

DATA NOTE

The genome sequence of the European crab apple, Malus sylvestris (L.) Mill., 1768 [version 1; peer review: 2 approved]

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Abstract

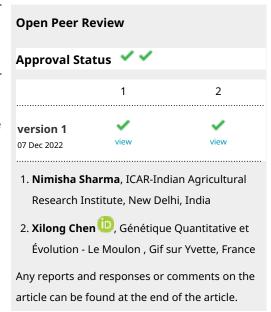
We present a genome assembly from an individual Malus sylvestris (the European or 'wild' crab apple; Streptophyta; Magnoliopsida; Rosales; Rosaceae). The genome sequence is 642 megabases in span. Most of the assembly (99.98%) is scaffolded into 17 chromosomal pseudomolecules. The mitochondrial and chloroplast genomes were also assembled, with respective lengths of 396.9 kilobases and 160.0 kilobases.

Keywords

Malus sylvestris, European crab apple, genome sequence, chromosomal, Rosaceae



This article is included in the Tree of Life gateway.



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Author roles: Ruhsam M: Investigation, Writing – Original Draft Preparation, Writing – Review & Editing; **Bell D**: Investigation, Resources, Writing – Original Draft Preparation, Writing – Review & Editing; **Hart M**: Investigation, Resources, Writing – Original Draft Preparation, Writing – Review & Editing; **Hollingsworth P**: Investigation, Resources, Writing – Original Draft Preparation, Writing – Review & Editing;

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Species taxonomy

Eukaryota; Viridiplantae; Streptophyta; Embryophyta; Tracheophyta; Spermatophyta; Magnoliopsida; eudicotyledons; Gunneridae; Pentapetalae; rosids; fabids; Rosales; Rosaceae; Amygdaloideae; Maleae; *Malus sylvestris* (L.) Mill., 1768 (NCBI:txid3752).

Background

Malus sylvestris (European wild or crab apple) belongs to the Rose family (Rosaceae) and is a small tree native across Europe. It reaches its north-western distribution limit in Scotland and Norway and is probably the UK's least understood and most under-appreciated tree, as even foresters sometimes do not seem to be aware of its native status. There is evidence of its longstanding use, as wild apples have been found at Neolithic and Bronze Age archaeological sites across Europe (Cornille et al. 2014). Additionally, without the European wild apple, the eating apples so many people love would not be quite the same. It is one of the main contributors to the domesticated apple, M. domestica (Cornille et al., 2014; Cornille et al., 2019). The widely planted cultivated varieties of M. domestica continue to hybridise with the wild apple trees, causing concern that the genetic integrity of M. sylvestris might be eroded in the long run (Cornille et al., 2013; Feurtey et al., 2017; Ruhsam et al., 2019; Schnitzler et al., 2014).

In a recent study, nearly 30% of the wild apple trees genotyped turned out to be of hybrid origin (Ruhsam *et al.*, 2019). As morphological identification of hybrid trees is difficult, access to the full genome of both *M. sylvestris* and *M. domestica* will facilitate the development of genome-wide species-specific markers, enabling the reliable assessment of levels of introgression in the European wild apple.

The genome of the European crab apple, *M. sylvestris*, was sequenced as part of the Darwin Tree of Life Project, a collaborative effort to sequence all named eukaryotic species in the Atlantic Archipelago of Britain and Ireland.

Genome sequence report

The genome was sequenced from a single *M. sylvestris* (hermaphroditic) (Figure 1), collected from Glen Falloch,



Figure 1. Image of the *Malus sylvestris* specimen from which samples used for sequencing were taken. The drMalSylv7 leaf samples were used to generate PacBio, 10X genomics and Hi-C data.

Scotland, UK. A total of 25-fold coverage in Pacific Biosciences single-molecule HiFi long reads and 87-fold coverage in 10X Genomics read clouds were generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 64 missing/misjoins and removed 10 haplotypic duplications, reducing the assembly size by 0.17% and the scaffold number by 36.0%.

