Neurodevelopmental and Epilepsy Phenotypes in Individuals With Missense Variants in the Voltage-Sensing and Pore Domains of *KCNH5*

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Abstract

Background and Objectives
KCNH5 encodes the voltage-gated potassium channel EAG2/Kv10.2. We aimed to delineate the neurodevelopmental and epilepsy phenotypic spectrum associated with de novo *KCNH5* variants.

Methods
We screened 893 individuals with developmental and epileptic encephalopathies for *KCNH5* variants using targeted or exome sequencing. Additional individuals with *KCNH5* variants were identified through an international collaboration. Clinical history, EEG, and imaging data were analyzed; seizure types and epilepsy syndromes were classified. We included 3 previously published individuals including additional phenotypic details.

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Glossary

ACMG = American College of Medical Genetics and Genomics; **DEE** = developmental and epileptic encephalopathy; **EAG** = ether-a-go-go; **ID** = intellectual disability; **GOF** = gain of function; **MIP** = molecular inversion probe; **NDD** = neurodevelopmental disorder; **VUS** = variants of uncertain significance.

Results

We report a cohort of 17 patients, including 9 with a recurrent de novo missense variant p.Arg327His, 4 with a recurrent missense variant p.Arg333His, and 4 additional novel missense variants. All variants were located in or near the functionally critical voltage-sensing or pore domains, absent in the general population, and classified as pathogenic or likely pathogenic using the American College of Medical Genetics and Genomics criteria. All individuals presented with epilepsy with a median seizure onset at 6 months. They had a wide range of seizure types, including focal and generalized seizures. Cognitive outcomes ranged from normal intellect to profound impairment. Individuals with the recurrent p.Arg333His variant had a self-limited drug-responsive focal or generalized epilepsy and normal intellect, whereas the recurrent p.Arg327His variant was associated with infantile-onset DEE. Two individuals with variants in the pore domain were more severely affected, with a neonatal-onset movement disorder, early-infantile DEE, profound disability, and childhood death.

Discussion

We describe a cohort of 17 individuals with pathogenic or likely pathogenic missense variants in the voltage-sensing and pore domains of Kv10.2, including 14 previously unreported individuals. We present evidence for a putative emerging genotype-phenotype correlation with a spectrum of epilepsy and cognitive outcomes. Overall, we expand the role of EAG proteins in human disease and establish *KCNH5* as implicated in a spectrum of neurodevelopmental disorders and epilepsy.

The voltage-gated potassium channels are a large group of transmembrane proteins critical for controlling neuronal excitability and regulating electrophysiologic properties in the brain.[1] The ether-a-go-go (EAG) subfamily consists of 2 members, EAG1/Kv10.1 encoded by *KCNH1* and EAG2/Kv10.2 encoded by *KCNH5*. Pathogenic gain-of-function (GOF) variants in *KCNH1* are implicated in Temple-Baraitser and Zimmermann-Laband syndromes[2-4]; epilepsy is a common feature of these overlapping syndromes.[5] Individuals with variants in *KCNH5* have been reported in large exome sequencing cohort studies of individuals with neurodevelopmental disorders (NDDs). Of the 3 previously reported individuals, 2 had a recurrent missense variant (p.Arg327His) associated with a developmental and epileptic encephalopathy (DEE) with onset prior to 1 year of age.[6,7] The third had a neonatal-onset DEE and a different de novo missense variant (p.Ile463Thr).[8] Despite these case reports, the *KCNH5*-associated phenotypes are not well delineated.

KCNH5 is expressed in excitatory neurons in upper layer IV of the cerebral cortex and the hippocampus.[9,10] *KCNH5* encodes for Kv10.2, which localizes to the somatodendritic region where it plays a role in controlling the electrical coupling between cell bodies and distal dendrites.[10] Like most potassium channels, it consists of 4 transmembrane domains (S1–S4) that make up the voltage-sensing domain and 2 transmembrane domains and a reentrant loop (S5–S6) that comprise the pore module (Figure 1, A and C). The recurrent Kv10.2 p.Arg327His variant localizes to S4, where voltage-clamp analysis in a heterologous expression system demonstrated a hyperpolarized shift of voltage dependence of activation and

acceleration of activation, consistent with a gain-of-function pathogenic mechanism.[11]

Here we report a cohort of 17 individuals with de novo or inherited (n = 1) missense variants in the voltage-sensing and pore domains of *KCNH5*. We describe 14 new patients and provide additional phenotypic information for the 3 published individuals from large cohort studies.[6-8] We analyze the spectrum of *KCNH5*-associated neurodevelopment disorders, particularly the epilepsy phenotypes, and discuss genotype-phenotype correlations.

