Supplementary Table 1: Principal component analysis (PCA)-derived scores of inherent resistance to single-shot radiotherapy (RTX), fractionated RTX, temozolomide (TMZ), and the combination thereof in human glioblastoma cell lines A172, LN18, LN229, T98G, U87, U138, and U251

Supplementary Table 2: Overview of composite karyotypes in human glioblastoma cell lines A172, LN18, LN229, T98G, U87, U138, and U251

The second column describes the predominant clonal karyotype including all chromosomal aberrations. The third column shows whether subclones are present in the cell lines.

Supplementary Table 3: Overview of molecular glioblastoma subtype-specific chromosomal amplifications and deletions, and subtype-specific expression of relevant driver genes

Supplementary Table 4: Genes with overlapping positive (R \geq 0.9, red) or negative (R \leq -0.9, blue) correlation with inherent therapy resistance in human glioblastoma cell lines as retrieved by global mRNA expression microarray analysis

Supplementary Table 5: CGC genes with overlapping positive ($R \ge 0.7$) correlation with inherent therapy resistance in human glioblastoma cells, for which drugs are readily available

Supplementary Table 6: Intersect leading edge genes for which drugs are readily available

Leading edge genes at the SSIR and TMZ intersection of the mammalian target of rapamycin complex 1 (mTORC1) pathway gene set for which inhibitors are readily available

References

- 1. Szymczak-Kulus, K., et al., Human Gb3/CD77 synthase produces P1 glycotope-capped N-glycans, which mediate Shiga toxin 1 but not Shiga toxin 2 cell entry. J Biol Chem, 2021. 296: p. 100299.
- 2. Piazzesi, A., S.Y. Afsar, and G. van Echten-Deckert, Sphingolipid metabolism in the development and progression of cancer: one cancer's help is another's hindrance. Mol Oncol, 2021. **15**(12): p. 3256-3279.
- 3. Jacob, F., et al., Transition of Mesenchymal and Epithelial Cancer Cells Depends on alpha1-4 Galactosyltransferase-Mediated Glycosphingolipids. Cancer Res, 2018. 78(11): p. 2952-2965.
- 4. Pellegrini, L., The Pol alpha-primase complex. Subcell Biochem, 2012. 62: p. 157-69.
- 5. Pollok, S., et al., Regulation of eukaryotic DNA replication at the initiation step. Biochem Soc Trans, 2003. 31(Pt 1): p. 266-9.
- 6. Ercilla, A., et al., Physiological Tolerance to ssDNA Enables Strand Uncoupling during DNA Replication. Cell Rep, 2020. 30(7): p. 2416-2429 e7.
- 7. O'Donnell, M., L. Langston, and B. Stillman, Principles and concepts of DNA replication in bacteria, archaea, and eukarya. Cold Spring Harb Perspect Biol, 2013. 5(7).
- 8. Starokadomskyy, P., et al., *DNA polymerase-alpha regulates the activation of type I interferons* through cytosolic RNA:DNA synthesis. Nat Immunol, 2016. 17(5): p. 495-504.
- 9. Yamaguchi, A., et al., DNA polymerase alpha positive-cell rate in colorectal cancer and its relationship to prognosis. Br J Cancer, 1992. 65(3): p. 421-4.
- 10. Tateishi, M., et al., DNA polymerase alpha as an independent prognostic parameter in non-small cell lung cancer--an immunohistochemical study. Eur J Surg Oncol, 1994. 20(4): p. 461-6.
- 11. Heitzer, E. and I. Tomlinson, Replicative DNA polymerase mutations in cancer. Curr Opin Genet Dev, 2014. 24: p. 107-13.
- 12. Berdis, A.J., Inhibiting DNA Polymerases as a Therapeutic Intervention against Cancer. Front Mol Biosci, 2017. 4: p. 78.
- 13. Han, T., et al., The antitumor toxin CD437 is a direct inhibitor of DNA polymerase alpha. Nat Chem Biol, 2016. 12(7): p. 511-5.
- 14. Abdel-Samad, R., et al., Mechanism of action of the atypical retinoid ST1926 in colorectal cancer: DNA damage and DNA polymerase alpha. Am J Cancer Res, 2018. 8(1): p. 39-55.
- 15. El Hajj, H., et al., Preclinical efficacy of the synthetic retinoid ST1926 for treating adult T-cell leukemia/lymphoma. Blood, 2014. 124(13): p. 2072-80.
- 16. Dallavalle, S., et al., Antitumor activity of novel POLA1-HDAC11 dual inhibitors. Eur J Med Chem, 2022. 228: p. 113971.
- 17. Park, S.Y. and X. Guo, Adaptor protein complexes and intracellular transport. Biosci Rep, 2014. $34(4)$.
- 18. Freire-Beneitez, V., et al., Elucidation of the BMI1 interactome identifies novel regulatory roles in glioblastoma. NAR Cancer, 2021. 3(1): p. zcab009.
- 19. Danan-Gotthold, M., et al., Identification of recurrent regulated alternative splicing events across human solid tumors. Nucleic Acids Res, 2015. 43(10): p. 5130-44.
- 20. Rangel, R., et al., Identification of New Tumor Suppressor Genes in Triple-Negative Breast Cancer. Cancer Res, 2017. 77(15): p. 4089-4101.
- 21. Beautrait, A., et al., A new inhibitor of the beta-arrestin/AP2 endocytic complex reveals interplay between GPCR internalization and signalling. Nat Commun, 2017. 8: p. 15054.
- 22. Yang, S. and N.B. Hecht, Translin associated protein X is essential for cellular proliferation. FEBS Lett, 2004. 576(1-2): p. 221-5.
- 23. Wang, Y., et al., Development of a nomogram for prognostic prediction of lower-grade glioma based on alternative splicing signatures. Cancer Med, 2020. 9(24): p. 9266-9281.
- 24. Yu, Y., et al., ZZEF1 is a Histone Reader and Transcriptional Coregulator of Kruppel-Like Factors. J Mol Biol, 2021. 433(2): p. 166722.
- 25. Giunti, L., et al., Genome-wide copy number analysis in pediatric glioblastoma multiforme. Am J Cancer Res, 2014. 4(3): p. 293-303.
- 26. Joo, J.E., et al., Heritable DNA methylation marks associated with susceptibility to breast cancer. Nat Commun, 2018. 9(1): p. 867.
- 27. Cuenda, A., Mitogen-activated protein kinase kinase 4 (MKK4). Int J Biochem Cell Biol, 2000. 32(6): p. 581-7.
- 28. Davis, S.J., et al., Analysis of the mitogen-activated protein kinase kinase 4 (MAP2K4) tumor suppressor gene in ovarian cancer. BMC Cancer, 2011. **11**: p. 173.
- 29. Ahn, Y.H., et al., Map2k4 functions as a tumor suppressor in lung adenocarcinoma and inhibits tumor cell invasion by decreasing peroxisome proliferator-activated receptor gamma2 expression. Mol Cell Biol, 2011. 31(21): p. 4270-85.
- 30. Spillman, M.A., et al., Regulation of the metastasis suppressor gene MKK4 in ovarian cancer. Gynecol Oncol, 2007. 105(2): p. 312-20.
- 31. Robinson, V.L., et al., Mitogen-activated protein kinase kinase 4/c-Jun NH2-terminal kinase kinase 1 protein expression is subject to translational regulation in prostate cancer cell lines. Mol Cancer Res, 2008. 6(3): p. 501-8.
- 32. Yamada, S.D., et al., Mitogen-activated protein kinase kinase 4 (MKK4) acts as a metastasis suppressor gene in human ovarian carcinoma. Cancer Res, 2002. 62(22): p. 6717-23.
- 33. Kim, H.L., et al., Mitogen-activated protein kinase kinase 4 metastasis suppressor gene expression is inversely related to histological pattern in advancing human prostatic cancers. Cancer Res, 2001. 61(7): p. 2833-7.
- 34. Chae, K.S., et al., Expression and mutation analyses of MKK4, a candidate tumour suppressor gene encoded by chromosome 17p, in human gastric adenocarcinoma. Eur J Cancer, 2002. 38(15): p. 2048-57.
- 35. Su, G.H., et al., Mutation rate of MAP2K4/MKK4 in breast carcinoma. Hum Mutat, 2002. 19(1): p. 81.
- 36. Xin, W., et al., MAP2K4/MKK4 expression in pancreatic cancer: genetic validation of immunohistochemistry and relationship to disease course. Clin Cancer Res, 2004. 10(24): p. 8516- 20.
- 37. Huang, C., et al., Overexpression of mitogen-activated protein kinase kinase 4 and nuclear factorkappaB in laryngeal squamous cell carcinoma: a potential indicator for poor prognosis. Oncol Rep, 2009. 22(1): p. 89-95.
- 38. Finegan, K.G. and C. Tournier, The mitogen-activated protein kinase kinase 4 has a pro-oncogenic role in skin cancer. Cancer Res, 2010. 70(14): p. 5797-806.
- 39. Pavese, J.M., et al., Mitogen-activated protein kinase kinase 4 (MAP2K4) promotes human prostate cancer metastasis. PLoS One, 2014. 9(7): p. e102289.
