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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section

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n/a	Confirmed
	$oxed{oxed}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🔀 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes	A description of all covariates tested
\boxtimes	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	\boxtimes Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
,	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
Sof	ftware and code

Policy information about availability of computer code

Data collection

Flow cytometry data was collected 5-laser Cytek Aurora flow cytometer (Cytek Biosciences) and BD LSRII (BD Biosciences). FluroSpot plates were counted and data nalyzed using the AID Autoimmune Diagnostica GmbH ELISpot reader.

Data analysis

Flow cytometry data was acquired by SpectroFlo software and BD FACS DIVA software and analyzed using Flow Jo software v 10 (BD), Graph Pad Prsim v. 9.4.1 was used for statistical analysis

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Data generated or analyzed during this study are included in this article and its supplementary information files.

Human rese	arch parti	cipants		
Policy information	about <u>studies i</u>	nvolving human research participants and Sex and Gender in Research.		
Reporting on sex	sex and gender N/A			
Population chara	cteristics	N/A		
Recruitment		N/A		
Ethics oversight		N/A		
Note that full informa	ation on the appr	oval of the study protocol must also be provided in the manuscript.		
Field-spe	ecific re	porting		
Please select the o	ne below that i	s the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
Life sciences	E	sehavioural & social sciences		
For a reference copy of t	the document with	all sections, see nature.com/documents/nr-reporting-summary-flat.pdf		
Life scier	nces sti	udy design		
All studies must dis	sclose on these	points even when the disclosure is negative.		
Sample size	In order to assure statistical significance in the results at-least N=5 mice/group/time point were taken and the experiments were repeated atleast twice independently. These have been described clearly in the methods section.			
Data exclusions	No data were e	No data were excluded		
Replication	Experiments were repeated atleast twice independently and clearly explained in the methods. Each sample was run in triplicates for ELISA and FluoroSpot.			
Randomization	All animals wer	re randomly assigned to vaccination groups.		
Blinding	Blinding was not performed, however the experiments were conducted in an unbiased manner to prevent potential biases in the experimental groups			
Reportin	g for si	pecific materials, systems and methods		
We require informati	on from authors	about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & ex	perimental s	ystems Methods		
n/a Involved in th	ne study	n/a Involved in the study		
☐ ☐ Antibodies ☐ ChIP-seq				
□ □ Eukaryotic cell lines □ □ Flow cytometry				
	logy and archaeo			
	nd other organisr	15		
Antibodies				

Antibodies used All the antibodies used in this study are described in the supplementary tables. All the antibodies are validated by their respective manufacturer and were titrated prior to use

Validation

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Policy information about <u>cell lines</u>	and Sex and Gender in Research
Cell line source(s)	P388D1 cells (ATCC); mammalian Expi293 cells (Thermo Fisher Scientific); HEK293 (Integral molecular)
Authentication	Commercially purchased and previously used in several experiments.
Mycoplasma contamination	Negative for mycoplasma
Commonly misidentified lines (See <u>ICLAC</u> register)	None were utilized

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals	Female C57BL/6 mice (5–6 weeks of age) were obtained from The Jackson Laboratory
Wild animals	N/A
Reporting on sex	Female
Field-collected samples	N/A
Ethics oversight	Animal studies were carried out in accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The protocols were approved by the Institutional Animal Care and Use Committee at the Walter Reed Army Institute of Research [Assurance number D16-00596 (A4117-01)].

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Flow Cytometry

Plots

Con	firm	that:

The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

All plots are contour plots with outliers or pseudocolor plots.

A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	Cryopreserved or fresh splenocytes or lymph node cells were used for the flow cytometry
Instrument	Cytek Aurora (Cytek Biosciences) and BD LSR II (BD Biosciences)
Software	SpectroFlo, BD FACS DIVA, FLOW JO
Cell population abundance	No sorting was performed
Gating strategy	provided in supplementary figures

| Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.