Methods

Standard Protocol Approvals, Registrations, and Patient Consents

We performed *KCNH5* targeted resequencing in 680 individuals with DEEs and exome sequencing in 213 individuals with DEEs, totaling 893 individuals. All individuals underwent clinical assessment, and medical records, EEG, and imaging data were obtained. Seizure types and epilepsy syndromes were classified according to the ILAE classification.[12,13] Individuals with DEEs had been previously tested for pathogenic variants in the majority of known DEE genes ([14] and unpublished data). The study was approved by the Austin Health Human Research Ethics Committee (H2007/02961), the New Zealand Health and Disability Ethics Committee (NTY/12/06/053/AM15), and the University Hospital of Lyon and Aix-Marseille University in France Ethics Committee (05/78, CPP Strasbourg Alsace 1). Written informed consent was provided by the

Figure 1 Location and Conservation of the 6 Unique De Novo *KCNH5* Missense Variants
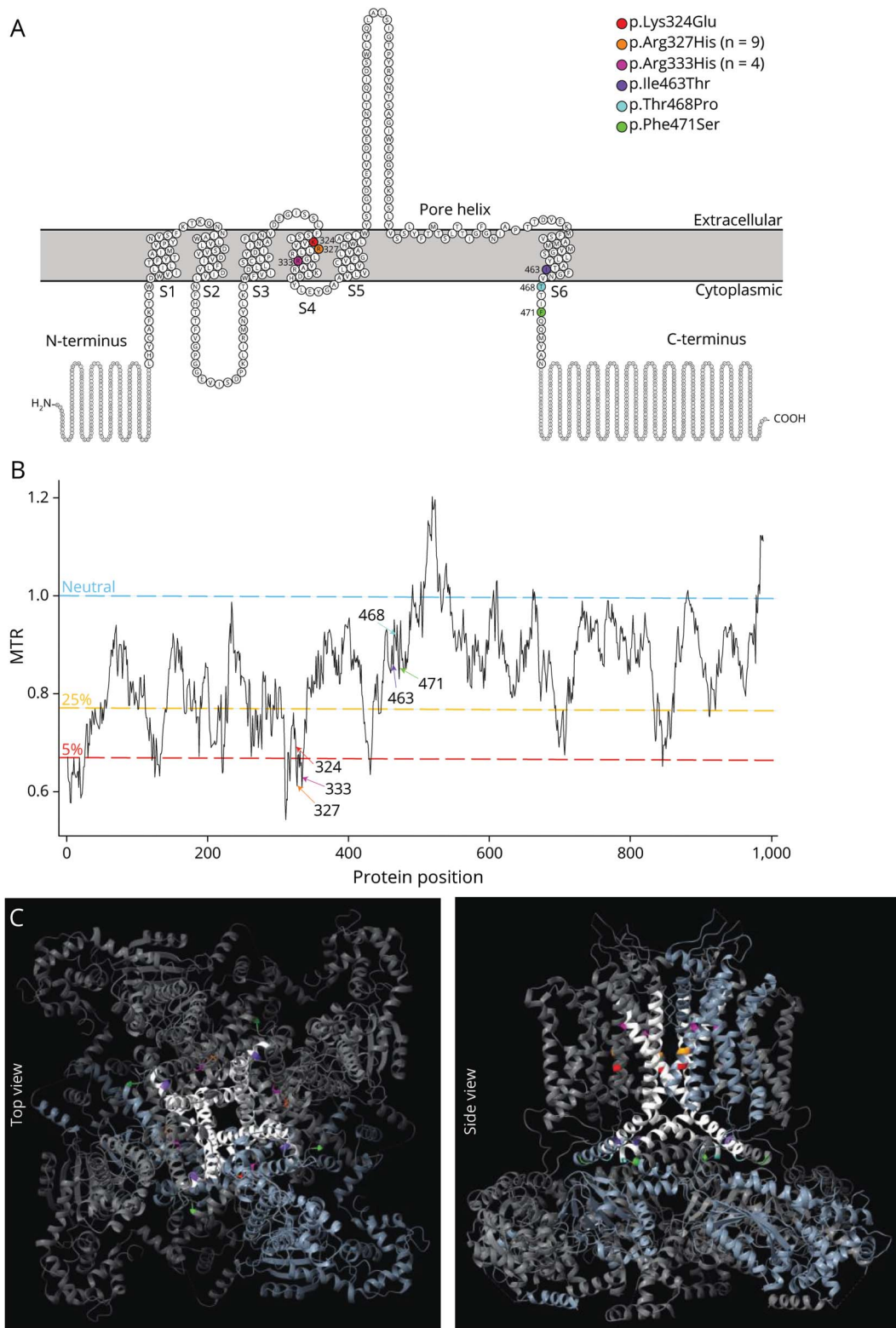
(A) Schematic of Kv10.2 (EAG2), modified from Protter[35] plot of Q8NCM2 (KCNH5_Human). Of the 6 transmembrane domains, S1-S4 make up the voltage-sensing domain, and S5-6 form the pore helix. The *KCNH5* patient-specific variants identified in this study are indicated by their amino acid change. (B) Graph of missense tolerance ratio (MTR; y-axis) and protein position (x-axis), with variants indicated. MTR[36] is a measure of protein-encoding cDNA sequence intolerance to missense variants. Variant positions with a value greater than 1 (blue line) is considered neutral; values below 1 are under constraint. Two variants (p.Arg327His and p.Arg333His) are in the 5th percentile of least tolerated missense alterations in the exome. The 5th and 25th percentiles are highlighted in red and yellow, respectively. (C) Locations of variants (color coded) are mapped onto the crystal structure of homotetrameric assembly of Kv10.1 (PDB5K7L); all variants are perfectly conserved between KCNH5 and KCNH1 and are mapped to the corresponding amino acid. The pore domain is highlighted in white, and a single tetrameric subunit is colored light blue. EAG = ether-a-go-go