- 40. Whitmarsh, A.J. and R.J. Davis, Role of mitogen-activated protein kinase kinase 4 in cancer. Oncogene, 2007. 26(22): p. 3172-84.
- 41. Xue, Z., et al., MAP3K1 and MAP2K4 mutations are associated with sensitivity to MEK inhibitors in multiple cancer models. Cell Res, 2018. 28(7): p. 719-729.
- 42. Chen, X., et al., Activation of JNK and p38 MAPK Mediated by ZDHHC17 Drives Glioblastoma Multiforme Development and Malignant Progression. Theranostics, 2020. 10(3): p. 998-1015.
- 43. Akiyama, T., et al., Genistein, a specific inhibitor of tyrosine-specific protein kinases. J Biol Chem, 1987. 262(12): p. 5592-5.
- 44. Liu, X., et al., Genistein inhibits radiation-induced invasion and migration of glioblastoma cells by blocking the DNA-PKcs/Akt2/Rac1 signaling pathway. Radiother Oncol, 2021. 155: p. 93-104.
- 45. Atefeh, Z., et al., Combination Treatment of Glioblastoma by Low-Dose Radiation and Genistein. Curr Radiopharm, 2016. 9(3): p. 258-263.
- 46. Hubner, M., et al., Intronic miR-744 Inhibits Glioblastoma Migration by Functionally Antagonizing Its Host Gene MAP2K4. Cancers (Basel), 2018. 10(11).
- 47. Kwong, A.J., et al., Rational Design, Optimization, and Biological Evaluation of Novel MEK4 Inhibitors against Pancreatic Adenocarcinoma. ACS Med Chem Lett, 2021. 12(10): p. 1559-1567.
- 48. Deibler, K.K., et al., Synthesis and Biological Evaluation of 3-Arylindazoles as Selective MEK4 Inhibitors. ChemMedChem, 2019. 14(6): p. 615-620.
- 49. Jiang, J., et al., Discovery of Covalent MKK4/7 Dual Inhibitor. Cell Chem Biol, 2020. 27(12): p. 1553-1560 e8.
- 50. Mathew, S. and C.G. Vazhappilly, Recent pharmacological advances on genistein in clinical trials. EXCLI J, 2020. 19: p. 1120-1123.
- 51. Mardian, E.B., R.M. Bradley, and R.E. Duncan, The HRASLS (PLA/AT) subfamily of enzymes. J Biomed Sci, 2015. 22: p. 99.
- 52. Uyama, T., et al., Characterization of the human tumor suppressors TIG3 and HRASLS2 as phospholipid-metabolizing enzymes. Biochim Biophys Acta, 2009. 1791(12): p. 1114-24.
- 53. Chen, C., et al., Identification of key genes in glioblastoma-associated stromal cells using bioinformatics analysis. Oncol Lett, 2016. 11(6): p. 3999-4007.
- 54. Zhou, J., et al., Activity-Based Protein Profiling Identifies alpha-Ketoamides as Inhibitors for Phospholipase A2 Group XVI. ACS Chem Biol, 2019. 14(2): p. 164-169.
- 55. Zhou, J., et al., Structure-Activity Relationship Studies of alpha-Ketoamides as Inhibitors of the Phospholipase A and Acyltransferase Enzyme Family. J Med Chem, 2020. 63(17): p. 9340-9359.
- 56. Wu, L., et al., Regulation of PLC-mediated signalling in vivo by CDP-diacylglycerol synthase. Nature, 1995. 373(6511): p. 216-22.
- 57. Shen, H. and W. Dowhan, Regulation of phospholipid biosynthetic enzymes by the level of CDPdiacylglycerol synthase activity. J Biol Chem, 1997. 272(17): p. 11215-20.
- 58. Yeh, K.T., et al., Methylation Inactivates Expression of CDP-diacylglycerol Synthase 1 (CDS1) in Hepatocellular Carcinoma. Cancer Genomics Proteomics, 2006. 3(3-4): p. 231-238.
- 59. Wang, H., et al., Integrative analysis of somatic mutations and differential expression profiles in glioblastoma based on aging acceleration. Int J Clin Exp Pathol, 2021. 14(5): p. 582-595.
- 60. Kucharzewska, P., H.C. Christianson, and M. Belting, Global profiling of metabolic adaptation to hypoxic stress in human glioblastoma cells. PLoS One, 2015. 10(1): p. e0116740.
- 61. Monne, M., et al., The mitochondrial oxoglutarate carrier: from identification to mechanism. J Bioenerg Biomembr, 2013. 45(1-2): p. 1-13.
- 62. Lee, J.S., et al., Loss of SLC25A11 causes suppression of NSCLC and melanoma tumor formation. EBioMedicine, 2019. 40: p. 184-197.
- 63. Pan, G., et al., SLC25A11 serves as a novel prognostic biomarker in liver cancer. Sci Rep, 2020. 10(1): p. 9871.
- 64. Buffet, A., et al., Germline Mutations in the Mitochondrial 2-Oxoglutarate/Malate Carrier SLC25A11 Gene Confer a Predisposition to Metastatic Paragangliomas. Cancer Res, 2018. **78**(8): p. 1914-1922.
- 65. Wu, L.L. and X.F. Zhou, *Huntingtin associated protein 1 and its functions.* Cell Adh Migr, 2009. 3(1): p. 71-6.
- 66. Zhao, X., et al., Biological functions and potential therapeutic applications of huntingtin-associated protein 1: progress and prospects. Clin Transl Oncol, 2022. 24(2): p. 203-214.
- 67. Wu, J., et al., HAP1 gene expression is associated with radiosensitivity in breast cancer cells. Biochem Biophys Res Commun, 2015. 456(1): p. 162-6.
- 68. Prieto-Alamo, M.J. and F. Laval, Overexpression of the human HAP1 protein sensitizes cells to the lethal effect of bioreductive drugs. Carcinogenesis, 1999. 20(3): p. 415-9.
- 69. Herring, C.J., et al., Expression levels of the DNA repair enzyme HAP1 do not correlate with the radiosensitivities of human or HAP1-transfected rat cell lines. Br J Cancer, 1999. 80(7): p. 940-5.
- 70. Amen, A.M., et al., Cancer-specific loss of TERT activation sensitizes glioblastoma to DNA damage. Proc Natl Acad Sci U S A, 2021. 118(13).
- 71. Walker, L.J., et al., A role for the human DNA repair enzyme HAP1 in cellular protection against DNA damaging agents and hypoxic stress. Nucleic Acids Res, 1994. 22(23): p. 4884-9.
- 72. Chatterji, A. and R. Sengupta, Cellular S-denitrosylases: Potential role and interplay of Thioredoxin, TRP14, and Glutaredoxin systems in thiol-dependent protein denitrosylation. Int J Biochem Cell Biol, 2021. 131: p. 105904.
- 73. Kyani, A., et al., Discovery and Mechanistic Elucidation of a Class of Protein Disulfide Isomerase Inhibitors for the Treatment of Glioblastoma. ChemMedChem, 2018. 13(2): p. 164-177.
- 74. Kudo, L.C., et al., Puromycin-sensitive aminopeptidase (PSA/NPEPPS) impedes development of neuropathology in hPSA/TAU(P301L) double-transgenic mice. Hum Mol Genet, 2011. 20(9): p. 1820-33.
- 75. Ren, G., et al., Cu, Zn-superoxide dismutase 1 (SOD1) is a novel target of Puromycin-sensitive aminopeptidase (PSA/NPEPPS): PSA/NPEPPS is a possible modifier of amyotrophic lateral sclerosis. Mol Neurodegener, 2011. 6: p. 29.
- 76. Agrawal, N. and M.A. Brown, Genetic associations and functional characterization of M1 aminopeptidases and immune-mediated diseases. Genes Immun, 2014. 15(8): p. 521-7.
- 77. Verano-Braga, T., et al., SuperQuant-assisted comparative proteome analysis of glioblastoma subpopulations allows for identification of potential novel therapeutic targets and cell markers. Oncotarget, 2018. 9(10): p. 9400-9414.
- 78. Dhanoa, B.S., et al., Update on the Kelch-like (KLHL) gene family. Hum Genomics, 2013. 7: p. 13.
- 79. Panieri, E., et al., NRF2 and Mitochondrial Function in Cancer and Cancer Stem Cells. Cells, 2022. 11(15).
- 80. Shahcheraghi, S.H., et al., The Role of NRF2/KEAP1 Pathway in Glioblastoma: Pharmacological Implications. Med Oncol, 2022. 39(5): p. 91.
- 81. Kanamori, M., et al., Activation of the NRF2 pathway and its impact on the prognosis of anaplastic glioma patients. Neuro Oncol, 2015. 17(4): p. 555-65.
- 82. Olivier, C., et al., Drug Resistance in Glioblastoma: The Two Faces of Oxidative Stress. Front Mol Biosci, 2020. 7: p. 620677.
- 83. Godoy, P., et al., Targeting NRF2, Regulator of Antioxidant System, to Sensitize Glioblastoma Neurosphere Cells to Radiation-Induced Oxidative Stress. Oxid Med Cell Longev, 2020. 2020: p. 2534643.
- 84. Robledinos-Anton, N., et al., Activators and Inhibitors of NRF2: A Review of Their Potential for Clinical Development. Oxid Med Cell Longev, 2019. 2019: p. 9372182.
