



Review

The Vascular-Immune Hypothesis of Alzheimer's Disease

Rashi I. Mehta ^{1,2,*} and Rupal I. Mehta ^{3,4,*}

¹ Department of Neuroradiology, Rockefeller Neuroscience Institute, West Virginia University, Morgantown, WV 26506, USA

² Department of Neuroscience, Rockefeller Neuroscience Institute, West Virginia University, Morgantown, WV 26506, USA

³ Rush Alzheimer's Disease Center, Rush University Medical Center, Chicago, IL 60612, USA

⁴ Department of Pathology, Rush University Medical Center, Chicago, IL 60612, USA

* Correspondence: rashi.mehta@hsc.wvu.edu (R.I.M.); rupal_mehta@rush.edu (R.I.M.)

Abstract: Alzheimer's disease (AD) is a devastating and irreversible neurodegenerative disorder with unknown etiology. While its cause is unclear, a number of theories have been proposed to explain the pathogenesis of AD. In large part, these have centered around potential causes for intracerebral accumulation of beta-amyloid (β A) and tau aggregates. Yet, persons with AD dementia often exhibit autopsy evidence of mixed brain pathologies including a myriad of vascular changes, vascular brain injuries, complex brain inflammation, and mixed protein inclusions in addition to hallmark neuropathologic lesions of AD, namely insoluble β A plaques and neurofibrillary tangles (NFTs). Epidemiological data demonstrate that overlapping lesions diminish the β A plaque and NFT threshold necessary to precipitate clinical dementia. Moreover, a subset of persons who exhibit AD pathology remain resilient to disease while other persons with clinically-defined AD dementia do not exhibit AD-defining neuropathologic lesions. It is increasingly recognized that AD is a pathologically heterogeneous and biologically multifactorial disease with uncharacterized biologic phenomena involved in its genesis and progression. Here, we review the literature with regard to neuropathologic criteria and incipient AD changes, and discuss converging concepts regarding vascular and immune factors in AD.

Keywords: Alzheimer's disease; beta-amyloid (β A); glymphatic-lymphatic system; mixed pathologies; neurovascular unit (NVU); perivascular unit (PVU); tau



Citation: Mehta, R.I.; Mehta, R.I. The Vascular-Immune Hypothesis of Alzheimer's Disease. *Biomedicines* **2023**, *11*, 408. <https://doi.org/10.3390/biomedicines11020408>

Academic Editor: Carolina Alquezar Burillo

Received: 2 January 2023

Revised: 26 January 2023

Accepted: 27 January 2023

Published: 30 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Alzheimer's disease (AD) is a chronic neuropathological condition that manifests from a host of biological factors that collectively cause damage and loss of neurons, synapses, and supporting cells of the brain (i.e., glia), ultimately manifesting with brain atrophy [1]. Clinically, AD dementia features memory and executive function deficits, with subsequent progressive, global cognitive decline [1,2]. Overall, AD is the sixth leading cause of death in the United States. Approximately 6 million Americans are currently afflicted with AD dementia and in the absence of effective therapies, this figure is expected to expand to approximately 14 million by 2050 [2].

Early-onset AD (EOAD) occurs before the age of 65 years and is often associated with autosomal dominant inheritance, whereas late-onset AD (LOAD) manifests after age 65 years and does not typically exhibit Mendelian transmission [3]. EOAD is caused by highly penetrant mutations in amyloid precursor protein (APP), presenilin 1 (PSEN1), or presenilin 2 (PSEN2) genes, while nonfamilial cases may involve trisomy of chromosome 21 that harbors the APP gene [4–6]. These genetic aberrations influence brain levels and quality of beta-amyloid (β A) peptide, the primary constituent of β A plaques. Apolipoprotein E (ApoE) is linked with LOAD and distinct ApoE isoforms have been shown to

divergently regulate β A production and aggregation [7]. Both EOAD and LOAD are characterized by preclinical and prodromal, i.e., mild cognitive impairment (MCI), stages that together span decades [2].

Despite knowledge on genetic associations and incipient phases of disease, the precise etiology of AD remains uncertain [8]. Investigation is complicated by the fact that many patients with AD also harbor comorbid diseases [9]. Nevertheless, a panoply of explanations has been put forth regarding the potential causes of this disease [10]. These hypotheses have been a driving force for clinical trials, yet disease-modifying treatments remain limited. Given the observation of widespread β A aggregation in diseased brains, β A has generally been regarded as the primary pharmacological target for AD [11] while other targets and theories continue to be debated. Here, we review the neuropathology of AD, early and incipient brain changes, and evidence in support of a vascular-immune basis of disease genesis. Furthermore, we summarize elements of the neurovascular and perivascular unit, which are essential to brain health and metabolism, and describe knowledge gaps and emerging directions in the field.

1.1. Neuropathologic Criteria for AD

Alzheimer's disease neuropathologic change (AD-NC) encompasses extracellular β A deposits and intraneuronal tau protein inclusions. Spatiotemporal propagation of these lesions generally follows stereotyped patterns in the brain, as summarized in National Institute on Aging–Alzheimer's Association (NIA-AA) guidelines [12,13].

β A derives from the amyloid precursor protein (APP) [14] and forms intracerebral senile plaques (SP) that are morphologically heterogeneous and graded according to the Thal scheme [13]. β A is loosely arranged in diffuse plaques (DP), diffuse amyloid lakes and subpial bands. When harboring thick neurites, SPs are called neuritic plaques (NP) [15]; these are separately graded by CERAD scheme [13]. Another NP subtype, the dense-core plaque, displays a central compact β A concretion that is surrounded by a loose zone of diffuse β A deposit [15]. In EOAD, β A plaques are also often arranged in so-called cotton wool plaques that are large, wispy, spherical structures that lack central β A concretions. Per current neuropathologic scoring schemes, neocortical NP densities are semiquantitatively graded [13]. The initial stage of β A deposition involves neocortical brain regions (stage I), but with disease progression, there is involvement of allocortical brain (stage II), subcortical nuclei (stage III), medulla oblongata and midbrain (stage IV) and, eventually, pons and cerebellum (stage V) [13].

In contrast to β A, tau derives from the MAPT gene (chromosome 17). Tau protein is normally found throughout the nervous system and is primarily expressed by neurons. Though its role is not fully elucidated, it is thought to have a role, in part, in neuronal microtubule stabilization. When abnormally hyperphosphorylated, tau is prone to aggregate as intraneuronal paired helical filament inclusions. Within neuronal soma, hyperphosphorylated tau protein forms neurofibrillary tangles (NFT), whereas in distal neuronal processes (i.e., axons and dendrites) it forms neurites and neuropil threads (NT). NFTs primarily localize to limbic regions during early or mild course of disease, but with disease progression the NFTs also aggregate in association cortices and subcortical gray matter nuclei, in a pattern that is distinct from that of β A. A consensus guideline suggests the Braak topographical scheme for NFT staging: stage 0 corresponds to an absence of NFT; stage I/II corresponds to NFT accumulation in transentorhinal regions; stage III/IV corresponds to NFT accumulation within limbic regions; stage V/VI corresponds to wider dispersal of NFT throughout neocortical brain regions [13].

1.2. Theories on AD Pathogenesis

While the causes of AD-NC are unproven, age is an important risk factor and a number of theories have been posited to explain the gradual age-associated accumulation of β A, NFT and NT inclusions. The **cholinergic hypothesis** proposes that AD symptoms arise due to deficiency of intracerebral acetylcholine [10]. This seminal hypothesis evolved

from knowledge that the acetylcholine neurotransmitter and cholinergic neuron loss in limbic and cerebral cortical regions are fundamental features of disease. The **amyloid (or amyloid cascade) hypothesis** theorizes that aberrant β A peptides produced by sequential cleavage of APP by APP-cleavage enzymes (i.e., BACE1 and β/γ -secretases) aggregate into oligomers and insoluble extracellular plaques that damage brain neurons and lead to a range of detrimental secondary phenomena [10,16,17]. The **tau propagation hypothesis** proposes that development and deposition of aberrant hyperphosphorylated tau protein is the initiating pathological event in AD [18], whereas the **inflammation hypothesis** emphasizes the importance of activated microglia and astrocytes in disease genesis [10,19]. Meanwhile, the **oxidative stress hypothesis** asserts that a variety of conditions that cause free radical generation lead to peroxidation of membrane polyunsaturated fatty acids, thus precipitating AD lesions [10]. Similarly, the **mitochondrial hypothesis** proposes that mitochondrial dysfunction occurs upstream of the above phenomena, influencing expression and processing of APP and thereby causing aberrant β A accumulation and an associated cascade of secondary brain changes [10]. The **infectious hypothesis** proposes that a pathogen, i.e., virus, bacterium or prion, underpins disease while the **calcium homeostasis hypothesis** and **metal ion hypothesis** conjecture that calcium or metal ion dysregulation, respectively, are the initial culprit [10]. The **ion channel** [10], **cell cycle** [20], **autoimmune** [21], **epigenetic** [22] and **granuloma hypotheses** [23], among others, have also been propositioned [19]. Altogether, the diversity and number of diverging theories for AD highlight the complexity of this neurological condition and the magnitude of uncertainty regarding initial pathogenetic events.

1.3. Therapeutics and Evidence in Support of Current Theories

Over the course of a century, only seven medications have been formally approved by the Food and Drug Administration (FDA) for AD treatment [11]. These include donepezil (Aricept[®]), galantamine (Razadyne[®]), rivastigmine (Exelon[®]), and tacrine (Cognex[®]), which are acetylcholinesterase inhibitors (the first drug in this class was first approved in 1996) [14]; memantine (Namenda[®]), a N-methyl-D-aspartate receptor (NMDA) receptor antagonist that was approved by the FDA in 2013; aducanumab (Aduhelm[®]), an anti- β A monoclonal IgG1 antibody that was approved in 2021; lecanemab (Leqembi[™]), a monoclonal anti- β A IgG1 antibody that selectively targets soluble aggregated β A species, that was just approved this month [24]. Both acetylcholinesterase inhibitors and NMDA receptor antagonists are regarded as supportive therapies, only, whereas anti- β A agents have overall shown mixed results with the overwhelming majority of drugs in this class failing to show cognitive improvement in humans [25].

Given that genetic, pathological, and functional evidence in humans points toward imbalanced β A production and removal as potential causes for early β A aggregation in various brain regions, β A peptides have generally been considered the primary pharmacological target for AD [10]. Consequently, the amyloid hypothesis has been the prevailing theory and has driven AD drug development strategies for over a quarter century. Initially, β A species in the form of soluble oligomers, intraneuronal aggregates, and insoluble plaques were suggested to be neurotoxic in AD subjects. Data show that β A deposits appear decades prior to dementia symptoms in persons with EOAD and LOAD [25,26]. Yet, clinical trials involving immune-mediated β A removal have been met with limited success [25,27]. Aducanumab, which targets the aggregated plaque form of β A, was approved for use in persons with mild AD symptoms and showed reduction of β A plaque burden but did not translate to significant cognitive improvement. This, too, was observed in only one large-scale trial that subjected patients to prolonged duration of therapy [28]. Meanwhile, high rates of amyloid-related imaging abnormalities (ARIA; up to 43% rate of edema or effusions (ARIA-E) among ApoE ϵ 4 carriers and a 20% rate of microhemorrhages (ARIA-H) in the EMERGE high-dose group) were noted, whereas chronic impairment was documented in 1–2% of patients and therapy-related death was suspected in at least one subject [29,30]. Due to concerns regarding its risk–benefit profile, aducanumab was ulti-

mately discontinued. Likewise, hundreds of prior candidate drugs targeting β A, including BACE1 and β/γ -secretase inhibitors have been ineffective and associated with adverse effects [16,31]. While the trial failures may be attributable to various factors, the failure of innumerable anti- β A drugs has raised questions and doubts regarding presumed roles of β A in AD [17].

