

Supporting Information for

Cryo-EM structure of the four-subunit *Rhodobacter sphaeroides* cytochrome *bc*₁ complex in styrene maleic acid nanodiscs

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This PDF file includes:

Tables S1 to S3

Figures S1 to S9

Table S1. Peptides detected by mass spectrometry analysis of purified four-subunit *cyt bc₁* following digestion with pepsin. Orange shading highlights the four *cyt bc₁* subunits resolved in the structure.

Protein	Uniprot ID	P value			Peptide spectrum matches			Sequence coverage (%)		
		Rep1	Rep2	Rep3	Rep1	Rep2	Rep3	Rep1	Rep2	Rep3
Cytochrome <i>b</i>	Q3IY10	0.000	0.000	0.000	166	171	202	55.51	56.18	56.85
Cytochrome <i>c₁</i>	Q3IY11	0.000	0.000	0.000	194	223	218	65.96	66.67	69.12
Ubiquinol-cytochrome <i>c</i> reductase iron-sulfur subunit	Q3IY09	0.000	0.000	0.000	21	22	24	47.06	49.73	59.36
Cytochrome <i>bc₁</i> subunit IV	Q3J2Z2	0.000	0.000	0.000	11	11	15	48.39	41.94	35.48
NADH dehydrogenase subunit L	Q3J3E6	0.000	0.000	0.000	10	13	15	8.89	10.97	13.06
Uncharacterized protein	Q3J5M7	0.000	nd	nd	2	nd	nd	9.63	nd	nd
Reaction center protein M chain	Q3J1A6	0.000	0.011	0.000	3	1	3	7.79	3.25	9.74
Probable cytosol aminopeptidase	Q3J2C5	0.001	0.000	0.000	8	7	11	9.61	10.22	14.93
Uncharacterized protein	U5NRE4	nd	0.001	0.003	nd	2	1	nd	40.91	22.73
Adenylosuccinate synthetase	Q3J0Z3	0.001	nd	0.033	2	nd	1	3.95	nd	2.33
Flagellar biosynthesis protein	Q3J1Y1	0.002	nd	0.026	2	nd	2	0.99	nd	0.99
Succinate dehydrogenase flavoprotein subunit A	Q3JZ75	0.002	0.000	0.005	4	6	6	6.00	8.17	9.83
Cytochrome <i>c_y</i>	Q3J003	0.003	nd	nd	2	nd	nd	9.20	nd	nd
Putative tape measure protein	Q3IWX5	0.007	0.012	0.041	1	1	1	1.03	1.03	1.03
Sec-independent protein translocase protein TatA	Q3J3D6	0.010	0.000	0.001	2	2	2	30.00	30.00	30.00
Non-homologous end joining protein Ku	Q3J0I8	0.010	nd	nd	1	nd	nd	4.87	nd	nd
ABC oligo/dipeptide transporter, inner membrane subunit	Q3J2X9	0.021	0.000	nd	5	4	nd	5.99	5.99	nd
Reaction center protein H chain	Q3J170	0.022	0.009	nd	3	3	nd	5.77	9.23	nd
L-threonine 3-dehydrogenase	Q3J3U9	0.047	nd	nd	11	nd	nd	2.05	nd	nd
Uncharacterized protein	Q3J6L2	nd	0.005	nd	nd	1	nd	nd	1.60	nd
Arginine utilization protein RocB	Q3JZA1	nd	0.009	nd	nd	1	nd	nd	1.31	nd
Transcriptional regulator, winged helix family	Q3IYWQ0	nd	0.009	nd	nd	1	nd	nd	3.18	nd
Glycosyltransferase	Q3IYT3	nd	0.019	nd	nd	2	nd	nd	1.45	nd
Uncharacterized protein	Q3IVS8	nd	0.019	nd	nd	1	nd	nd	4.11	nd
Cytochrome <i>c</i> oxidase subunit Cox1	Q3J5A7	nd	0.023	0.000	nd	2	2	nd	3.00	2.65
YkuD domain-containing protein	Q3J5N3	nd	0.028	0.041	nd	2	2	nd	3.57	3.57
Protein RdxB	P54932	nd	0.045	nd	nd	4	nd	nd	4.61	nd
H-NS histone family protein	U5NMX2	nd	0.048	0.007	nd	2	2	nd	9.78	9.78
Na ⁺ /solute symporter	Q3J0D4	nd	0.050	nd	nd	1	nd	nd	1.65	nd
ATP synthase subunit beta 2	Q3HKH4	nd	nd	0.021	nd	nd	2	nd	nd	2.16
Ysc84 domain-containing protein	Q3J2M1	nd	nd	0.032	nd	nd	1	nd	nd	4.76
Amino acid/amide ABC transporter substrate-binding protein, HAAT family	Q3J5H5	nd	nd	0.032	nd	nd	2	nd	nd	4.60
Propionyl-CoA carboxylase regulator	Q3J4E6	nd	nd	0.038	nd	nd	3	nd	nd	3.47
Phage-related protein, putative phage tail tape measure protein, lambda family	Q3J604	nd	nd	0.043	nd	nd	4	nd	nd	1.80
UvrABC system protein B	Q3J4P6	nd	nd	0.045	nd	nd	2	nd	nd	2.06
ABC Fe ³⁺ -siderophore transporter, inner membrane subunit	Q3IWR6	nd	nd	0.046	nd	nd	2	nd	nd	2.89

