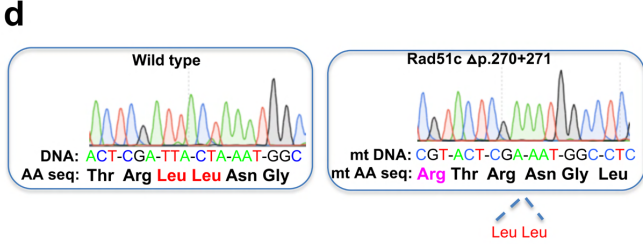
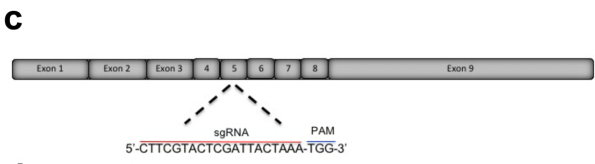
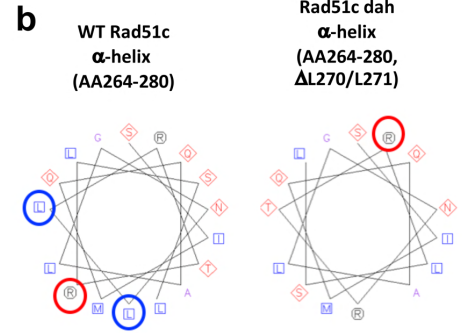


a You have run SYMPRED using the Dynamic Programming strategy with no weighting.

SEQUENCE NAME: NP_001278369.1DNArepairproteinRAD51homolog3isoform1[Musmusculus]
SEQUENCE LENGTH: 384

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Sequence	MLYRVHLAWLPSRLRPLFLFLCSLSGYIRNVTRTSETRRQPYMKPVCGISSAARPVQVIGSKEEALETL
PROF	EEEEEE HHHHHHHH HHHHHHHHHHHH EEEE HHH HHHHHHHH HH
SSPRO	EEEEH HHHHHH HHHHHHHHHHHH HHH HHHHHHHHHHHH
YASPIN	EEEEEEEE EEEEE HHH EEEEEEE HHHHHHHH HHHHHHHH
PSIPRED	EEEEEEEE HHHHHHHHHHHH HHHHHHHHHHHH HHH HH HHHHHHHH
SYMPRED	EEEEEE HHHHHH HHHHHHHHHHHHEE HHH HHH HHHHHHHHHH
Ruler81.....91.....101.....111.....121.....131.....
Sequence	QILRRECLTNKPRCAGTSVANEKCTALELLEQEHQTGGFIITFCSALDNILGGGIPLMKTTEVCGVPGVK
PROF	HHHHHHHHHHHHHHHHH HHHHHHHH EEEE HHHHHHHH HHHH
SSPRO	HHHHHHHHHHHHHHHHH HHHHHHHHHHHH HHHHHHHH EEEEE
YASPIN	HHHHHHHHHHHHHHHHH HHHHHHHH EEEE HHHHHHHH EEEEE HH
PSIPRED	HHHHHHHHH HHHHHHHH HHHHHHHHHH HHHHHHHH EEEEE HH
SYMPRED	HHHHHHHHHHHHHHHHH HHHHHHHH EEE HHHHHHHH EEEEE
Ruler151.....161.....171.....181.....191.....201.....
Sequence	TQLCMQLAVDVQIPECFGVGAGEAVFIDTEGSFMVDRVVSALATACIQHLHLIAGTHTHEEHQKALKDFTL
PROF	EEEEEEEEEEEE EEEEE HHHHHHHHHHHHHHHHHH HHHHHHHH H
SSPRO	EEEEEEEEEEEE H EEEEE HHHHHHHHHHHHHHHHHH HHHHHHHH H
YASPIN	HHHHHHHHHHHHH EEEEE HHHHHHHHHHHHHHHHHH HHHHHHHH H
PSIPRED	HHHHHHHHHHH HHHHHH EEEEE HHHHHHHHHHHHHHHH HHHHHHHH H
SYMPRED	HHHHHHHHHHHHH EEEEE HHHHHHHHHHHHHHHHHH HHHHHHHH H
Ruler221.....231.....241.....251.....261.....271.....
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PROF	HHHHHHHHHHH HHHHHHHHHHHHHH EEEEE HHHH HHHHHHHHHHHHHHHH
SSPRO	HHHHHHHHHHH HHHHHHHHHHHHHH EEEEEHHHHHHHHHHHHHHHHHHHHH
YASPIN	HHHH EEEEE HHHHHHHHHHHHHHHH EEEEE HHHHHHHH HHHHHHHHHHHHHH
PSIPRED	HHH EEEE HHHHHHHHHHHHHHHH EEEE HHHHHHHH HHHHHHHHHHHHHH
SYMPRED	HHHHHHHEEH HHHHHHHHHHHHHHHH EEEEE HHHHHHHH HHHHHHHHHHHHHH
Ruler291.....301.....311.....321.....331.....341.....
Sequence	LANNHRLAVILTNQMTTKIDKNQALLVPALGESWGHAAATIRLIFHWQKQRFATLYKSPSQKESTIPFQI
PROF	HHH EEEEE EEE HHHHHHEEEEE EEEEE EEEEE EEEEE
SSPRO	HHH EEEEE EEEE EEE HHEEEEE EEEEE EEEEE
YASPIN	HHH EEEEEEEEE EEE HHH EEEEE EEEEE EEEEE
PSIPRED	HHH EEEE EEE HHHHHHH EEEEE EEEEE EEEEE
SYMPRED	HHH EEEEEEEEE HHH EEEEE EEEEE EEEEE
Ruler361.....371.....381.....
Sequence	TPQGFRDAVVTAASSQTTESSLNFRKRSREPEEE
PROF	
SSPRO	H
YASPIN	HHH
PSIPRED	EEEE HHH
SYMPRED	



e

NP_001278369.1 DNA repair protein RAD51 homolog 3 isoform 1 [Mus musculus]
Sequence ID: Query_58491 Length: 384 Number of Matches: 1

Range 1: 58 to 383 [Graphics](#) [Next Match](#) [Previous](#)

Score	Expect	Method	Identities	Positives	Gaps
591 bits(1524)	0.0	Compositional matrix adj.	284/327(87%)	306/327(93%)	1/327(0%)
Query 49	EVGISKAELTQLIIRRECLTNKPRYAGTSESHKCTALELLEQEHQTGGFIITFCSALD	108			
Sbjct 58	+VGISK EALETQLI+RRECLTNKPR AGTS +++KCTALELLEQEHQTGGFIITFCSALD	117			
Query 109	DILGGVPLMKTTEICGAPGVGKTLQCMQLAVDVQIPECFGVGAGEAVFIDTEGSFMVDR	168			
Sbjct 118	+ILGGG+PLMKTTE+CG PGVGTQLCMQLAVDVQIPECFGVGAGEAVFIDTEGSFMVDR	177			
Query 169	VVDLATACIQHLIAEKHKKEEHRKALEDFLTDNLISHIYFRCRDYTELLAQVYLLPD	228			
Sbjct 178	VV LATACIQHL LIA H EEH+KAL+DFTL+NILSHIYFRC DYTELLAQVYLLPD	237			
Query 229	FLSEHSKVLVVDGIAFPFRHDLDLSLRLRLNGLAQQMISLANNHRLAVILTNQMTT	288			
Sbjct 238	FLS+H KV+LVI+DGIAPFRHDL+DLSLRLRLNGLAQQMISLANNHRLAVILTNQMTT	297			
Query 289	KIDRNQALLVPALGESWGHAAATIRLIFHWDRKORLATLYKSPSQKECTVLFQIKPQGRD	348			
Sbjct 298	KIDRNQALLVPALGESWGHAAATIRLIFHWDRKORLATLYKSPSQKECTVLFQIKPQGRD	357			
Query 349	TVVTSACSLQTEGSLSTRKRSRDPREE VVT+A S QTE SL+ RKRSR+PEEE	375			
Sbjct 358	AVVTAASS-QTESSLNFRKRSREPEEE 383				

Supplementary Fig.1. Generation of Rad51c mutant mice with 6bp deletion.

a, Schematic diagram of sgRNA guide and targeting sites in the mouse *Rad51c* locus.

