INVITED ARTICLE



REVIEWS OF ANTI-INFECTIVE AGENTS: Louis D. Saravolatz, Section Editor

Cefiderocol: A New Cephalosporin Stratagem Against Multidrug-Resistant Gram-Negative Bacteria

Sharon Ong'uti,¹ Mary Czech,² Elizabeth Robilotti,³ and Marisa Holubar^{4,0}

¹Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ²Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ³ Memorial Sloan Kettering Cancer Center, New York, New York, USA; and ⁴Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ³ Memorial Sloan Kettering Cancer Center, New York, New York, USA; and ⁴Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ³ Memorial Sloan Kettering Cancer Center, New York, New York, USA; and ⁴Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ⁴ Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA; ⁴ Division of Infectious Diseases and Geographic Medicine, Stanford, California, USA; ⁴ Division of Infectious Diseases and Geographic Medicine, Stanford, California, USA; ⁴ Division of Medicine, Stanford, California, USA; ⁴ Division of Infectious Diseases and Geographic Medicine, Stanford, California, USA; ⁴ Division of Medicine, Stanford, California, USA; ⁴ D

Cefiderocol is a novel injectable siderophore cephalosporin that hijacks the bacterial iron transport machinery to facilitate cell entry and achieve high periplasmic concentrations. It has broad in vitro activity against gram-negative bacteria, including multidrug-resistant (MDR) organisms such as carbapenem-resistant *Enterobacterales*, carbapenem-resistant *Pseudomonas aeruginosa*, and *Acinetobacter baumannii*. It was approved by the US Food and Drug Administration for the treatment of complicated urinary tract infections and nosocomial pneumonia based on clinical trials that demonstrated noninferiority to comparators. In this review, we summarize the available in vitro and clinical data, including recent evidence from 2 phase 3 clinical trials (APEKS-NP and CREDIBLE-CR), and discuss the place of cefiderocol in the clinician's armamentarium against MDR gram-negative infections.

Keywords. cefiderocol; multidrug resistant; gram-negative bacteria; carbapenem resistant *Enterobacterales*; extended spectrum beta-lactamase producing (ESBL).

The World Health Organization has declared antimicrobial resistance a top global public health threat [1]. The Centers for Disease Control and Prevention estimates that at least 2.8 million infections due to multidrug-resistant organisms (MDROs) occur in the United States annually, resulting in more than 35 000 deaths [2]. Treatment choices are limited for MDRO infections, particularly for those caused by carbapenem-resistant Enterobacterales, extended-spectrum beta-lactamase-producing Enterobacterales, carbapenemresistant Pseudomonas aeruginosa, and Acinetobacter baumannii. Carbapenem-sparing antibiotics are critically needed as resistance to this antibiotic class is increasingly common. Cefiderocol is a novel injectable siderophore cephalosporin with broad-spectrum in vitro activity that was specifically developed as a treatment option for challenging MDRO infections [3].

CHEMISTRY AND MECHANISM OF ACTION

Cefiderocol contains a cephalosporin core with 2 side chains similar to those of ceftazidime and cefepime (Figure 1)

Clinical Infectious Diseases® 2022;74(7):1303–12

[4–6]. The C-7 side chain has an aminothiazole ring and a carboxypropyl-oxyimino group (Figure 1), both of which improve transport across the outer membrane of gram-negative bacteria and confer stability against hydrolysis by several beta-lactamases. The C-3 side chain has a pyrrolidinium group that enhances its water solubility by conferring zwitterionic properties.

The catechol moiety on the C-3 position distinguishes cefiderocol from cefepime and ceftazidime and functions as a siderophore, which chelates extracellular iron, forming a cefiderocol-ferric complex. As a consequence, while cefiderocol, like other beta-lactams, transits the outer cell membrane by passive diffusion through porins, it is also actively transported into organism by its iron uptake system [4, 5]. Once cefiderocol is transported into the periplasmic space, it dissociates from the iron and binds penicillin-binding proteins (PBP), primarily PBP3, to inhibit peptidoglycan synthesis, causing cell death.

The potent activity of cefiderocol against MDR gram-negative bacteria is due, at least in part, to its ability to overcome resistance caused by porin changes and achieve high concentrations in the periplasmic space via its active transport. Other factors include its high affinity for PBP3, its relative indifference to increased efflux pump expression [4, 7], and its resistance to hydrolysis by many beta-lactamases, including most serine carbapenemases and some metallo beta-lactamases, likely due to its C-3 side chain [4, 8]. Cefiderocol lacks significant activity against gram-positive organisms and anaerobes (the latter of which have lesser dependence on siderophore-iron transport systems for growth) [7, 9].

Received 16 February 2021; editorial decision 30 August 2021; published online 7 September 2021.

Correspondence: S. Onguti, Stanford University School of Medicine, Division of Infectious Diseases and Geographic Medicine, 300 Pasteur Drive, Room L-134, Stanford, CA 94305-5105 (onguti@stanford.edu).

[©] The Author(s) 2021. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com. https://doi.org/10.1093/cid/ciab757



Figure 1. Cefiderocol's structural components responsible for antibacterial activity. *Image source: https://en.wikipedia.org/wiki/Cefiderocol#/media/File:Cefiderocol. svg. I, the copyright holder of this work, release this work into the public domain. This applies worldwide. In some countries this may not be legally possible; if so: I grant anyone the right to use this work for any purpose, without any conditions, unless such conditions are required by law.

ANTIMICROBIAL ACTIVITY IN VITRO

The proposed cefiderocol minimum inhibitory concentration (MIC) breakpoints per the Clinical and Laboratory Standards Institute (CLSI), US Food and Drug Administration (FDA), and European Committee on Antimicrobial Susceptibility Testing (EUCAST) are listed in Table 1. CLSI published investigational MIC breakpoints for research purposes. The FDA released more conservative breakpoints when it first approved cefiderocol for complicated urinary tract infections (cUTIs) in November 2019; these were subsequently revised in September 2020 based on clinical data from the CREDIBLE-CR [10] and APEKS-NP [11] trials. A CLSI breakpoint update is anticipated in 2021; some advocate for use of FDA and EUCAST breakpoints for clinical care in the interim [9].