Table S2. Cryo-EM data collection, refinement, and validation statistics.

	Cyt <i>bc</i> ₁ consensus (b-b) (EMD-15616) (PDB-8ASI)	Cyt <i>bc</i> ₁ focussed refine (c-b) (EMD-15617) (PDB-8ASJ)
Data collection and processing		
Magnification	130 000 x	-
Electron exposure (e ⁻ /Å ²)	40	-
Defocus range	-0.8 to -2.0 μm	-
Pixel size (Å)	0.651	-
Symmetry imposed	C1	C1
Initial particle images (no.)	4 060 135	-
Final particle images (no.)	282 636 (7%)	72 118
Initial model used (PDB code)	RELION <i>de novo</i> model	-
Map resolution (Å)	2.9	3.75
FSC threshold	0.143	0.500
Model resolution range (Å)	2.6 – 4	Not determined
Map sharpening <i>B</i> factor (Å ²)	-117.187 (unsharpened map)	-114.875 (unsharpened map)
Model Composition and validation		
Non-hydrogen atoms	14726	14726
Protein residues	1789	1789
Ligands	21	21
Bond lengths (Å)	0.004 (0)	0.003 (0)
Bond angles (°)	1.079 (12)	0.804 (22)
Validation		
MolProbity score	1.38	1.40
Clashscore	5.03	3.80
Rotamer outliers (%)	0.00	0.21
Ramachandran plot		
Favoured (%)	97.41	96.50
Allowed (%)	2.54	3.38
Outliers (%)	0.06	0.11

Table S3: Primers used for generation of strains. Restriction sites used for cloning are underlined.

Primer Name	Sequence
<i>znuC</i> TGATGA UF	CCGGAATTCGAGGTGGCGGACCGTCAG
<i>znuC</i> TGATGA F	CACGACCATACGtgatgaCACGATCATTCC
<i>znuC</i> TGATGA R	GGAATGATCGTGtcatcaCGTATGGTCGTG
<i>znuC</i> KO DR	CGGCAAGCTTGCGAGGATGTCCGCGAAGAG
RSP2687 UF	CCGGAATTCGATCATCTCGCGCCTCAAGATGGACGCCGACATGTTCTCATTATCGTC TATCCCATC
RSP2687 UR	GATGGGATAGACGATGAATGAGAACATGTCCGGCGTC
RSP2687 DF	TTCTCATTATCGTCTATCCCATCGAATGACCCGGGCTACAAATGAGCACGCGTCTCAA GCTTGCCG
RSP2687 DR	CGGCAAGCTTGAGACGCGTGCTCATTGTAGC

MFS **F**I**D**D**I**P**S**F**E**Q**I**K**AR**VR**D**D**L**R**KH**GW**EK****R**
 WNDSRL **V**Q**K**S**R**E**L**L**N**D**E**E**L**K**I**D**P**A**T**W****I**W**K**R**
 MPSREEVAARRQRDFETV **W**K**Y**R**L**G**G**F**A**S**
GA**L**L**A**L**A**G**I**F**S**T**G**N**F**G**G**S**S**D**A**G**N**R**P**S**V**
YP**I**E******************

Figure S1. Sequence of SIV (Uniprot ID Q3J2Z2). Peptides identified by mass spectrometry of purified *cyt bc₁* highlighted green and resolved residues underlined in bold text.

