

Supplementary Material

1 Supplementary Figures and Tables

1.1 Supplementary Figures

CLUSTAL 0(1.2.4) multiple sequence alignment

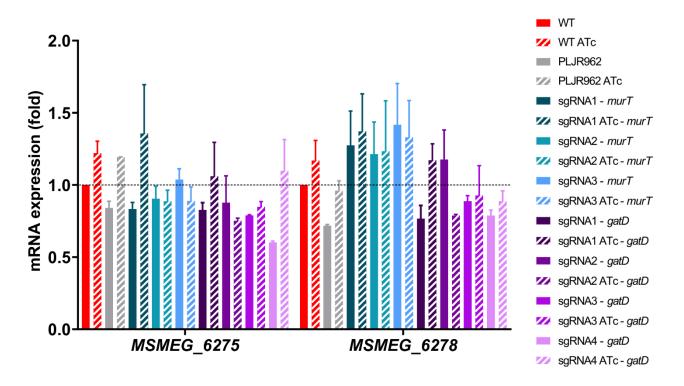
sp A0A0H3JUU7 MURT_STAANMRQWTAIHLAKLARKASRAVGKRGTDLPGQIAsp Q8DNZ9 MURT_STRR6MNLKTTLGLLAGRSSHFVLSRLGRGSTLPGKVAtr A0R5Q7 A0R5Q7_MYCS2-MLTVRGRAALAAGAAARWASRVTGRGAGAMIGGLVAtr I6Y4C7 I6Y4C7_MYCTU-WVTTRARLALAAGAGAARWASRVTGRGAGAMIGGLVA	32 33 36 36
RKVDTDVLRKLAEQVDDIVFISGTNGKTTTSNLIGHTLKANNIQIIHNNEGANMAAGITSAFIMQST LQFDKDILQSLAKNY-EIVVVTGTNGKTLTTALTVGILKEVYGQVLTNPSGANMITGIATTFLTAKS MTLDRSVLRQLGQGR-RTAIVTGTNGKSTTTRMIAAALA-PLGPVASNTEGANMDAGLVSALAANR- MTLDRSILRQLGMGR-RTVVVT <u>GTNGKSTT</u> TRMIAAALG-TLGAVATNAEGANMDAGLVAALAAHR- .* .:*:.*:*****: *: : * : * .* .****	99 99 100 100
PKTKIAVIEIDEGSIPRVLKEVTPSMMVFTNFFRDOMDRFGEIDIMVNNIAETISNKG-IKLLLN SKTGKNIAVLEIDEASLSRICDYIQPSLFVITNIFRDOMDRFGEIYTTYNMILDAIRKVPTATVLLN DAGLAALEVDEMHVPHVSDAVSPSVIVLLNLSRDOLDRVGEINHIERTLRAGLARHPDAVIVAN DAELAVLEVDEMHVPHISDAVDPAVVVLLNLSRDOLDRVGEINVIERTLRAGLARHPDAVVVAN :*.:*:** ::: :: :: :: :: :: :: :: :: :: ::	163 166 164 164
ADDPFVSRLKIASDTIVYYGMKAHAHEFEQ-STMNESRYCPNCGRLLQYDYIHYNQIGHYHCQ-CGF GDSPLFYKPTI-PNPIEYFGFDLEKGPAQLAHYNTEGILCPDCQGILKYEHNTYANLGAYICEGCGC CDDVLMTSAAYDNPDVVWVAAGGTWANDSVSCPRSGEVIVRDGRDWYSTGTDF CDDVLMTSAAYDSPNVVWVAAGGAWSNDSVSCPRSGEVIVRAPSQEDHWYSTGADF *. :. :: :: :: :: :: :: :: :: :: :: :: ::	228 232 217 221
KREQAKYEISSFDVAPFLYLNINDEKYDMKIAGDFNAYNALAAYTVLRELGLNEQTIKNGFETYT KRPDLDYRLTKLVELTNNRSRFVIDGQEYGIQIGGLYNIYNALAAVAIARFLGADSQLIKQGFDKSR KRPSPQWWFDETHIHGPDGLSVPMELALPGTVNRGNATQAVAAAVALGADPVAAVAAVSTVD KRPAPHWWFDDATLYGPDGLALPMRLALPGSVNRGNAAQAVAAAVALGADPAAVAAAVCQVD ** :: * ** : ** :	293 299 279 283
I SDNGRMQYFKKER-KEAMINLAKNPAGMNASLSVGEQLEGEKVYVISLNDNAADGRDTSWIYDADFE AVFORQETFHIGD-KECTLVLIKNPVGATQAIEMIKLAPYPFSLSVLLNANYADGIDTSWIWDADFE EVACRYRTVQVGPRHTARVLLAKNPAGWQEALSMVDRTGAGVVIAVNGQVPDGEDLSWLWDVRFE EVACRYRTVRIGA-HQARILLAKNPAGWQEALAMVDKHADGVVIAVNGRVPDGEDLSWLWDVRFE **: : * ***** :: : . : * ***:** III I	359 365 344 347
KLSKQQIEAIIVTGTRAEELQLRKLAEVEVPIIVERDIYKATAKTMDYKGFTVAIPNYTSIAPM QITDMDIPEINAGGVRHSEIARRLRVTGYPAEKITETSNLEQVLKTIENQDCKHAYILATYTAMLEF DFSGVQVVAAGERGTDLAVRLGYADVEHTLVHDTISAIKSCPPGHVEVIANYTAFLQL HFEKTRVVAA <u>GERGTDLAVRLGYA</u> GVEHTLVHDTVAAIASCPPGRVEV <u>VANYTAF</u> LQL .: : . * * :: ** : : : : : : : : : : : :	424 432 402 405
LEQLNRSFEGGQS 437 RELLASRQIVRKEMN 447 NRRLS 407 QRALARRG 413	

