ORIGINAL ARTICLE



Humoral response to mRNA-based COVID-19 vaccine and booster effect of a third dose in patients with mature T cell and NK-cell neoplasms

Mirei Kobayashi¹ · Akio Mori¹ · Masahiro Onozawa² · Shihori Tsukamoto¹ · Hajime Senjo^{1,2} · Takashi Ishio¹ · Emi Yokoyama¹ · Minoru Kanaya¹ · Koh Izumiyama¹ · Makoto Saito¹ · Haruna Muraki^{3,4} · Masanobu Morioka¹ · Takanori Teshima² · Takeshi Kondo¹

Received: 4 January 2023 / Accepted: 15 February 2023 / Published online: 2 March 2023 © The Author(s) 2023

Abstract

Patients with lymphoid malignancies have impaired humoral immunity caused by the disease itself and its treatment, placing them at risk for severe coronavirus disease-19 (COVID-19) and reduced response to vaccination. However, data for COVID-19 vaccine responses in patients with mature T cell and NK-cell neoplasms are very limited. In this study of 19 patients with mature T/NK-cell neoplasms, anti-severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) spike antibodies were measured at 3 months, 6 months, and 9 months after the second mRNA-based vaccination. At the time of the second and third vaccinations, 31.6% and 15.4% of the patients were receiving active treatment. All patients received the primary vaccine dose and the third vaccination rate was 68.4%. In patients with mature T/NK-cell neoplasms, both seroconversion rate (p < 0.01) and antibody titers (p < 0.01) after the second vaccination were significantly lower than those in healthy controls (HC). In individuals who received the booster dose, patients had significantly lower antibody titers than those in HC (p < 0.01); however, the seroconversion rate in patients was 100%, which was the same as that in HC. The booster vaccine resulted in a significant increase of antibodies in elderly patients who had shown a response that was inferior to that in younger patients after two doses of vaccination. Since higher antibody titers and higher seroconversion rate reduced the

Mirei Kobayashi and Masahiro Onozawa contributed equally to this work.

Condensed abstract

In patients with mature T/NK-cell neoplasms, both the seroconversion rate and antibody titer after the second dose of mRNA-based COVID-19 vaccine were significantly lower than those in healthy controls, but the seroconversion rate of the patients increased to 100% after the booster dose.
The booster vaccine resulted in a significant increase of antibodies in elderly patients who had shown a response that was inferior to that in younger patients after two doses of vaccination.

Lay summary In patients with mature T/NK-cell neoplasms, both the seroconversion rate and antibody titer after the second dose of COVID-19 vaccine were significantly lower than those in healthy controls. In contrast, in patients who received the booster dose, the median antibody titer was significantly lower than that in healthy controls who received the booster; however, the seroconversion rate increased to 100% in the patients after the booster. Of note, the booster vaccine resulted in a significant increase of antibodies in elderly patients. Therefore, vaccination more than three times should be considered in patients with mature T/NK-cell neoplasms, especially in elderly patients.

Extended author information available on the last page of the article

incidence of infection and mortality, vaccination more than three times may have the advantage for patients with mature T/NK-cell neoplasms, especially in elderly patients. Clinical trial registration number: UMIN 000,045,267 (August 26th, 2021), 000,048,764 (August 26th, 2022).

Keywords COVID-19 · SARS-CoV-2 · Mature T cell and NK-cell neoplasms · Vaccine · Booster vaccine effect

Introduction

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the virus responsible for coronavirus disease-19 (COVID-19), has had devastating consequences globally and the end of the outbreak is not yet in sight. mRNA-based vaccines, BNT162b2 or mRNA-1273, were developed and these are useful in preventing infection and disease severity from COVID-19 [1-3]. However, previous pivotal studies conducted on only healthy individuals and did not include patients with hematological malignancies [1, 2, 4-6]. Therefore, data for COVID-19 vaccine responses in patients with hematological malignancies, particularly mature T cell and NK-cell neoplasms, are limited. In contrast, previous studies have reported that patients with hematological malignancies have a high mortality rate from COVID-19 [7–14]. Patients with hematological malignancies have impaired humoral immunity secondary to their malignancy and its treatment, placing them at risk for severe COVID-19 and reduced response to vaccination [15-18]. Furthermore, recent studies have reported that patients with hematological malignancies have lower seroconversion rate compared to that in healthy controls (HC) [19-25]. Homologous and heterologous booster vaccines with BNT162b2, mRNA-1273, and Ad26. COV2.S were shown to be immunogenic in healthy adults [26]. In contrast, the booster vaccine efficacy in patients with hematological malignancies seems to be inferior to that in healthy individuals [27-31]. In this study, we measured the antibody titers of COVID-19 in patients with mature T/NKcell neoplasms who received more than at least two doses of an mRNA-based COVID-19 vaccine and compared them to those in HC. We also investigated the antibody titers against SARS-CoV2 in patients with mature T/NK-cell neoplasms who received the third mRNA-based COVID-19 vaccine dose (booster).

