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### DATA NOTE

# The genome sequence of the Atlantic horse mackerel,

# *Trachurus trachurus* (Linnaeus 1758) [version 1; peer review: 3

# approved]

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### Abstract

We present a genome assembly from an individual *Trachurus trachurus* (the Atlantic horse mackerel; Chordata; Actinopteri; Carangiformes; Carangidae). The genome sequence is 801 megabases in span. The majority of the assembly, 98.68%, is scaffolded into 24 chromosomal pseudomolecules. Gene annotation of this assembly on Ensembl has identified 25,797 protein coding genes.

### **Keywords**

Trachurus trachurus, Atlantic horse mackerel, genome sequence, chromosomal



This article is included in the Tree of Life

gateway.

 Open Peer Review

 Approval Status

 1
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 3

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 Adelaide Rhodes D, Tufts University, Medford, USA

Any reports and responses or comments on the article can be found at the end of the article.

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Author roles: Genner M: Investigation, Resources, Writing – Original Draft Preparation, Writing – Review & Editing; Collins R: Investigation, Resources, Writing – Original Draft Preparation, Writing – Review & Editing;

**Competing interests:** No competing interests were disclosed.

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The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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### Species taxonomy

Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Actinopterygii; Neopterygii; Teleostei; Neoteleostei; Acanthomorphata; Carangaria; Carangiformes; Carangidae; Trachurus; *Trachurus trachurus* (Linnaeus, 1758) (NCBI txid:36212).

### Background

The Atlantic horse mackerel Trachurus trachurus (Linnaeus, 1758), also known as European horse mackerel or common scad, is northern Europe's only resident representative of the Carangidae, a ray-finned fish family that includes the jacks, pompanos and trevallies. Trachurus trachurus is a benthopelagic shoaling species and is typically found at depths of less than 200 m. The species has a broad distribution, including Iceland, Northeast Atlantic continental shelf waters, the Mediterranean, and north-western African coastal waters at least as far as Ghana (Healey et al., 2020). Atlantic horse mackerel are targeted by commercial fisheries using trawls, purse seines and long-lines. Major fished stocks are managed regionally. Those in Northeast Atlantic continental shelf waters are separated into a southern stock (Atlantic waters of the Iberian Peninsula), a western stock (shelf-edge seas from Bay of Biscay to the Norwegian coast, including spawning grounds of the Celtic Sea), and a North Sea stock (central and southern North Sea, including the Skagerrak and Kattegat) (ICES, 2019). Total landings of 140,000 metric tonnes were reported in 2018, down from catches of over 450,000 metric tonnes in the mid 1990s. On the basis of declining abundance over sections of the species range, it has been listed as Vulnerable by the International Union for the Conservation of Nature (Smith-Vaniz et al., 2015).

### **Genome sequence report**

The genome was sequenced from a single *T. trachurus* of unknown sex collected from Southampton Water, off the coast of Hampshire, UK. A total of 105-fold coverage in Pacific Biosciences single-molecule long reads (N50 23 kb) and 64-fold coverage in 10X Genomics read clouds (from molecules with an estimated N50 of 22 kb) were generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 141 missing/misjoins and removed 43 haplotypic duplications, reducing the scaffold number by 22.14%, increasing the scaffold N50 by 19.37% and decreasing the assembly length by 1.57%.

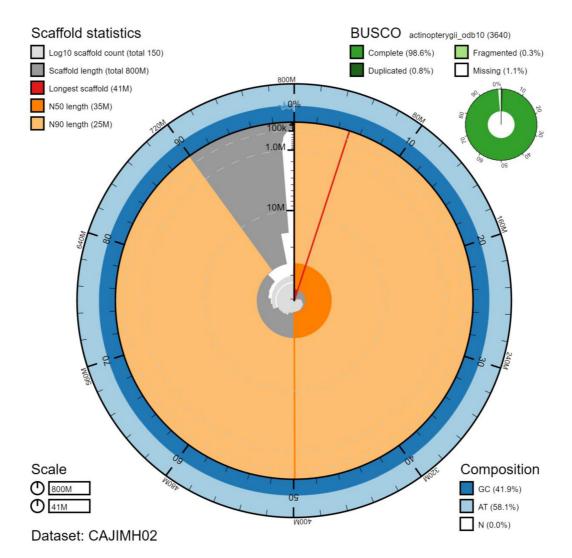
The final assembly has a total length of 801 Mb in 152 sequence scaffolds with a scaffold N50 of 35.4 Mb (Table 1). The majority, 98.68%, of the assembly sequence was assigned to 24 chromosomal-level scaffolds, representing 24 autosomes (numbered by synteny to *Oryzias latipes* (Japanese medaka); GCF\_002234675.1) (Figure 1–Figure 4; Table 2). The assembly has a BUSCO v5.1.2 (Manni *et al.*, 2021) completeness of 98.6% using the actinopterygii\_odb10 reference set. While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited.

