

## Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

### Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size ( $n$ ) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided  
*Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g.  $F$ ,  $t$ ,  $r$ ) with confidence intervals, effect sizes, degrees of freedom and  $P$  value noted  
*Give  $P$  values as exact values whenever suitable.*
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's  $d$ , Pearson's  $r$ ), indicating how they were calculated

*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

Data collection No software was used for data collection.

Data analysis

RNA-seq analysis

Cleaned short reads were aligned to reference genome taair10 by Bowtie2 (v2.1.0), and expression abundance was calculated by RSEM (v1.3.1) with default parameters. Heatmaps were visualized with the R package pheatmap (v1.0.12). To reduce false positive of differential expression, transcripts with less than 5 reads of all replicates in total were regarded as lowly expressed genes and have been removed in subsequent analysis. Differential expression analysis was conducted using edgeR (v3.32.1). A threshold of p value < 0.05 and Fold Change > 2 were used to decide whether significant expression difference exists between samples.

ChIP-seq analysis

ChIP-seq fastq reads were aligned to the TAIR10 reference genome with Bowtie2 (v2.1.0), allowing only uniquely mapping reads with 0 mismatches. Duplicated reads were removed by Samtools. ChIP-seq peaks were called by MACS2 (v2.1.1) and annotated with CHIPseeker (v1.28.3). Differential peaks were called by bdgdiff function in MACS2. ChIP-seq data metaplots were plotted by deeptools (v2.5.1). For Pol II 5' occupancy analysis, Pol II occupancy was calculated based normalized reads count (RPKM) on a TSS +/- 200 bp region and a TSS +500 bp to TTS gene body region by bedtools. Detailed information for published ChIP-seq datasets is listed in Supplementary Table 2.

Trim\_galore ([http://www.bioinformatics.babraham.ac.uk/projects/trim\\_galore/](http://www.bioinformatics.babraham.ac.uk/projects/trim_galore/)) was used to trim adapters after filtering low quality reads. BS-seq reads were aligned to TAIR10 reference genome by Bismark (v0.18.2) with default settings. Reads with three or more consecutive CHH sites were considered as unconverted reads and filtered. DNA methylation levels were defined as #C/ (#C + #T). DMRs (Differentially Methylated Regions) were called by DMRcaller with p < 0.01 for where the differences in CG, CHG, and CHH methylation were at least 0.4, 0.2, and 0.1, respectively.

## BS-PCR analysis

BS-PCR data were trimmed with primer sequences and mapped to TAIR10 reference genome with bsmmap (v2.90) allowing 2 mismatches and 1 best hit (-v 2 -w 1).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

## Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

All high-throughput sequencing data generated in this study are accessible at NCBI's Gene Expression Omnibus (GEO) via GEO Series accession number GSE197063(<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE197063>). Tair10 genome is available at <https://www.arabidopsis.org/index.jsp>

## Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences  Behavioural & social sciences  Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://nature.com/documents/nr-reporting-summary-flat.pdf)

## Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	No sample size calculation was performed. The applied sample sizes for RNA-seq, WGBS, ChIP-seq, and BS-PCR etc al, were selected according to public standards in the field.
Data exclusions	No data exclusion in the study.
Replication	Two replicates for ChIP-seq. Two replicates for BS-PCR. Three replicates for RNA-seq samples. Three technical replicates for qRT-PCR. All replicates were performed independently and produced high reproducible results.
Randomization	For all experiments, treatment and control samples were grown side by side, each replicate on separate plate. Allocation of samples were not random, because it is not relevant to the study.
Blinding	No blinding used because it was largely not relevant to our study. All data were collected based on the genotype of plants, while blinding the samples during the experiments will increases the risk of mislabeling and wrong results.