The final assembly has a total length of 641 Mb in 32 sequence scaffolds with a scaffold N50 of 36.9 Mb (Table 1). Most of the assembled sequence (99.98%) was assigned to 17 chromosomal-level scaffolds numbered by synteny based on *Malus domestica* (apple) GCA_004115385.1 (Figure 2–Figure 5; Table 2).

Table 1. Genome data for Malus sylvestris, drMalSylv7.2.

Project accession data			
Assembly identifier	mbly identifier drMalSylv7.2		
Species	Malus sylvestris		
Specimen	drMalSylv7 (genome assembly, Hi-C)		
NCBI taxonomy ID	3752		
BioProject	PRJEB47316		
BioSample ID	SAMEA9197672		
Isolate information	Leaf tissue		
Raw data accessions			
PacificBiosciences SEQUEL II	ERR6808038, ERR6808039		
10X Genomics Illumina	ERR6688721-ERR6688723; ERR6688403		
Hi-C Illumina	ERR6688724		
Standard Illumina libraries	ERR6909395-ERR6909404		
Genome assembly			
Assembly accession	GCA_916048215.2		
Accession of alternate haplotype	GCA_916049865.1		
Span (Mb)	641		
Number of contigs	133		
Contig N50 length (Mb)	8.3		
Number of scaffolds	32		
Scaffold N50 length (Mb)	36.9		
Longest scaffold (Mb)	54.9		
BUSCO* genome score	C:98.5%[S:59.4%,D:39.1%], F:0.6%,M:0.9%,n:2,326		

*BUSCO scores based on the eudicots_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/drMalSylv7.1/dataset/CAJZHM01/busco. Page 3 of 12

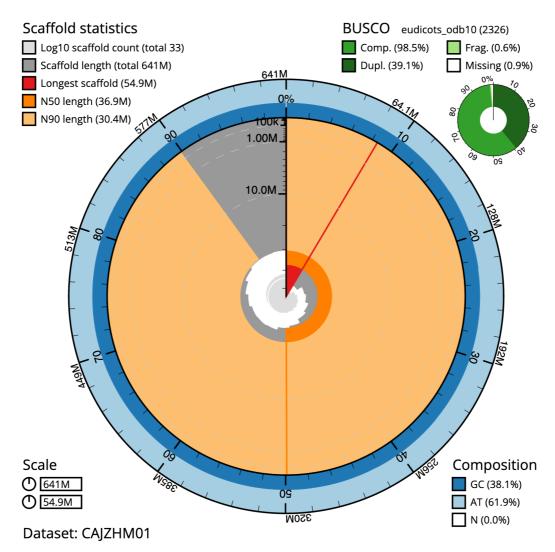


Figure 2. Genome assembly of *Malus sylvestris*, **drMalSylv7.1: metrics.** The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 640,986,597 bp assembly. The distribution of chromosome lengths is shown in dark grey with the plot radius scaled to the longest chromosome present in the assembly (54,898,662 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 chromosome lengths (36,902,754 and 30,380,474 bp), respectively. The pale grey spiral shows the cumulative chromosome count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the eudicots_odb10 set is shown in the top right. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/drMalSylv7.1/dataset/CA|ZHM01/snail.

Shared sequences between chromosomes are visible in the Hi-C map in agreement with the findings of (Velasco *et al.*, 2010) for *Malus domestica*. The Hi-C map provides evidence of inversions between haplotypes in chromosome 2 (24.82–26.19 Mb), and chromosome 11 (17.06–19.05 Mb). There are several scaffolds that it was possible to localise to a chromosome but not place, and these have been labelled as 'unloc'. Two of the largest unlocalised scaffolds – SUPER_4_unloc_1 and SUPER_14_unloc_1 – appear to be larger haplotypes of two specific loci: Chromosome 4 at ~26.13–26.31 Mb and

chromosome 14 at \sim 782 kb respectively. From the Hi-C it appears that these loci currently represent the shorter haplotype. As there is some uncertainty over these unlocalised scaffolds, they were left in the primary assembly.