patient or their parent or legal guardian in the case of minors or those with intellectual disability (ID).

We identified an additional 14 individuals with KCNH5 variants through GeneMatcher.[15] These individuals were consented using research protocols approved by their local ethics committees. Their clinicians provided clinical information. For the 3 previously published individuals (probands 2, 10, and 15), we obtained additional clinical information. In total, the cohort consists of 19 individuals with KCNH5 missense variants, with 17 having variants localized in or near the voltage-sensing or pore domains.

KCNH5 Variant Identification and Analysis

We used molecular inversion probes (MIPs) to capture all exons and 5 base pairs of flanking intronic KCNH5 sequence; next-generation sequencing and data analysis were performed as described previously.[14] We resequenced KCNH5, covering 95% of the gene at a depth of 50X (median across the cohort of 680 individuals). Exome sequencing (n = 213) was performed via Epi25.[16] We considered only variants that were nonsynonymous, altered the acceptor/donor splice sites, or indels that disrupted the coding frame. Only variants that were not present in gnomAD[17] and TOPMed[18] were considered for segregation analysis. The variants in the 14 individuals recruited by the matchmaker exchange network[15] were identified via clinical or research genome (n = 2), exome (n = 10), or gene panel (n = 2) sequencing.[16] Library preparation and data analysis were performed using local protocols and computational pipelines. Variants were classified according to the American College of Medical Genetics and Genomics (ACMG) guidelines for variant classification.[19]

KCNH1 (EAG1/Kv10.1) and KCNH5 (EAG2/Kv10.2) have 72% identity at the amino acid level. The 3-dimensional structure of Kv10.2 has not yet been resolved; thus, to model the location of each of the identified missense variants, we used the Kv10.2 homology model generated from the Kv10.1 cryo-EM structure (5k7l.1.A) using UCSF ChimeraX.[20]

Data Availability

All data from this study are available in the manuscript or supplementary materials.

Results

Rare Missense Variants in KCNH5

We identified 2 individuals with the previously reported de novo missense variant p.Arg327His in 2 DEE cohorts (1/680 by MIPs and 1/213 by exome[16]). Through our international collaboration, we identified 5 additional individuals with this recurrent variant, which arose de novo in 4 individuals (Table 1). We also identified 5 additional missense variants in the voltage-sensing and pore domains of Kv10.2 (Figure 1). One variant (p.Arg333His) was present in 4 individuals; it arose de novo in 3 and was inherited from an affected mother in the fourth proband. The 4 other de novo missense variants were each observed once in single individuals.

The amino acids at all 6 variant positions (2 recurrent, 4 singletons) were conserved between Kv10 proteins. The missense variants had CADD[21] scores of 26–32, conferring a high likelihood of deleteriousness (Figure 1B and Table 1). All variants were absent from the general population (gnomAD and TOPMed) and were classified as either pathogenic or likely pathogenic (P/LP) by the ACMG criteria (eTable 1, links.lww.com/WNL/C453).[19]

We also identified 2 de novo variants of uncertain significance (VUS) in the N-terminal (p.Leu181Pro) and C-terminal (p.Ile606Thr) of Kv10.2 in 2 individuals with NDDs but without seizures (eAppendix 1, links.lww.com/WNL/C453). These VUS were located outside of the functionally critical pore or voltage-sensing domains. One variant (p.Leu181Pro) did not segregate with the NDD in the family, and the individual with the p.Ile606Thr variant also carried a pathogenic 16p11.2 microdeletion that may explain some or all of their clinical features. Although these variants are included here for completeness, this study focuses on the 17 individuals with variants in the voltage-sensing or pore-forming domains.

