Phylodynamic analysis of HIV-1 subtypes B, C and CRF 02_AG in Senegal

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Supplementary Material

² Parameter Estimation

We estimated the parameters of our mathematical model using the differential-evolution Markov 3 chain Monte Carlo (MCMC) zs sampler (MCMC-DEzs) (ter Braak and Vrugt, 2008). We initially run 4 few MCMC-DEzs for 3,000 to 4,000 iterations, and we chose one run per analyses to provide initial 5 conditions for subsequent longer runs. Because of computational resources, we run several longer 6 MCMC-DEzs ranging from 10,000 to 15,000 iterations using different initial conditions depending 7 on the analyses. These longer MCMC-DEzs were run in parallel using the computing resources of the Open Science Grid (OSG) (Pordes et al., 2007; Sfiligoi et al., 2009). We merged from 4 to 15 9 independent runs (also depending on the analyses) in order to have two sets of runs to compare 10 posterior distributions for each parameter and assess convergence of the chains. We also used the 11 Gelman diagnostics to check for convergence. 12 The calculation of the likelihood used in the MCMC-DEzs was carried out using the function *colik* 13

¹⁴ in R package *phydynR* version 0.1 (Volz, 2017). This function implements the structured coalescent ¹⁵ model (SCM) (Volz, 2012) which model each HIV-1 lineage in the phylogeny assuming that each ¹⁶ node in the phylogenetic tree corresponds to a single transmission event. For the calculation of ¹⁷ the likelihood we provided a demographic model using the *build.demographic.process* function in

 $_{18}$ phydynR, and we set the initial and end time of the calculations to 1978 and 2014, respectively.

¹⁹ For a list of parameters we estimated and their corresponding prior see Table S1.

Parameter	Prior
Transmission rate parameter of linear function gp0	Gamma(3, 3/0.1)
Transmission rate parameter of linear function gp1	Gamma(3, 3/0.1)
Transmission rate parameter of linear function gp2	Gamma(3, 3/0.1)
Linear function interval (time) for gp	U(1978, 2014)
Transmission rate parameter of linear function msm0	Gamma(3, 3/0.1)
Transmission rate parameter of linear function msm1	Gamma(3, 3/0.1)
Transmission rate parameter of linear function msm2	Gamma(3, 3/0.1)
Linear function interval (time) for msm	U(1978, 2014)
Risk ratio of gpm to transmit to $gpf(\psi)$	U(0.5, 2)
Importation rate	Exp(30)
Effective population size of src	Exp(1/100)
Probability of infected gpf to transmit to a gpm (p)	Beta(16, 4)
Probability of infected msm to transmit to a msm (q)	Beta(16, 4)
Initial number of infected msm	$\operatorname{Exp}(1/3)$
Initial number of infected gp	$\operatorname{Exp}(1/3)$
Removal rate (γ)	Fixed at $1/10$

Table S1: Summary of parameters estimated and MCMC priors

²⁰ Prevalence and Likelihood Calculation

We computed a statistic to calculate the proportion of infected heterosexual reproductive aged men (gpm) that are msm that we could compare to our mathematical model. For that we used available HIV-1 prevalence data for gpm in 2010 (4.0%, 95% CI: 14% – 80%) and for msm in 2016 (29.7%, 95% CI: 21.3% – 38.1%) in Dakar, Senegal (Mukandavire *et al.*, 2018). We also used surveillance data on the proportion of men who are msm (1.2%) (Mukandavire *et al.*, 2018), and we assumed that this proportion was independent of the estimated HIV prevalence for gpm and msm. Using this information, we have:

$$X = q \times p_{msm} / (q \times p_{msm} + (1 - q) \times p_m) \tag{1}$$

²⁸ Where q is the proportion of males who are msm; p_{msm} is HIV prevalence in msm; and p_m is ²⁹ HIV prevalence in gpm. We also extrapolated and assumed that msm HIV prevalence in 2010 was ³⁰ the same as in 2016.

³¹ We then approximate the standard deviation of X to a normal distribution, and recomputed ³² X for many replicates of q, p_m and p_{msm} using the differential-evolution Markov chain Monte ³³ Carlo zs sampler. We also calculated the "observed" X (X_{OBS}) for 2010 in our phylodynamic ³⁴ analysis, and the mean and standard deviation of X. Using this information we added the term ³⁵ dnorm(Xobs, Xmean, Xsd, log = TRUE) to the calculation of the likelihood. See scripts available ³⁶ at https://github.com/thednainus/senegalHIVmodel for further information on how we imple-³⁷ mented these calculations in R.

38 Statistical Analyses

After estimating the parameters of our mathematical model (Table S1), we calculated statistics of 39 interest in molecular epidemiology. We calculated these statistics using the MCMC-DEzs posterior 40 distribution, after removing the burnin, and for maximum a posteriori (MAP) estimates. Note 41 that in SCM as implemented in phydynR, we provided the ordinary differential equations (ODEs) as 42 matrices for birth and migration rates, and a vector for the removal rate (Volz, 2012). The birth 43 44 matrix represents the number of HIV transmissions within the different sub-populations or demes (*qpf, qpm, msm* and *src*). Similarly, the migration matrix represents allowed transmissions from one 45 sub-population/deme to another sub-population/deme, for example, transmissions from *qpm* to *qpf*. 46 For each parameter estimates in the posterior distribution, and for the MAP, we solved a de-47 mographic model using phydynR and estimated the birth and migration matrices, and the effective 48 number of infections for each time point from 1978 to 2014. 49

⁵⁰ In summary, the statistics we calculated for the dynamics of HIV in Senegal were:

- The population attributable fraction (PAF) for each group: *gpf*, *gpm* and *msm*, which are represented by each row in the birth matrix;
- The recent proportion of infections in one group attributable to another group. For example: ⁵⁴ proportion of infections in *gpf* attributable to *msm*;
- The effective number of infections for each group: *gpm*, *gpf* and *msm*.