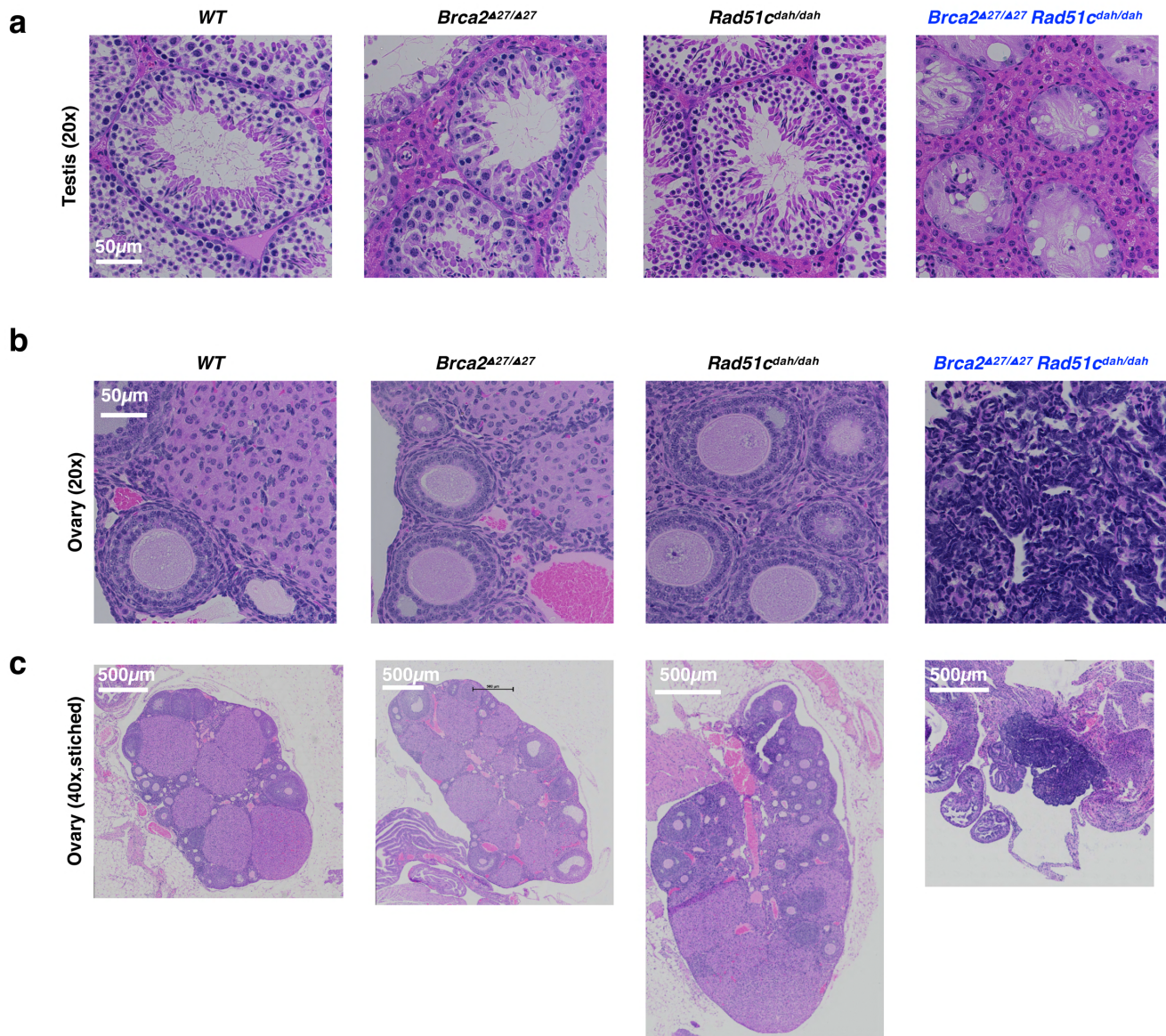
PAM = protospacer adjacent motif.

b, wheel projection of alpha helix amino acids 264-280 of murine Rad51c (left), and wheel projection of the same alpha helix in the Rad51c dah mutant missing L270/L271.

c, Sanger sequencing of PCR products encompassing the *Rad51c* targeting region from a representative wild-type (WT) mice and *Rad51c^{dah/dah}* mouse. The two leucine residues that are deleted in *Rad51c^{dah/dah}* mouse are marked in red, the adjacent Arginine that is substituted by a histidine in the FANCO patient is marked in magenta.

d, Blast sequence alignment of human (NP_478123.1) and murine RAD51C (NP_001278369) shows high similarity and evolutionary conservation.

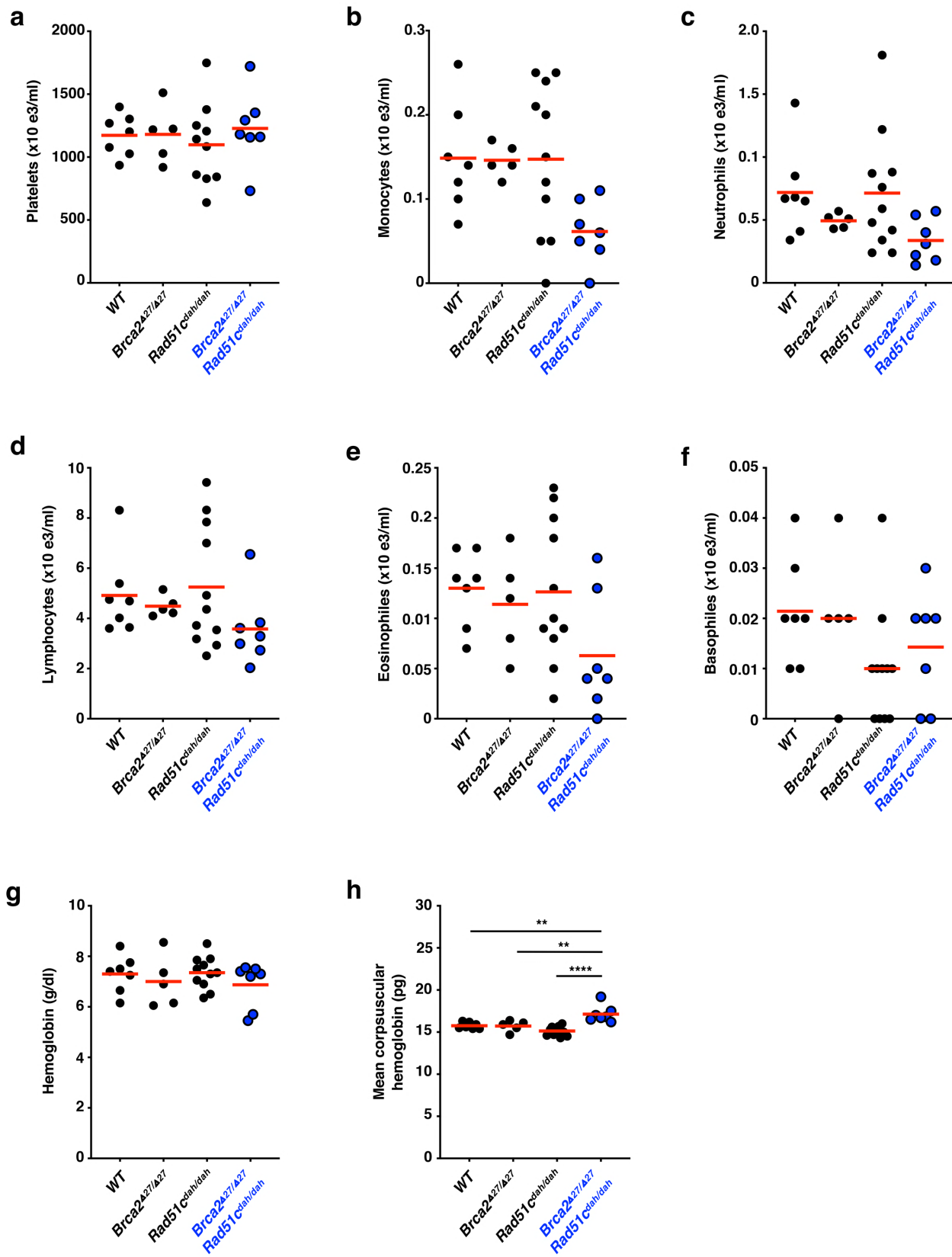
e, SympRED protein secondary structure prediction of RAD51C showing R258 (blue square, mutated to histidine in the FANCO patient) adjacent to the two leucines (red square) that are deleted in the mouse model within the same a-helix



Supplementary Fig.2. Polygenic mutant FANC mice show impaired gonad development.

a, Representative images of hematoxylin and eosin (H&E)-stained tissue cross-sections of testis from animals with indicated genotypes at 8-12 weeks. Seminiferous tubuli of *Brca2*^{Δ27/Δ27} mice can show decreased spermatocytes (arrow), while those of *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice show a Sertoli cell only phenotype with a complete loss in spermatocytes and spermatogonia. Scale bars indicate 50 μm. Similar results were obtained by analyzing three independent tissues of each genotype.

b-c, Representative images of H&E-stained tissue cross-sections of ovaries from animals with indicated genotypes at 12 weeks. Scale bars indicate 50 μm **b**, and 500 μm **c**. Similar results were obtained by analyzing three independent tissues of each genotype.