More recently, lecanemab has shown efficacy in slowing cognitive decline in early-stage AD subjects enrolled in a large, phase III clinical trial [32,33]. While lecanemab elicited brain swelling and effusions (ARIA-E) in a subset of persons, this drug was approved under FDA's accelerated approval pathway, based on phase II trials that incorporated 18F florbetapir positron emission tomography (PET) imaging of β A as a surrogate endpoint. Additional clinical treatment effects of lecanemab are yet to be determined and a number of other anti- β A drugs remain under investigation [34].

2. Discordance between Neuropathologic Grading Schemes and Theories of AD

Despite the requirement of β A plaques and NFTs for diagnosis of AD-NC and the use of β A as a surrogate endpoint of disease, these markers are of limited utility in disease grading. For instance, precise form(s) and toxic β A species are ambiguous and associations of DP density with cognitive impairment are weak. On the other hand, the abundance and distribution of NFTs and NPs are thought to be predictors for cognitive impairment [13,35]. Yet, NFTs and NPs are also commonly observed in individuals who are cognitively intact [35]. An investigation into risk factors associated with rapid clinical progression, from MCI to dementia, highlights the significance of cerebrospinal fluid (CSF) total tau and phosphorylated tau, but not β A¹⁻⁴² levels, as pertinent to disease [36]. Moreover, abnormal tau protein inclusions also arise in association with other neurodegenerative conditions (i.e., tauopathies), including Pick disease, progressive supranuclear palsy, corticobasal degeneration, chronic traumatic encephalopathy, aging-related tau astroglialopathy, primary age-related tauopathy, and some forms of frontotemporal dementia [1]. Thus, NFTs and NPs are nonspecific lesions that do not accurately predict elderly persons with AD dementia [37,38]. Meanwhile, a number of alternate brain changes and protein inclusions not encompassed by AD-NC criteria are commonly associated with AD. While AD-NC, vascular diseases, and brain vascular injuries are recognized as the most common neuropathologic lesions associated with AD dementia, these entities also often overlap [39]. For a variety of reasons, it has increasingly been thought that β A is associated with, but not causative of AD. It is also increasingly accepted that significant subsets of individuals with AD dementia exhibit heterogeneous tissue substrates and unique combinations of tissue lesions [40]. As chronic systemic and neuropathologic diseases propagate in the oldest old (i.e., persons > 90 years of age), the relationship between AD dementia and established AD-NC criteria diminishes further relative to younger persons, even after correcting for mixed pathologies, suggesting that heterogeneous unknown biological factors apart from β A, NFTs, and other recognized neuropathologies are determinants of the disease [41–43].

Shifting Perspectives: Inconsistencies of the Amyloid Cascade Hypothesis

Despite intensive investigation, direct mechanistic links between β A and AD dementia remain unproven and β A has never been universally accepted as the underlying cause for AD. Over the past decade, a number of studies have suggested that β A toxicity is dependent on the presence of phosphorylated tau inclusions [18]. Moreover, the ApoE genotype has been recognized to modify the risk of AD and cognitive decline through both β A-dependent and β A-independent mechanisms [7]. Increasing evidence highlighting discrepancies between AD-NC and established diagnostic criteria for AD dementia has led to reappraisals of the amyloid hypothesis. A revised amyloid hypothesis suggests that lower-order soluble β A polymers that are imperceptible on histology are the true neurotoxic species in AD [17]. Some researchers maintain that β A and NFTs co-occur, but suggest that disease progression is mediated by tau pathology [18]. Several investigators now believe that tau pathology is an integral substrate of LOAD and propose that tau toxicity is in some

way triggered by β A or β A-related biology. Recent data also uncovers marked variability of tau strains and conformations, and suggests that selective tau recognition by chaperones may differentially influence the accumulation and effects of proteotoxic tau species [38]. Though tau pathology is largely characterized according to NFT burden, research also highlights a diversity of post-translational tau modifications prior to NFT formation [44]. As with β A, soluble tau forms are increasingly thought to represent the true neurotoxic species. Some recent data also suggest that AD pathogenesis occurs due to the effects of long-term elevation of β A concentration, rather than by transient overproduction or impaired clearance [45]. Other studies stress aberrant protein aggregation properties [46]. Still, epidemiological evidence implies that other phenomena, lesions and/or brain “hits” are contributory [41,47,48] and highlight perplexing questions about AD: why are some mutation carriers from AD kindred resistant to β A plaques and NFTs [48]? Why does cerebral amyloid angiopathy (CAA) variably co-occur with AD-NC? Why are non- β A and non-tau protein inclusions observed in heterogeneous combination with disease [40]? Why are vascular pathologies so frequent in association with disease [41–43]? Why do SuperAgers exhibit AD-NC, but no clinical evidence of disease [1,40]? In light of recent literature, these inconsistencies emphasize the need for distinctions between diagnostic biomarkers and molecular targets for AD therapies [14].

3. New Theories: Convergence on Vascular and Neuroimmune Homeostatic Factors

Over the past two decades, investigation into alternative and early-stage AD biomarkers has underscored potential new AD targets [17,49]. Given the convergence of a number of early biological phenomena around cerebral vessels, it is now clear that cerebrovascular contributions to AD have been underrecognized. Increasing data show that inflammatory changes and loss of vascular wall integrity are the earliest identifiable lesions in persons with AD, preceding β A and tau accumulation [50,51]. Interestingly, β A has also been reported to have cytokine-like immune functions, to act as an early responder to diverse stress stimuli, and to demonstrate antimicrobial properties that have not been fully explored [52–56].

In parallel, improvements in microscopy and imaging technologies have led to revised concepts in neuroanatomy, neuroimmunology, and brain barrier biology. The meningeal lymphatic system has emerged as a critical player in neurophysiology [57–59]. This new literature challenges long-held assumptions regarding the brain’s “immune privilege”, as it reveals dynamic functional cross-talk between the brain and peripheral cells and tissues through various routes and a myriad of cellular and molecular mechanisms [60,61]. Moreover, studies into intracerebral fluid physiology in mammalian models have demonstrated that a brain-wide fluid system, generally referred to by some as the glymphatic system, transports solutes and waste metabolites through the cerebrum by way of interstitial and perivascular spaces (PVS) [62–64]. When the glymphatic and lymphatic systems are dysfunctional, CSF tracer and solute transport is impeded [62,65]. Furthermore, this brain drainage system links the health of cerebral vasculature with brain inflammation, stressing critical inter-relationships between brain blood vessels and neuroimmunity. Given these recent neuroscientific advances and knowledge on lymphatic diseases in peripheral tissues, a vascular–immune basis for AD is conceivable and evidence in support of specific pathways and mechanisms of brain fluid and metabolite clearance is now emerging rapidly across species.

3.1. Brain Border Macrophages in AD

Pivotal roles of macrophages at brain surfaces and their significance in brain homeostasis are highlighted in recent literature [66–70]. Within the cranium, but outside brain parenchyma, macrophages known as central nervous system (CNS)-associated macrophages (CAMs) or parenchymal border macrophages (PBMs) reside within meninges, PVS, and choroid plexus and are involved in regulating essential exchange between CNS parenchyma and the periphery. These macrophages are thought to support and maintain

brain barrier properties, control drainage of CNS antigens, and aid in clearance of waste metabolites [66,67]. Moreover, macrophage populations at brain borders are shown to be phenotypically diverse [66,68]. Due to their strategical positioning as well as roles in CSF processing and coordination of reciprocal cross-talk with other cell types, CAMs/PBMs act as critical regulators of brain metabolism and gatekeepers of general neuroimmune processing [67,68]. Evidence suggests that they control the environment and tune entry of immune cells from the CSF and blood into the brain parenchyma and, in turn, regulate the exchange of diverse molecules between the bloodstream, periphery, and brain [67]. PVS macrophages are increasingly thought to have underrecognized roles in CAA and AD [68,69]. In a transgenic TgCRND8 AD mouse model, their depletion by clodronate liposome resulted in a prominent increase in PVS βA^{1-42} deposits [70]. Moreover, CAA pathology was mitigated in this model by chitin-mediated PVS macrophage enrichment [70]. Separate works demonstrated that PVS macrophage density is associated with arterial motion, brain extracellular matrix components, CSF flow dynamics, and clearance of proteins [68,69]. Moreover, the number of PVS macrophages and efficiency of phagocyte activity was shown to be highly associated with perivascular βA deposits [69]. Collectively, these studies suggest that activity and manipulation of PVS macrophages could be integral in the regulation of CAA and AD.

3.2. Peripheral Monocytes and Macrophage Infiltration in AD

While microglia are the principal innate immune cell of the brain and originate from yolk sac erythromyeloid progenitor cells from which they migrate, propagate, and spread during embryogenesis, it has been shown that peripheral monocytes are continuously replenished in adult brains [71,72]. This includes healthy adult brains, in which the recruited cells engraft and differentiate into parenchymal microglia, i.e., new brain resident cells, or remain a distinct population [73–75]. While this recruitment occurs in limited numbers in healthy subjects, experimental AD subjects exhibit accelerated recruitment of monocytes from the periphery [73–75]. Invading monocytes and macrophages that engraft the brain have been shown to derive from circulating Ly-6C^{hi} monocytes or from bone marrow-derived progenitors such as granulocyte–macrophage progenitors (GMPs), or other hematopoietic stem cell progeny [66,73,74]. In a study that employed a green fluorescence protein (GFP)-bone marrow chimeric APP transgenic mouse model (APP23 mice) to study CNS invasion by hematopoietic cells, it was discovered in nontransgenic control mice that the majority of recruited GFP-positive cells localized to brain borders (i.e., PVS, meninges, choroid plexus, and ventricular ependyma) [75]. In contrast, in βA -depositing APP23 mice that develop CAA, a significant portion of recruited GFP-positive cells localized to brain parenchyma, particularly to neocortical areas that exhibit high βA load [75]. These data illustrate critical differences in peripheral myeloid cell recruitment patterns between AD and non-AD subjects, suggesting a role in disease evolution and implying that their targeting may modulate disease course.

3.3. Microgliosis and Other Vascular-Immune Factors in AD

Reactive microglia, i.e., resident brain phagocytes, invariably accompany AD-NC [76–83]. In fact, activated microglia and βA deposits spatially overlap in cerebral cortices of subjects with MCI [77,82], while baseline brain microglial activation and activated hippocampal microglia are significantly increased in subjects with AD [71]. In response to the accrual of βA plaques, adjacent microglia increase their expression of CD11b, CD68, and complement receptor 3 [82,84]. Moreover, *in vitro* analyses demonstrate the ability of βA to stimulate pro-inflammatory cytokine (including interleukin (IL)-1 β , IL-6, IL-8, tumor necrosis factor (TNF)), chemokine, and neurotoxin production by human microglia, causing neuronal damage [85–87]. In response to βA , microglia also secrete proteolytic enzymes and express receptors to promote βA phagocytosis and clearance. This includes the upregulation of class A scavenger receptors (SR-A), the receptor for advanced glycation end-products (RAGE), CD36, toll-like receptors (TLRs) and b1 integrins [82,88]. Therefore, βA may signal

in combination with a host of immune factors to facilitate the recruitment of peripheral immune cells into brains of persons with AD, which may impact on brain function [75]. Some data also suggest that microglia form a protective barrier around β A plaques, compacting β A fibrils into tightly packed and less toxic forms [71,80].

