## **Supplementary information**

## Fast and sensitive GCaMP calcium indicators for imaging neural populations

In the format provided by the authors and unedited

| Cycle | Frequency | Off  | On  | Trials | Volt (Lights Off) | Volt (Lights On) | Time (s) | Description |
|-------|-----------|------|-----|--------|-------------------|------------------|----------|-------------|
| 1     | 0.5       |      |     | 10     | 0                 | 5                | 20       | Frequency   |
| 2     |           | 25   | 500 | 25     | 0                 | 5                | 14       | Dark Flash  |
| 3     |           | 8    | 500 | 25     | 0                 | 5                | 12       | Dark Flash  |
| 4     |           | 4    | 500 | 25     | 0                 | 5                | 12       | Dark Flash  |
| 5     | 0.5       |      |     | 10     | 0                 | 5                | 20       | Frequency   |
| 6     | 30        |      |     | 600    | 0                 | 5                | 20       | Frequency   |
| 7     | 28        |      |     | 560    | 0                 | 5                | 20       | Frequency   |
| 8     | 26        |      |     | 520    | 0                 | 5                | 20       | Frequency   |
| 9     | 24        |      |     | 480    | 0                 | 5                | 20       | Frequency   |
| 10    | 22        |      |     | 440    | 0                 | 5                | 20       | Frequency   |
| 11    | 20        |      |     | 400    | 0                 | 5                | 20       | Frequency   |
| 12    | 18        |      |     | 360    | 0                 | 5                | 20       | Frequency   |
| 13    | 16        |      |     | 320    | 0                 | 5                | 20       | Frequency   |
| 14    | 14        |      |     | 280    | 0                 | 5                | 20       | Frequency   |
| 15    | 12        |      |     | 240    | 0                 | 5                | 20       | Frequency   |
| 16    | 10        |      |     | 200    | 0                 | 5                | 20       | Frequency   |
| 17    | 8         |      |     | 160    | 0                 | 5                | 20       | Frequency   |
| 18    | 6         |      |     | 120    | 0                 | 5                | 20       | Frequency   |
| 19    | 4         |      |     | 80     | 0                 | 5                | 20       | Frequency   |
| 20    | 2         |      |     | 40     | 0                 | 5                | 20       | Frequency   |
| 21    | 1         |      |     | 20     | 0                 | 5                | 20       | Frequency   |
| 22    | 0.5       |      |     | 10     | 0                 | 5                | 20       | Frequency   |
| 23    |           | 2000 | 500 | 15     | Ramp              | 5                | 45       | Ramp        |
| 24    |           | 1000 | 500 | 15     | Ramp              | 5                | 23       | Ramp        |
| 25    |           | 3000 | 500 | 15     | Ramp              | 5                | 67.5     | Ramp        |
| 26    |           | 25   | 500 | 25     | 2.5               | 5                | 12       | Dark Flash  |
| 27    |           | 25   | 500 | 25     | 0                 | 2.5              | 12       | Dark Flash  |
| 28    |           | 8    | 500 | 25     | 2.5               | 5                | 14       | Dark Flash  |
| 29    | 1         | 8    | 500 | 25     | 0                 | 2.5              | 14       | Dark Flash  |
| 30    |           | 500  | 25  | 25     | 0                 | 5                | 14       | Light Flash |
| 31    |           | 500  | 8   | 25     | 0                 | 5                | 12       | Light Flash |
| 32    |           | 500  | 4   | 25     | 0                 | 5                | 12       | Light Flash |
| 33    | 0.5       | 1    |     | 10     | 0                 | 5                | 20       | Frequency   |

## Supp. Table 3. Adult *Drosophila* L2 assay protocol

## Supp. Table 4. Summary of changes to the field stimulation pipeline between jGCaMP7 publication<sup>1</sup> and current manuscript:

|                             | jGCaMP7 publication <sup>1</sup>                               | Current publication   | Reason for change   |
|-----------------------------|--|---|---|
| Cell segmentation algorithm | Script with manually set thresholds                            | Cell segmentation with Ilastik <sup>2</sup> ,<br>a machine learning-based<br>algorithm                      | Better separation of somata from neuropil   |
| Source of neurons           | Neonatal rat<br>hippocampus                                    | Neonatal rat cortex   | More neurons can be<br>harvested from the cortex,<br>enabling higher<br>throughput          |
| Imaging speed               | 35 Hz  | 200 Hz  | Focus on kinetics in<br>current publication   |
| Quality control             | Wells with low<br>expression manually<br>removed from analysis | Wells with low expression<br>manually removed from<br>analysis; dim outlier cells<br>automatically excluded | Dim cells ( $F_0$ near<br>background) produce<br>artificially high $\Delta F/F_0$<br>signal |

1. Dana, H. *et al.* High-performance calcium sensors for imaging activity in neuronal populations and microcompartments. *Nat Methods* **16**, 649–657 (2019).

2. Berg, S. *et al.* ilastik: interactive machine learning for (bio)image analysis. *Nat Methods* **16**, 1226–1232 (2019).