Phenotypic Features in Individuals With KCNH5 Pathogenic or Likely Pathogenic Variants

All 17 individuals with pathogenic or likely pathogenic missense variants in the voltage-sensing or pore domains had epilepsy (Table 1). Moreover, the mother of proband 14 who carries the recurrent p.Arg333His variant also has epilepsy, although detailed clinical information was not available. The median age at seizure onset was 6 months (range: 1–12 months). A wide range of seizure types were seen: focal impaired awareness seizures (8), focal to bilateral tonic-clonic seizures (9), focal motor seizures (8), generalized myoclonic (4), generalized tonic-clonic seizure (2), myoclonic-atonic (1), absence seizure (1), unknown onset tonic-clonic seizures (6), and epileptic spasms (2). Nine (53%) individuals had a DEE. The remainder had focal epilepsy (3), generalized epilepsy (2), or there was inadequate information to make an epilepsy type diagnosis (3). Nine individuals (53%) achieved seizure freedom, with median seizure offset at 2.5 years (range: 3 months to 8 years).

Developmental concerns were noted at a median age of 15 months in 14 individuals (range birth to 3.5 years), predominantly affecting language (11). One child had normal development, and for 2 individuals, aged 61 years, early developmental information was limited. Cognitive outcome for the 10 individuals over 5 years of age ranged from normal (3/10) to mild (3/10), moderate (2/10), severe (1/10), and profound (1/10) ID. Two individuals died at ages 13 months and 9 years, respectively.

There were no consistent or significant dysmorphic features noted in these individuals. Proband 1 was reported to have long eyelashes and mild ptosis, whereas proband 2 had prominent ears, small hands, and a flexion contracture of the 4th and 5th digits of one hand. Significant sleep disorders were not noted in any individual.

Table 1 Clinical Presentation of Our Cohort and Previously Published Individuals With P/LP *KCNH5* Missense Variants

Proband (age at study, sex)	Protein change (CADD)	Inheritance	Syndrome	Age at sz onset: type	Other sz type(s)	Sz offset	Development (age of first concern); regression (age)	Developmental and behavioral outcome	EEG	MRI	ASMs (positive response underlined)
1 (4y, M)	p.Lys324Glu (27)	De novo	DEE	3m: FMS (Hc)	FBTCS, SE, and TCS	N/A	GDD (18m); no	Sev ID (single words, amb, and assisted feeds; ADHD	3m: N; 10m: FD 11m, 18m: N	3m: N	<u>LEV</u>, <u>PHT</u>, <u>CLB</u>, <u>VPA</u>, and <u>LTG</u>
2[Veeramah] (13y, M)	p.Arg327His (32)	De novo	FDEE	6m: TCS	TCS and FMS (Hc)	N/A	GDD (6m); yes (3y)	Sev ID (NV); ASD	Freq MFD (sleep activation)	NP	<u>VPA</u> (partial), others unknown
3 (5.5y, M)		De novo	DEE: D/EE-SWAS	5m: FIAS	FBTCS and FMS (Hc)	3y	Language delay (2.5y); yes (2.5y)	Mild/Mod ID (sentences and amb); ADHD	6m: FMS; 9m, 11m: MFD; 2.8y CSWS; 3y: MFD (sleep activation); 11y: focal slow	6m: N, 4y: N	PB, <u>CBZ</u>, CLB, LEV, <u>STR</u>, VPA, and TPM
4 (61y, F)		Parents unavailable	Epilepsy: unknown type	6m: Unknown	GTCS, Abs, and FAMS	N/A	Unknown; no	N (learning difficulties at school; sentences, amb, and self-feeds)	56y: focal slow, FD, and GSW	55y: N	CBZ, LEV, PB, PHT, TPM, VPA, CBZ, <u>PGB</u>, and <u>LTG</u>
5 (5y, F)		De novo	FDEE	6m: FS	FMS and FBTCS	N/A	Language delay (16m); no	Mod ID (2-word sentences, amb, and self-feeds); autistic trait	8m: N; 12m: MFD; 2y 9m: slow	NP	<u>OXC and LEV</u>
6 (10y, F)		De novo	FDEE	7m: FIAS	FBTCS and FMS (Hc)	N/A	Language and behavior delay (3.5y); no	Mild ID (sentences, amb, and self-feeds); behavioral difficulties	7m: N; 3y: FD; 5y, 6y, 8y, 10y: slow, Freq MFD (sleep activation)	1y, 3y, 5y: N	VPA (partial), LEV, <u>LCS</u> (partial), TPM, LTG, CLB, CBZ, ETX, GBP, CBD, and <u>KD</u>
7 (9y, M)		De novo	DEE	10m: FIAS	MyAS, My, and TCS	6y	Language delay (15m); no	Mod ID (sentences, amb, and self-feeds)	1y: slow; 5y, 6y: Freq MFD (sleep activation)	10m, 4y: N	CBZ (partial) and <u>LTG</u>
8 (3.8y F)		De novo	FDEE	2m: FMS	FBTCS and FIAS	N/A	Language delay (15m); no	Sev ID (2-word sentences, amb, and assisted feeds); ASD	2y 9m: Slow, FD (sleep activation)	11m, 18m: N	LEV, LTG, CLB, and <u>CBZ</u> (partial)
9 (19m, M)		De novo	CE	7m: GTCS	FBTCS	14 m	Language delay (14m); no	Language delay (single words, amb, and self-feeds)	9m: GSW; 11m, 13m: N	14m: N	LEV and <u>OXC</u>
10[Minardi] (61y, M)		Parents unavailable	DEE	<1 y: TCS	None	8y	GDD (unknown); no	Mild ID (sentences, amb, and self-feeds)	54, 56-59y: slow, diffuse MFD (sleep activation)	52y, 53y: multiple nonenhancing white matter abnormalities; arachnoid cyst; mineral deposits in the orbit (eFigure 1, links.lww.com/WNL/C453)	LEV and <u>LTG</u>