Supplementary Fig.3. Complete blood count analysis of *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice.

a-h, Complete blood count analysis of 8–12 weeks old *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} and control mice. Data represent biologically independent samples for wild-type (n=7), *Brca2* ^{$\Delta 27/\Delta 27$} (n=5), *Rad51c*^{*dah/dah*} (n=11, except a, platelet count only contain n=10) and *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} (n=7) mice.

a, Platelet count.

b, Monocyte count.

c, Neutrophils count.

d, Lymphocytes count.

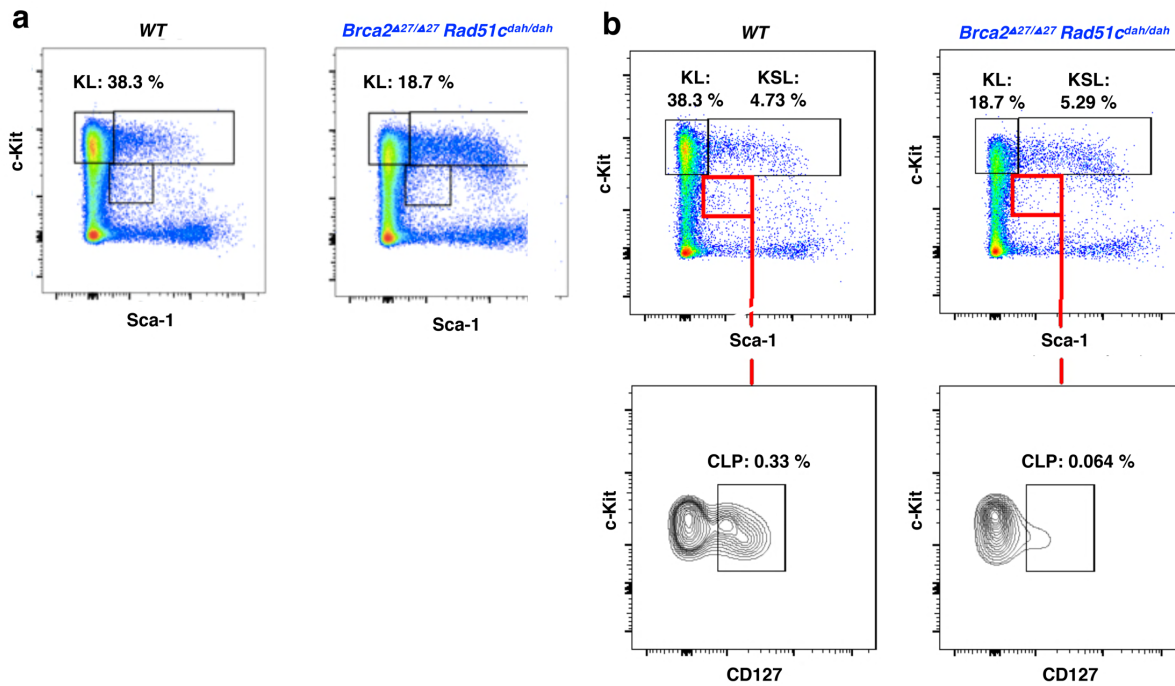
e, Eosinophil count.

f, Basophil count.

g, Hemoglobin concentrations.

h, Mean corpuscular hemoglobin concentration. *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} against WT: p=0.0026; *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} against *Brca2* ^{$\Delta 27/\Delta 27$} : p=0.0053; ; *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} against *Rad51c*^{*dah/dah*}: p<0.0001.

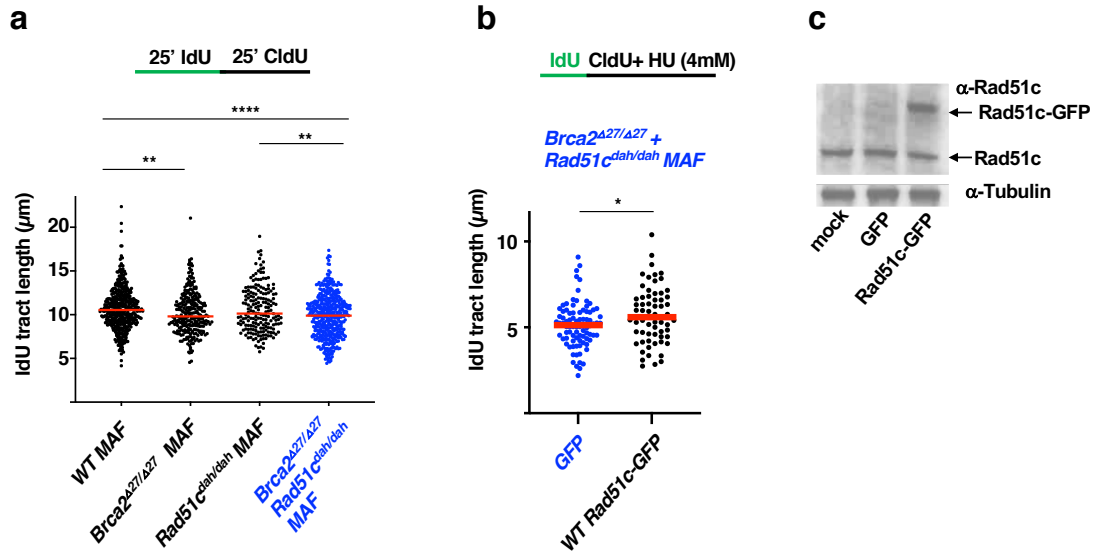
Red bar in scatter graphs represent the mean. *p*-values are derived using the one-way ANOVA test. ** *p* < 0.01, **** *p* < 0.0001.



Supplementary Fig.4. Hematological FACS analysis of *Brca2*^{Δ27/Δ27} +*Rad51*^{c^{dah/dah}} mice.

a, Representative FACS plot showing gating for hematological lineage analysis performed for bone marrow cells collected from 8-12 weeks old mice with indicated genotypes (n=5-6). Lineage negative cells (20,000 events) are gated for CD117 (c-Kit) positive and Sca-1 negative populations to identify the KL subset designating myeloid progenitor cells.

b, Representative FACS plot showing gating for identification of the CLP subset, designating common lymphoid progenitors. Lineage negative cells (20,000 events) are gated for c-Kit positive and Sca-1 moderate expression (KSL), which is further gated for CD127 positive cells to identify the CLP subpopulation designating the lymphoid population.