Patients and methods

Patients

Patients with mature T/NK-cell neoplasms under follow up at the Blood Disorders Center at Aiiku Hospital during the period from May 26, 2021, to August 5, 2022, were included in this study. All of the patients were vaccinated with at least two doses of an mRNA-based COVID-19 vaccine, either BNT162b2 or mRNA-1273. BNT162b2 and mRNA-1273 were administered 21 and 28 days apart, respectively. All disease statuses of patients were determined at the time of the second vaccination. We recruited healthcare workers aged 50 years or older who had received at least two doses of BNT162b2 vaccine as HC. They were unlikely to be transmitted from inpatients since our hospital had not accepted COVID-19 patients. Individuals with a known history of COVID-19 were excluded from both cohorts of patients and HC. This study was a part of a prospective observation study (UMIN 000,045,267, 000,048,764) and it was conducted in compliance with ethical principles based on the Helsinki Declaration and was approved by the institutional review board of Aiiku Hospital. All patients provided written informed consent.

Assessment of serological response

Anti-SARS-CoV-2 spike (S) immunoassays were performed at 3 months \pm 4 weeks, 6 months \pm 4 weeks, and 9 months \pm 4 weeks after the second vaccine dose as previously described [32–35]. This assay has a minimum measurement value of 0.4 U/ml and a maximum measurement value of 25,000 U/ml. A value of 0.8 U/ml or higher is considered as a positive result [36]. An antibody titer less than 0.4 U/ml or more than 25,000 U/ml was calculated as 0.4 U/ml or 25,000 U/ml for convenience, respectively. All individuals who received a third dose were administered the third dose between 3-month and 9-month blood samplings, and the choice of cross-vaccination was made available.

Statistical analysis

Median antibody titers in paired samples and in unpaired samples were compared using the Wilcoxon signed-rank test and the Mann–Whitney U test, respectively. Differences between the means of two different groups were analyzed using the *t*-test. We used Fisher's exact test for categorical variables. p values less than 0.05 for statistical analyses were considered significant. All statistical analyses were performed with EZR (Jichi Medical University, Saitama, Japan) [37].

Results

Characteristics of patients and healthy controls

Nineteen patients with mature T/NK-cell neoplasms whose median age was 74 (range: 43–87) years were enrolled in this study. The characteristics of the patients are shown in Table 1. The patients included 6 patients with angio-immunoblastic T cell lymphoma (AITL), 5 patients with

peripheral T cell lymphoma, not otherwise specified (PTCL-NOS), 2 patients with anaplastic large cell lymphoma (ALCL), 2 patients with nodal peripheral T cell lymphoma with T follicular helper phenotype (PTCL with Tfh phenotype), one patient with adult T cell leukemia/ lymphoma (ATL/L), one patient with T cell large granular lymphocytic leukemia (T-LGLL), one patient with extranodal NK/T cell lymphoma, nasal type (ENKL), and one patient with monomorphic epitheliotropic intestinal T cell lymphoma (MEITL). HC included 29 individuals with a median age of 55 (50–72) years, and 62.1% were female. None of the patients with mature T/NK-cell neoplasms following at our hospital had a history of SARS-CoV-2 infection during the follow-up period.