Continued

Table 1 Clinical Presentation of Our Cohort and Previously Published Individuals With P/LP *KCNH5* Missense Variants *(continued)*

Proband (age at study, sex)	Protein change (CADD)	Inheritance	Syndrome	Age at sz onset: type	Other sz type(s)	Sz offset	Development (age of first concern); regression (age)	Developmental and behavioral outcome	EEG	MRI	ASMs (positive response underlined)
11 (3.5y, F)	p.Arg333His (30)	De novo	FE	8m: FIAS	FBTCS	12m	N; no	N (sentences, amb, and self-feeds)	11m, 3y: N	13m: N	VPA
12 (7y, F)		De novo	Epilepsy: unknown type	9m: TCS	None	5y	Language delay (18m); no	N (sentences, amb, and self-feeds)	15m: N; 7y: FD	15m, 7y: N	LEV, TPM, and ZNS
13 (10y, F)		De novo	FE	10m: FIAS	FBTCS	1y	Language and gross motor delay (18m); no	N (sentences, amb, and self-feeds); behavioral difficulties	12m:, 2y,: focal slow, 3y:N	4y, 9y: frontal atrophy (not progressive); arachnoid cyst; Chiari I malformation (eFigure 2)	VPA, weaned at 4y
14 (1.3y, F)		Inherited from affected mother	GE	2m: My	None	3m	Hypotonia (4m); no	N (single words and walks with support)	2m: GSW; 3m: N	4m: N	VPA
15[Imafidon] (11m, F) (deceased)	p.Ile463Thr (28)	De novo	DEE: EIDEE	3m: TCS	ES and My	N/A	Hypotonia, GDD (birth); yes (4m)	Profound DD	1d: N; 3m, 5m and 7m: FD; 10m: slow, MFD (continuous)	9d: N; 3m: delayed myelination and mild cerebral atrophy	VPA, CLN, and LEV
16 (9y, F) (deceased)	p.Thr468Pro (26)	De novo	DEE: EIDEE	1m: FTS	ES, TCS, and My	N/A	Hypotonia, GDD (birth); no	Profound ID: NV, NA, and G-tube	1w: N; 1m: FD; 1y: BS; 2y: slow, MFD, GSW; 5y, 6y: MFD, BS	3w: increased signal posterior fossa; 2y (CT): cerebral atrophy	VPA, KD, VGB, STR, PN, PLP, TPM, LEV (partial), and PB (partial)
17 (12y, F)	p.Phe471Ser (31)	De novo	DE + FE	1y: TCS	FIAS	5y	GDD (9m); no	Mod ID (sentences, amb, and self-feeds)	1y, 4y, 7y: FD (sleep activation)	1y, 3y: N	VPA, weaned at 11y

Abbreviations: Abs = absence seizures; amb = ambulatory; ADHD = attention-deficit hyperactivity disorder; ASD = autism spectrum disorder; ASM = antiseizure medication; BS = burst suppression; CBD = cannabidiol; CBZ = carbamazepine; CE = combined focal and generalized epilepsy; CLB = clobazam; CLN = clonazepam; CSWS = continuous spike and wave in slow wave sleep; d = day; DD = developmental delay; DEE = developmental and epileptic encephalopathy; D/EE-SWAS = developmental and/or epileptic encephalopathy with spike-wave activation in sleep; DE + FE = developmental encephalopathy and focal epilepsy; EIDEE = early infantile DEE; ES = epileptic spasm; ETX = ethosuximide; FAMS = focal aware motor seizure; FBTCS = focal to bilateral tonic-clonic seizure; FD = focal discharge; FDEE = focal DEE; FE = focal epilepsy; FIAS = focal impaired awareness seizure; FMS = focal motor seizure; FMy = focal myoclonic seizure; FTS = focal tonic seizure; Freq = frequent; FS = febrile seizure; GBP = gabapentin; GDD = global developmental delay; GE = generalized epilepsy; GSW = generalized spike and wave; GTCS = generalized tonic-clonic seizure; Hc = hemiclonic; ID = intellectual disability; LEV = levetiracetam; KD = ketogenic diet; LCS = lacosamide; LTG = lamotrigine; m = months; MFD = multifocal discharge; Mod = moderate; My = myoclonic; MyAS = myoclonic-atonic seizure; N = normal; N/A = not applicable; NA = nonambulatory; NP = not performed; NV = nonverbal; OXC = oxcarbazepine; PB = phenobarbital; PGB = pregabalin; PHT = phenytoin; PLP = pyridoxal-5-phosphate; P/LP = pathogenic/likely pathogenic; PM = primary motor; PN = pyridoxine; SE = status epilepticus; Sev = severe; STR = steroid; Sz = seizure; TCS = tonic-clonic seizure; TPM = topiramate; VAF = variant allele frequency; VGB = vigabatrin; VPA = valproate; VUS = variants of uncertain significance; w = weeks; y = years; ZNS = zonisamide.
References indicate previously published.