- 85. Yagishita, Y., et al., Current Landscape of NRF2 Biomarkers in Clinical Trials. Antioxidants (Basel), 2020. 9(8).
- 86. Pouremamali, F., et al., An update of Nrf2 activators and inhibitors in cancer prevention/promotion. Cell Commun Signal, 2022. 20(1): p. 100.
- 87. Alavi, M., et al., Resveratrol mediates its anti-cancer effects by Nrf2 signaling pathway activation. Cancer Cell Int, 2021. 21(1): p. 579.
- 88. Ashrafizadeh, M., et al., Curcumin Activates the Nrf2 Pathway and Induces Cellular Protection Against Oxidative Injury. Curr Mol Med, 2020. 20(2): p. 116-133.
- 89. Olayanju, A., et al., Brusatol provokes a rapid and transient inhibition of Nrf2 signaling and sensitizes mammalian cells to chemical toxicity-implications for therapeutic targeting of Nrf2. Free Radic Biol Med, 2015. 78: p. 202-12.
- 90. Ramos-Gomez, M., et al., Sensitivity to carcinogenesis is increased and chemoprotective efficacy of enzyme inducers is lost in nrf2 transcription factor-deficient mice. Proc Natl Acad Sci U S A, 2001. 98(6): p. 3410-5.
- 91. Reisman, S.A., et al., Topical application of the synthetic triterpenoid RTA 408 activates Nrf2 and induces cytoprotective genes in rat skin. Arch Dermatol Res, 2014. 306(5): p. 447-54.
- 92. Probst, B.L., et al., RTA 408, A Novel Synthetic Triterpenoid with Broad Anticancer and Anti-Inflammatory Activity. PLoS One, 2015. 10(4): p. e0122942.
- 93. Singh, A., et al., Small Molecule Inhibitor of NRF2 Selectively Intervenes Therapeutic Resistance in KEAP1-Deficient NSCLC Tumors. ACS Chem Biol, 2016. 11(11): p. 3214-3225.
- 94. Hu, L., et al., Discovery of a small-molecule inhibitor and cellular probe of Keap1-Nrf2 protein-protein interaction. Bioorg Med Chem Lett, 2013. 23(10): p. 3039-43.
- 95. Bard, J.A.M., et al., Structure and Function of the 26S Proteasome. Annu Rev Biochem, 2018. 87: p. 697-724.
- 96. Manasanch, E.E. and R.Z. Orlowski, Proteasome inhibitors in cancer therapy. Nat Rev Clin Oncol, 2017. 14(7): p. 417-433.
- 97. Adams, J., Proteasome inhibition: a novel approach to cancer therapy. Trends Mol Med, 2002. 8(4 Suppl): p. S49-54.
- 98. Hideshima, T., et al., The proteasome inhibitor PS-341 inhibits growth, induces apoptosis, and overcomes drug resistance in human multiple myeloma cells. Cancer Res, 2001. 61(7): p. 3071-6.
- 99. LeBlanc, R., et al., Proteasome inhibitor PS-341 inhibits human myeloma cell growth in vivo and prolongs survival in a murine model. Cancer Res, 2002. 62(17): p. 4996-5000.
- 100. Kuhn, D.J., et al., Potent activity of carfilzomib, a novel, irreversible inhibitor of the ubiquitinproteasome pathway, against preclinical models of multiple myeloma. Blood, 2007. 110(9): p. 3281- 90.
- 101. Yang, H., et al., Celastrol, a triterpene extracted from the Chinese "Thunder of God Vine," is a potent proteasome inhibitor and suppresses human prostate cancer growth in nude mice. Cancer Res, 2006. 66(9): p. 4758-65.
- 102. Sethi, G., et al., Celastrol, a novel triterpene, potentiates TNF-induced apoptosis and suppresses invasion of tumor cells by inhibiting NF-kappaB-regulated gene products and TAK1-mediated NFkappaB activation. Blood, 2007. 109(7): p. 2727-35.
- 103. Piva, R., et al., CEP-18770: A novel, orally active proteasome inhibitor with a tumor-selective pharmacologic profile competitive with bortezomib. Blood, 2008. 111(5): p. 2765-75.
- 104. Kupperman, E., et al., Evaluation of the proteasome inhibitor MLN9708 in preclinical models of human cancer. Cancer Res, 2010. 70(5): p. 1970-80.
- 105. Tsubuki, S., et al., Differential inhibition of calpain and proteasome activities by peptidyl aldehydes of di-leucine and tri-leucine. J Biochem, 1996. 119(3): p. 572-6.
- 106. Raedler, L., Velcade (Bortezomib) Receives 2 New FDA Indications: For Retreatment of Patients with Multiple Myeloma and for First-Line Treatment of Patients with Mantle-Cell Lymphoma. Am Health Drug Benefits, 2015. 8(Spec Feature): p. 135-40.
- 107. Vander Ark, A., J. Cao, and X. Li, TGF-beta receptors: In and beyond TGF-beta signaling. Cell Signal, 2018. 52: p. 112-120.
- 108. Song, K., et al., ERBB3, IGF1R, and TGFBR2 expression correlate with PDGFR expression in glioblastoma and participate in PDGFR inhibitor resistance of glioblastoma cells. Am J Cancer Res, 2018. 8(5): p. 792-809.
- 109. Jun, F., et al., Epithelial membrane protein 3 regulates TGF-beta signaling activation in CD44-high glioblastoma. Oncotarget, 2017. 8(9): p. 14343-14358.
- 110. Kim, B.G., et al., Novel therapies emerging in oncology to target the TGF-beta pathway. J Hematol Oncol, 2021. 14(1): p. 55.
- 111. Bueno, L., et al., Semi-mechanistic modelling of the tumour growth inhibitory effects of LY2157299, a new type I receptor TGF-beta kinase antagonist, in mice. Eur J Cancer, 2008. 44(1): p. 142-50.
- 112. Son, J.Y., et al., EW-7197, a novel ALK-5 kinase inhibitor, potently inhibits breast to lung metastasis. Mol Cancer Ther, 2014. 13(7): p. 1704-16.
- 113. Jin, C.H., et al., Discovery of N-((4-([1,2,4]triazolo[1,5-a]pyridin-6-yl)-5-(6-methylpyridin-2-yl)-1Himidazol-2 -yl)methyl)-2-fluoroaniline (EW-7197): a highly potent, selective, and orally bioavailable inhibitor of TGF-beta type I receptor kinase as cancer immunotherapeutic/antifibrotic agent. J Med Chem, 2014. 57(10): p. 4213-38.
- 114. Li, H.Y., et al., Dihydropyrrolopyrazole transforming growth factor-beta type I receptor kinase domain inhibitors: a novel benzimidazole series with selectivity versus transforming growth factor-beta type II receptor kinase and mixed lineage kinase-7. J Med Chem, 2006. 49(6): p. 2138-42.
- 115. Liao, F., et al., LSKL peptide alleviates subarachnoid fibrosis and hydrocephalus by inhibiting TSP1 mediated TGF-beta1 signaling activity following subarachnoid hemorrhage in rats. Exp Ther Med, 2016. 12(4): p. 2537-2543.
- 116. Wick, A., et al., Phase 1b/2a study of galunisertib, a small molecule inhibitor of transforming growth factor-beta receptor I, in combination with standard temozolomide-based radiochemotherapy in patients with newly diagnosed malignant glioma. Invest New Drugs, 2020. 38(5): p. 1570-1579.
- 117. Fedele, M., E. Crescenzi, and L. Cerchia, The POZ/BTB and AT-Hook Containing Zinc Finger 1 (PATZ1) Transcription Regulator: Physiological Functions and Disease Involvement. Int J Mol Sci, 2017. 18(12).
- 118. Quintero-Rivera, F., et al., Disruption of a synaptotagmin (SYT14) associated with neurodevelopmental abnormalities. Am J Med Genet A, 2007. 143A(6): p. 558-63.
- 119. Richter, W.F., et al., The Mediator complex as a master regulator of transcription by RNA polymerase II. Nat Rev Mol Cell Biol, 2022.
- 120. Adler, D., et al., MED15, encoding a subunit of the mediator complex, is overexpressed at high frequency in castration-resistant prostate cancer. Int J Cancer, 2014. 135(1): p. 19-26.
- 121. Klumper, N., et al., Differential expression of Mediator complex subunit MED15 in testicular germ cell tumors. Diagn Pathol, 2015. 10: p. 165.
- 122. Offermann, A., et al., MED15 overexpression in prostate cancer arises during androgen deprivation therapy via PI3K/mTOR signaling. Oncotarget, 2017. 8(5): p. 7964-7976.
- 123. Syring, I., et al., The knockdown of the Mediator complex subunit MED15 restrains urothelial bladder cancer cells' malignancy. Oncol Lett, 2018. 16(3): p. 3013-3021.
- 124. White, D.N. and M.H.B. Stowell, Room for Two: The Synaptophysin/Synaptobrevin Complex. Front Synaptic Neurosci, 2021. 13: p. 740318.
- 125. Chen, D.H., et al., SYPL1 overexpression predicts poor prognosis of hepatocellular carcinoma and associates with epithelial-mesenchymal transition. Oncol Rep, 2017. 38(3): p. 1533-1542.