Supplementary Figure 1. Multiple amino acid sequence alignment of MurT from *S. aureus* (MURT_STAAN), *S. pneumoniae* (MURT_STRR6), *M. smegmatis* (A0R5Q7_MYCS2), and *M. tuberculosis* (I6Y4C7_MYCTU) using CLUSTAL version 1.2.4 (https://www.ebi.ac.uk/Tools/msa/clustalo/). The MurT central domain is displayed in black while the C-terminal domain (DUF1727) is shown in orange. The ATP and Mg²⁺ binding regions are presented in blue and green rectangles, respectively. A cysteine-rich region is shown in purple, with *S. aureus* and *S. pneumoniae* conserved cysteines in **bold**. The four conserved regions (I, II, III, and IV) of the MurT DUF1727 domain are indicated in red. The two residues and the DNAAD motif involved in the recognition and channeling of the ammonia produced by GatD, in *S. aureus*, are shown in grey.

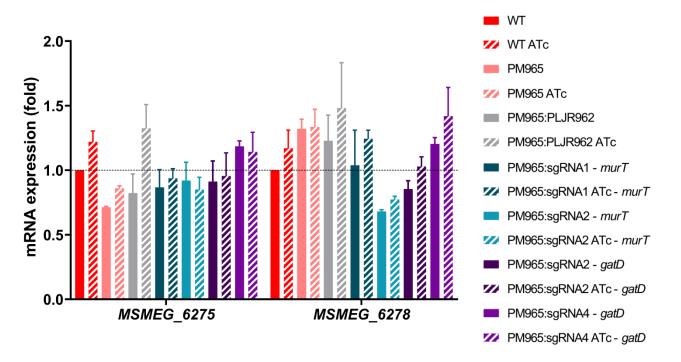
CLUSTAL 0(1.2.4) multiple sequence alignment

