.; Tang, W.H.W.; Hazen, S.L. Intestinal microbiota-generated metabolite Trimethylamine-N-oxide and 5-year mortality risk in stable coronary artery disease: The contributory role of intestinal microbiota in a COURAGE-like patient cohort. *J. Am. Heart Assoc.* **2016**, *5*, e002816. [[CrossRef](#)]
81. Wang, Z.; Klipfell, E.; Bennett, B.J.; Koeth, R.; Levison, B.S.; Dugar, B.; Feldstein, A.E.; Britt, E.B.; Fu, X.; Chung, Y.M.; et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* **2011**, *472*, 57–63. [[CrossRef](#)]

82. Chistiakov, D.A.; Melnichenko, A.A.; Myasoedova, V.A.; Grechko, A.V.; Orekhov, A.N. Mechanisms of foam cell formation in atherosclerosis. *J. Mol. Med.* **2017**, *95*, 1153–1165. [[CrossRef](#)]
83. Thon, M.P.; Hemmler, A.; Glinzer, A.; Mayr, M.; Wildgruber, M.; Zerneck-Madsen, A.; Gee, M.W. A multiphysics approach for modeling early atherosclerosis. *Biomech. Model. Mechanobiol.* **2018**, *17*, 617–644. [[CrossRef](#)]
84. Geng, J.; Yang, C.; Wang, B.; Zhang, X.; Hu, T.; Gu, Y.; Li, J. Trimethylamine N-oxide promotes atherosclerosis via CD36-dependent MAPK/JNK pathway. *Biomed. Pharmacother.* **2018**, *97*, 941–947. [[CrossRef](#)] [[PubMed](#)]
85. Mohammadi, A.; Najar, A.G.; Yaghoobi, M.M.; Jahani, Y.; Vahabzadeh, Z. Trimethylamine-N-Oxide Treatment Induces Changes in the ATP-Binding Cassette Transporter A1 and Scavenger Receptor A1 in Murine Macrophage J774A.1 cells. *Inflammation* **2016**, *39*, 393–404. [[CrossRef](#)] [[PubMed](#)]
86. Zhu, Y.; Li, Q.; Jiang, H. Gut microbiota in atherosclerosis: Focus on trimethylamine N-oxide. *Apmis* **2020**, *128*, 353–366. [[CrossRef](#)]
87. Sun, X.; Jiao, X.; Ma, Y.; Liu, Y.; Zhang, L.; He, Y.; Chen, Y. Trimethylamine N-oxide induces inflammation and endothelial dysfunction in human umbilical vein endothelial cells via activating ROS-TXNIP-NLRP3 inflammasome. *Biochem. Biophys. Res. Commun.* **2016**, *481*, 63–70. [[CrossRef](#)]
88. Ren, D.; Liu, Y.; Zhao, Y.; Yang, X. Hepatotoxicity and endothelial dysfunction induced by high choline diet and the protective effects of phloretin in mice. *Food Chem. Toxicol.* **2016**, *94*, 203–212. [[CrossRef](#)]
89. Chou, R.H.; Chen, C.Y.; Chen, I.C.; Huang, H.L.; Lu, Y.W.; Kuo, C.S.; Chang, C.C.; Huang, P.H.; Chen, J.W.; Lin, S.J. Trimethylamine N-Oxide, Circulating Endothelial Progenitor Cells, and Endothelial Function in Patients with Stable Angina. *Sci. Rep.* **2019**, *9*, 4249. [[CrossRef](#)]
90. Ingram, D.A.; Mead, L.E.; Tanaka, H.; Meade, V.; Fenoglio, A.; Mortell, K.; Pollok, K.; Ferkowicz, M.J.; Gilley, D.; Yoder, M.C. Identification of a novel hierarchy of endothelial progenitor cells using human peripheral and umbilical cord blood. *Blood* **2004**, *104*, 2752–2760. [[CrossRef](#)]
91. Basile, D.P.; Yoder, M.C. Circulating and Tissue Resident Endothelial Progenitor Cells. *J. Cell. Physiol.* **2014**, *229*, 10–16. [[CrossRef](#)]
92. Chopra, H.; Hung, M.K.; Kwong, D.L.; Zhang, C.F.; Pow, E.H.N. Insights into endothelial progenitor cells: Origin, classification, potentials, and prospects. *Stem Cells Int.* **2018**, *2018*, 9847015. [[CrossRef](#)]
93. Prater, D.N.; Case, J.; Ingram, D.A.; Yoder, M.C. Working hypothesis to redefine endothelial progenitor cells. *Leukemia* **2007**, *21*, 1141–1149. [[CrossRef](#)]
94. Hur, J.; Yoon, C.H.; Kim, H.S.; Choi, J.H.; Kang, H.J.; Hwang, K.K.; Oh, B.H.; Lee, M.M.; Park, Y.B. Characterization of Two Types of Endothelial Progenitor Cells and Their Different Contributions to Neovascularization. *Arterioscler. Thromb. Vasc. Biol.* **2004**, *24*, 288–293. [[CrossRef](#)] [[PubMed](#)]
95. Davignon, J.; Ganz, P. Role of endothelial dysfunction in atherosclerosis. *Circulation* **2004**, *109*, III27–III32. [[CrossRef](#)]
96. Seldin, M.M.; Meng, Y.; Qi, H.; Zhu, W.F.; Wang, Z.; Hazen, S.L.; Lusis, A.J.; Shih, D.M. Trimethylamine N-oxide promotes vascular inflammation through signaling of mitogen-activated protein kinase and nuclear factor- κ B. *J. Am. Heart Assoc.* **2016**, *5*, e002767. [[CrossRef](#)]
97. Chen, H.; Li, J.; Li, N.; Liu, H.; Tang, J. Increased circulating trimethylamine N-oxide plays a contributory role in the development of endothelial dysfunction and hypertension in the RUPP rat model of preeclampsia. *Hypertens. Pregnancy* **2019**, *38*, 96–104. [[CrossRef](#)]