- 126. Yang, C. and Y. Wang, Identification of differentiated functional modules in papillary thyroid carcinoma by analyzing differential networks. J Cancer Res Ther, 2018. 14(Supplement): p. S969- S974.
- 127. Pinzon Martin, S., P.H. Seeberger, and D. Varon Silva, Mucins and Pathogenic Mucin-Like Molecules Are Immunomodulators During Infection and Targets for Diagnostics and Vaccines. Front Chem, 2019. 7: p. 710.
- 128. Sun, X., et al., C1GALT1 in health and disease. Oncol Lett, 2021, 22(2): p. 589.
- 129. Lin, M.C., et al., C1GALT1 predicts poor prognosis and is a potential therapeutic target in head and neck cancer. Oncogene, 2018. 37(43): p. 5780-5793.
- 130. Dong, X., et al., A novel mechanism for C1GALT1 in the regulation of gastric cancer progression. Cell Biosci, 2021. 11(1): p. 166.
- 131. Grummt, I., Life on a planet of its own: regulation of RNA polymerase I transcription in the nucleolus. Genes Dev, 2003. 17(14): p. 1691-702.
- 132. Drygin, D., W.G. Rice, and I. Grummt, The RNA polymerase I transcription machinery: an emerging target for the treatment of cancer. Annu Rev Pharmacol Toxicol, 2010. 50: p. 131-56.
- 133. Rossetti, S., A.J. Wierzbicki, and N. Sacchi, Mammary epithelial morphogenesis and early breast cancer. Evidence of involvement of basal components of the RNA Polymerase I transcription machinery. Cell Cycle, 2016. 15(18): p. 2515-26.
- 134. Ferreira, R., et al., Targeting the RNA Polymerase I Transcription for Cancer Therapy Comes of Age. Cells, 2020. 9(2).
- 135. Peltonen, K., et al., Identification of novel p53 pathway activating small-molecule compounds reveals unexpected similarities with known therapeutic agents. PLoS One, 2010. 5(9): p. e12996.
- 136. Drygin, D., et al., Anticancer activity of CX-3543: a direct inhibitor of rRNA biogenesis. Cancer Res, 2009. 69(19): p. 7653-61.
- 137. Drygin, D., et al., Targeting RNA polymerase I with an oral small molecule CX-5461 inhibits ribosomal RNA synthesis and solid tumor growth. Cancer Res, 2011. 71(4): p. 1418-30.
- 138. Hourcade, D., V.M. Holers, and J.P. Atkinson, The regulators of complement activation (RCA) gene cluster. Adv Immunol, 1989. 45: p. 381-416.
- 139. Parente, R., et al., Complement factor H in host defense and immune evasion. Cell Mol Life Sci, 2017. 74(9): p. 1605-1624.
- 140. Zhai, L., et al., Tumor Cell IDO Enhances Immune Suppression and Decreases Survival Independent of Tryptophan Metabolism in Glioblastoma. Clin Cancer Res, 2021. 27(23): p. 6514-6528.
- 141. Bian, A., et al., Circular RNA Complement Factor H (CFH) Promotes Glioma Progression by Sponging miR-149 and Regulating AKT1. Med Sci Monit, 2018. 24: p. 5704-5712.
- 142. Ajona, D., et al., Expression of complement factor H by lung cancer cells: effects on the activation of the alternative pathway of complement. Cancer Res, 2004. 64(17): p. 6310-8.
- 143. Junnikkala, S., et al., Secretion of soluble complement inhibitors factor H and factor H-like protein (FHL-1) by ovarian tumour cells. Br J Cancer, 2002. 87(10): p. 1119-27.
- 144. Jin, H.J., J. Kim, and J. Yu, Androgen receptor genomic regulation. Transl Androl Urol, 2013. 2(3): p. 157-177.
- 145. Matsumoto, T., et al., The androgen receptor in health and disease. Annu Rev Physiol, 2013. 75: p. 201-24.
- 146. Davey, R.A. and M. Grossmann, Androgen Receptor Structure, Function and Biology: From Bench to Bedside. Clin Biochem Rev, 2016. 37(1): p. 3-15.
- 147. Hu, C., et al., The androgen receptor expression and association with patient's survival in different cancers. Genomics, 2020. 112(2): p. 1926-1940.
- 148. Farina-Jeronimo, H., et al., Androgen Receptor Activity Is Associated with Worse Survival in Glioblastoma. J Integr Neurosci, 2022. 21(3): p. 86.
- 149. Zalcman, N., et al., Androgen receptor: a potential therapeutic target for glioblastoma. Oncotarget, 2018. 9(28): p. 19980-19993.
- 150. Lysiak, M., et al., The sex-dependent role of the androgen receptor in glioblastoma: results of molecular analyses. Mol Oncol, 2022.
- 151. Yang, J.D., et al., Contribution of the Testosterone Androgen Receptor-PARD3B Signaling Axis to Tumorigenesis and Malignance of Glioblastoma Multiforme through Stimulating Cell Proliferation and Colony Formation. J Clin Med, 2022. 11(16).
- 152. Chen, T.C., et al., AR ubiquitination induced by the curcumin analog suppresses growth of temozolomide-resistant glioblastoma through disrupting GPX4-Mediated redox homeostasis. Redox Biol, 2020. 30: p. 101413.
- 153. Rodriguez-Lozano, D.C., et al., Testosterone Promotes Glioblastoma Cell Proliferation, Migration, and Invasion Through Androgen Receptor Activation. Front Endocrinol (Lausanne), 2019. 10: p. 16.
- 154. Rodriguez-Lozano, D.C., et al., Dihydrotestosterone Induces Proliferation, Migration, and Invasion of Human Glioblastoma Cell Lines. Onco Targets Ther, 2020. 13: p. 8813-8823.
- 155. Yu, X., et al., Androgen receptor signaling regulates growth of glioblastoma multiforme in men. Tumour Biol, 2015. 36(2): p. 967-72.
- 156. Werner, C.K., et al., *Expression of the Androgen Receptor Governs Radiation Resistance in a Subset* of Glioblastomas Vulnerable to Antiandrogen Therapy. Mol Cancer Ther, 2020. 19(10): p. 2163- 2174.
- 157. Zhao, N., et al., Androgen Receptor, Although Not a Specific Marker For, Is a Novel Target to Suppress Glioma Stem Cells as a Therapeutic Strategy for Glioblastoma. Front Oncol, 2021. 11: p. 616625.
- 158. Orozco, M., et al., Dutasteride combined with androgen receptor antagonists inhibit glioblastoma U87 cell metabolism, proliferation, and invasion capacity: Androgen regulation. Steroids, 2020. 164: p. 108733.
- 159. Kim, T.J., Y.H. Lee, and K.C. Koo, Current Status and Future Perspectives of Androgen Receptor Inhibition Therapy for Prostate Cancer: A Comprehensive Review. Biomolecules, 2021. 11(4).
- 160. Kono, M., et al., Androgen Receptor Function and Androgen Receptor-Targeted Therapies in Breast Cancer: A Review. JAMA Oncol, 2017. 3(9): p. 1266-1273.
- 161. Chen, Y., et al., Second generation androgen receptor antagonists and challenges in prostate cancer treatment. Cell Death Dis, 2022. 13(7): p. 632.
- 162. Clegg, N.J., et al., ARN-509: a novel antiandrogen for prostate cancer treatment. Cancer Res, 2012. 72(6): p. 1494-503.
- 163. Furr, B.J., et al., ICI 176,334: a novel non-steroidal, peripherally selective antiandrogen. J Endocrinol, 1987. 113(3): p. R7-9.
- 164. Furr, B.J., ICI 176,334: a novel non-steroidal, peripherally-selective antiandrogen. Prog Clin Biol Res, 1988. 260: p. 13-26.
- 165. Moilanen, A.M., et al., Discovery of ODM-201, a new-generation androgen receptor inhibitor targeting resistance mechanisms to androgen signaling-directed prostate cancer therapies. Sci Rep, 2015. 5: p. 12007.
- 166. Tran, C., et al., Development of a second-generation antiandrogen for treatment of advanced prostate cancer. Science, 2009. 324(5928): p. 787-90.
- 167. Katchen, B. and S. Buxbaum, Disposition of a new, nonsteroid, antiandrogen, alpha,alpha,alphatrifluoro-2-methyl-4'-nitro-m-propionotoluidide (Flutamide), in men following a single oral 200 mg dose. J Clin Endocrinol Metab, 1975. 41(2): p. 373-9.
- 168. Harris, M.G., et al., Nilutamide. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic efficacy in prostate cancer. Drugs Aging, 1993. 3(1): p. 9-25.
- 169. Belanger, A., A. Dupont, and F. Labrie, Inhibition of basal and adrenocorticotropin-stimulated plasma levels of adrenal androgens after treatment with an antiandrogen in castrated patients with prostatic cancer. J Clin Endocrinol Metab, 1984. 59(3): p. 422-6.
- 170. Wollman, A.L. and J.B. Hamilton, Prevention by cyproterone acetate of androgenic, but not of gonadotrophic, elicitation of persistent estrus in rats. Endocrinology, 1967. 81(2): p. 350-6.