sp A0A0H3JN63 GATD_STAAN sp Q8DNZ8 GATD_STRR6 tr A0A0D6J2S9 A0A0D6J2S9_MYCSM tr I6XI14 I6XI14_MYCTU	MHELTIYHFMSDKLNLYS MVYTSLSSKDGNYPYQLNIAHLYGNLMNTYG MTHPSTVRIGLVLPDVMGTYG MVRIGLVLPDVMGTYG : * . : :. *.	18 31 21 16
DIGNIIALRQRAKKRNIKVNVVEINETEGITFDEC DNGNILMLKYVAEKLGAHVTVDIVSLHDDFDENHY DGGNAVVLRQRLRLRGIDAEIVEITLDDPV-PESL DGGNAVVLRQRLLLRGIAAEIVEITLADPV-PDSL * ** : *: : : . :	DIAFFGGGQDFEQSIIADDLPAKKESIDNYIQ DLYTLGGAEDYAQRLATKHLIRH-PGLQRAAE DLYTLGGAEDYAQRLATRHLRRY-PGLQRAAG	85 98 86 81
DGMPGLTICGGYQFLGKKYITPDGTELEGLGILDF NDGVVLAICGGFQLLGQYYVEASGKRIEGLGVMGH RGAPILAICAAIQVLGHWYETSAGERVEGVGLLDA RGAPVLAIQAAIQVLGHWYETSSGDRVDGVGLLDV . *:** *.**: * * .::*:*:*	YTLNQTNNRFIGDIKIHNEDFDETYYGFEN TTSPQ-EARTIGEVASRPLPDGLDQPLTGFEN	148 163 152 147
HGGRTYHDFGTLGHVTFGYGNNDEDKKEGIHYK HQGRTFLSDDQKPLGQVVYGNGNNEEKVGEGVHYK HRGGTLLGSDARPLGAVTKGAGNRAGDGFDGAVQG HRGGTVLGPGTSPLGAVVKGAGNRAGDGFDGAVAG * * *	NVFGSYFHGPILSRNANLAYRLVTTALKKKYG SVVATYMHGPCLARNPQLADHLLSRVVGDLP- SVVATYMHGPCLARNPELADLLLSKVVGELA-	212 230 218 213
IPFEPKEIDNEAEIQAKQVLIDRANRQKKSR QDIQLPAYEDILSQEIAEEYSDVKSKADFS -PLELPEVERLRSERL-AAPRRV -PLDLPEVDLLRRERL-SAR :: : ::	243 260 239 231	

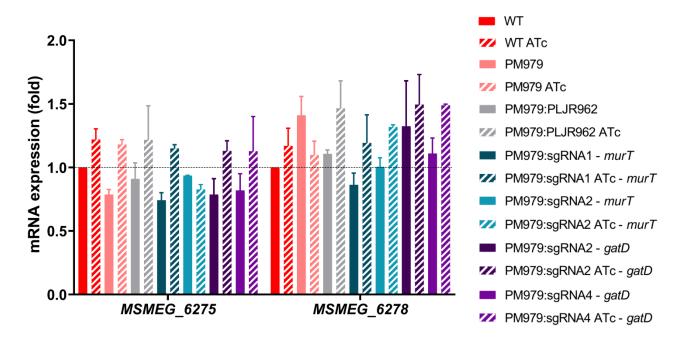
Supplementary Figure 2. Multiple amino acid sequence alignment of GatD from *S. aureus* (MURT_STAAN), *S. pneumoniae* (MURT_STRR6), *M. smegmatis* (A0R5Q7_MYCS2), and *M. tuberculosis* (I6Y4C7_MYCTU) using CLUSTAL version 1.2.4 (https://www.ebi.ac.uk/Tools/msa/clustalo/). Two conserved regions important for glutamine amidotransferase activity are presented in red. The two conserved residues responsible for the catalytic activity (C94, H189) of GatD are indicated in blue. The Y17 residue that interacts with MurT protein is displayed in a green rectangle. The D32 of *S. pneumoniae* that substitutes the missing glutamate in the catalytic triad of GatD is shown in purple.