### Table 1. Genome data for Trachurus trachurus, fTraTra1.2.

Project accession data	
Assembly identifier	fTraTra1.2
Species	Trachurus trachurus
Specimen	BMNH 2021.3.19.1; fTraTra1
NCBI taxonomy ID	NCBI:txid36212
BioProject	PRJEB42240
BioSample ID	SAMEA7524396
Isolate information	Muscle
Raw data accessions	
PacificBiosciences SEQUEL II	ERR6445210
10X Genomics Illumina	ERX5643309, ERX5643310, ERX5693250, ERX5693251
Hi-C Illumina	ERR6054366-ERR6054368
Genome assembly	
Assembly accession	GCA_905171665.2
Accession of alternate haplotype	GCA_905171655.2
Span (Mb)	801
Number of contigs	374
Contig N50 length (Mb)	6.49
Number of scaffolds	152
Scaffold N50 length (Mb)	35.45
Longest scaffold (Mb)	40.75
BUSCO* genome score	C:98.6%[S:97.8%,D:0.8%], F:0.3%,M:1.1%,n:3640
Genome annotation**	
Number of protein-coding genes	25,797
Average length of protein-coding gene (bp)	1,811
Average number of exons per gene	12
Average exon size (bp)	178
Average intron size (bp)	1,755

\*BUSCO scores based on the actinopterygii\_odb10 BUSCO set using v5.1.2. C= complete [S= single copy, D=duplicated], F=fragmented, M=missing, n=number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/fTraTra1.2/dataset/ CAJIMH02/busco.

\*\*Genome annotation provided for assembly fTraTra1.1.



**Figure 1. Genome assembly of** *Trachurus trachurus*, **fTraTra1.2: metrics.** The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 801,243,942 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (40,754,244 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (35,447,499 and 25,462,409 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the actinopterygii\_odb10 set is shown in the top right. An interactive version of this figure is available at https://blobtoolkit.genomehubs. org/view/fTraTra1.2/dataset/CAJIMH02/snail.

### **Gene annotation**

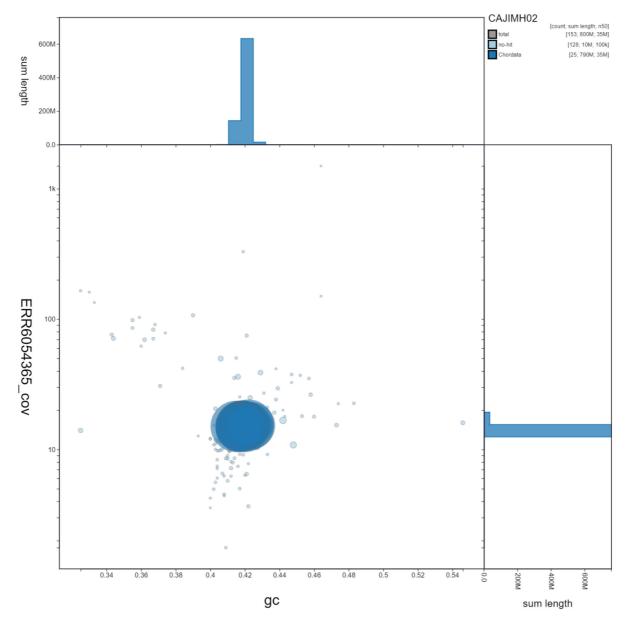
The Ensembl gene annotation system (Aken *et al.*, 2016) was used to generate annotation for the *T. trachurus* assembly (GCA\_905171665.1). Annotation was created primarily through alignment of transcriptomic data to the genome, with gap filling via protein to-genome alignments of a select set of vertebrate proteins from UniProt (UniProt Consortium, 2019) and coordinate mapping of GENCODE (Frankish *et al.*, 2019) mouse reference annotations via a pairwise whole genome alignment. The resulting Ensembl annotation includes 60,310

transcripts assigned to 25,797 coding and 2,264 non-coding genes (Trachurus trachurus - Ensembl Rapid Release).