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

### Materials & experimental systems

n/a	Involved in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

### Methods

n/a	Involved in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

## Antibodies

Antibodies used

## Antibodies used

Anti-FLAG M2-Peroxidase (HRP) Millipore Sigma Sigma-Aldrich Cat# A8592, RRID:AB\_439702  
 Anti-FLAG® M2 Magnetic Beads Millipore Sigma Cat# M8823, RRID:AB\_2637089  
 Anti-Histone H3 Abcam Cat# ab1791, RRID:AB\_302613  
 Anti-trimethyl-Histone H3 (Lys4)~ Millipore Sigma Cat# 04-745, RRID:AB\_1163444  
 Anti-Histone H3 (acetyl K9) Abcam Cat# ab4441, RRID:AB\_2118292  
 Anti-Histone H3 (acetyl K14) Abcam Cat# ab52946, RRID:AB\_880442  
 Anti-Histone H3 (acetyl K27) Abcam Cat# ab4729, RRID:AB\_2118291  
 Anti-trimethyl-Histone H3 (Lys27) Millipore Sigma Cat# 07-449, RRID:AB\_310624  
 Anti-Histone H3 (di methyl K36) Abcam Cat# ab9049, RRID:AB\_1280939  
 Anti-Histone H3 (tri methyl K36) Abcam Cat# ab9050, RRID:AB\_306966  
 Anti-Histone H4 (acetyl K16) Abcam Cat# ab109463, RRID:AB\_10858987  
 Anti-RNA polymerase II CTD repeat YSPTSPS (phospho S5) antibody Abcam Cat# ab5131, RRID:AB\_449369

## Validation

anti-FLAG M2 (Sigma): the antibodies have been validated by the manufacturer, <https://www.sigmaaldrich.com/catalog/product/sigma/fl804>  
 Anti-FLAG M2-Peroxidase (HRP)(Sigma): the antibodies have been validated by the manufacturer, <https://www.sigmaaldrich.com/US/en/product/sigma/a8592>  
 Anti-Pol II Ser 5 (Abeam ab5131): the antibodies have been validated by the manufacturer, <https://www.abcam.com/rna-polymerase-ii-ctd-repeat-ysptsp-phospho-s5-antibody-ab5131.html>  
 anti-H3 (Ab1791, Abeam): the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-antibody-nuclear-marker-and-chip-grade-ab1791.html>  
 anti-H3K36me2 (Ab9049, Abeam): the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-dimethyl-k36-antibody-chip-grade-ab9049.html>  
 anti-H3K36me3 (Ab9050, Abeam): the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-trimethyl-k36-antibody-chip-grade-ab9050.html>  
 Anti-H3K27me3 (07-449, Millipore Sigma): the antibodies have been validated by the manufacturer, [https://www.emdmillipore.com/US/en/product/Anti-trimethyl-Histone-H3-Lys27-Antibody,MM\\_NF-07-449](https://www.emdmillipore.com/US/en/product/Anti-trimethyl-Histone-H3-Lys27-Antibody,MM_NF-07-449)  
 Anti-H3K4me3 (04-745, Millipore Sigma): the antibodies have been validated by the manufacturer, [https://www.emdmillipore.com/US/en/product/Anti-trimethyl-Histone-H3-Lys4-Antibodyclone-MC315-rabbit-monoclonal,MM\\_NF-04-745](https://www.emdmillipore.com/US/en/product/Anti-trimethyl-Histone-H3-Lys4-Antibodyclone-MC315-rabbit-monoclonal,MM_NF-04-745)  
 Anti-H3K27ac (ab4729, Abcam) : the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-acetyl-k27-antibody-chip-grade-ab4729.html>  
 Anti-H3K9ac (ab4441, Abcam) :the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-acetyl-k9-antibody-chip-grade-ab4441.html>  
 Anti-H3K14ac (ab82501, abcam): the antibodies have been validated by the manufacturer, <https://www.abcam.com/histone-h3-acetyl-k14-antibody-ab82501.html>

## Data deposition

- Confirm that both raw and final processed data have been deposited in a public database such as [GEO](https://www.ncbi.nlm.nih.gov/geo/).
- Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

## Data access links

*May remain private before publication.*

All high-throughput sequencing data generated in this study are accessible at NCBI's Gene Expression Omnibus (GEO) via GEO Series accession number GSE197063(<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE197063>). Enter token qvetmgogbnqxmp into the box.

## Files in database submission

ChIPseq-SET1-H3-ELF7-ZF-rep1.bw  
 ChIPseq-SET1-H3-fwa-rep1.bw  
 ChIPseq-SET1-H3-JMJ14-ZF-rep1.bw  
 ChIPseq-SET1-H3-LHP1-ZF-rep1.bw  
 ChIPseq-SET1-H3-MSI1-ZF-rep1.bw  
 ChIPseq-SET1-H3K27me3-ELF7-ZF-rep1.bw  
 ChIPseq-SET1-H3K27me3-fwa-rep1.bw  
 ChIPseq-SET1-H3K27me3-JMJ14-ZF-rep1.bw  
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ChIPseq-SET3-H3-fwa2-rep1.bw  
ChIPseq-SET3-H3-HD2A-ZF-rep1.bw  
ChIPseq-SET3-H3-HD2B-ZF-rep1.bw  
ChIPseq-SET3-H3-HD2C-ZF-rep1.bw