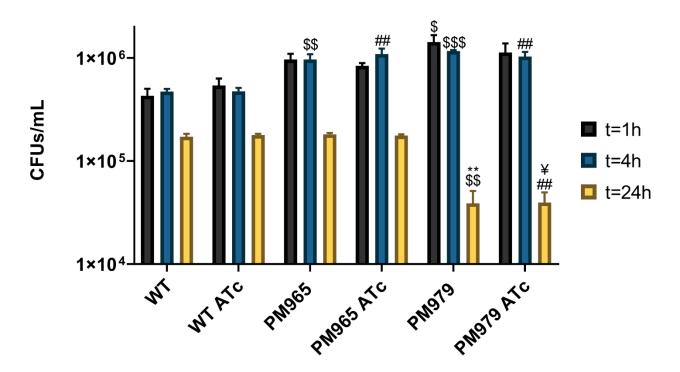
Supplementary Figure 3. Results of qRT-PCR assays for the *murT* (blue shade bars) and *gatD* (purple shade bars) knockdown mutants constructed in *M. smegmatis* WT (n=2). The graph shows the mean of the relative mRNA expression of *MSMEG_6275* and *MSMEG_6278* normalized to *sigA*, at 6 hours post-induction, with (stripped bars) and without (smooth bars) ATc. The dashed lines show the WT sample as calibrator. Error bars show the standard error of the mean. Multiple comparisons were made using one-way ANOVA. No significant differences were found.



Supplementary Figure 4. Results of qRT-PCR assays for the double mutants, that is, the *murT* (blue shade bars) and *gatD* (purple shade bars) knockdown mutants constructed in the *M*. *smegmatis* PM965 strain (n=2). The graph shows the mean of the relative mRNA expression of $MSMEG_{6275}$ and $MSMEG_{6278}$ normalized to *sigA*, at 6 hours post-induction, with (stripped bars) and without (smooth bars) ATc. The dashed lines show the WT sample as calibrator. Error bars show the standard error of the mean. Multiple comparisons were made using one-way ANOVA. No significant differences were found.



Supplementary Figure 5. Results of qRT-PCR assays for the triple mutants, that is, the *murT* (blue shade bars) and *gatD* (purple shade bars) knockdown mutants constructed in the *M*. *smegmatis* PM979 strain (n=2). The graph shows the mean of the relative mRNA expression of $MSMEG_{6275}$ and $MSMEG_{6278}$ normalized to *sigA*, at 6 hours post-induction, with (stripped bars) and without (smooth bars) ATc. The dashed lines show the WT sample as calibrator. Error bars show the standard error of the mean. Multiple comparisons were made using one-way ANOVA. No significant differences were found.



Supplementary Figure 6. Logarithmic representation of the mean of the bacterial survival of the parental strains *M. smegmatis* WT, PM965 and PM979, with and without ATc, in CFUs/mL, after infection and disruption of J774.A1 macrophages (n=3). Error bars show the standard error of the mean. Multiple comparisons were made using one-way ANOVA, with significance levels: * P < 0.05; ** P < 0.01; *** P < 0.001. Significant differences are indicated with symbols: \$ comparing to the WT strain; # comparing to the induced WT strain (WT ATc), * comparing to the PM965 strain; ¥ comparing to the induced PM965 ATc).



1.2 Supplementary Tables

Table S1 – sgRNAs used to target *murT*, *gatD* and *namH* in *M. smegmatis*.^a - mRNA knockdown caused by the electroporation of the indicated sgRNA in the *M. smegmatis* WT strain was measured by qRT-PCR after ATc induction. Fold knockdown efficiency is relative to an uninduced control plasmid containing the same sgRNA.