98. Mostafa Mtairag, E.; Chollet-Martin, S.; Oudghiri, M.; Laquay, N.; Jacob, M.P.; Michel, J.B.; Feldman, L.J. Effects of interleukin-10 on monocyte/endothelial cell adhesion and MMP-9/TIMP-1 secretion. *Cardiovasc. Res.* **2001**, *49*, 882–890. [[CrossRef](#)]
99. Li, T.; Chen, Y.; Gua, C.; Li, X. Elevated circulating trimethylamine N-oxide levels contribute to endothelial dysfunction in aged rats through vascular inflammation and oxidative stress. *Front. Physiol.* **2017**, *8*, 350. [[CrossRef](#)]
100. Herrera, M.D.; Mingorance, C.; Rodríguez-Rodríguez, R.; Alvarez de Sotomayor, M. Endothelial dysfunction and aging: An update. *Ageing Res. Rev.* **2010**, *9*, 142–152. [[CrossRef](#)]
101. Donato, A.J.; Morgan, R.G.; Walker, A.E.; Lesniewski, L.A. Cellular and molecular biology of aging endothelial cells. *J. Mol. Cell. Cardiol.* **2015**, *89*, 122–135. [[CrossRef](#)]
102. Seals, D.R.; Jablonski, K.L.; Donato, A.J. Aging and vascular endothelial function in humans. *Clin. Sci.* **2011**, *120*, 357–375. [[CrossRef](#)]
103. Ma, G.H.; Pan, B.; Chen, Y.; Guo, C.X.; Zhao, M.M.; Zheng, L.M.; Chen, B.X. Trimethylamine N-oxide in atherogenesis: Impairing endothelial self-repair capacity and enhancing monocyte adhesion. *Biosci. Rep.* **2017**, *37*, BSR20160244. [[CrossRef](#)]
104. Hai, X.; Landeras, V.; Dobro, M.A.; DeOreo, P.; Meyer, T.W.; Hostetter, T.H. Mechanism of prominent trimethylamine oxide (TMAO) accumulation in hemodialysis patients. *PLoS ONE* **2015**, *10*, e0143731. [[CrossRef](#)] [[PubMed](#)]
105. Missailidis, C.; Hällqvist, J.; Qureshi, A.R.; Barany, P.; Heimbürger, O.; Lindholm, B.; Stenvinkel, P.; Bergman, P. Serum trimethylamine-N-Oxide is strongly related to renal function and predicts outcome in chronic kidney disease. *PLoS ONE* **2016**, *11*, e0141738. [[CrossRef](#)] [[PubMed](#)]
106. Kim, R.B.; Morse, B.L.; Djurdjev, O.; Tang, M.; Muirhead, N.; Barrett, B.; Holmes, D.T.; Madore, F.; Clase, C.M.; Rigatto, C.; et al. Advanced chronic kidney disease populations have elevated trimethylamine N-oxide levels associated with increased cardiovascular events. *Kidney Int.* **2016**, *89*, 1144–1152. [[CrossRef](#)]
107. Robbiano, L.; Mereto, E.; Corbu, C.; Brambilla, G. DNA damage induced by seven N-nitroso compounds in primary cultures of human and rat kidney cells. *Mutat. Res.* **1996**, *368*, 41–47. [[CrossRef](#)] [[PubMed](#)]

108. Teft, W.A.; Morse, B.L.; Leake, B.F.; Wilson, A.; Mansell, S.E.; Hegele, R.A.; Ho, R.H.; Kim, R.B. Identification and Characterization of Trimethylamine-N-oxide Uptake and Efflux Transporters. *Mol. Pharm.* **2017**, *14*, 310–318. [[CrossRef](#)]
109. Zixin, Y.; Lulu, C.; Xiangchang, Z.; Qing, F.; Binjie, Z.; Chunyang, L.; Tai, R.; Dongsheng, O. TMAO as a potential biomarker and therapeutic target for chronic kidney disease: A review. *Front. Pharmacol.* **2022**, *12*, 929262. [[CrossRef](#)]
110. Kalagi, N.A.; Thota, R.N.; Stojanovski, E.; Alburikan, K.A.; Garg, M.L. Association between Plasma Trimethylamine N-Oxide Levels and Type 2 Diabetes: A Case Control Study. *Nutrients* **2022**, *14*, 2093. [[CrossRef](#)]
111. Zhuang, R.; Ge, X.; Han, L.; Yu, P.; Gong, X.; Meng, Q.; Zhang, Y.; Fan, H.; Zheng, L.; Liu, Z.; et al. Gut microbe-generated metabolite trimethylamine N-oxide and the risk of diabetes: A systematic review and dose-response meta-analysis. *Obes. Rev.* **2019**, *20*, 883–894. [[CrossRef](#)]
112. Tang, W.H.W.; Wang, Z.; Li, X.S.; Fan, Y.; Li, D.S.; Wu, Y.; Hazen, S.L. Increased Trimethylamine N-Oxide Portends High Mortality Risk Independent of Glycemic Control in Patients with Type 2 Diabetes Mellitus. *Clin. Chem.* **2017**, *63*, 297–306. [[CrossRef](#)]
113. Dambrova, M.; Latkovskis, G.; Kuka, J.; Strele, I.; Konrade, I.; Grinberga, S.; Hartmane, D.; Pugovics, O.; Erglis, A.; Liepinsh, E. Diabetes is Associated with Higher Trimethylamine N-oxide Plasma Levels. *Exp. Clin. Endocrinol. Diabetes* **2016**, *124*, 251–256. [[CrossRef](#)]
114. Gao, X.; Liu, X.; Xu, J.; Xue, C.; Xue, Y.; Wang, Y. Dietary trimethylamine N-oxide exacerbates impaired glucose tolerance in mice fed a high fat diet. *J. Biosci. Bioeng.* **2014**, *118*, 476–481. [[CrossRef](#)]