Replicates

2

Sequencing depth

Name	Total_reads	Unique_reads	Reads_length	Reads_type
ChIPseq-SET1-H3-ELF7-ZF-rep1_S20_L003	34692333	30138805	50	PE
ChIPseq-SET1-H3-fwa-rep1_S22_L003	34615984	29849162	50	PE
ChIPseq-SET1-H3-JMJ14-ZF-rep1_S35_L003	30624027	24609533	50	PE
ChIPseq-SET1-H3-LHP1-ZF-rep1_S16_L003	36532278	31760176	50	PE
ChIPseq-SET1-H3-MSI1-ZF-rep1_S19_L003	34773551	29846276	50	PE
ChIPseq-SET1-H3K27me3-ELF7-ZF-rep1_S85_L003	16857198	13565042	50	PE
ChIPseq-SET1-H3K27me3-fwa-rep1_S48_L003	24850552	19542262	50	PE
ChIPseq-SET1-H3K27me3-JMJ14-ZF-rep1_S91_L003	13614616	9023791	50	PE
ChIPseq-SET1-H3K27me3-LHP1-ZF-rep1_S70_L003	19171383	14191867	50	PE
ChIPseq-SET1-H3K27me3-MSI1-ZF-rep1_S47_L003	25550724	19876895	50	PE
ChIPseq-SET1-H3K4me3-ELF7_ZF-rep1	17892582	14665344	50	PE
ChIPseq-SET1-H3K4me3-fwa-rep1	26780511	22388582	50	PE
ChIPseq-SET1-H3K4me3-JMJ14-ZF-rep1	16201931	12494941	50	PE
ChIPseq-SET10-H3-CPL2-ZF-rep1	45106998	40540319	50	PE
ChIPseq-SET10-H3-fwa-rep1	54359212	48847610	50	PE
ChIPseq-SET10-PolIII-CPL2-ZF-rep1	33785877	28604913	50	PE
ChIPseq-SET10-PolIII-fwa-rep1	32512367	28918814	50	PE
ChIPseq-SET11-H3-CPL2-ZF-rep2	30282941	27328876	50	PE
ChIPseq-SET11-H3-fwa-rep2	34762270	31665246	50	PE
ChIPseq-SET11-PolIII-CPL2-ZF-rep2	33302491	28278239	50	PE
ChIPseq-SET11-PolIII-fwa-rep2	29869149	24921260	50	PE
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ChIPseq-SET12-H3-elf7-rep1	30482761	26831959	50	PE
ChIPseq-SET12-PolIII-Col-0-rep1	21776017	18795866	50	PE
ChIPseq-SET12-PolIII-elf7-rep1	21635631	18731688	50	PE
ChIPseq-SET13-H3-Col-0-rep2	45824394	41912020	50	PE
ChIPseq-SET13-H3-elf7-rep2	37223321	33652002	50	PE
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ChIPseq-SET13-PolIII-elf7-rep2	19988675	17608335	50	PE
ChIPseq-SET14-H3-ELF7-ZF-rep1	65351053	53822838	50	PE
ChIPseq-SET14-H3-EYFP-ZF-rep1_S3_L003	60530650	44916068	50	PE
ChIPseq-SET14-H3-EYFP-ZF-rep2_S4_L003	59588234	44961384	50	PE
ChIPseq-SET14-H3-fwa-rep1	72788551	57831743	50	PE
ChIPseq-SET14-H3-fwa-rep2	57686916	48104153	50	PE
ChIPseq-SET14-H3K27me3-EYFP-ZF-rep1_S16_L003	31084636	17847186	50	PE
ChIPseq-SET14-H3K27me3-EYFP-ZF-rep2	40418814	18323190	50	PE
ChIPseq-SET14-H3K27me3-fwa-rep1	40315526	18877298	50	PE
ChIPseq-SET14-H3K27me3-fwa-rep2	35664069	24163516	50	PE
ChIPseq-SET14-H3K36me2-ELF7-ZF-rep1	32259193	23676068	50	PE
ChIPseq-SET14-H3K36me2-fwa-rep1_S25_L003	30263355	22711628	50	PE
ChIPseq-SET14-H3K36me3-ELF7-ZF-rep1	51708205	39152947	50	PE
ChIPseq-SET14-H3K36me3-fwa-rep1	56600189	39809288	50	PE
ChIPseq-SET14-H3K4me3-EYFP-ZF-rep1	36933595	22418852	50	PE
ChIPseq-SET14-H3K4me3-EYFP-ZF-rep2	36490756	23447340	50	PE
ChIPseq-SET14-H3K4me3-fwa-rep1	41475420	28572652	50	PE
ChIPseq-SET14-H3K4me3-fwa-rep2	27195069	19783991	50	PE
ChIPseq-SET14-PolIII-EYFP-ZF-rep1	29680365	20620976	50	PE
ChIPseq-SET14-PolIII-EYFP-ZF-rep2	44560864	31033634	50	PE
ChIPseq-SET14-PolIII-fwa-rep1	30298421	22873363	50	PE
ChIPseq-SET14-PolIII-fwa-rep2	28536886	21535184	50	PE
ChIPseq-SET15-H3-EYFP-ZF-rep1	52324342	41816662	50	PE
ChIPseq-SET15-H3-EYFP-ZF-rep2	44051707	35836694	50	PE
ChIPseq-SET15-H3-fwa-rep1	76132941	64435746	50	PE
ChIPseq-SET15-H3-fwa-rep2	53395517	41430947	50	PE
ChIPseq-SET15-H3K14ac-EYFP-ZF-rep1	48349170	34032629	50	PE
ChIPseq-SET15-H3K14ac-EYFP-ZF-rep2	44200092	32570333	50	PE
ChIPseq-SET15-H3K14ac-fwa-rep1	44075233	34082299	50	PE
ChIPseq-SET15-H3K14ac-fwa-rep2	53313785	35890145	50	PE
ChIPseq-SET15-H3K27ac-EYFP-ZF-rep1	43602512	32254342	50	PE
ChIPseq-SET15-H3K27ac-EYFP-ZF-rep2	49632364	38520371	50	PE
ChIPseq-SET15-H3K27ac-fwa-rep1	50226011	39972700	50	PE
ChIPseq-SET15-H3K27ac-fwa-rep2	52110188	36735932	50	PE
ChIPseq-SET15-H3K9ac-EYFP-ZF-rep1_S41_L003	28632649	21285354	50	PE
ChIPseq-SET15-H3K9ac-EYFP-ZF-rep2	29897263	22824816	50	PE
ChIPseq-SET15-H3K9ac-fwa-rep1_S39_L003	29327321	23464787	50	PE
ChIPseq-SET15-H3K9ac-fwa-rep2	30198630	18853902	50	PE
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Genome browser session  
(e.g. [UCSC](#))