Target gene	Name of the oligos	РАМ	Base-pairing region of the sgRNA (the 12 bp seed region is underlined)	BLASTed sequence in the genome of <i>M. smegmatis</i> (seed region + PAM)	Fold knockdown efficiency ^a
	sgRNA1	5 '- GCAGAAC - 3'	5' - GCTGATCAC <u>GCGACAGATTGA</u> - 3'	5' - GCGACAGATTGAGCAGAAC - 3'	57
murT	sgRNA2	5' - CAGGAAC - 3'	5' - GACAGCCG <u>CCGGTTCAGTTG</u> - 3'	5' - CCGGTTCAGTTGCAGGAAC - 3'	1.6
	sgRNA3	5' - GCAGGAC - 3'	5' - GCGGCCCT <u>GGCCGAGCTGCC</u> - 3'	5' - GGCCGAGCTGCCGCAGGAC - 3'	21.1
	sgRNA1	5' - CGAGAAG - 3'	5' - GAGCGGAC <u>GTGCGTCGGAAC</u> - 3'	5' - GTGCGTCGGAACCGAGAAG - 3'	1.7
gatD	sgRNA2	5' - CGAGGAT - 3'	5' - ACCTGGATGG <u>CCGCGCAGATCG</u> - 3'	5' - CCGCGCAGATCGCGAGGAT - 3'	2.4
guiD	sgRNA3	5' - CGAGCAG - 3'	5' - GCCTCCTGCGGAG <u>AGGTGGTGGCGT</u> - 3'	5' - AGGTGGTGGCGTCGAGCAG - 3'	3.3
	sgRNA4	5' - AGAGCAG - 3'	5' - GCAGATCGC <u>CGACCACGCGGG</u> - 3'	5' - CGACCACGCGGGAGAGCAG - 3'	2.5
	sgRNA1	5' - CTGGAAC - 3'	5' - AGAGTGCT <u>TCGCCAGCCTAG</u> - 3'	5' - TCGCCAGCCTAGCTGGAAC - 3'	7.5
namH	sgRNA2	5' - GAAGAAG - 3'	5' - GCTTCACCGA <u>GTCGGTGGTCTC</u> - 3'	5' - GTCGGTGGTCTCGAAGAAG - 3'	5.6
	sgRNA3	5' - GGGGAAG - 3'	5' - GGTTCGCG <u>GACAACCCGTTT</u> - 3'	5' - GACAACCCGTTTGGGGGAAG - 3'	5.3

Table S2 - Primers designed and synthesized to clone the sgRNAs into PLJR962 for CRISPRi-mediated targeting in *M. smegmatis*. ^a – PAM strength was described in accordance with the table of functional PAMs *in vivo* for dCas9_{Sth1}-mediated targeting in mycobacteria defined by Rock *et al.*; ^b – Calculated with the Eurofins Genomics Melting temperature (Tm) formula.

Target gene	Name of the oligos	РАМ	PAM strength ^a	Gene Location 5'-3' (bp)	Primers		Length (bp)	% GC	Tm ^b (°C)
	sgRNA1	5 '- GCAGAAC - 3'	5	406	PFwd			56	66
	sgrinai	5 - UCAUAAC - 5	5	400	PRv			48	63
murT	sgRNA2	5' - CAGGAAC - 3'	14	1220	PFwd	5' - GGGAGACAGCCGCCGGTTCAGTTG - 3'	24	67	70
mui 1	sginiaz	5 - CAUGAAC - 5	14	1220	PRv	5' - AAACCAACTGAACCGGCGGCTGTC - 3'	24	58	66
		5' - GCAGGAC - 3'	15	153	PFwd	5' - GGGAGCGGCCCTGGCCGAGCTGCC - 3'	24	83	76
	sgRNA3	5 - OCAOUAC - 5	15	155	PRv	5' - AAACGGCAGCTCGGCCAGGGCCGC - 3'	24	75	73
	sgRNA1 5' - CGAGAAG - 3'		1	498	PFwd	5' - GGGAGAGCGGACGTGCGTCGGAAC - 3'	24	71	71
	sgRNA1	J - CUAUAAO - J	I	490	PRv	5' - AAACGTTCCGACGCACGTCCGCTC - 3'	27	63	68
	sgRNA2 5'	2 5' - CGAGGAT - 3'	9	299	PFwd	5' - GGGAACCTGGATGGCCGCGCAGATCG - 3'	26	69	73
gatD	sginaz	5 - CUAUUAT - 5)	2))	PRv	5' - AAACCGATCTGCGCGGCCATCCAGGT - 3'	20	62	70
guiD	sgRNA3 5' - CGAGCAG - 3'		13	383	PFwd	5' - GGGAGCCTCCTGCGGAGAGGTGGTGGCGT - 3'	29	72	77
	sgiuns	J - CUAUCAU - J	15	383	PRv	PRv 5' - AAACACGCCACCACCTCTCCGCAGGAGGC -		66	74
	sgRNA4	5' - AGAGCAG - 3'	13	652	PFwd	5' - GGGAGCAGATCGCCGACCACGCGGG - 3'	25	76	75
	Sgittinne	J - AGAGEAG - J	15	032	PRv	5' - AAACCCCGCGTGGTCGGCGATCTGC - 3'	25	68	71
	sgRNA1	5' - CTGGAAC - 3'	11	-44	PFwd	5' - GGGAAGAGTGCTTCGCCAGCCTAG - 3'	24	63	68
	sgittal	5 - CTOOAAC - 5	11	-44	PRv	5' - AAACCTAGGCTGGCGAAGCACTCT - 3'	24	54	64
namH	sgRNA?	5' - GAAGAAG - 3'	1	318	PFwd	5' - GGGAGCTTCACCGAGTCGGTGGTCTC - 3'	26	65	71
num11	SERINAL	J - UAAUAAU - J	1	510	PRv	5' - AAACGAGACCACCGACTCGGTGAAGC - 3'	20	58	68
	sgRNA3	5' - GGGGAAG - 3'	4	1142	PFwd	5' - GGGAGGTTCGCGGACAACCCGTTT - 3'	24	63	68
sgKINA3	5 - 0000AA0 - 5	+	1142	PRv	5' - AAACAAACGGGTTGTCCGCGAACC - 3'	24	54	64	

