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A 68-Year-Old Man with a Cytogenetic Diagnosis of Chronic Myeloid Leukemia and Bone Marrow Findings of Philadelphia Chromosome Translocation Between the Long Arm of Chromosomes 9 and 22, Leading to the BCR-ABL1 Fusion Gene and V617F Mutation in the JAK2 Gene

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Data Collection B
Statistical Analysis C
Data Interpretation D
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Patient: Male, 68-year-old
Final Diagnosis: CML
Symptoms: Leukocytosis • splenomegaly
Clinical Procedure: Bone marrow biopsy
Specialty: Hematology
Objective: Rare disease
Background: Breakpoint cluster region (BCR)-Abelson murine leukemia (ABL1) and Janus Kinase-2 (JAK2) mutations have been thought to be mutually exclusive in myeloproliferative neoplasms (MNP), but recent data suggest that they can occur together.
Case report: A 68-year-old man was referred to the hematology clinic because of an elevated white blood cell count. His medical history included type II diabetes mellitus, hypertension, and retinal hemorrhage. Fluorescence in situ hybridization analysis of the bone marrow was positive for BCR-ABL1 in 66/100 cells. Conventional cytogenetics was positive for the Philadelphia chromosome in 16/20 counted cells. The percentage of BCR-ABL1 was 12%. Considering the patient's age and medical comorbidities, he was started on imatinib 400 mg once daily. Further tests showed JAK2 V617F mutation positivity and absence of acquired von Willebrand disease. He was then started on aspirin 81 mg and hydroxyurea 500 mg once daily, which was later increased to 1000 mg daily. The patient achieved a major molecular response after 6 months of treatment, with undetectable BCR-ABL1 levels.
Conclusions: BCR-ABL1 and JAK2 mutations can co-existence in MNP. Physicians must suspect the presence of one of the MNP in chronic myeloid leukemia (CML) patients with persistent or increased thrombocytosis, an atypical course of the disease, or hematological abnormalities despite evidence of response or remission of CML. Therefore, testing for JAK2 should be performed accordingly. Combining cytoreductive therapy with tyrosine kinase inhibitors (TKIs) is a therapeutic option when both mutations are present, and TKI alone is not sufficient to control peripheral blood cell counts.
Keywords: Janus Kinase 2 • EXOSC5 Protein, Human • Thrombocytosis
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Background

Chronic myeloid leukemia (CML) is a clonal hematopoietic stem cell disorder that causes bone marrow hyperplasia. It is a myeloproliferative neoplasm (MPN) with an estimated incidence of 1.9 cases per 100,000 adults and a death rate of 0.3 cases per 100,000 adults recorded in 2019 in the United States [1]. It is characterized by the presence of fusion between the Abelson murine leukemia (ABL1) gene on chromosome 9 and the breakpoint cluster region (BCR) gene on chromosome 22, leading to the expression of the oncoprotein BCR-ABL1. The derived chromosome is termed Philadelphia chromosome. BCR-ABL1 is a constitutively active tyrosine kinase that promotes growth and replication through downstream signaling pathways involving RAF, RAS, JAN kinase, STAT, and MYC [2]. Other MPNs include polycythemia vera (PV), essential thrombocythemia (ET), and primary myelofibrosis (PMF), which are collectively known as Philadelphia-negative MPNs. Diagnostic criteria for all MPNs have been developed by the World Health Organization [3]. The presence of recurrent genetic mutations is central to diagnosis. JAK2 mutation is the most common mutation detected in Philadelphia-negative MPNs, which also causes cell proliferation through the JAK-RAS-STAT pathway [4]. JAK2 mutations are present in up to 95% of PV cases and 65% of ET and PMF cases [5]. BCR-ABL1 and JAK2 mutations are thought to be mutually exclusive, but recent data and case reports have described cases in which these mutations occur concomitantly [6-13]. In this report, we describe

a unique coincidence presentation of CML and JAK2-positive thrombocytosis one month after initiating tyrosine kinase inhibitor (TKI) therapy.

