

Sequence and structural motifs controlling the broad substrate specificity of the mycobacterial hormone sensitive lipase LipN

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Running title: Substrate specificity of LipN

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Table S1: Kinetic characterization of LipN with fluorogenic esters.

	k_{cat} (s^{-1})	K_{m} (μM)	$k_{\text{cat}}/K_{\text{m}}$ ($\text{M}^{-1}\text{s}^{-1}$)
Alkyl Esters			
1^a	0.10 ± 0.038	0.22 ± 0.12	460000 ± 150000
2	0.18 ± 0.06	4.2 ± 0.5	42000 ± 15000
3	0.062 ± 0.029	6.2 ± 0.1	10000 ± 5000
4	0.024 ± 0.001	3.8 ± 0.5	6400 ± 1400
5	0.0074 ± 0.0047	4.4 ± 0.9	1700 ± 600
6	0.017 ± 0.014	9.5 ± 2.2	1800 ± 800
Polar Esters			
7	0.11 ± 0.12	2.6 ± 1.1	40000 ± 25000
8	0.093 ± 0.048	2.8 ± 0.5	34000 ± 9000
9	0.12 ± 0.12	3.0 ± 1.1	39000 ± 21000
10	0.0005 ± 0.0004	21 ± 4	26 ± 11
11	0.015 ± 0.008	3.1 ± 0.6	4900 ± 1400
α/β Unsaturated Esters			
12	0.020 ± 0.030	3.4 ± 1.7	6100 ± 4700
13	0.0093 ± 0.0050	0.032 ± 0.08	29000 ± 9000
14	0.0029 ± 0.0023	0.051 ± 0.19	5700 ± 2500
15	0.0061 ± 0.0047	1.8 ± 0.5	3300 ± 1400
Cycloalkyl Esters			
16	0.025 ± 0.017	3.0 ± 0.7	8300 ± 3100
17	0.0012 ± 0.0001	33.9 ± 7.71	29 ± 4
18	0.00070 ± 0.00060	29 ± 5	25 ± 11
19	0.0030 ± 0.0048	0.84 ± 0.60	3500 ± 3100
20	0.0030 ± 0.0001	0.033 ± 0.016	91000 ± 22000
21	0.0003 ± 0.0013	1.2 ± 1.7	310 ± 690
Tertiary Esters			
22	0.000024 ± 0.000030	0.71 ± 0.41	34 ± 23
23	0.000026 ± 0.000020	0.049 ± 0.027	530 ± 250
24	0.000011 ± 0.000030	0.87 ± 1.1	12 ± 19
25	0.0012 ± 0.0018	1.7 ± 1.1	670 ± 590
26	0.000058 ± 0.00014	13 ± 8	4.4 ± 5.4
27	0.0029 ± 0.0016	0.40 ± 0.10	7400 ± 2200
28	0.000014 ± 0.000050	0.084 ± 0.15	170 ± 330
29	0.00090 ± 0.0011	0.37 ± 0.20	2600 ± 1700
30	0.0011 ± 0.0018	1.7 ± 1.1	660 ± 590
Fluorinated Esters			
31	0.013 ± 0.004	1.1 ± 0.2	11000 ± 2000
32	0.00023 ± 0.00020	47 ± 8	4.8 ± 2.2
33	0.000037 ± 0.000030	0.37 ± 0.13	98 ± 43

^aKinetic constants for substrates **1–33** were determined by measuring the increase in fluorogenic enzyme substrate fluorescence over time. Data were fitted to a standard Michaelis-Menten equation to determine the values for k_{cat} , K_{M} , and $k_{\text{cat}}/K_{\text{M}}$. Kinetic measurements for each substrate were repeated at least three times and the values are given \pm SD.

Table S2: Kinetic characterization of LipN with *p*-nitrophenyl esters.

Substrate	k_{cat} (s ⁻¹)	K_{m} (mM)	$k_{\text{cat}}/K_{\text{m}}$ (M ⁻¹ s ⁻¹)
C2^a	94 ± 8	2.4 ± 1.2	39000 ± 10000
C4	190 ± 10	0.020 ± 0.009	9300000 ± 2100000
C6	15 ± 1	0.25 ± 0.07	59000 ± 8000
C8	1.1 ± 0.2	0.41 ± 0.33	2700 ± 1100
C14	0.28 ± 0.02	0.12 ± 0.05	2300 ± 400

^aKinetic constants for *p*-nitrophenyl substrates were determined by measuring the change in A₄₁₂ due to ester hydrolysis. Data were fitted to a standard Michaelis-Menten equation to determine the values for k_{cat} , K_{M} , and $k_{\text{cat}}/K_{\text{M}}$. Kinetic measurements for each substrate were repeated at least three times and the values are given ± SD. Substrates represent different carbon chain lengths: *p*-nitrophenyl acetate (C2), *p*-nitrophenyl butyrate (C4), *p*-nitrophenyl valerate (C6), *p*-nitrophenyl octanoate (C8), and *p*-nitrophenyl myristate (C14).

Table S3: Biochemical characterization of active site and binding pocket variants of LipN.

LipN	k_{cat} (10^{-3} s^{-1}) ^a	K_{m} (μM)	$k_{\text{cat}}/K_{\text{m}}$ ($\text{M}^{-1}\text{s}^{-1}$)	T_{m} ($^{\circ}\text{C}$) ^b
<i>Wild-type</i>				52.0 ± 0.6
1	0.10 ± 0.038	0.22 ± 0.12	460000 ± 150000	
7	0.11 ± 0.12	2.6 ± 1.1	40000 ± 25000	
20	0.0030 ± 0.0001	0.033 ± 0.016	91000 ± 22000	
<i>G144A</i>				52.3 ± 0.3
1	0.017 ± 0.001	5.4 ± 0.5	3300 ± 200	
7	0.066 ± 0.024	60 ± 15	1100 ± 200	
20	0.0029 ± 0.0001	1.5 ± 0.4	2000 ± 300	
<i>G145A</i>				50.7 ± 0.3
1	0.045 ± 0.012	3.4 ± 1.3	13000 ± 3000	
7	0.18 ± 0.11	120 ± 60	1500 ± 600	
20	0.0019 ± 0.0005	0.30 ± 0.16	6200 ± 1800	
<i>H154A</i>				42.7 ± 0.7
1	0.030 ± 0.004	32 ± 4	940 ± 80	
7	0.28 ± 0.15	694 ± 422	410 ± 170	
20	0.0024 ± 0.0002	3.4 ± 0.4	700 ± 50	
<i>D215A</i>				49.0 ± 0.6
1	0.038 ± 0.012	2.5 ± 1.2	16000 ± 4000	
7	0.039 ± 0.017	3.3 ± 2.1	12000 ± 4000	
20	0.0023 ± 0.0003	0.49 ± 0.13	4600 ± 700	
<i>S216G</i>				63.0 ± 0.5
1	0.00070 ± 0.00027	21 ± 16	34 ± 15	
7	0.00018 ± 0.00006	22 ± 2	82 ± 4	
20	0.000021 ± 0.000003	3.6 ± 1.8	5.7 ± 1.5	
<i>R246A</i>				49.0 ± 0.6
1	0.054 ± 0.011	21 ± 4	2500 ± 370	
7	0.10 ± 0.02	150 ± 40	680 ± 100	
20	0.089 ± 0.010	120 ± 20	760 ± 80	
<i>D269A</i>				46.0 ± 0.6
1	0.0055 ± 0.0003	5.8 ± 0.4	960 ± 50	
7	0.032 ± 0.017	15 ± 8	2200 ± 800	
20	0.046 ± 0.0001	5.7 ± 1.9	1700 ± 300	
<i>D312A</i>				52.7 ± 0.3
1	0.19 ± 0.06	28 ± 7	6700 ± 1400	
7	0.31 ± 0.29	35 ± 27	9000 ± 5500	
20	0.0039 ± 0.0008	5.7 ± 1.9	4300 ± 800	
<i>H342A</i>				51.3 ± 0.3
1	0.0000032 ± 0.000001	0.92 ± 1.15	3.5 ± 2.2	
7	0.000053 ± 0.000017	20 ± 6	2.6 ± 0.6	
20	0.0000033 ± 0.0000033	0.31 ± 0.63	11 ± 12	

^aKinetic constants for substrates **1**, **7**, and **20** were determined by measuring the increase in fluorogenic enzyme substrate fluorescence over time. Data were fitted to a standard Michaelis-Menten equation to determine the values for k_{cat} , K_{M} , and $k_{\text{cat}}/K_{\text{M}}$. Kinetic measurements for each substrate were repeated at least three times and the values are given \pm SD.

^bValues for T_{m} were determined by following the change in Sypro Orange fluorescence with increasing temperature. Melting curves were repeated at least three times for each variant and the T_{m} values reported \pm SD.

Table S4: Biochemical characterization of Ser216 variants of LipN.

LipN	k_{cat} (10^{-3} s^{-1}) ^a	K_{m} (μM)	$k_{\text{cat}}/K_{\text{m}}$ ($\text{M}^{-1}\text{s}^{-1}$)	T_{m} ($^{\circ}\text{C}$) ^b
<i>Wild-type</i>				52.0 ± 0.6
1	0.10 ± 0.038	0.22 ± 0.12	460000 ± 150000	
7	0.11 ± 0.12	2.6 ± 1.1	40000 ± 25000	
20	0.0030 ± 0.0001	0.033 ± 0.016	91000 ± 22000	
<i>S216C</i>				56.7 ± 0.2
1	0.0031 ± 0.0019	82 ± 68	37 ± 19	
7	0.0032 ± 0.0003	60 ± 9	53 ± 5	
20	0.000061 ± 0.000022	120 ± 70	0.53 ± 0.17	
<i>S216G</i>				63.0 ± 0.5
1	0.00070 ± 0.00027	21 ± 16	34 ± 15	
7	0.00018 ± 0.00006	22 ± 2	82 ± 4	
20	0.000021 ± 0.000003	3.6 ± 1.8	5.7 ± 1.5	
<i>S216N</i>				49.5 ± 1.3
1	0.00082 ± 0.0003	26 ± 3	31 ± 2	
7	0.0021 ± 0.0010	41 ± 33	51 ± 24	
20	0.00012 ± 0.00003	26 ± 15	15 ± 5	
<i>S177A</i>				57.2 ± 0.3
1	1.3 ± 0.1	1.1 ± 0.1	110000 ± 10000	
7	1.8 ± 0.1	6.4 ± 0.6	270000 ± 30000	
20	0.019 ± 0.001	2.0 ± 0.7	9400 ± 1700	
<i>S226A</i>				53.0 ± 0.5
1	3.6 ± 0.1	20 ± 2	180000 ± 20000	
7	2.3 ± 0.1	31 ± 4	73000 ± 5000	
20	0.017 ± 0.001	1.1 ± 0.1	15000 ± 1000	

^aKinetic constants for substrates **1**, **7**, and **20** were determined by measuring the increase in fluorogenic enzyme substrate fluorescence over time. Data were fitted to a standard Michaelis-Menten equation to determine the values for k_{cat} , K_{M} , and $k_{\text{cat}}/K_{\text{M}}$. Kinetic measurements for each substrate were repeated at least three times and the values are given \pm SD.