### Methods

project.

Sample acquisition, DNA extraction and sequencing A single *T. trachurus* of unknown sex was collected in January 2017 from near Marchwood Power Station in Southampton Water, off the coast of Hampshire, UK (latitude 50.901563, longitude -1.440836) by Rupert Collins as part of the SeaDNA

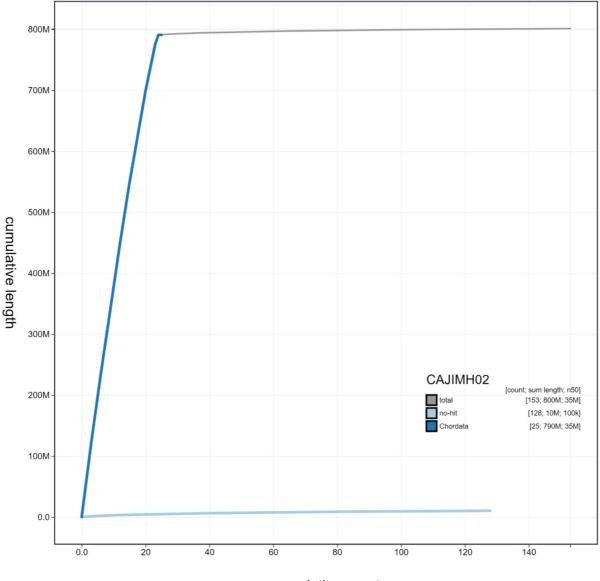


**Figure 2. Genome assembly of Trachurus trachurus, fTraTra1.2: GC coverage.** BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/fTraTra1.2/dataset/CAJIMH02/blob.

DNA was extracted using an agarose plug extraction from muscle tissue following the Bionano Prep Animal Tissue DNA Isolation Soft Tissue Protocol. Pacific Biosciences CLR long read and 10X Genomics read cloud sequencing libraries were constructed according to the manufacturers' instructions. Sequencing was performed by the Scientific Operations core at the Wellcome Sanger Institute on Pacific Biosciences SEQUEL II and Illumina HiSeq X instruments. Hi-C data were generated from muscle tissue using the Arima Hi-C kit and sequenced using a HiSeq X instrument.

### Genome assembly

Assembly was carried out following the Vertebrate Genome Project pipeline (Rhie *et al.*, 2020) with Falcon-unzip (Chin *et al.*, 2016). Haplotypic duplication was identified and removed with purge\_dups (Guan *et al.*, 2020) and a first round of scaffolding carried out with 10X Genomics read clouds using scaff10x. Scaffolding with Hi-C data (Rao *et al.*, 2014) was carried out with SALSA2 (Ghurye *et al.*, 2019). The Hi-C scaffolded assembly was polished with arrow using the PacBio data, then polished with the 10X Genomics Illumina data by

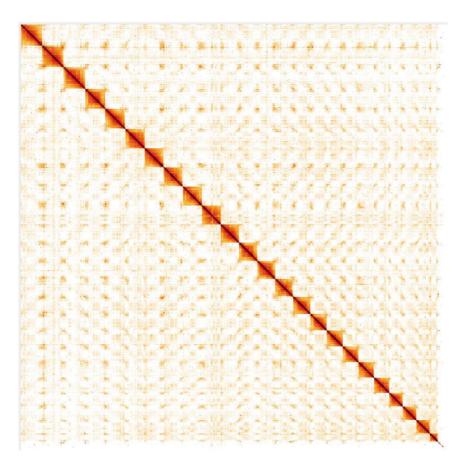


### cumulative count

**Figure 3. Genome assembly of** *Trachurus trachurus*, **fTraTra1.2: cumulative sequence.** BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/fTraTra1.2/dataset/ CAJIMH02/cumulative.