- 171. Brave, M., et al., An FDA Review of Drug Development in Nonmetastatic Castration-resistant Prostate Cancer. Clin Cancer Res, 2020. 26(18): p. 4717-4722.
- 172. Kanai, T., et al., *Identification of STAT5A and STAT5B target genes in human T cells.* PLoS One, 2014. 9(1): p. e86790.
- 173. Kanai, T., J. Jenks, and K.C. Nadeau, The STAT5b Pathway Defect and Autoimmunity. Front Immunol, 2012. 3: p. 234.
- 174. Dubois, N., et al., STAT5b is a marker of poor prognosis, rather than a therapeutic target in glioblastomas. Int J Oncol, 2022. 61(4).
- 175. Roos, A., et al., EGFRvIII-Stat5 Signaling Enhances Glioblastoma Cell Migration and Survival. Mol Cancer Res, 2018. 16(7): p. 1185-1195.
- 176. Latha, K., et al., Nuclear EGFRvIII-STAT5b complex contributes to glioblastoma cell survival by direct activation of the Bcl-XL promoter. Int J Cancer, 2013. 132(3): p. 509-20.
- 177. Moyama, C., et al., Stat5b inhibition blocks proliferation and tumorigenicity of glioblastoma stem cells derived from a de novo murine brain cancer model. Am J Cancer Res, 2022. 12(3): p. 1129-1142.
- 178. Liang, Q.C., et al., Inhibition of transcription factor STAT5b suppresses proliferation, induces G1 cell cycle arrest and reduces tumor cell invasion in human glioblastoma multiforme cells. Cancer Lett, 2009. 273(1): p. 164-71.
- 179. Loh, C.Y., et al., Signal Transducer and Activator of Transcription (STATs) Proteins in Cancer and Inflammation: Functions and Therapeutic Implication. Front Oncol, 2019. 9: p. 48.
- 180. Muller, J., et al., Discovery of chromone-based inhibitors of the transcription factor STAT5. Chembiochem, 2008. 9(5): p. 723-7.
- 181. Chien, C.M., et al., Novel indoloquinoline derivative, IQDMA, suppresses STAT5 phosphorylation and induces apoptosis in HL-60 cells. Chem Biol Interact, 2008. 176(1): p. 40-7.
- 182. Nelson, E.A., et al., The STAT5 inhibitor pimozide decreases survival of chronic myelogenous leukemia cells resistant to kinase inhibitors. Blood, 2011. 117(12): p. 3421-9.
- 183. Wingelhofer, B., et al., Pharmacologic inhibition of STAT5 in acute myeloid leukemia. Leukemia, 2018. 32(5): p. 1135-1146.
- 184. Maranto, C., et al., Prospects for Clinical Development of Stat5 Inhibitor IST5-002: High Transcriptomic Specificity in Prostate Cancer and Low Toxicity In Vivo. Cancers (Basel), 2020. 12(11).
- 185. Liu, Y., et al., BD750, a benzothiazole derivative, inhibits T cell proliferation by affecting the JAK3/STAT5 signalling pathway. Br J Pharmacol, 2013. 168(3): p. 632-43.
- 186. Kruger, N.J. and A. von Schaewen, The oxidative pentose phosphate pathway: structure and organisation. Curr Opin Plant Biol, 2003. 6(3): p. 236-46.
- 187. Luzzatto, L., M. Ally, and R. Notaro, Glucose-6-phosphate dehydrogenase deficiency. Blood, 2020. 136(11): p. 1225-1240.
- 188. Song, J., et al., The Multiple Roles of Glucose-6-Phosphate Dehydrogenase in Tumorigenesis and Cancer Chemoresistance. Life (Basel), 2022. 12(2).
- 189. Xu, X., et al., Rewiring of purine metabolism in response to acidosis stress in glioma stem cells. Cell Death Dis, 2021. 12(3): p. 277.
- 190. Stanke, K.M., C. Wilson, and S. Kidambi, High Expression of Glycolytic Genes in Clinical Glioblastoma Patients Correlates With Lower Survival. Front Mol Biosci, 2021. 8: p. 752404.
- 191. Yang, C.A., et al., G6PD as a predictive marker for glioma risk, prognosis and chemosensitivity. J Neurooncol, 2018. 139(3): p. 661-670.
- 192. Sun, M., et al., PIKE-A promotes glioblastoma growth by driving PPP flux through increasing G6PD expression mediated by phosphorylation of STAT3. Biochem Pharmacol, 2021. 192: p. 114736.
- 193. Carmona, A. and R.A. Freedland, *Effect of 6-aminonicotinamide on pentose cycle activity in isolated* hepatocytes. Int J Biochem, 1990. 22(6): p. 595-9.
- 194. Scicinski, J., et al., Preclinical evaluation of the metabolism and disposition of RRx-001, a novel investigative anticancer agent. Drug Metab Dispos, 2012. 40(9): p. 1810-6.
- 195. Das, D.S., et al., A novel hypoxia-selective epigenetic agent RRx-001 triggers apoptosis and overcomes drug resistance in multiple myeloma cells. Leukemia, 2016. 30(11): p. 2187-2197.
- 196. Oronsky, B., T. Reid, and P. Cabrales, Vascular priming with RRx-001 to increase the uptake and accumulation of temozolomide and irinotecan in orthotopically implanted gliomas. J Drug Target, 2021. 29(9): p. 998-1003.
- 197. Oronsky, B., et al., Discovery of RRx-001, a Myc and CD47 Downregulating Small Molecule with Tumor Targeted Cytotoxicity and Healthy Tissue Cytoprotective Properties in Clinical Development. J Med Chem, 2021. 64(11): p. 7261-7271.
- 198. Mele, L., et al., A new inhibitor of glucose-6-phosphate dehydrogenase blocks pentose phosphate pathway and suppresses malignant proliferation and metastasis in vivo. Cell Death Dis, 2018. 9(5): p. 572.
- 199. Ghergurovich, J.M., et al., A small molecule G6PD inhibitor reveals immune dependence on pentose phosphate pathway. Nat Chem Biol, 2020. 16(7): p. 731-739.
- 200. Luo, Z., et al., Discovery and characterization of a novel glucose-6-phosphate dehydrogenase (G6PD) inhibitor via high-throughput screening. Bioorg Med Chem Lett, 2021. 40: p. 127905.
- 201. Lu, S.C., Glutathione synthesis. Biochim Biophys Acta, 2013. 1830(5): p. 3143-53.
- 202. Griffith, O.W., Biologic and pharmacologic regulation of mammalian glutathione synthesis. Free Radic Biol Med, 1999. 27(9-10): p. 922-35.
- 203. Bu, X., et al., CD147 confers temozolomide resistance of glioma cells via the regulation of beta-TrCP/Nrf2 pathway. Int J Biol Sci, 2021. 17(12): p. 3013-3023.
- 204. Landras, A., et al., CD147 Is a Promising Target of Tumor Progression and a Prognostic Biomarker. Cancers (Basel), 2019. 11(11).
- 205. Trautwein, C., et al., Tissue metabolites in diffuse glioma and their modulations by IDH1 mutation, histology, and treatment. JCI Insight, 2022. 7(3).
- 206. Fack, F., et al., Altered metabolic landscape in IDH-mutant gliomas affects phospholipid, energy, and oxidative stress pathways. EMBO Mol Med, 2017. 9(12): p. 1681-1695.
- 207. Griffith, O.W. and A. Meister, Potent and specific inhibition of glutathione synthesis by buthionine sulfoximine (S-n-butyl homocysteine sulfoximine). J Biol Chem, 1979. 254(16): p. 7558-60.
- 208. Backos, D.S., et al., Glycation of glutamate cysteine ligase by 2-deoxy-d-ribose and its potential impact on chemoresistance in glioblastoma. Neurochem Res, 2013. 38(9): p. 1838-49.
- 209. Ran, S., et al., NaAsO2 decreases GSH synthesis by inhibiting GCLC and induces apoptosis through Hela cell mitochondrial damage, mediating the activation of the NF-kappaB/miR-21 signaling pathway. Ecotoxicol Environ Saf, 2022. 234: p. 113380.
- 210. Villablanca, J.G., et al., A Phase I New Approaches to Neuroblastoma Therapy Study of Buthionine Sulfoximine and Melphalan With Autologous Stem Cells for Recurrent/Refractory High-Risk Neuroblastoma. Pediatr Blood Cancer, 2016. 63(8): p. 1349-56.
- 211. Anderson, C.P., et al., Pilot study of intravenous melphalan combined with continuous infusion L-S,R-buthionine sulfoximine for children with recurrent neuroblastoma. Pediatr Blood Cancer, 2015. 62(10): p. 1739-46.
- 212. Lee, S., S.M. Kim, and R.T. Lee, Thioredoxin and thioredoxin target proteins: from molecular mechanisms to functional significance. Antioxid Redox Signal, 2013. **18**(10): p. 1165-207.
- 213. Mohammadi, F., et al., The thioredoxin system and cancer therapy: a review. Cancer Chemother Pharmacol, 2019. 84(5): p. 925-935.
- 214. Erdi, F., et al., New Clues in the Malignant Progression of Glioblastoma: Can the Thioredoxin System Play a Role? Turk Neurosurg, 2018. 28(1): p. 7-12.
