

Supplemental Materials

A short-chain acyl-CoA synthetase that supports branched-chain fatty acid synthesis in *Staphylococcus aureus*

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**Tables S1-S3.
Figures S1-S5.**

Table S1**Fatty acid compositions of strains JE2 (wild-type) and NE1896 (*lpdA::φNΣ*).**

Fatty acid ^a	Strain		
	JE2 ^b	NE1896 ^b	NE1896 ^c
<i>iso</i> 14:0	0.6 ± 0.2	5.2 ± 1.2	0.9 ± 0.4
14:0	ND ^d	ND	0.1 ± 0.1
<i>iso</i> 15:0	6.4 ± 1.0	ND	ND
<i>anteiso</i> 15:0	26.8 ± 1.2	17.7 ± 3.4	24.2 ± 2.3
15:0	ND	ND	ND
<i>iso</i> 16:0	2.3 ± 0.2	12.8 ± 1.9	3.4 ± 0.6
16:0	1.3 ± 0.1	2.6 ± 0.5	2.2 ± 0.4
<i>iso</i> 17:0	7.4 ± 0.0	ND	ND
<i>anteiso</i> 17:0	27.9 ± 0.7	4.1 ± 0.6	18.9 ± 0.2
17:0	ND	0.9 ± 0.2	0.4 ± 0.1
<i>iso</i> 18:0	1.7 ± 0.1	8.5 ± 0.5	3.0 ± 0.1
18:0	7.5 ± 1.2	20.1 ± 2.0	16.4 ± 2.3
<i>iso</i> 19:0	3.6 ± 0.8	ND	ND
<i>anteiso</i> 19:0	10.0 ± 1.9	1.6 ± 0.5	8.5 ± 1.6
19:0	0.4 ± 0.1	2.9 ± 0.4	2.5 ± 0.1
<i>iso</i> 20:0	0.4 ± 0.1	2.9 ± 0.7	1.0 ± 0.2
20:0	3.6 ± 0.6	20.7 ± 3.9	18.4 ± 1.0

^aFatty acid composition is in weight percent.^bStrains JE2 and NE1896 were grown in LB^cStrain NE1896 was grown in LB supplemented with 1 mM αC5.^dND means not detected.

Table S2
Bacterial strains used in this study.

Strain	Genotype	Gene name	Description	Source
JE2			parent wild-type strain	Fey, <i>et al.</i> ^a
NE1829	JE2 SAUSA300_1465:: ϕ N Σ	<i>bkdA2</i>	branched-chain alpha-keto acid dehydrogenase E1	Fey, <i>et al.</i> ^a
NE1896	JE2 SAUSA300_1467:: ϕ N Σ	<i>lpdA</i>	dihydrolipoamide dehydrogenase	Fey, <i>et al.</i> ^a
NE162	JE2 SAUSA300_0559:: ϕ N Σ		putative CoA ligase	Fey, <i>et al.</i> ^a
NE385	JE2 SAUSA300_1679:: ϕ N Σ	<i>acsA</i>	acetyl-CoA synthetase	Fey, <i>et al.</i> ^a
NE539	JE2 SAUSA300_0228:: ϕ N Σ	<i>fadE</i>	acyl-CoA synthetase	Fey, <i>et al.</i> ^a
NE1036	JE2 SAUSA300_2542:: ϕ N Σ	<i>mbcS</i>	putative acetate-CoA ligase	Fey, <i>et al.</i> ^a
SW658	NE1036/pMbcS (pPJ658)		<i>mbcS</i> complement strain	This study

^aFey, P. D., Endres, J. L., Yajjala, V. K., Widhelm, T. J., Boissy, R. J., Bose, J. L., and Bayles, K. W. (2013) A genetic resource for rapid and comprehensive phenotype screening of nonessential *Staphylococcus aureus* genes. *MBio* **4**, e00537-00512

Table S3
Plasmids used in this study.