115. Jia, J.; Dou, P.; Gao, M.; Kong, X.; Li, C.; Liu, Z.; Huang, T. Assessment of Causal Direction Between Gut Microbiota-Dependent Metabolites and Cardiometabolic Health: A Bidirectional Mendelian Randomization Analysis. *Diabetes* **2019**, *68*, 1747–1755. [[CrossRef](#)]
116. Rea, D.; Coppola, G.; Palma, G.; Barbieri, A.; Luciano, A.; Del Prete, P.; Rossetti, S.; Berretta, M.; Facchini, G.; Perdonà, S.; et al. Microbiota effects on cancer: From risks to therapies. *Oncotarget* **2018**, *9*, 17915–17927. [[CrossRef](#)]
117. Sender, R.; Fuchs, S.; Milo, R. Revised Estimates for the Number of Human and Bacteria Cells in the Body. *PLoS Biol.* **2016**, *14*, e1002533. [[CrossRef](#)]
118. Cani, P.D. Metabolism in 2013: The gut microbiota manages host metabolism. *Nat. Rev. Endocrinol.* **2014**, *10*, 74–76. [[CrossRef](#)]
119. Ianiro, G.; Bruno, G.; Lopetuso, L.; Beghella, F.; Laterza, L.; D’Aversa, F.; Gigante, G.; Cammarota, G.; Gasbarrini, A. Role of Yeasts in Healthy and Impaired Gut Microbiota: The Gut Mycome. *Curr. Pharm. Des.* **2014**, *20*, 4565–4569. [[CrossRef](#)]
120. Lozupone, C.A.; Stombaugh, J.I.; Gordon, J.I.; Jansson, J.K.; Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* **2012**, *489*, 220–230. [[CrossRef](#)]
121. Columpsi, P.; Sacchi, P.; Zuccaro, V.; Cima, S.; Sarda, C.; Mariani, M.; Gori, A.; Bruno, R. Beyond the gut bacterial microbiota: The gut virome. *J. Med. Virol.* **2016**, *88*, 1467–1472. [[CrossRef](#)]
122. Mills, S.; Stanton, C.; Lane, J.A.; Smith, G.J.; Ross, R.P. Precision nutrition and the microbiome, part I: Current state of the science. *Nutrients* **2019**, *11*, 923. [[CrossRef](#)]
123. Cenit, M.C.; Sanz, Y.; Codoñer-Franch, P. Influence of gut microbiota on neuropsychiatric disorders. *World J. Gastroenterol.* **2017**, *23*, 5486–5498. [[CrossRef](#)]
124. Slyepchenko, A.; Maes, M.; Jacka, F.N.; Köhler, C.A.; Barichello, T.; McIntyre, R.S.; Berk, M.; Grande, I.; Foster, J.A.; Vieta, E.; et al. Gut Microbiota, Bacterial Translocation, and Interactions with Diet: Pathophysiological Links between Major Depressive Disorder and Non-Communicable Medical Comorbidities. *Psychother. Psychosom.* **2016**, *86*, 31–46. [[CrossRef](#)]
125. Hold, G.L.; Smith, M.; Grange, C.; Watt, E.R.; El-Omar, E.M.; Mukhopadhyay, I. Role of the gut microbiota in inflammatory bowel disease pathogenesis: What have we learnt in the past 10 years? *World J. Gastroenterol.* **2014**, *20*, 1192–1210. [[CrossRef](#)] [[PubMed](#)]
126. Llorente, C.; Schnabl, B. The Gut Microbiota and Liver Disease. *Cell. Mol. Gastroenterol. Hepatol.* **2015**, *1*, 275–284. [[CrossRef](#)]
127. Vindigni, S.M.; Zisman, T.L.; Suskind, D.L.; Damman, C.J. The intestinal microbiome, barrier function, and immune system in inflammatory bowel disease: A tripartite pathophysiological circuit with implications for new therapeutic directions. *Therap. Adv. Gastroenterol.* **2016**, *9*, 606–625. [[CrossRef](#)] [[PubMed](#)]
128. Fukui, H. Increased Intestinal Permeability and Decreased Barrier Function: Does It Really Influence the Risk of Inflammation? *Inflamm. Intest. Dis.* **2016**, *1*, 135–145. [[CrossRef](#)]
129. Meng, C.; Bai, C.; Brown, T.D.; Hood, L.E.; Tian, Q. Human Gut Microbiota and Gastrointestinal Cancer. *Genom. Proteom. Bioinform.* **2018**, *16*, 33–49. [[CrossRef](#)]
130. Mima, K.; Ogino, S.; Nakagawa, S.; Sawayama, H.; Kinoshita, K.; Krashima, R.; Ishimoto, T.; Imai, K.; Iwatsuki, M.; Hashimoto, D.; et al. The role of intestinal bacteria in the development and progression of gastrointestinal tract neoplasms. *Surg. Oncol.* **2017**, *26*, 368–376. [[CrossRef](#)] [[PubMed](#)]
131. Bhandari, A.; Woodhouse, M.; Gupta, S. Colorectal cancer is a leading cause of cancer incidence and mortality among adults younger than 50 years in the USA: A SEER-based analysis with comparison to other young-onset cancers. *J. Investig. Med.* **2017**, *65*, 311–315. [[CrossRef](#)]
132. Zackular, J.P.; Baxter, N.T.; Iverson, K.D.; Sadler, W.D.; Petrosino, J.F.; Chen, G.Y.; Schloss, P.D. The gut microbiome modulates colon tumorigenesis. *MBio* **2013**, *4*, e00692-13. [[CrossRef](#)]