Two large studies, SIDERO 2014 [12] and SIDERO 2015 [13], established that cefiderocol exhibits potent in vitro activity against a broad spectrum of gram-negative bacilli, including

meropenem-nonsusceptible isolates of the *Enterobacteriaceae* family, *P. aeruginosa*, and *A. baumannii*. Cumulatively, more than 99% of the more than 18 000 gram-negative bacilli isolates tested showed a cefiderocol MIC <4 μ g/mL. In addition, SIDERO 2015 [13] further demonstrated that cefiderocol retained potent in vitro activity against both *Enterobacteriaceae* and *Pseudomonas* isolates that were nonsusceptible to ceftazidime-avibactam and ceftolozone-tazobactam, all with MIC_{ap} values <4 μ g/mL.

Cefiderocol additionally exhibits in vitro activity against *Stenotrophomonas maltophilia*, despite its intrinsic beta-lactam resistance (L1 metallo-beta-lactamase and L2 serine-beta-lactamase) [12, 13]. A limited amount of data suggest that cefiderocol may be active against some *Burkholderia cepacia* isolates [12, 13].

Cefiderocol Resistance Stratified by Beta-Lactamases

Cefiderocol's in vitro activity against MDR *Enterobacterales*, *P. aeruginosa*, and *A. baumannii* has been further characterized according to the Ambler classification of beta-lactamases. Within the *Enterobacterales* order, resistance did not differ by species [14]. Overall, cefiderocol retains potent activity against *Enterobacterales*, *Pseudomonas*, and *Acinetobacter* isolates that produce serine-beta-lactamases, cephalosporinases, and oxacillinases. However, although cefiderocol retains activity in the presence of most metallo-beta-lactamases, in vitro data are concerning for reduced potency against New Delhi Metallo- β -Lactamase (NDM)-producing isolates (Table 2).

Data describing cefiderocol's activity in the presence of NDM enzymes are primarily limited to isolates from the *Enterobacterales* order (Table 2). In these studies, cefiderocol MICs were variable, and a substantial portion of these NDM-producing isolates were cefiderocol-nonsusceptible [12, 14–16] (Table 2). Similarly, more limited data of NDM-producing *Pseudomonas* and *Acinetobacter* isolates demonstrate variable in vitro cefiderocol activity with wide MIC ranges (Table 2). These data suggest that susceptibility testing will be important

Organism	Clinical ards In: tional I	and Laborato stitute MIC I Breakpoints	ory Stand- nvestiga- (ug/mL) ^a	US Foo tion M	od and Drug Ac	lministra- (ua/mL) ^b	European Committee on Anti- microbial Susceptibility Testing MIC Breakpoints (ug/mL) ^c		
	S	1	R	S	1	R	S		R
Enterobacterales	<4	8	>16	<4	8	>16	<2		>2
Pseudomonas aeruginosa	<4	8	>16	<1	2	>4	<2	-	>2
Acinetobacter baumannii	<4	8	>16	<1	2	>4	NA	NA	NA
Stenotrophomonas maltophilia	<4	8	>16	NA	NA	NA	NA	NA	NA

Table 1. Cefiderocol Minimum Inhibitory Concentration Breakpoints Per the Clinical and Laboratory Standards Institute, US Food and Drug Administration, and European Committee on Antimicrobial Susceptibility Testing

Investigational.

MICs are determined by broth microdilution using iron-depleted and cation-adjusted Mueller-Hinton broth.

Abbreviations: I, Intermediate; MIC, minimum inhibitory concentration; NA, Not applicable; R, Resistant; S, Sensitive

^aClinical and Laboratory Standards Institute (CLSI) investigational MIC breakpoints are intended for research purposes and should not be used for routine clinical care purposes. CLSI breakpoints published June 2018; update anticipated 2021.

^bUS Food and Drug Administration breakpoints updated September 2020.

^cEuropean Committee on Antimicrobial Susceptibility Testing breakpoints published April 2020

Table 2. In Vitro Cefiderocol Activity Against Gram-Negative Bacilli With Differing Resistance Mechanisms