Plasmid	Description	Source
pET-28a	IPTG inducible protein expression in <i>E. coli</i>	Novagen
pPJ656	<i>mbcS</i> in pET-28a	This study
pCN51	Low copy number, cadmium inducible expression vector	Charpentier, <i>et al.</i> ^a
pPJ657	pCN51 vector with chloramphenicol selection	This study
pMbcS	<i>mbcS</i> in pPJ658	This study

^aCharpentier, E., Anton, A. I., Barry, P., Alfonso, B., Fang, Y., and Novick, R. P. (2004) Novel cassette-based shuttle vector system for gram-positive bacteria. *Appl. Environ. Microbiol* **70**, 6076-6085

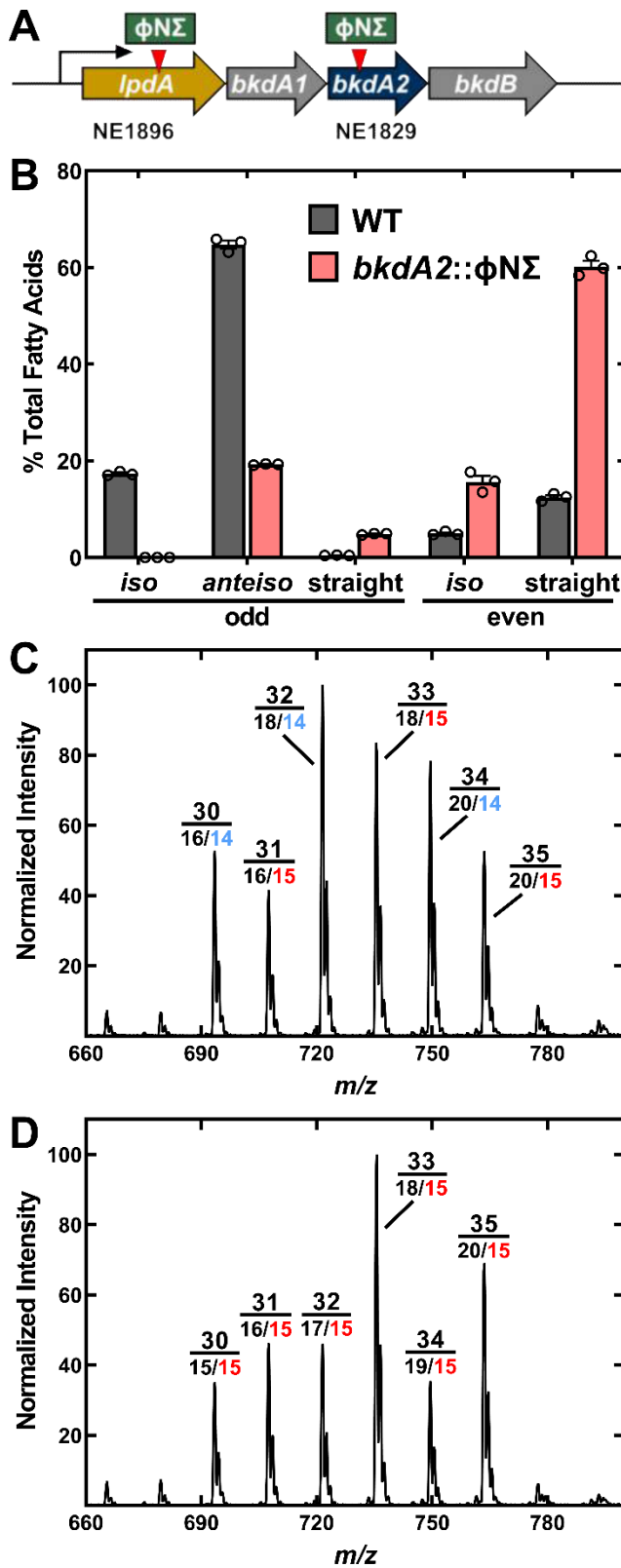
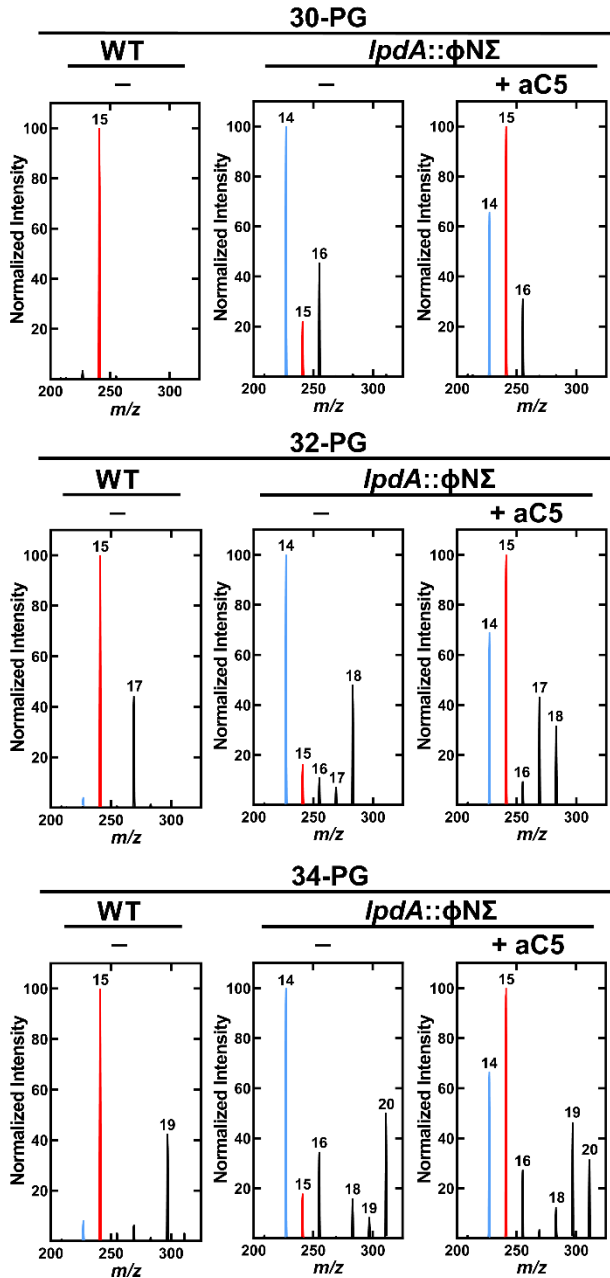