aligning to the assembly with longranger align, calling variants with freebayes (Garrison & Marth, 2012) and applying homozygous non-reference edits using bcftools consensus. Two rounds of the Illumina polishing were applied. The mitochondrial genome was assembled using the mitoVGP pipeline (Formenti *et al.*, 2021). The assembly was checked for contamination and corrected using the gEVAL system

(Chow *et al.*, 2016) as described previously (Howe *et al.*, 2021). Manual curation (Howe *et al.*, 2021) was performed using gEVAL, HiGlass (Kerpedjiev *et al.*, 2018) and Pretext. The genome was analysed and BUSCO scores generated using BlobToolKit (Challis *et al.*, 2020). Table 3 contains a list of all software tool versions used, where appropriate.



**Figure 4. Genome assembly of** *Trachurus trachurus*, **fTraTra1.2: Hi-C contact map.** Hi-C contact map of the fTraTra1.2 assembly, visualised in HiGlass. Chromosomes are given in order of size from top to bottom and left to right. The interactive Hi-C map can be viewed here.

INSDC accession	Chromosome	Size (Mb)	GC%
LR991628.1	1	40.75	41.8
LR991651.1	2	15.47	42.7
LR991630.1	3	40.66	41.5
LR991635.1	4	36.09	41.9
LR991633.1	5	36.89	41.7
LR991634.1	6	36.28	41.9
LR991638.1	7	35.45	42.0
LR991642.1	8	33.05	42.4
LR991632.1	9	37.40	41.9
LR991631.1	10	38.57	41.8
LR991641.1	11	33.39	42.1
LR991645.1	12	29.58	41.6

INSDC accession	Chromosome	Size (Mb)	GC%
LR991629.1	13	40.70	42.2
LR991640.1	14	33.71	41.8
LR991643.1	15	31.05	41.9
LR991639.1	16	35.01	41.9
LR991637.1	17	35.71	42.0
LR991646.1	18	29.25	42.0
LR991650.1	19	25.33	42.2
LR991648.1	20	25.46	42.2
LR991636.1	21	35.83	41.7
LR991644.1	22	30.60	42.2
LR991649.1	23	25.35	42.1
LR991647.1	24	29.13	42.0
LR991652.1	MT	0.02	46.5
-	Unplaced	10.50	31.6

Table 2 Chusmasser	ماييم من مماريم ما معرفا معرفا م
lable 2. Chromosoma	al pseudomolecules in the
nonomo occombly of	Trachurus trachurus, fTraTra1.2.
genome assembly of	Truchurus truchurus, Tiratra 1.2.

Software tool	Version	Source
Falcon unzip	1.4.2	Chin <i>et al.</i> , 2016
purge_dups	1.0.1	Guan <i>et al.</i> , 2020
scaff10x	4.1	https://github.com/wtsi-hpag/Scaff10X
SALSA2	2.2	Ghurye <i>et al.</i> , 2019
arrow	GCpp-1.9.0	https://github.com/PacificBiosciences/GenomicConsensus
freebayes	1.3.1-17-gaa2ace8	Garrison & Marth, 2012
mitoVGP	2.2	Formenti <i>et al.</i> , 2021
gEVAL	N/A	Chow <i>et al.</i> , 2016
HiGlass	1.11.6	Kerpedjiev <i>et al.,</i> 2018
PretextView	0.1.x	https://github.com/wtsi-hpag/PretextView
BlobToolKit	2.6.4	Challis <i>et al.</i> , 2020

#### Table 3. Software tools used.

### **Data availability**

European Nucleotide Archive: Trachurus trachurus (Atlantic horse mackerel). Accession number PRJEB42240; https://identifiers.org/ena.embl/PRJEB42240.

The genome sequence is released openly for reuse. The *T. trachurus* genome sequencing initiative is part of the Darwin Tree of Life (DToL) project and Vertebrate Genome Project (VGP). The specimen has been frozen and deposited with the Natural History Museum, London under registration number BMNH 2021.3.19.1, where it will remain accessible to the research community for posterity. All raw sequence data and the assembly have been deposited in INSDC databases. Raw data and assembly accession identifiers are reported in Table 1.