Reference	Resistance Mechanism	Isolate Number	Range	MIC ₅₀	MIC ₉₀
Enterobacterales					
	Ambler class A – serine-beta-lactamases				
Kohira et al [16]	ESBL	92	<0.125–4	0.125	0.5
Mushtaq et al [14]*	ESBL + porin loss	26	0.125–32	2	8
Kazmierczak et al [12]	KPC	75	0.03–4	1	2
Kohira et al [16]	KPC	47	<0.125-4	0.125	0.5
Mushtaq et al [14]	KPC	56	<0.03-8	0.25	2
Jacobs et al [15]*	KPC-2	355	<0.03–32	1	8
Delgado-Valverde et al [39]	ST512/KPC-3	25	0.25–4	2	4
Delgado-Valverde et al [39]	ST258/KPC-3	25	0.06–4	2	2
Jacobs et al [15]	KPC-3	380	<0.03–64	0.25	2
	Ambler class B – metallo-beta-lactamases				
Kazmierczak et al [12]	VIM	27	0.12-4	1	4
Kohira et al [16]	VIM	12	<0.125–16	0.125	0.25
Mushtaq et al [14]	VIM	47	<0.03–8	0.5	4
Kanazawa et al [40]	IMP-6	82	<0.03-2	0.06	1
Kohira et al [16]	IMP	8	<0.125–16	-	-
Mushtaq et al [14]	IMP	15	<0.03–4	0.25	2
Kazmierczak et al [12]*	NDM	12	1–8	4	8
Kohira et al [16]*	NDM	49	<0.125->16	1	16
Jacobs et al [15]*	NDM	28	0.25->64	2	8
Mushtaq et al [14]*	NDM	61	0.25–32	4	8
	Ambler class C – cephalosporinases				
Mushtaq et al [14]	AmpC + porin loss	25	0.06-2	0.5	2
	Ambler class D – oxacillinases				
Delgado-Valverde et al [39]	OXA-48	3	0.06-0.5	-	-
Kazmierczak et al [12]	OXA-48-like	32	0.03–4	0.5	4
Mushtaq et al [14]	OXA-48-like	56	<0.03-8	0.25	2
Jacobs et al [15]	OXA-48-like	7	<0.03–1	0.25	1
	Combination resistance mechanisms				
Delgado-Valverde et al [39]	ST11/OXA-48 + CTX-M-15	25	<0.03-4	0.25	2
Delgado-Valverde et al [39]	ST15/OXA-48 + CTX-M-15	25	<0.03–4	0.25	4
Delgado-Valverde et al [39]	ST392/OXA-48 + CTX-M-15	4	0.06-1	-	-
Kazmierczak et al [12]	Carbapenemase negative, meropenem resistant	13	0.008–4	0.12	2
Pseudomonas aeruginosa					
	Ambler class A – serine-beta-lactamases				
Kazmierczak et al [12]	GES	4	0.12-0.25	-	-
Mushtaq et al [14]	GES	20	0.06–4	0.25	2
Mushtaq et al [14]*	PER	15	0.06–16	1	16
Mushtaq et al [14]	VEB	10	0.5–8	-	-
	Ambler class B – metallo-beta-lactamases				
Kazmierczak et al [12]	VIM	26	0.008–2	0.25	2
Mushtaq et al [14]	VIM	30	<0.03->128	0.25	1
Kazmierczak et al [12]	IMP	4	1–2	-	-
Mushtaq et al [14]*	IMP	25	0.06–16	0.25	8
Mushtaq et al [14]*	NDM	11	1->128	4	16
Kazmierczak et al [12]	Carbapenemase-negative, meropenem non-susceptible	319	<0.002-4	0.12	0.5
Jacobs et al [15]	Carbapenem-resistant NOS	27	<0.03-1	0.25	0.5
Acinetobacter baumannii					
	Ambler class A – serine-beta-lactamases				
Kazmierczak et al [12]	GES-type	7	0.25–8	-	-
	Ambler class B – metallo-beta-lactamases				
Kazmierczak et al [12]	NDM	2	1–1	-	-
Mushtaq et al [14]*	NDM	20	1->128	2	16
	Ambler class D – oxacillinases				
Delgado-Valverde et al [39]	OXA-23	25	0.06-1	0.25	0.5
Kazmierczak et al [12]	OXA-23	543	<0.002-16	0.12	1

Table 2. Continued

Reference	Resistance Mechanism	Isolate Number	Range	MIC ₅₀	MIC ₉₀
Mushtaq et al [14]*	OXA-23	41	0.06->128	0.25	16
Delgado-Valverde et al [39]*	OXA-24	25	0.5–16	2	16
Kazmierczak et al [12]	OXA-24	124	0.004–64	0.12	1
Mushtaq et al [14]	OXA-51	19	0.06–16	0.125	0.5
Kazmierczak et al [12]	OXA-58	14	0.06–1	0.06	1
Delgado-Valverde et al [39]	ST2/OXA-58	25	0.06-0.5	0.125	0.5
Delgado-Valverde et al [39]	ST745/OXA-58	5	0.06-0.25	-	-
Mushtaq et al [14]	OXA-58	10	0.06->128	-	-
Mushtaq et al [14]	OXA-24/40	9	0.25–4		
Kazmierczak et al [12]*	Carbapenemase-negative, meropenem non-susceptible	86	0.008-8	0.25	2
Jacobs et al [15]	Carbapenem-resistant NOS	101	<0.03->63	0.25	1

MICs are determined by broth microdilution using iron-depleted and cation-adjusted Mueller-Hinton broth. MIC₅₀, MIC at which cefiderocol inhibits growth of 50% of tested isolates.

MIC₉₀ MIC at which cefiderocol inhibits growth of 90% of tested isolates. MIC₅₀ and MIC₉₀ not determined if <10 isolates. *MIC 90 exceeding either FDA or EUCAST breakpoint, whichever is higher.

Abbreviations: CTX-M, group of class A extended-spectrum β-lactamases; ESBL, extended-spectrum beta-lactamase; GES, Guiana extended spectrum β-lactamase; IMP, imipenemhydrolyzing β-lactamase; KPC, *Klebsiella pneumoniae* carbapenemase; MIC, minimum inhibitory concentration; NDM, New Delhi metallo β-lactamase; NOS, not otherwise specified; OXA, oxacillin-hydrolyzing β-lactamases; PER: extended-spectrum β-lactamase first discovered in strains of *P. aeruginosa*; ST, sequence type; VEB, Vietnamese extended-spectrum β-lactamase; VIM, Verona integron-encoded metallo-β-lactamase.

before use of cefiderocol as a single agent to treat infections due to NDM-producing organisms.

In vitro testing data pooled from 6 studies suggest cefiderocol does retain activity against most *Enterobacterales*, *Pseudomonas*, *and Acinetobacter* isolates producing non-NDM beta-lactamase enzymes (Table 2). However, there are reports of reduced cefiderocol potency without a clear pattern across Ambler class or beta-lactamase enzyme type. For instance, among *Enterobacterales* isolates producing serine-beta-lactamases, there is a wide distribution of MIC ranges. Furthermore, a large study showed variable cefiderocol potency against *Enterobacterales* within and between differing *Klebsiella pneumoniae* carbapenemase (KPC) enzyme subtypes (Table 2) [15]. These inconsistent resistance patterns within and among beta-lactamases suggest cefiderocol resistance is not exclusively determined by the presence of beta-lactamase enzymes.

Despite limited data, studies have started to shed light on mechanisms of cefiderocol resistance. Although it seems cefiderocol resistance is not caused by the presence of betalactamases alone (potential exception being NDM-producing organisms), some studies suggest that cefiderocol activity may be potentiated by the addition of beta-lactamase inhibitors [14, 17]. These data suggest cefiderocol resistance is likely mediated by a combination of resistance mechanisms, and the addition of beta-lactamase inhibitors may be sufficient to restore the drug's activity against some of these mechanisms. Other proposed mechanisms of cefiderocol resistance include mutations in iron transport channels, which inhibit cefiderocol's novel mechanism of entry into bacteria [7]. Unlike other gram-negative bacilli, cefiderocol resistance does not seem to be mediated by porin and efflux mutations [7]. To further elucidate nuances of cefiderocol resistance, future studies should pay particular attention to NDM enzymes and combinations of resistance mechanisms, including beta-lactamases and mutations in iron transport.