Figure S1. The *S. aureus* *bkd* operon and lipidomic characteristics of strain NE1829 (*bkdA2::φNΣ*). A, diagram of the 4-gene *bkd* operon showing the locations of the $\phi N\Sigma$ transposon insertions in the *lpdA* and *bkdA2* genes (red triangles). B, fatty acid composition of strain NE1829 (*bkdA2::φNΣ*) compared to the wild-type parent strain JE2 grown in LB. The strain JE2 data are the same as in Fig. 2A for comparison. C, PG molecular species in strain NE1829 (*bkdA2::φNΣ*) grown in LB. D, PG molecular species of strain NE1829 (*bkdA2::φNΣ*) grown in LB supplemented with 1 mM aC5.

A Even number PG



B Odd number PG

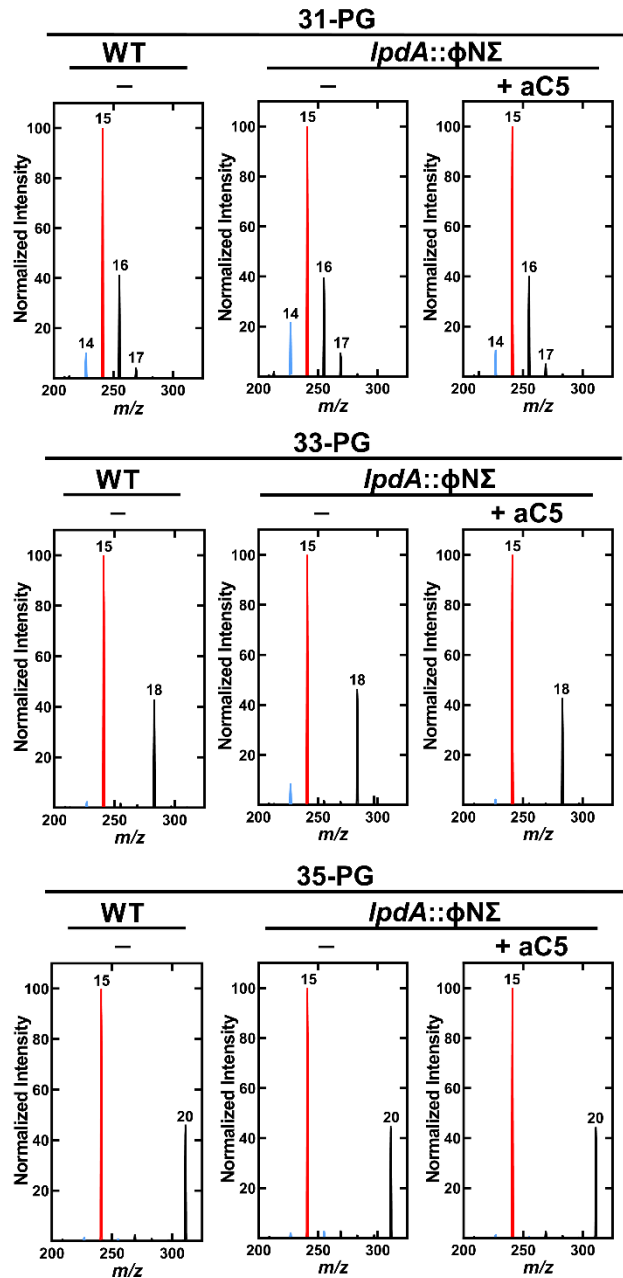


Figure S2. Fragmentation of the PG molecular species in strain NE1896 (*lpdA::φNΣ*) grown in LB with and without a 1 mM aC5 supplement. The individual PG peaks in the mass spectra shown in Figs. 2B, 2C and 2D were fragmented to determine the identity and positional distribution of the acyl chains in each PG peak. The 2-position acyl chain is characteristically ~2 times more intense than the 1-position acyl chain (see text). Identity of the BCFA chains is deduced based on knowledge of the fatty acid composition (Table S1). The i14 acyl chain is cyan and the a15 acyl chain is red.