TREM2 (triggering receptor expressed in myeloid cells 2) has also emerged as a key player in microglial and AD biology [71,89,90]. This protein has been shown to function as a receptor for β A and has affinity for its oligomeric forms [89]. The TREM2 cell surface receptor promotes microglial association with β A plaques and via its interaction with the activating adaptor protein DAP12, it plays critical roles in chemotaxis, survival, and proliferation of myeloid cell populations and phagocytosis of a variety of substrates, including apoptotic neurons, bacteria, low-density lipoprotein (LDL) and other lipoproteins, and β A [71]. Loss of TREM2 function diminishes microglial responses to β A plaques, enabling a more toxic state [90]. Rare TREM2 variants are now known to increase the risk of LOAD by two- to four-fold [78].

Variances in analytes from TNF, complement, and coagulation pathways are also demonstrated in persons with MCI [79]. These data demonstrate that neuroinflammatory sequelae manifest early in the course of AD and suggest that they may represent cardinal features of disease. In line with this, plaque-independent inflammation has been demonstrated in neurons that harbor increased levels of soluble and oligomeric β A [83]. Cross-sectional studies further support the hypothesis that neuroinflammation may influence AD pathology at early stages of disease [79,91,92], while epidemiological evidence from several studies highlight decreased prevalence of AD and delayed disease progression in populations treated with long-term anti-inflammatory therapies [93,94].

Adding to this evidence, a majority of recently identified AD risk genes discovered on genome wide association studies such as ABCA7, BIN1, CD33, CR1, INPP5D, MS4A6A, PLCG2SHIP1, and TREM2 are highly or exclusively expressed by macrophages and microglia [71,76–79]. Additional risk genes such as SORL1, CD2AP, PICALM, PTK2B involve sorting receptors, cell membranes, and lipid metabolism [77,95]. These findings underscore the potential critical roles of myeloid cells, intracranial transport, and transport molecules in AD pathogenesis [80–95].

3.4. *Imaging Evidence of Early Hypometabolism and Vascular and Perivascular Dysfunction*

In support of a vascular-immune theory of AD, clinical imaging evidence demonstrates that subtle, asymptomatic vascular changes and blood flow aberrations precede β A and NFT formation, cerebral cortical atrophy, and cognitive decline [51,96–98]. 18F-FDG-PET imaging studies document decreased glucose uptake in specific brain regions during prodromal stages of AD [99]. As abnormal FDG uptake is an indicator of resting-state brain glucose hypometabolism, this represents a biomarker of brain vascular dysfunction. Longitudinal 18F-FDG-PET studies have shown that reduced hippocampal FDG uptake predicts cognitive decline with high sensitivity and specificity years in advance of clinical AD symptoms [100]. Furthermore, high resolution dynamic contrast-enhanced (DCE) MRI analyses demonstrate blood-brain barrier (BBB) injury in the hippocampi of individuals with MCI, suggesting vascular tunica intima damage as an early independent biomarker of cognitive impairment [50,51]. DCE MRI analyses also suggest that BBB breakdown contributes to ApoE4-associated cognitive decline, independent of AD-NC [98].

Enlarged PVS have also emerged as an early imaging biomarker of dementia and AD. While the physiology of these fluid channels remains an area of intensive investigation, it is clear that PVS play underrecognized roles in interstitial fluid and metabolite homeostasis [68,69,101]. PVS surround intracerebral and meningeal vessels and, as described above, provide a unique niche for CAMs/PBMs which are intricately involved in brain health and PVS function [68,69,102]. PVS structures enlarge with age, BBB permeability, hypertension, CAA and other small vessel diseases [101–103], though the causes and effects of this phenomenon are unclear. Various PVS metrics, including number, diameter, and

volume, may be quantified in vivo on structural MRI, though further investigations are needed to optimize and harmonize MRI quantitation techniques [103–105].

3.5. Cardiac and Cardiovascular Disease Effects in AD

Clinicopathologic investigations document a variety of cardiac and extracranial and intracranial vascular changes and pathologies in persons with AD [1,106–114]. Cardiac and extracranial cardiovascular diseases associated with AD include structural or functional heart diseases such as arrhythmic disorders and coronary artery, valvular, and hypertensive diseases [1,112]. Associated extracranial carotid artery changes include stenosis, occlusion, and emboli, whereas intracranial cerebral large vessel pathologies include circle of Willis atherosclerosis, vasculitis and vasculopathies [1,112]. Intracerebral small vessel diseases encompass a multitude of processes, most notably CAA and arteriolosclerosis [1,112].

Microscopic and macroscopic infarcts, i.e., secondary effects of cardiovascular diseases, are predictive of brain neuron and axon density as well as AD dementia risk [1,106]. Longitudinal community-based studies confirm that macroscopic and microscopic infarcts are independent predictors for cognitive decline, though pathophysiological mechanisms are not fully defined [104,107,108]. Other cardiovascular risk factors such as atrial fibrillation and congestive heart failure are also linked to frequency of AD-NC [105,106,108].

CAA is characterized by the deposition of βA within the tunica media and tunica adventitia of cerebral arteries, arterioles, capillaries, and rarely venules. CAA preferentially involves deposition of βA^{1-40} species and may be associated with microhemorrhages or overt, lobar hemorrhages [1,109]. Clinicopathological studies demonstrate significant associations between moderate-to-severe CAA with age and ischemic pathologies [115]. Community-based studies notably document an association between CAA and cognitive impairment, even in the absence of AD-NC [115]. Data show that CAA, arteriolosclerosis, and atherosclerosis severity independently predict increased odds of AD, even after adjusting for NPs, NFTs, and other common age-related neuropathologies [41,114,116].

Interestingly, recent studies also document associations between watershed brain pathology and hypoperfusion injury, and NFTs [106,117]. Mechanisms of brain cell damage and loss associated with vascular diseases are diverse, and may include apoptotic, oncotic, and mixed cell death pathways; however, pathogenetic mechanisms associated with vascular diseases are not fully elucidated. In both large vessel diseases and small vessel diseases, endothelial dysfunction, oxidative stress, and inflammation may contribute to vascular-mediated injuries. A study that employed a time-varying effect model showed that while some vascular diseases (i.e., atherosclerosis and CAA) are associated with a lower level of cognition, their detrimental effects are relatively stable over time, whereas others (e.g., macroscopic infarcts) are associated with progressive deleterious effect on cognition [42]. On the other hand, microinfarcts and arteriolosclerosis were not associated with temporal cognitive changes. These data indicate that age-related vascular pathologies are differentially related to cognitive trajectories over the lifespan [42]. Due to several complexities, no formal criteria exist currently for pathological staging of vascular diseases in AD.

4. Potential Roles of the Glymphatic–Lymphatic System in AD

Since the re-discovery of pseudolymphatic [62,117] and lymphatic [118,119] channels in mice, glymphatic and lymphatic transport mechanisms have been increasingly characterized as highly organized fluid transport systems, though investigations in humans are limited [63,101]. Literature on glymphatic system demonstrates that perivascular channels lined by aquaporin 4, i.e., water channels on the end-feet of perivascular astrocytes, provide a major driving force for fluid flow across the brain while the meningeal lymphatic system effluxes intracranial fluid to cervical lymph nodes [58,59], thus promoting elimination of soluble proteins and metabolites from brain parenchyma [63]. Although the concept of lymphatic-mediated βA and metabolite clearance in humans is not novel [117,118], specific anatomical routes and mechanisms for fluid flow in human brain have been more clearly

defined in recent years [60,64,69,120–124]. While aspects of intracerebral fluid dynamics remain controversial, knowledge on intraparenchymal and peri-parenchymal fluid pathways revises concepts of brain physiology, in that interstitial fluid (ISF) and CSF are increasingly accepted to function as the brain's lymphatic fluid [62,120–122]. The glymphatic and lymphatic systems, originally characterized as separate parallel systems involved in intracerebral [62] and extracerebral [58,59] fluid transport, respectively, are now understood to be anatomically coupled [60] and functionally linked [61]. Furthermore, extracellular β A and tau clearance are significantly hampered in the rodent brain in the absence of a functionally draining dural lymphatic system [57]. Altogether, these data suggest the need for re-examination of mechanisms of macromolecule and metabolite clearance, brain homeostasis, and intracranial fluid regulation in humans. Although the role of β A in the context of AD pathogenesis is unclear, perivascular β A deposition patterns notably recapitulate intracerebral fluid inflow and outflow paths at brain borders, suggesting that intracerebral β A accumulation may occur as a result of flow or clearance failure or, alternatively, may cause glymphatic–lymphatic impedance [69]. Thus, more detailed histological, physiological, and epidemiological investigations of β A risks and deposition patterns along the brain and cerebrovasculature in the context of CAA and AD are needed.

4.1. Clearance Mechanisms at the Neurovascular-Perivascular Interface

Aside from permitting nutrient transfer into brain, the neurovascular–perivascular unit (NVU/PVU) is a critical brain border and vital interface responsible for the transfer of metabolic wastes, immune cells, immune signals, and other small molecules into and out of brain [102,104]. Within the brain, the NVU is comprised of endothelial cells, pericytes, smooth muscle cells, reticular cells, macrophages, and astrocytes, among other cell types, whereas PVUs are situated along the perimeter of intracerebral blood vasculature (Figure 1). Though cellular and molecular mechanisms involving the NVU/PVU remain under investigation, knowledge on this brain barrier is evolving swiftly and available data suggest that anatomic, physiologic and pathologic changes at this interface are essential to brain homeostasis [69,104]. Brain metabolites, including β A, are cleared from the brain by ISF, CSF and blood [125]; through phagocytosis by pericytes, vascular smooth muscle cells, macrophages, and glial cells; through transcytotic processes across the BBB; and through receptor-mediated endocytosis involving various cell types [14,126,127] (Figure 2). Detection of tau and other brain proteins within blood implies that they are also cleared from the brain to the periphery through the NVU/PVU [125,128]. In combination with preclinical evidence on glymphatic–lymphatic functions [58,59,62], knowledge on NVU/PVU biology highlights the potential significance of cerebral vascular and perivascular tissue change and pathology in AD, as NVU/PVU alterations are likely associated with impaired brain metabolite clearance and hampered brain–brain and brain–periphery signaling, in addition to inefficient brain nutrient delivery [129–131]. Thus, NVU/PVU dysfunction may have regional and/or global brain effects, and may lead to generalized protein clearance failure and dysregulated brain–body communication.

4.2. Compound Proteinopathies in AD

As highlighted in recent literature, significant subsets of AD dementia cases are attributable to TDP-43 inclusions and/or α -synuclein-positive Lewy bodies [1,132,133]. Studies show that non- β A and non-tau proteinopathies are not mutually exclusive of one another, nor with AD-NC [132,133]. In fact, cohort studies demonstrate that TDP-43 pathology combined with AD-NC accounts for 35–37% of pathology in elderly subjects with AD dementia [134,135], while Lewy body inclusions overall manifest in more than 50% [135,136]. Thus, common mechanisms may exist for proteinopathic diseases [1,137]. Increasing evidence supports the hypotheses that generalized protein misfolding phenomena in combination with impaired waste clearance efficiency and genetic predispositions may place persons at higher risk for dementing diseases (Figure 3) [44,54,137]. The stereotyped accumulation of protein inclusions from ventral to dorsal brain regions may further impli-

cate perivascular glymphatic–lymphatic transport failure as a common driver of aberrant protein aggregate deposition or cognitive impairment [137].