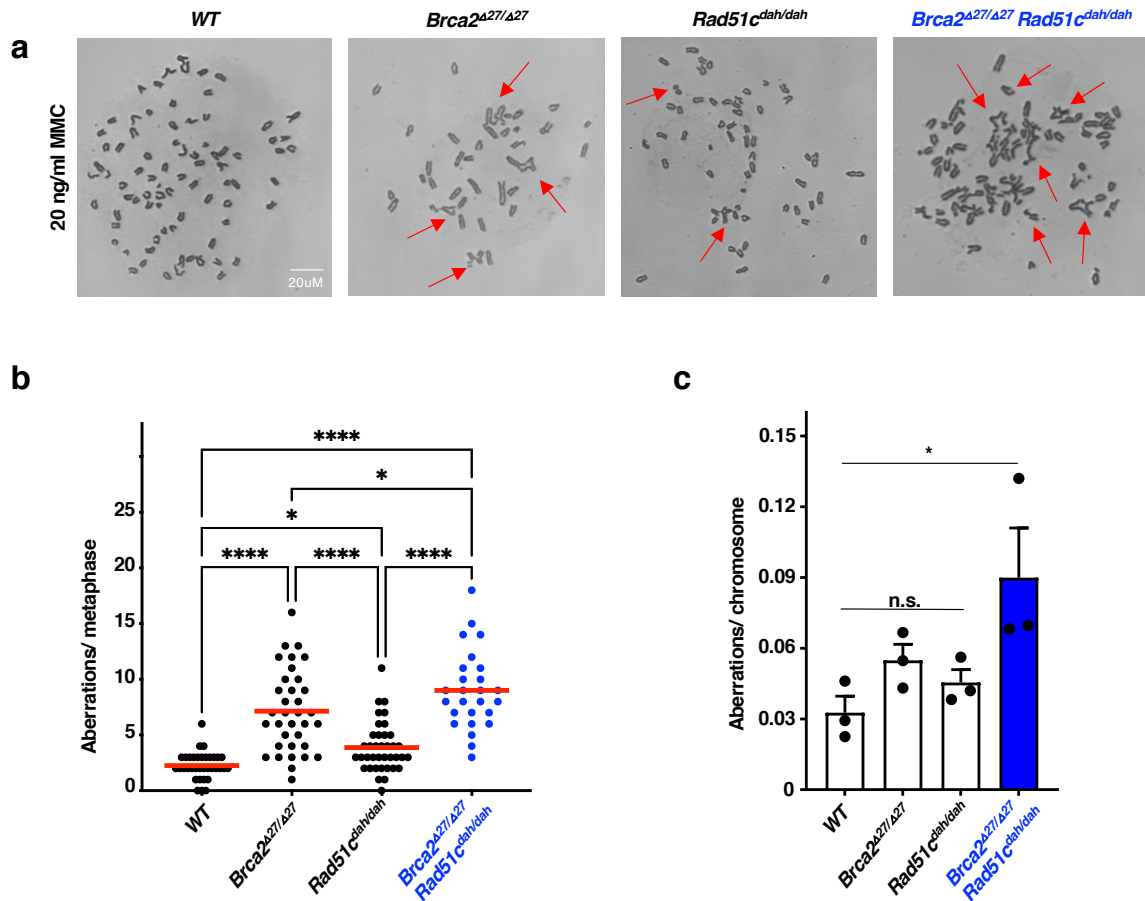
Extended Data Fig. 5. Cells with polygenic *FANC* mutations show reduced DNA replication speeds.

a, Scatter plot of DNA fiber analysis of nascent IdU tract lengths incorporated without external replication stalling, measuring DNA replication progression. Data represent biologically independent fiber experiments from wild-type ($n=5$), *Brca2* ^{$\Delta 27/\Delta 27$} ($n=4$), *Rad51c*^{*dah/dah*} ($n=2$) and *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} ($n=4$) mouse embryonic fibroblasts (MAF). In total, analysis was performed for wild-type ($n_{\text{fibers}}=469$), *Brca2* ^{$\Delta 27/\Delta 27$} ($n_{\text{fibers}}=270$), *Rad51c*^{*dah/dah*} ($n_{\text{fibers}}=190$) and *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} ($n_{\text{fibers}}=462$) fibers. *Brca2* ^{$\Delta 27/\Delta 27$} against WT : $p = 0.0019$, *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} against WT : $p < 0.0001$ and *Rad51c*^{*dah/dah*} against *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} : $p = 0.0023$.

b, Scatter plot of DNA fiber analysis of nascent IdU tract lengths before 4mM hydroxyurea in *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} MAFs transfected with empty GFP vector or wild-type *Rad51c*-GFP. Experiment was performed twice and representative fiber analysis for MAFs transfected with empty GFP vector ($n=86$) or wild-type *Rad51c*-GFP ($n=64$) with $p = 0.0244$ between the two groups.

c, Western blot of mock transfected, empty GFP vector transfected, and wild-type *Rad51c*-GFP transfected *Brca2* ^{$\Delta 27/\Delta 27$} + *Rad51c*^{*dah/dah*} MAFs. Experiment was performed once.

p -values for DNA fiber analysis are derived using the two-tailed Mann-Whitney test, * $p < 0.05$, *** $p < 0.001$, **** $p < 0.0001$, n.s. $p > 0.05$.

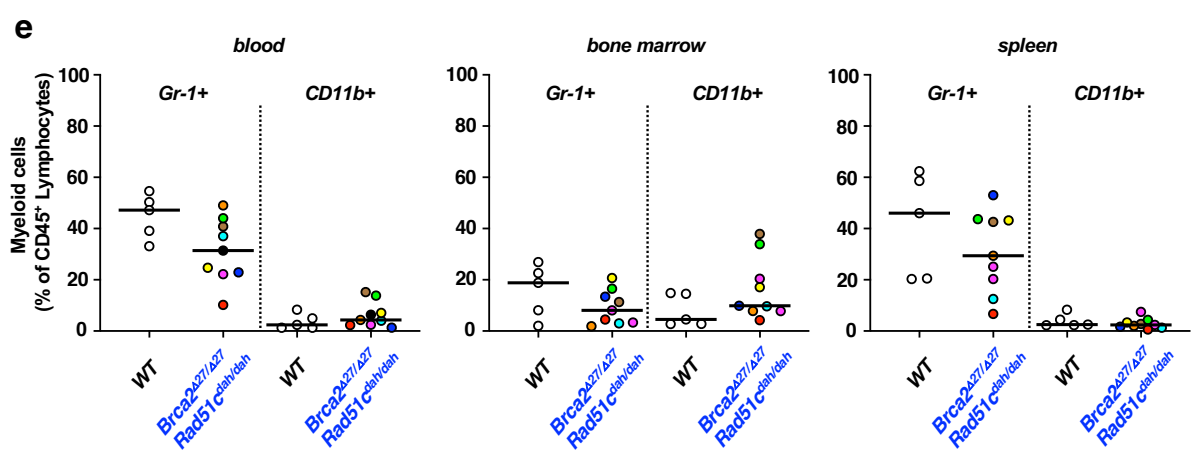
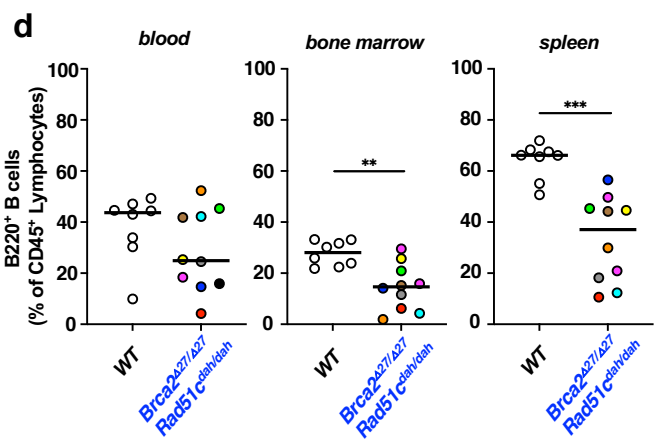
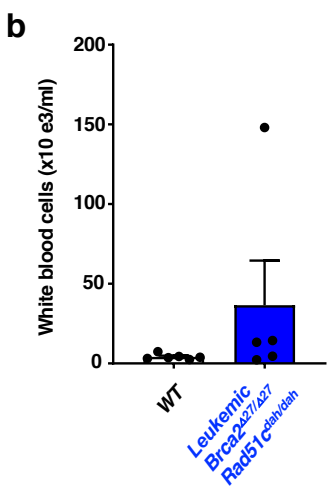
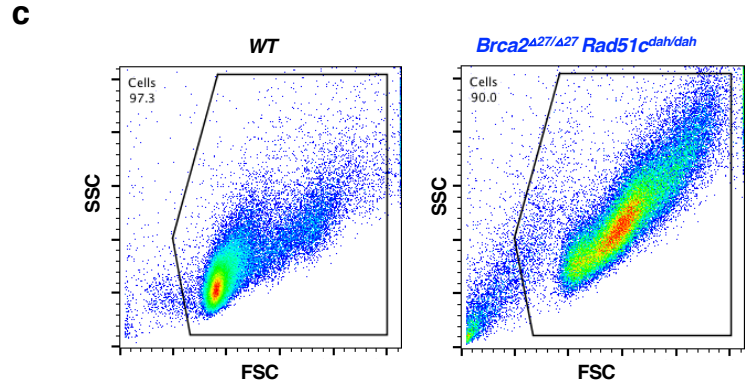
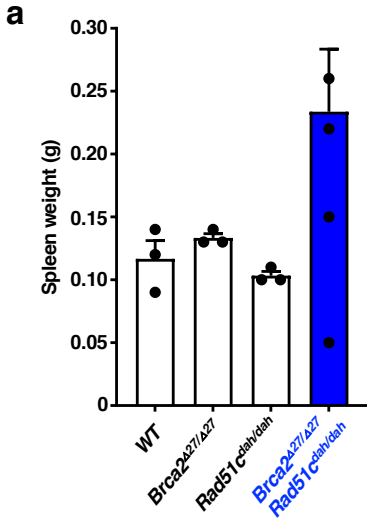