133. Zackular, J.P.; Baxter, N.T.; Chen, G.Y.; Schloss, P.D. Manipulation of the Gut Microbiota Reveals Role in Colon Tumorigenesis. *mSphere* **2016**, *1*, e00001-15. [[CrossRef](#)]
134. Bullman, S.; Peadarallu, C.S.; Sicinska, E.; Clancy, T.E.; Zhang, X.; Cai, D.; Neuberg, D.; Huang, K.; Guevara, F.; Nelson, T.; et al. Analysis of Fusobacterium persistence and antibiotic response in colorectal cancer. *Science* **2017**, *358*, 1443–1448. [[CrossRef](#)]

135. Drewes, J.L.; Housseau, F.; Sears, C.L. Sporadic colorectal cancer: Microbial contributors to disease prevention, development and therapy. *Br. J. Cancer* **2016**, *115*, 273–280. [[CrossRef](#)]
136. Gagnière, J.; Raisch, J.; Veziant, J.; Barnich, N.; Bonnet, R.; Buc, E.; Bringer, M.A.; Pezet, D.; Bonnet, M. Gut microbiota imbalance and colorectal cancer. *World J. Gastroenterol.* **2016**, *22*, 501–518. [[CrossRef](#)]
137. Sun, J.; Kato, I. Gut microbiota, inflammation and colorectal cancer. *Genes Dis.* **2016**, *3*, 130–143. [[CrossRef](#)]
138. Youssef, O.; Lahti, L.; Kokkola, A.; Karla, T.; Tikkanen, M.; Ehsan, H.; Carpelan-Holmström, M.; Koskensalo, S.; Böhling, T.; Rautelin, H.; et al. Stool Microbiota Composition Differs in Patients with Stomach, Colon, and Rectal Neoplasms. *Dig. Dis. Sci.* **2018**, *63*, 2950–2958. [[CrossRef](#)]
139. Sears, C.L.; Garrett, W.S. Microbes, microbiota, and colon cancer. *Cell Host Microbe* **2014**, *15*, 317–328. [[CrossRef](#)]
140. Chen, J.; Domingue, J.C.; Sears, C.L. Microbiota dysbiosis in select human cancers: Evidence of association and causality. *Semin. Immunol.* **2017**, *32*, 25–34. [[CrossRef](#)]
141. Arthur, J.C.; Perez-Chanona, E.; Mühlbauer, M.; Tomkovich, S.; Uronis, J.M.; Fan, T.J.; Campbell, B.J.; Abujamel, T.; Dogan, B.; Rogers, A.B.; et al. Intestinal inflammation targets cancer-inducing activity of the microbiota. *Science* **2012**, *338*, 120–123. [[CrossRef](#)]
142. Huycke, M.M.; Abrams, V.; Moore, D.R. Enterococcus faecalis produces extracellular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. *Carcinogenesis* **2002**, *23*, 529–536. [[CrossRef](#)]
143. Sze, M.A.; Baxter, N.T.; Ruffin, M.T.; Rogers, M.A.M.; Schloss, P.D. Normalization of the microbiota in patients after treatment for colonic lesions. *Microbiome* **2017**, *5*, 150. [[CrossRef](#)]
144. Deng, X.; Li, Z.; Li, G.; Li, B.; Jin, X.; Lyu, G. Comparison of microbiota in patients treated by surgery or chemotherapy by 16S rRNA sequencing reveals potential biomarkers for colorectal cancer therapy. *Front. Microbiol.* **2018**, *9*, 1607. [[CrossRef](#)] [[PubMed](#)]
145. Sánchez-Alcoholado, L.; Ordóñez, R.; Otero, A.; Plaza-Andrade, I.; Laborda-Illanes, A.; Medina, J.A.; Ramos-Molina, B.; Gómez-Millán, J.; Queipo-Ortuño, M.I. Gut Microbiota-Mediated Inflammation and Gut Permeability in Patients with Obesity and Colorectal Cancer. *Int. J. Mol. Sci.* **2020**, *21*, 6782. [[CrossRef](#)]
146. Liu, X.; Liu, H.; Yuan, C.; Zhang, Y.; Wang, W.; Hu, S.; Liu, L.; Wang, Y. Preoperative serum TMAO level is a new prognostic marker for colorectal cancer. *Biomarkers Med.* **2017**, *11*, 443–447. [[CrossRef](#)] [[PubMed](#)]
147. Chen, M.L.; Zhu, X.H.; Ran, L.; Lang, H.-D.; Yi, L.; Mi, M.-T. Trimethylamine-N-Oxide Induces Vascular Inflammation by Activating the NLRP3 Inflammasome Through the SIRT3-SOD2-mtROS Signaling Pathway. *J. Am. Heart Assoc.* **2017**, *6*, e006347. [[CrossRef](#)] [[PubMed](#)]
148. Yue, C.; Yang, X.; Li, J.; Chen, X.; Zhao, X.; Chen, Y.; Wen, Y. Trimethylamine N-oxide prime NLRP3 inflammasome via inhibiting ATG16L1-induced autophagy in colonicepithelial cells. *Biochem. Biophys. Res. Commun.* **2017**, *490*, 541–551. [[CrossRef](#)]
149. Huang, C.-F.; Chen, L.; Li, Y.-C.; Wu, L.; Yu, G.-T.; Zhang, W.-F.; Sun, Z.-J. NLRP3 inflammasome activation promotes inflammation-induced carcinogenesis in head and neck squamous cell carcinoma. *J. Exp. Clin. Cancer Res.* **2017**, *36*, 116. [[CrossRef](#)]
150. Wang, H.; Luo, Q.; Feng, X.; Zhang, R.; Li, J.; Chen, F. NLRP3 promotes tumor growth and metastasis in human oral squamous cell carcinoma. *BMC Cancer* **2018**, *18*, 500. [[CrossRef](#)]
151. He, Q.; Fu, Y.; Tian, D.; Yan, W. The contrasting roles of inflammasomes in cancer. *Am. J. Cancer Res.* **2018**, *8*, 566–583.