Table 1 Patients' characteristics

	Mature T-cell and NK-cell neoplasms (n=19) 74 (43–87)	
Age median (range)		
Sex male/female	9/10	
Period from diagnosis to second vaccination, months, median (range)	23 (0–176)	
Period from second dose to third dose*, days, median (range)	228 (182–315)	
	First/second dose	Third dose
Patients with seroconversion, n (%)	14/19 (73.7)	13/13 (100)
On active therapy at the time of vaccination		
Yes	6	2
СНОР	1	0
CHOEP	1	0
Romidepsin monotherapy	1	0
Etoposide monotherapy	0	1
Prednisolone + cyclosporine	1	0
Prednisolone monotherapy	2	1
No	13	11
Treatment off in CR	10	10
Terminated active treatment	1	1
Previously untreated (treatment-naïve)	2	0
Status at the time of vaccination		
CR	12	10
PR	3	2
SD	1	1
Relapse/refractory	1	0
Previously untreated (treatment-naive)	2	0
Vaccination rate, n (%)	19/19 (100)	13/19 (68.4)
Vaccine subtype		
BNT162b2	17	8
mRNA-1273	2	5
Not yet vaccinated	0	6

*Patients who did not receive the third dose were excluded. Abbreviations: CHOP, (cyclophosphamide, doxorubicin, vincristine, and prednisolone); CHOEP, CHOP plus etoposide; CR, complete response; PR, partial response; SD, stable disease

Seroconversion rates and antibody titers at 3 months after the second vaccination

Six of the 19 patients were receiving treatment at the time of the second vaccination. The seroconversion rate after the second vaccination in patients was 73.7%, which was significantly lower than that in HC (100%) (p < 0.01). At 3 months after the second vaccination, patients showed a significantly lower antibody titer than that in HC (67.8 (interquartile range (IQR): 0.8–893.0) U/ml vs. 974.0 (608.0–1466.0) U/ml, p < 0.01) (Fig. 1A).

Seroconversion rates and antibody titers after the booster dose

Third vaccination rates were 89.7% and 68.4% in HC and patients, respectively. Two of the 13 patients who received the third dose were receiving active treatment at the time of the third vaccination. Seroconversion rates after the third vaccination in patients and HC were both 100%.

Next, we examined seroconversion rates at 9 months in patients who received the booster vaccine (n = 13) and those who did not receive the booster vaccine (n = 5). The seroconversion rate at 9 months was significantly higher in patients who received the booster vaccine than in patients who did not receive the booster vaccine (100% vs. 40.0%) p < 0.05). The antibody titer in patients who did not receive the booster dose (0.4 (0.4-42.7) U/ml vs. 15,324.0 (10,122.5-24,361.3) U/ml, p < 0.001) and that in patients who received the booster dose (4515.0 (273.0-15,373.0) U/ml, p < 0.01) at 9 months were both lower than that in healthy individuals who received the booster dose (Fig. 1B). Furthermore, patients who received the booster vaccine had a significantly higher median antibody titer at 9 months than that in patients who did not receive the booster vaccine (4515.0 (273.0-15,373.0) U/ml vs. 0.4 (0.4-42.7) U/ml, p < 0.05) (Fig. 1B). The mean age of the five patients who did not receive the booster vaccine was $62.2 (\pm 15.8)$ years old. In contrast, the mean age of the 13 patients who had received the booster vaccine was 74.2 (± 9.9) years old, which was older than that of the patients who had not received the booster vaccine although the difference was not significant (p = 0.0692).

In contrast, four of the five patients who had not received the booster vaccine were receiving active treatment at the time of the second vaccination. The other patient who was not receiving active treatment at the time of the second vaccination had a low antibody titer of less than 0.4 U/ml at 9 months. Only one of the 13 patients who received the booster vaccine was receiving active treatment at the time of the second vaccination. The antibody titer of this patient at 9 months was relatively high at 4500 U/ml.



Fig. 1 (**A**) Anti-SARS-CoV-2 S antibody titers at 3 months after the second vaccination in healthy controls and in patients with mature T/NK-cell neoplasms. (**B**) Anti-SARS-CoV-2 S antibody titers at 9 months after the second vaccination in healthy controls and patients with mature T/NK-cell neoplasms who received the booster vaccine

or those who did not receive the booster vaccine. The Mann–Whitney U test was used to compare medians of antibody titers. The two short lines show interquartile range (IQR) and the center long line shows the median. HC, healthy controls

Time-dependent decrease in antibody titer and booster vaccine response

The median antibody titers in 11 patients and 26 healthy individuals who had all 3-month, 6-month, and 9-month paired samples and who also received the third dose were compared using the Wilcoxon signed-rank test. Since patients who received third dose before the 6-month blood sampling were excluded, all third doses were administered between 6-month and 9-month blood samplings in this analysis. In HC, the median antibody titer at 6 months was significantly lower than that at 3 months (582.5 (263.3-1067.0) U/ml vs. 913.5 (514.3–1456.3) U/ml, *p* < 0.001). Even in patients, the median antibody titer at 6 months was also lower than that at 3 months, but the difference was marginally tendency (p=0.0665). After the booster vaccine, there was a significant increase in antibody titers in both HC (582.5 (263.3-1067.0) U/ml vs. 15,324.0 (10,122.5-24,361.3) U/ ml, p < 0.001) and patients (142.0 (0.7-478.0) U/ml vs. 4515.0 (157.3–12,938.5) U/ml, *p* < 0.01) (Fig. 2).