Supplementary Fig.6. Cells with polygenic FANC mutations show increased genomic aberrations.

a, Representative images of metaphase chromosome spreads in mouse adult fibroblasts (MAF) of indicated genotypes with 20 ng/ml mitomycin C (MMC). Examples for chromosomal aberrations are indicated with an arrow.

b, Scatter dot blot of chromosomal aberrations in metaphase spreads of MAFs after treatment with 20ng/ml MMC as indicated. Data represent 2 biologically independent experiments. In total, chromosome analysis was performed for 20ng/ml MMC treated wild-type (n=35), *Brca2*^{Δ27/Δ27} (n=35), *Rad51c*^{dah/dah} (n=38) and *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} (n=25) cells. *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} against WT and *Rad51c*^{dah/dah}: $p < 0.0001$; *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} against *Brca2*^{Δ27/Δ27}: $p = 0.0268$; *Brca2*^{Δ27/Δ27} against WT and *Rad51c*^{dah/dah}: $p < 0.0001$; *Rad51c*^{dah/dah} against WT: $p = 0.0403$

c, Bar graph of spontaneous chromosome aberrations per chromosome (without MMC) in metaphase spreads in MAFs. Data represent the average aberrations/chromosome of three biologically independent experiments. In total, analysis was performed for wild-type (n=50), *Brca2*^{Δ27/Δ27} (n=52), *Rad51c*^{dah/dah} (n=46) and *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} (n=53) cells. *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} against WT: $p = 0.0379$. Error bars represent the standard error of the mean. The data represent the mean of compiled data from biological repeats. p -values are derived using the one-way ANOVA test. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$



Supplementary Fig.7. *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice show abnormalities in tissues of the immune system.

a, Bar-graph of enlarged spleen quantification in malade *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} and age-matched control mice. Data represent biologically independent samples for wild-type (n=3), *Brca2*^{Δ27/Δ27} (n=3), *Rad51c*^{dah/dah} (n=3) and *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} (n=5) mice.

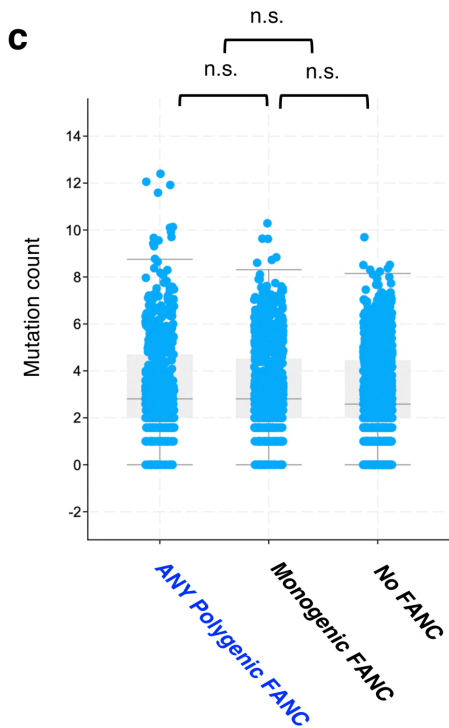
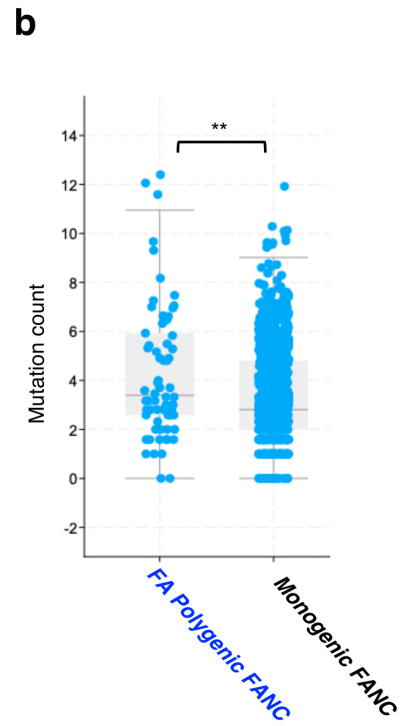
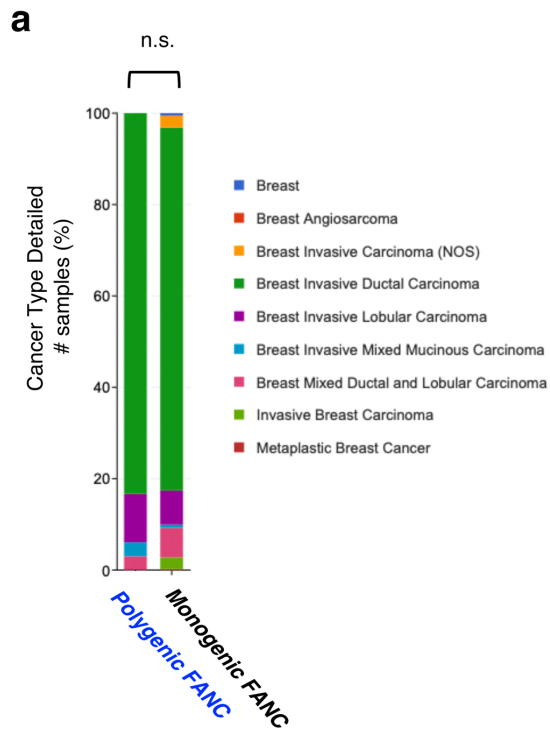
b, Representative forward and side-ward scatter FACS plot image of thymus cells from WT and malade *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice show enlargement of the mutant cells.

c, White blood cell count of leukemic *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice shows increased counts compared to age-matched wild-type (WT) mice. Data represent biologically independent blood samples from wild-type (n=7) and leukemic *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} (n=5) mice.

Error bars represent the standard error of the mean.

d, e, Flow cytometry analysis of blood, bone marrow and spleen cells from malade *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice identifying B-Cells (**d**, B220+) or a myeloide population (**e**, Gr-1 or CD11b) as percentage of total lymphocytes (CD45+). Data represent biologically independent samples for wild-type (n=8) and *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} (n=10) mice for identifying B-Cells (**d**, B220+). Data represent biologically independent samples for wild-type (n=5) and *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} (n=9) mice for identifying myeloide populations (**e**, Gr-1 or CD11b). *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} against WT for bone marrow: $p=0.0021$; and for spleen $p=0.0002$.

p -values for flow cytometry analysis are derived using the two-tailed Mann-Whitney test. * $p < 0.1$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

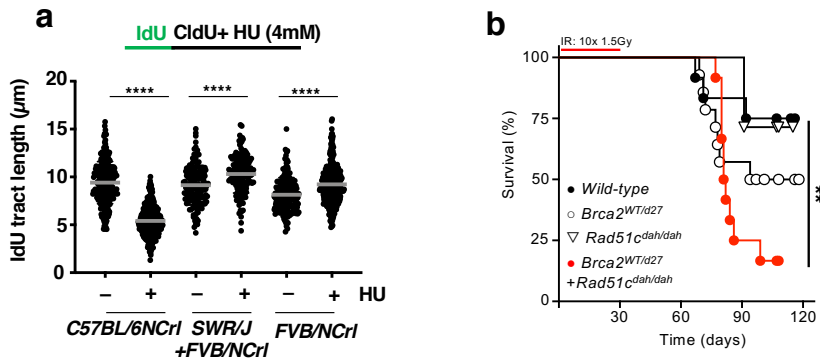


Supplementary Fig.8. Breast cancer patients with polygenic FANC tumor mutations show increased mutagenic count.