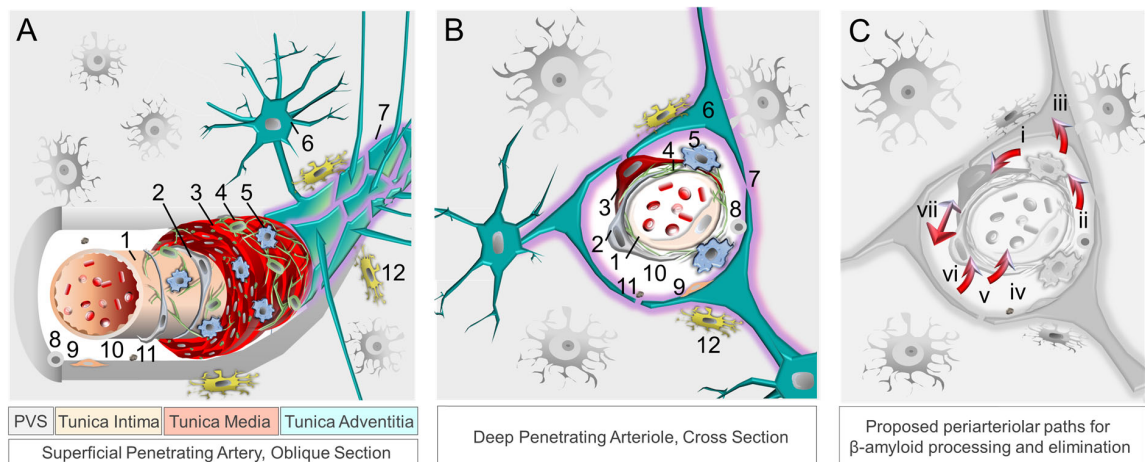


Figure 1. The neurovascular–perivascular unit and recognized β A and brain metabolite clearance mechanisms. Neurovascular and perivascular elements facilitate nutrient exchange as well as metabolism and removal of metabolic wastes, including β A. The neurovascular unit, depicted in oblique section (A) and cross section (B), is composed of (1) endothelial cells, (2) pericytes, (3) smooth muscle cells, (4) reticular cells, (5) macrophages, and (6) astrocytes comprised of (7) aquaporin-4 expressing astrocytic end-feet. The perivascular space harbors (8) other immune cells, (9) fibroblasts, (10) apolipoprotein E, (11) and extracellular matrix components, while (12) microglia are present in adjacent brain tissue. Metabolites, including β A and tau, are ultimately cleared from the brain by interstitial fluid flow in combination with other mechanisms (C), including phagocytosis [by (i) vascular smooth muscle cells, (ii) macrophages, (iii) glial cells, and (vi) pericytes]; (iv) apolipoprotein E-mediated processes; (v) transcytosis across the blood–brain barrier; receptor-mediated endocytosis (involving various immune and other cell types); (vii) perivascular glymphatic mechanisms [62,63,121]. Abbreviation: β , beta; PVS, perivascular space.

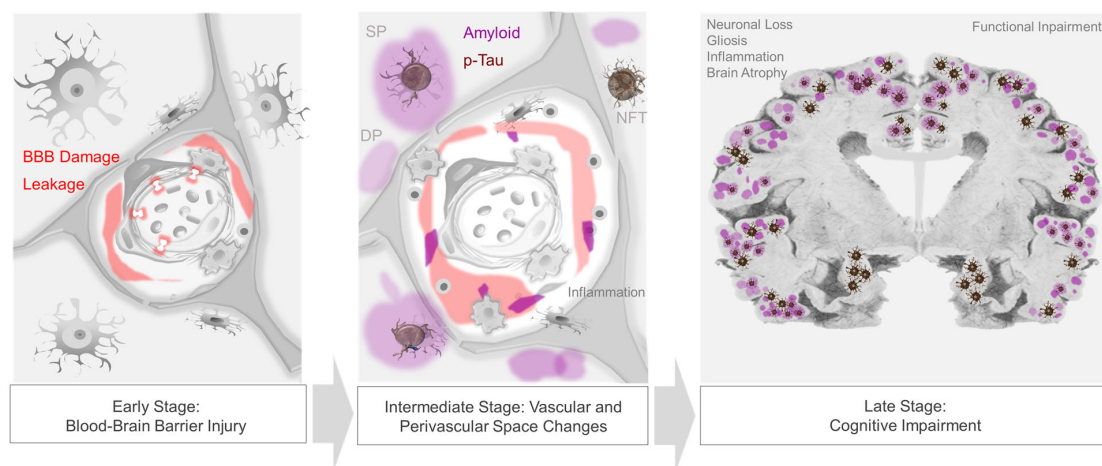


Figure 2. Progression of neurovascular–perivascular and parenchymal brain changes in AD. Blood–brain barrier (BBB) damage and infiltration of serum and blood cellular elements occur in prodromal, i.e., early-stage AD (left panel). Vascular and/or inflammatory insults may further induce transudates in perivascular spaces, impede interstitial fluid drainage, and cause impaired nutrient exchange, edema, cytokine and metabolic waste accumulation, and reactive inflammation and dysregulation (middle panel). In later stages of disease, extracellular β A plaques and intraneuronal neurofibrillary tangles, as well as other protein inclusions, are frequent and are associated with progressive cognitive impairment (right panel).

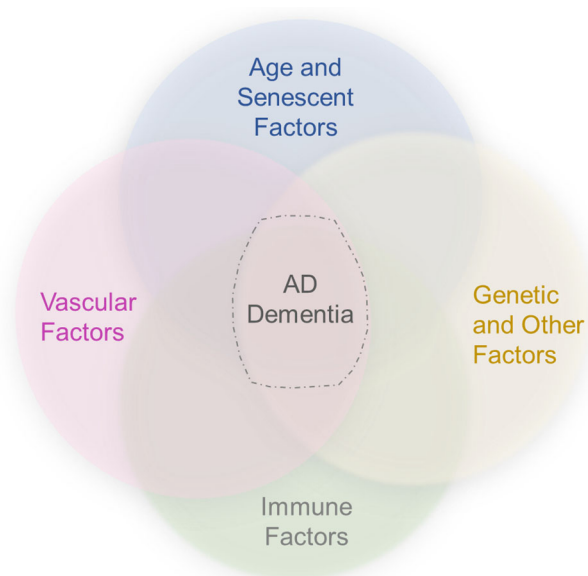


Figure 3. An updated framework suggests the convergence of vascular, immune, and other risk factors in AD. In combination with age, genetic and other factors, vascular and immune factors may converge to precipitate β A plaque and intraneuronal NFT formation, and induce other biological phenomena to cause dementia.

4.3. Blood, Peripheral Signaling and Brain-Body Connections in AD

Intracranial vessels (i.e., lymphatic and blood vascular) and biofluids (i.e., ISF, CSF, and blood) convey innumerable signals, in addition to cytokines and chemokines discussed above. Secretory mechanisms at the BBB impart endocrine-like properties of BBB endothelium [138,139]. Moreover, neurons and neuroglial cells communicate with themselves, with one another, and with the periphery via receptors and extracellular molecules involving autocrine, paracrine, and endocrine signals that transmit in extracellular spaces [138–140]. Other serum-derived factors and biomolecules derived from brain and periphery are also transported through intracranial fluids [141–147]. Recent literature demonstrates that exosomes and ectosomes, small extracellular vesicles (EV) released by exocytosis and plasma membrane shedding, have critical effects on brain homeostasis [142–146]. Exosomes and ectosomes transport a diverse array of cargo including proteins, glycoproteins, lipids, nucleic acids, metabolites, and other biomolecules following their release into the microenvironment [142–146]. As they are capable of inducing phenotypic changes of cells upon fusion and uptake, they are predicted to participate in AD pathogenesis through various mechanisms and pathways [148].

Recent studies also indicate that microbiomes influence brain structure and function across aging [147]. Microbiota (i.e., bacteria, archae, viruses, protists and fungi) and host immune systems co-evolve across the lifespan and homeostasis of the brain–gut axis is reliant upon their efficient coupling and communication [147,148]. The host microbiome influences brain function and is increasingly recognized to modulate both peripheral and central immune responses, as pathogen-associated molecular patterns (PAMPs)/microbe-associated molecular patterns (MAMPs) trigger endogenous molecules such as damage-associated molecular patterns (DAMPs) that are recognized by tissue-resident immune cells via host pattern recognition receptors (PRRs), and cause cell activation while eliciting innate immune responses through a variety of mechanisms [147,148]. This includes increased production of cytokines, chemokines, neuropeptides, metabolic and other signals, as well as complement activation [147,148]. It has been shown that microbiota influence BBB permeability and infiltration of peripheral immune cells, including cytotoxic T-cells into brains of AD subjects, where they are associated with synaptic dysfunction and cognitive changes [147–150]. Imbalances of microbiome homeostasis impact peripheral immune cell trafficking and cause

microglial activation within the brain, impacting brain function [147–150]. Likewise, brain–heart, brain–lung, and other brain–body connections are additionally influenced through bloodborne and immune factors that signal within the brain and at brain borders [150–154], and are subject to effects of the fluid and neuroimmune dysregulation [155].

5. Conclusions

Despite considerable advances in knowledge regarding brain structure and physiology and the dire need for new AD therapies, appropriate AD drug development strategies remain undefined. While aging, genetic and biochemical factors influence the formation of intracerebral β A and tau aggregates that constitute core AD-NC criteria, the bases for cognitive decline in persons with AD dementia are variable and complex. Together, the cerebral vasculature and brain lymphatic system supply the brain with oxygen and nutrients while removing waste metabolites and permitting transvascular and perivascular molecular exchange, signaling, and processing that are critical for brain health and homeostasis. These roles of the cerebral vascular–immune complex are fundamental to sensing and controlling the brain’s microenvironment.

A vascular–immune basis of disease, incorporating dysfunctional brain clearance mechanisms and signaling pathways, centered upon the NVU and PVU, may in part account for inconsistencies in AD pathology and grading (Figure 3). However, basic knowledge on the brain vascular and lymphatic structure and function in humans and their associations with AD remains incomplete. Further clinical investigation into biomarkers that serve as indicators of abnormal intracerebral fluid flow, immune dysfunction, and general protein transport efficiency in aging may yield a better understanding of appropriate therapeutic targets and etiologies involved in cognitive decline in persons afflicted with AD. While prior theories insufficiently explain AD pathobiology, recognition of a brain-wide fluid signaling and lymphatic drainage system raises novel concepts. Disorder of the brain’s vascular–immune system may encompass several previously proposed hypotheses of AD. Future priorities and challenges in understanding this system will include further defining healthy and diseased brain vasculature characteristics across the arteriovenous axis, age spectrum, and brain regions in humans and elucidating divergent effects of immune-related changes and vascular pathologies across comorbidities and stages of disease progression.