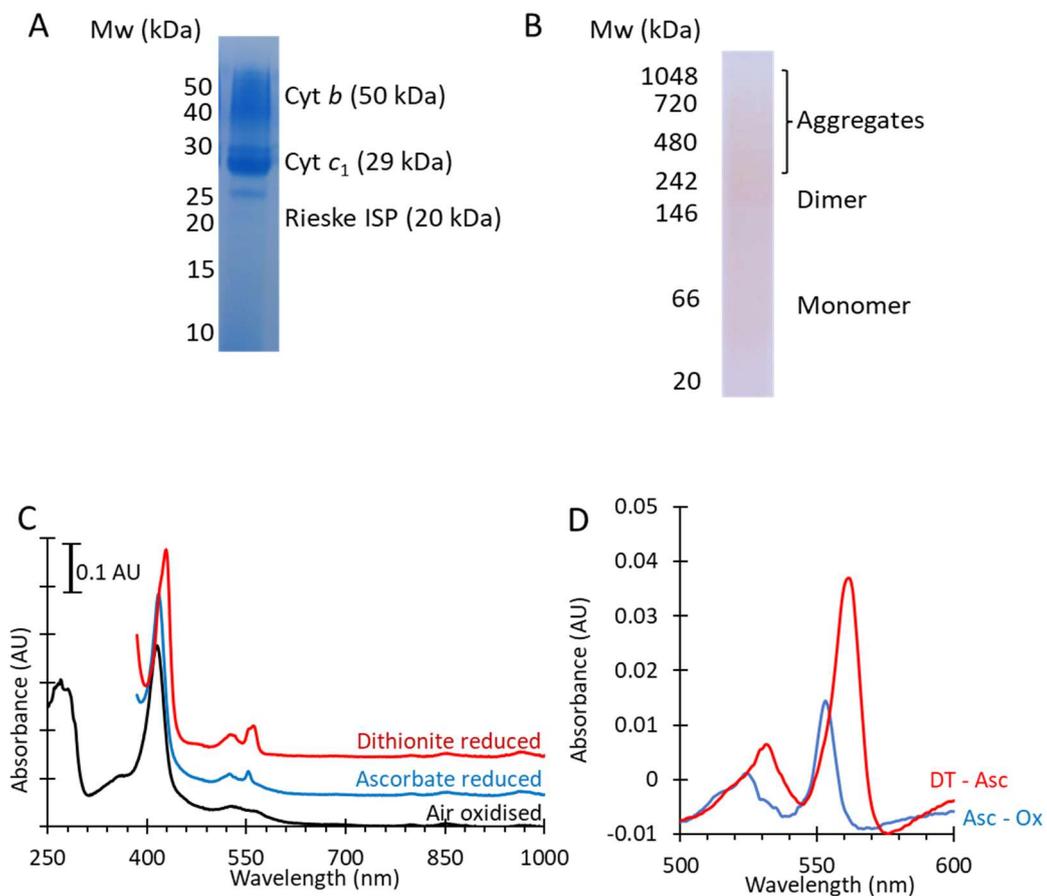


Figure S2. Gels and spectra of purified *cyt bc₁* complexes from the Δ SIV background. (A) Coomassie stained SDS page gel of purified Δ SIV *cyt bc₁* complexes. Each band is labelled with its identity and mass. (B) CN-PAGE gel of purified Δ SIV *cyt bc₁* complexes with the oligomeric state indicated. (C) UV/Vis/NIR spectra of Δ SIV *cyt bc₁* complexes as prepared (air oxidised) and following the addition of sodium ascorbate to reduce the *c* heme, and sodium dithionite to reduce all heme. (D) Ascorbate reduced minus ferricyanide oxidised (Asc – Ox), and dithionite reduced minus ascorbate reduced (DT – Asc) spectra in the 500–600 nm region used for determination of *c* and *b* heme concentrations, respectively. The heme *b* to heme *c* ratio was 1.7:1.