^bValues for T_{m} were determined by following the change in Sypro Orange fluorescence with increasing temperature. Melting curves were repeated at least three times for each variant and the T_{m} values reported \pm SD.

Table S5: Biochemical characterization of Trp146 variants of LipN.

LipN	k_{cat} (10^{-3} s^{-1}) ^a	K_{m} (μM)	$k_{\text{cat}}/K_{\text{m}}$ ($\text{M}^{-1}\text{s}^{-1}$)	T_{m} ($^{\circ}\text{C}$) ^b
<i>Wild-type</i>				52.0 ± 0.6
1	0.10 ± 0.038	0.22 ± 0.12	460000 ± 150000	
7	0.11 ± 0.12	2.6 ± 1.1	40000 ± 25000	
20	0.0030 ± 0.0001	0.033 ± 0.016	91000 ± 22000	
<i>W146A</i>				40.0 ± 0.7
1	0.075 ± 0.006	2.0 ± 0.8	37000 ± 7000	
7	0.067 ± 0.004	1.1 ± 0.3	62000 ± 10000	
20	0.0013 ± 0.0001	0.71 ± 0.11	1800 ± 200	
<i>W146C</i>				45.0 ± 0.4
1	0.013 ± 0.006	23 ± 26	570 ± 340	
7	0.15 ± 0.01	13 ± 1	11000 ± 1000	
20	0.000061 ± 0.000022	120 ± 70	0.53 ± 0.17	
<i>W146Q</i>				45.0 ± 0.5
1	0.064 ± 0.004	1.4 ± 0.4	45000 ± 7000	
7	0.075 ± 0.0023	2.2 ± 0.3	34000 ± 2000	
20	0.0020 ± 0.0002	3.5 ± 1.3	550 ± 110	
<i>W146S</i>				33.0 ± 0.6
1	0.091 ± 0.007	23 ± 5	4000 ± 500	
7	0.17 ± 0.01	4.1 ± 0.5	42000 ± 3000	
20	0.0017 ± 0.0001	3.5 ± 0.4	500 ± 30	

^aKinetic constants for substrates **1**, **7**, and **20** were determined by measuring the increase in fluorogenic enzyme substrate fluorescence over time. Data were fitted to a standard Michaelis-Menten equation to determine the values for k_{cat} , K_{M} , and $k_{\text{cat}}/K_{\text{M}}$. Kinetic measurements for each substrate were repeated at least three times and the values are given \pm SD.

^b Values for T_{m} were determined by following the change in Sypro Orange fluorescence with increasing temperature. Melting curves were repeated at least three times for each variant and the T_{m} values reported \pm SD.

Table S6: Substitutions in substrate binding and catalytic residues among hydrolases similar to LipN from *M. marinum*.

Organism	144	145	146	154	177	215	216	226	246	269	312	342	% Identity
<i>M. marinum</i>	G	G	W	H	S	D	S	S	R	D	D	H	-
<i>M. kansasii</i>				A	A			C					78
<i>M. palustre</i>					A			C					75
<i>M. tuberculosis</i> (LipN)					A			C					73
<i>M. avium</i>			F		A			A	V				64
<i>M. smegmatis</i>			F		A				I				53
<i>M. kubicæ</i>			F		A				G				56
<i>M. abscessus</i>					A			A	A	A			53
<i>B. cereus</i>				M	A			T	T	Q			36
<i>S. acidocaldarius</i>			F	Y	A				A	Q			39
<i>L. monocytogenes</i>			F		A			T	A	S			36
<i>S. pombe</i>	S			D	A	A			T	I			30
<i>E. coli</i>			F					A	L				26
<i>P. aeruginosa</i>					A			C	L				37
<i>M. tuberculosis</i> (LipW)			Y	D	A			A	M	A			37
<i>S. multispecies</i>						A		T	V	A			35
<i>S. oneidensis</i>		C	F		A			C	M	T			31
<i>D. discoideum</i>			F	V	A			A	I	F			32
<i>P. putida</i>			F		A				V	M			44
<i>P. fluorescens</i>			F		A				V	M			44
<i>D. radiodurans</i>			F		A			A	A	R			44
<i>B. multivorans</i>			F		A			A	V	A			40
<i>B. cenocepacia</i>			F		A			A	V	A			40

^aThe amino acid sequence of *MmLipN* was aligned using Clustal Omega (EMBL EBI). The sequences used in the alignment were from *Mycobacterium marinum* (WP_036455371.1), *Mycobacterium kansasii* (ORB86246.1), *Mycobacterium palustre* (WP_085077243.1), *Mycobacterium tuberculosis* LipN (COV27923.1), *Mycobacterium avium* (WP_029248963.1), *Mycobacterium smegmatis* (WP_011728324.1), *Mycobacterium kubicæ* (WP_085075701.1), *Mycobacteroides abscessus* (WP_145044121.1), *Bacillus cereus* (WP_098523648.1), *Sulfolobus acidocaldarius* (WP_011277970.1), *Listeria monocytogenes* (WP_099183763.1), *Schizosaccharomyces pombe* (NP_593998.1), *Escherichia coli* str. K-12 (NP_415009.1), *Pseudomonas aeruginosa* PAO1 (NP_254071.1), *Mycobacterium tuberculosis* LipW (NP_214731.1), *Streptomyces multispecies* (WP_003972012.1), *Shewanella oneidensis* (WP_011071097.1), *Dictyostelium discoideum* (XP_638888.1), *Pseudomonas putida* (WP_110963922.1), *Pseudomonas fluorescens* (WP_150715891.1), *Deinococcus radiodurans* (WP_027480227.1), *Burkholderia multivorans* (WP_105844151.1), *Burkholderia cenocepacia* (WP_060264533.1). Sequences for alignment were chosen based on protein BLAST analysis of *MmLipN* and extracting unique protein sequences from model organisms with significant percent similarity (> 20%).

^bCatalytic triad residues are highlighted in blue.

Table S7: Substitutions in bHSL motifs among hydrolases similar to LipN from *M. marinum*.

Organism	142	143	144	145	146	214	215	216	217	218	% Identity
<i>M. marinum</i>	H	G	G	G	W	G	D	S	A	G	-
<i>M. kansasii</i>											78
<i>M. palustre</i>											75
<i>M. tuberculosis</i> (LipN)											73
<i>M. avium</i>					F						64
<i>M. smegmatis</i>					F						53
<i>M. kubicae</i>					F						56
<i>M. abscessus</i>											53
<i>B. cereus</i>									V		36
<i>S. acidocaldarius</i>					F						39
<i>L. monocytogenes</i>					F				V		36
<i>S. pombe</i>		S	S				A				30
<i>E. coli</i>					F						26
<i>P. aeruginosa</i>											37
<i>M. tuberculosis</i> (LipW)		A			Y						37
<i>S. multispecies</i>							A		V		35
<i>S. oneidensis</i>				C	F						31
<i>D. discoideum</i>		S			F				V		32
<i>P. putida</i>					F						44
<i>P. fluorescens</i>					F						44
<i>D. radiodurans</i>					F						44
<i>B. multivorans</i>					F						40
<i>B. cenocepacia</i>					F						40

^aThe amino acid sequence of *MmLipN* was aligned using Clustal Omega (EMBL EBI). The sequences used in the alignment were from *Mycobacterium marinum* (WP_036455371.1), *Mycobacterium kansasii* (ORB86246.1), *Mycobacterium palustre* (WP_085077243.1), *Mycobacterium tuberculosis* LipN (COV27923.1), *Mycobacterium avium* (WP_029248963.1), *Mycobacterium smegmatis* (WP_011728324.1), *Mycobacterium kubicae* (WP_085075701.1), *Mycobacteroides abscessus* (WP_145044121.1), *Bacillus cereus* (WP_098523648.1), *Sulfolobus acidocaldarius* (WP_011277970.1), *Listeria monocytogenes* (WP_099183763.1), *Schizosaccharomyces pombe* (NP_593998.1), *Escherichia coli* str. K-12 (NP_415009.1), *Pseudomonas aeruginosa* PAO1 (NP_254071.1), *Mycobacterium tuberculosis* LipW (NP_214731.1), *Streptomyces multi-species* (WP_003972012.1), *Shewanella oneidensis* (WP_011071097.1), *Dictyostelium discoideum* (XP_638888.1), *Pseudomonas putida* (WP_110963922.1), *Pseudomonas fluorescens* (WP_150715891.1), *Deinococcus radiodurans* (WP_027480227.1), *Burkholderia multivorans* (WP_105844151.1), *Burkholderia cenocepacia* (WP_060264533.1). Sequences for alignment were chosen based on protein BLAST analysis of *MmLipN* and extracting unique protein sequences from model organisms with significant percent similarity (> 20%).

^bHGGGW motif shaded gray.

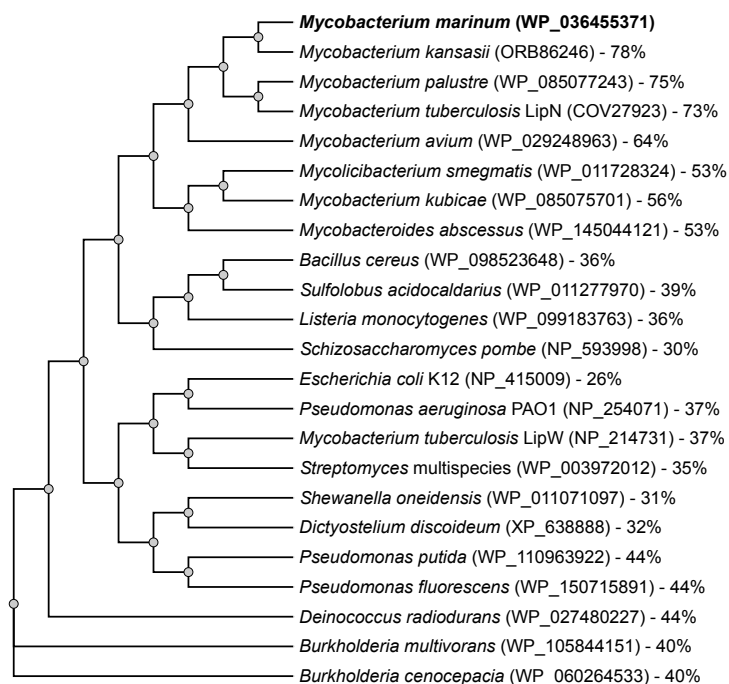


Figure S1: Phylogenetic relationship between *MmLipN* and homologous serine hydrolases across various model organisms. The amino acid sequence of LipN was aligned with the 22 homologues and a cladogram of the aligned proteins was constructed with percent similarities from Clustal Omega. A linear dendrogram with decreasing ladderizing was then constructed using FastTree from NgPhylogeny.fr. Sequences for alignment were chosen based on protein BLAST analysis of *MmLipN* and extracting unique protein sequences from model organisms with significant percent similarity (> 20%). Detailed sequence analysis is given in Tables S6 and S7.