The assembly has a BUSCO v5.3.2 (Manni *et al.*, 2021) completeness of 98.5% (single 59.4%, duplicated 39.1%) using the eudicots_odb10 reference set (n = 2,326). While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited.

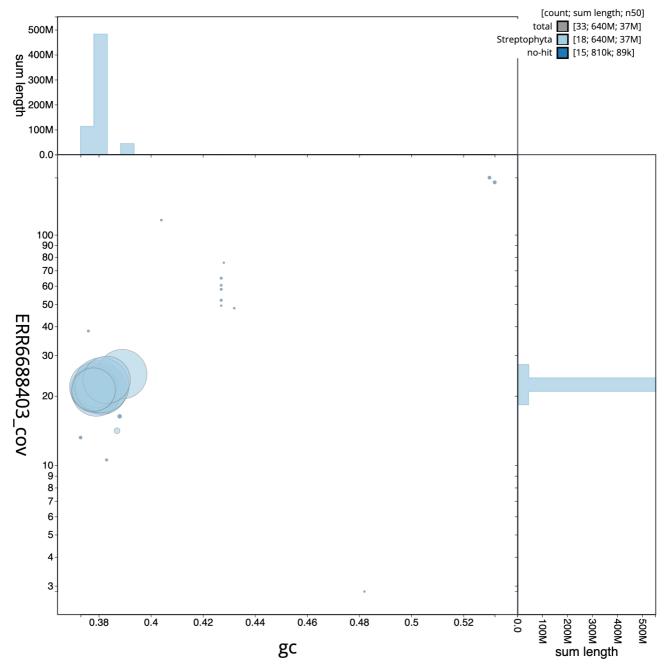


Figure 3. Genome assembly of *Malus sylvestris*, **drMalSylv7.1: GC coverage.** BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/drMalSylv7.1/dataset/CAJZHM01/blob.

Methods

Sample acquisition and nucleic acid extraction

Leaf samples from a single *M. sylvestris* specimen (drMalSylv7; genome assembly, Hi-C) were collected from Glen Falloch, Scotland, UK (latitude 56.343794, longitude –4.7078054) by Markus Ruhsam (Royal Botanic Garden Edinburgh), who also identified the specimen. The leaf samples were picked by hand, cut and put into FluidX tubes, then snap-frozen in liquid nitrogen.

The drMalSylv7 leaf sample was weighed and dissected on dry ice with tissue set aside for Hi-C sequencing. Leaf tissue was cryogenically disrupted to a fine powder using a Covaris cryoPREP Automated Dry Pulveriser, receiving multiple impacts. High molecular weight (HMW) DNA was extracted using the Qiagen Plant MagAttract HMW DNA extraction kit. Low molecular weight DNA was removed from a 200 ng aliquot of extracted DNA using 0.8X AMpure XP purification kit prior to 10X Chromium sequencing; a minimum of 50 ng DNA

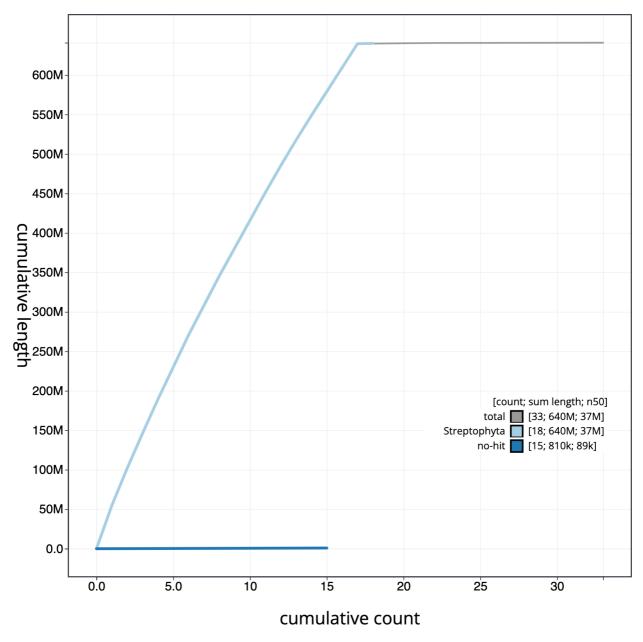


Figure 4. Genome assembly of *Malus sylvestris***, drMalSylv7.1: cumulative sequence.** BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/drMalSylv7.1/dataset/CAJZHM01/cumulative.

