### Thioflavin Staining and Amyloid Formation are Unique to Mixed Tauopathies

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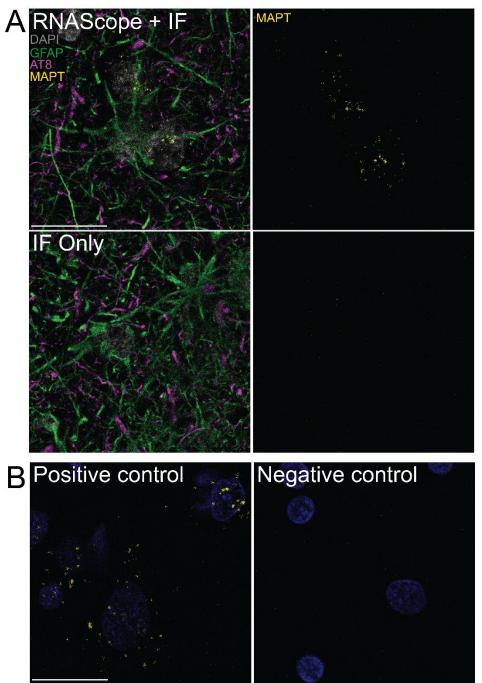
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# Supplementary Figure 1.



### Supplementary Figure 1.

Validation of RNAscope and immunofluorescence staining techniques. (*A*) Immunofluorescence staining without *MAPT* probe (yellow) shows no *in situ* hybridization signal compared to adjacent region on the same slide of human brain tissue stained with probe. (*B*) Further validation using manufacturer provided positive (yellow) and negative control probes on adjacent regions on the same slide of human brain tissue. DAPI (blue) was used to highlight nuclei. Scale bar=  $20\mu m$  for both (*A*) and (*B*).

# Supplementary Figure 2. Quantification code

ds=imageDatastore('Binarized Images\'); %create an image store of Cy5 images file names=ds.Files; %read file names [a,b]=size(file names); %determine how many files there are Cy5=zeros([a,1]); %initialize the vector (saves processing time) for i=1:a I=readimage(ds,i); %read image i x=nnz(I); %calculate the number of non-zero pixels in the %thresholded image Cy5(i,1)=x; %add to vector

```
end
```

%create a vector for the diagnoses, transpose it to a vertical, and turn it into a categorical Dx=["AD" "PSP" "CBD" "Control" "AD" "Control" "AD" "CBD" "CBD" "PSP" "PSP"]; Dx=Dx'; Dx=categorical(Dx);

```
%%create indices for each diagnosis
idx_AD=Dx=="AD";
idx_CBD=Dx=="CBD";
idx_PSP=Dx=="PSP";
idx_Con=Dx=="Control";
```

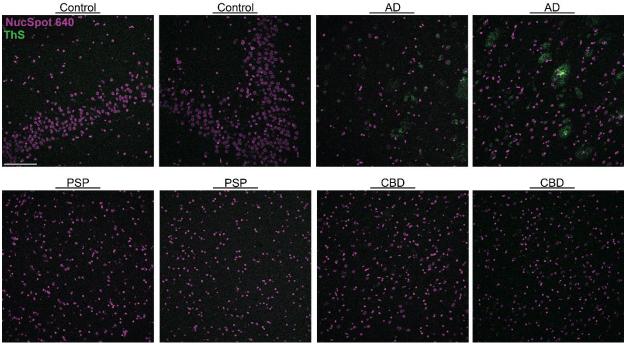
%%run ANOVA to check if any comparison is significant. [~,~,stats]=anova1(Cy5,Dx);

```
%%run multcompare to figure out which
[c,~,~,gnames]=multcompare(stats);
tbl = array2table(c,"VariableNames", ...
["Group A","Group B","Lower Limit","A-B","Upper Limit","P-value"]);
tbl.("Group A") = gnames(tbl.("Group A"));
tbl.("Group B") = gnames(tbl.("Group B"))
```

%generate the plot scatter(1,Cy5(idx\_Con),"black","filled") hold on scatter(2,Cy5(idx\_AD),"black","filled") scatter(3,Cy5(idx\_CBD),"black","filled") scatter(4,Cy5(idx\_PSP),"black","filled") ylabel("Number of Positive Pixels") Thioflavin S is Unique to Mixed Tauopathies

```
xlabel("")
xticks([1 2 3 4])
xticklabels({'Control' 'AD' 'CBD' 'PSP'})
xlim([0.5 4]);
line([1 2],[29000 29000])
text(1.5,30000,"p="+round(tbl{2,6},4,"significant"),'HorizontalAlignment', 'center')
line([2 3],[28000 28000])
text(2.5,29000,"p="+round(tbl{3,6},4,"significant"),'HorizontalAlignment', 'center')
line([2 4],[30000 30000])
text(3.5,3.1e4,"p="+round(tbl{1,6},4,"significant"),'HorizontalAlignment', 'center')
hold off
```

# Supplementary Figure 3.



Supplementary Figure 3.

Validation of thioflavin S staining pattern using alternative source of thioflavin S (see methods). Thioflavin staining (green) is present in AD but is not found in control, PSP, or CBD. NucSpot 640 (magenta) was used to highlight nuclei. Two cases are shown per disease (total n=8). Scale bar= 50µm. Abbreviations: *AD* Alzheimer disease; *PSP* progressive supranuclear palsy; *CBD* corticobasal degeneration; *ThS* thioflavin S

**Supplementary Table 1.** Quantification of colocalization using FIJI BIOP JACOP to calculate Mander's M1/M2 and Pearson's correlation coefficients for images shown in main text Fig. 2.

Case ID	Pathologic Diagnosis	Area A	Area B	Area Overlap	Pearson's Coefficient
#3	Control	0.012	0.856	0	-3.15E-04
#8	AD	81.496	18.834	9.876	0.288
#13	PSP	5.346	2.067	0.047	0.016
#20	CBD	6.455	8.827	0.163	0.02
#22	PiD	10.668	0.093	3.86E-04	-8.35E-04
#24	ALS and	9.992	8.778	1.684	0.245
	PART				

Case ID	M1	M2	Threshold A	Threshold B	Thresholded M1	Thresholded M1
#3	1.11E-04	1.53E-04	92	37	0	0
#8	0.331	0.641	116	97	0.126	0.531
#13	0.049	0.039	98	37	0.01	0.023
#20	0.128	0.032	111	37	0.027	0.019
#22	4.47E-04	0.029	116	31	3.23E-05	0.003
#24	0.272	0.203	139	37	0.176	0.302

AD Alzheimer's disease; PSP Progressive supranuclear palsy; CBD Corticobasal degeneration; PiD Pick's disease; ALS Amyotrophic lateral sclerosis; PART Primary age-related tauopathy