- 215. Yao, A., et al., Thioredoxin System Protein Expression Is Associated with Poor Clinical Outcome in Adult and Paediatric Gliomas and Medulloblastomas. Mol Neurobiol, 2020. 57(7): p. 2889-2901.
- 216. Kaya, B., et al., Intratumoral hemorrhage-related differences in the expression of vascular endothelial growth factor, basic fibroblast growth factor and thioredoxin reductase 1 in human glioblastoma. Mol Clin Oncol, 2016. 5(4): p. 343-346.
- 217. Branco, V., et al., Thioredoxin, Glutathione and Related Molecules in Tumors of the Nervous System. Curr Med Chem, 2020. 27(12): p. 1878-1900.
- 218. Jovanovic, M., et al., Novel TrxR1 Inhibitors Show Potential for Glioma Treatment by Suppressing the Invasion and Sensitizing Glioma Cells to Chemotherapy. Front Mol Biosci, 2020. 7: p. 586146.
- 219. Zhang, Y., et al., TIGAR knockdown radiosensitizes TrxR1-overexpressing glioma in vitro and in vivo via inhibiting Trx1 nuclear transport. Sci Rep, 2017. 7: p. 42928.
- 220. Wang, H., et al., Auranofin radiosensitizes tumor cells through targeting thioredoxin reductase and resulting overproduction of reactive oxygen species. Oncotarget, 2017. 8(22): p. 35728-35742.
- 221. Patwardhan, R.S., D. Sharma, and S.K. Sandur, Thioredoxin reductase: An emerging pharmacologic target for radiosensitization of cancer. Transl Oncol, 2022. 17: p. 101341.
- 222. Yao, A., et al., Cytotoxic and Radiosensitising Effects of a Novel Thioredoxin Reductase Inhibitor in Brain Cancers. Mol Neurobiol, 2022. 59(6): p. 3546-3563.
- 223. Sutton, B.M., et al., Oral gold. Antiarthritic properties of alkylphosphinegold coordination complexes. J Med Chem, 1972. 15(11): p. 1095-8.
- 224. Rigobello, M.P., et al., *Effect of auranofin on the mitochondrial generation of hydrogen peroxide.* Role of thioredoxin reductase. Free Radic Res, 2005. 39(7): p. 687-95.
- 225. Gandin, V., et al., Cancer cell death induced by phosphine gold(I) compounds targeting thioredoxin reductase. Biochem Pharmacol, 2010. 79(2): p. 90-101.
- 226. Zhang, X., et al., Repurposing of auranofin: Thioredoxin reductase remains a primary target of the drug. Biochimie, 2019. 162: p. 46-54.
- 227. Javvadi, P., et al., Thioredoxin reductase-1 mediates curcumin-induced radiosensitization of squamous carcinoma cells. Cancer Res, 2010. 70(5): p. 1941-50.
- 228. Kalin, S.N., A. Altay, and H. Budak, Diffractaic acid, a novel TrxR1 inhibitor, induces cytotoxicity, apoptosis, and antimigration in human breast cancer cells. Chem Biol Interact, 2022. 361: p. 109984.
- 229. Wang, L., et al., Ethaselen: a potent mammalian thioredoxin reductase 1 inhibitor and novel organoselenium anticancer agent. Free Radic Biol Med, 2012. 52(5): p. 898-908.
- 230. Rice, K.P., et al., Thioredoxin reductase is inhibited by the carbamoylating activity of the anticancer sulfonylhydrazine drug laromustine. Mol Cell Biochem, 2012. 370(1-2): p. 199-207.
- 231. Lu, J., et al., Inhibition of Mammalian thioredoxin reductase by some flavonoids: implications for myricetin and quercetin anticancer activity. Cancer Res, 2006, 66(8); p. 4410-8.
- 232. Zhang, Q., et al., Piperlongumine, a Novel TrxR1 Inhibitor, Induces Apoptosis in Hepatocellular Carcinoma Cells by ROS-Mediated ER Stress. Front Pharmacol, 2019. 10: p. 1180.
- 233. Zhang, J., et al., Inhibition of Thioredoxin Reductase by Santamarine Conferring Anticancer Effect in HeLa Cells. Front Mol Biosci, 2021. 8: p. 710676.
- 234. Stafford, W.C., et al., Irreversible inhibition of cytosolic thioredoxin reductase 1 as a mechanistic basis for anticancer therapy. Sci Transl Med, 2018. 10(428).
- 235. Dos Santos, D.C., et al., IP-Se-06, a Selenylated Imidazo[1,2-a]pyridine, Modulates Intracellular Redox State and Causes Akt/mTOR/HIF-1alpha and MAPK Signaling Inhibition, Promoting Antiproliferative Effect and Apoptosis in Glioblastoma Cells. Oxid Med Cell Longev, 2022. 2022: p. 3710449.
- 236. Arner, E.S. and A. Holmgren, The thioredoxin system in cancer. Semin Cancer Biol, 2006. 16(6): p. 420-6.
- 237. Karlenius, T.C. and K.F. Tonissen, Thioredoxin and Cancer: A Role for Thioredoxin in all States of Tumor Oxygenation. Cancers (Basel), 2010. 2(2): p. 209-32.
- 238. Tian, Y., et al., Diallyl trisulfide sensitizes radiation therapy on glioblastoma through directly targeting thioredoxin 1. Free Radic Biol Med, 2022. 189: p. 157-168.
- 239. Lu, J., E.H. Chew, and A. Holmgren, Targeting thioredoxin reductase is a basis for cancer therapy by arsenic trioxide. Proc Natl Acad Sci U S A, 2007. 104(30): p. 12288-93.
- 240. Welsh, S.J., et al., The thioredoxin redox inhibitors 1-methylpropyl 2-imidazolyl disulfide and pleurotin inhibit hypoxia-induced factor 1alpha and vascular endothelial growth factor formation. Mol Cancer Ther, 2003. 2(3): p. 235-43.
- 241. Ramanathan, R.K., et al., A randomized phase II study of PX-12, an inhibitor of thioredoxin in patients with advanced cancer of the pancreas following progression after a gemcitabine-containing combination. Cancer Chemother Pharmacol, 2011. 67(3): p. 503-9.
- 242. Ramanathan, R.K., et al., A phase I trial of PX-12, a small-molecule inhibitor of thioredoxin-1. administered as a 72-hour infusion every 21 days in patients with advanced cancers refractory to standard therapy. Invest New Drugs, 2012. 30(4): p. 1591-6.
- 243. Baker, A.F., et al., A phase IB trial of 24-hour intravenous PX-12, a thioredoxin-1 inhibitor, in patients with advanced gastrointestinal cancers. Invest New Drugs, 2013, 31(3): p. 631-641.
- 244. Padavannil, A., et al., The Mysterious Multitude: Structural Perspective on the Accessory Subunits of Respiratory Complex I. Front Mol Biosci, 2021. 8: p. 798353.
- 245. Pan, Y.B., et al., Transcriptome analyses reveal molecular mechanisms underlying phenotypic differences among transcriptional subtypes of glioblastoma. J Cell Mol Med, 2020. 24(7): p. 3901-3916.
- 246. Bachvarov, D., et al., Gene expression patterns of chemoresistant and chemosensitive serous epithelial ovarian tumors with possible predictive value in response to initial chemotherapy. Int J Oncol, 2006. 29(4): p. 919-33.
- 247. Chang, Z., X. Miao, and W. Zhao, Identification of Prognostic Dosage-Sensitive Genes in Colorectal Cancer Based on Multi-Omics. Front Genet, 2019. 10: p. 1310.
- 248. Sotgia, F. and M.P. Lisanti, Mitochondrial biomarkers predict tumor progression and poor overall survival in gastric cancers: Companion diagnostics for personalized medicine. Oncotarget, 2017. 8(40): p. 67117-67128.
- 249. Sterne, J., [Treatment of diabetes mellitus with N,N-dimethylguanylguanidine (LA. 6023, glucophage)]. Therapie, 1959. 14: p. 625-30.
- 250. El-Mir, M.Y., et al., Dimethylbiguanide inhibits cell respiration via an indirect effect targeted on the respiratory chain complex I. J Biol Chem, 2000. 275(1): p. 223-8.
- 251. OWEN, M.R., E. DORAN, and A.P. HALESTRAP, Evidence that metformin exerts its anti-diabetic effects through inhibition of complex 1 of the mitochondrial respiratory chain. Biochemical Journal, 2000. 348(3): p. 607-614.
- 252. Fontaine, E., Metformin-Induced Mitochondrial Complex I Inhibition: Facts, Uncertainties, and Consequences. Front Endocrinol (Lausanne), 2018. 9: p. 753.
- 253. Kawada, M., et al., Intervenolin, a new antitumor compound with anti-Helicobacter pylori activity, from Nocardia sp. ML96-86F2. J Antibiot (Tokyo), 2013. 66(9): p. 543-8.
- 254. Yoshida, J., et al., Mitochondrial complex I inhibitors suppress tumor growth through concomitant acidification of the intra- and extracellular environment. iScience, 2021. 24(12): p. 103497.
- 255. Shi, Y., et al., Gboxin is an oxidative phosphorylation inhibitor that targets glioblastoma. Nature, 2019. 567(7748): p. 341-346.