Case Report

A 68-year-old man was referred to the hematology clinic because of an elevated white blood cell (WBC) count. He was diagnosed with type II diabetes mellitus and hypertension, after which he developed retinal hemorrhage. Prior to retinal laser photocoagulation, he was found to have a high white blood cell count. He was started empirically on hydroxyurea (500 mg once daily) and was referred to the clinic for further evaluation. Physical examination revealed splenomegaly (spleen size: 13 cm). His initial complete blood count (CBC) revealed the following: hemoglobin (Hb), 14.1 g/dL; WBCs, $34.6 \times 10^9/L$; platelets, $539 \times 10^9/L$; basophils, $0.34 \times 10^9/L$; eosinophils, $0.85 \times 10^9/L$; and neutrophils, $27.47 \times 10^9/L$. Bone marrow biopsy showed hypercellular marrow (cellularity estimated at 100%) with increased tri-lineage hematopoiesis, prominent megakaryopoiesis, and increased reticulin staining. The percentage of blasts was <1% as shown in **Figure 1**. Fluorescence in situ hybridization (FISH) analysis of the bone marrow was positive for BCR-ABL1 in 66/100 cells, with **Table 1** showing only 1 fusion signal, 1 signal for bcr, and 2 signals for abl, which indicates BCR/ABL rearrangement, representing a Philadelphia translocation with loss of 1 BCR copy and **Figure 2** showing 2 Green (BCR),

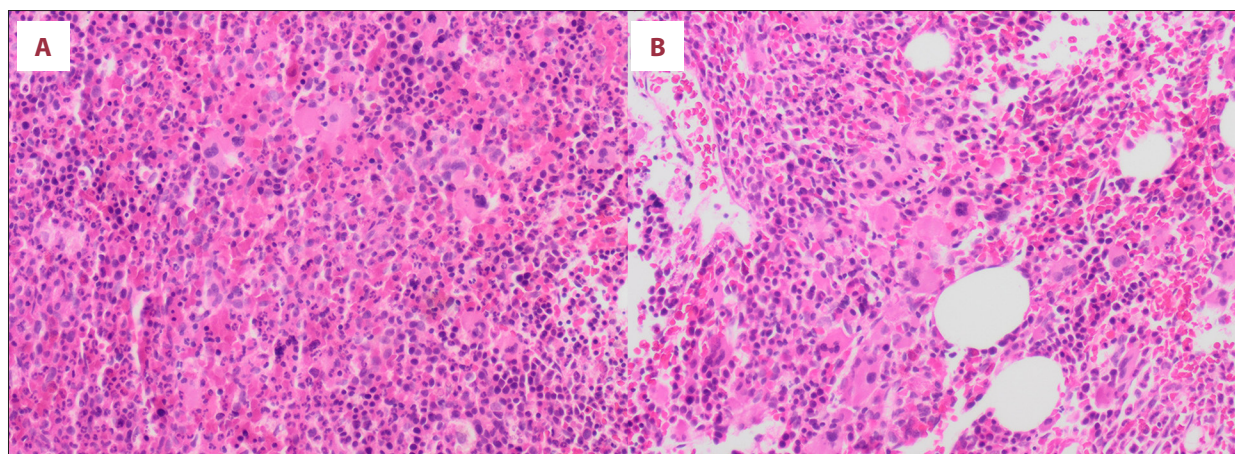


Figure 1. (A) Bone marrow biopsy showed hypercellularity ~100% (H&E stain, x50) with myeloid hyperplasia. Megakaryocytes are seen with variable sizes, and morphology includes dwarf forms and multiple erythroid islands. (B) Megakaryocytes are increased and show small loose clusters.

Table 1. Signals per probe of an analysis of 100 interphase fluorescence in situ hybridization (FISH) nuclei with probe for BCR/ABL.

BCR	ABL	Fusion signals	Number of cells (%)
2	2	0	34 (34)
2	3	1	66 (66)

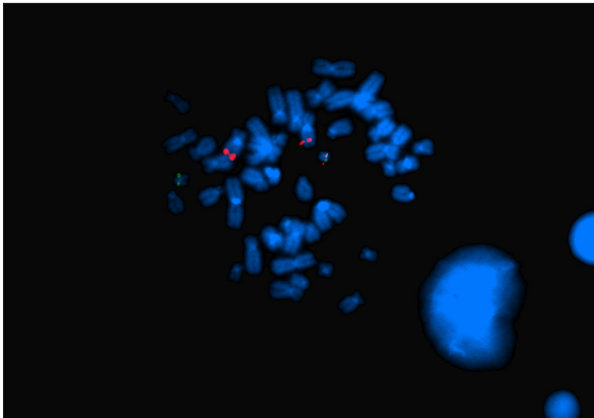


Figure 2. FISH analysis of a metaphase cell with LSI BCR/ABL revealed the following signal distribution: 2 signals for ABL on both chromosomes 9 (red), 1 signal for BCR on 1 chromosome 22 (green), and 1 fusion signal on the derivative chromosome 22.