Considerations for Clinical Laboratory In Vitro Testing

Both broth microdilution (using iron-depleted media) and agar disc diffusion methods have been approved and validated for evaluation of cefiderocol's in vitro activity. Broth microdilution allows for determination of MICs, whereas agar disc diffusion does not allow precise determination of the amount of antimicrobial agent diffused into agar. However, agar disc diffusion is likely the most practical method for clinical laboratories to adopt until cefiderocol testing can be included on commercially available automated antimicrobial susceptibility testing panels [9].

PHARMACOKINETICS AND PHARMACODYNAMICS

Animal Studies

Similar to other cephalosporins, the activity of cefiderocol is best described by time-dependent killing [18], which is enhanced when cefiderocol is administered as an extended 3-hour infusion compared with a 1-hour infusion [19]. Humanized exposures of cefiderocol in murine thighs (dosed at an equivalent of 2 g intravenous every 8 hours, 3-hour infusion) induced bacterial stasis or >1 log₁₀ reduction in bacterial colony-forming units in the majority of gram-negative bacilli with a MIC $<4 \mu g/mL$; however, the probability of target attainment was significantly reduced against isolates with MIC >8 μ g/mL [20, 21]. These in vivo studies demonstrate the effectiveness of cefiderocol and support consideration of an MIC of 4 µg/mL as the susceptibility breakpoint for Enterobacteriaceae, Pseudomonas, Acinetobacter, and Stenotrophomonas. Additionally, in vivo studies show repeat doses of cefiderocol were not associated with the development of resistance [22], but results of these animal studies need correlation with clinical trial data.

Human Studies

Cefiderocol displays linear kinetics. It is primarily renally excreted and does not undergo significant hepatic metabolism

Table 3. Cefiderocol Dose Regimens Based on Renal Function

Renal Function	Renal Function Estimates	Dose Regimen
Augmented ^a	CG-CL _{cr} , >120 mL/min	2 g every 6 hours, 3-hour infusion
Normal	MDRD-eGFR, 90 to <120 mL/min/1.73 m^2	2 g every 8 hours, 3-hour infusion
Mild impairment	MDRD-eGFR, 60 to <90 mL/min/1.73 m^2	2 g every 8 hours, 3-hour infusion
Moderate impairment	MDRD-eGFR, 30 to <60 mL/min/1.73 m^2	1.5 g every 8 hours, 3-hour infusion
Severe impairment	MDRD-eGFR, 15 to <30 mL/min/1.73 m^2	1 g every 8 hours, 3-hour infusion
End-stage renal disease	MDRD-eGFR, <15 mL/min/1.73 m ²	0.75 g every 12 hours, 3-hour infusion
Requiring intermittent hemodialysis		0.75 g every 12 hours, 3-hour infusion, with consideration for supplemental (third) dose of 0.75 g administered after hemodialysis on dialysis days [25]
Requiring continuous renal	Dosed based on effluent flow rate	
replacement therapy	≤2 L/hour	1.5 g every 12 hours
	2.1 to 3 L/hour	2 g every 12 hours
	3.1 to 4 L/hour	1.5 g every 8 hours
	≥4.1 L/hour	2 g every 8 hours

Abbreviations: CG-CL_{CR}, Cockcroft-Gault creatinine clearance; eGFR, estimated glomerular filtration rate; MDRD, Modification of Diet in Renal Disease. ^aPatients with hypermetabolic states due to sepsis [31].

[23, 24]. Dose adjustments are required for renal impairment but not for hepatic impairment [25].

Early human pharmacokinetic studies support the use of cefiderocol for treatment of pulmonary, urinary tract, and possibly bloodstream infections. Phase 1 studies show cefiderocol achieves intrapulmonary concentrations similar to plasma concentrations [26]. Phase 2 studies in cUTI and uncomplicated pyelonephritis show modestly increased drug clearance with infection, but plasma levels of cefiderocol remained adequate to treat cUTIs [27]. The largest study used plasma cefiderocol concentrations collected from 516 patients, including healthy volunteers and those enrolled in clinical trials [10, 11, 28], to develop a population pharmacokinetics model. Renal function was the most significant variable in the model. Analysis using the model showed a >95% probability of target attainment (75% time above MIC) for gram-negative isolates with MICs <4 regardless of infection site. The model predicted a >90% probability of target attainment (100% time above MIC) for isolates with MICs <4 for pneumonia and cUTI but dropped to 85% for bloodstream infection (BSI) [29]. No data are yet available on cerebrospinal fluid penetration.

DOSAGE AND ADMINISTRATION

Recommended cefiderocol dosing for patients with normal renal function is 2 g administered intravenously over 3 hours every 8 hours. Dosing recommendations based on renal function are described in Table 3. Intermittent hemodialysis can remove up to 60% of cefiderocol, so there may be a role for administering a supplemental dose of cefiderocol following hemodialysis [25, 30, 31]. Presently, there are no recommendations for weight-based dosing in obesity and no data regarding cerebral spinal fluid penetration. With these sophisticated dosing regimens and minimal pharmacokinetic data in disease states, therapeutic drug monitoring may be helpful.

CLINICAL EXPERIENCE

Complicated Urinary Tract Infection

The APEKS-cUTI study [28] was a phase 2, multicenter, double-blind, parallel-group noninferiority trial that evaluated cefiderocol vs imipenem-cilastatin for the treatment of cUTI. Patients were randomized to receive 1-hour intravenous infusions of cefiderocol (2 g) or imipenem-cilastatin (1 g) every 8 hours for 7–14 days. The primary efficacy end point was the composite of clinical and microbiological response 7 days (±2 days) after the end of antibiotic treatment.