Factors affecting vaccine responses in patients with mature T/NK-cell neoplasms

Since the median age of the patients in our cohort was 74 years, we divided the patients by age of 75 years for clinical usefulness. The seroconversion rate at 3 months after the second vaccination in patients aged 75 years or older was lower than that in patients younger than 75 years of age, but the difference was not statistically significant (55.6% vs. 90.0%, p = 0.1410). After the booster vaccination, the seroconversion rates increased to 100% in

both patients younger than 75 years of age and patients aged 75 years or older. In contrast, the median antibody titer at 3 months after the second dose in patients aged 75 years or older was significantly lower than that in patients younger than 75 years of age (0.8 (0.7–4.1) U/ml vs. 415.5 (118.1–1192.0) U/ml, p < 0.05) (Fig. 3). Of note, after the booster dose, the median antibody titer in patients aged 75 years or older was comparable to that in patients younger than 75 years of age (2278.3 (36.0–9006.8) U/ml vs. 7679.0 (3632.5–15,961.5) U/ml, p = 0.2340).

The vaccine response was examined in 6 patients who were receiving active treatment and 13 patients who were not receiving active treatment at the time of the second vaccination. There was no significant difference in the seroconversion rate at 3 months after the second vaccination between patients who were receiving and those who were not receiving active treatment (66.7% vs. 76.9%, p = 1.0000). The median antibody titer in patients receiving active treatment was lower than that in untreated patients at 3 months after the second vaccination, but the difference was not statistically significant (Supplemental Fig. 1). Seroconversion rates after the third vaccination in patients receiving and those not receiving active treatment were both 100%, and there was no significant difference in the median antibody titers after the booster dose between these patients (Supplemental Fig. 1). There were no significant differences in seroconversion rate (complete response (CR) vs. non-CR: 66.7% vs. 100%, p = 0.2610at 3 months, both 100% at 9 months) and antibody titers depending on disease status at the second vaccination both at 3 months and 9 months after the second vaccination (Supplemental Fig. 2).

Fig. 2 Anti-SARS-CoV-2 S antibody titers after the initial vaccination and booster vaccination in healthy controls and patients with mature T/NK-cell neoplasms. Individuals who did not receive a third dose were excluded from this analysis. The Wilcoxon signed-rank test was used to compare medians of antibody titers in paired individuals. The two short lines show interquartile range (IQR) and the center long line shows the median. HC, healthy controls. M. months



Fig. 3 Anti-SARS-CoV-2 S antibody titers at 3 months and 9 months after the second vaccination in patients 75 years of age or older and in patients younger than 75 years of age. Individuals who did not receive a third dose prior to the 9-month blood sampling were excluded from the analysis of 9-month blood sampling. The Mann-Whitney U test was used to compare medians of antibody titers. The two short lines show interquartile range (IQR) and the center long line shows the median. M, months



Antibody titers in each patient

The dynamics of antibody titers in each patient are shown in Supplemental Fig. 3. Five patients had antibody titers of more than 1000 U/ml at 3 months. Their ages varied from 53 to 85 years old. Four of the five patients remained in CR for more than one year after the end of treatment until the time of initial vaccination. Two of them remained in CR for more than 8 years after the end of treatment until the time of initial vaccination. These two patients had high antibody titers of more than 5000 U/ml at 6 months, but they had received the booster vaccine 5 and 9 days before the day of 6-month blood sampling, respectively. As we mentioned before, these patients were excluded from the Wilcoxon signed-rank analysis in Fig. 2.