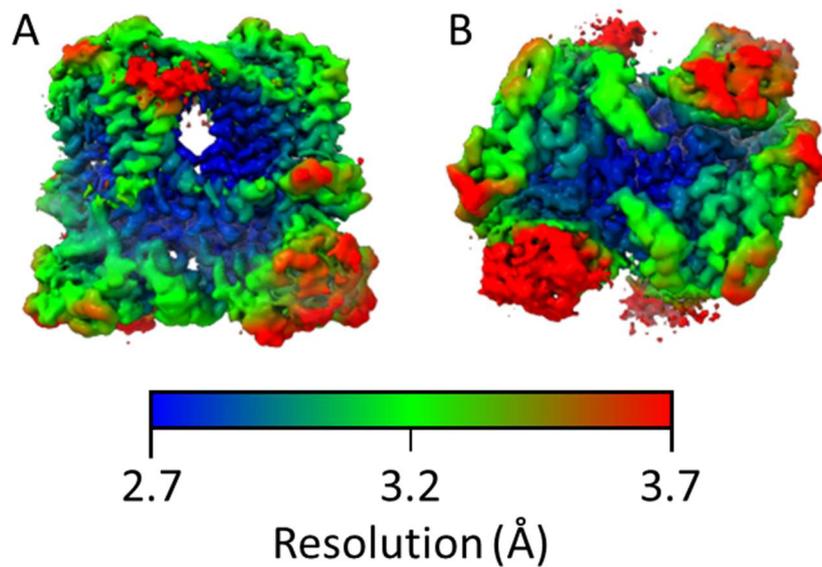


Figure S3. Locally sharpened maps viewed in the plain of the membrane (A) and from the periplasmic face (B). The scale bar indicates the local resolution.