A total of 452 patients were randomized to receive cefiderocol (n = 303) and imipenem-cilastatin (n = 149). Of these, 252 in the cefiderocol group and 119 in the imipenem-cilastatin group were included in the modified intention-to-treat (mITT) population. The most common baseline uropathogens were Escherichia coli and Klebsiella pneumoniae, and close to half were pan-susceptible (cefiderocol n = 117 of 252, 46%; imipenem-cilastatin n = 60 of 119, 50%.) Cefiderocol was deemed noninferior to impenem-cilastatin as the primary efficacy end point was achieved by 73% (n = 183) in the cefiderocol group and 55%(n = 65) in the imipenem-cilastatin group with an adjusted treatment difference of 18.58% (95% confidence interval [CI], 8.23% to 28.92%). The clinical response rates were similar between the treatment groups (the preset lower bound to conclude noninferiority was 15%). However, microbiological response at test of cure was higher in the cefiderocol arm (73% vs 56%; difference 17.25%; 6.92% to 27.58%). Sensitivity analyses of composite and microbiological outcomes were consistent with those of the mITT cohort.

Nosocomial Pneumonia

The APEKS-NP study [11] was a phase 3, randomized, double-blind, multicenter, noninferiority trial that compared cefiderocol vs high-dose, extended infusion meropenem for

adults with hospital-acquired pneumonia (HAP), ventilatorassociated pneumonia (VAP), or healthcare-associated pneumonia (HCAP) due to gram-negative pathogens. Patients were randomized 1:1 to receive 3-hour intravenous infusions of either cefiderocol 2 g or meropenem 2 g every 8 hours for 7–14 days. All patients also received open-label intravenous linezolid (600 mg every 12 hours) for at least 5 days. The primary end point was all-cause mortality at day 14 in the mITT population.

A total of 300 patients were randomized to receive cefiderocol (n = 148) or meropenem (n = 152). Of these, 145 in the cefiderocol group and 147 in the meropenem group were included in the mITT population (all patients who received at least 1 study drug dose, excluding those with gram-positive monomicrobial infections). Cefiderocol was noninferior to high-dose extended infusion meropenem as the primary outcome (all-cause mortality at day 14) was similar in the 2 groups (12.4% cefiderocol vs 11.6% meropenem; adjusted difference, 0.8%; 95% CI, -6.6% to 8.2%) based on a 12.5% noninferiority margin. The proportion of patients with clinical cure and microbiological eradication at test of cure was similar in both groups. The most common pathogen was K. pneumoniae followed by P. aeruginosa and A. baumannii. Predefined subgroup analyses showed more deaths in the cefiderocol arm in patients with HCAP (9 vs 2 deaths), but mortality was similar in both arms in all other subgroups. Sensitivity analyses for the all-cause mortality at days 14 and 28 in the mITT population were similar to the analyses of the microbiologically evaluable per-protocol population (all patients in the mITT population who followed the study protocol and had a positive culture for gram-negative bacilli).

Severe Carbapenem-Resistant Infections

The CREDIBLE-CR study [10] differs from the previous 2 studies in its design and study population. This phase 3, randomized, open-label, multicenter, descriptive study assessed cefiderocol vs clinician-directed best available therapy (BAT) in adults with serious carbapenem-resistant (CR) gram-negative infections. Investigators included patients hospitalized with nosocomial pneumonia (HAP, VAP, HCAP), BSI, cUTI, or sepsis not otherwise specified (NOS) caused by a suspected or proven carbapenem-resistant gram-negative pathogen. Participants were assigned 2:1 to receive either a 3-hour intravenous infusion of cefiderocol 2 g every 8 hours or BAT for an expected 7-14 days (could be extended to 21 days per clinician discretion). For patients with nosocomial pneumonia, BSI, or sepsis NOS, the primary end point was clinical cure after treatment completion (7 days \pm 2 days after end of therapy). The primary end point for patients with cUTI was microbiological eradication at test of cure.

A total of 152 patients were randomized to receive cefiderocol (n = 101) or BAT (n = 51). The most common diagnosis upon

enrollment was nosocomial pneumonia (n = 67, 45%) followed by BSI/sepsis NOS (n = 47, 31%) and cUTI (n = 36, 24%). The primary analysis included 118 patients (cefiderocol n = 80, BAT n = 38) who had a confirmed carbapenem-resistant infection. In the cefiderocol group, 83% (66 of 80) of patients received monotherapy, while in the BAT arm, 71% (27 of 38) received combination therapy (the majority of which were colistin-based regimens). The most common carbapenemresistant pathogens were *A. baumannii*, *K. pneumoniae*, and *P. aeruginosa* (cefiderocol MIC₉₀ of 1 µg/mL, 4 µg/mL, and 2 µg/ mL, respectively).

Clinical cure at test of cure was similar in each group for those with nosocomial pneumonia (cefiderocol 50% [20 of 40], 95% CI, 33.8% to 66.2%; BAT 53% [10 of 19], 95% CI, 28.9% to 75.6%) and BSI or sepsis NOS (cefiderocol 43% [10 of 23], 95% CI, 23.2% to 65.5%; BAT 43% [6 of 14], 95% CI, 17.7% to 71.1%). For patients with cUTI, microbiological eradication at test of cure was 53% (9 of 17, 95% CI, 27.8% to 77.0%) in the cefiderocol group and 20% (1 of 5, 95% CI, 0.5% to 71.6%) in the BAT group.

More patients died in the cefiderocol arm compared with those treated with BAT (33.7% [34 of 101] vs 18.3% [9 of 49]). Post hoc all-cause mortality in the cefiderocol arm was higher at day 28 (difference 6.4%; 95% CI, -8.6% to 19.2%) and at day 49 (difference 13.3%; 95% CI, -2.5% to 27.8%). Exploratory analyses also found that the mortality rate differed by underlying infection and infecting organism. A study adjudication committee attributed more deaths in the cefiderocol arm to treatment failure compared with the BAT arm (15.8% [16 of 101] vs 8.2% [4 of 49]), most of which occurred within 15 days of study initiation [32]; the remainder of the deaths were attributed to underlying comorbidities. Most of the treatment failure deaths in the cefiderocol arm occurred in patients with Acinetobacter infections (13 of 16) compared with only 1 death (1 of 4) in the BAT arm. Fifteen patients who received cefiderocol had evidence suggesting treatment emergent in vitro resistance, with a 4-fold increase in cefiderocol MIC from baseline; 10 of these patients experienced treatment failure.