3 Red (ABL) and 1 Fusion (BCR/ABL). Conventional cytogenetics was positive for the Philadelphia chromosome in 16/20 counted cells, showing 3-way translocation 46,XY,t(4;9;22)(q31;q34;q11.2)[17]/46,XY [4] as described in **Figure 3**. Real-time quantitative polymerase chain reaction (RT-qPCR) showed

that the percentage of BCR-ABL1 was 12%. Considering the patient's age and medical comorbidities, he was thus started on imatinib 400 mg once daily.

After 1 month of therapy, the patient's WBC count normalized and BCR-ABL1 percentage by RT-qPCR decreased to 9.5%, but his platelet count increased to $1543 \times 10^9/L$. Repeat bone marrow biopsy was negative for accelerated phase/blast phase CML. Further tests showed that JAK2 V617F mutation was positive, and the genomic DNA from the material obtained was screened with the Mutascreen Kit from IPSOGEN for presence of the mutation c. 1849G<T (p.V617F). Exon 14 of the Janus Kinase 2 gene (JAK2) was analyzed using real-time PCR. The measured results were compared with a reference sample, but the allele burden was not assessed. Tests for acquired von Willebrand disease yielded negative results. He was then started on aspirin 81 mg and hydroxyurea 500 mg once daily, which was later increased to 1000 mg daily. Throughout his treatment, the patient achieved all the CML treatment milestones, and he achieved a major molecular response after 6 months of treatment. His platelet count decreased to $400\text{-}500 \times 10^9/L$, but his spleen remained enlarged at 13 cm. During follow-up, the patient remained in major molecular response with undetectable levels of BCR-ABL1.