### Author information

Members of the Wellcome Sanger Institute Tree of Life programme are listed here: https://doi.org/10.5281/zenodo. 6125027.

Members of Wellcome Sanger Institute Scientific Operations: DNA Pipelines collective are listed here: https://doi.org/10.5281/zenodo.5746904.

Members of the Tree of Life Core Informatics collective are listed here: https://doi.org/10.5281/zenodo.6125046.

Members of the Darwin Tree of Life Consortium are listed here: https://doi.org/10.5281/zenodo.5638618.

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PubMed Abstract | Publisher Full Text | Free Full Text

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Guan D, McCarthy SA, Wood J, et al.: Identifying and Removing Haplotypic Duplication in Primary Genome Assemblies. *Bioinformatics*. 2020; 36(9): 2896-2898

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Healey AJE, Farthing MW, Nunoo FKE, et al.: Genetic Analysis Provides Insights into Species Distribution and Population Structure in East Atlantic Horse Mackerel (Trachurus Trachurus and T. Capensis). J Fish Biol. 2020; 96(3): 795-805

#### PubMed Abstract | Publisher Full Text | Free Full Text

Howe K, Chow W, Collins J, *et al.*: **Significantly Improving the Quality of Genome Assemblies through Curation**. *Gigascience*. 2021; **10**(1): giaa153.

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# 19(1): 125. PubMed Abstract | Publisher Full Text | Free Full Text

Manni M, Berkeley MR, Seppey M, et al.: BUSCO Update: Novel and Streamlined Workflows along with Broader and Deeper Phylogenetic Coverage for Scoring of Eukaryotic, Prokaryotic, and Viral Genomes. *Mol* Biol Evol. 2021; 38(10): 4647-54.

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### **Publisher Full Text**

Smith-Vaniz WF, Sidibe A, Nunoo F, et al.: Trachurus Trachurus. IUCN Red List of Threatened Species. 2015. Publisher Full Text

UniProt Consortium: UniProt: A Worldwide Hub of Protein Knowledge. Nucleic Acids Res. 2019; 47(D1): D506-15.

PubMed Abstract | Publisher Full Text | Free Full Text

# **Open Peer Review**

## Current Peer Review Status:

Version 1

Reviewer Report 10 January 2023

https://doi.org/10.21956/wellcomeopenres.19721.r53698

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### Adelaide Rhodes 匝

Tufts University, Medford, MA, USA

This genome report is an important contribution to the development of genome information an important fisheries stock from the family Carangidae, as the second published chromosome level assembly within the family and the third chromosome level assembly for the order Carangiformes. The paper does a comprehensive and complete job of laying out the methodology and results of the genome assembly. It is also written in a FAIR (findable, accessible, interoperable and reusable) format which will assist future researchers who may be responsible for managing the stocks. The final assembly has a total length of 801 Mb, found in 24 autosomes and 128 unplaced scaffolds with a scaffold N50 of 35.4 Mb. The prediction of 24 autosomal chromosomes is consistent with the only other chromosome level genome published for the the Carangidae family, *Seriola aureovittata*, which is smaller than the *T. trachurus* genome by approximately 151 Mb.

Only one minor error was detected, the link to the Hi-C interactive map in Figure 4 did not work at the time of this review. It would be nice to have a fixed link in regards to examining the data in more detail.

Another small issue is that the genetic material came from a single sample of unknown sex. Sex determination in fish as a whole is complicated, it would be helpful in future genome studies to find some confirmation of the sex of the sample if possible through gonadal cannulation or biopsy.

In addition the genome report finds after generating the 24 autosomes based on synteny to *Oryzias latipes* (Japanese medaka) that 128 scaffolds of N50 104,180 and length 10.5 Mb fell into the unplaced sequence. It would be interesting to determine if some polishing of the genome or the pseudo-alternative haplotype could resolve these unplaced scaffolds, perhaps by choosing a different fish model for syntenic comparison.

An alternative haplotype was generated containing 1,049 scaffolds with total length 797 Mb and an N50 of 1.6 Mb. It would be helpful, in this reviewer's opinion, to provide a few more details about the alignments of the alternate locus reference sequences to the main chromosome

sequences in the assembly to put these alternative loci into the context of the reference genome.