152. Chen, J.; Pitmon, E.; Wnag, K. Micorbiome, inflammation and colorec5toal cancer. *Semin. Immunol.* **2017**, *32*, 43–53. [[CrossRef](#)]
153. Wu, D.; Cao, M.; Peng, J.; Li, N.; Yi, S.; Song, L.; Wang, X.; Zhang, M.; Zhao, J. The effect of trimethylamine N-oxide on Helicobacter pylori-induced changes of immunoinflammatory genes expression in gastric epithelial cells. *Int. Immunopharmacol.* **2017**, *43*, 172–178. [[CrossRef](#)]
154. Kumari, N.; Dwarakanath, B.S.; Das, A.; Bhatt, A.N. Role of interleukin-6 in cancer progression and therapeutic resistance. *Tumor Biol.* **2016**, *37*, 11553–11572. [[CrossRef](#)]
155. Tilg, H.; Adolph, T.E.; Gerner, R.R.; Moschen, A.R. The Intestinal Microbiota in Colorectal Cancer. *Cancer Cell* **2018**, *33*, 954–964. [[CrossRef](#)]
156. McEntyre, C.J.; Lever, M.; Chambers, S.T.; George, P.M.; Slow, S.; Elmslie, J.L.; Florkowski, C.M.; Lunt, H.; Krebs, J.D. Variation of betaine, N,N-dimethylglycine, choline, glycerophosphorylcholine, taurine and trimethylamine-N-oxide in the plasma and urine of overweight people with type 2 diabetes over a two-year period. *Ann. Clin. Biochem.* **2015**, *52 Pt 3*, 352–360. [[CrossRef](#)]
157. Reuter, S.; Gupta, S.C.; Chaturvedi, M.M.; Aggarwal, B.B. Oxidative stress, inflammation, and cancer: How are they linked? *Free Radic. Biol. Med.* **2010**, *49*, 1603–1616. [[CrossRef](#)]
158. Federico, A.; Morgillo, F.; Tuccillo, C.; Ciardiello, F.; Loguercio, C. Chronic inflammation and oxidative stress in human carcinogenesis. *Int. J. Cancer* **2007**, *121*, 2381–2386. [[CrossRef](#)]
159. Liu, Y.; Dai, M. Trimethylamine N-Oxide Generated by the Gut Microbiota Is Associated with Vascular Inflammation: New Insights into Atherosclerosis. *Mediat. Inflamm.* **2020**, *2020*, 4634172. [[CrossRef](#)]
160. Gao, X.; Xu, J.; Jiang, C.; Zhang, Y.; Xue, Y.; Li, Z.; Wang, J.; Xue, C.; Wang, Y. Fish oil ameliorates trimethylamine N-oxide-exacerbated glucose intolerance in high-fat diet-fed mice. *Food Funct.* **2015**, *6*, 1117–1125. [[CrossRef](#)]
161. Hernandez-Alonso, P.; Canueto, D.; Giardina, S.; Salas-Salvado, J.; Canellas, N.; Correig, X.; Bullo, M. Pistachio intake modulates urinary gut microbiota-related metabolites in pre-diabetic subjects. *Obes. Facts* **2017**, *10*, 198. [[CrossRef](#)]
162. Fiber Intervention on Gut Microbiota in Children with Prader-Willi Syndrome. *Case Med. Res.* **2019**, *4*, 991. [[CrossRef](#)]

163. Velazquez, R.; Ferreira, E.; Knowles, S.; Fux, C.; Rodin, A.; Winslow, W.; Oddo, S. Lifelong choline supplementation ameliorates Alzheimer's disease pathology and associated cognitive deficits by attenuating microglia activation. *Aging Cell* **2019**, *18*, e13037. [CrossRef]
164. Leermakers, E.T.M.; Moreira, E.M.; Kieffe-de Jong, J.C.; Darweesh, S.K.L.; Visser, T.; Voortman, T.; Bautista, P.K.; Chowdhury, R.; Gorman, D.; Bramer, W.M.; et al. Effects of choline on health across the life course: A systematic review. *Nutr. Rev.* **2015**, *73*, 500–522. [CrossRef] [PubMed]
165. Sarter, M.; Parikh, V. Choline transporters, cholinergic transmission and cognition. *Nat. Rev. Neurosci.* **2005**, *6*, 48–56. [CrossRef] [PubMed]
166. Zeisel, S.H. The fetal origins of memory: The role of dietary choline in optimal brain development. *J. Pediatr.* **2006**, *149*, S131–S136. [CrossRef]
167. Wessler, I.; Kirkpatrick, C.J. Acetylcholine beyond neurons: The non-neuronal cholinergic system in humans. *Br. J. Pharmacol.* **2008**, *154*, 1558–1571. [CrossRef]
168. Vance, D.E.; Ridgway, N.D. The methylation of phosphatidylethanolamine. *Prog. Lipid Res.* **1988**, *27*, 61–79. [CrossRef]
169. Jiang, X.; West, A.A.; Caudill, M.A. Maternal choline supplementation: A nutritional approach for improving offspring health? *Trends Endocrinol. Metab.* **2014**, *25*, 263–273. [CrossRef]
170. Shronts, E.P. Essential nature of choline with implications for total parenteral nutrition. *J. Am. Diet. Assoc.* **1997**, *97*, 639–649. [CrossRef]
171. European Food Safety Authority (EFSA). *Panel on Dietetic Products Nutrition and Allergies (NDA) Overview on Dietary Reference Values for the EU Population as Derived by the EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA)*; EFSA: Parma, Italy, 2017.
172. Zaina, S.; Heyn, H.; Carmona, F.J.; Varol, N.; Sayols, S.; Condom, E.; Ramírez-Ruz, J.; Gomez, A.; Gonçalves, I.; Moran, S.; et al. DNA methylation map of human atherosclerosis. *Circ. Cardiovasc. Genet.* **2014**, *7*, 692–700. [CrossRef]
173. Zaina, S.; Lindholm, M.W.; Lund, G. Nutrition and aberrant DNA methylation patterns in atherosclerosis: More than just hyperhomocysteinemia? *J. Nutr.* **2005**, *135*, 5–8. [CrossRef]
174. Da Costa, K.A.; Gaffney, C.E.; Fischer, L.M.; Zeisel, S.H. Choline deficiency in mice and humans is associated with increased plasma homocysteine concentration after a methionine load. *Am. J. Clin. Nutr.* **2005**, *81*, 440–444. [CrossRef]
175. Clarke, R.; Collins, R.; Lewington, S.; Donald, A.; Alftan, G.; Tuomilehto, J.; Arnesen, E.; Bona, K.; Blacher, J.; Boers, G.H.J.; et al. Homocysteine and risk of ischemic heart disease and stroke: A meta-analysis. *J. Am. Med. Assoc.* **2002**, *288*, 2015–2022.