- 256. Zampieri, L.X., et al., Olaparib Is a Mitochondrial Complex I Inhibitor That Kills Temozolomide-Resistant Human Glioblastoma Cells. Int J Mol Sci, 2021. 22(21).
- 257. Luna Yolba, R., et al., EVT-701 is a novel selective and safe mitochondrial complex 1 inhibitor with potent anti-tumor activity in models of solid cancers. Pharmacol Res Perspect, 2021. 9(5): p. e00854.
- 258. Pant, S., et al., A first-in-human dose-escalation study of ME-143, a second generation NADH oxidase inhibitor, in patients with advanced solid tumors. Invest New Drugs, 2014. 32(1): p. 87-93.
- 259. Lim, S.C., K.T. Carey, and M. McKenzie, Anti-cancer analogues ME-143 and ME-344 exert toxicity by directly inhibiting mitochondrial NADH: ubiquinone oxidoreductase (Complex I). Am J Cancer Res, 2015. 5(2): p. 689-701.
- 260. Bendell, J.C., et al., Phase 1, open-label, dose escalation, safety, and pharmacokinetics study of ME-344 as a single agent in patients with refractory solid tumors. Cancer, 2015. **121(7): p. 1056-63.**
- 261. Molina, J.R., et al., An inhibitor of oxidative phosphorylation exploits cancer vulnerability. Nat Med, 2018. 24(7): p. 1036-1046.
- 262. Helbig, L., et al., BAY 87-2243, a novel inhibitor of hypoxia-induced gene activation, improves local tumor control after fractionated irradiation in a schedule-dependent manner in head and neck human xenografts. Radiat Oncol, 2014. 9: p. 207.
- 263. Schockel, L., et al., Targeting mitochondrial complex I using BAY 87-2243 reduces melanoma tumor growth. Cancer Metab, 2015. 3: p. 11.
- 264. Degli Esposti, M., Inhibitors of NADH-ubiquinone reductase: an overview. Biochim Biophys Acta, 1998. 1364(2): p. 222-35.
- 265. Ashton, T.M., et al., Oxidative Phosphorylation as an Emerging Target in Cancer Therapy. Clin Cancer Res, 2018. 24(11): p. 2482-2490.
- 266. Pombo, C.M., et al., MST Kinases and Metabolism. Endocrinology, 2019. 160(5): p. 1111-1118.
- 267. Mahlapuu, M., et al., GCKIII kinases in lipotoxicity: Roles in NAFLD and beyond. Hepatol Commun, 2022.
- 268. Lim, S., et al., Identification of the kinase STK25 as an upstream activator of LATS signaling. Nat Commun, 2019. 10(1): p. 1547.
- 269. Wu, F., et al., STK25-induced inhibition of aerobic glycolysis via GOLPH3-mTOR pathway suppresses cell proliferation in colorectal cancer. J Exp Clin Cancer Res, 2018. 37(1): p. 144.
- 270. Zhang, Y., et al., STK25 enhances hepatocellular carcinoma progression through the STRN/AMPK/ACC1 pathway. Cancer Cell Int, 2022. 22(1): p. 4.
- 271. Kurhe, Y., et al., Antagonizing STK25 Signaling Suppresses the Development of Hepatocellular Carcinoma Through Targeting Metabolic, Inflammatory, and Pro-Oncogenic Pathways. Cell Mol Gastroenterol Hepatol, 2022. 13(2): p. 405-423.
- 272. Zhu, Z., et al., Glutathione reductase mediates drug resistance in glioblastoma cells by regulating redox homeostasis. J Neurochem, 2018. 144(1): p. 93-104.
- 273. McLoughlin, M.R., et al., TrxR1, Gsr, and oxidative stress determine hepatocellular carcinoma malignancy. Proc Natl Acad Sci U S A, 2019. 116(23): p. 11408-11417.
- 274. Baity, M., et al., Glutathione reductase (GSR) gene deletion and chromosome 8 aneuploidy in primary lung cancers detected by fluorescence in situ hybridization. Am J Cancer Res, 2019. 9(6): p. 1201-1211.
- 275. Gopi, L.K. and B.L. Kidder, Integrative pan cancer analysis reveals epigenomic variation in cancer type and cell specific chromatin domains. Nat Commun, 2021. 12(1): p. 1419.
- 276. Chung, P.M., R.E. Cappel, and H.F. Gilbert, Inhibition of glutathione disulfide reductase by glutathione. Arch Biochem Biophys, 1991. 288(1): p. 48-53.
- 277. Seefeldt, T., et al., Characterization of a novel dithiocarbamate glutathione reductase inhibitor and its use as a tool to modulate intracellular glutathione. J Biol Chem, 2009. 284(5): p. 2729-2737.
- 278. Johnson, F.D., et al., Characterization of a small molecule inhibitor of disulfide reductases that induces oxidative stress and lethality in lung cancer cells. Cell Rep, 2022. 38(6): p. 110343.
- 279. Cole, S.P., Targeting multidrug resistance protein 1 (MRP1, ABCC1): past, present, and future. Annu Rev Pharmacol Toxicol, 2014. 54: p. 95-117.
- 280. Dean, M., K. Moitra, and R. Allikmets. The human ATP-binding cassette (ABC) transporter superfamily. Hum Mutat, 2022. 43(9): p. 1162-1182.
- 281. Kunicka, T. and P. Soucek, Importance of ABCC1 for cancer therapy and prognosis. Drug Metab Rev, 2014. 46(3): p. 325-42.
- 282. Xiao, H., et al., Clinically-Relevant ABC Transporter for Anti-Cancer Drug Resistance. Front Pharmacol, 2021. 12: p. 648407.
- 283. Li, Y. and X. Gao, LINC00883 Promotes Drug Resistance of Glioma Through a microRNA-136/NEK1-Dependent Mechanism. Front Oncol, 2021. 11: p. 692265.
- 284. Mahinfar, P., et al., The Role of microRNAs in Multidrug Resistance of Glioblastoma. Cancers (Basel), 2022. 14(13).
- 285. Wu, C., et al., LINC00470 promotes tumour proliferation and invasion, and attenuates chemosensitivity through the LINC00470/miR-134/Myc/ABCC1 axis in glioma. J Cell Mol Med, 2020. 24(20): p. 12094-12106.
- 286. Li, Y., et al., miR-1268a regulates ABCC1 expression to mediate temozolomide resistance in glioblastoma. J Neurooncol, 2018. 138(3): p. 499-508.
- 287. Chen, X.R., Y.G. Zhang, and Q. Wang, miR-9-5p Mediates ABCC1 to Elevate the Sensitivity of Glioma Cells to Temozolomide. Front Oncol, 2021. 11: p. 661653.
- 288. Jones, T.R., et al., Pharmacology of L-660,711 (MK-571): a novel potent and selective leukotriene D4 receptor antagonist. Can J Physiol Pharmacol, 1989. 67(1): p. 17-28.
- 289. Burkhart, C.A., et al., Small-molecule multidrug resistance-associated protein 1 inhibitor reversan increases the therapeutic index of chemotherapy in mouse models of neuroblastoma. Cancer Res, 2009. 69(16): p. 6573-80.
- 290. Silbermann, K., et al., Identification of Thienopyrimidine Scaffold as an Inhibitor of the ABC Transport Protein ABCC1 (MRP1) and Related Transporters Using a Combined Virtual Screening Approach. J Med Chem, 2019. 62(9): p. 4383-4400.
- 291. Hirano, Y., et al., A heterodimeric complex that promotes the assembly of mammalian 20S proteasomes. Nature, 2005. 437(7063): p. 1381-5.
- 292. Voutsadakis, I.A., Proteasome expression and activity in cancer and cancer stem cells. Tumour Biol, 2017. 39(3): p. 1010428317692248.
- 293. Arlt, A., et al., Increased proteasome subunit protein expression and proteasome activity in colon cancer relate to an enhanced activation of nuclear factor E2-related factor 2 (Nrf2). Oncogene, 2009. 28(45): p. 3983-96.
- 294. Lee, D., et al., miR-484 is associated with disease recurrence and promotes migration in prostate cancer. Biosci Rep, 2020. 40(5).
- 295. Li, K. and T. Liu, Evaluation of Oncogene NUP37 as a Potential Novel Biomarker in Breast Cancer. Front Oncol, 2021. 11: p. 669655.
- 296. Alshabi, A.M., et al., Identification of Crucial Candidate Genes and Pathways in Glioblastoma Multiform by Bioinformatics Analysis. Biomolecules, 2019. 9(5).
- 297. Tanaka, K., The proteasome: overview of structure and functions. Proc Jpn Acad Ser B Phys Biol Sci, 2009. 85(1): p. 12-36.
- 298. Yongjun Zhang, M.M., et al., Association between TGM5, PPAP2B and PSMA4 polymorphisms and NSCLC in never-smoking Chinese population. J Cancer Res Ther, 2013. 9(4): p. 660-3.
- 299. Guo, M., et al., The Study on the Clinical Phenotype and Function of HPRT1 Gene. Child Neurol Open, 2022. 9: p. 2329048X221108821.
- 300. Townsend, M.H., R.A. Robison, and K.L. O'Neill, A review of HPRT and its emerging role in cancer. Med Oncol, 2018. 35(6): p. 89.