a, Bar-graph of cancer types of patients with polygenic and monogenic FANC tumor mutations ($p=0.262$). p -values were derived using the two-sided Chi-squared test.

b, Scatter plot of mutation counts of tumor samples from 3593 patients with polygenic and monogenic FANC tumor mutations, whereby only polygenic FANC mutation combinations were considered that are found in FA patients. Box centres indicate median, boundaries represent 25th and 75th percentiles, and error bars represent maximum and minimum values. p -values were derived using the two-sided Kruskal-Wallis test, $**p=0.014$.

c, Scatter plot of mutation counts of tumor samples from 3593 patients with polygenic and monogenic FANC tumor mutations, whereby any combination of two FANC tumor mutations in the 23 FANC genes were considered as polygenic FANC mutation. Box centres indicate median, boundaries represent 25th and 75th percentiles, and error bars represent maximum and minimum values. p -values were derived using the two-sided Kruskal-Wallis test.



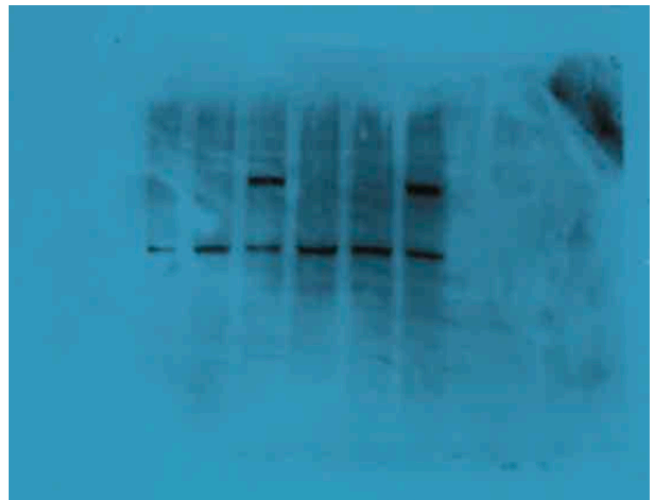
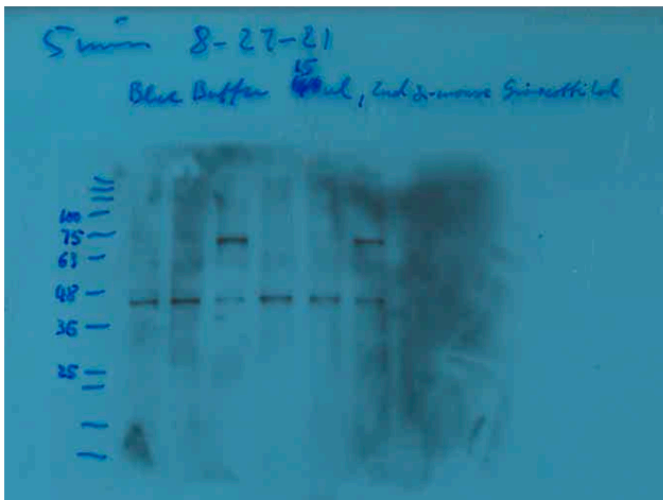
Supplementary Fig.9. Mouse strain backgrounds vary in intrinsic fork protection defects.

a, Scatter plot of DNA fiber analysis of nascent IdU tract lengths incorporated before replication stalling with high concentrations of hydroxyurea (HU, 4mM) and without, measuring DNA fork protection as the reduction of IdU tract lengths with HU in MAFs in various mouse strain backgrounds as indicated. Data represent two independent experiments. In total, fiber analysis was performed for untreated (n=245) and HU treated (282) C57BL/6NCrI MAFs, untreated (n=140) and HU treated (148) SWR/J+ FVB/NCrI mixed background MAFs and untreated (n=189) and HU treated (n=310) FVB/NCrI MAFs. *p*-values for DNA fiber analysis are derived using the two-tailed Mann-Whitney test, **** *p* < 0.0001 for all comparison of untreated and treated samples in each group.

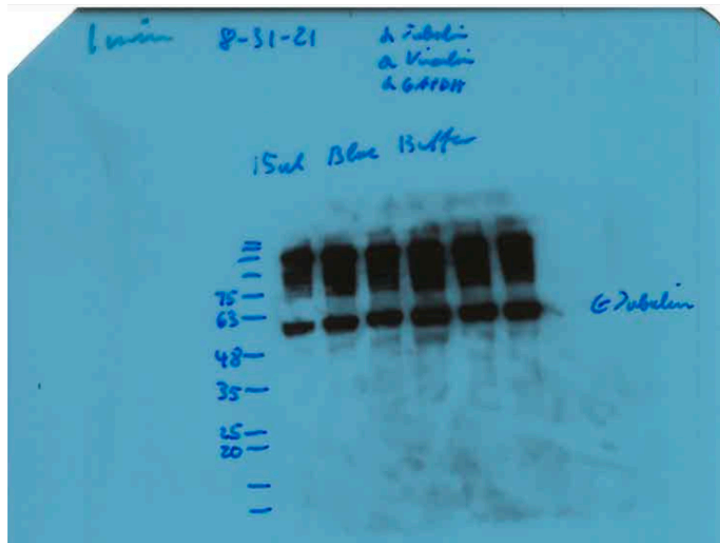
b, Kaplan-Meier curves for overall survival of male wild-type (WT), *Brca2*^{WT/Δ27}, *Rad51*^{dah/dah}, and *Brca2*^{WT/Δ27}+*Rad51*^{dah/dah} mice after ten irradiations with 1.5 Gy daily. *Brca2*^{WT/Δ27}+*Rad51*^{dah/dah} against WT: *p*=0.0091. *p*-values for comparing survival of two groups are derived using the Mantel-Cox test,

Uncropped Western blots of Supplementary Fig. 5c.

Anti-Rad51c blot: lane 1: protein marker, lane 2: mock transfected, lane 3: mock-GFP transfected, lane 4: wild-type Rad51c-GFP transfected *Brca2*^{Δ27/Δ27}+*Rad51c*^{dah/dah} MAFs. Higher exposure blot to the right.



Anti- α -tubulin blot: lane 1: protein marker, lane 2: mock transfected, lane 3: mock-GFP transfected, lane 4: wild-type Rad51c-GFP transfected *Brca2* ^{Δ 27/ Δ 27}+*Rad51c*^{*dah/dah*} MAFs.



Supplementary Tables and References

ID	FA Subtype	Additional FANC mutations	Reference
HSC230	FA-B	‡ <i>BRCA2/FANCD1</i> c.2805_2808delAAAC, p.(Ala938Profs*21)	(1)
EUFA867	FA-A	‡ <i>FANCM</i> c.2171C>A, p.(Ser724*); <i>FANCM</i> c.4222+1978_4300del, p.(fs*) [EX15del]	(2), (3)
FA4	FA-C	<i>BRCA2/FANCD1</i> c.951A>G, p.(N317S)	(4)
FA26	FA-L	<i>FANCI</i> c.3737C>T, p.(P1246L)	(4)
Fa-004	FA-D2	<i>BRCA2/FANCD1</i> chr13:31809499C>G (NCBI36/hg18), p.(H1003D)	(5)
Fa-001	FA-A	<i>FANCI</i> chr15:87636941A > G, missense I > V	(5)
Fa-002	FA-A	‡ <i>FANCB</i> chrX:14781120C > T, missense V > I; ‡ <i>BRCA2</i> chr13:31871012A > C, 3'-UTR	(5)
Fa-003	FA-M	‡ <i>BRCA2</i> chr13:31804480A > C, missense N > H	(5)

Supplementary Table 1. Previously reported *FANC* passenger mutations. Literature search of previously reported FA patients with germline *FANC* gene mutations in addition to a biallelic or hemizygous *FANC* inactivation. **‡** Denotes homozygous or biallelic additional *FANC* mutation.