176. Wu, L.L.; Wu, J.T. Hyperhomocysteinemia is a risk factor for cancer and a new potential tumor marker. *Clin. Chim. Acta* **2002**, *322*, 21–28. [CrossRef] [PubMed]
177. Seshadri, S.; Beiser, A.; Selhub, J.; Jacques, P.F.; Rosenberg, I.H.; D'Agostino, R.B.; Wilson, P.W.F.; Wolf, P.A. Plasma Homocysteine as a Risk Factor for Dementia and Alzheimer's Disease. *N. Engl. J. Med.* **2002**, *346*, 476–483. [CrossRef] [PubMed]
178. Van Meurs, J.B.J.; Dhonukshe-Rutten, R.A.M.; Pluijm, S.M.F.; van der Klift, M.; de Jonge, R.; Lindemans, J.; de Groot, L.C.P.G.M.; Hofman, A.; Witteman, J.C.M.; van Leeuwen, J.P.T.M.; et al. Homocysteine Levels and the Risk of Osteoporotic Fracture. *N. Engl. J. Med.* **2004**, *350*, 2033–2041. [CrossRef] [PubMed]
179. Zeisel, S.H. Metabolic crosstalk between choline/1-carbon metabolism and energy homeostasis. *Clin. Chem. Lab. Med.* **2013**, *51*, 467–475. [CrossRef]
180. Gao, X.; Wang, Y.; Randell, E.; Pedram, P.; Yi, Y.; Gulliver, W.; Sun, G. Higher Dietary Choline and Betaine Intakes are Associated with Better Body Composition in the Adult Population of Newfoundland, Canada. *PLoS ONE* **2016**, *11*, e0155403. [CrossRef]
181. Craig, S.A.S. Betaine in human nutrition. *Am. J. Clin. Nutr.* **2004**, *80*, 539–549. [CrossRef]
182. Tsuge, H.; Nakano, Y.; Onishi, H.; Futamura, Y.; Ohashi, K. A novel purification and some properties of rat liver mitochondrial choline dehydrogenase. *Biochim. Biophys. Acta* **1980**, *614*, 274–284. [CrossRef]
183. Mödinger, Y.; Schön, C.; Wilhelm, M.; Hals, P.A. Plasma kinetics of choline and choline metabolites after a single dose of SuperbaBoost™ krill oil or choline bitartrate in healthy volunteers. *Nutrients* **2019**, *11*, 2548. [CrossRef]
184. Cho, C.E.; Aardema, N.D.J.; Bunnell, M.L.; Larson, D.P.; Aguilar, S.S.; Bergeson, J.R.; Malysheva, O.V.; Caudill, M.A.; Lefevre, M. Effect of choline forms and gut microbiota composition on trimethylamine-n-oxide response in healthy men. *Nutrients* **2020**, *12*, 2220. [CrossRef]
185. Lemos, B.S.; Medina-Vera, I.; Malysheva, O.V.; Caudill, M.A.; Fernandez, M.L. Effects of Egg Consumption and Choline Supplementation on Plasma Choline and Trimethylamine-N-Oxide in a Young Population. *J. Am. Coll. Nutr.* **2018**, *37*, 716–723. [CrossRef] [PubMed]
186. Missimer, A.; Fernandez, M.L.; DiMarco, D.M.; Norris, G.H.; Blesso, C.N.; Murillo, A.G.; Vergara-Jimenez, M.; Lemos, B.S.; Medina-Vera, I.; Malysheva, O.V.; et al. Compared to an Oatmeal Breakfast, Two Eggs/Day Increased Plasma Carotenoids and Choline without Increasing Trimethyl Amine N-Oxide Concentrations. *J. Am. Coll. Nutr.* **2018**, *37*, 140–148. [CrossRef] [PubMed]
187. Bremer, J. Carnitine. Metabolism and functions. *Physiol. Rev.* **1983**, *63*, 1420–1480. [CrossRef] [PubMed]
188. Rebouche, C.J. Kinetics, pharmacokinetics, and regulation of L-Carnitine and acetyl-L-carnitine metabolism. *Ann. N. Y. Acad. Sci.* **2004**, *1033*, 30–41. [CrossRef]
189. Koeth, R.A.; Lam-Galvez, B.R.; Kirsop, J.; Wang, Z.; Levison, B.S.; Gu, X.; Copeland, M.F.; Bartlett, D.; Cody, D.B.; Dai, H.J.; et al. L-Carnitine in omnivorous diets induces an atherogenic gut microbial pathway in humans. *J. Clin. Investig.* **2019**, *129*, 373–387. [CrossRef]

190. Wang, Z.; Bergeron, N.; Levison, B.S.; Li, X.S.; Chiu, S.; Xun, J.; Koeth, R.A.; Lin, L.; Wu, Y.; Tang, W.H.W.; et al. Impact of chronic dietary red meat, white meat, or non-meat protein on trimethylamine N-oxide metabolism and renal excretion in healthy men and women. *Eur. Heart J.* **2019**, *40*, 583–594. [[CrossRef](#)]
191. Park, J.Y.; von Karsa, L.; Herrero, R. Prevention strategies for gastric cancer: A global perspective. *Clin. Endosc.* **2014**, *47*, 478–489. [[CrossRef](#)]
192. Griffin, L.E.; Djuric, Z.; Angiletta, C.J.; Mitchell, C.M.; Baugh, M.E.; Davy, K.P.; Neilson, A.P. A Mediterranean diet does not alter plasma trimethylamine: N -oxide concentrations in healthy adults at risk for colon cancer. *Food Funct.* **2019**, *10*, 2138–2147. [[CrossRef](#)]
193. Martin, F.P.J.; Wang, Y.; Sprenger, N.; Yap, I.K.S.; Lundstedt, T.; Lek, P.; Rezzi, S.; Ramadan, Z.; Van Bladeren, P.; Fay, L.B.; et al. Probiotic modulation of symbiotic gut microbial-host metabolic interactions in a humanized microbiome mouse model. *Mol. Syst. Biol.* **2008**, *4*, 157. [[CrossRef](#)]