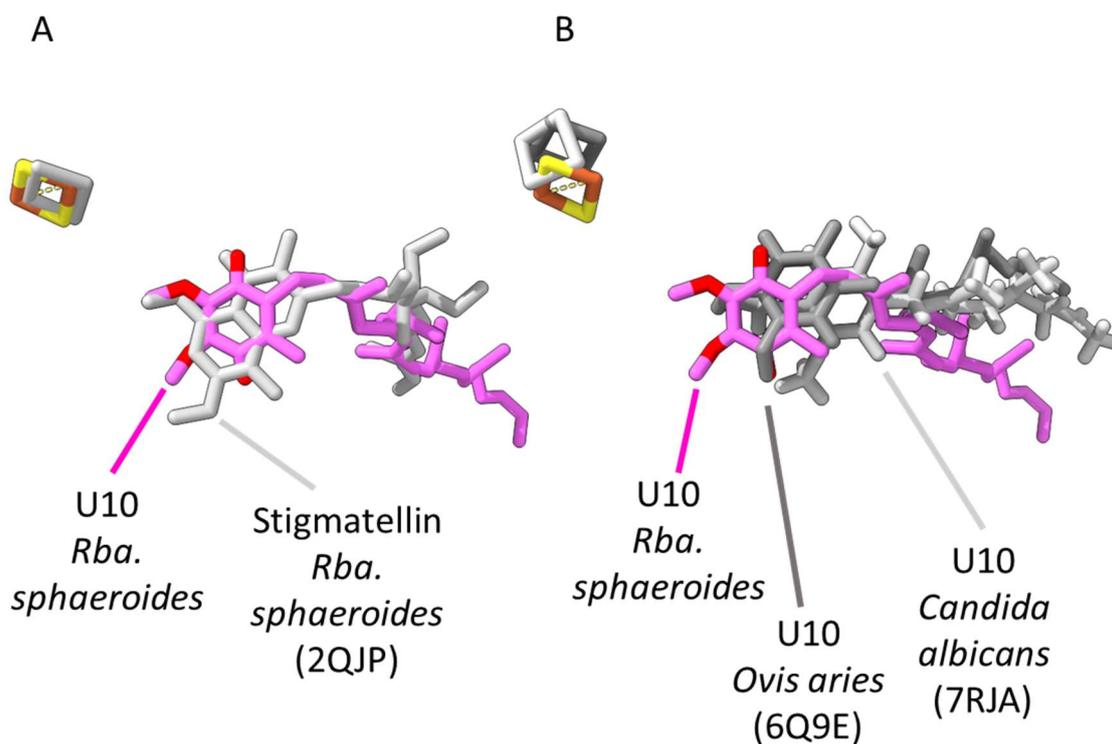


Figure S4. Comparison of the position of ubiquinone (U10) and stigmatellin in the Q_o sites of *cyt bc*₁ complexes. (A) Overlay of U10 in our structure with stigmatellin bound to the *Rba. sphaeroides* complex (PDB ID: 2QJP). (B) Overlay of U10 in our structure with Q_o quinones bound to complex III from *Ovis aries* (PDB ID:6Q9E, Dark grey) and *Candida albicans* (PDB ID:7RJA, light grey).