Supplemental Methods:

The syntheses of compounds (**1**, **2**, **12**, **16**, **22**)¹, (**7**, **8**, **9**, **13**, **17**, **18**, **19**, **21**, **31**, **32**)², and (**3**, **4**, **5**)³ have been previously described. The synthesis of the common starting material for divergent synthesis, Fluorescein bis(chloromethyl ether), has been previously described⁴. Carboxylic acids were obtained from Sigma Aldrich, Alfa Aesar, Enamine, Matrix Scientific, and Chem-Bridge. Compounds **24** and **30** are each mixtures of stereoisomers that presumably resemble the isomeric mixture of the corresponding carboxylic acid starting material in distribution. All other chemicals were purchased from Sigma Aldrich and used without further purification. Nuclear magnetic resonance (NMR) spectra were obtained using a Bruker Biospin Avance III HD 400 operating at 400.19 MHz for ¹H and 100.64 MHz for ¹³C. High resolution mass spectrometry (HRMS) was performed with electrospray ionization (ESI) by the Mass Spectrometry facility at the Department of Chemistry, Indiana University using an Agilent 1200 HPLC-6130 MSD mass spectrometer.

The following procedure is representative for the synthesis of all compounds.

Synthesis of Fluorescein bis((2-methyl)propionyloxymethyl ether) (6). Fluorescein bis(chloromethyl ether) (30.0 mg, 69.9 μmol, 1.0 Eq), (2-methyl)propanoic acid (20.1 mg, 280 μmol, 4.0 Eq.), and Cs₂CO₃ (91.1 mg, 279.6 μmol, 4.0 Eq.) were dissolved in dry CH₃CN (1 mL). Molecular sieves (100 mg) were added, and the reaction was covered in foil and allowed to stir for 24 h at ambient temperature. The reaction mixture was adsorbed onto celite and purified via silica column chromatography.

Fluorescein bis((2-methyl)propionyloxymethyl ether) (6)

Data for **6**: (79%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.16 (d, 1H), 6.96 (m, 2H), 6.73 (m, 4H), 5.79 (s, 4H), 2.67-2.56 (sept., 2H), 1.15 (d, 12 H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 175.8, 169.2, 158.4, 152.8, 152.2, 135.1, 129.8, 129.3, 126.6, 125.1, 123.9, 113.1, 112.7, 103.5, 84.9, 82.5, 33.9, 18.7 ppm. HRMS (ESI): calc'd for MH⁺ C₃₀H₂₉O₉: 533.1806; found: 533.1801.

Fluorescein bis(3-(2-methoxyethoxy)propionyloxymethyl ether) (10)

Data for **10**: (27%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.61-7.70 (m, 2H), 7.16 (d, 1H), 6.96 (t, 2H), 6.73 (d, 4H), 5.80 (s, 4H), 3.77 (t, 4H), 3.53 (dq, 8H), 3.34 (s, 6H), 2.69 (t, 4H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 170.4, 169.2, 158.4, 153.0, 152.2, 135.1, 129.9, 129.4, 126.6, 125.2, 123.9, 113.3, 112.8, 103.6, 85.0, 82.4, 71.8, 70.4, 66.2, 59.1, 35.0 ppm. HRMS (ESI): calc'd for MNa⁺ C₃₄H₃₆O₁₃Na: 675.2054; found: 675.2041.

Fluorescein bis(cyanoacetyloxymethyl ether) (11)

Data for **11**: (6%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.16 (d, 1H), 6.75 (m, 2H), 6.72 (m, 4H), 5.87(s, 4H), 3.55 (s, 4H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 169.5, 162.0, 157.8, 152.9, 152.3, 135.3, 129.9, 129.5, 126.6, 125.2, 123.9, 113.9, 112.7, 112.2, 103.7, 86.5, 82.8, 24.8 ppm. HRMS (ESI): calc'd for MH⁺ C₂₈H₁₉N₂O₉: 527.1085; found: 527.1091.

Fluorescein bis(crotonyloxymethyl ether) (14)

Data for **14**: (65%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.70-7.61 (m, 2H), 7.16 (d, *J* = 8.05 Hz, 1H), 7.14-7.05 (m, 2H), 6.98 (s, 2H), 6.74 (s, 4H), 5.89 (d, *J* = 16.8 Hz, 2H), 5.85 (s, 4H), 1.91 (d, *J* = 7.02 Hz, 6H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 169.2, 164.9, 158.5, 152.9, 152.2, 147.2, 135.1, 129.9, 129.4, 126.7, 125.2, 123.9, 121.7, 113.2, 112.7, 103.5, 84.9, 82.5, 18.2 ppm. HRMS (ESI): calc'd for MH⁺ C₃₀H₂₅O₉: 529.1498; found: 529.1516.

Fluorescein bis((3-methyl)propargyloxymethyl ether) (15)

Data for **15**: (68%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.15 (d, 1H), 6.98 (m, 2H), 6.75 (d, 4H), 5.83 (s, 4H), 2.01 (s, 6H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 170.6,

169.2, 158.2, 153.0, 152.2, 135.1, 129.9, 129.4, 126.5, 125.1, 123.9, 113.6, 112.7, 103.7, 88.1, 85.6, 82.3, 71.7, 3.9 ppm. HRMS (ESI): calc'd for MH^+ $C_{30}H_{21}O_9$: 525.1180; found: 525.1183.

Fluorescein bis((5-oxazolyl)carboxymethyl ether) (20)

Data for **20**: (59%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.08 (s, 2H), 8.05 (d, 1H), 7.89 (s, 2H), 7.72-7.64 (m, 2H), 7.17 (d, 1H), 7.04 (d, 2H), 6.83-6.77 (m, 4H), 6.04 (s, 4H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 169.1, 158.1, 156.1, 154.0, 152.8, 152.2, 141.8, 135.2, 134.9, 130.0, 129.6, 126.5, 125.2, 123.8, 113.8, 112.7, 103.7, 85.6, 82.1 ppm. HRMS (ESI): calc'd for MH^+ $C_{30}H_{19}N_2O_{11}$: 583.0989; found: 583.0981.

Fluorescein bis(((1-(2-pyridinyl))cyclopropyl)carboxymethyl ether) (23)

Data for **23**: (56%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.53 (d, 2H), 8.04 (d, 1H), 7.7-7.6 (m, 2H), 7.58 (dd, 2H), 7.41 (d, 2H), 7.17-7.13 (dd, 2H), 7.16 (s, 1H), 6.69 (m, 2H), 6.63-6.61 (m, 4H), 5.75 (s, 4H), 1.74-1.71 (m, 4H), 1.53-1.52 ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 172.3, 169.0, 158.1, 157.0, 152.7, 151.9, 148.8, 136.0, 134.9, 129.7, 129.0, 126.4, 124.9, 124.7, 123.7, 122.0, 113.0, 112.5, 103.5, 85.0, 82.2, 30.4, 17.5 ppm. HRMS (APCI): calc'd for MH^+ = $C_{40}H_{31}N_2O_9$: 683.2029; found: 683.2024.

Fluorescein bis(((2,2-dichloro-1-methyl)cyclopropyl)carboxymethyl ether) (24)

Data for **24**: (57%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.04 (d, 1H), 7.7-7.6 (m, 2H), 7.18 (d, 1H), 6.98 (m, 2H), 6.75 (m, 4H), 5.92-5.82 (d, 4H), 2.31 (m, 2H), 1.60 (s, 6H), 1.47 (m, 2H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 169.2, 168.2, 158.3, 152.9, 152.2, 135.2, 129.9, 129.5, 126.6, 125.2, 123.9, 113.59, 113.56, 112.8, 112.6, 103.8, 103.7, 86.2, 82.3, 62.4, 35.3, 31.0, 18.1 ppm. HRMS (APCI): calc'd for MH^+ = $C_{32}H_{25}Cl_4O_9$: 693.0247; found = 693.0253; calc'd for MNa^+ = $C_{32}H_{24}Cl_4O_9Na$: 715.0067; found = 715.0071.

Fluorescein bis(((1-cyano)cyclopropyl)carboxymethyl ether) (25)

Data for **25**: (78%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 7.98 (d, 1H), 7.6-7.5 (m, 2H), 7.12 (d, 1H), 6.95 (m, 2H), 6.71 (m, 4H), 5.79 (s, 4H), 1.75-1.65 (m, 8H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 169.2, 167.0, 158.1, 152.8, 152.2, 135.3, 130.0, 129.6, 126.6, 125.2, 123.9, 117.9, 113.9, 112.7, 112.4, 104.0, 103.1, 86.5, 82.2, 19.9, 13.4 ppm. HRMS (APCI): calc'd for MH^+ = $C_{32}H_{23}N_2O_9$: 579.1398, found: 579.1399; calc'd for MNa^+ for $C_{32}H_{22}N_2O_9Na$: 601.1218, found: 601.1218

Fluorescein bis(((1-phenyl)cyclopropyl)carboxymethyl ether) (26)

Data for **26**: (29%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.05 (d, 1H), 7.7-7.6 (m, 2H), 7.29-7.26 (m, 10H), 7.16 (d, 1H), 6.78 (m, 2H), 6.68-6.58 (m, 4H), 5.70 (s, 4H), 1.67-1.63 (m, 4H), 1.28-1.26 (m, 4H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 173.4, 169.3, 158.4, 153.0, 152.1, 138.6, 135.1, 130.5, 129.9, 129.2, 128.3, 127.5, 126.6, 125.2, 123.9, 113.2, 112.8, 104.0, 85.4, 82.4, 29.2, 17.2 ppm. HRMS (APCI): calc'd for MH^+ = $C_{42}H_{33}O_9$: 681.2119; found: 681.2125.

Fluorescein bis(((1-hydroxy)cyclopropyl)carboxymethyl ether) (27)

Data for **27**: (43%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.16 (d, 1H), 6.92 (m, 2H), 6.73 (s, 4H), 5.84 (m, 4H), 2.97 (s, 2H), 1.36-1.33 (m, 4H), 1.24-1.21 (m, 4H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 174.0, 169.2, 158.2, 152.87, 152.2, 135.2, 130.0, 129.5, 126.6, 125.2, 123.9, 113.5, 112.6, 103.5, 85.6, 82.3, 54.9, 17.4 ppm. HRMS (APCI): calc'd for MH^+ = $C_{30}H_{25}O_{11}$: 561.1391; found = 561.1392; calc'd for MNa^+ = $C_{30}H_{24}O_{11}Na$: 583.1211; found = 583.1212.

Fluorescein bis(((1-methanesulfonyl)cyclopropyl)carboxymethyl ether) (28)

Data for **28**: (30%, white solid). 1H NMR ($CDCl_3$, 400 MHz): δ = 8.04 (d, 1H), 7.7-7.6 (m, 2H), 7.17 (d, 1H), 6.96 (m, 2H), 6.75 (m, 4H), 5.88 (s, 4H), 3.17 (s, 6H), 1.89-1.86 (m, 4H), 1.73-1.70 (m, 4H) ppm. ^{13}C NMR ($CDCl_3$, 100 MHz): δ = 169.1, 166.47, 158.0, 152.6, 152.2, 135.3, 130.1, 129.7, 126.6, 125.3, 123.9, 114.0, 112.6, 103.8, 86.2, 82.1, 42.9, 41.1, 16.3 ppm. HRMS (ESI): calc'd for MH^+ = $C_{32}H_{29}O_{13}S_2$: 685.1044; found: 685.1047; calc'd for MNa^+ = $C_{32}H_{28}O_{13}S_2Na$: 707.0864; found: 707.0867.