Table S3 - Primers used to quantify the mRNA expression levels of target genes by qRT-PCR in *M. smegmatis.* ^a - qRT-PCR primers were designed to avoid primer secondary structures and sequence repeats, have at least 50% of GC content and have a Tm between 57-63 $^{\circ}$ C; ^b - Calculated with the Eurofins Genomics Tm formula; ^c – AE designates the amplification efficiency calculated for each pair of primers in a single-run qPCR amplification of the calibrator sample (*M. smegmatis* WT), performed in triplicates.

qRT-PCR primers ^a									
Target gene		Primer sequences	Length (bp)	% GC	Tm ^b (° C)	Product size (bp)	AE ^c (%)		
sigA	PFwd	5' - ACACCGACCTGGAACTCG - 3'	18	61	58	152	80		
(MSMEG_2758)	PRv	5' - GACGCCTTGTCCTTCTCG - 3'	18	61	58	132	80		
murT	PFwd	5' - AATCGACCACCACCAGGAT - 3'	19	53	57	213	105		
(<i>MSMEG_6276</i>)	PRv	5' - CGACAGATTGAGCAGAACGA - 3'	20	50	57	213	105		
gatD	PFwd	5' - GCGACGGTTTCGACGGTG - 3'	18	67	61	167	95		
(<i>MSMEG_6277</i>)	PRv	5' - AGGCGCTCAGAACGCAAAC - 3'	19	58	59	107	93		
MSMEC (275	PFwd	5' - GAAGCTGTAGTCGAACCCCG - 3'	20	60	61	160	04		
MSMEG_6275	PRv	5' - CTCAACCCTGGTGTCGATCC - 3'	20	60	61	160	84		
MCMEC (270	PFwd	5' - TTGAACACCCCGACGATCAC - 3'	20	55	59	154	90		
MSMEG_6278	PRv	5' - CGATGTCCTCGACTTCGCA - 3'	19	58	59	154	80		
namH	PFwd	5' - CGTTCTTCAAGTGCCTCACC - 3'	20	55	59	1.6.	07		
(<i>MSMEG_6410</i>)	PRv	5' - GTTGCCTTCCACCACACC - 3'	18	61	58	165	87		
MOMEC (199	PFwd	5' - CCGCAGCTTCGAGATCAAGG - 3'	20	60	61	127	02		
MSMEG_6409	PRv	5' - CCGGTACAGGTTCTTCGGAC - 3'	20	60	61	137	83		
MSMEC (411	PFwd	5' - GTGCCAGAAGCTGTGACCG - 3'	19	63	61	120	05		
MSMEG_6411	PRv	5' - GGGATCACCTTCGATGTGCC - 3'	20	60	61	120	85		

Table S4 – The EUCAST non-species related PK/PD breakpoints for amoxicillin (AMX), amoxicillin-clavulanate (AMX+CLA), cefotaxime (CTX) and meropenem (MEM). These breakpoints, used when there are no species-specific breakpoints, are based on version 12 of the EUCAST guidelines on susceptibility testing (<u>https://www.eucast.org/clinical_breakpoints/</u>). For the AMX+CLA breakpoint, the concentration of clavulanate used was fixed at 2 µg/mL.