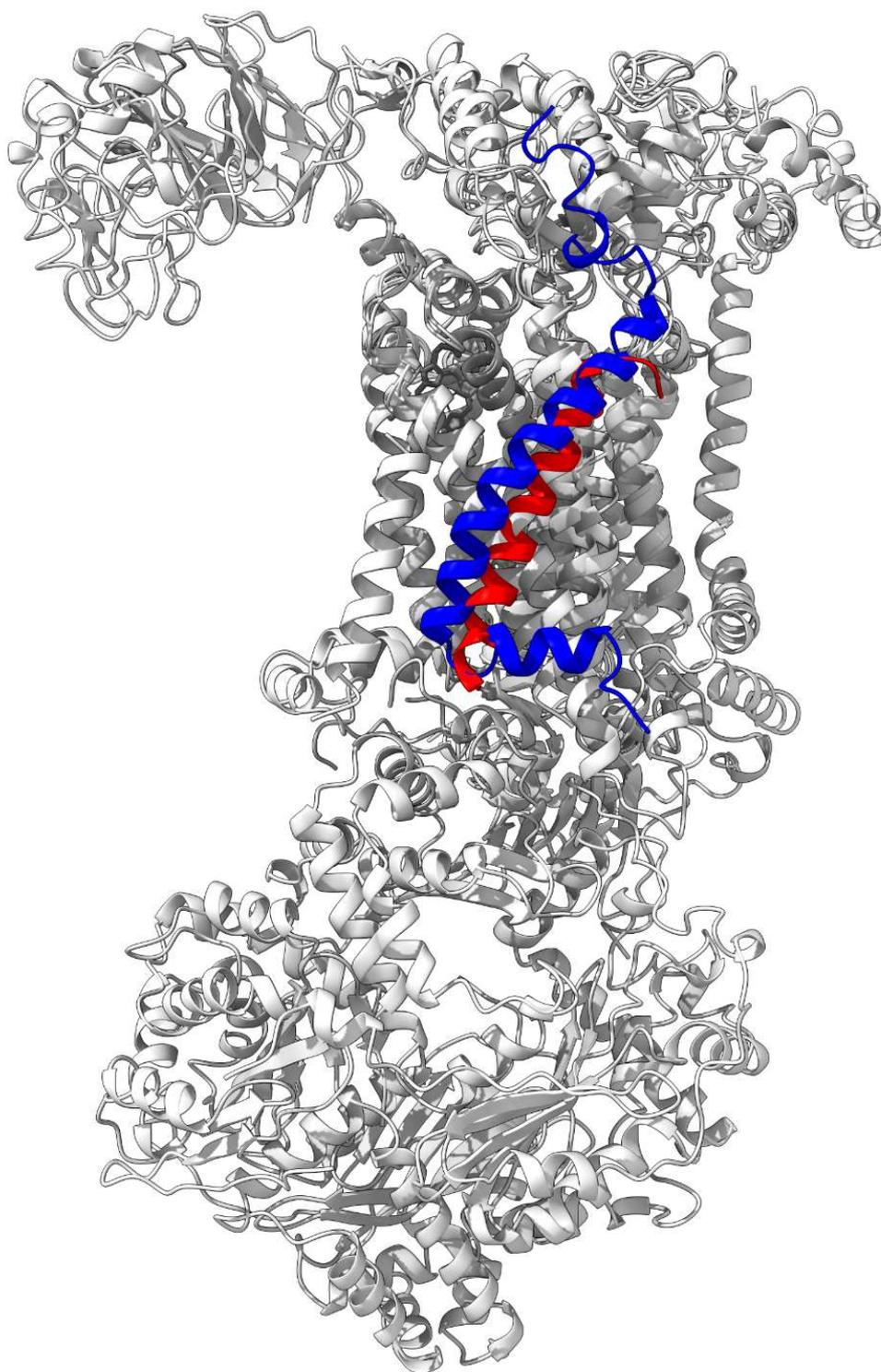


Figure S5. Comparison of the position of *Rba. sphaeroides* SIV with the UQCR10 subunit in the bovine complex. One monomer from the four subunit *Rba. sphaeroides* cyt bc_1 and the eleven-subunit bovine complex III (PDB ID: 1BE3) aligned to their catalytic subunits. Subunits for both complexes are shown in grey except *Rba. sphaeroides* SIV (red) and bovine UQCR10 (blue).

