



Draft Genome Sequence of *Venenivibrio stagnispumantis* CP.B2^T, Isolated from Champagne Pool, Waiotapu, Aotearoa-New Zealand

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ABSTRACT *Venenivibrio stagnispumantis* strain CP.B2^T is a thermophilic, chemolithoautotrophic bacterium from the family *Hydrogenothermaceae* (phylum *Aquificota*), isolated from Champagne Pool in the Waiotapu geothermal field, Aotearoa-New Zealand. The genome consists of 1.73 Mbp in 451 contigs with a 30.8 mol% G+C content.

The bacterial genus *Venenivibrio*, within the phylum *Aquificota* (family *Hydrogenothermaceae*), is characterized by thermophilic microaerophilic hydrogenotrophs (1–3). The type strain for the genus, *Venenivibrio stagnispumantis* CP.B2^T, was isolated from Champagne Pool, Waiotapu, Aotearoa-New Zealand, a geothermal system under the *kaitiakitanga* (guardianship) of Māori *iwi* (tribe) Ngāti Tahu-Ngāti Whaoa. The *V. stagnispumantis* genome has previously been sequenced as part of the Genomic Encyclopedia of Bacteria and Archaea (GEBA) project KMG-II (4) (IMG genome ID 2724679818). However, we have resequenced the genome on behalf of Ngāti Tahu-Ngāti Whaoa to recognize indigenous data sovereignty and to assert *mana whenua* (customary rights) over the bacterium, its genome, and the location in which it was isolated.

Venenivibrio stagnispumantis CP.B2^T (obtained from the University of Waikato, Aotearoa-New Zealand) was cultivated using a modified MSH medium (70°C, pH 5.5) within a headspace consisting of N₂/H₂/CO₂/O₂ at ratios of 50:40:7.5:2.5 (vol/vol), respectively (1). A total DNA extract (3.5 ng μl⁻¹) of CP.B2^T was performed using a NucleoSpin Soil kit (Macherey-Nagel, Düren, Germany) according to the manufacturer's protocols. Whole-genome sequencing (Macrogen, Inc., Seoul, South Korea) was undertaken using the Illumina HiSeq 2500 platform with TruSeq DNA Nano (550) library preparation for paired-end 101-bp reads (Illumina, Inc., San Diego, CA, USA). Default parameters were used for all assembly, quality control (QC), and annotation software unless otherwise specified. A total of 3.8-Gbp raw sequences (37.32 million reads) were assembled using SPAdes v3.12.0 (5) with the “-careful” option to minimize mismatches and kmer sizes of 21, 33, and 55. The assembly was evaluated using QUAST v5.0.0 (6). The genome was annotated via the NCBI Prokaryotic Genome Annotation Pipeline (PGAP; v6.3 [7]) (GenBank accession no. [JAPEIW010000000](https://www.ncbi.nlm.nih.gov/nuccore/JAPEIW010000000)) using the GeneMarkS-2+ protein reference set. A functionally equivalent annotation of 47 scaffolds using the Integrated Microbial Genomes annotation pipeline v4.16.4 (8) is also available (IMG genome ID 2799112217).

The *V. stagnispumantis* CP.B2^T genome comprised 1,731,156 bp containing 2,034 protein-coding genes, with 1,963 having predicted function. The genome consisted of 451 contigs, with a total G+C content of 30.8% mol. The *N*₅₀ value was 91,033, the *L*₅₀ was 8, and genome coverage was ~2,242×. The genome encodes 49 RNAs, including single complete copies of the 5S, 16S, and 23S rRNAs; 36 tRNAs; and 3 noncoding RNAs (ncRNAs).

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A pairwise average nucleotide identity (ANI) comparison using FastANI v0.1.3 (9) of the *V. stagnispumantis* CP.B2^T, *Sulfurihydrogenibium yellowstonense* SS-5^T, and *Persephonella marina* EX-H1^T genomes gave <80% similarity for each, confirming the designation of *Venenivibrio* as a separate genus to sister genera *Sulfurihydrogenibium* and *Persephonella*.

Data availability. Raw sequences for the *V. stagnispumantis* CP.B2^T genome were deposited in the European Nucleotide Archive under the BioProject accession no. [PRJEB55610](https://www.ebi.ac.uk/bioproject/155610). The assembled genome was deposited in the Genomes Online Database (10) (GOLD analysis ID [Ga0591133](https://www.genomesonline.org/analysis/Ga0591133)), for associated annotation with the Integrated Microbial Genomes & Microbiomes platform (8) (IMG genome ID [2799112217](https://img.jgi.doe.gov/2799112217)). This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under accession no. [JAPEIW010000000](https://www.ncbi.nlm.nih.gov/nuccore/JAPEIW010000000). The version described in this paper is version [JAPEIW010000000.1](https://www.ncbi.nlm.nih.gov/nuccore/JAPEIW010000000.1).

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REFERENCES

- Hetzer A, McDonald IR, Morgan HW. 2008. *Venenivibrio stagnispumantis* gen. nov., sp. nov., a thermophilic hydrogen-oxidizing bacterium isolated from Champagne Pool, Waiotapu, New Zealand. *Int J Syst Evol Microbiol* 58:398–403. <https://doi.org/10.1099/ijs.0.64842-0>.
- Hetzer A, Morgan HW, McDonald IR, Daughney CJ. 2007. Microbial life in Champagne Pool, a geothermal spring in Waiotapu, New Zealand. *Extremophiles* 11:605–614. <https://doi.org/10.1007/s00792-007-0073-2>.
- Power JF, Carere CR, Lee CK, Wakerley GLJ, Evans DW, Button M, White D, Climo MD, Hinze AM, Morgan XC, McDonald IR, Cary SC, Stott MB. 2018. Microbial biogeography of 925 geothermal springs in New Zealand. *Nature Comms* 9:2876. <https://doi.org/10.1038/s41467-018-05020-y>.
- Whitman WB, Woyke T, Klenk HP, Zhou Y, Lilburn TG, Beck BJ, De Vos P, Vandamme P, Eisen JA, Garrity G, Hugenholtz P, Kyrpides NC. 2015. Genomic encyclopedia of bacterial and archaeal type strains, phase III: the genomes of soil and plant-associated and newly described type strains. *Stand Genomic Sci* 10:26. <https://doi.org/10.1186/s40793-015-0017-x>.
- Prijbelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes *de novo* assembler. *Curr Protoc Bioinformatics* 70:e102. <https://doi.org/10.1002/cpbi.102>.
- Mikheenko A, Prijbelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>.
- Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Chen IA, Chu K, Palaniappan K, Ratner A, Huang J, Huntemann M, Hajek P, Ritter S, Varghese N, Seshadri R, Roux S, Woyke T, Eloe-Fadrosh EA, Ivanova NN, Kyrpides NC. 2021. The IMG/M data management and analysis system v.6.0: new tools and advanced capabilities. *Nucleic Acids Res* 49:D751–D763. <https://doi.org/10.1093/nar/gkaa939>.
- Jain C, Rodriguez RL, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>.
- Mukherjee S, Stamatis D, Bertsch J, Ovchinnikova G, Sundaramurthi JC, Lee J, Kandimalla M, Chen IA, Kyrpides NC, Reddy TBK. 2021. Genomes OnLine Database (GOLD) v.8: overview and updates. *Nucleic Acids Res* 49:D723–D733. <https://doi.org/10.1093/nar/gkaa983>.