194. Tripolt, N.J.; Leber, B.; Triebel, A.; Köfeler, H.; Stadlbauer, V.; Sourij, H. Effect of Lactobacillus casei Shirota supplementation on trimethylamine-N-oxide levels in patients with metabolic syndrome: An open-label, randomized study. *Atherosclerosis* **2015**, *242*, 141–144. [[CrossRef](#)]
195. Qiu, L.; Tao, X.; Xiong, H.; Yu, J.; Wei, H. Lactobacillus plantarum ZDY04 exhibits a strain-specific property of lowering TMAO via the modulation of gut microbiota in mice. *Food Funct.* **2018**, *9*, 4299–4309. [[CrossRef](#)]
196. Qiu, L.; Yang, D.; Tao, X.; Yu, J.; Xiong, H.; Wei, H. Enterobacter aerogenes ZDY01 attenuates choline-induced trimethylamine N-oxide levels by remodeling gut microbiota in mice. *J. Microbiol. Biotechnol.* **2017**, *27*, 1491–1499. [[CrossRef](#)]
197. Dridi, B.; Fardeau, M.L.; Ollivier, B.; Raoult, D.; Drancourt, M. *Methanomassiliicoccus luminyensis* gen. nov., sp. nov., a methanogenic archaeon isolated from human faeces. *Int. J. Syst. Evol. Microbiol.* **2012**, *62*, 1902–1907. [[CrossRef](#)]
198. Poesen, R.; Evenepoel, P.; De Loor, H.; Delcour, J.A.; Courtin, C.M.; Kuypers, D.; Augustijns, P.; Verbeke, K.; Meijers, B. The influence of prebiotic arabinoxylan oligosaccharides on microbiota derived uremic retention solutes in patients with chronic kidney disease: A randomized controlled trial. *PLoS ONE* **2016**, *11*, e0153893. [[CrossRef](#)]
199. Chaplin, A.; Carpené, C.; Mercader, J. Resveratrol, metabolic syndrome, and gut microbiota. *Nutrients* **2018**, *10*, 1651. [[CrossRef](#)]
200. Jung, C.M.; Heinze, T.M.; Schnackenberg, L.K.; Mullis, L.B.; Elkins, S.A.; Elkins, C.A.; Steele, R.S.; Sutherland, J.B. Interaction of dietary resveratrol with animal-associated bacteria. *FEMS Microbiol. Lett.* **2009**, *297*, 266–273. [[CrossRef](#)]
201. Estaki, M.; Pither, J.; Baumeister, P.; Little, J.P.; Gill, S.K.; Ghosh, S.; Ahmadi-Vand, Z.; Marsden, K.R.; Gibson, D.L. Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions. *Microbiome* **2016**, *4*, 42. [[CrossRef](#)]
202. Codella, R.; Luzi, L.; Terruzzi, I. Exercise has the guts: How physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig. Liver Dis.* **2018**, *50*, 331–341. [[CrossRef](#)]
203. Munukka, E.; Ahtiainen, J.P.; Puigbó, P.; Jalkanen, S.; Pakkala, K.; Keskitalo, A.; Kujala, U.M.; Pietilä, S.; Hollmén, M.; Elo, L.; et al. Six-week endurance exercise alters gut metagenome that is not reflected in systemic metabolism in over-weight women. *Front. Microbiol.* **2018**, *9*, 2323. [[CrossRef](#)]
204. Taniguchi, H.; Tanisawa, K.; Sun, X.; Kubo, T.; Hoshino, Y.; Hosokawa, M.; Takeyama, H.; Higuchi, M. Effects of short-term endurance exercise on gut microbiota in elderly men. *Physiol. Rep.* **2018**, *6*, e13935. [[CrossRef](#)]
205. Allen, J.M.; Mailing, L.J.; Niemiro, G.M.; Moore, R.; Cook, M.D.; White, B.A.; Holscher, H.D.; Woods, J.A. Exercise Alters Gut Microbiota Composition and Function in Lean and Obese Humans. *Med. Sci. Sports Exerc.* **2018**, *50*, 747–757. [[CrossRef](#)] [[PubMed](#)]
206. Jäger, R.; Mohr, A.E.; Carpenter, K.C.; Kerksick, C.M.; Purpura, M.; Moussa, A.; Townsend, J.R.; Lamprecht, M.; West, N.P.; Black, K.; et al. International Society of Sports Nutrition Position Stand: Probiotics. *J. Int. Soc. Sports Nutr.* **2019**, *16*, 1–44. [[CrossRef](#)] [[PubMed](#)]
207. Ticinesi, A.; Lauretani, F.; Tana, C.; Nouvenne, A.; Ridolo, E.; Meschi, T. Exercise and immune system as modulators of intestinal microbiome: Implications for the gut-muscle axis hypothesis. *Exerc. Immunol. Rev.* **2019**, *25*, 84–95. [[PubMed](#)]
208. Karl, J.P.; Margolis, L.M.; Madslien, E.H.; Murphy, N.E.; Castellani, J.W.; Gundersen, Y.; Hoke, A.V.; Levangie, M.W.; Kumar, R.; Chakraborty, N.; et al. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2017**, *312*, G559–G571. [[CrossRef](#)]
209. Mills, S.; Lane, J.A.; Smith, G.J.; Grimaldi, K.A.; Ross, R.P.; Stanton, C. Precision nutrition and the microbiome part ii: Potential opportunities and pathways to commercialisation. *Nutrients* **2019**, *11*, 1468. [[CrossRef](#)]
210. Shin, N.R.; Lee, J.C.; Lee, H.Y.; Kim, M.S.; Whon, T.W.; Lee, M.S.; Bae, J.W. An increase in the Akkermansia spp. population induced by metformin treatment improves glucose homeostasis in diet-induced obese mice. *Gut* **2014**, *63*, 727–735. [[CrossRef](#)]
211. Effect of Metformin on Metabolic Improvement and Gut Microbiota. *Appl. Environ. Microbiol.* **2014**, *80*, 5935–5943. [[CrossRef](#)]