Ser.#	Subtype	<i>FANC</i> passenger Mutation (non-synonymous)
1	FA-A	<i>FANCG</i> c.418C>T, p.(His140Tyr)
2	FA-F	<i>BRCA2/FANCD1</i> c.3854del, p.(Asn1287Ilefs*6)
3	FA-C	none
4	FA-A	none
5	FA-N	none
6	FA-C	none
7	FA-C	none
8	FA-A	none
9	FA-A	none
10	FA-A	none
11	FA-A	none
12	FA-N	none
13	FA-A	none
14	FA-G	none
15	FA-A	none
16	FA-A	none
17	FA-A	<i>SLX4/FANCP</i> c.734C>T, p.Pro245Leu,
18	FA-A	none
19	FA-A	<i>BRCAl/FANCS</i> c.301+7G>A
20	FA-Q	none
21	FA-A	none
22	FA-A	none
23	FA-A	none
24	FA-A	none
25	FA-A	none
26	FA-A	none
27	FA-D2	none
28	FA-C	none
29	FA-A	<i>FANCG</i> c.77A>G, p.Gln26Arg; <i>FANCI</i> c.1305G>A, p.(Met435Ile)
30	FA-G	none
31	FA-M	<i>FANCA</i> c.2151G>T, p.(Met717Ile); <i>RAD51C/FANCO</i> c.29T>G, p.(Met10Arg)
32	FA-B	<i>BRCA2/FANCD1</i> c.6725A>G, p.(Asp2242Gly); <i>BRCA2/FANCD1</i> c.9104A>C, p.(Tyr3035Ser)
33	FA-B	none
34	FA-A	<i>SLX4/FANCP</i> c.3334A>C, p.(Lys1112Gln)
35	FA-D2	none
36	FA-A	<i>FANCE</i> c.1099C>A, p.(Leu367Ile)

Supplementary Table 2. *FANC* passenger mutations in whole-exome sequences of 36 FA patients. Whole exome sequencing of FA patient DNAs with previously diagnosed biallelic or hemizygous *FANC* inactivation (subtype) was performed and filtered for additional heterozygous non-synonymous, ACMG class 3 or higher variants in any one of the so far identified 23 *FANC* genes. Only exomes that could be re-evaluated unambiguously were used.

Phenotype	<i>BRCA2</i> mutation	<i>RAD51C</i> mutation	Reference
*Fanconi Anemia	Hom. BRCA2 V2466A Het. BRCA2 N372H	Het. RAD51C G264S	(6) (7) (8)
Breast cancer susceptibility	Het. BRCA2 Δ ex26	Het. RAD51C R258H	(9) (10)

Supplementary Table 3. Polygenic *BRCA2+RAD51C* mutations are found in a Fanconi Anemia patient and families with a history of breast cancer. BRCA2 V2466A destabilizes the protein, BRCA2 N372H is a low-penetrance variant significantly associated with an increased risk of overall cancer, RAD51C G264S is a deleterious mutation predicted by SIFT and Polygen, BRCA2 Δ ex26 (L3216L) causes C-terminal internal deletions of exon 26, RAD51C R258H is a damaging mutation found in FA and breast cancer patients.

* The data/analyses presented in the current publication are based on the use of study data downloaded from the dbGaP web site, under phs001481.v1.p1, https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs001481.v1.p1

a

Parental genotype		Offspring genotype				
Father	Mother	<i>Brca2</i>	+/+	+/d27	d27/d27	Statistical Significance
		<i>Rad51c</i>	dah/dah	dah/dah	dah/dah	
<i>Brca2</i> ^{d27/+} <i>Rad51c</i> ^{dah/dah}	<i>Brca2</i> ^{d27/+} <i>Rad51c</i> ^{dah/dah}	Expected frequency	25.00%	50.00%	25.00%	p=0.0001, n=132
		Observed frequency	32.58%	56.82%	10.61%	

b

Parental genotype		Offspring genotype										
Father	Mother	<i>Brca2</i>	+/+	+/+	+/+	+/d27	+/d27	+/d27	d27/d27	d27/d27	d27/d27	Statistical Significance
		<i>Rad51c</i>	+/+	+/dah	dah/dah	+/+	+/dah	dah/dah	+/+	+/dah	dah/dah	
<i>Brca2</i> ^{d27/+} <i>Rad51c</i> ^{dah/+}	<i>Brca2</i> ^{d27/+} <i>Rad51c</i> ^{dah/+}	Expected frequency	6.25%	12.5%	6.25%	12.5%	25.00%	12.5%	6.25%	12.5%	6.25%	p=0.01, n=472
		Observed frequency	5.93%	11.86%	4.24%	18.01%	29.45%	7.42%	8.47%	11.44%	3.18%	

Supplementary Table 4. Polygenic *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice are born at sub-Mendelian frequency. Observed genotypes of *Brca2*^{Δ27/Δ27} + *Rad51c*^{dah/dah} mice was determined by genotyping offspring of *Brca2*^{Δ27/WT} + *Rad51c*^{dah/dah} (**a**, top) and *Brca2*^{Δ27/WT} + *Rad51c*^{dah/WT} (**b**, bottom) intercross. Expected genotypes are given using Mendelian genetics. *p*-value was calculated using chi-squared test (χ^2 -test).

Sample ID	Gene 1	mutation type	AA	zygosity	CNA	LOH	Gene 2	Mutation type	AA	zygosity	CNA	LOH
TCGA-C8-A12T-01	BRCA1	mut	D1344H	hetero	shallowdel	yes	FANCA	CNA	n/a	n/a	DeepDel	no
TCGA-C8-A12T-01	BRCA2	mut	E3177Q	hetero	shallowdel	yes	FANCA	CNA	n/a	n/a	DeepDel	no
TCGA-AN-A046-01	BRCA2	mut	E3342K	hetero	shallowdel	yes	FANCC	mut	R555Q	hetero	diploid	no
TCGA-AN-A046-01	BRCA2	mut	E3342K	hetero	shallowdel	yes	FANCM	mut	S1808Y	hetero	diploid	no
TCGA-AN-A046-01	BRCA2	mut	K985I	hetero	shallowdel	yes	FANCC	mut	R555Q	hetero	diploid	no
TCGA-AN-A046-01	BRCA2	mut	K985I	hetero	shallowdel	yes	FANCM	mut	S1808Y	hetero	diploid	no
TCGA-A8-A085-01	BRCA2	mut	E260Sfs*15	homo	shallowdel	yes	RAD51C	cna	n/a	n/a	AMP	no
TCGA-D8-A1JP-01	BRCA2	mut	L1965Ffs*39	hetero	shallowdel	yes	FANCC	struc var	n/a	n/a	fusion	no
TCGA-D8-A27V-01	BRCA2	mut	K936Nfs*24	hetero	shallowdel	yes	FANCC	mut	X511 splice	hetero	gain	no
TCGA-D8-A27V-01	BRCA2	mut	K936Nfs*24	hetero	shallowdel	yes	FANCC	mut	P482Lfs*5	hetero	gain	no
TCGA-A2-A0ET-01	BRCA2	struc var	n/a	n/a	BR fusion	no	FANCB	mut	I418Nfs*18	hetero	diploid	no
TCGA-A8-A07I-01	BRCA2	mut	D1355Y	hetero	shallowdel	yes	RAD51C	cna	n/a	n/a	AMP	no
TCGA-D8-A27G-01	BRCA2	mut	E2175Q	hetero	diploid	no	RAD51C	cna	n/a	n/a	AMP	no
TCGA-AC-A23H-01	BRCA2	mut	D687H	hetero	diploid	no	FANCA	mut	D931H	hetero	gain	no
TCGA-AC-A23H-01	BRCA2	mut	D687H	hetero	diploid	no	FANCA	mut	E38K	homo	gain	no
TCGA-AC-A23H-01	BRCA2	mut	D1033H	hetero	diploid	no	FANCA	mut	D931H	hetero	gain	no
TCGA-AC-A23H-01	BRCA2	mut	D1033H	hetero	diploid	no	FANCA	mut	E38K	homo	gain	no
TCGA-AC-A23H-01	BRCA2	mut	D687H	hetero	diploid	no	RAD51C	mut	D141H	hetero	AMP	no
TCGA-AC-A23H-01	BRCA2	mut	D1033H	hetero	diploid	no	RAD51C	mut	D141H	hetero	AMP	no
TCGA-AC-A23H-01	BRCA2	mut	D687H	hetero	diploid	no	RAD51C	cna	n/a	n/a	AMP	no
TCGA-AC-A23H-01	BRCA2	mut	D1033H	hetero	diploid	no	RAD51C	cna	n/a	n/a	AMP	no
TCGA-AC-A23H-01	FANCA	mut	D931H	hetero	gain	no	FANCG	mut	E566K	hetero	gain	no
TCGA-AC-A23H-01	FANCA	mut	E38K	homo	gain	no	FANCG	mut	E566K	hetero	gain	no
TCGA-AC-A23H-01	FANCA	mut	D931H	hetero	gain	no	FANCI	mut	S361L	hetero	diploid	no
TCGA-AC-A23H-01	FANCA	mut	E38K	homo	gain	no	FANCI	mut	S361L	hetero	diploid	no
TCGA-EW-A1PE-01	BRCA2	mut	E1158Q	hetero	diploid	no	FANCD2	mut	E1108*	hetero	diploid	no
TCGA-EW-A1PE-01	BRCA2	mut	K1191N	hetero	diploid	no	FANCD2	mut	E1108*	hetero	diploid	no
TCGA-A2-A04P-01	BRCA2	mut	Q3206E	hetero	gain	no	FANCA	cna	n/a	n/a	AMP	no
TCGA-AN-A0XW-01	BRCA2	mut	P527T	hetero	shallowdel	yes	RAD51C	struc var	n/a	n/a	fusion	no
TCGA-AN-A0XW-01	BRCA2	mut	P527T	hetero	shallowdel	yes	RAD51C	cna	n/a	n/a	AMP	no
TCGA-A8-A06R-01	BRCA2	cna	n/a	n/a	AMP	no	FANCB	cna	n/a	n/a	DeepDel	no