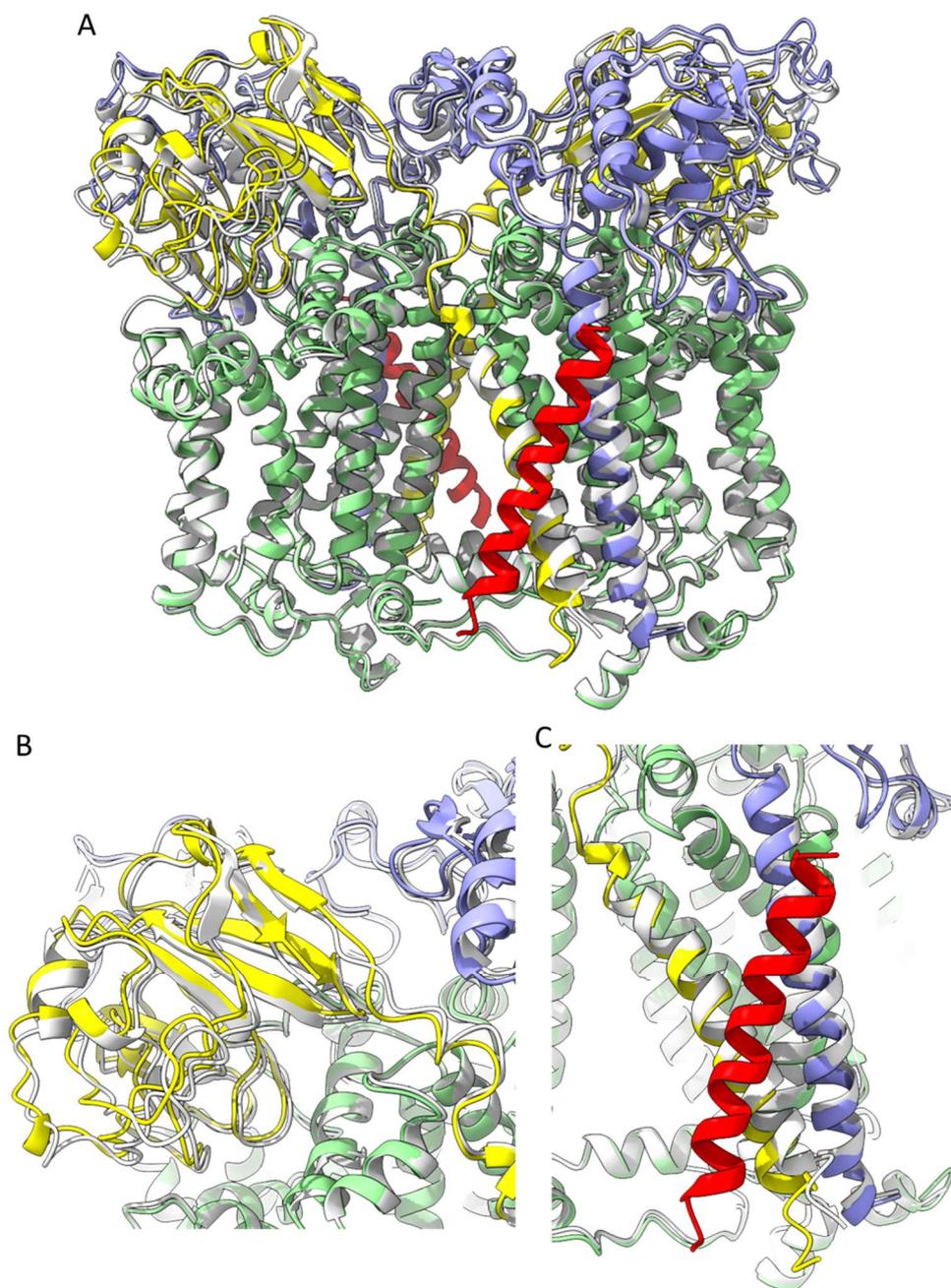
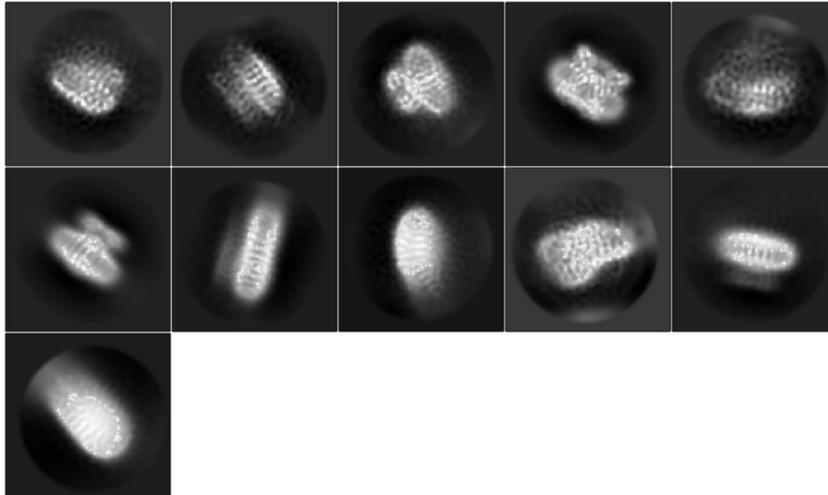


Figure S6. Alignment of the four- and three-subunit *Rba. sphaeroides* cyt bc_1 complexes. Alignment of our cryo-EM structure (coloured as in Figure 1 of the main paper) with the crystal structure of the complex lacking SIV (grey) (PDB ID: 2QJP). (A) Complete views of both complexes. (B) Zoomed in view of one Rieske head domain. (C) View of the SIV, cyt c_1 and Rieske TMHs.