212. Wu, M.; Yang, S.; Wang, S.; Cao, Y.; Zhao, R.; Li, X.; Xing, Y.; Liu, L. Effect of Berberine on Atherosclerosis and Gut Microbiota Modulation and Their Correlation in High-Fat Diet-Fed ApoE^{-/-} Mice. *Front. Pharmacol.* **2020**, *11*, 223. [[CrossRef](#)]
213. Zhu, L.; Zhang, D.; Zhu, H.; Zhu, J.; Weng, S.; Dong, L.; Liu, T.; Hu, Y.; Shen, X. Berberine treatment increases Akkermansia in the gut and improves high-fat diet-induced atherosclerosis in ApoE^{-/-} mice. *Atherosclerosis* **2018**, *268*, 117–126. [[CrossRef](#)]
214. Zhang, W.; Xu, J.H.; Yu, T.; Chen, Q.K. Effects of berberine and metformin on intestinal inflammation and gut microbiome composition in db/db mice. *Biomed. Pharmacother.* **2019**, *118*, 109131. [[CrossRef](#)]

215. Wang, H.; Guan, L.; Li, J.; Lai, M.; Wen, X. The effects of berberine on the gut microbiota in APC min/+ mice fed with a high fat diet. *Molecules* **2018**, *23*, 2298. [[CrossRef](#)]
216. Li, C.N.; Wang, X.; Lei, L.; Liu, M.Z.; Li, R.C.; Sun, S.J.; Liu, S.N.; Huan, Y.; Zhou, T.; Liu, Q.; et al. Berberine combined with stachyose induces better glycometabolism than berberine alone through modulating gut microbiota and fecal metabolomics in diabetic mice. *Phyther. Res.* **2020**, *34*, 1166–1174. [[CrossRef](#)]
217. Zhang, T.; Li, Q.; Cheng, L.; Buch, H.; Zhang, F. Akkermansia muciniphila is a promising probiotic. *Microb. Biotechnol.* **2019**, *12*, 1109–1125. [[CrossRef](#)]
218. Crovesy, L.; Masterson, D.; Rosado, E.L. Profile of the gut microbiota of adults with obesity: A systematic review. *Eur. J. Clin. Nutr.* **2020**, *74*, 1251–1262. [[CrossRef](#)]
219. Wang, Z.; Roberts, A.B.; Buffa, J.A.; Levison, B.S.; Zhu, W.; Org, E.; Gu, X.; Huang, Y.; Zamanian-Daryoush, M.; Culley, M.K.; et al. Non-lethal Inhibition of Gut Microbial Trimethylamine Production for the Treatment of Atherosclerosis. *Cell* **2015**, *163*, 1585–1595. [[CrossRef](#)]
220. Roberts, A.B.; Gu, X.; Buffa, J.A.; Hurd, A.G.; Wang, Z.; Zhu, W.; Gupta, N.; Skye, S.M.; Cody, D.B.; Levison, B.S.; et al. Development of a gut microbe-targeted nonlethal therapeutic to inhibit thrombosis potential. *Nat. Med.* **2018**, *24*, 1407–1417. [[CrossRef](#)]
221. Dambrova, M.; Skapare-makarova, E.; Konrade, I.; Pugovics, O.; Grinberga, S.; Tirzite, D.; Petrovska, R.; Kalvins, I.; Liepins, E. Meldonium decreases the diet-increased plasma levels of trimethylamine n-oxide, a metabolite associated with atherosclerosis. *J. Clin. Pharmacol.* **2013**, *53*, 1095–1098. [[CrossRef](#)]
222. Kuka, J.; Liepinsh, E.; Makrecka-Kuka, M.; Liepins, J.; Cirule, H.; Gustina, D.; Loza, E.; Zharkova-Malkova, O.; Grinberga, S.; Pugovics, O.; et al. Suppression of intestinal microbiota-dependent production of pro-atherogenic trimethylamine N-oxide by shifting L-carnitine microbial degradation. *Life Sci.* **2014**, *117*, 84–92. [[CrossRef](#)]
223. Konop, M.; Radkowski, M.; Grochowska, M.; Perlejewski, K.; Samborowska, E.; Ufnal, M. Enalapril decreases rat plasma concentration of TMAO, gut bacteria-derived cardiovascular marker. *Biomarkers* **2018**, *23*, 380–385. [[CrossRef](#)]
224. Velebova, K.; Hoang, T.; Veleba, J.; Belinova, L.; Kopecky, J.; Kuda, O.; Segetova, M.; Kopecky, J.; Melenovsky, V.; Pelikanova, T. The effect of metformin on serum levels of Trimethylamine-N-oxide in patients with type 2 diabetes/prediabetes and chronic heart failure. *Diabetologia* **2016**, *59*, S533.
225. Kuka, J.; Videja, M.; Makrecka-Kuka, M.; Liepins, J.; Grinberga, S.; Sevostjanovs, E.; Vilks, K.; Liepinsh, E.; Dambrova, M. Metformin decreases bacterial trimethylamine production and trimethylamine N-oxide levels in db/db mice. *Sci. Rep.* **2020**, *10*, 14555. [[CrossRef](#)] [[PubMed](#)]
226. Kalagi, N.A.; Abbott, K.A.; Alburikan, K.A.; Alkofide, H.A.; Stojanovski, E.; Garg, M.L. Modulation of Circulating Trimethylamine N-Oxide Concentrations by Dietary Supplements and Pharmacological Agents: A Systematic Review. *Adv. Nutr.* **2019**, *10*, 876–887. [[CrossRef](#)] [[PubMed](#)]
227. Jalandra, R.; Dalal, N.; Yadav, A.K.; Verma, D.; Sharma, M.; Singh, R.; Khosla, A.; Kumar, A.; Solanki, P.R. Emerging role of trimethylamine-N-oxide (TMAO) in colorectal cancer. *Appl. Microbiol. Biotechnol.* **2021**, *105*, 7651–7660. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.