



Published in final edited form as:

Am J Med Genet A. 2022 March ; 188(3): 959–964. doi:10.1002/ajmg.a.62584.

Identification and validation of a novel pathogenic variant in *GDF2* (BMP9) responsible for hereditary hemorrhagic telangiectasia and pulmonary arteriovenous malformations

Srimmitha Balachandar¹, Tamara J. Graves¹, Anika Shimonty¹, Katie Kerr², Jill Kilner², Sihao Xiao^{3,4}, Richard Slade^{3,4}, Manveer Sroya⁵, Mary Alikian^{4,6}, Emanuel Curetean⁶, Ellen Thomas^{6,7}, Vivienne P. M. McConnell⁸, Shane McKee⁸, Freya Boardman-Pretty⁷, Andrew Devereau⁷, Tom A. Fowler^{7,9}, Mark J. Caulfield^{7,9}, Eric W. Alton^{3,4}, Teena Ferguson⁶, Julian Redhead⁶, Amy J. McKnight^{2,4}, Geraldine A. Thomas⁵, Genomics England Research Consortium⁷, Micheala A. Aldred^{1,4}, Claire L. Shovlin^{3,4,6}

¹Division of Pulmonary, Critical Care, Sleep and Occupational Medicine, Department of Medicine, Indiana University School of Medicine, Indianapolis, Indiana, USA

²School of Medicine, Dentistry and Biomedical Sciences, Queen's University Belfast, Belfast, UK

³National Heart and Lung Institute, Imperial College London, London, UK

⁴Genomics England Respiratory Clinical Interpretation Partnership (GeCIP), London, UK

⁵Department of Surgery and Cancer, Imperial College London, London, UK

⁶West London Genomic Medicine Centre, Imperial College Healthcare NHS Trust, London, UK

⁷Genomics England, London, UK

⁸Regional Genetics Service, Belfast Health and Social Care Trust, Belfast, UK

⁹William Harvey Research Institute, Queen Mary University of London, London, UK

Abstract

Correspondence: Claire L. Shovlin, NHLI Cardiovascular Sciences, Imperial Centre for Translational and Experimental Medicine, Imperial College London, Hammersmith Campus, Du Cane Road, London W12 0NN, UK. c.shovlin@imperial.ac.uk, Micheala A. Aldred, Division of Pulmonary, Critical Care, Sleep & Occupational Medicine, Department of Medicine, Indiana University School of Medicine, 980 W Walnut St, Indianapolis, IN 46202, USA. maaldred@iu.edu.

AUTHOR CONTRIBUTIONS

Micheala A. Aldred and Claire L. Shovlin wrote the manuscript. Srimmitha Balachandar, Tamara J. Graves, and Anika Shimonty performed in vitro functional studies. Katie Kerr and Jill Kilner performed recall sample preparations. Manveer Sroya, Mary Alikian, Emanuel Curetean, Eric W. Alton, Teena Ferguson, Julian Redhead, Geraldine A. Thomas, and Claire L. Shovlin contributed to project set up at West London Genomic Medicine Centre, family recruitment; and sample preparations. Vivienne P. M. McConnell, Shane McKee, Amy J. McKnight, and Claire L. Shovlin set up recall sampling. Genomics England Research Consortium performed WGS. Srimmitha Balachandar, Tamara J. Graves, Anika Shimonty, Sihao Xiao, Richard Slade, Ellen Thomas, Micheala A. Aldred, and Claire L. Shovlin contributed to specific data analyses. Freya Boardman-Pretty, Andrew Devereau, Tom A. Fowler and Mark J. Caulfield contributed to specific project set up project at Genomics England. All authors have reviewed and approved the final manuscript.

CONFLICT OF INTEREST

The authors declare no potential conflict of interest.

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.