The FDA published its analysis of the CREDIBLE data [10] and described aspects of the study design that limit its interpretation, including the open-label design, small sample size, imbalance between study groups, and limited descriptive analytic plan [32]. Notably, FDA approval of cefiderocol is based solely on APEKS-cUTI [28] and APEKS-NP [11] data, which did not show increased mortality in patients who received cefiderocol except for the HCAP subgroup in the APEKS-NP trial. Regardless, the FDA recommends that clinicians closely monitor patients who receive cefiderocol for evidence of treatment failure. Of note, the European Medical Agency, the European Union's pharmaceutical governing body, has issued a pathogenfocused approval based, in part, on CREDIBLE trial data, authorizing cefiderocol use for aerobic gram-negative infections in patients with limited treatment options [3, 33].

	APEK	S-NP, %	APE	KS-cUTI, %	CRE	DIBLE-CR, %
Adverse Event	Cefiderocol (n = 148)	Meropenem (n = 150)	Cefiderocol (n = 300)	lmipenem-cilastam (n = 148)	Cefiderocol ($n = 101$)	Best Available Therapy (n = 49)
All TEAEs	88	86	41	51	91	96
Mild	22	25	NR	NR	23	18
Moderate	28	31	NR	NR	26	33
Severe	38	30	NR	NR	43	45
TEAEs	Reported >5% in either	group	Reported >2% in either	group	Reported >5% in either	group
Diarrhea	0	0	4	6	19	12
Constipation	Q	4	e	4	ω	6
Nausea	NR	NR	2	4	7	4
Vomiting	NR	NR	2	1	13	14
Headache	NR	NR	2	5	NR	NR
Infusion site pain	NR	NR	e	ო	NR	NR
Rash	1	0	NR	NR	т	00
Hypokalemia	11	15	2	ო	ი	14
Elevated LFTs	2.7	6.7			ω	2
TEAE leading to drug discontinuation	ω	6	2	2	e	4
Drug-related TEAEs	Q	11	6	11	15	22
Serious adverse events	36	30	5	8	50	47
Clostridium difficile-associated disease	3	3	<1	З	-	0
Numbers of patients included in the safety analy	/sis of each study.					

Table 4. Adverse Events Reported in Clinical Trials

Abbreviations: LFT, Liver function test; NR, not reported; TEAE, treatment emergent adverse events.

		וו טו טושים-חפאנ						
				In Vitro Activity				
		Enterobacter	ales					
Agent	Ambler Class A	Ambler Class B	Ambler Class D	Pseudomonas aeruginosa	Acinetobacter baumannii	Stenotrophomonas maltophilia	US Food and Drug Adminis- tration-Approved Indications	Daily Cost for Normal Renal Function
Cefiderocol	Yes	Yes	Partial	Yes	Yes	Yes	cUTI/AP, HABP/VABP	\$1320.00
Ceftazidime-avibactam	Yes	No	Yes	Yes	No	No	cUTI/AP, cIAI, HABP/VABP	\$1291.71
Ceftolozane-tazobactam	No	No	No	Yes	No	No	cUTI/AP, cIAI	\$450.78
Meropenem-vaborbactam	Yes	No	No	Yes	No	No	cUTI/AP	\$1283.04
Imipenem-cilastatin-relebactam	Yes	No	No	Yes	No	No	cUTI/AP, cIAI	\$1322.56
Dosing: Cefiderocol: 2 g intravenous	(IV) every 8 hour:	s. Ceftazidime-avib	actam: 2.5 g (cefta.	zidime 2 g/avibactam 0.5 (g) IV every 8 hours. Ceftol	ozane-tazobactam: 1.5 g (ceftolo	zane 1 g/tazobactam 0.5 g) IV every 8 hours. Meropener	m-vaborbactam: 4 g

Abbreviations: AP, acute pyelonephritits; cIAI, complicated intraabdominal inflection; cUTI, complicated urinary tract inflection; HABP, hospital-acquired bacterial pneuronia; VABP, ventilator-associated bacterial pneuronia (meropenem 2 g/vaborbactam 2 g) IV every 8 hours. Imipenem-cilastatin-relebactam: 1.25 g (imipenem 500 mg/cilastatin 500 mg/relebactam 250 mg) IV every 6 hours.

Cases Reports and Case Series

Limited case reports demonstrate variable success of off-label use of cefiderocol. Although CREDIBLE data are concerning for treatment failure with the use of cefiderocol for Carbapenemresistant (CR) Acinetobacter, case reports illustrate mixed success in treating extensively resistant Acinetobacter infections including orthopedic infections [34-37].

Safety and Adverse Events

Phase 1 studies in healthy humans demonstrated cefiderocol was safe and well tolerated [23]. In phase 2 and 3 clinical trials, adverse events were reported at similar rates in the cefiderocol arm and in the comparator arm, and the majority were mild or moderate in severity (Table 4). The most common adverse events reported across these trials were nausea, diarrhea, rash, elevated liver function tests, and hypokalemia. There were no reported laboratory abnormalities in these studies to suggest cefiderocol impacted patients' iron hemostasis.

The FDA approval includes a warning of an increased risk of all-cause mortality for treatment of carbapenem-resistant gram-negative bacterial infections based on data from the CREDIBLE trial (see above) [10]. It also includes warnings applicable to other beta-lactams including the risk of Clostridioides difficile-associated diarrhea, cephalosporin-associated risk of potentiating seizure activity (similar to cefepime), and hypersensitivity reactions. Cefiderocol's R1 side chain is exactly the same as that of ceftazidime and aztreonam and is similar to that of ceftaroline. These similarities confer potential cross-reactivity of allergic reactions, and clinicians should be especially cognizant of these similarities when prescribing alternate betalactams in patients with allergies [38].