Author Contributions: Conceptualization, draft writing and editing, schematic rendering, finalization and approval of the manuscript, R.I.M. (Rashi I. Mehta) and R.I.M. (Rupal I. Mehta). All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by an Alzheimer’s Association Grant (AARGD-22-973935), RADC Development Award, and Grant R21AG079221 from the National Institutes of Health (National Institute on Aging) to Rupal I. Mehta. The funding sources had no role in the preparation, review, or approval of the manuscript; or decision to submit the manuscript for publication.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Mehta, R.I.; Schneider, J.A. Neuropathology of the Common Forms of Dementia. *Clin. Geriatr. Med.* **2023**, *39*, 91–107. [[CrossRef](#)] [[PubMed](#)]
2. Alzheimer’s Association. 2020 Alzheimer’s disease facts and figures. *Alzheimers Dement.* **2020**, *16*, 391–460. [[CrossRef](#)] [[PubMed](#)]
3. Bekris, L.M.; Yu, C.E.; Bird, T.D.; Tsuang, D.W. Genetics of Alzheimer disease. *J. Geriatr. Psychiatry Neurol.* **2010**, *23*, 213–227. [[CrossRef](#)] [[PubMed](#)]
4. Schupf, N.; Sergievsky, G.H. Genetic and host factors for dementia in Down’s syndrome. *Br. J. Psychiatry* **2002**, *180*, 405–410. [[CrossRef](#)]

5. Wiseman, F.K.; Pulford, L.J.; Barkus, C.; Liao, F.; Portelius, E.; Webb, R.; Chávez-Gutiérrez, L.; Cleverley, K.; Noy, S.; Sheppard, O.; et al. Trisomy of human chromosome 21 enhances amyloid- β deposition independently of an extra copy of APP. *Brain* **2018**, *141*, 2457–2474. [[CrossRef](#)]
6. Soto-Ospina, A.; Araque Marín, P.; Bedoya, G.J.; Villegas Lanau, A. Structural Predictive Model of Presenilin-2 Protein and Analysis of Structural Effects of Familial Alzheimer's Disease Mutations. *Biochem. Res. Int.* **2021**, *2021*, 9542038. [[CrossRef](#)]
7. Liu, C.C.; Kanekiyo, T.; Xu, H.; Bu, G. Apolipoprotein E and Alzheimer disease: Risk, mechanisms and therapy. *Nat. Rev. Neurol.* **2013**, *9*, 106–118. [[CrossRef](#)]
8. Dickson, D.W. The pathogenesis of senile plaques. *J. Neuropathol. Exp. Neurol.* **1997**, *56*, 321–339. [[CrossRef](#)]
9. Surguchov, A. Caveolin: A New Link Between Diabetes and AD. *Cell. Mol. Neurobiol.* **2020**, *40*, 1059–1066. [[CrossRef](#)]
10. Liu, P.P.; Xie, Y.; Meng, X.Y.; Kang, J.S. History and progress of hypotheses and clinical trials for Alzheimer's disease. *Signal Transduct. Target Ther.* **2019**, *4*, 29. [[CrossRef](#)]
11. Cummings, J.; Lee, G.; Zhong, K.; Fonseca, J.; Taghva, K. Alzheimer's disease drug development pipeline: 2021. *Alzheimers Dement.* **2021**, *7*, e12179. [[CrossRef](#)]
12. Hyman, B.T.; Trojanowski, J.Q. Consensus recommendations for the postmortem diagnosis of Alzheimer disease from the National Institute on Aging and the Reagan Institute Working Group on diagnostic criteria for the neuropathological assessment of Alzheimer disease. *J. Neuropathol. Exp. Neurol.* **1997**, *56*, 1095–1097. [[CrossRef](#)]
13. Hyman, B.T.; Phelps, C.H.; Beach, T.G.; Bigio, E.H.; Cairns, N.J.; Carrillo, M.C.; Dickson, D.W.; Duyckaerts, C.; Frosch, M.P.; Masliah, E.; et al. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement.* **2012**, *8*, 1–13. [[CrossRef](#)]
14. Kumar, A.; Nemeroff, C.B.; Cooper, J.J.; Widge, A.; Rodriguez, C.; Carpenter, L.; McDonald, W.M. Amyloid and Tau in Alzheimer's Disease: Biomarkers or Molecular Targets for Therapy? Are We Shooting the Messenger? *Am. J. Psychiatry* **2021**, *178*, 1014–1025. [[CrossRef](#)] [[PubMed](#)]
15. Dickson, T.C.; Vickers, J.C. The morphological phenotype of beta-amyloid plaques and associated neuritic changes in Alzheimer's disease. *Neuroscience* **2001**, *105*, 99–107. [[CrossRef](#)]
16. Das, B.; Yan, R. A Close Look at BACE1 Inhibitors for Alzheimer's Disease Treatment. *CNS Drugs* **2019**, *33*, 251–263. [[CrossRef](#)]
17. Selkoe, D.J.; Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol. Med.* **2016**, *8*, 595–608. [[CrossRef](#)]
18. Arnsten, A.F.T.; Datta, D.; Del Tredici, K.; Braak, H. Hypothesis: Tau pathology is an initiating factor in sporadic Alzheimer's disease. *Alzheimers Dement.* **2021**, *17*, 115–124. [[CrossRef](#)]
19. Qiu, C.; Kivipelto, M.; von Strauss, E. Epidemiology of Alzheimer's disease: Occurrence, determinants, and strategies toward intervention. *Dialogues Clin. Neurosci.* **2009**, *11*, 111–128. [[CrossRef](#)]
20. Woods, J.; Snape, M.; Smith, M.A. The cell cycle hypothesis of Alzheimer's disease: Suggestions for drug development. *Biochim. Biophys. Acta* **2007**, *1772*, 503–508. [[CrossRef](#)]
21. Arshavsky, Y.I. Alzheimer's Disease: From Amyloid to Autoimmune Hypothesis. *Neuroscientist* **2020**, *26*, 455–470. [[CrossRef](#)]
22. Penner, M.R.; Roth, T.L.; Barnes, C.A.; Sweatt, J.D. An epigenetic hypothesis of aging-related cognitive dysfunction. *Front. Aging Neurosci.* **2010**, *2*, 9. [[CrossRef](#)] [[PubMed](#)]
23. Lemke, G.; Huang, Y. The dense-core plaques of Alzheimer's disease are granulomas. *J. Exp. Med.* **2022**, *219*, e20212477. [[CrossRef](#)] [[PubMed](#)]
24. Reardon, S. FDA approves Alzheimer's drug lecanemab amid safety concerns. *Nature* **2023**, *613*, 227–228. [[CrossRef](#)]
25. Makin, S. The amyloid hypothesis on trial. *Nature* **2018**, *559*, S4–S7. [[CrossRef](#)]
26. Villemagne, V.L.; Burnham, S.; Bourgeat, P.; Brown, B.; Ellis, K.A.; Salvado, O.; Szoëke, C.; Macaulay, S.L.; Martins, R.; Maruff, P.; et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: A prospective cohort study. *Lancet Neurol.* **2013**, *12*, 357–367. [[CrossRef](#)]
27. Panza, F. Are antibodies directed against amyloid- β ($A\beta$) oligomers the last call for the βA hypothesis of Alzheimer's disease? *Immunotherapy* **2019**, *11*, 3–6. [[CrossRef](#)]
28. Rabinovici, G.D. Controversy and Progress in Alzheimer's Disease—FDA Approval of Aducanumab. *N. Engl. J. Med.* **2021**, *385*, 771–774. [[CrossRef](#)]
29. Haerberlein, S.B.; Aisen, P.S.; Barkhof, F.; Chalkias, S.; Chen, T.; Cohen, S.; Dent, G.; Hansson, O.; Harrison, K.; von Hehn, C.; et al. Two Randomized Phase 3 Studies of Aducanumab in Early Alzheimer's Disease. *J. Prev. Alzheimers Dis.* **2022**, *9*, 197–210. [[CrossRef](#)]
30. Woloshin, S.; Kesselheim, A.S. What to Know About the Alzheimer Drug Aducanumab (Aduhelm). *JAMA Intern. Med.* **2022**, *182*, 892. [[CrossRef](#)]
31. Coric, V.; Salloway, S.; van Dyck, C.H.; Dubois, B.; Andreasen, N.; Brody, M.; Curtis, C.; Soininen, H.; Thein, S.; Shiovitz, T.; et al. Targeting Prodromal Alzheimer Disease with Avagacestat: A Randomized Clinical Trial. *JAMA Neurol.* **2015**, *72*, 1324–1333. [[CrossRef](#)] [[PubMed](#)]
32. Van Dyck, C.H.; Swanson, C.J.; Aisen, P.; Bateman, R.J.; Chen, C.; Gee, M.; Kanekiyo, M.; Li, D.; Reyderman, L.; Cohen, S.; et al. Lecanemab in Early Alzheimer's Disease. *N. Engl. J. Med.* **2023**, *388*, 9–21. [[CrossRef](#)] [[PubMed](#)]
33. McDade, E.; Cummings, J.L.; Dhadda, S.; Swanson, C.J.; Reyderman, L.; Kanekiyo, M.; Koyama, A.; Irizarry, M.; Kramer, L.D.; Bateman, R.J. Lecanemab in patients with early Alzheimer's disease: Detailed results on biomarker, cognitive, and clinical effects from the randomized and open-label extension of the phase 2 proof-of-concept study. *Alzheimers Res. Ther.* **2022**, *14*, 191. [[CrossRef](#)] [[PubMed](#)]