Available at GEO

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	<p>ChIPseq-SET8-H3-fwa-rep1 33391447 30985333 50 PE  ChIPseq-SET8-H3-HD2A-ZF-rep1 28408697 26352421 50 PE  ChIPseq-SET8-PolIII-ELF7-ZF-rep1_S54_L003 22931326 19386760 50 PE  ChIPseq-SET8-PolIII-fwa-rep1 21615655 17293168 50 PE  ChIPseq-SET8-PolIII-HD2A-ZF-rep1 26388420 22619253 50 PE  ChIPseq-SET9-H3-ELF7-ZF-rep2_S2_L001 26619337 24867631 50 PE  ChIPseq-SET9-H3-fwa-rep2 26871683 25416722 50 PE  ChIPseq-SET9-H3-HD2A-ZF-rep2 23227253 21677325 50 PE  ChIPseq-SET9-PolIII-ELF7-ZF-rep2 8780828 7854858 50 PE  ChIPseq-SET9-PolIII-fwa-rep2 9302660 8406014 50 PE  ChIPseq-SET9-PolIII-HD2A-ZF-rep2 7529608 6408422 50 PE</p>
Antibodies	<p>Anti-H3K27me3 (Millipore Sigma)  Anti-H3 (Abcam)  Anti-H3K4me3 (Millipore Sigma)  Anti-H3K27ac (Abcam)  Anti-H3K9ac (Abcam)  Anti-H4K16ac (Abcam)  Anti-H3K36me2 (Abcam)  Anti-H3K14ac (Abcam)  anti-Pol II Ser 5 (Abcam)  anti-FLAG M2 (Sigma)  Anti-H3K36me3 (Abcam)</p>
Peak calling parameters	<p>MACS2: '-f BAM -g 1.3e+8 -q 0.05 --extsize 147'</p>
Data quality	<p>All identified peaks in the study were called with a qual threshold of 0.01 ( FDR 1%).</p>
Software	<p>Bowtie (v1.1.2),  Samtools (v1.9)  MACS2 (v2.1.1)  ChIPseeker  deeptools (v2.5.1).  bedtools (v2.26.0)</p>