Place in Therapy and Formulary Considerations

Cefiderocol's role in treating MDR gram-negative infections is still uncertain. Although its in vitro spectrum of activity is promising, cefiderocol's clinical trial performance thus far has only demonstrated results similar to comparators (namely, carbapenems) at a greater cost (Table 5). Although unexplained, the increased mortality observed in the CREDIBLE trial casts doubt on cefiderocol's role in treating carbapenemresistant A. baumanii and perhaps other nonfermenting gram-negative bacilli infections unless no other viable option exists. The lack of official CLSI breakpoints and challenges in susceptibility testing also hinder its clinical use. Until more data become available, cefiderocol will most likely be reserved for second-line or salvage therapy in selected MDR gram-negative infections.

CONCLUSIONS

Cefiderocol is a novel injectable siderophore cephalosporin with broad in vitro activity against gram-negative bacteria, including carbapenem-resistant pathogens. It is FDA approved for the treatment of cUTIs and nosocomial pneumonia, requires renally adjusted dosing, and appears to be fairly well tolerated. Early data suggest cefiderocol is a promising alternative agent for some carbapenem-resistant infections, but its role in the management of carbapenem-resistant *Pseudomonas* and *Acinetobacter* infections is uncertain. Pathogen-specific trials would help to refine cefiderocol's place in therapy.

Notes

Acknowledgments. The authors thank Dr Stan Deresinski for his careful review of this manuscript.

Potential conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest.

References

- World Health Organization. Ten threats to global health in 2019. 2019. Available at: https://www.who.int/news-room/spotlight/ten-threats-to-globalhealth-in-2019. Accessed 22 September 2021.
- Centers for Disease Control and Prevention. Antibiotic resistance threats in the United States 2019. 2019. Available at: https://www.cdc.gov/drugresistance/pdf/ threats-report/2019-ar-threats-report-508.pdf. Accessed 22 September 2021.
- Echols R, Ariyasu M, Nagata TD. Pathogen-focused clinical development to address unmet medical need: cefiderocol targeting carbapenem resistance. Clin Infect Dis 2019; 69:559–64.
- Sato T, Yamawaki K. Cefiderocol: discovery, chemistry, and in vivo profiles of a novel siderophore cephalosporin. Clin Infect Dis 2019; 69:538–43.
- Zhanel GG, Golden AR, Zelenitsky S, et al. Cefiderocol: a siderophore cephalosporin with activity against carbapenem-resistant and multidrug-resistant gram-negative bacilli. Drugs 2019; 79:271–89.
- Wu JY, Srinivas P, Pogue JM. Cefiderocol: a novel agent for the management of multidrug-resistant gram-negative organisms. Infect Dis Ther 2020; 9:17–40.
- Ito A, Sato T, Ota M, et al. In vitro antibacterial properties of cefiderocol, a novel siderophore cephalosporin, against gram-negative bacteria. Antimicrob Agents Chemother 2018; 62::e01454-17.
- Giacobbe DR, Ciacco E, Girmenia C, et al.; Italian Study Group on Resistant Infections of the Italian Society of Anti-infective Therapy. Evaluating cefiderocol in the treatment of multidrug-resistant gram-negative bacilli: a review of the emerging data. Infect Drug Resist 2020; 13:4697–711.
- Simner PJ, Patel R. Cefiderocol antimicrobial susceptibility testing considerations: the Achilles' heel of the trojan horse? J Clin Microbiol 2020; 59:e00951-20.
- Bassetti M, Echols R, Matsunaga Y, et al. Efficacy and safety of cefiderocol or best available therapy for the treatment of serious infections caused by carbapenemresistant gram-negative bacteria (CREDIBLE-CR): a randomised, open-label, multicentre, pathogen-focused, descriptive, phase 3 trial. Lancet Infect Dis 2020; 21:226–40.
- Wunderink RG, Matsunaga Y, Ariyasu M, et al. Cefiderocol versus high-dose, extended-infusion meropenem for the treatment of gram-negative nosocomial pneumonia (APEKS-NP): a randomised, double-blind, phase 3, non-inferiority trial. Lancet Infect Dis 2021; 21:213–25.
- Kazmierczak KM, Tsuji M, Wise MG, et al. In vitro activity of cefiderocol, a siderophore cephalosporin, against a recent collection of clinically relevant carbapenemnon-susceptible gram-negative bacilli, including serine carbapenemase- and metallo-β-lactamase-producing isolates (SIDERO-WT-2014 Study). Int J Antimicrob Agents 2019; 53:177–84.
- Karlowsky JA, Hackel MA, Tsuji M, Yamano Y, Echols R, Sahm DF. In vitro activity of cefiderocol, a siderophore cephalosporin, against gram-negative bacilli isolated by clinical laboratories in North America and Europe in 2015-2016: SIDERO-WT-2015. Int J Antimicrob Agents 2019; 53:456–66.
- Mushtaq S, Sadouki M, Vickers A, Livermore DM, Woodford N. In vitro activity of cefiderocol, a siderophore cephalosporin, against multidrug-resistant gram-negative bacteria. Antimicrob Agents Chemother 2020; 64:e01582-20.
- 15. Jacobs MR, Abdelhamed AM, Good CE, et al. ARGONAUT-I: activity of cefiderocol (S-649266), a siderophore cephalosporin, against gram-negative bacteria, including carbapenem-resistant nonfermenters and Enterobacteriaceae with defined extended-spectrum beta-lactamases and carbapenemases. Antimicrob Agents Chemother **2019**; 63:e01801-18.
- Kohira N, West J, Ito A, et al. In vitro antimicrobial activity of a siderophore cephalosporin, S-649266, against Enterobacteriaceae clinical isolates, including carbapenem-resistant strains. Antimicrob Agents Chemother 2016; 60:729–34.