Fluorescein bis(cyclopropyl)carboxymethyl ether (**29**)

Data for **29**: (21%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.17 (d, 1H), 6.98 (m, 2H), 6.74 (d, 4H), 5.78 (s, 4H), 1.70-1.63 (m, 2H), 1.08-1.06 (m, 4H), 0.96-0.93 (m, 4H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 174.0, 169.5, 158.7, 153.2, 152.5, 135.4, 130.1, 129.6, 126.9, 125.4, 124.1, 113.5, 113.0, 103.8, 85.1, 82.7, 13.1, 9.5 ppm. HRMS (ESI): calc'd for MH⁺ = C₃₀H₂₅O₉: 529.1493; found: 529.1494.

Fluorescein bis(((2,2-dimethyl)cyclopropyl)carboxymethyl ether) (**30**)

Data for **30**: (73%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.03 (d, 1H), 7.7-7.6 (m, 2H), 7.17 (d, 1H), 6.98 (m, 2H), 6.74 (m, 4H), 5.80-5.75 (dd, 4H), 1.56-1.53 (m, 2H), 1.21 (s, 6H), 1.17 (s, 6H), 1.17-1.15 (m, 2H), 0.95-0.92 (m, 2H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 171.5, 169.1, 158.3, 152.8, 152.1, 134.9, 129.7, 129.2, 126.5, 125.0, 123.7, 113.0, 112.9, 112.7, 112.6, 103.4, 103.3, 84.6, 82.4, 26.7, 26.4, 24.3, 22.8, 18.6 ppm. HRMS (ESI): calc'd for MH⁺ = C₃₄H₃₃O₉: 585.2119; found: 585.2121.

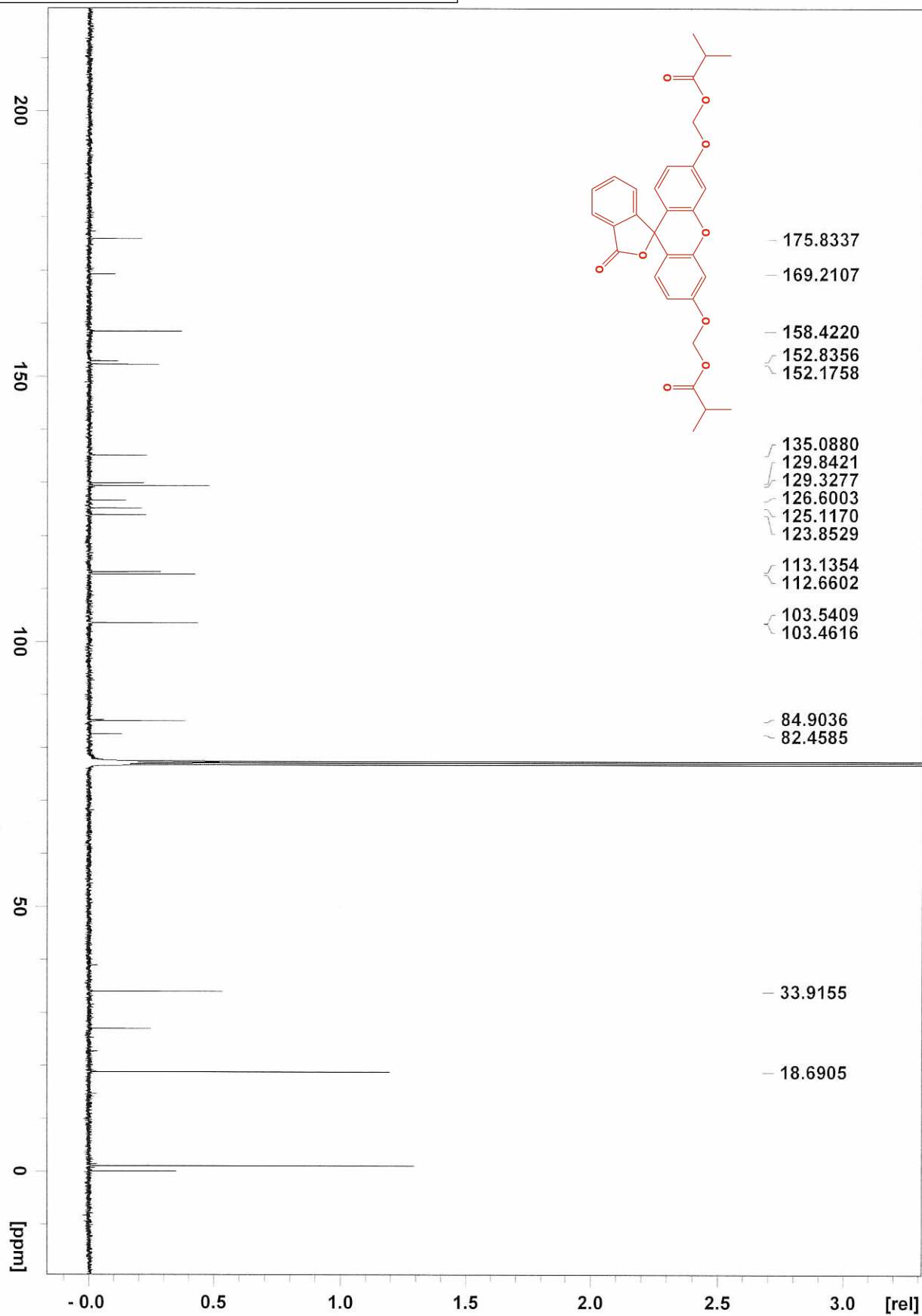
Fluorescein bis(((1-trifluoromethyl)cyclopropyl)carboxymethyl ether) (**33**)

Data for **33**: (39%, white solid). ¹H NMR (CDCl₃, 400 MHz): δ = 8.05 (d, 1H), 7.7-7.6 (m, 2H), 7.18 (d, 1H), 6.98 (m, 2H), 6.75 (s, 4H), 5.84 (m, 4H), 1.50 (s, 4H), 1.42 (m, 4H) ppm. ¹³C NMR (CDCl₃, 100 MHz): δ = 169.1, 167.4, 158.2, 152.7, 152.1, 135.1, 129.8, 129.3, 126.5, 125.4, 125.1, 123.8, 122.7, 113.6, 112.6, 103.8, 85.8, 82.2, 27.0-26.0 (q), 13.80/13.79 ppm. HRMS (ESI): calc'd for MH⁺ = C₃₂H₂₃F₆O₉: 665.1241; Found: 665.1245.

References for Supplemental Methods:

1. Tian, Lin; Yang, Yunlei; Wysocki, Laura M.; Arnold, Alma C; Hu, Amy; Ravichandran, Balaji; Sternson, Scott M.; Looger, Loren L. & Lavis, Luke D.; Selective esterase-ester pair for targeting small molecules with cellular specificity. *Proc. Natl. Acad. Sci. U. S. A.* **109** 4756–4761 (2012)
2. Ellis, E. E.; Adkins, C. T.; Galovska, N. M.; Lavis, L. D. & Johnson, R. J.; Decoupled roles for the atypical, bifurcated binding pocket of the ybfF hydrolase. *ChemBioChem* **14**, 1134-1144 (2013).
3. Hedge, Matthew K; Gehring, Alexandra M; Adkins, Chinessa T; Weston, Leigh A.; Lavis, Luke D. & Johnson, R. Jeremy; The structural basis for the narrow substrate specificity of an acetyl esterase from *Thermotoga maritima*. *Biochim. Biophys. Acta* **1824** 1024–1030 (2012)
4. Bassett, Braden; Waibel, Brent; White, Alex; Hansen, Heather; Stephens, Dominique; Koelper, Andrew; Larsen, Erik M.; Kim, Charles; Glanzer, Adam; Lavis, Luke D.; Hoops, Geoffrey C. & Johnson, R. Jeremy; Measuring the global substrate specificity of Mycobacterial serine hydrolases using a library of fluorogenic ester substrates. *ACS Infect. Dis.* **4** 904-911 (2018)

Figure S2



"MAM Fluorescein and isobutyric acid C1 P1" 10 1 C:\Bruker\TopSpin3.2\data\routine\nmr