A



B

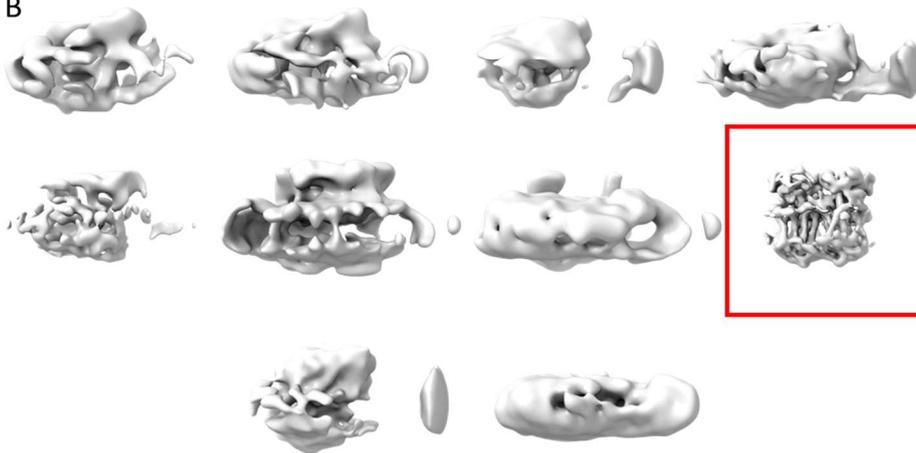


Figure S7. 2D and 3D classes from cryo-EM data processing. (A) Selected 2D classes after the fifth round of 2D classification. (B) 3D classes after two rounds of 3D classification. Class 8, indicated with a red square, was taken forward for refinement.

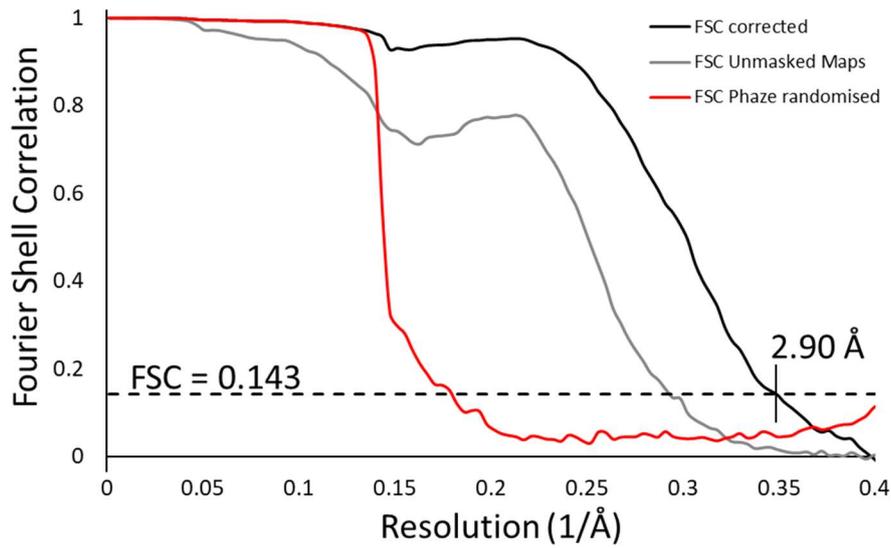


Figure S8. FSC curves for the final map. The resolution was determined with FSC = 0.143.

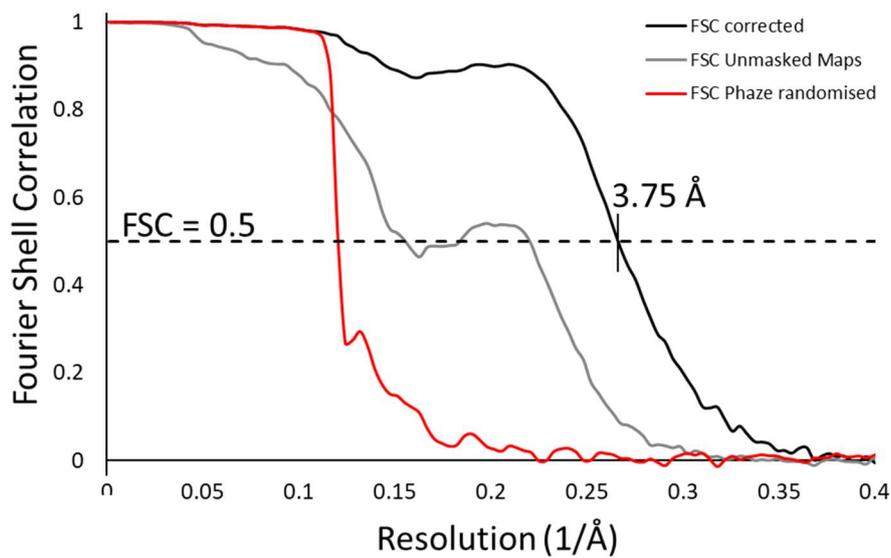


Figure S9. FSC curves for the focussed class showing an alternative conformation for the Rieske subunit. The resolution was estimated with FSC=0.5.