- 301. M, J.S., et al., Hypoxanthine Phosphoribosyl Transferase 1 Is Upregulated, Predicts Clinical Outcome and Controls Gene Expression in Breast Cancer. Cancers (Basel), 2020. 12(6).
- 302. Ahmadi, M., et al., Overexpression of HPRT1 is associated with poor prognosis in head and neck squamous cell carcinoma. FEBS Open Bio, 2021. 11(9): p. 2525-2540.
- 303. Wu, T., et al., HPRT1 Promotes Chemoresistance in Oral Squamous Cell Carcinoma via Activating MMP1/PI3K/Akt Signaling Pathway. Cancers (Basel), 2022. 14(4).
- 304. Lane, A.N. and T.W. Fan, Regulation of mammalian nucleotide metabolism and biosynthesis. Nucleic Acids Res, 2015. 43(4): p. 2466-85.
- 305. Sun, H., et al., RRM2 is a potential prognostic biomarker with functional significance in glioma. Int J Biol Sci, 2019. 15(3): p. 533-543.
- 306. Rasmussen, R.D., et al., BRCA1-regulated RRM2 expression protects glioblastoma cells from endogenous replication stress and promotes tumorigenicity. Nat Commun, 2016. 7: p. 13398.
- 307. Jiang, H., et al., RRM2 Mediates the Anti-Tumor Effect of the Natural Product Pectolinarigenin on Glioblastoma Through Promoting CDK1 Protein Degradation by Increasing Autophagic Flux. Front Oncol, 2022. 12: p. 887294.
- 308. Li, C., et al., RRM2 promotes the progression of human glioblastoma. J Cell Physiol, 2018. 233(10): p. 6759-6767.
- 309. Li, J., et al., Suppression of RRM2 inhibits cell proliferation, causes cell cycle arrest and promotes the apoptosis of human neuroblastoma cells and in human neuroblastoma RRM2 is suppressed following chemotherapy. Oncol Rep, 2018. 40(1): p. 355-360.
- 310. Shi, S.C., Y. Zhang, and T. Wang, High RRM2 expression has poor prognosis in specific types of breast cancer. PLoS One, 2022. 17(3): p. e0265195.
- 311. Zhuang, S., et al., RRM2 elicits the metastatic potential of breast cancer cells by regulating cell invasion, migration and VEGF expression via the PI3K/AKT signaling. Oncol Lett, 2020. 19(4): p. 3349-3355.
- 312. Zhan, Y., et al., Inhibiting RRM2 to enhance the anticancer activity of chemotherapy. Biomed Pharmacother, 2021. 133: p. 110996.
- 313. Jiang, X., et al., RRM2 silencing suppresses malignant phenotype and enhances radiosensitivity via activating cGAS/STING signaling pathway in lung adenocarcinoma. Cell Biosci, 2021. 11(1): p. 74.
- 314. Abdel-Rahman, M.A., M. Mahfouz, and H.O. Habashy, RRM2 expression in different molecular subtypes of breast cancer and its prognostic significance. Diagn Pathol, 2022. 17(1): p. 1.
- 315. Ohmura, S., et al., Translational evidence for RRM2 as a prognostic biomarker and therapeutic target in Ewing sarcoma. Mol Cancer, 2021. 20(1): p. 97.
- 316. Zhou, B., et al., A small-molecule blocking ribonucleotide reductase holoenzyme formation inhibits cancer cell growth and overcomes drug resistance. Cancer Res, 2013. 73(21): p. 6484-93.
- 317. Liu, X., et al., Inhibition of hepatitis B virus replication by targeting ribonucleotide reductase M2 protein. Biochem Pharmacol, 2016. 103: p. 118-28.
- 318. Xie, Y., et al., Preclinical validation and phase I trial of 4-hydroxysalicylanilide, targeting ribonucleotide reductase mediated dNTP synthesis in multiple myeloma. J Biomed Sci, 2022. 29(1): p. 32.
- 319. Koppula, P., L. Zhuang, and B. Gan, Cystine transporter SLC7A11/xCT in cancer: ferroptosis, nutrient dependency, and cancer therapy. Protein Cell, 2021. 12(8): p. 599-620.
- 320. Koppula, P., et al., Amino acid transporter SLC7A11/xCT at the crossroads of regulating redox homeostasis and nutrient dependency of cancer. Cancer Commun (Lond), 2018. 38(1): p. 12.
- 321. Liu, J., X. Xia, and P. Huang, xCT: A Critical Molecule That Links Cancer Metabolism to Redox Signaling. Mol Ther, 2020. 28(11): p. 2358-2366.
- 322. Robert, S.M., et al., SLC7A11 expression is associated with seizures and predicts poor survival in patients with malignant glioma. Sci Transl Med, 2015. 7(289): p. 289ra86.
- 323. Polewski, M.D., et al., SLC7A11 Overexpression in Glioblastoma Is Associated with Increased Cancer Stem Cell-Like Properties. Stem Cells Dev, 2017. 26(17): p. 1236-1246.
- 324. Jyotsana, N., K.T. Ta, and K.E. DelGiorno, The Role of Cystine/Glutamate Antiporter SLC7A11/xCT in the Pathophysiology of Cancer. Front Oncol, 2022. 12: p. 858462.
- 325. Dixon, S.J., et al., Pharmacological inhibition of cystine-glutamate exchange induces endoplasmic reticulum stress and ferroptosis. Elife, 2014. 3: p. e02523.
- 326. Jin, C., et al., Inhibition of SLC7A11 by Sulfasalazine Enhances Osteogenic Differentiation of Mesenchymal Stem Cells by Modulating BMP2/4 Expression and Suppresses Bone Loss in Ovariectomized Mice. J Bone Miner Res, 2017. 32(3): p. 508-521.
- 327. Zhang, Y., et al., Imidazole Ketone Erastin Induces Ferroptosis and Slows Tumor Growth in a Mouse Lymphoma Model. Cell Chem Biol, 2019. 26(5): p. 623-633 e9.
- 328. Hu, K., et al., Suppression of the SLC7A11/glutathione axis causes synthetic lethality in KRASmutant lung adenocarcinoma. J Clin Invest, 2020. 130(4): p. 1752-1766.
- 329. Scalise, M., et al., The Human SLC1A5 (ASCT2) Amino Acid Transporter: From Function to Structure and Role in Cell Biology. Front Cell Dev Biol, 2018. 6: p. 96.
- 330. Cormerais, Y., et al., The glutamine transporter ASCT2 (SLC1A5) promotes tumor growth independently of the amino acid transporter LAT1 (SLC7A5). J Biol Chem, 2018. 293(8): p. 2877-2887.
- 331. Luo, Y., et al., ASCT2 overexpression is associated with poor survival of OSCC patients and ASCT2 knockdown inhibited growth of glutamine-addicted OSCC cells. Cancer Med, 2020. 9(10): p. 3489- 3499.
- 332. Zhang, Z., et al., ASCT2 (SLC1A5)-dependent glutamine uptake is involved in the progression of head and neck squamous cell carcinoma. Br J Cancer, 2020. 122(1): p. 82-93.
- 333. Zhang, H., et al., Comprehensive molecular and clinical characterization of SLC1A5 in human cancers. Pathol Res Pract, 2021. 224: p. 153525.
- 334. Zhu, D., et al., Ferroptosis-related gene SLC1A5 is a novel prognostic biomarker and correlates with immune infiltrates in stomach adenocarcinoma. Cancer Cell Int, 2022. 22(1): p. 124.
- 335. Zhao, J., et al., Correlation Between Prognostic Biomarker SLC1A5 and Immune Infiltrates in Various Types of Cancers Including Hepatocellular Carcinoma. Front Oncol, 2021. 11: p. 608641.
- 336. Yoo, H.C., et al., A Variant of SLC1A5 Is a Mitochondrial Glutamine Transporter for Metabolic Reprogramming in Cancer Cells. Cell Metab, 2020. 31(2): p. 267-283 e12.
- 337. Alves, M.J.F., et al., The expression of the aminoacid transporters ASCT2 (SLC1A5) and LAT1 (SLC7A5) in astrocytomas. Medical Express, 2016. 3(6).
- 338. Sidoryk, M., et al., Increased expression of a glutamine transporter SNAT3 is a marker of malignant gliomas. Neuroreport, 2004. 15(4): p. 575-8.
- 339. Esslinger, C.S., K.A. Cybulski, and J.F. Rhoderick, Ngamma-aryl glutamine analogues as probes of the ASCT2 neutral amino acid transporter binding site. Bioorg Med Chem, 2005. 13(4): p. 1111-8.
- 340. Schulte, M.L., et al., Pharmacological blockade of ASCT2-dependent glutamine transport leads to antitumor efficacy in preclinical models. Nat Med, 2018. 24(2): p. 194-202.
- 341. Ndaru, E., et al., Novel alanine serine cysteine transporter 2 (ASCT2) inhibitors based on sulfonamide and sulfonic acid ester scaffolds. J Gen Physiol, 2019. 151(3): p. 357-368.
- 342. Garibsingh, R.A., et al., Rational design of ASCT2 inhibitors using an integrated experimentalcomputational approach. Proc Natl Acad Sci U S A, 2021. 118(37).