Hereditary hemorrhagic telangiectasia (HHT) is an autosomal dominant multisystemic vascular dysplasia, characterized by arteriovenous malformations (AVMs), mucocutaneous telangiectasia and nosebleeds. HHT is caused by a heterozygous null allele in *ACVRL1*, *ENG*, or *SMAD4*, which encode proteins mediating bone morphogenetic protein (BMP) signaling. Several missense and stop-gain variants identified in *GDF2* (encoding BMP9) have been reported to cause a vascular anomaly syndrome similar to HHT, however none of these patients met diagnostic criteria for HHT. HHT families from UK NHS Genomic Medicine Centres were recruited to the Genomics England 100,000 Genomes Project. Whole genome sequencing and tiering protocols identified a novel, heterozygous *GDF2* sequence variant in all three affected members of one HHT family who had previously screened negative for *ACVRL1*, *ENG*, and *SMAD4*. All three had nosebleeds and typical HHT telangiectasia, and the proband also had severe pulmonary AVMs from childhood. In vitro studies showed the mutant construct expressed the proprotein but lacked active mature BMP9 dimer, suggesting the mutation disrupts correct cleavage of the protein. Plasma BMP9 levels in the patients were significantly lower than controls. In conclusion, we propose that this heterozygous *GDF2* variant is a rare cause of HHT associated with pulmonary AVMs.

Keywords

arteriovenous malformations; bone morphogenetic protein 9; hereditary hemorrhagic telangiectasia

1 | INTRODUCTION

Hereditary hemorrhagic telangiectasia (HHT; syn. Osler–Weber–Rendu syndrome) is a multisystemic vascular dysplasia inherited as an autosomal dominant trait. It is diagnosed clinically by the presence of at least three diagnostic criteria, known as the Curaçao criteria: spontaneous, recurrent nose bleeds; multiple telangiectases at characteristic sites (lips, oral cavity, fingers, nose); visceral lesions such as gastrointestinal telangiectasia (with or without bleeding); arteriovenous malformations (AVMs) in the lung, liver, brain or spine; and a first degree relative with HHT according to these criteria (Shovlin et al., 2000). Nosebleeds are the most common symptom in HHT and frequently lead to iron deficiency anemia. Pulmonary AVMs (PAVMs) affect at least 50% of HHT patients, are usually silent, but cause low blood oxygen levels (hypoxemia), paradoxical ischemic strokes, and cerebral abscess (Shovlin, 2014). The majority of genotyped HHT patients have a heterozygous pathogenic DNA sequence variant in *ENG*, encoding endoglin (*ENG*, *HHT1*) (McAllister et al., 1994), or *ACVRL1*, encoding activin receptor-like kinase (*ALK1*, *HHT2*) (Johnson et al., 1996). A smaller proportion have HHT due to a heterozygous pathogenic variant in *SMAD4*, which also cause other pathologies, including juvenile polyposis and aortopathy (Gallione et al., 2004; Heald et al., 2015). Current sequencing strategies do not detect causal variants in ~10% of genotyped HHT patients (Shovlin et al., 2020).

BMP9 is a specific ALK1 ligand encoded by *GDF2* (David et al., 2007). BMP9/10 inhibition in neonatal mice leads to hyper-vascularization and induces retinal AVMs (Ruiz et al., 2016). Heterozygous missense substitutions in *GDF2* were first identified in three of 191 unrelated individuals with nosebleeds and skin telangiectasia, who did not have a pathogenic

variant in any of the established HHT genes (*ACVRL1*, *ENG*, or *SMAD4*) (Wooderchak-Donahue et al., 2013). However, the telangiectases were atypical for HHT and considered more consistent with a capillary-malformation AVM syndrome (Wooderchak-Donahue et al., 2013). With the recent expansion of population-scale whole genome sequencing (WGS), one of these variants (p.Arg333Trp) is found in 0.1% of control chromosomes, and thus its pathogenicity is now uncertain (Table S1). More recently, two children with PAVMs and other features suggestive of HHT were identified with homozygous *GDF2* mutations (one frameshift, one nonsense) (Hodgson et al., 2021; Liu et al., 2020), while a third patient with a homozygous nonsense mutation initially presented with pulmonary arterial hypertension (PAH) (Wang et al., 2016), and was later found to have multiple spider-like telangiectases on the face (Hodgson et al., 2021). Again, the telangiectases in these three cases were not typical of HHT (Hodgson et al., 2021; Liu et al., 2020). Remarkably, all of the heterozygous parents and one homozygous sibling had few or no HHT features. In summary, none of the patients previously reported with *GDF2* variants met Curaçao criteria (Hodgson et al., 2021; Liu et al., 2020; Wooderchak-Donahue et al., 2013).