Figure S3

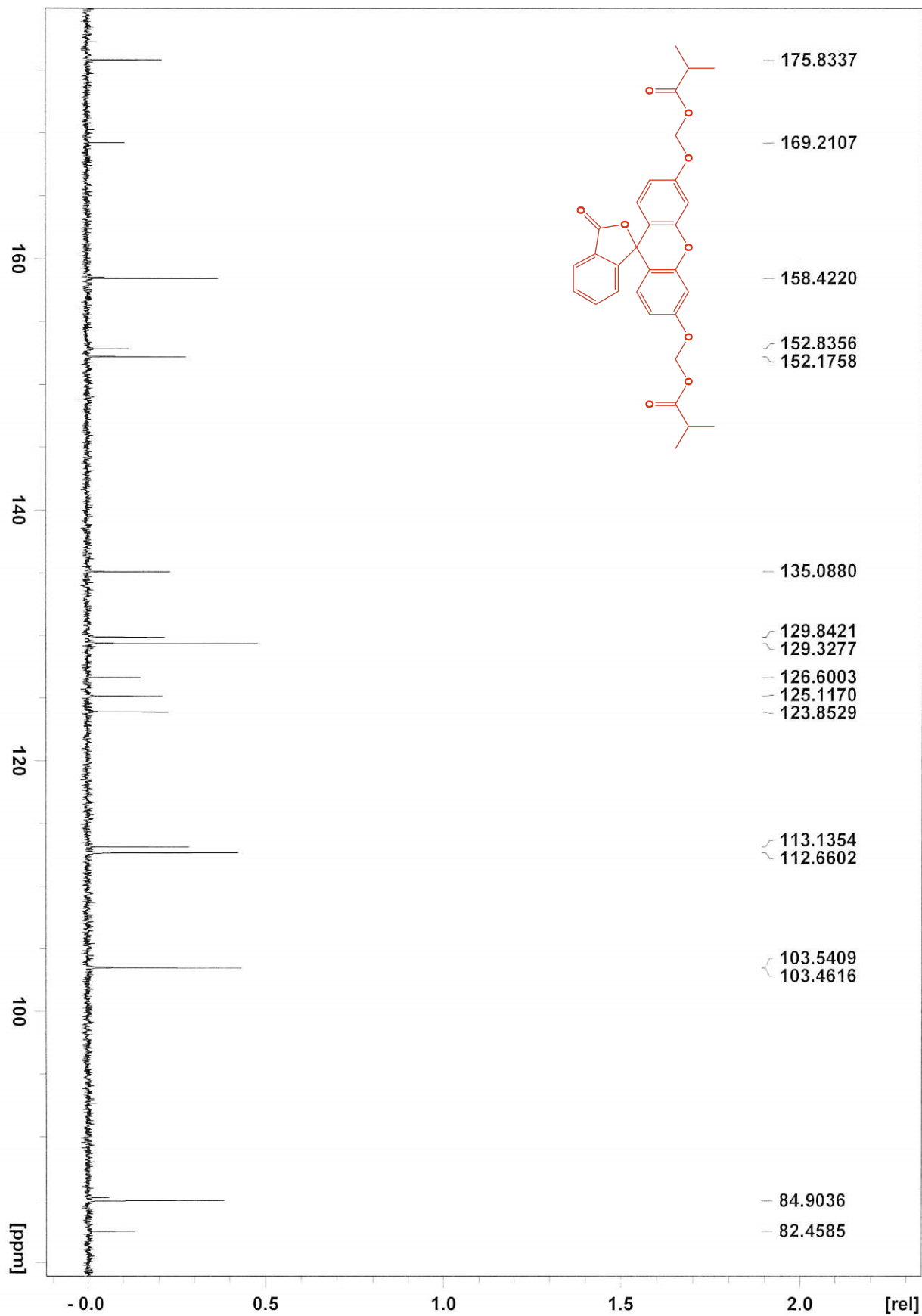


Figure S4

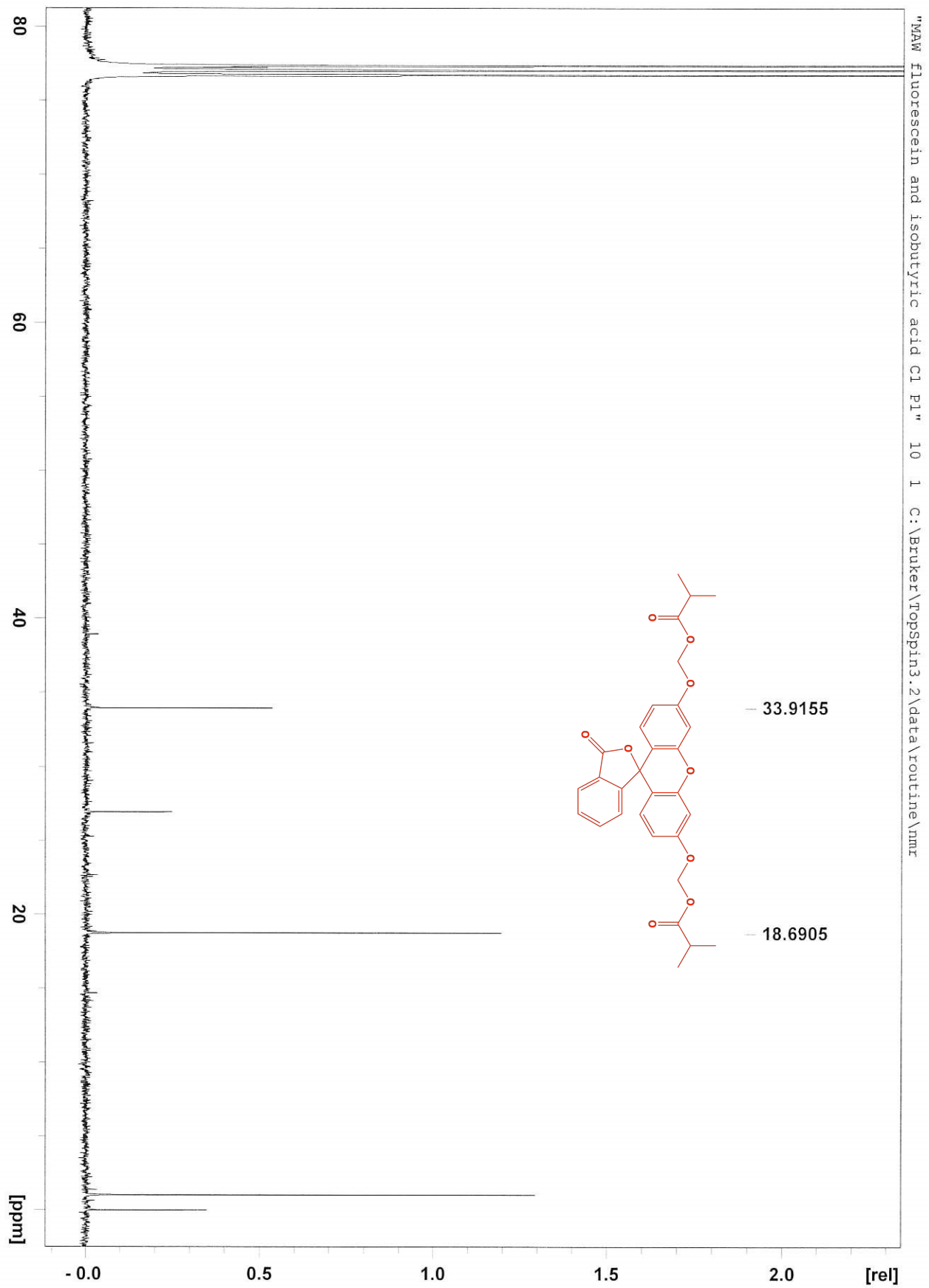


Figure S5

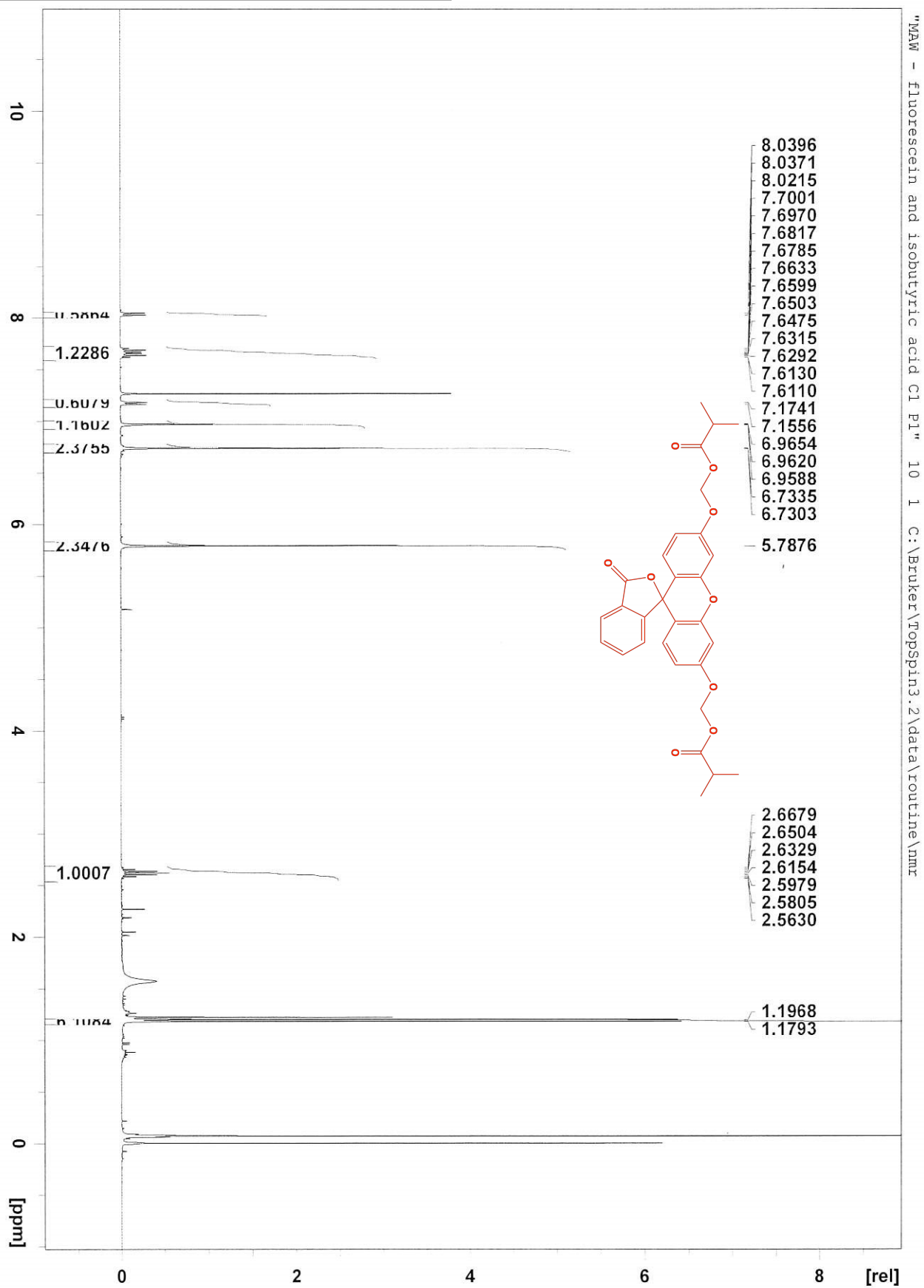


Figure S6

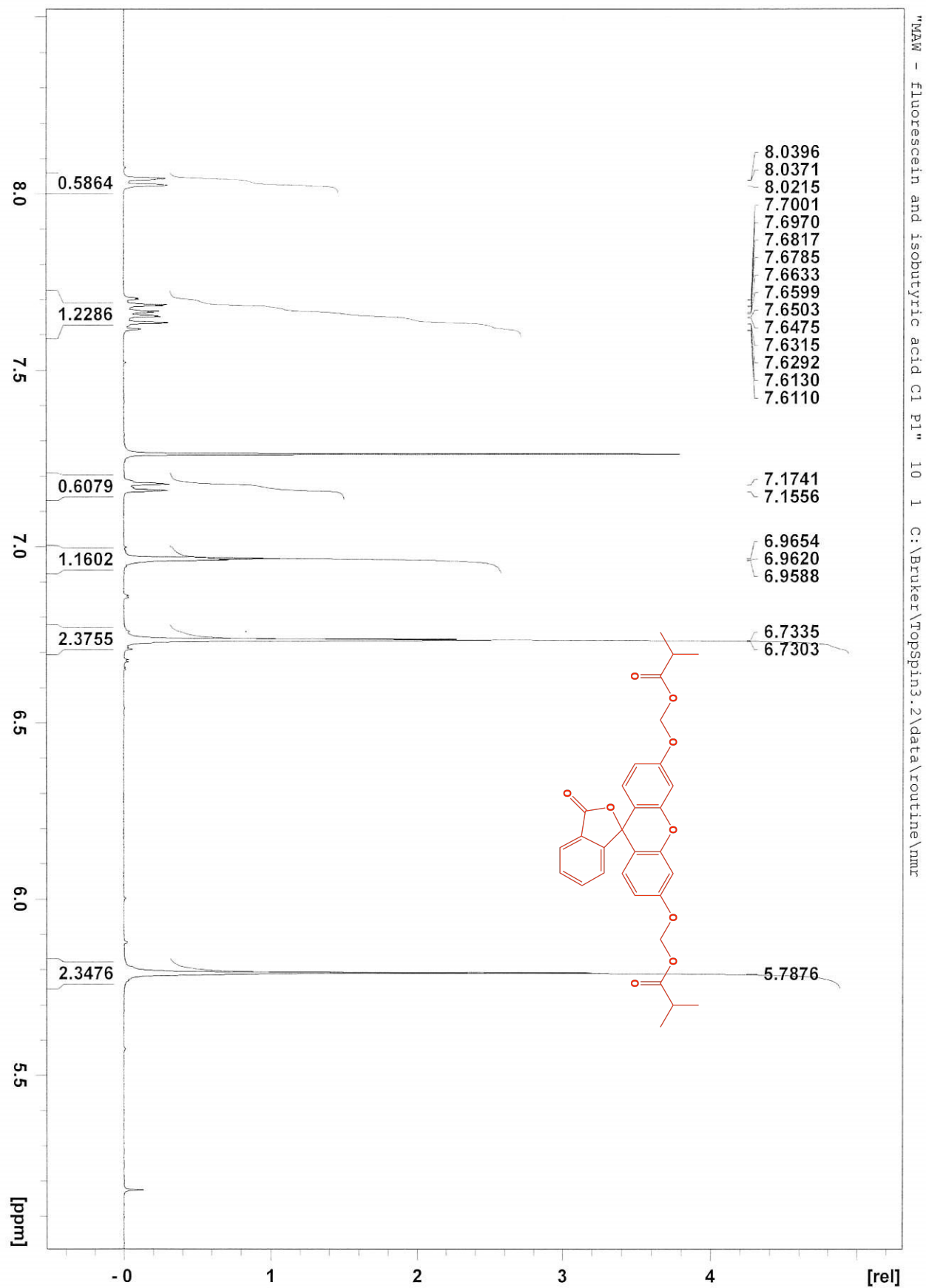