34. Cao, Y.; Yu, F.; Lyu, Y.; Lu, X. Promising candidates from drug clinical trials: Implications for clinical treatment of Alzheimer's disease in China. *Front. Neurol.* **2022**, *13*, 1034243. [[CrossRef](#)]
35. Crystal, H.; Dickson, D.; Fuld, P.; Masur, D.; Scott, R.; Mehler, M.; Masdeu, J.; Kawas, C.; Aronson, M.; Wolfson, L. Clinico-pathologic studies in dementia: Nondemented subjects with pathologically confirmed Alzheimer's disease. *Neurology* **1988**, *38*, 1682–1687. [[CrossRef](#)]
36. Van Rossum, I.A.; Visser, P.J.; Knol, D.L.; van der Flier, W.M.; Teunissen, C.E.; Barkhof, F.; Blankenstein, M.A.; Scheltens, P. Injury markers but not amyloid markers are associated with rapid progression from mild cognitive impairment to dementia in Alzheimer's disease. *J. Alzheimers Dis.* **2012**, *29*, 319–327. [[CrossRef](#)]
37. Gao, Y.L.; Wang, N.; Sun, F.R.; Cao, X.P.; Zhang, W.; Yu, J.T. Tau in neurodegenerative disease. *Ann. Transl. Med.* **2018**, *6*, 175. [[CrossRef](#)]
38. Ryder, B.D.; Wydorski, P.M.; Hou, Z.; Joachimiak, L.A. Chaperoning shape-shifting tau in disease. *Trends Biochem. Sci.* **2022**, *47*, 301–313. [[CrossRef](#)]
39. Jack, C.R.; Bennett, D.A.; Blennow, K.; Carrillo, M.C.; Dunn, B.; Haeberlein, S.B.; Holtzman, D.M.; Jagust, W.; Jessen, F.; Karlawish, J.; et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement.* **2018**, *14*, 535–562. [[CrossRef](#)]
40. Kawas, C.H.; Corrada, M.M. Successful cognitive aging: What the oldest-old can teach us about resistance and resilience. *Neurology* **2020**, *95*, 329–330. [[CrossRef](#)]
41. Boyle, P.A.; Wilson, R.S.; Yu, L.; Barr, A.M.; Honer, W.G.; Schneider, J.A.; Bennett, D.A. Much of late life cognitive decline is not due to common neurodegenerative pathologies. *Ann. Neurol.* **2013**, *74*, 478–489. [[CrossRef](#)]
42. Boyle, P.A.; Yang, J.; Yu, L.; Leurgans, S.E.; Capuano, A.W.; Schneider, J.A.; Wilson, R.S.; Bennett, D.A. Varied effects of age-related neuropathologies on the trajectory of late life cognitive decline. *Brain* **2017**, *140*, 804–812. [[CrossRef](#)] [[PubMed](#)]
43. Boyle, P.A.; Yu, L.; Wilson, R.S.; Leurgans, S.E.; Schneider, J.A.; Bennett, D.A. Person-specific contribution of neuropathologies to cognitive loss in old age. *Ann. Neurol.* **2018**, *83*, 74–83. [[CrossRef](#)]
44. Carroll, T.; Guha, S.; Nehrke, K.; Johnson, G.V.W. Tau Post-Translational Modifications: Potentiators of Selective Vulnerability in Sporadic Alzheimer's Disease. *Biology* **2021**, *10*, 1047. [[CrossRef](#)]
45. Zhou, B.; Lu, J.G.; Siddu, A.; Wernig, M.; Südhof, T.C. Synaptogenic effect of APP-Swedish mutation in familial Alzheimer's disease. *Sci. Transl. Med.* **2022**, *14*, eabn9380. [[CrossRef](#)]
46. Condello, C.; Merz, G.E.; Aoyagi, A.; DeGrado, W.F.; Prusiner, S.B. β A and Tau Prions Causing Alzheimer's Disease. *Methods Mol. Biol.* **2023**, *2561*, 293–337.
47. Satizabal, C.L.; Beiser, A.S.; Chouraki, V.; Chêne, G.; Dufouil, C.; Seshadri, S. Incidence of Dementia over Three Decades in the Framingham Heart Study. *N. Engl. J. Med.* **2016**, *374*, 523–532. [[CrossRef](#)]
48. Arboleda-Velasquez, J.F.; Lopera, F.; O'Hare, M.; Delgado-Tirado, S.; Marino, C.; Chmielewska, N.; Saez-Torres, K.L.; Amarnani, D.; Schultz, A.P.; Sperling, R.A.; et al. Resistance to autosomal dominant Alzheimer's disease in an APOE3 Christchurch homozygote: A case report. *Nat. Med.* **2019**, *25*, 1680–1683. [[CrossRef](#)]
49. Haage, V.; De Jager, P.L. Neuroimmune contributions to Alzheimer's disease: A focus on human data. *Mol. Psychiatry* **2022**, *27*, 3164–3181. [[CrossRef](#)]
50. Nation, D.A.; Sweeney, M.D.; Montagne, A.; Sagare, A.P.; D'Orazio, L.M.; Pachicano, M.; Seppehrband, F.; Nelson, A.R.; Buennagel, D.P.; Harrington, M.G.; et al. Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat. Med.* **2019**, *25*, 270–276. [[CrossRef](#)]
51. Montagne, A.; Barnes, S.R.; Sweeney, M.D.; Halliday, M.R.; Sagare, A.P.; Zhao, Z.; Toga, A.W.; Jacobs, R.E.; Liu, C.Y.; Amezcua, L.; et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **2015**, *85*, 296–302. [[CrossRef](#)] [[PubMed](#)]
52. Bisette, G. Does Alzheimer's disease result from attempts at repair or protection after transient stress? *J. Alzheimers Dis.* **2009**, *18*, 371–380. [[CrossRef](#)] [[PubMed](#)]
53. Kagan, B.L.; Jang, H.; Capone, R.; Teran Arce, F.; Ramachandran, S.; Lal, R.; Nussinov, R. Antimicrobial properties of amyloid peptides. *Mol. Pharm.* **2012**, *9*, 708–717. [[CrossRef](#)] [[PubMed](#)]
54. LaRocca, T.J.; Cavalier, A.N.; Roberts, C.M.; Lemieux, M.R.; Ramesh, P.; Garcia, M.A.; Link, C.D. Amyloid beta acts synergistically as a pro-inflammatory cytokine. *Neurobiol. Dis.* **2021**, *159*, 105493. [[CrossRef](#)] [[PubMed](#)]
55. Kizawa, M.; Iwasaki, Y. Amyloid β -related angiitis of the central nervous system occurring after COVID-19 vaccination: A case report. *World J. Clin. Cases* **2022**, *10*, 12617–12622. [[CrossRef](#)]
56. Bratt, B.M.; Kolb, W.P.; Cooper, N.R. Complement-dependent proinflammatory properties of the Alzheimer's disease beta-peptide. *J. Exp. Med.* **1998**, *188*, 431–438. [[CrossRef](#)]
57. Da Mesquita, S.; Fu, Z.; Kipnis, J. The Meningeal Lymphatic System: A New Player in Neurophysiology. *Neuro* **2018**, *100*, 375–388. [[CrossRef](#)]
58. Louveau, A.; Smirnov, I.; Keyes, T.J.; Eccles, J.D.; Rouhani, S.J.; Peske, J.D.; Derecki, N.C.; Castle, D.; Mandell, J.W.; Lee, K.S.; et al. Structural and functional features of central nervous system lymphatic vessels. *Nature* **2015**, *523*, 337–341. [[CrossRef](#)]
59. Aspelund, A.; Antila, S.; Proulx, S.T.; Karlsen, T.V.; Karaman, S.; Detmar, M.; Wiig, H.; Alitalo, K. A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. *J. Exp. Med.* **2015**, *212*, 991–999. [[CrossRef](#)]

60. Shah, T.; Leurgans, S.E.; Mehta, R.I.; Yang, J.; Galloway, C.A.; de Mesy Bentley, K.L.; Schneider, J.A.; Mehta, R.I. Arachnoid granulations are lymphatic conduits that communicate with bone marrow and dura-arachnoid stroma. *J. Exp. Med.* **2023**, *220*, e20220618. [[CrossRef](#)]
61. Rustenhoven, J.; Drieu, A.; Mamuladze, T.; de Lima, K.A.; Dykstra, T.; Wall, M.; Papadopoulos, Z.; Kanamori, M.; Salvador, A.F.; Baker, W. Functional characterization of the dural sinuses as a neuroimmune interface. *Cell* **2021**, *184*, 1000–1016.e27. [[CrossRef](#)]
62. Iliif, J.J.; Wang, M.; Liao, Y.; Plogg, B.A.; Peng, W.; Gundersen, G.A.; Benveniste, H.; Vates, G.E.; Deane, R.; Goldman, S.A.; et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β . *Sci. Transl. Med.* **2012**, *4*, 147ra111. [[CrossRef](#)]
63. Mestre, H.; Kostrikov, S.; Mehta, R.I.; Nedergaard, M. Perivascular spaces, glymphatic dysfunction, and small vessel disease. *Clin. Sci.* **2017**, *131*, 2257–2274. [[CrossRef](#)]
64. Mestre, H.; Du, T.; Sweeney, A.M.; Liu, G.; Samson, A.J.; Peng, W.; Mortensen, K.N.; Stæger, F.F.; Bork, P.A.R.; Bashford, L.; et al. Cerebrospinal fluid influx drives acute ischemic tissue swelling. *Science* **2020**, *367*, eaax7171. [[CrossRef](#)]
65. Da Mesquita, S.; Papadopoulos, Z.; Dykstra, T.; Brase, L.; Farias, F.G.; Wall, M.; Jiang, H.; Kodira, C.D.; de Lima, K.A.; Herz, J.; et al. Meningeal lymphatics affect microglia responses and anti-A β immunotherapy. *Nature* **2021**, *593*, 255–260. [[CrossRef](#)]
66. Prinz, M.; Priller, J.; Sisodia, S.S.; Ransohoff, R.M. Heterogeneity of CNS myeloid cells and their roles in neurodegeneration. *Nat. Neurosci.* **2011**, *14*, 1227–1235. [[CrossRef](#)]
67. Kierdorf, K.; Masuda, T.; Jordão, M.J.C.; Prinz, M. Macrophages at CNS interfaces: Ontogeny and function in health and disease. *Nat. Rev. Neurosci.* **2019**, *20*, 547–562. [[CrossRef](#)]
68. Drieu, A.; Du, S.; Storck, S.E.; Rustenhoven, J.; Papadopoulos, Z.; Dykstra, T.; Zhong, F.; Kim, K.; Blackburn, S.; Mamuladze, T.; et al. Parenchymal border macrophages regulate the flow dynamics of the cerebrospinal fluid. *Nature* **2022**, *611*, 585–593. [[CrossRef](#)]
69. Mestre, H.; Verma, N.; Greene, T.D.; Lin, L.A.; Ladron-de-Guevara, A.; Sweeney, A.M.; Liu, G.; Thomas, V.K.; Galloway, C.A.; de Mesy Bentley, K.L.; et al. Periaxonal spaces modulate cerebrospinal fluid transport into brain and demonstrate altered morphology in aging and Alzheimer’s disease. *Nat. Commun.* **2022**, *13*, 3897. [[CrossRef](#)]
70. Hawkes, C.A.; McLaurin, J. Selective targeting of perivascular macrophages for clearance of beta-amyloid in cerebral amyloid angiopathy. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 1261–1266. [[CrossRef](#)]
71. Hansen, D.V.; Hanson, J.E.; Sheng, M. Microglia in Alzheimer’s disease. *J. Cell Biol.* **2018**, *217*, 459–472. [[CrossRef](#)] [[PubMed](#)]
72. El Khoury, J.; Toft, M.; Hickman, S.E.; Means, T.K.; Terada, K.; Geula, C.; Luster, A.D. Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimer-like disease. *Nat. Med.* **2007**, *13*, 432–438. [[CrossRef](#)] [[PubMed](#)]
73. Malm, T.M.; Koistinaho, M.; Pärepallo, M.; Vatanen, T.; Ooka, A.; Karlsson, S.; Koistinaho, J. Bone-marrow-derived cells contribute to the recruitment of microglial cells in response to beta-amyloid deposition in APP/PS1 double transgenic Alzheimer mice. *Neurobiol. Dis.* **2005**, *18*, 134–142. [[CrossRef](#)] [[PubMed](#)]
74. Simard, A.R.; Soulet, D.; Gowing, G.; Julien, J.P.; Rivest, S. Bone marrow-derived microglia play a critical role in restricting senile plaque formation in Alzheimer’s disease. *Neuron* **2006**, *49*, 489–502. [[CrossRef](#)]
75. Stalder, A.K.; Ermini, F.; Bondolfi, L.; Krenger, W.; Burbach, G.J.; Deller, T.; Coomaraswamy, J.; Staufenbiel, M.; Landmann, R.; Jucker, M. Invasion of hematopoietic cells into the brain of amyloid precursor protein transgenic mice. *J. Neurosci.* **2005**, *25*, 11125–11132. [[CrossRef](#)]
76. Efthymiou, A.G.; Goate, A.M. Late onset Alzheimer’s disease genetics implicates microglial pathways in disease risk. *Mol. Neurodegener.* **2017**, *12*, 43. [[CrossRef](#)]
77. Gao, C.; Shen, X.; Tan, Y.; Chen, S. Pathogenesis, therapeutic strategies and biomarker development based on “omics” analysis related to microglia in Alzheimer’s disease. *J. Neuroinflamm.* **2022**, *19*, 215. [[CrossRef](#)]
78. Ulland, T.K.; Colonna, M. TREM2—A key player in microglial biology and Alzheimer disease. *Nat. Rev. Neurol.* **2018**, *14*, 667–675. [[CrossRef](#)]
79. Pillai, J.A.; Maxwell, S.; Bena, J.; Bekris, L.M.; Rao, S.M.; Chance, M.; Lamb, B.T.; Leverenz, J.B.; Alzheimer’s Disease Neuroimaging Initiative. Key inflammatory pathway activations in the MCI stage of Alzheimer’s disease. *Ann. Clin. Transl. Neurol.* **2019**, *6*, 1248–1262. [[CrossRef](#)]
80. Condello, C.; Yuan, P.; Schain, A.; Grutzendler, J. Microglia constitute a barrier that prevents neurotoxic protofibrillar A β 42 hotspots around plaques. *Nat. Commun.* **2015**, *6*, 6176. [[CrossRef](#)]
81. Fan, Z.; Brooks, D.J.; Okello, A.; Edison, P. An early and late peak in microglial activation in Alzheimer’s disease trajectory. *Brain* **2017**, *140*, 792–803. [[CrossRef](#)]
82. Wyss-Coray, T.; Rogers, J. Inflammation in Alzheimer disease—a brief review of the basic science and clinical literature. *Cold Spring Harb. Perspect. Med.* **2012**, *2*, a006346. [[CrossRef](#)]
83. Welikovitsh, L.A.; Carmo, S.D.; Maglóczy, Z.; Malcolm, J.C.; Löke, J.; Klein, W.L.; Freund, T.; Cuellar, A.C. Early intraneuronal amyloid triggers neuron-derived inflammatory signaling in APP transgenic rats and human brain. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 6844–6854. [[CrossRef](#)]
84. Herber, D.L.; Mercer, M.; Roth, L.M.; Symmonds, K.; Maloney, J.; Wilson, N.; Freeman, M.J.; Morgan, D.; Gordon, M.N. Microglial activation is required for Abeta clearance after intracranial injection of lipopolysaccharide in APP transgenic mice. *J. Neuroimmune Pharmacol.* **2007**, *2*, 222–231. [[CrossRef](#)]