2 | CLINICAL DESCRIPTION

The pedigree for the family is indicated in Figure 1a. The proband (III.1) was diagnosed with HHT during childhood due to frequent nosebleeds, mucocutaneous telangiectasia, and PAVMs. He had presented at the age of 8 years with cyanosis, and was diagnosed with multiple PAVMs that were too diffuse for treatment. He initially deteriorated steadily, with oxygen saturations (SaO₂) less than 80% at rest, and by the age of 11 years was only able to walk a few steps. A possible heart–lung transplant was declined by the family. He was commenced on nifedipine, based on theoretical considerations by his pediatrician, and improved functionally, though remained cyanosed and clubbed. At the time of recruitment to the 100,000 Genomes Project when classical HHT telangiectasia were confirmed, he was 38 years old, working full time, with a low normal exercise capacity (Medical Research Council dyspnea scale 1 [normal], New York Heart Association Class I [normal], VSAQ varying between 8 and 9 corresponding to metabolic equivalents of ~10–11 kcal/kg/min) (Fletcher, 1960; Myers et al., 1994; New York Heart Association, 1964). This corresponded to the upper tertile of a recent PAVM patient cohort (range 1.33–15.55 kcal/kg/h [median 8.84, interquartile range 6.64–11.86 kcal/kg/h]) (Gawecki et al., 2019). The SaO₂ was 85% standing at rest, and he was compensating for this by polycythemia (hemoglobin 201 g/L, hematocrit 0.558). On cardiology workup, he was considered to have no significant pulmonary hypertension. His DNA had been sequenced for *ACVRL1*, *ENG*, and *SMAD4* with no variants identified.

The proband's sister (III.2) realized she was likely to be affected with HHT when nosebleeds developed in her 30s. She received a definite clinical diagnosis aged 41 years due to the affected sibling, nosebleeds, and typical tiny HHT telangiectasia over the lips and face (none on finger pads, mouth, ears, or conjunctiva). A thoracic CT scan identified no pulmonary AVMs, and SaO₂ was normal. The proband's mother (II.2) received a positive clinical diagnosis at the age of 76 years due to severe nosebleeds as a child, subtle telangiectasia on the lips, and affected children. Again SaO₂ was normal. An uncle (II.3) had died during

a childhood tonsillectomy due to excessive bleeding. The proband's maternal grandmother (I.2) died of an unusual stroke in her 50s.

3 | METHODS

3.1 | Editorial policies and ethical considerations

This study was approved by the East of Scotland Research Ethics Service (16/ES/0095). All participants provided written informed consent.

3.2 | Sequencing and analysis pipelines

Detailed methodology is provided in the Supporting Information. Due to the absence of a variant in the known HHT genes, the family was identified through the UK National Health Service (NHS) as suitable for the 100,000 Genomes Project. For HHT, Tier 1 variants were defined as clear loss of function variants in *ENG*, *ACVRL1*, and *SMAD4*, Tier 2 as other variants in these genes, and Tier 3 as protein truncating, de novo or protein altering variants in other genes. Genomics England performed all WGS, data alignments, and initial variant tiering using Illumina pipeline alignments and variant calling. Separately, a Respiratory GeCIP (Clinical Interpretation Partnership) was established to analyze full WGS/phenotypic datasets. Customized scripts were written in Python to extract all variants in HHT patients' variant call format files for single nucleotide variants, small indels and larger copy number variants. Data were exported under Project Code RR42.

3.3 | In vitro functional analysis

Functional consequences of the *GDF2* variant were studied by in vitro expression of mutant and wild-type plasmids. Briefly, site-directed mutagenesis was used to introduce the variant into a full-length human *GDF2* cDNA clone. Plasmids were transfected into 293T cells and cultured for 5 days. Western blot analysis was performed on cell lysates and conditioned media, using antibodies that specifically target the mature BMP9 protein or the proprotein. Enzyme-linked immunosorbent assay (ELISA) was performed on conditioned media samples using the DuoSet Human BMP9 kit (R&D Systems) according to the manufacturer's instructions. To determine if the secreted BMP9 was biologically active, control pulmonary artery endothelial cells were incubated with conditioned medium for 4 h, then harvested for RNA extraction and analyzed by realtime quantitative RT-PCR for expression of the BMP-responsive genes *ID1* and pre-miR-21.