EUCAST breakpoi	EUCAST breakpoints (µg/mL)							
	S≤	R >						
AMX	2	8						
AMX+CLA	2	8						
СТХ	1	2						
MEM	2	8						

Table S5 - Single nucleotide polymorphisms (SNPs) found in the target genes *murT*, *gatD* or *namH* through the whole genome sequencing of 172 clinical strains of *Mtb*, with the description ID of the strains in which mutations were found. Locus tags, gene names, products and positions refer to the H37Rv reference genome (AL123456.3).

Locus Tag	Gene Name	Genomic Position	Mutation	Type of Variant	Effect in Product	Frequency	Strains
Rv3712	murT	4158032	A>C	Missense	Lys351Thr	1/172	PT_TB0303
		4158361	A>G	Synonymous	Pro45Pro	1/172	PT_TB0303
Rv3713	gatD	4158493	C>T	Synonymous	Ile89Ile	10/172	PT_TB0067; PT_TB0057; PT_TB0029; PT_TB0069; PT_TB0328; PT_TB0319; PT_TB0276; PT_TB0265; PT_TB0288; PT_TB0214
		4158865	G>A	Synonymous	Ala213Ala	4/172	PT_TB0311; PT_TB0314; PT_TB0305; PT_TB0242
		4282589	T>C	Synonymous	Ala47Ala	1/172	PT_TB0245
D2 010		4282707	C>G	Missense	Pro87Ala	1/172	PT_TB0070
Rv3818	namH	4283319	G>A	Missense	Ala291Thr	2/172	PT_TB0313; PT_TB0250
		4283424	G>C	Missense	Ala326Pro	1/172	PT_TB0350

Table S6 - Information of the strain ID, Sequence Read Archive (SRA)/European Nucleotide Archive (ENA) accession numbers, drug-resistance profile and *in silico* TB-profiler predictions of lineages, spoligotypes and regions of difference (RD) of the clinical isolates of *Mtb* for which mutations in the target genes *murT*, *gatD* or *namH* were found through whole genome sequencing.

Starin ID	SRA/ENA		Drug-resistance	e TB profiler prediction			
Strain ID	Accession #	Classification	Profile	Lineage	Spoligotype	Regions of difference (RD)	
PT_TB0303	ERR7764425	Susceptible	Susceptible	lineage6	AFRI 1	RD702	
PT_TB0067	ERR2864255	Resistant	MDR	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0057	ERR2864286	Resistant	MDR	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0029	ERR2864231	Resistant	MDR	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0069	ERR2864228	Resistant	MDR	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0328	ERR7764343	Susceptible	Susceptible	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0319	ERR7764417	Susceptible	Susceptible	lineage2.2.1.1	Beijing-RD150	RD105; RD207; RD181; RD150	
PT_TB0276	ERR7764390	Susceptible	Susceptible	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0265	ERR7764438	Susceptible	Susceptible	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0288	ERR7764362	Susceptible	Susceptible	lineage2.2.1	Beijing-RD181	RD105; RD207; RD181	
PT_TB0214	ERR4781456	Resistant	Pre-XDR	lineage2.2.1.1	Beijing-RD150	RD105; RD207; RD181; RD150	
PT_TB0311	ERR7764372	Susceptible	Susceptible	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0314	ERR7764364	Susceptible	Susceptible	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0305	ERR7764375	Resistant	Mono-DR	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0242	ERR7764378	Susceptible	Susceptible	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0245	ERR7764426	Susceptible	Susceptible	lineage4.1.2.1	T1; H1	RD182	
PT_TB0070	ERR2864232	Resistant	MDR	lineage3	CAS	RD750	
PT_TB0313	ERR7764325	Susceptible	Susceptible	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0250	ERR7764333	Susceptible	Susceptible	lineage4.8	T1; T2; T3; T5	RD219	
PT_TB0350	ERR7764370	Resistant	MDR	lineage4.3.4.2	LAM1; LAM4; LAM11	RD174	