85. McGeer, P.L.; Itagaki, S.; Tago, H.; McGeer, E.G. Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. *Neurosci. Lett.* **1987**, *79*, 195–200. [[CrossRef](#)]
86. McGeer, E.G.; McGeer, P.L. Innate immunity in Alzheimer's disease: A model for local inflammatory reactions. *Mol. Interv.* **2001**, *1*, 22–29.
87. Rogers, J.; Lue, L.F. Microglial chemotaxis, activation, and phagocytosis of amyloid beta-peptide as linked phenomena in Alzheimer's disease. *Neurochem. Int.* **2001**, *39*, 333–340. [[CrossRef](#)]
88. Doens, D.; Fernández, P.L. Microglia receptors and their implications in the response to amyloid β for Alzheimer's disease pathogenesis. *J. Neuroinflamm.* **2014**, *11*, 48. [[CrossRef](#)]
89. Zhao, Y.; Wu, X.; Li, X.; Jiang, L.L.; Gui, X.; Liu, Y.; Sun, Y.; Zhu, B.; Piña-Crespo, J.C.; Zhang, M.; et al. TREM2 Is a Receptor for β -Amyloid that Mediates Microglial Function. *Neuron* **2018**, *97*, 1023–1031.e7. [[CrossRef](#)]
90. Shi, Y.; Holtzman, D.M. Interplay between innate immunity and Alzheimer disease: APOE and TREM2 in the spotlight. *Nat. Rev. Immunol.* **2018**, *18*, 759–772. [[CrossRef](#)]
91. Albrecht, D.S.; Sagare, A.; Pachicano, M.; Sweeney, M.D.; Toga, A.; Zlokovic, B.; Chui, H.; Joe, E.; Schneider, L.; Morris, J.C.; et al. Early neuroinflammation is associated with lower amyloid and tau levels in cognitively normal older adults. *Brain Behav. Immun.* **2021**, *94*, 299–307. [[CrossRef](#)] [[PubMed](#)]
92. Toppala, S.; Ekblad, L.L.; Tuisku, J.; Helin, S.; Johansson, J.J.; Laine, H.; Löyttyniemi, E.; Marjamäki, P.; Blennow, K.; Zetterberg, H.; et al. Association of Early β -Amyloid Accumulation and Neuroinflammation Measured With [¹¹C]PBR28 in Elderly Individuals Without Dementia. *Neurology* **2021**, *96*, e1608–e1619. [[CrossRef](#)] [[PubMed](#)]
93. Zhang, C.; Wang, Y.; Wang, D.; Zhang, J.; Zhang, F. NSAID Exposure and Risk of Alzheimer's Disease: An Updated Meta-Analysis From Cohort Studies. *Front. Aging Neurosci.* **2018**, *10*, 83. [[CrossRef](#)] [[PubMed](#)]
94. In 't Veld, B.A.; Ruitenber, A.; Hofman, A.; Launer, L.J.; van Duijn, C.M.; Stijnen, T.; Breteler, M.M.; Stricker, B.H. Nonsteroidal antiinflammatory drugs and the risk of Alzheimer's disease. *N. Engl. J. Med.* **2001**, *345*, 1515–1521. [[CrossRef](#)]
95. Hung, C.; Tuck, E.; Stubbs, V.; van der Lee, S.J.; Aalfs, C.; van Spaendonk, R.; Scheltens, P.; Hardy, J.; Holstege, H.; Livesey, F.J. SORL1 deficiency in human excitatory neurons causes APP-dependent defects in the endolysosome-autophagy network. *Cell Rep.* **2021**, *35*, 109259. [[CrossRef](#)]
96. Sweeney, M.D.; Kisler, K.; Montagne, A.; Toga, A.W.; Zlokovic, B.V. The role of brain vasculature in neurodegenerative disorders. *Nat. Neurosci.* **2018**, *21*, 1318–1331. [[CrossRef](#)]
97. Sweeney, M.D.; Montagne, A.; Sagare, A.P.; Nation, D.A.; Schneider, L.S.; Chui, H.C.; Harrington, M.G.; Pa, J.; Law, M.; Wang, D.J.J.; et al. Vascular dysfunction—The disregarded partner of Alzheimer's disease. *Alzheimers Dement.* **2019**, *15*, 158–167. [[CrossRef](#)]
98. Montagne, A.; Nation, D.A.; Sagare, A.P.; Barisano, G.; Sweeney, M.D.; Chakhoyan, A.; Pachicano, M.; Joe, E.; Nelson, A.R.; D'Orazio, L.M.; et al. APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature* **2020**, *581*, 71–76. [[CrossRef](#)]
99. Hunt, A.; Schönknecht, P.; Henze, M.; Seidl, U.; Haberkorn, U.; Schröder, J. Reduced cerebral glucose metabolism in patients at risk for Alzheimer's disease. *Psychiatry Res.* **2007**, *155*, 147–154. [[CrossRef](#)]
100. Mosconi, L.; Tsui, W.H.; Herholz, K.; Pupi, A.; Drzezga, A.; Lucignani, G.; Reiman, E.M.; Holthoff, V.; Kalbe, E.; Sorbi, S.; et al. Multicenter standardized 18F-FDG PET diagnosis of mild cognitive impairment, Alzheimer's disease, and other dementias. *J. Nucl. Med.* **2008**, *49*, 390–398. [[CrossRef](#)]
101. Javierre-Petit, C.; Schneider, J.A.; Kapasi, A.; Makinejad, N.; Tamhane, A.A.; Leurgans, S.E.; Mehta, R.I.; Barnes, L.L.; Bennett, D.A.; Arfanakis, K. Neuropathologic and Cognitive Correlates of Enlarged Perivascular Spaces in a Community-Based Cohort of Older Adults. *Stroke* **2020**, *51*, 2825–2833. [[CrossRef](#)]
102. MacLulich, A.M.; Wardlaw, J.M.; Ferguson, K.J.; Starr, J.M.; Seckl, J.R.; Deary, I.J. Enlarged perivascular spaces are associated with cognitive function in healthy elderly men. *J. Neurol. Neurosurg. Psychiatry* **2004**, *75*, 1519–1523. [[CrossRef](#)]
103. Niazi, M.; Karaman, M.; Das, S.; Zhou, X.J.; Yushkevich, P.; Cai, K. Quantitative MRI of Perivascular Spaces at 3T for Early Diagnosis of Mild Cognitive Impairment. *AJNR Am. J. Neuroradiol.* **2018**, *39*, 1622–1628. [[CrossRef](#)]
104. Brown, R.; Benveniste, H.; Black, S.E.; Charpak, S.; Dichgans, M.; Joutel, A.; Nedergaard, M.; Smith, K.J.; Zlokovic, B.V.; Wardlaw, J.M. Understanding the role of the perivascular space in cerebral small vessel disease. *Cardiovasc. Res.* **2018**, *114*, 1462–1473. [[CrossRef](#)]
105. Boyle, P.A.; Yu, L.; Leurgans, S.E.; Wilson, R.S.; Brookmeyer, R.; Schneider, J.A.; Bennett, D.A. Attributable risk of Alzheimer's dementia attributed to age-related neuropathologies. *Ann. Neurol.* **2019**, *85*, 114–124. [[CrossRef](#)]
106. Kapasi, A.; Yu, L.; Petyuk, V.; Arfanakis, K.; Bennett, D.A.; Schneider, J.A. Association of small vessel disease with tau pathology. *Acta Neuropathol.* **2022**, *143*, 349–362. [[CrossRef](#)]
107. Wang, M.; Iliff, J.J.; Liao, Y.; Chen, M.J.; Shinseki, M.S.; Venkataraman, A.; Cheung, J.; Wang, W.; Nedergaard, M. Cognitive deficits and delayed neuronal loss in a mouse model of multiple microinfarcts. *J. Neurosci.* **2012**, *32*, 17948–17960. [[CrossRef](#)]
108. Wang, M.; Ding, F.; Deng, S.; Guo, X.; Wang, W.; Iliff, J.J.; Nedergaard, M. Focal Solute Trapping and Global Glymphatic Pathway Impairment in a Murine Model of Multiple Microinfarcts. *J. Neurosci.* **2017**, *37*, 2870–2877. [[CrossRef](#)]
109. Attems, J.; Jellinger, K.A. The overlap between vascular disease and Alzheimer's disease—lessons from pathology. *BMC Med.* **2014**, *12*, 206. [[CrossRef](#)]