A few more comments on the significance of this genome report:

The horse mackerel is not a true "mackerel", which represents another important fisheries stock. It may be helpful, therefore, to point out that true mackerels are in the family Scombridae which holds many highly migratory species with unique challenges and adaptations. In contrast, the horse mackerel has two significant populations. The Northern stock of the horse mackerel spawns in the North Sea and heads back to Northern waters, whereas the Western stock spawns in the Bay of Biscay and heads west as the fish mature.

The family Carangidae contains 30 genera and approximately 152 species, which have very few whole genome assemblies as a group. At the time of this review, in National Center for Biotechnoloy Information (NCBI), National Library of Medicine (NLM), hosted by the National Institutes of Health (NIH), USA (https://www.ncbi.nlm.nih.gov/data-hub/taxonomy/1489907/), there are currently fifteen published Carangidae genomes other than the two from this genome report. Nine genomes are from the genus *Seriola*, which contains the amberjacks; two are from the genus *Caranx* (giant trevally and bluefin trevally) and three are from the genus from *Trachinotaus*, all three are from the golden pompano *Trachinotus ovatus*. *Seriola aureovittata* ( the great amberjack, synonymous with *Seriola lalandi*) is the only other species in the family Carangidae with a chromosome level assembly. Within the order Carangiformes, only one other fish, the live sharksucker *Echeneis naucrates* has a chromosome level assembly.

Overall, this genome report is a significant contribution which will help in the management of important fisheries stocks that are under pressure from climate change, habitat reduction and increasing economic demand for food from the oceans.

### Is the rationale for creating the dataset(s) clearly described?

Yes

### Are the protocols appropriate and is the work technically sound?

Yes

# Are sufficient details of methods and materials provided to allow replication by others? $\ensuremath{\mathsf{Yes}}$

### Are the datasets clearly presented in a useable and accessible format?

Partly

*Competing Interests:* No competing interests were disclosed.

*Reviewer Expertise:* Marine biology, Bioinformatics, Whole genome assemblies of non-model organisms, Metagenomics.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 03 January 2023

### https://doi.org/10.21956/wellcomeopenres.19721.r53697

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### Josephine Paris 问

Department of Health, Life and Environmental Sciences, University of L'Aquila, Coppito, Italy

This Data Note article presents a chromosome-level assembly of *Trachurus trachurus* using longread, short-read and Hi-C data for assembly. Being a commercially-important species, and the only resident member of the Carangidae family, the rationale for genomically enabling this species is very clear. The protocol for assembly follows the well-established VGP pipeline and the annotation is performed by Ensembl using the latest available proteomic analysis. Regarding the annotation, I miss some more detail on where the transcriptomic data is from (i.e. publicly available or generated in this project?) and also from which tissues the data are derived from (e.g. a broader range of tissues = a better chance of capturing all the protein-coding genes). I suggest this information is added to the article. Methods are clear and replicable. The genome, and its raw data, are available via the appropriate channels (i.e. NCBI / ENA), as well as the annotation via Ensembl's Rapid Release.

### Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound? Yes

# Are sufficient details of methods and materials provided to allow replication by others? $\ensuremath{\mathsf{Yes}}$

## Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Population genomics, genome assembly, transcriptomics

# I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 21 April 2022

https://doi.org/10.21956/wellcomeopenres.19721.r49622

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### Luís Filipe C. Castro 匝

CIIMAR-Interdisciplinary Centre of Marine and Environmental Research, U. Porto-University of Porto, Porto, Portugal

This genome provides a valuable resource in the context of teleost species that represent a fishery resource. The lack of a fully phased genome is not a significant problem. This will provide an opportunity for comparative genomics approaches with the ever growing number of high-quality genomes currently available for this range of taxa.

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?  $\ensuremath{\mathsf{Yes}}$ 

Are sufficient details of methods and materials provided to allow replication by others?  $\ensuremath{\mathsf{Yes}}$ 

Are the datasets clearly presented in a useable and accessible format?  $\ensuremath{\mathsf{Yes}}$ 

Competing Interests: No competing interests were disclosed.

**Reviewer Expertise:** Comparative Genomics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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