Discussion

In our institution, we routinely recommend COVID-19 vaccination to our patients, especially these with malignant lymphoma who were receiving active treatment. Recent studies have shown that the seroconversion rates after two doses of mRNA-based COVID-19 vaccines were 34–58% in patients with aggressive non-Hodgkin lymphoma (NHL) and 43–61% in patients with indolent NHL [28, 38, 39]. In patients with lymphoid malignancies, the seroconversion rate and antibody titer were significantly lower than those in HC after the second vaccination [39]. In a study of only 8 patients with T cell lymphoma, the seroconversion rate was 75% after two doses of mRNA-based vaccines, which is consistent with our results [39]. Furthermore, the seroconversion rate after a booster vaccine in patients with B-NHL in whom two previous doses were ineffective was 29.5% [30]. Those studies also showed that recent administration of anti-CD20 monoclonal antibodies reduces the vaccine response [28, 30, 39]. Furthermore, in patients with hematological malignancies, the vaccine responses to the primary and booster doses were reported to be lower in elderly patients [20–23, 25, 27–29]. However, data for COVID-19 vaccine responses in patients with mature T/NK-cell neoplasms, especially data for the booster effect, are very limited.

This study was conducted in patients with mature T/ NK-cell neoplasms, and although none of the patients had received anti-CD20 monoclonal antibodies at the time of vaccinations, the seroconversion rate and median antibody titer after two doses were lower than those in HC, as previously reported for patients with B cell malignancies [24, 25, 28, 30, 38, 39]. Furthermore, patients who received the booster dose had lower antibody titers at 9 months than those in healthy individuals who received the booster dose. These results may have been influenced by CHOP (cyclophosphamide, doxorubicin, vincristine, and prednisolone) therapy and other chemotherapy as well as the inherent immunodeficiency of patients with malignant lymphoma itself. However, since the number of patients was small in our study, it seemed to be difficult to perform further sub-analysis for association between treatment and booster vaccine efficacy. On the other hand, since patients who were receiving active treatment tend to have lower antibody titers than those who were not receiving active treatment, we recommend tixagevimab and cilgavimab for these patients [40].

In patients with mature T/NK-cell neoplasms, the seroconversion rate after the second dose was 73.7%, which was significantly lower than that in HC, but the seroconversion rate after the booster dose in the patients was 100%, which was the same as that in HC. In patients with hematological malignancies, although no significant association was found between the seroconversion rate after vaccination and the incidence of COVID-19, no patient with seroconversion died from COVID-19 [28]. Furthermore, as shown in Fig. 2 and Supplemental Fig. 3, it should be noted that the acquired antibody titer decreases in a time-dependent manner. Therefore, vaccination more than three times should be considered in patients with mature T/NK-cell neoplasms.

The median antibody titer in patients aged 75 years or older was significantly lower than that in patients younger than 75 years of age at 3 months after the primary vaccination. However, the median antibody titer after administration of the booster vaccine in patients aged 75 years or older increased to a level comparable to that in patients younger than 75 years of age. Although the preventive antibody titer against SARS-CoV-2 infection is unknown, it has been reported that higher antibody titers contributed to protection against SARS-CoV-2 infection [41, 42]. Therefore, since a booster dose may be more beneficial in elderly patients, administration of a booster vaccine should be considered especially in the elderly.

Our study has several limitations. HC were health care workers. Although our hospital had not accepted COVID-19 patients, the possibility of individuals with asymptomatic COVID-19 cannot be ruled out. The median age of the patient was 74 (range: 43–87) years, whereas the median age of HC was 55 (50-72) years. Thus, there appeared to be differences in the age distribution of patients and HC. This study included a heterogenous patient population and only humoral immunity was evaluated. T cell immunity, which is also important for preventing a severe course of COVID-19, was not evaluated [43]. Our study included both patients who had completed the treatment and those undergoing the treatment. The proportion of patients who were receiving active treatment at the time of the second vaccination in the group of patients who had not received the booster vaccine was higher than the group of patients who had received the booster vaccine. Therefore, as shown in Supplemental Fig. 1, it cannot be ruled out that treatment may have caused an underestimation of antibody titers at 9 months in the group of patients who had not received the booster vaccine. Finally, the number of patients was small. Therefore, our results should be confirmed by further prospective large-scale studies.

In conclusion, in patients with mature T/NK-cell neoplasms, both the seroconversion rate and antibody titer after the second vaccination were significantly lower than those in HC. In contrast, in patients who received the booster dose, the median antibody titer was significantly lower than that in HC who received the booster dose, but the seroconversion rate increased to 100% in the patients after the booster. Of note, the booster vaccine resulted in a significant increase of antibodies in elderly patients who had shown a response that was inferior to that in younger patients after two doses of vaccination. Therefore, vaccination more than three times may have the advantage for patients with mature T/NK-cell neoplasms, especially in elderly patients.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00277-023-05142-4.