110. Laing, K.K.; Simoes, S.; Baena-Caldas, G.P.; Lao, P.J.; Kothiya, M.; Igwe, K.C.; Chesebro, A.G.; Houck, A.L.; Pedraza, L.; Hernández, A.I.; et al. Cerebrovascular disease promotes tau pathology in Alzheimer's disease Alzheimer's Disease Neuroimaging Initiative. *Brain Commun.* **2020**, *2*, fcaa132. [[CrossRef](#)]
111. Esiri, M.M.; Nagy, Z.; Smith, M.Z.; Barnettson, L.; Smith, A.D. Cerebrovascular disease and threshold for dementia in the early stages of Alzheimer's disease. *Lancet* **1999**, *354*, 919–920. [[CrossRef](#)] [[PubMed](#)]
112. Arvanitakis, Z.; Capuano, A.W.; Leurgans, S.E.; Bennett, D.A.; Schneider, J.A. Relation of cerebral vessel disease to Alzheimer's disease dementia and cognitive function in elderly people: A cross-sectional study. *Lancet Neurol.* **2016**, *15*, 934–943. [[CrossRef](#)] [[PubMed](#)]
113. Arvanitakis, Z.; Capuano, A.W.; Leurgans, S.E.; Buchman, A.S.; Bennett, D.A.; Schneider, J.A. The Relationship of Cerebral Vessel Pathology to Brain Microinfarcts. *Brain Pathol.* **2017**, *27*, 77–85. [[CrossRef](#)] [[PubMed](#)]
114. Kapasi, A.; Schneider, J.A. Vascular contributions to cognitive impairment, clinical Alzheimer's disease, and dementia in older persons. *Biochim. Biophys. Acta* **2016**, *1862*, 878–886. [[CrossRef](#)] [[PubMed](#)]
115. Reijmer, Y.D.; van Veluw, S.J.; Greenberg, S.M. Ischemic brain injury in cerebral amyloid angiopathy. *J. Cereb. Blood Flow Metab.* **2016**, *36*, 40–54. [[CrossRef](#)]
116. Boyle, P.A.; Yu, L.; Nag, S.; Leurgans, S.; Wilson, R.S.; Bennett, D.A.; Schneider, J.A. Cerebral amyloid angiopathy and cognitive outcomes in community-based older persons. *Neurology* **2015**, *85*, 1930–1936. [[CrossRef](#)]
117. Rennels, M.L.; Gregory, T.F.; Blaumanis, O.R.; Fujimoto, K.; Grady, P.A. Evidence for a 'paravascular' fluid circulation in the mammalian central nervous system, provided by the rapid distribution of tracer protein throughout the brain from the subarachnoid space. *Brain Res.* **1985**, *326*, 47–63. [[CrossRef](#)]
118. Bucchieri, F.; Farina, F.; Zummo, G.; Cappello, F. Lymphatic vessels of the dura mater: A new discovery? *J. Anat.* **2015**, *227*, 702–703. [[CrossRef](#)]
119. Zamboni, P. The discovery of the brain lymphatic system. *Veins Lymphat.* **2015**, *4*, 5360. [[CrossRef](#)]
120. Piehl, N.; van Olst, L.; Ramakrishnan, A.; Teregulova, V.; Simonton, B.; Zhang, Z.; Tapp, E.; Channappa, D.; Oh, H.; Losada, P.M.; et al. Cerebrospinal fluid immune dysregulation during healthy brain aging and cognitive impairment. *Cell* **2022**, *185*, 5028–5039.e13. [[CrossRef](#)]
121. Ishida, K.; Yamada, K.; Nishiyama, R.; Hashimoto, T.; Nishida, I.; Abe, Y.; Yasui, M.; Iwatsubo, T. Glymphatic system clears extracellular tau and protects from tau aggregation and neurodegeneration. *J. Exp. Med.* **2022**, *219*, e20211275. [[CrossRef](#)]
122. Absinta, M.; Ha, S.K.; Nair, G.; Sati, P.; Luciano, N.J.; Palisoc, M.; Louveau, A.; Zaghoul, K.A.; Pittaluga, S.; Kipnis, J.; et al. Human and nonhuman primate meninges harbor lymphatic vessels that can be visualized noninvasively by MRI. *eLife* **2017**, *6*, e29738. [[CrossRef](#)]
123. Mehta, R.I.; Carpenter, J.S.; Mehta, R.I.; Haut, M.W.; Ranjan, M.; Najib, U.; Lockman, P.; Wang, P.; D'haese, P.F.; Rezai, A.R. Blood-Brain Barrier Opening with MRI-guided Focused Ultrasound Elicits Meningeal Venous Permeability in Humans with Early Alzheimer Disease. *Radiology* **2021**, *298*, 654–662. [[CrossRef](#)]
124. Kanda, T. Central nervous system has a lymphatic system: What's next? *Clin. Exp. Neuroimmunol.* **2015**, *6*, 347–348. [[CrossRef](#)]
125. Rauch, J.N.; Luna, G.; Guzman, E.; Audouard, M.; Challis, C.; Sibih, Y.E.; Leshuk, C.; Hernandez, I.; Wegmann, S.; Hyman, B.T.; et al. LRP1 is a master regulator of tau uptake and spread. *Nature* **2020**, *580*, 381–385. [[CrossRef](#)]
126. Michaud, J.P.; Bellavance, M.A.; Préfontaine, P.; Rivest, S. Real-time in vivo imaging reveals the ability of monocytes to clear vascular amyloid beta. *Cell Rep.* **2013**, *5*, 646–653. [[CrossRef](#)]
127. Droujinine, I.A.; Perrimon, N. Defining the interorgan communication network: Systemic coordination of organismal cellular processes under homeostasis and localized stress. *Front. Cell Infect. Microbiol.* **2013**, *3*, 82. [[CrossRef](#)]
128. Palmqvist, S.; Tideman, P.; Cullen, N.; Zetterberg, H.; Blennow, K.; Alzheimer's Disease Neuroimaging Initiative; Dage, J.L.; Stomrud, E.; Janelidze, S.; Mattsson-Carlsson, N.; et al. Prediction of future Alzheimer's disease dementia using plasma phospho-tau combined with other accessible measures. *Nat. Med.* **2021**, *27*, 1034–1042. [[CrossRef](#)]
129. Julius, S.; Gudbrandsson, T.; Jamerson, K.; Andersson, O. The interconnection between sympathetics, microcirculation, and insulin resistance in hypertension. *Blood Press.* **1992**, *1*, 9–19. [[CrossRef](#)]
130. Louveau, A.; Plog, B.A.; Antila, S.; Alitalo, K.; Nedergaard, M.; Kipnis, J. Understanding the functions and relationships of the glymphatic system and meningeal lymphatics. *J. Clin. Investig.* **2017**, *127*, 3210–3219. [[CrossRef](#)]
131. Rahimi, J.; Kovacs, G.G. Prevalence of mixed pathologies in the aging brain. *Alzheimers Res. Ther.* **2014**, *6*, 82. [[CrossRef](#)] [[PubMed](#)]
132. De Reuck, J.; Deramecourt, V.; Cordonnier, C.; Pasquier, F.; Leys, D.; Maurage, C.A.; Bordet, R. The incidence of post-mortem neurodegenerative and cerebrovascular pathology in mixed dementia. *J. Neurol. Sci.* **2016**, *366*, 164–166. [[CrossRef](#)] [[PubMed](#)]
133. James, B.D.; Wilson, R.S.; Boyle, P.A.; Trojanowski, J.Q.; Bennett, D.A.; Schneider, J.A. TDP-43 stage, mixed pathologies, and clinical Alzheimer's-type dementia. *Brain* **2016**, *139*, 2983–2993. [[CrossRef](#)] [[PubMed](#)]
134. Nelson, P.T.; Dickson, D.W.; Trojanowski, J.Q.; Jack, C.R.; Boyle, P.A.; Arfanakis, K.; Rademakers, R.; Alafuzoff, I.; Attems, J.; Brayne, C.; et al. Limbic-predominant age-related TDP-43 encephalopathy (LATE): Consensus working group report. *Brain* **2019**, *142*, 1503–1527. [[CrossRef](#)]
135. Beach, T.G.; Malek-Ahmadi, M. Alzheimer's Disease Neuropathological Comorbidities are Common in the Younger-Old. *J. Alzheimers Dis.* **2021**, *79*, 389–400. [[CrossRef](#)]
136. Uchikado, H.; Lin, W.L.; DeLucia, M.W.; Dickson, D.W. Alzheimer disease with amygdala Lewy bodies: A distinct form of alpha-synucleinopathy. *J. Neuropathol. Exp. Neurol.* **2006**, *65*, 685–697. [[CrossRef](#)]

137. Nedergaard, M.; Goldman, S.A. Glymphatic failure as a final common pathway to dementia. *Science* **2020**, *370*, 50–56. [[CrossRef](#)]
138. Banks, W.A. The blood-brain barrier as an endocrine tissue. *Nat. Rev. Endocrinol.* **2019**, *15*, 444–455. [[CrossRef](#)]
139. Castellani, G.; Schwartz, M. Immunological Features of Non-neuronal Brain Cells: Implications for Alzheimer’s Disease Immunotherapy. *Trends Immunol.* **2020**, *41*, 794–804. [[CrossRef](#)]
140. Kim, H.; Li, Q.; Hempstead, B.L.; Madri, J.A. Paracrine and autocrine functions of brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) in brain-derived endothelial cells. *J. Biol. Chem.* **2004**, *279*, 33538–33546. [[CrossRef](#)]
141. Seshadri, S.; Beiser, A.; Selhub, J.; Jacques, P.F.; Rosenberg, I.H.; D’Agostino, R.B.; Wilson, P.W.; Wolf, P.A. Plasma homocysteine as a risk factor for dementia and Alzheimer’s disease. *N. Engl. J. Med.* **2002**, *346*, 476–483. [[CrossRef](#)]
142. Coleman, B.M.; Hill, A.F. Extracellular vesicles—Their role in the packaging and spread of misfolded proteins associated with neurodegenerative diseases. *Semin. Cell Dev. Biol.* **2015**, *40*, 89–96. [[CrossRef](#)]
143. Garcia-Contreras, M.; Thakor, A.S. Extracellular vesicles in Alzheimer’s disease: From pathology to therapeutic approaches. *Neural Regen. Res.* **2023**, *18*, 18–22.
144. Alexandrov, P.N.; Dua, P.; Hill, J.M.; Bhattacharjee, S.; Zhao, Y.; Lukiw, W.J. microRNA (miRNA) speciation in Alzheimer’s disease (AD) cerebrospinal fluid (CSF) and extracellular fluid (ECF). *Int. J. Biochem. Mol. Biol.* **2012**, *3*, 365–373.
145. Su, H.; Rustam, Y.H.; Masters, C.L.; Makalic, E.; McLean, C.A.; Hill, A.F.; Barnham, K.J.; Reid, G.E.; Vella, L.J. Characterization of brain-derived extracellular vesicle lipids in Alzheimer’s disease. *J. Extracell. Vesicles* **2021**, *10*, e12089. [[CrossRef](#)]
146. Ruan, Z.; Pathak, D.; Venkatesan Kalavai, S.; Yoshii-Kitahara, A.; Muraoka, S.; Bhatt, N.; Takamatsu-Yukawa, K.; Hu, J.; Wang, Y.; Hersh, S.; et al. Alzheimer’s disease brain-derived extracellular vesicles spread tau pathology in interneurons. *Brain* **2021**, *144*, 288–309. [[CrossRef](#)]
147. Ma, Q.; Xing, C.; Long, W.; Wang, H.Y.; Liu, Q.; Wang, R.F. Impact of microbiota on central nervous system and neurological diseases: The gut-brain axis. *J. Neuroinflamm.* **2019**, *16*, 53. [[CrossRef](#)]
148. Bairamian, D.; Sha, S.; Rolhion, N.; Sokol, H.; Dorothée, G.; Lemere, C.A.; Krantic, S. Microbiota in neuroinflammation and synaptic dysfunction: A focus on Alzheimer’s disease. *Mol. Neurodegener.* **2022**, *17*, 19. [[CrossRef](#)]
149. Haqqani, A.S.; Delaney, C.E.; Tremblay, T.L.; Sodja, C.; Sandhu, J.K.; Stanimirovic, D.B. Method for isolation and molecular characterization of extracellular microvesicles released from brain endothelial cells. *Fluids Barriers CNS* **2013**, *10*, 4. [[CrossRef](#)]
150. Kowalski, K.; Mulak, A. Brain-Gut-Microbiota Axis in Alzheimer’s Disease. *J. Neurogastroenterol. Motil.* **2019**, *25*, 48–60. [[CrossRef](#)]
151. Kim, M.S.; Kim, J.J. Heart and brain interconnection—Clinical implications of changes in brain function during heart failure. *Circ. J.* **2015**, *79*, 942–947. [[CrossRef](#)] [[PubMed](#)]
152. Li, C.; Chen, W.; Lin, F.; Li, W.; Wang, P.; Liao, G.; Zhang, L. Functional Two-Way Crosstalk Between Brain and Lung: The Brain-Lung Axis. *Cell. Mol. Neurobiol.* **2022**, *9*, 1–13. [[CrossRef](#)] [[PubMed](#)]
153. Lau, W.L.; Fisher, M.; Greenia, D.; Floriolli, D.; Fletcher, E.; Singh, B.; Sajjadi, S.A.; Corrada, M.M.; Whittle, C.; Kawas, C.; et al. Cystatin C, cognition, and brain MRI findings in 90+-year-olds. *Neurobiol. Aging* **2020**, *93*, 78–84. [[CrossRef](#)] [[PubMed](#)]
154. Uno, K. Roles of the interorgan neuronal network in the development of metabolic syndrome. *Diabetol. Int.* **2016**, *7*, 205–211. [[CrossRef](#)]
155. Franceschi, C.; Bonafè, M.; Valensin, S.; Olivieri, F.; De Luca, M.; Ottaviani, E.; De Benedictis, G. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann. N. Y. Acad. Sci.* **2000**, *908*, 244–254. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.