3.4 | BMP9 concentration in human plasma

Plasma samples were obtained from the three affected family members, one unaffected relative (II.1), and four unrelated healthy controls. Circulating BMP9 levels were measured by ELISA as described above, with the addition of 0.1% heat inactivated goat serum to avoid nonspecific binding.

4 | RESULTS

In keeping with the earlier clinical sequencing studies, WGS and tiering protocols identified no Tier 1 or Tier 2 variants in the family. The family had 76 Tier 3 variants, that is, protein

truncating or protein altering variants in other genes, present in the affected mother and children, but absent in the unaffected father. Inspection of these Tier 3 variants identified a novel, heterozygous *GDF2* missense variant (c.1282T>C, p.Cys428Arg). This variant is predicted to be deleterious (SIFT) and probably damaging (PolyPhen), with a Combined Annotation Dependent Depletion (CADD) score of 28.8 (Rentzsch et al., 2019). It has been reported once in 152,092 WGS alleles (population frequency of 6.58×10^{-6}) in gnomAD v3.1.1 (<https://gnomad.broadinstitute.org>). Sanger sequencing confirmed the presence of this variant in all three HHT-affected family members (Figure 1b). Six other Tier 3 variants were identified that probably modify function, but were not likely candidate genes for HHT on the basis of their known biology (Table S2). Through the 100,000 Genomes Project Data Release 6.0, WGS data were available for 160 HHT participants from 126 families. No other *GDF2* variants were identified in these families.

Given previous reports of *GDF2* variants in patients with HHT-like features, this variant was prioritized for further analysis. The concentration of circulating mature BMP9 protein measured in plasma was significantly lower for the three HHT patients who carry the variant than in samples from five unaffected individuals (the father (II.1) and four unrelated controls), which were collected and processed in parallel (median 2.65 vs. 7.43 pg/ml, $p = 0.036$) (Figure 1c). Notably, the severely affected proband had the lowest BMP9 level of all. He had become increasingly unwell in the months preceding plasma collection, and had been diagnosed with decompensated cirrhosis without evidence of hepatic AVMs. It is unclear whether the very low circulating BMP9 and his liver failure are related.

BMP9 is synthesized as a proprotein, which then dimerizes and is cleaved to produce the mature secreted protein. Western blotting of cells transfected with the wild-type or mutant plasmid demonstrated expression of the proprotein in all cells, whereas the mature protein was not detected from the mutant construct, suggesting the c.1282T>C variant disrupts processing of BMP9 (Figure 2a). This was supported by analysis with the proprotein antibody, where two species were detected in wild-type cells—the uncleaved and cleaved proprotein—whereas in mutant cells, the uncleaved protein predominated (Figure S1). Western blotting of conditioned media confirmed secretion of the mature BMP9 dimer from the wild-type plasmid (20 kDa), together with a larger partially cleaved form detected at about 57 kDa, whereas neither were detected in medium from the mutant construct (Figure S2). ELISA confirmed that mature BMP9 was undetectable in conditioned media from the mutant construct, even at a 1 in 10 dilution, whereas the concentration in wild-type medium was 973 ng/ml (Figure S3). Furthermore, whereas wild-type medium induced a fivefold increase in *IDI* expression in cultured endothelial cells and a threefold increase in pre-miR-21, conditioned media from the mutant construct failed to upregulate these BMP-responsive target genes (Figure 2b). *IDI* is a target of canonical BMP signaling, whereas miR-21 is upregulated via a noncanonical *SMAD4*-independent pathway that regulates microRNA maturation (Davis et al., 2008).

5 | DISCUSSION

We identified a missense *GDF2* variant in three affected members of a family with HHT. Results of our in vitro studies suggest this mutation disrupts normal cleavage of BMP9

proprotein, leading to a reduction in the active mature dimer form. Consistent with this, the level of mature protein detectable in plasma from the three mutation-positive patients was more than 2.5-fold lower than controls. Although *GDF2* mutations have previously been identified in individuals with a vascular anomaly syndrome similar to HHT, this is the first family that meets Curaçao criteria. Table S1 shows the characteristics of missense *GDF2* variants reported in patients with features of HHT. Of note, the c.1282T>C variant in this family has the highest CADD pathogenicity score, whereas some variants now appear likely benign on the basis of low pathogenicity scores, and population frequencies greater than 1/10,000 (Table S1).