Acknowledgements The authors thank the nurses and medical staff at Aiiku Hospital who took care of the patients. We gave special thanks to Ms. Yuko Fujimaki, the nursing manager, Mr. Masayuki Kudo, the vice general manager, and Ms. Kaori Shin for her work in data entry and management. This work was supported by Yahoo Japan Foundation.

Author contribution Contribution: M.K. designed the study, analyzed the data, and wrote the manuscript. A.M. designed the study and revised and approved the manuscript. S.T., H.S., T.I., E.Y., M.K., K.I., M.S., and M.M. performed recruitment and treatment of patients and opined on the manuscript. H.M. performed experiments and opined on the manuscript. M.O. revised the manuscript. T.T. supervised and revised the manuscript. All authors read and approved the final manuscript.

Data availability The datasets generated and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations

Informed consent Informed consent was obtained from all patients for being included in the study.

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Polack FP, Thomas SJ, Kitchin N et al (2020) Safety and efficacy of the BNT162b2 mRNA COVID-19 vaccine. N Engl J Med 383:2603–2615. https://doi.org/10.1056/NEJMoa2034577
- Baden LR, El Sahly HME, Essink B et al (2021) Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. N Engl J Med 384:403–416. https://doi.org/10.1056/NEJMoa2035389
- Dagan N, Barda N, Kepten E et al (2021) BNT162b2 mRNA COVID-19 vaccine in a nationwide mass vaccination setting. N Engl J Med 384:1412–1423. https://doi.org/10.1056/NEJMoa2101765
- Vasileiou E, Simpson CR, Shi T et al (2021) Interim findings from first-dose mass COVID-19 vaccination roll-out and COVID-19

hospital admissions in Scotland: a national prospective cohort study. Lancet 397:1646–1657. https://doi.org/10.1016/S0140-6736(21)00677-2

- Heath PT, Galiza EP, Baxter DN et al (2021) Safety and efficacy of NVX-CoV2373 COVID-19 vaccine. N Engl J Med 385:1172– 1183. https://doi.org/10.1056/NEJMoa2107659
- Voysey M, Clemens SAC, Madhi SA et al (2021) Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet 397:99–111. https://doi.org/10.1016/S0140-6736(20)32661-1
- Dai M, Liu D, Liu M et al (2020) Patients with cancer appear more vulnerable to SARS-CoV-2: a multicenter study during the COVID-19 outbreak. Cancer Discov 10:783–791. https://doi.org/ 10.1158/2159-8290.CD-20-0422
- Mehta V, Goel S, Kabarriti R et al (2020) Case fatality rate of cancer patients with COVID-19 in a New York Hospital System. Cancer Discov 10:935–941. https://doi.org/10.1158/2159-8290. CD-20-0516
- Lee LYW, Cazier JB, Starkey T et al (2020) COVID-19 prevalence and mortality in patients with cancer and the effect of primary tumour subtype and patient demographics: a prospective cohort study. Lancet Oncol 21:1309–1316. https://doi.org/10.1016/ S1470-2045(20)30442-3
- Mato AR, Roeker LE, Lamanna N et al (2020) Outcomes of COVID-19 in patients with CLL: a multicenter international experience. Blood 136:1134–1143. https://doi.org/10.1182/blood. 2020006965
- Hultcrantz M, Richter J, Rosenbaum CA et al (2020) COVID-19 infections and clinical outcomes in patients with multiple myeloma in New York City: a cohort study from five academic centers. Blood Cancer Discov 1:234–243. https://doi.org/10.1158/ 2643-3230.BCD-20-0102
- Chari A, Samur MK, Martinez-Lopez J et al (2020) Clinical features associated with COVID-19 outcome in multiple myeloma: first results from the International Myeloma Society data set. Blood 136:3033–3040. https://doi.org/10.1182/blood.2020008150
- Sharafeldin N, Bates B, Song Q et al (2021) Outcomes of COVID-19 in patients with cancer: report from the National COVID Cohort Collaborative (N3C). J Clin Oncol 39:2232–2246. https:// doi.org/10.1200/JCO.21.01074
- 14. Vijenthira A, Gong IY, Fox TA et al (2020) Outcomes of patients with hematologic malignancies and COVID-19: a systematic review and meta-analysis of 3377 patients. Blood 136:2881–2892. https://doi.org/10.1182/blood.2020008824
- Ollila TA, Lu S, Masel R et al (2021) Antibody response to COVID-19 vaccination in adults with hematologic malignant disease. JAMA Oncol 7:1714–1716. https://doi.org/10.1001/jamao ncol.2021.4381
- Ribas A, Dhodapkar MV, Campbell KM et al (2021) How to provide the needed protection from COVID-19 to patients with hematologic malignancies. Blood Cancer Discov 2:562–567. https:// doi.org/10.1158/2643-3230.BCD-21-0166
- Bedognetti D, Zoppoli G, Massucco C et al (2011) Impaired response to influenza vaccine associated with persistent memory B cell depletion in non-Hodgkin's lymphoma patients treated with rituximab-containing regimens. J Immunol 186:6044–6055. https://doi.org/10.4049/jimmunol.1004095
- Teh JSK, Coussement J, Neoh ZCF et al (2022) Immunogenicity of COVID-19 vaccines in patients with hematological malignancy: a systematic review and meta-analysis. Blood Adv 6:2014– 2034. https://doi.org/10.1182/bloodadvances.2021006333
- Oekelen OV, Gleason CR, Agte S et al (2021) Highly variable SARS-CoV-2 spike antibody responses to two doses of COVID-19 RNA vaccination in patients with multiple myeloma. Cancer Cell 39:1028–1030. https://doi.org/10.1016/j.ccell.2021.06.014