was submitted for 10X sequencing. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

Sequencing

Pacific Biosciences HiFi circular consensus and 10X Genomics Chromium read cloud sequencing libraries were constructed according to the manufacturers' instructions. Sequencing was performed by the Scientific Operations core at the Wellcome Sanger Institute on Pacific Biosciences SEQUEL II (HiFi) and Illumina NovaSeq 6000 (10X) instruments. Hi-C data were generated in the Tree of Life laboratory from remaining leaf tissue of drMalSylv7 using the Arima v2 kit and sequenced on a NovaSeq 6000 instrument. Standard read sequencing

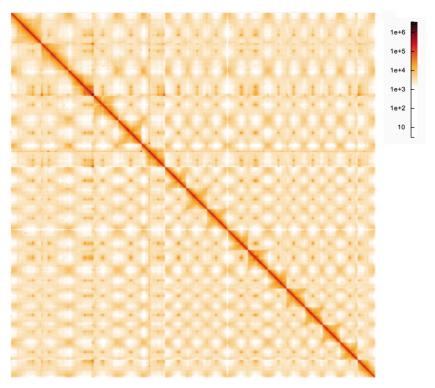


Figure 5. Genome assembly of *Malus sylvestris***, drMalSylv7.2: Hi-C contact map.** Hi-C contact map of the drMalSylv7.2 assembly, visualised in HiGlass. Chromosomes are arranged in size order from left to right and top to bottom. The interactive Hi-C map can be viewed at https://genome-note-higlass.tol.sanger.ac.uk/l/?d=FgGkT9v7SR2otUnBK-HkfQ.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Malus sylvestris*, drMalSylv7.2. Chromosome-scale scaffolds are numbered by synteny based on *Malus domestica* (apple) GCA_004115385.1.

INSDC accession	Name	Size (Mb)	GC%
OU696503.1	1	30.13	38.3
OU696504.1	2	37.96	37.8
OU696505.1	3	36.9	38
OU696506.1	4	31.14	38
OU696507.1	5	47.13	38.1
OU696508.1	6	34.96	38
OU696509.1	7	35.53	37.9
OU696510.1	8	30.37	37.9
OU696511.1	9	35.18	38.1
OU696512.1	10	43.2	37.9
OU696513.1	11	41.35	37.8
OU696514.1	12	32.55	37.9
OU696515.1	13	44.07	38.9
OU696516.1	14	30.38	37.9

INSDC accession	Name	Size (Mb)	GC%
OU696517.1	15	54.9	38.1
OU696518.1	16	40.12	38.3
OU696519.1	17	33.87	37.8
-	MT	0.4	45.4
-	Pltd	0.16	36.6
-	unplaced	1.23	41.7

libraries were generated using an Illumina NovaSeq 6000 (10X) instrument, as per the manufacturer's instructions.

Genome assembly

Assembly was carried out with Hifiasm (Cheng et al., 2021) and haplotypic duplication was identified and removed with purge_dups (Guan et al., 2020). One round of polishing was performed by aligning 10X Genomics read data to the assembly with longranger align, calling variants with freebayes (Garrison & Marth, 2012). The assembly was then scaffolded with Hi-C data (Rao et al., 2014) using SALSA2 (Ghurye et al., 2019). The assembly was checked for contamination as described previously (Howe et al., 2021). Manual curation was performed using HiGlass (Kerpedjiev et al., 2018) and Pretext (Harry, 2022). The mitochondrial and plastid genomes

were assembled using MBG (Rautiainen & Marschall, 2021) from PacBio HiFi reads mapping to related genomes. A representative circular sequence was selected for each from the graph based on read coverage. The genome was analysed and BUSCO scores were generated within the BlobToolKit environment (Challis *et al.*, 2020). Table 3 contains a list of the software tool versions used, where appropriate.