56 **Results**

Below are the plots for the effective number of infections and population attributable fraction (PAF)
for each variation of the model not depicted in the main text.

- ⁵⁹ For the individual analyses for subtype CRF 02_AG and subtype C, the following applies:
- Model 1: We assigned each sequence to its respective risk-group in the phylogenetic tree a value of 1.0 (100% in the respective self-reported risk group);
- Model 2: We assumed some uncertainty in the self-reported *gpm* by assigning to every *gpm* sequence a value of 0.5 (50%) of being *gpm* and 0.5 (50%) of being *msm*;
- Model 3: We assigned each sequence to its respective risk-group in the phylogenetic tree a value of 1.0 (100% in the respective self-reported risk group) and added the prevalence term to the calculation of the likelihood. This plot is only shown in the main text only;
- Model 4: We assumed some uncertainty in the self-reported *gpm* by assigning to every *gpm* sequence a value of 0.5 (50%) of being *gpm* and 0.5 (50%) of being *msm*. We also added the prevalence term to the calculation of the likelihood.
- ⁷⁰ For the combined analyses using data from subtypes B, C and CRF 02_AG the following applies:
- Model 1: We assigned each sequence to its respective risk-group in the phylogenetic tree a value of 1.0 (100% in the respective self-reported risk group);
- Model 2: We removed all *gpm* sequences from the phylogenetic tree;
- Model 3: We assumed some uncertainty in the self-reported *gpm* by assigning to every *gpm* sequence a value of 0.5 (50%) of being *gpm* and 0.5 (50%) of being *msm*;

- Model 4: We assigned each sequence to its respective risk-group in the phylogenetic tree a 76 value of 1.0 (100% in the respective self-reported risk group) and added the prevalence term 77 to the calculation of the likelihood. This plot is only shown in the main text only; 78
- Model 5: We removed all *gpm* sequences from the phylogenetic tree and added the prevalence 79 term to the calculation of the likelihood; 80
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• Model 6: We assumed some uncertainty in the self-reported gpm by assigning to every gpm sequence a value of 0.5 (50%) of being gpm and 0.5 (50%) of being msm and added the prevalence term to the calculation of the likelihood.

Subtype C: Model 2 84

For subtype C model 2 we noticed that the MCMC runs for three parameters representing the linear 85 function for *msm* did not converge to the same posterior distribution. This could be attributable 86 to non-identifiability of these parameters. For MCMC posterior probability plots, see analyses/ 87 results/plots/mcmc_runs within the GitHub repository for the Senegal analyses. https://github. 88 com/thednainus/senegalHIVmodel. 89

To better understand if this was a potential non-identifiability problem, we solved PAF and effec-90 tive number of infections using both results from the MCMC posterior distributions. Results were 91 very similar as observed at results/plots/plots_withMaleX/subtypeC_model2 in the GitHub 92 repository https://github.com/thednainus/senegalHIVmodel. 93

Effective Number of Infections 94



Figure S1: Effective number of infections for subtype CRF 02_AG. Plots showing the proportion of the effective number of infections for gpf, gpm and msm. MAP = maximum a posteriori.



Figure S2: Effective number of infections for subtype C. Plots showing the proportion of the effective number of infections for gpf, gpm and msm. MAP = maximum a posteriori.



Figure S3: Effective number of infections for combined analyses. Plots showing the proportion of the effective number of infections for gpf, gpm and msm. MAP = maximum a posteriori.



Figure S4: Effective number of infections for combined analyses. Plots showing the proportion of the effective number of infections for gpf, gpm and msm. MAP = maximum a posteriori.

95 Population attributable fraction

Figure S5: **Population attributable fraction for subtype CRF 02_AG.** Plots showing the population attributable fraction for *gpf*, *gpm* and *msm*. Point estimates and error bars in the last plot represents 1-year PAF estimated for MSM in Mukandavire *et al.* (2018). MAP = maximum a posteriori.



Figure S6: **Population attributable fraction for subtype C.** Plots showing the population attributable fraction for *gpf*, *gpm* and *msm*. Point estimates and error bars in the last plot represents 1-year PAF estimated for MSM in Mukandavire *et al.* (2018). MAP = maximum a posteriori.



Figure S7: **Population attributable fraction for the combined analyses.** Plots showing the population attributable fraction for gpf, gpm and msm. Point estimates and error bars in the last plot represents 1-year PAF estimated for MSM in Mukandavire *et al.* (2018). MAP = maximum a posteriori.



Figure S8: **Population attributable fraction for the combined analyses.** Plots showing the population attributable fraction for gpf, gpm and msm. Point estimates and error bars in the last plot represents 1-year PAF estimated for MSM in Mukandavire *et al.* (2018). MAP = maximum a posteriori.



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