Highly variable phenotypes between HHT family members affected by the same pathogenic variant are the norm in HHT, although the discrepancy in HHT phenotype between the proband and other two cases in this pedigree is unusually marked. With the exception of the proband, the clinical phenotype in the other HHT-affected individuals was mild, and in the absence of the proband's history it is unlikely that their diagnosis would have been suspected. Nevertheless, they both had three Curaçao Criteria, unlike other *ACVRL*^{+/−} and *ENG*^{+/−} confirmed HHT cases in our cohort (Sharma et al., 2021), and were not “atypical” as seen by us for patients with *EPHB4*^{+/−} and other vasculopathies (Shovlin et al., 2021). WGS identified the scale of genetic variation between the family members, and studies to identify variants potentially relevant to the pulmonary AVM and cirrhotic phenotypes are ongoing. Cirrhosis has been recognized in HHT patients but is not considered part of the direct HHT phenotype (Silvain et al., 2020; VASCERN, 2019), instead usually ascribed to infective or toxin-based etiologies not relevant to individual III.1. Notably, although BMP9 is synthesized in the liver, this is predominantly from nonparenchymal cells (Miller et al., 2000), and therefore it is not currently known if the cirrhosis in the proband was specifically related to the *GDF2* mutation.

The majority of *GDF2* variants reported to date have been found in patients with PAH (Gräf et al., 2018; Wang et al., 2019), a severe progressive lung vasculopathy that is also associated with mutations of the BMP pathway, especially the type-II BMP receptor (*BMPR2*), but has a distinct clinical spectrum. Many of the *GDF2* variants in PAH are also missense and have similar effects in reducing the secretion of mature BMP9 protein (Hodgson et al., 2020; Wang et al., 2019). Thus at present, there is no obvious genotype–phenotype correlation. Some phenotypic crossover between the two diseases is already known, since a small subset of HHT patients develop PAH, while at least two PAH patients with *BMPR2* mutations have been reported with PAVM (Rigelsky et al., 2008; Soon et al., 2014). *GDF2* now adds to this enigma, highlighting the importance of understanding potential modifier genes and second hits in the pathogenesis of both HHT and PAH.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGMENTS

The authors are indebted to the family for their participation in this study. This research was made possible through access to the data and findings generated by the 100,000 Genomes Project. The 100,000 Genomes Project

is managed by Genomics England Limited (a wholly owned company of the Department of Health and Social Care). The 100,000 Genomes Project is funded by the National Institute for Health Research (NIHR) and NHS England. The Wellcome Trust, Cancer Research UK, and the Medical Research Council have also funded research infrastructure. The 100,000 Genomes Project uses data provided by patients and collected by the National Health Service as part of their care and support. The authors thank the National Health Service staff of the UK Genomic Medicine Centre and the families for their willing participation. This research was co-funded by the NIHR Imperial Biomedical Research Centre, and by NHLBI grant R35HL140019 (to M.A.A.). The views expressed are those of the authors and not necessarily those of funders, the NHS, the NIHR, or the Department of Health and Social Care. This research was co-funded by the NIHR Imperial Biomedical Research Centre, and Medical Research Council – Northern Ireland Executive support of the Northern Ireland Genomic Medicine Centre (MC_PC_16018). K.K. was supported by a Department for the Economy Co-operative Awards in Science and Technology (DfE-CAST) PhD studentship award with Belfast Health and Social Care Trust.

Funding information

National Institute for Health Research (NIHR) and NHS England, Grant/Award Number: The 100,000 Genomes Project; National Heart, Lung, and Blood Institute, Grant/Award Number: R35HL140019; National Institute for Health Research, Grant/Award Number: Imperial Biomedical Research Centre 2017–2022; Medical Research Council – Northern Ireland Executive: Northern Ireland Genomic Medicine Centre, Grant/Award Number: MC_PC_16018; Department for the Economy Co-operative Awards in Science and Technology (DfE-CAST)

DATA AVAILABILITY STATEMENT

Primary data from the 100,000 Genomes Project, which are held in a secure Research Environment, are available to registered users. Please see <https://www.genomicsengland.co.uk/about-gecip/for-gecip-members/data-and-data-access> for further information.