For the 9 individuals with the recurrent voltage-sensing domain p.Arg327His variant, seizure onset occurred at a median age of 6 months (range 2–10 months). For the 8 individuals for whom an epilepsy syndrome diagnosis could be made, all but one, who was 19 months old, had a DEE (7/8), which was focal in 5 individuals. Proband 3 had the syndrome of epileptic encephalopathy with spike-and-wave activation in sleep, which was associated with language regression.[12] All patients had EEGs, which captured sleep. Sleep activation of epileptiform discharges was noted in 6 other patients but was not associated with developmental regression. The cognitive outcome for those over 5 years of age with this recurrent variant ranged from normal with learning difficulties (1/6) to mild (3/6), moderate (1/6), or severe ID (1/6). For those in whom the complete antiseizure medication history was available, their epilepsy was drug resistant (8/9), with only 4/9 eventually becoming seizure-free. Brain MRI was normal in all apart from proband 10. He had no early imaging, but an MRI performed at 52 years demonstrated nonenhancing periventricular white matter abnormalities suggestive of demyelinating lesions. The lesions were stable on a repeat MRI 6 months later. Note was also made of an arachnoid cyst and mineral deposits in the orbit (eFigure 1, links.lww.com/WNL/C453).

The phenotype was much less severe for the 4 individuals with the second recurrent voltage-sensing domain p.Arg333His variant. This group had onset at a similar age (2–10 months) with either focal or generalized seizures. In contrast, their seizures were drug responsive, particularly to valproate (3/4), and all became seizure-free (aged 3 months to 5 years). One of the 4 individuals had mild motor and language delay at presentation, 1 had mild language delay, and 1 had only mild motor delay. All had normal cognitive development as assessed by their clinician when last seen at ages 1–10 years. MRI was normal apart from proband 13 who had mild frontal cortical atrophy, an arachnoid cyst, and a Chiari I malformation (eFigure 2, links.lww.com/WNL/C453).

Strikingly, probands 15 (p.Ile463Thr) and 16 (p.Thr468Pro), with missense variants in or at the junction of the S6 transmembrane pore-forming domain, had a much more severe phenotype. Both individuals presented with persistent nonepileptic myoclonus on day 1 of life and developed an early-infantile DEE with drug-resistant seizures by 3 months and eventual epileptic spasms. They had very limited developmental progression. Neuroimaging showed cortical atrophy. Proband 15 died at 13 months due to respiratory failure secondary to increased seizure activity. Proband 16 died at 9 years due to pneumonia.

There were 2 additional individuals with likely pathogenic variants and epilepsy. Proband 1, with p.Lys324Glu variant in the voltage-sensing domain, had a similar phenotype to individuals with the nearby recurrent p.Arg327His variant. This individual had a focal DEE with focal seizures beginning at 3 months and developmental delay evident by 18 months, evolving to severe ID. Proband 17 had the p.Phe471Ser variant in the C-terminal

cytoplasmic domain, which was very close to the pore-forming domain. She had moderate ID, drug-responsive focal epilepsy, and a hyperkinetic movement disorder consisting of a tremor predominantly in the upper limbs and exacerbated when she was tired or unwell. She did not have an epileptic encephalopathy and presented a similar epilepsy phenotype to patients with the recurrent p.Arg333His variant. We provide clinical vignettes for 3 patients (probands 6, 13, and 16) to highlight the spectrum of phenotypes associated with *KCNH5* pathogenic variants (eAppendix 2, Clinical Vignettes, links.lww.com/WNL/C453).

Discussion

Potassium channel subunits are one of the most important groups of genes associated with epilepsy, in particular with the most severe group of epilepsies, the DEEs.[22] Here, we show that de novo pathogenic variants in *KCNH5* in 17 patients are frequently associated with a DEE but may also cause self-limited epilepsies. We describe the phenotypic spectrum, with particular emphasis on the phenotype-genotype correlations and the putative effect of these missense variants on Kv10.2 channel function.