- Kohira N, Hackel MA, Ishioka Y, et al. Reduced susceptibility mechanism to cefiderocol, a siderophore cephalosporin, among clinical isolates from a global surveillance programme (SIDERO-WT-2014). J Glob Antimicrob Resist 2020; 22:738–41.
- Nakamura R, Ito-Horiyama T, Takemura M, et al. In vivo pharmacodynamic study of cefiderocol, a novel parenteral siderophore cephalosporin, in murine thigh and lung infection models. Antimicrob Agents Chemother 2019; 63:e02031-18.
- Matsumoto S, Singley CM, Hoover J, et al. Efficacy of cefiderocol against carbapenem-resistant gram-negative bacilli in immunocompetent-rat respiratory tract infection models recreating human plasma pharmacokinetics. Antimicrob Agents Chemother 2017; 61:e00700-17.
- Monogue ML, Tsuji M, Yamano Y, Echols R, Nicolau DP. Efficacy of humanized exposures of cefiderocol (S-649266) against a diverse population of gram-negative bacteria in a murine thigh infection model. Antimicrob Agents Chemother 2017; 61:e01022-17.
- Chen IH, Kidd JM, Abdelraouf K, Nicolau DP. Comparative in vivo antibacterial activity of human-simulated exposures of cefiderocol and ceftazidime against *Stenotrophomonas maltophilia* in the murine thigh model. Antimicrob Agents Chemother **2019**; 63:e01558-19.
- 22. Stainton SM, Monogue ML, Tsuji M, Yamano Y, Echols R, Nicolau DP. Efficacy of humanized cefiderocol exposures over 72 hours against a diverse group of gram-negative isolates in the neutropenic murine thigh infection model. Antimicrob Agents Chemother 2019; 63:e01040-18.
- Saisho Y, Katsube T, White S, Fukase H, Shimada J. Pharmacokinetics, safety, and tolerability of cefiderocol, a novel siderophore cephalosporin for gram-negative bacteria, in healthy subjects. Antimicrob Agents Chemother 2018; 62:e02163-17.
- Miyazaki S, Katsube T, Shen H, Tomek C, Narukawa Y. Metabolism, excretion, and pharmacokinetics of [14 C]-cefiderocol (S-649266), a siderophore cephalosporin, in healthy subjects following intravenous administration. J Clin Pharmacol 2019; 59:958–67.
- Katsube T, Wajima T, Ishibashi T, Ferreira JCA, Echols R. Pharmacokinetic/pharmacodynamic modeling and simulation of cefiderocol, a parenteral siderophore cephalosporin, for dose adjustment based on renal function. Antimicrob Agents Chemother 2017; 61:e01381-16.
- Katsube T, Saisho Y, Shimada J, Furuie H. Intrapulmonary pharmacokinetics of cefiderocol, a novel siderophore cephalosporin, in healthy adult subjects. J Antimicrob Chemother 2019; 74:1971–4.
- 27. Kawaguchi N, Katsube T, Echols R, Wajima T. Population pharmacokinetic analysis of cefiderocol, a parenteral siderophore cephalosporin, in healthy subjects, subjects with various degrees of renal function, and patients with complicated urinary tract infection or acute uncomplicated pyelonephritis. Antimicrob Agents Chemother **2018**; 62:e01391-17.
- Portsmouth S, van Veenhuyzen D, Echols R, et al. Cefiderocol versus imipenemcilastatin for the treatment of complicated urinary tract infections caused by gram-negative uropathogens: a phase 2, randomised, double-blind, noninferiority trial. Lancet Infect Dis 2018; 18:1319–28.
- 29. Kawaguchi N, Katsube T, Echols R, Wajima T. Population pharmacokinetic and pharmacokinetic/pharmacodynamic analyses of cefiderocol, a parenteral siderophore cephalosporin, in patients with pneumonia, bloodstream infection/sepsis, or complicated urinary tract infection. Antimicrob Agents Chemother 2020; 65:e01437-20.
- Katsube T, Echols R, Arjona Ferreira JC, Krenz HK, Berg JK, Galloway C. Cefiderocol, a siderophore cephalosporin for gram-negative bacterial infections: pharmacokinetics and safety in subjects with renal impairment. J Clin Pharmacol 2017; 57:584–91.
- Fetroja (Cefiderocol). Osaka, Japan: Shionogi & Co., Ltd.. Available at: https:// www.accessdata.fda.gov/drugsatfda_docs/label/2019/209445s000lbl.pdf.
- Naseer S, Weinstein EA, Rubin DB, et al. US Food and Drug Administration (FDA): benefit-risk considerations for cefiderocol (Fetroja(R)). Clin Infect Dis 2020; 72:e1103-11.
- 33. Fetcroja. Summary of product characteristics [product information, labeling, package leaflet]. Shionogi & Co., Ltd. Available at: https://www.ema.europa.eu/en/documents/product-information/fetcroja-epar-product-information_en.pdf. Accessed 22 September 2021.
- 34. Falcone M, Tiseo G, Nicastro M, et al. Cefiderocol as rescue therapy for *Acinetobacter baumannii* and other carbapenem-resistant gram-negative infections in ICU patients. Clin Infect Dis 2020; 72:2021–4.
- 35. Oliva A, Ceccarelli G, De Angelis M, et al. Cefiderocol for compassionate use in the treatment of complicated infections caused by extensively and pan-resistant *Acinetobacter baumannii*. J Glob Antimicrob Resist 2020; 23:292–6.
- Dagher M, Ruffin F, Marshall S, et al. Case report: successful rescue therapy of extensively drug-resistant *Acinetobacter baumannii* osteomyelitis with cefiderocol. Open Forum Infect Dis 2020: 7:ofaa150.
- Zingg S, Nicoletti GJ, Kuster S, et al. Cefiderocol for extensively drug-resistant gram-negative bacterial infections: real-world experience from a case series and review of the literature. Open Forum Infect Dis 2020; 7:ofaa185.
- Chaudhry SB, Veve MP, Wagner JL. Cephalosporins: a focus on side chains and beta-lactam cross-reactivity. Pharmacy 2019; 7.

- Delgado-Valverde M, Conejo MDC, Serrano L, Fernández-Cuenca F, Pascual Á. Activity of cefiderocol against high-risk clones of multidrugresistant Enterobacterales, Acinetobacter baumannii, Pseudomonas aeruginosa and Stenotrophomonas maltophilia. J Antimicrob Chemother 2020; 75:1840–9.
- 40. Kanazawa S, Sato T, Kohira N, Ito-Horiyama T, Tsuji M, Yamano Y. Susceptibility of imipenem-susceptible but meropenem-resistant blaimp-6-carrying Enterobacteriaceae to various antibacterials, including the siderophore cephalosporin cefiderocol. Antimicrob Agents Chemother 2017; 61:e00576-17.