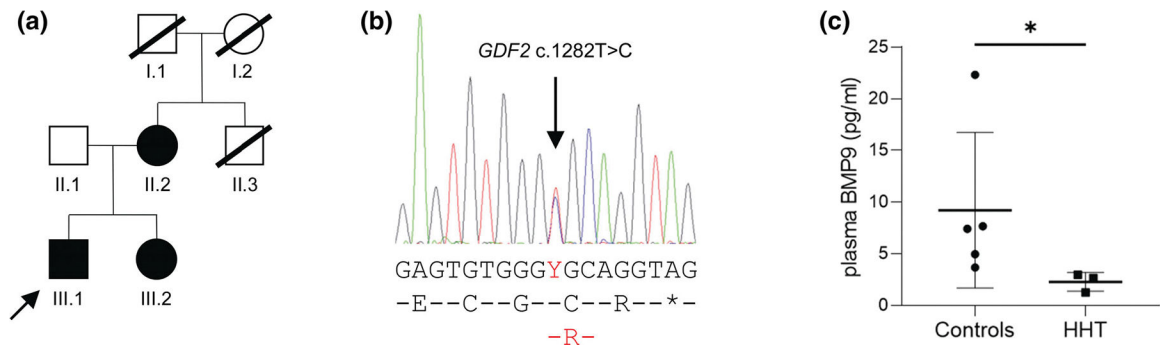
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**FIGURE 1.**

HHT family with *GDF2* missense variant and reduced plasma BMP9 levels. (a) Pedigree at the time of family recruitment. The proband (III.1) is arrowed. (b) Sanger sequencing trace from individual III.2 showing the heterozygous *GDF2* variant. Nucleotide sequence and protein translation are depicted below, with variants denoted in red font. (c) Circulating BMP9 levels in human plasma from the three individuals carrying the c.1282T>C variant were significantly lower than in five unaffected individuals collected and analyzed under the same conditions (* $p = 0.036$, Mann-Whitney two-tailed test)

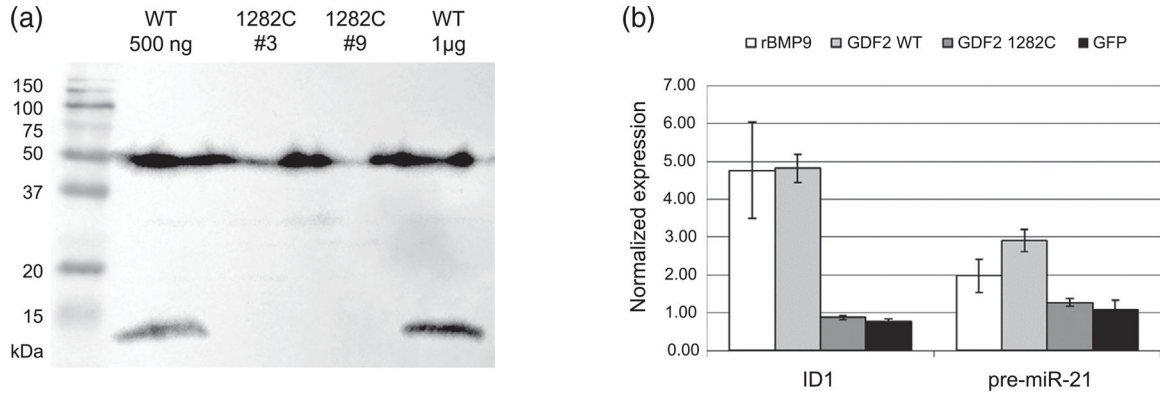


FIGURE 2.

GDF2 c.1282T>C disrupts processing of BMP9 protein. (a) Western blot of lysates from 293T cells transfected with wild-type (WT; 500 ng or 1 μg of DNA) or mutant (1282C, clones #3 and #9; 1 μg) *GDF2* plasmids, run under reducing conditions and probed with antibody to human BMP9. Full length protein (50 kDa) is expressed from the mutant plasmids, but is not correctly processed, leading to absence of the mature monomer (12 kDa). (b) Conditioned medium from these cells was applied to cultures of human pulmonary artery endothelial cells (PAEC). Medium from the wild-type (WT) but not mutant (c.1282C) plasmid induces mRNA expression of BMP-responsive genes, *ID1* and pre-miR-21. Recombinant BMP9 (rBMP9) was used as a positive control, and medium from GFP transfection as a negative control. Expression was normalized to GAPDH, and fold change was calculated relative to untreated PAEC