Figure S7

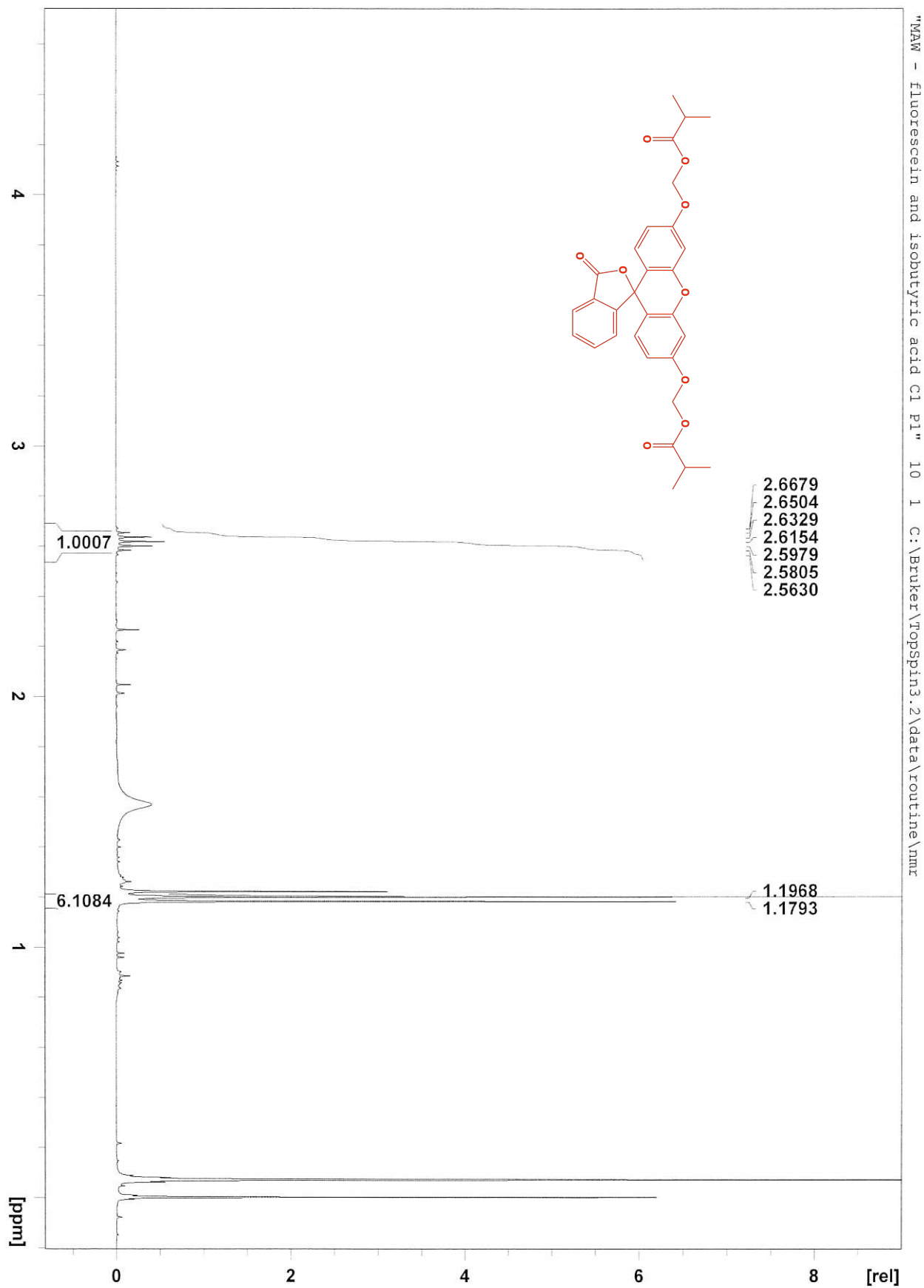
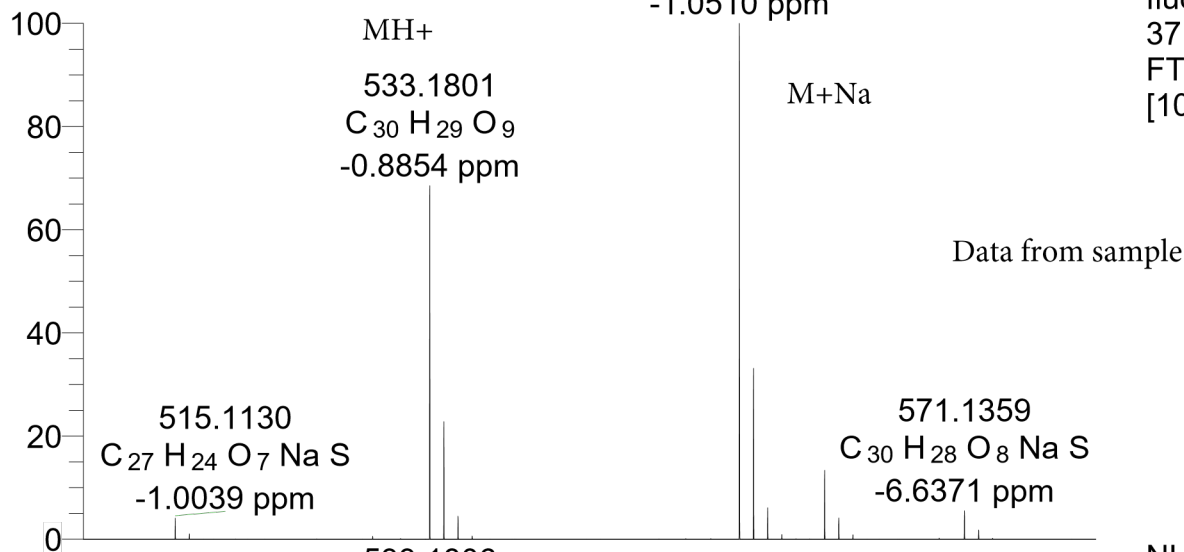
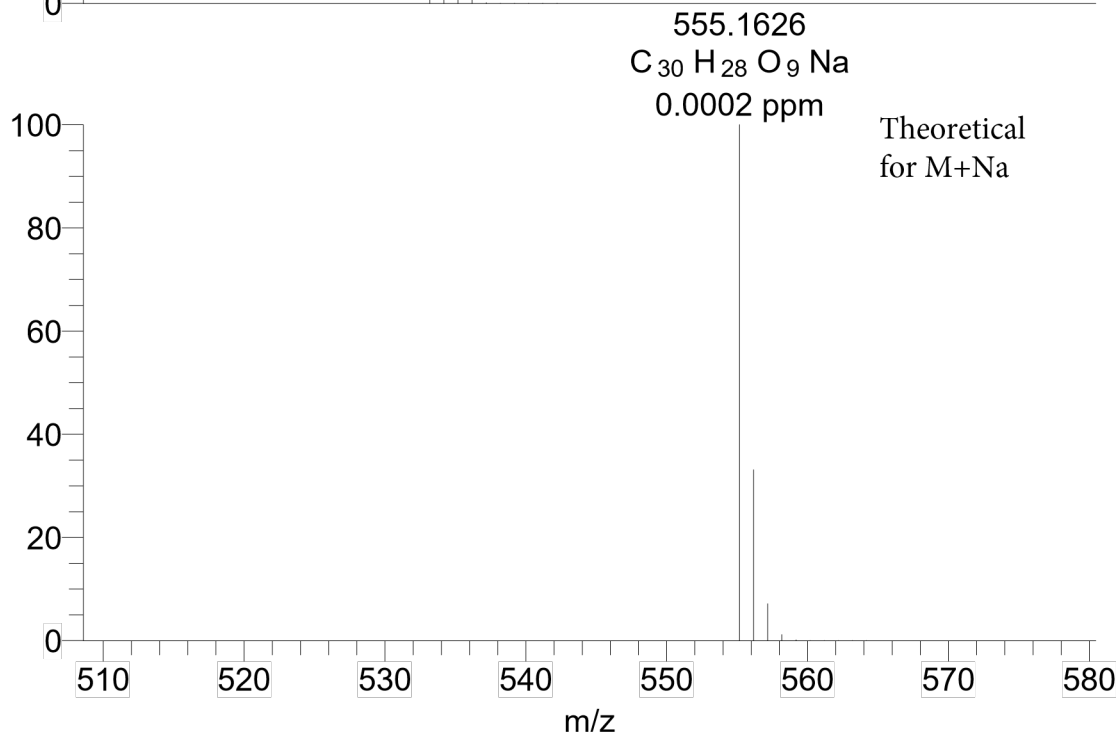
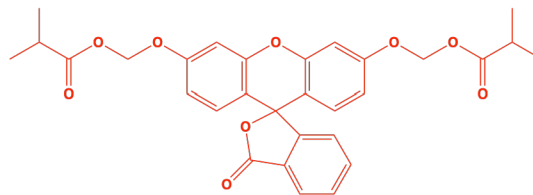
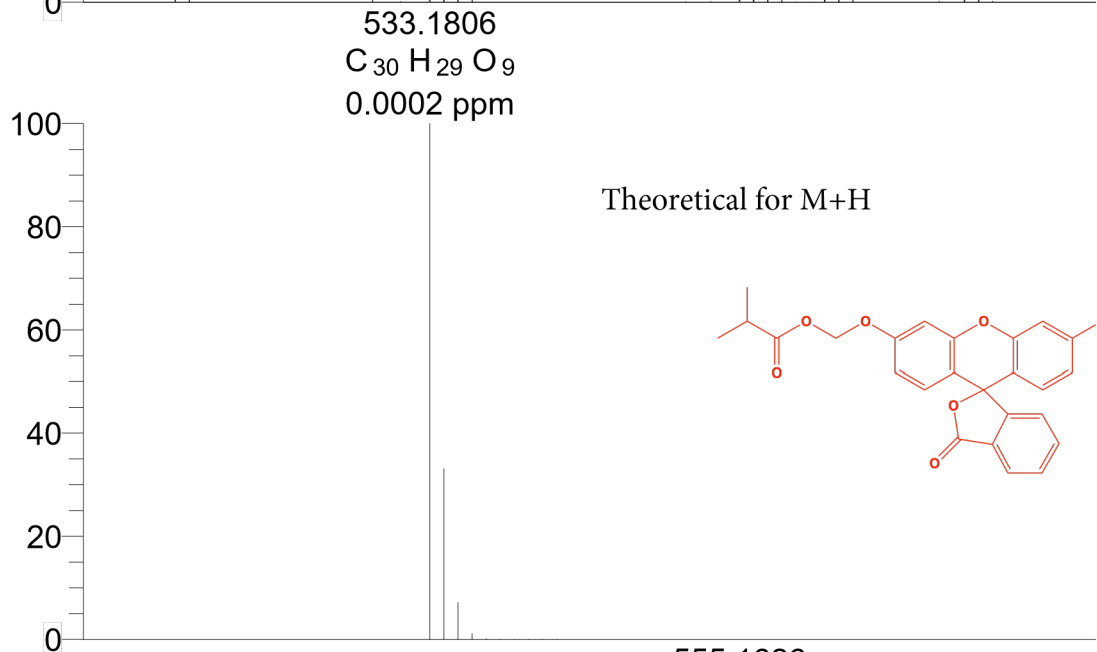