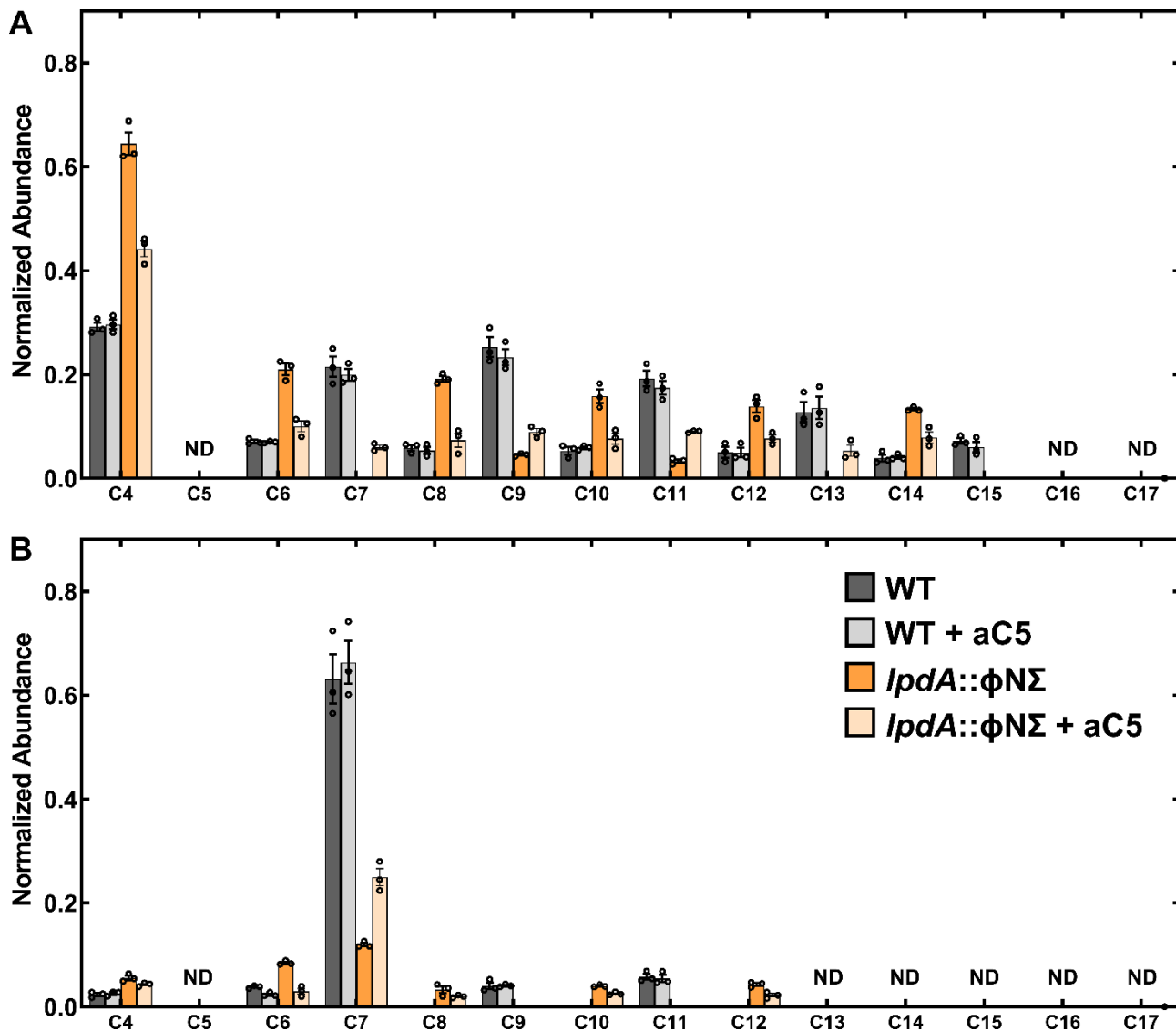


Figure S3. Impact of aC5 supplementation on the composition of the 3-hydroxy- and *trans*-2-acyl-ACP pools. Strains JE2 (wild-type) and NE1896 (*lpdA::φNΣ*) were grown in LB or LB supplemented with 1 mM aC5. The cells were collected by acid precipitation and protease digested. The FASII 3-hydroxy- and *trans*-2-acyl-ACP intermediates were detected by LC-MS/MS and the relative abundance of each peak was calculated based on the [¹³C₂]acetyl-ACP standard. *A*, the distribution of 3-hydroxy-acyl-ACP chain lengths. *B*, the distribution of *trans*-2-acyl-ACP chain lengths. *Inset*, the color key for both panels.