We highlight a genotype-phenotype correlation for variants across the protein (Figure 2). The p.Arg333His variant is associated with the mildest epilepsy phenotype, a drug-responsive self-limited focal or generalized epilepsy with normal cognitive outcome. Conversely, the nearby most common recurrent p.Arg327His variant has a far more severe epilepsy phenotype of DEE. Of interest, the nearby p.Lys324Glu voltage-sensing domain variant was also associated with a DEE. Individuals with these 2 variants predominantly presented with an infantile-onset DEE between 2 and 10 months of age with drug-resistant generalized and focal seizures. Early developmental concerns were noted with cognitive outcome ranging from learning difficulties to severe ID. Finally, 2 individuals with variants in the pore-forming S6 transmembrane domain (p.Ile463Thr and p.Thr468Pro) had an even more severe phenotype of neonatal-onset DEE with onset on day 1 of life resulting in profound disability and death.

The p.Arg327His variant, located in the voltage-sensing S4 transmembrane domain, acts in a GOF manner.[11] Multistate structural modeling demonstrated that the p.Arg327His variant favors channel opening, and voltage-clamp experiments showed a hyperpolarized shift of voltage dependence of activation and acceleration of activation.[11] These changes in electrophysiologic properties are consistent with the GOF mechanism that is typically observed for several potassium channels implicated in DEEs, such as *KCNH1*,[2] *KCNT1*,[23] and *KCNT2*.[24] A subset of epilepsy-associated variants in *KCNA2, KCNB1, KCNQ2,* and *KCNQ3* have similarly been shown to act in a GOF manner.[25-27] How these changes lead to epilepsy remains unresolved.[28] We hypothesize that the new *KCNH5* voltage-sensing and pore domain pathogenic variants described here also have a similar GOF mechanism to the p.Arg327His variant. This is supported by in silico data

Figure 2 Phenotypic Spectrum Associated With *KCNH5* Missense Variants
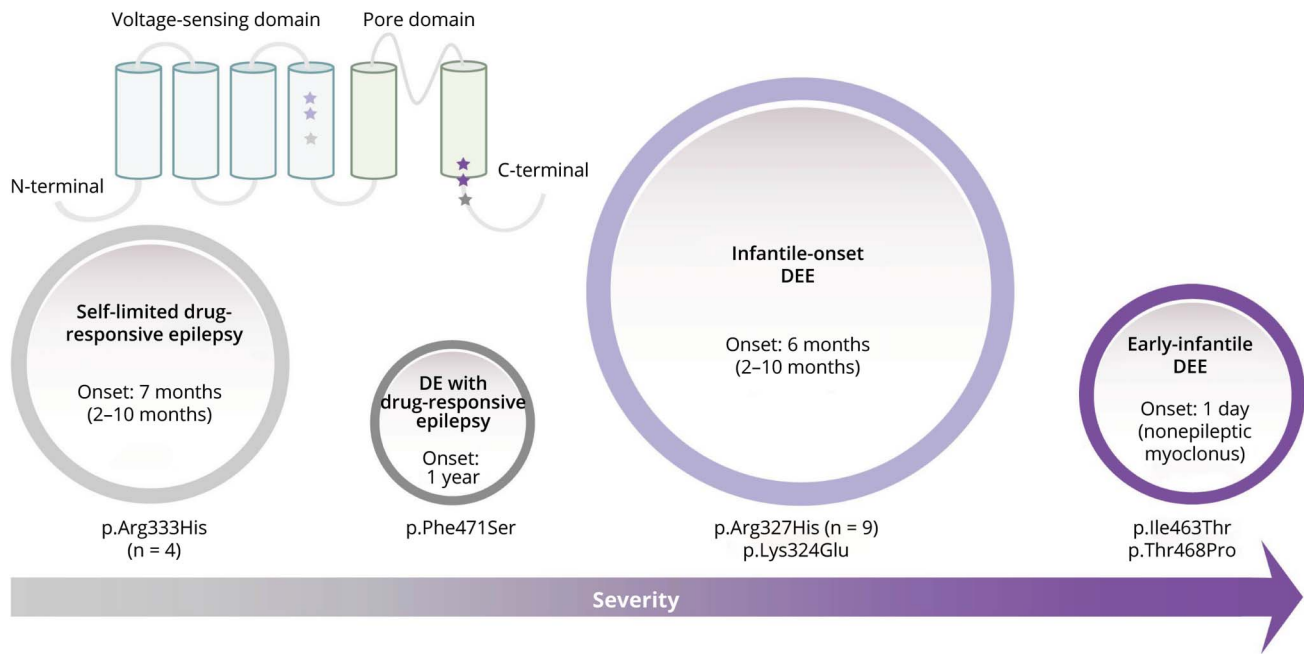
The number of individuals in the phenotype group is represented by circle size, and colors of each circle and star match the variant/phenotype class. The individuals with the p.Lys324Glu and p.Arg327His variants in the S4 transmembrane domain presented with an infantile-onset DEE with drug-resistant generalized and focal seizures, whereas the 4 individuals with the nearby recurrent p.Arg333His variant had drug-responsive seizures and became seizure-free. The 2 individuals with variants in or directly at the junction of the S6 transmembrane domain (p.Ile463Thr and p.Thr468Pro) are the most severely affected with a neonatal-onset movement disorder and early-infantile DEE. The single individual with the nearby p.Phe471Ser variant was less severely affected with moderate ID and drug-responsive seizures. Range is indicated in parentheses where applicable. DE = developmental encephalopathy; DEE = developmental and epileptic encephalopathy.