Figure S8

fluor-ester-isobutyric-acid#32-37 RT: 0.25-0.30 AV: 6 T: FTMS + p ESI Full ms [100.00-2000.00]

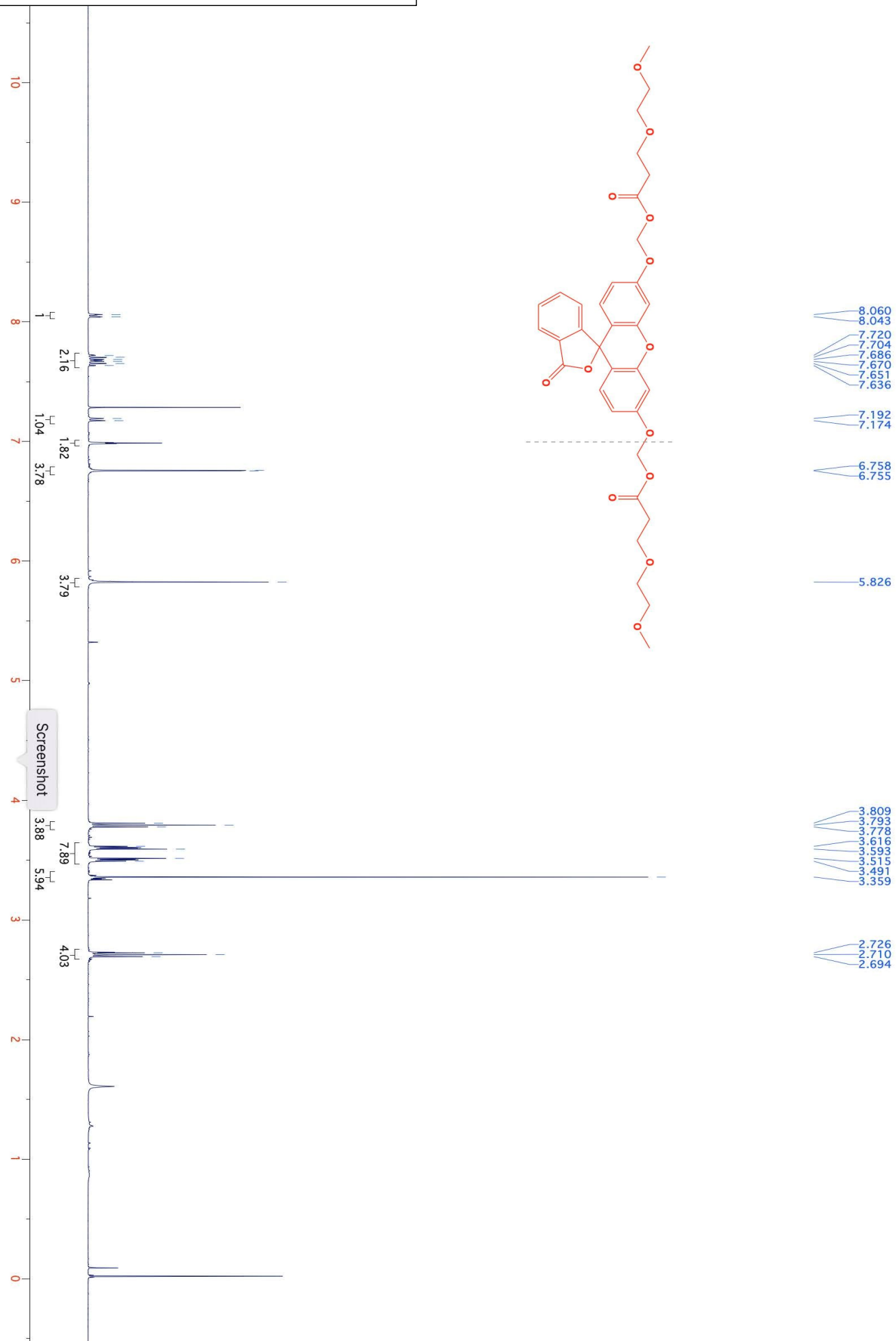


NL: 7.06E5
 C₃₀H₂₈O₉H:
 C₃₀H₂₉O₉
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res .Pwr . @FWHM



NL: 7.06E5
 C₃₀H₂₈O₉Na:
 C₃₀H₂₈O₉Na₁
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res .Pwr . @FWHM