- Herishanu Y, Avivi I, Aharon A et al (2021) Efficacy of the BNT162b2 mRNA COVID-19 vaccine in patients with chronic lymphocytic leukemia. Blood 137:3165–3173. https://doi.org/10. 1182/blood.2021011568
- Avivi I, Balaban R, Shragai T et al (2021) Humoral response rate and predictors of response to BNT162b2 mRNA COVID19 vaccine in patients with multiple myeloma. Br J Haematol 195:186– 193. https://doi.org/10.1111/bjh.17608
- Gurion R, Rozovski U, Itchaki G et al (2022) Humoral serologic response to the BNT162b2 vaccine is abrogated in lymphoma patients within the first 12 months following treatment with anti-CD2O antibodies. Haematologica 107:715–720. https://doi.org/ 10.3324/haematol.2021.279216
- Maneikis K, Šablauskas K, Ringelevičiūtė U et al (2021) Immunogenicity of the BNT162b2 COVID-19 mRNA vaccine and early clinical outcomes in patients with haematological malignancies in Lithuania: a national prospective cohort study. Lancet Haematol 8:e583–e592. https://doi.org/10.1016/S2352-3026(21)00169-1
- Perry C, Luttwak E, Balaban R et al (2021) Efficacy of the BNT162b2 mRNA COVID-19 vaccine in patients with B-cell non-Hodgkin lymphoma. Blood Adv 5:3053–3061. https://doi. org/10.1182/bloodadvances.2021005094
- Chung DJ, Shah GL, Devlin SM et al (2021) Disease- and therapy-specific impact on humoral immune responses to COVID-19 vaccination in hematologic malignancies. Blood Cancer Discov 2:568–576. https://doi.org/10.1158/2643-3230.BCD-21-0139
- Atmar RL, Lyke KE, Deming ME et al (2022) Homologous and heterologous COVID-19 booster vaccinations. N Engl J Med 386:1046–1057. https://doi.org/10.1056/NEJMoa2116414
- Mai AS, Lee ARYB, Tay RYK et al (2022) Booster doses of COVID-19 vaccines for patients with haematological and solid cancer: a systematic review and individual patient data meta-analysis. Eur J Cancer 172:65–75. https://doi.org/10.1016/j.ejca.2022.05.029
- Ollila TA, Masel RH, Reagan JL et al (2022) Seroconversion and outcomes after initial and booster COVID-19 vaccination in adults with hematologic malignancies. Cancer 128:3319–3329. https:// doi.org/10.1002/cncr.34354
- 29. Herishanu Y, Rahav G, Levi S et al (2022) Efficacy of a third BNT162b2 mRNA COVID-19 vaccine dose in patients with CLL who failed standard 2-dose vaccination. Blood 139:678–685. https://doi.org/10.1182/blood.2021014085
- 30. Avivi I, Luttwak E, Saiag E et al (2022) BNT162b2 mRNA COVID-19 vaccine booster induces seroconversion in patients with B-cell non-Hodgkin lymphoma who failed to respond to two prior vaccine doses. Br J Haematol 196:1329–1333. https://doi. org/10.1111/bjh.18029
- 31. Kohn M, Delord M, Chbat M et al (2022) A third anti-SARS-CoV-2 mRNA dose does not overcome the pejorative impact of anti-CD20 therapy and/or low immunoglobulin levels in patients with lymphoma or chronic lymphocytic leukemia. Haematologica 107:1454–1459. https://doi.org/10.3324/haematol.2021.280026
- 32. Kageyama T, Ikeda K, Tanaka S et al (2021) Antibody responses to BNT162b2 mRNA COVID-19 vaccine and their predictors among healthcare workers in a tertiary referral hospital in Japan. Clin Microbiol Infect 27:1861.e1-1861.e5. https://doi.org/10. 1016/j.cmi.2021.07.042
- Mori A, Onozawa M, Tsukamoto S et al (2022) Humoral response to mRNA-based COVID-19 vaccine in patients with myeloid malignancies. Br J Haematol 197:691–696. https://doi.org/10.1111/bjh.18138
- 34. Mori A, Onozawa M, Kobayashi M et al (2022) Humoral response to mRNA-based COVID-19 vaccine in patients with de novo and pre-existing immune thrombocytopenia with exacerbation of thrombocytopenia after vaccination. Br J Haematol 199:627–630. https://doi.org/10.1111/bjh.18447
- 35. Mori A, Onozawa M, Kobayashi M et al (2022) Humoral response to mRNA-based COVID-19 vaccine in patients with immune