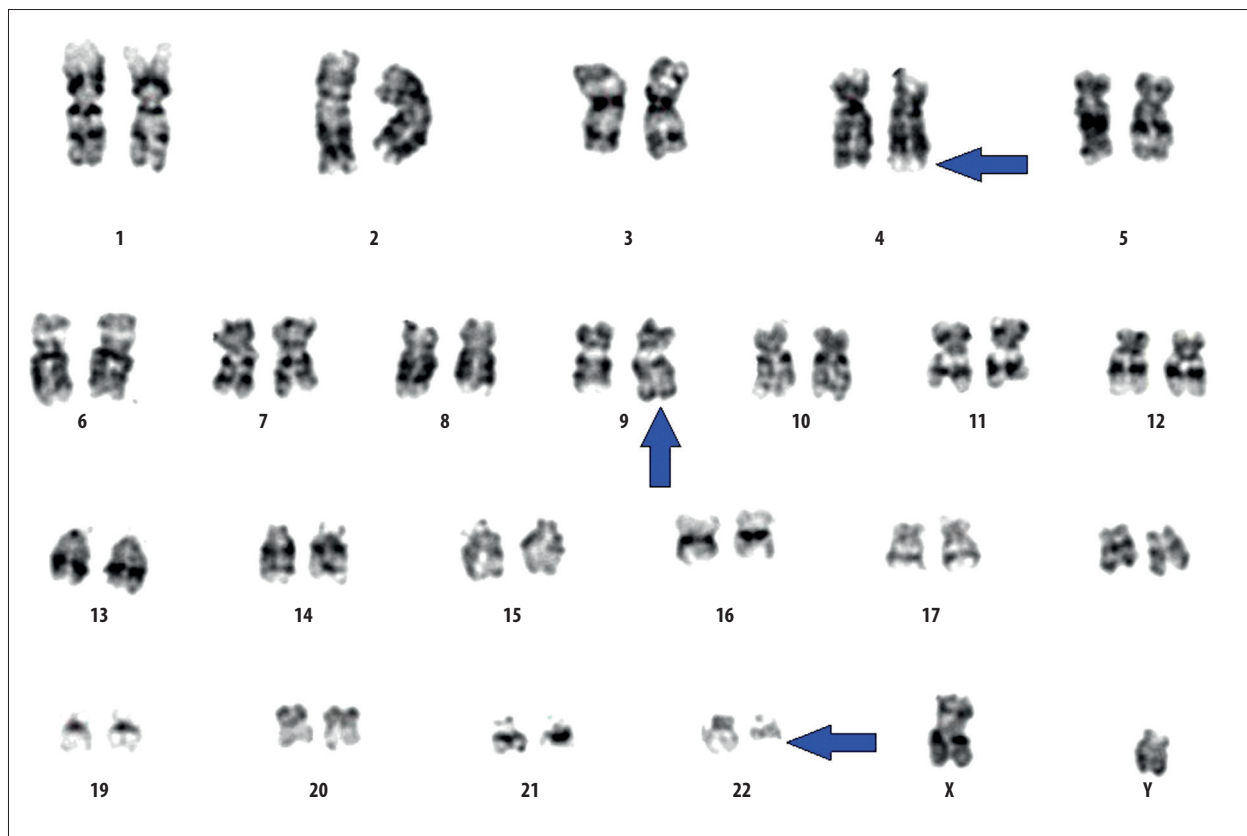


Figure 3. Chromosomal analysis revealed the presence of variant Philadelphia chromosome as a result of 3-way translocation between the long arms of chromosome 4,9 and 22 at bands 4q31,9q34, and 22q11.2 (arrows).

Discussion

The JAK2 V617F and BCR-ABL1 oncogenes are thought to be mutually exclusive; however, rare cases have been reported in which the occurrence of one may precede the occurrence of the other. Several attempts have been made to identify the prevalence of concomitant BCR-ABL1 and JAK2 mutations in patients with MPNs. Pieri et al tested 314 patients with BCR-ABL1-positive CML and found that 8 of them (2.5%) had the JAK2 V617F mutation [14]. Martin-Cabrera et al reported that among patients diagnosed with a myeloproliferative neoplasm by the presence of either JAK2 V617F, BCR-ABL1, or MPL mutations, 0.2% were found to be positive for both JAK2 V617F and BCR-ABL1 [15]. More recently, Soderquist et al reported a 0.4% prevalence of concurrent JAK2 V617F and BCR-ABL1 mutations when concomitant testing was performed on all patients suspected to have underlying MPN [10].

In our literature review of case reports, the diagnosis varied among CML preceding an MPN, an MPN preceding CML, or the co-existence of both disorders simultaneously [10,12]. The majority of the reviewed cases were initially diagnosed with MPN, ET, PV, or PMF, and had a JAK2 V617F mutation and later developed CML. The time from diagnosis of CML to MPN varied, ranging from 2 to 18 years after the initial diagnosis. The average duration from the diagnosis of MPN to CML later is longer than that from the diagnosis of CML to MPN later (7 vs 2 years) [12,16,17]. Although JAK2 and BCR-ABL were considered mutually exclusive drivers of mutation, it was repeatedly reported in the literature that they can co-exist. A case series by Soderquist et al indicated that both mutations can present in various manners [18-20].

Several theories attempt to explain the occurrence of both mutations simultaneously. One theory suggests the existence of 2 separate clones, in which one clone is suppressed by the proliferation of the other clone. Another theory suggests that there is a single clone that includes a relatively smaller subclone with both mutations. Other theories point to the possibility of a single clone harboring both mutations [21-25]. Ping et al proposed that the JAK clone will usually achieve dominance once the BCR-ABL clone is suppressed, although their patient had a recent diagnosis of PV [26].

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In our case, thrombocytosis became more pronounced after starting TKI (imatinib), with a decrease in BCR-ABL1 RT-qPCR levels. It is likely that these 2 mutations exist in different clones, and once the CML clone is suppressed, the JAK2 V617F clone becomes more active. The patient responded well to TKI and cytoreductive (hydroxyurea) therapy. The other options would include combining TKI and JAK inhibitors in such cases.

The limitations in our case report include not measuring the JAK2 V617F allele burden and inability to perform more complex testing for clonality, as we believe it is important to differentiate between clonal evolution vs selection.

Conclusions

Although BCR-ABL1 and JAK2 mutations are thought to be mutually exclusive, cases with co-existence of both mutations and acquiring new mutations have been reported. Physicians must suspect the presence of one of the MPNs in CML patients with persistent or increased thrombocytosis, an atypical course of the disease, or hematological abnormalities despite evidence of response or remission of CML. Therefore, testing for JAK2 should be performed. Combining cytoreductive therapy with TKI is a therapeutic option when both mutations are present, and TKI alone is not sufficient to control peripheral blood cell counts.

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Declaration of Figures' Authenticity

All figures submitted have been created by the authors who confirm that the images are original with no duplication and have not been previously published in whole or in part.

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