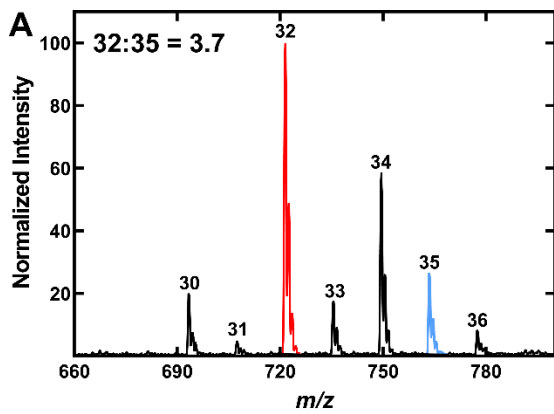
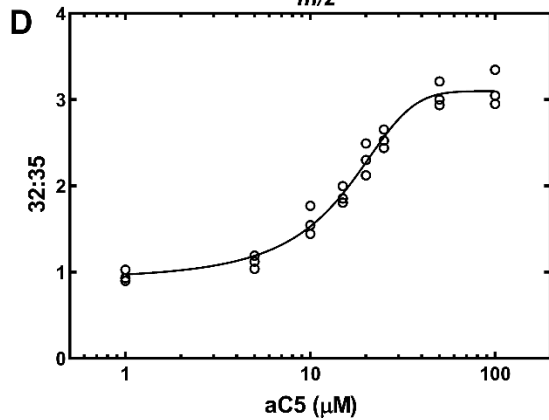
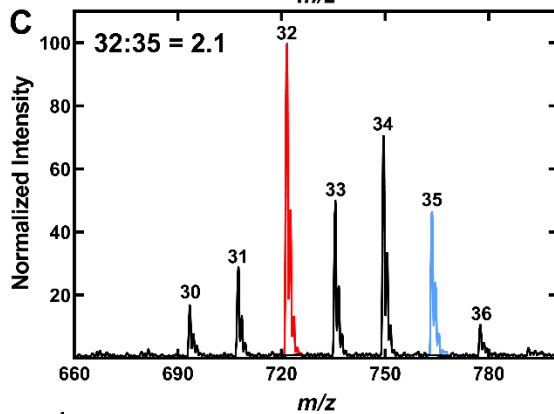
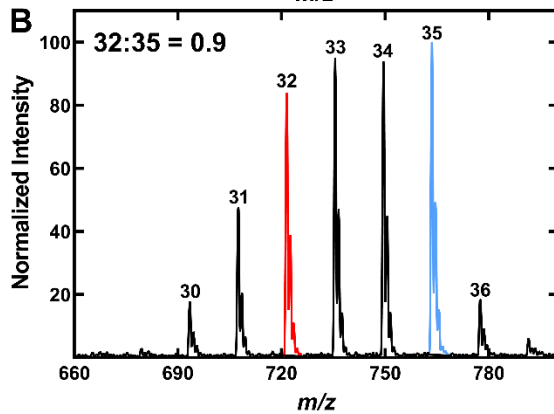


Figure S4. The impact of aC5 supplementation on the PG molecular species in wild-type strain JE2. *A*, strain JE2 grown in defined medium had a 32:35 PG ratio of 3.7. *B*, strain JE2 grown in defined medium lacking Ile and Leu had a 32:35-PG ratio of 0.9. *C*, strain JE2 grown in defined medium lacking Ile and Leu supplemented with 500 μM aC5 had a 32:35-PG ratio of 2.1. *D*, the 32:35-PG ratio in strain JE2 grown in defined medium lacking Ile and Leu plotted as a function of the aC5 concentration in the media.



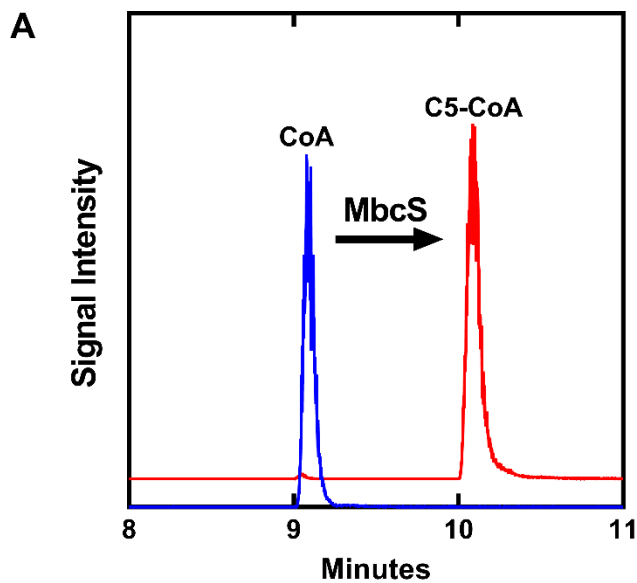


Figure S5. Assay for MbcS. MbcS assays were performed as described in Experimental Procedures and the reactions stopped by the addition of methanol. *A*, in this example, the CoA substrate (blue) was separated from the α C5-CoA product (red), and the peak areas analyzed with MultiQuant™ 3.0.2 software (Sciex). *B*, the C4-CoA calibration curve used to calculate the pmoles of acyl-CoA product formed in the MbcS reactions. *C*, the MbcS assay was linear with protein concentration. This example used 2-methylbutyric acid as the substrate.