thrombocytopenia. Br J Haematol 28: https://doi.org/10.1111/ bjh.18578

- Riester E, Findeisen P, Hegel JK et al (2021) Performance evaluation of the Roche Elecsys Anti-SARS-CoV-2 S immunoassay. J Virol Methods 297:114271. https://doi.org/10.1016/j.jviromet. 2021.114271
- Kanda Y (2013) Investigation of the freely available easy-to-use software "EZR" for medical statistics. Bone Marrow Transplant 48:452–458. https://doi.org/10.1038/bmt.2012.244
- Gagelmann N, Passamonti F, Wolschke C et al (2022) Antibody response after vaccination against SARS-CoV-2 in adults with hematological malignancies: a systematic review and meta-analysis. Haematologica 107:1840–1849. https://doi.org/10.3324/ haematol.2021.280163
- Marasco V, Carniti C, Guidetti A et al (2022) T cell immune response after mRNA SARS-CoV-2 vaccines is frequently detected also in the absence of seroconversion in patients with lymphoid malignancies. Br J Haematol 196:548–558. https://doi. org/10.1111/bjh.17877

- Levin MJ, Ustianowski A, Wit SD et al (2022) Intramuscular AZD7442 (tixagevimab-cilgavimab) for prevention of COVID-19. N Engl J Med 386:2188–2200. https://doi.org/10.1056/NEJMoa2116620
- Bergwerk M, Gonen T, Lustig Y et al (2021) COVID-19 breakthrough infections in vaccinated health care workers. N Engl J Med 385:1474–1484. https://doi.org/10.1056/NEJMoa2109072
- Gilbert PB, Montefiori DC, McDermott AB et al (2022) Immune correlates analysis of the mRNA-1273 COVID-19 vaccine efficacy clinical trial. Science 375:43–50. https://doi.org/10.1126/science.abm3425
- Bonifacius A, Tischer-Zimmermann S, Dragon AC et al (2021) COVID-19 immune signatures reveal stable antiviral T cell function despite declining humoral responses. Immunity 54:340–35. e46. https://doi.org/10.1016/j.immuni.2021.01.008

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Authors and Affiliations

Mirei Kobayashi¹ · Akio Mori¹ · Masahiro Onozawa² · Shihori Tsukamoto¹ · Hajime Senjo^{1,2} · Takashi Ishio¹ · Emi Yokoyama¹ · Minoru Kanaya¹ · Koh Izumiyama¹ · Makoto Saito¹ · Haruna Muraki^{3,4} · Masanobu Morioka¹ · Takanori Teshima² · Takeshi Kondo¹

Mirei Kobayashi miir0731@gmail.com

- ³ Division of Laboratory, Aiiku Hospital, Sapporo, Japan
- ⁴ Sapporo Clinical Laboratory Inc, Sapporo, Japan
- ¹ Blood Disorders Center, Aiiku Hospital, Sapporo, Japan
- ² Department of Hematology, Faculty of Medicine, Hokkaido University, Sapporo, Japan