TCGA-D8-A1J9-01	FANCA	mut	R52L	homo	shallowdel	yes	FANCG	cna	n/a	n/a	AMP	no
TCGA-C8-A26Y-01	FANCA	mut	D694Y	hetero	shallowdel	yes	SLX4	mut	S381G	hetero	gain	no
TCGA-A2-A04X-01	FANCM	mut	E1622del	hetero	diploid	no	RAD51C	cna	n/a	n/a	AMP	no
TCGA-D8-A1JB-01	BRCA2	cna	n/a	n/a	DeepDel	no	FANCM	mut	R1684*	hetero	diploid	no
TCGA-D8-A1JD-01	BRCA2	cna	n/a	n/a	DeepDel	no	RAD51C	mut	P21A	hetero	diploid	no
MB-0211	BRCA2	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-0218	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-0291	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-0313	FANCA	CNA	n/a	n/a	deep del	no	FANCM	CNA	n/a	n/a	amp	no
MB-0468	BRCA2	CNA	n/a	n/a	amp	no	FANCM	CNA	n/a	n/a	amp	no
MB-0664	BRCA2	Mut	C3069Lfs*5	n/a	diploid	no	FANCA	CNA	n/a	n/a	amp	no
MB-2513	BRCA2	CNA	n/a	n/a	amp	no	FANCC	CNA	n/a	n/a	amp	no
MB-2513	BRCA2	CNA	n/a	n/a	amp	no	FANCD2	CNA	n/a	n/a	amp	no
MB-2763	FANCA	CNA	n/a	n/a	amp	no	SLX4	CNA	n/a	n/a	amp	no
MB-2847	BRCA2	CNA,Mut	T3310Nfs*17	n/a	shallow del	yes	RAD51C	CNA	n/a	n/a	amp	no
MB-3277	BRIP1	CNA	n/a	n/a	amp	no	FANCL	CNA	n/a	n/a	amp	no
MB-3396	BRCA2	CNA	n/a	n/a	deep del	no	FANCD2	Mut	R1299C	n/a	diploid	no
MB-3871	FANCA	CNA,Mut	R212G	n/a	shallow del	yes	SLX4	CNA	n/a	n/a	amp	no
MB-4331	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-4351	BRIP1	CNA	n/a	n/a	amp	no	FANCL	CNA	n/a	n/a	amp	no
MB-4601	BRCA2	CNA	n/a	n/a	amp	no	FANCA	CNA, Mut	R714Q	n/a	shallow del	yes
MB-4828	BRCA2	CNA	n/a	n/a	deep del	no	RAD51C	CNA	n/a	n/a	amp	no
MB-4938	BRCA2	Mut	L2654Ffs*3	n/a	diploid	no	RAD51C	CNA	n/a	n/a	amp	no
MB-4938	BRCA2	Mut	L2654Ffs*3	n/a	diploid	no	FANCD2	CNA, Mut	X330 splice	n/a	gain	no
MB-4949	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5130	BRCA2	Mut	S3366Mfs*2, V3365 S3366ins	n/a	diploid	no	FANCM	CNA	n/a	n/a	amp	no
MB-5197	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5275	BRCA2	Mut	G4V	n/a	diploid	no	FANCA	Mut	Q153*,E1420K	n/a	diploid	no
MB-5275	BRCA2	Mut	G4V	n/a	diploid	no	FANCD2	Mut	E369Q, S409L	n/a	diploid	no
MB-5275	FANCA	Mut	Q153*,E1420K	n/a	diploid	no	FANCD2	Mut	E369Q, S409L	n/a	diploid	no
MB-5414	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5465	BRIP1	CNA	n/a	n/a	amp	no	FANCL	CNA	n/a	n/a	amp	no
MB-5518	BRCA2	CNA	n/a	n/a	amp	no	FANCD2	CNA	n/a	n/a	amp	no
MB-5540	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5556	BRCA2	CNA	n/a	n/a	deep del	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5593	BRCA2	Mut	V2687I	n/a	diploid	no	RAD51C	CNA	n/a	n/a	amp	no
MB-5604	BRCA1	CNA	n/a	n/a	amp	no	FANCA	CNA, Mut	R1144W	n/a	shallow del	yes
MB-5634	FANCA	CNA	n/a	n/a	amp	no	FANCG	CNA	n/a	n/a	amp	no
MB-6026	BRCA2	CNA	n/a	n/a	deep del	no	RAD51C	CNA	n/a	n/a	amp	no
MB-6100	BRCA2	Mut	E1514K	n/a	diploid	no	RAD51C	CNA	n/a	n/a	amp	no
MB-6100	BRCA2	Mut	E1514K	n/a	diploid	no	FANCF	CNA	n/a	n/a	amp	no
MB-6156	BRCA2	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-6178	BRCA1	Mut	V452A	n/a	diploid	no	FANCA	Mut	D79Y	n/a	diploid	no
MB-6217	FANCA	Mut	A152G	n/a	diploid	no	FANCE	CNA	n/a	n/a	amp	no
MB-6329	FANCA	Mut	H292Q	n/a	diploid	no	FANCM	CNA	n/a	n/a	amp	no
MB-7028	FANCM	CNA	n/a	n/a	amp	no	RAD51C	CNA	n/a	n/a	amp	no
MB-7186	FANCA	CNA	n/a	n/a	deep del	no	SLX4	CNA	n/a	n/a	amp	no
MB-7299	FANCA	Mut	D598E	n/a	diploid	no	FANCM	CNA	n/a	n/a	amp	no
MTS-T0697	BRCA2	CNA,Mut	Q3036*	n/a	shallow del	yes	RAD51C	CNA	n/a	n/a	amp	no
MTS-T2118	BRCA2	Mut	T2662M	n/a	not available	no	FANCD2	Mut	E719G	n/a	n/a	no
MTS-T2398	BRCA1	CNA	n/a	n/a	amp	no	FANCA	Mut	H544Q, P1392S	n/a	n/a	no
MB-5556	BRCA2	CNA	n/a	n/a	deep del	no	RAD51C	CNA	n/a	n/a	amp	no

Supplementary Table 5. Individual mutation data as defined by cBioportal.com of breast cancer patient with tumors containing polygenic FANC mutations in the sixteen FANC combinations determined in Supplementary Table 1 and 2. Deep del denotes deep deletions, shallow del denotes shallow deletions, Amp denotes amplifications, CNA denotes copy number aberration, Mut denotes mutation. Hetero/homozygosity and LOH were inferred from allele frequency and CNA.

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