including the tolerance of *KCNH5* to loss-of-function (truncating) variants in the general population. *KCNH5* has a probability of loss-of-function intolerant (pLI) score of 0.02[29]. Epilepsy-associated genes that confer pathogenicity via haploinsufficiency typically have pLI scores ~1, highlighting that loss of function is unlikely to be the pathogenic mechanism for *KCNH5* variants. Conversely, missense variants in the gene are not as well tolerated, with a z-score of 2.51 (Figure 1B), suggesting selection against certain missense variants throughout human evolution.

The 2 most severely affected individuals carried missense variants (p.Ile463Thr and p.Thr468Pro) that cluster right at the junction of the S6 transmembrane domain and the early C-terminal cytoplasmic domain that form the channel pore. The 2 missense variants are located within 5 amino acids of each other right at the channel pore. Missense variants are similarly found in and just outside the S6 domain of the EAG1/Kv10 pore encoded by *KCNH1* in individuals with Temple-Baraitser syndrome, which includes epilepsy as a key feature. Detailed electrophysiologic studies of these S6 missense variants in *KCNH1* showed delayed deactivation and decreased threshold of activation in both human cells and *Xenopus* oocytes, consistent with a GOF mechanism.[2,3] We hypothesize that the pore-forming *KCNH5* missense variants act in a similar GOF manner. Like other voltage-gated potassium channels, Kv10.2 comprises 4 subunits that form a homotetramer. Any given tetramer could have between zero and 4 subunits containing the variant residue, and it is possible that the function of the resulting channels could be differentially disrupted. This principle is a challenge in ion channel physiology generally, and in particular, for epilepsy, new tools and models are currently being developed to address this challenge. Moreover, future studies will also be necessary to confirm the functional effect and determine whether missense pathogenic variants in the most central pore region have a more profound GOF effect on Kv10.2 function than those in the voltage-sensing domain or located further away from the pore.

De novo missense variants in *KCNH1*/Kv10.1 cause Temple-Baraitser syndrome (TBS) and Zimmermann-Laband syndrome (ZLS).[2-4] Features of both include ID, epilepsy, facial dysmorphism, and nail hypoplasia.[30] GOF *KCNH1* missense variants have recently been reported in individuals with ID and epilepsy without the additional phenotypic features of TBS/ZLS.[31,32] Individuals with both syndromic and nonsyndromic *KCNH1*-associated disorders with epilepsy have both focal and generalized seizures in infancy.[5] Three of the 6 reported KCNH5 variants (p.Lys324Glu, p.Arg327His, and p.Ile463Thr) affect analogous KCNH1 amino acid residues that have been reported in individuals with KCNH1-associated NDDs,[2,4,32] further highlighting the importance of these positions on channel function. *KCNH1* and *KCNH5* are predominantly expressed in the adult CNS (Human Protein Atlas[33] and GTEx[34]); however, current expression data sets are largely limited to adult tissues.

Particularly as all 6 amino acid residues affected by the nucleotide changes described here are conserved between KCNH1 and KCNH5 (eFigure 3, links.lww.com/WNL/C453), future data sets such as the Developmental Genotype-Tissue Expression initiative will be valuable for evaluating both the multisystem involvement of *KCNH1*-associated disorders and the CNS-limited *KCNH5*-related phenotypes.

Overall, we establish *KCNH5* as a gene associated with epilepsy and neurodevelopmental phenotypes and identify an emerging genotype-phenotype correlation. We describe 17 individuals with pathogenic or likely pathogenic variants in *KCNH5*, including 4 novel variants. The p.Arg327His variant located in the voltage-sensing domain causes gain of channel function,[11] and we hypothesize that the additional *KCNH5* voltage-sensing and pore domain variants also lead to epilepsy via a similar GOF mechanism. We highlight an intriguing genotype-phenotype correlation with a spectrum of epilepsy and cognitive outcomes. These observations will be further supported by examining the functional effects associated with the location of different variants in Kv10.2, with potential therapeutic implications. Our study expands the role of EAG proteins in human disease, highlighting that *KCNH5* variants are implicated in a spectrum of NDD and epilepsy phenotypes.

Accession Numbers

KCNH5 mRNA NM_139318.4 and protein NP_647479.2 sequence.

KCNH1 protein NP_758872.1.

Disclosure

Publication History

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Appendix *(continued)*

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