Ethics/compliance issues

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the Darwin Tree of Life Project Sampling Code of Practice. By agreeing with and signing up to the Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project. Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited

Table 3. Software tools used.

Software tool	Version	Source
Hifiasm	0.15.3-r339	Cheng <i>et al.</i> , 2021
purge_dups	1.2.3	Guan et al., 2020
SALSA2	2.2	Ghurye et al., 2019
longranger align	2.2.2	https://support.10xgenomics.com/ genome-exome/software/pipelines/ latest/advanced/other-pipelines
freebayes	1.3.1-17- gaa2ace8	Garrison & Marth, 2012
HiGlass	1.11.6	Kerpedjiev et al., 2018
PretextView	0.2.x	Harry, 2022
BlobToolKit	3.2.9	Challis et al., 2020

(operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Malus sylvestris* (European crab apple). Accession number PRJEB47316; https://identifiers.org/ena.embl/PRJEB47316 (Wellcome Sanger Institute, 2022)

The genome sequence is released openly for reuse. The *M. sylvestris* genome sequencing initiative is part of the Darwin Tree of Life (DToL) project. All raw sequence data and the assembly have been deposited in INSDC databases. The genome will be annotated and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1.

Author information

Members of the Royal Botanic Garden Edinburgh Genome Acquisition Lab are listed here: https://doi.org/10.5281/zenodo.4786682.

Members of the Darwin Tree of Life Barcoding collective are listed here:

https://doi.org/10.5281/zenodo.4893703.

Members of the Wellcome Sanger Institute Tree of Life programme are listed here:

https://doi.org/10.5281/zenodo.4783585.

Members of Wellcome Sanger Institute Scientific Operations: DNA Pipelines collective are listed here: https://doi.org/10.5281/zenodo.4790455.

Members of the Tree of Life Core Informatics collective are listed here:

https://doi.org/10.5281/zenodo.5013541.

Members of the Darwin Tree of Life Consortium are listed here:

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Open Peer Review

Current Peer Review Status:







Reviewer Report 25 January 2023

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Xilong Chen 🗓

Génétique Quantitative et Évolution - Le Moulon , Gif sur Yvette, France

This article presents the assembly of the genome of the European wild apple (*Malus sylvestris*) . The authors combined data from Pacific Biosciences, Illumina HiSeq 10X, and Hi-C to assemble the genome. The result has good assembly metrics N50 and BUSCO. The graphical information and the software listings help to understand the data and the reproducibility studies. The authors also compared the assembly results with the cultivated apple (*Malus domestica*) genome. Some structural variants were found.

To conclude, this article is an important contribution to European wild apple genome research and the genomic data provide a basis for studying apple domestication and Rosaceae evolution. In addition, I hope that the authors will complete the annotation of the genes and make it publicly available.

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?

Yes

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Population genetics, fruit tree biology, next generation sequencing

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 06 January 2023

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Nimisha Sharma

Division of Fruits and Horticultural Technology, ICAR-Indian Agricultural Research Institute, New Delhi, India

Assessing the genome of crab apple is very much useful for the apple breeders, as *M. domestica* is cultivated apple and wild apple is one of the contributors, to know the molecular mechanism behind any trait specific breeding and tailoring of the crop accordingly would be easier and their genetic mechanism can be interpreted.

- I feel abbreviations must be provided so that it will be more understandable to every discipline person.
- Reference genome assembly could be generated.
- Many times wild species are not taken into consideration but they may be very informative for cultivated apples and if breeders want to introgress any gene from the wild species, it could be possible with this genome.
- Present manuscript related with the sequence of apple genome and it is just 10X which is very low coverage.
- Figure 1 must focus on the studied species rather than background.
- Is this genome mapped with other published genomes of the Rosaceae family?

 Overvier, after revision present manuscript can be accepted as it imparts knowledge for

However, after revision present manuscript can be accepted as it imparts knowledge for other species as well.

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?

Yes

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Molecular Biology of Fruit Crops

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.