Figure S9



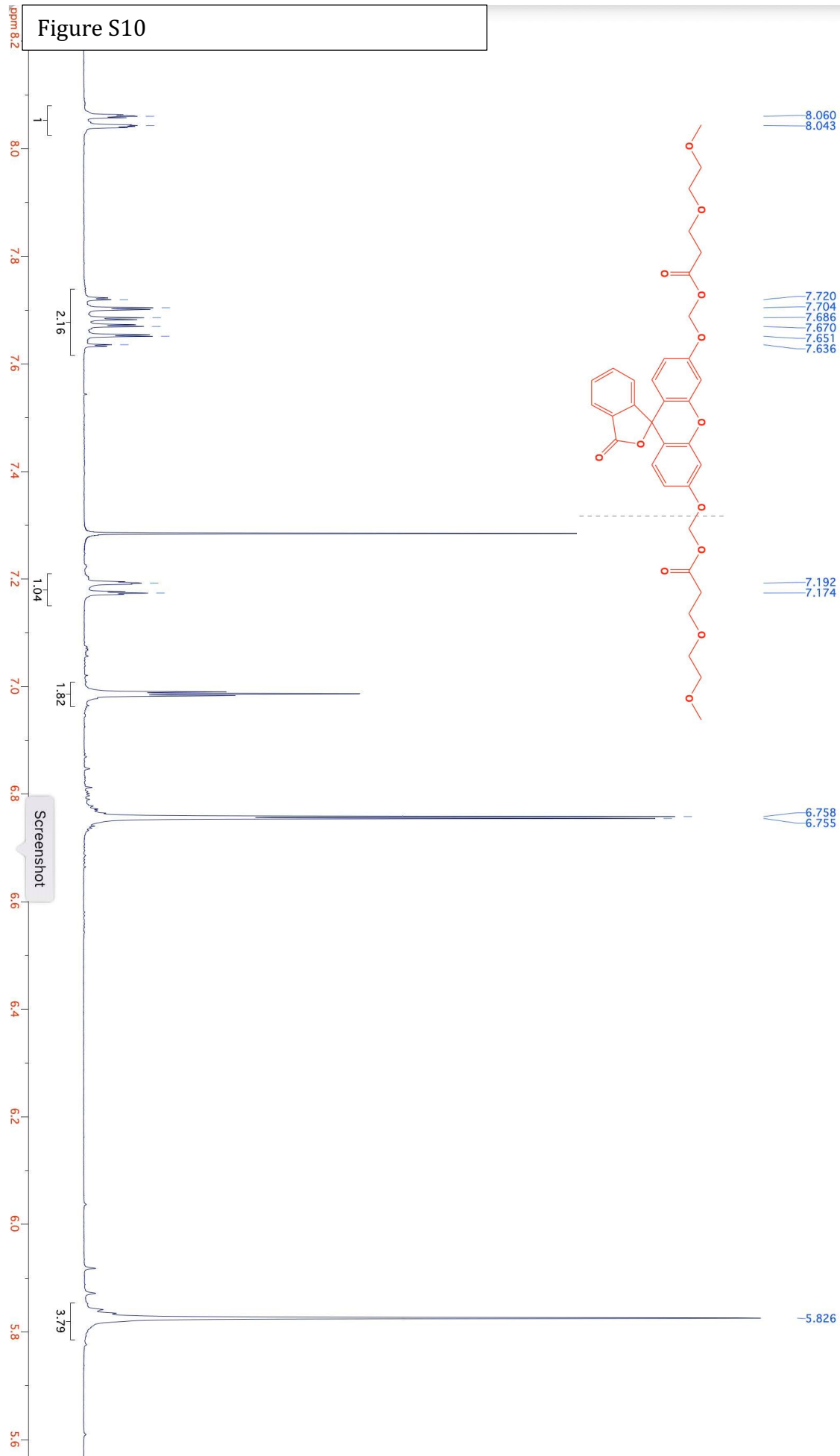


Figure S11

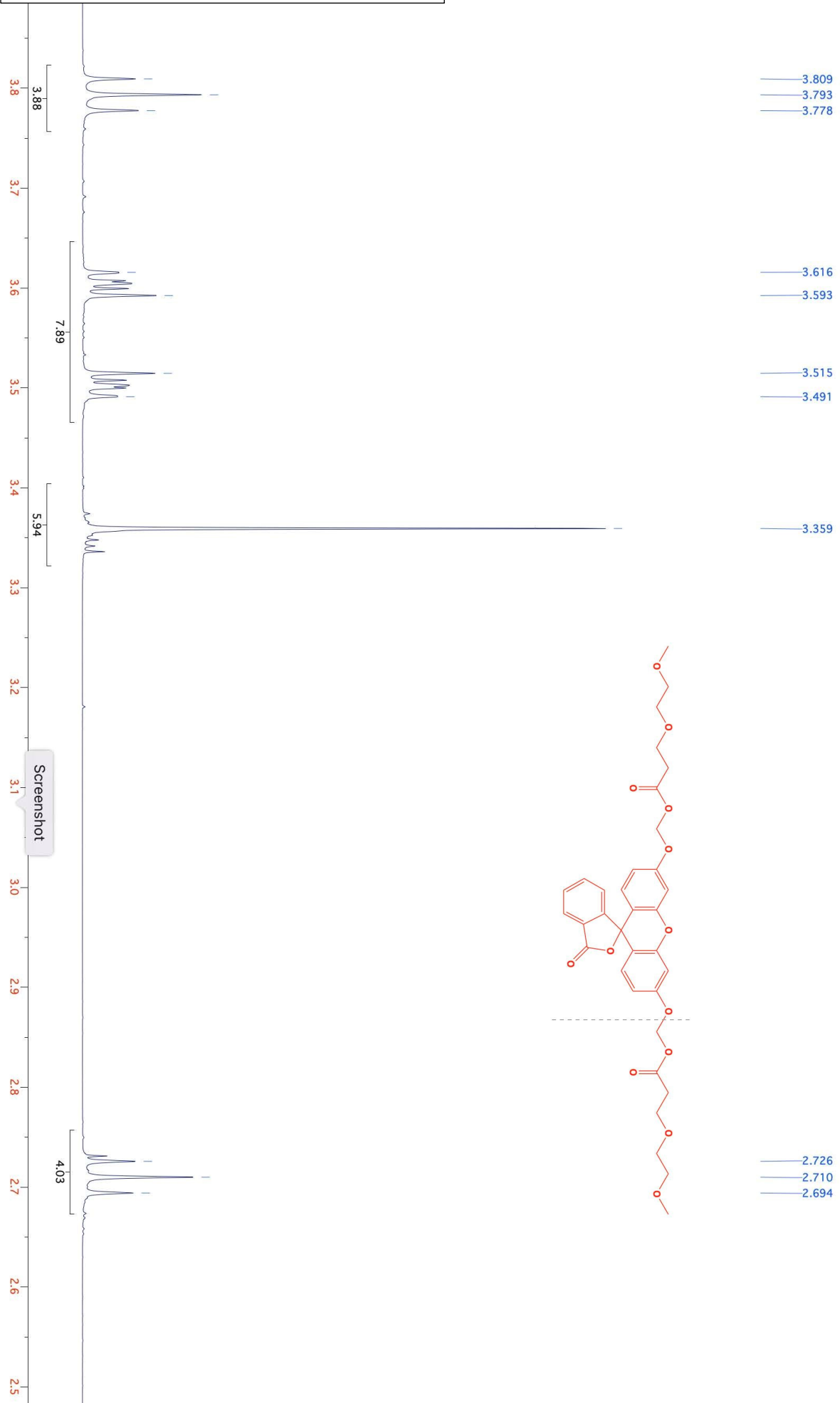
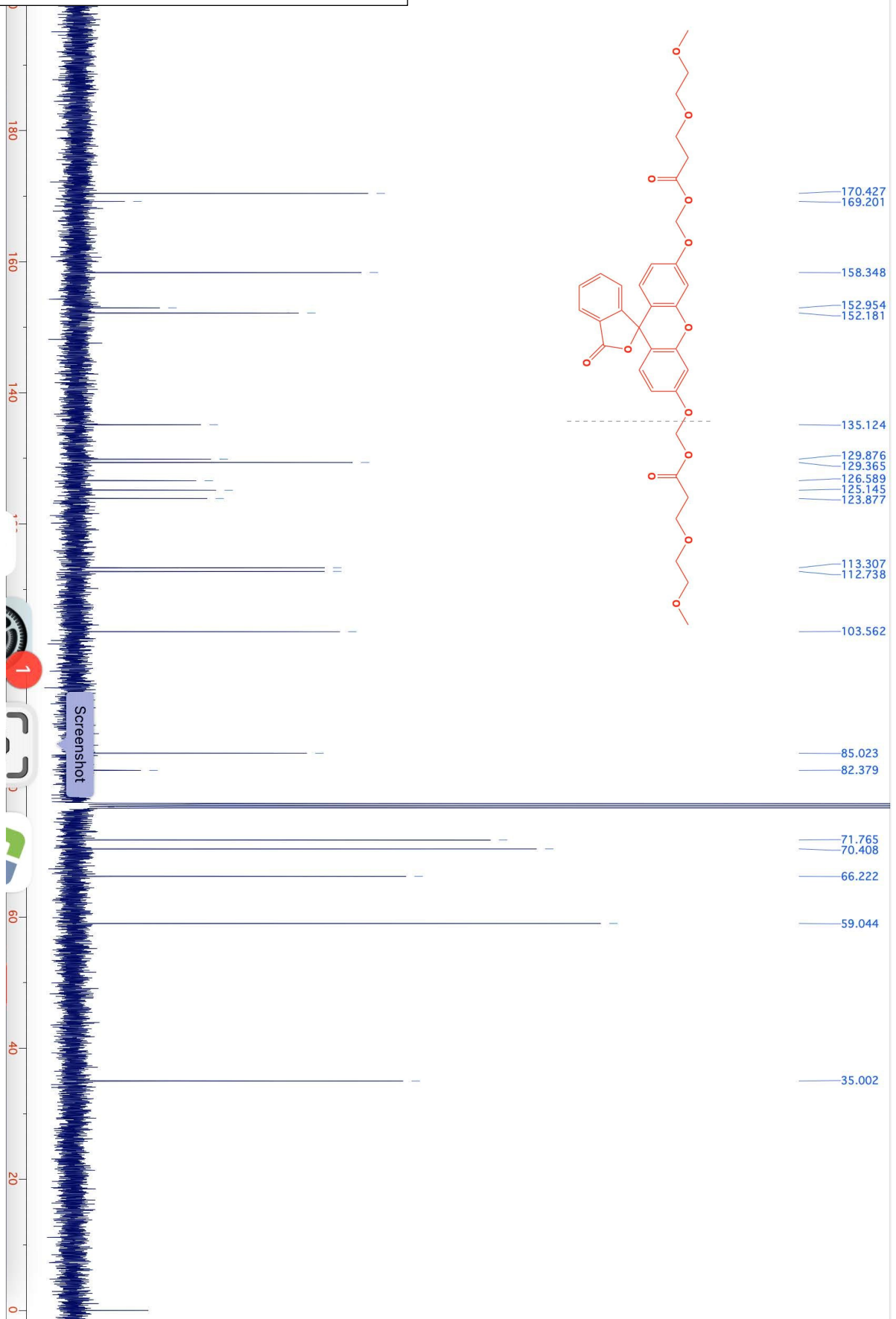


Figure S12



butler-071416-02

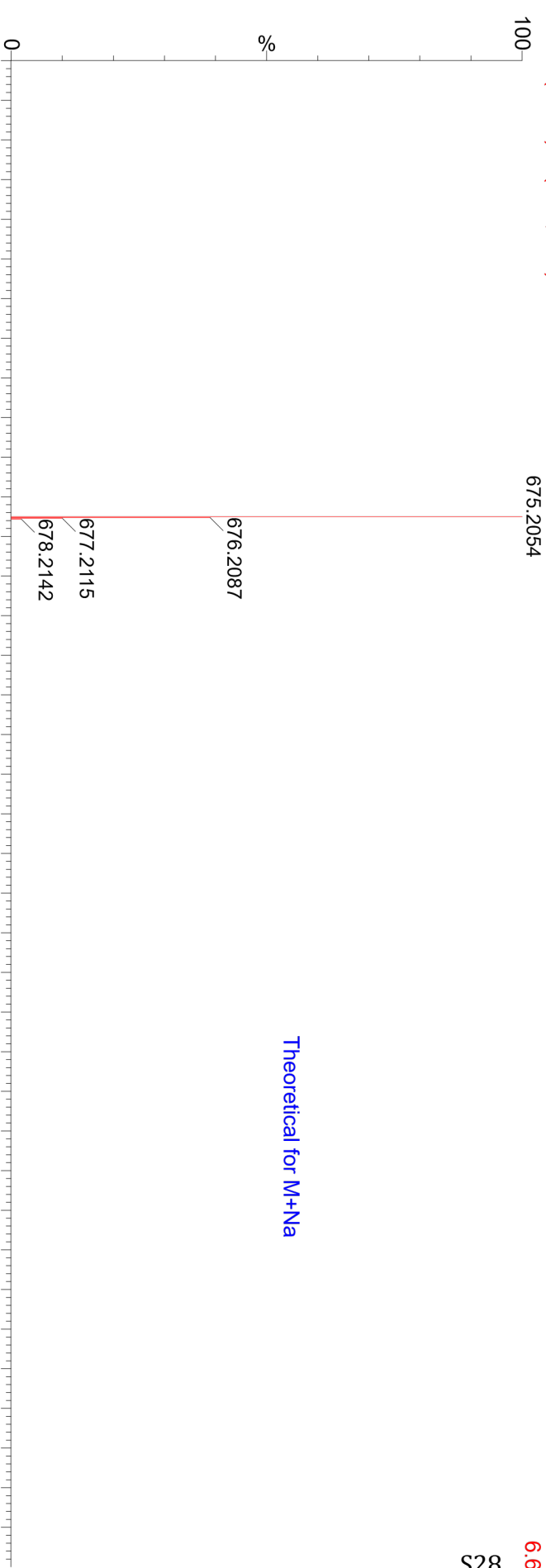
TPPD (0.070) Is (0.10,1.00) C34H36O13Na

8.000000000

1: TOF MS ES+

6.62e12

S28

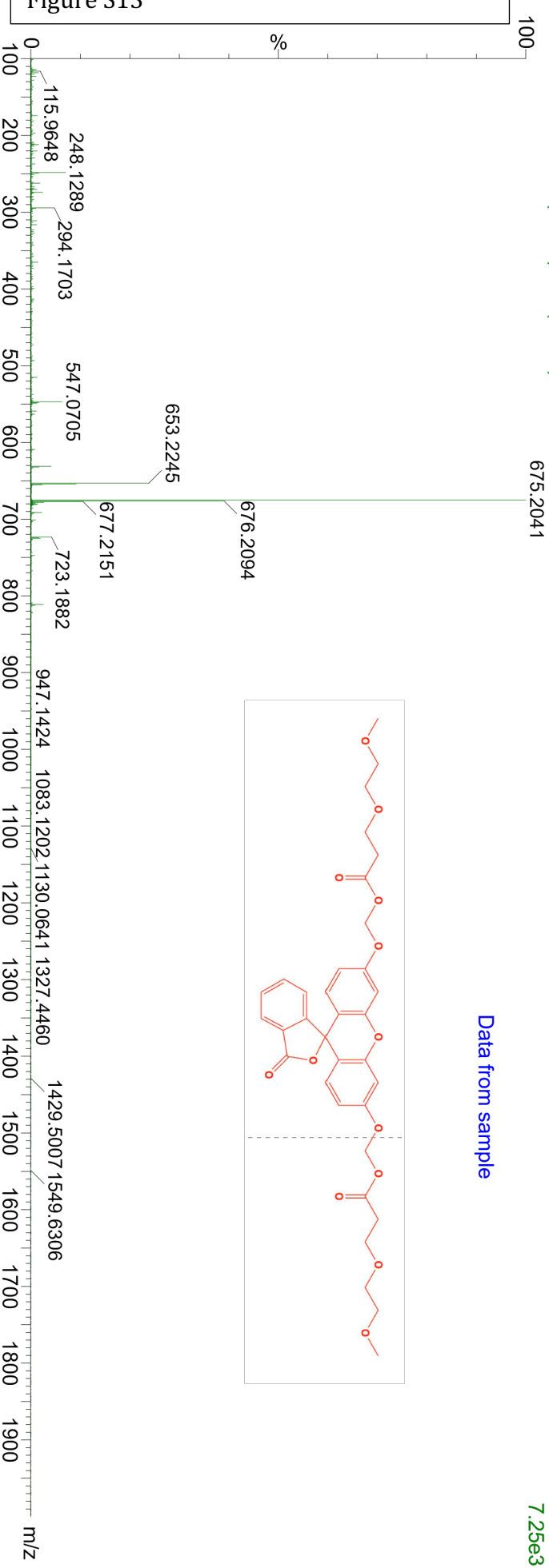


Theoretical for M+Na

butler-071416-02 33 (1.237) Cm (33.34)

1: TOF MS ES+

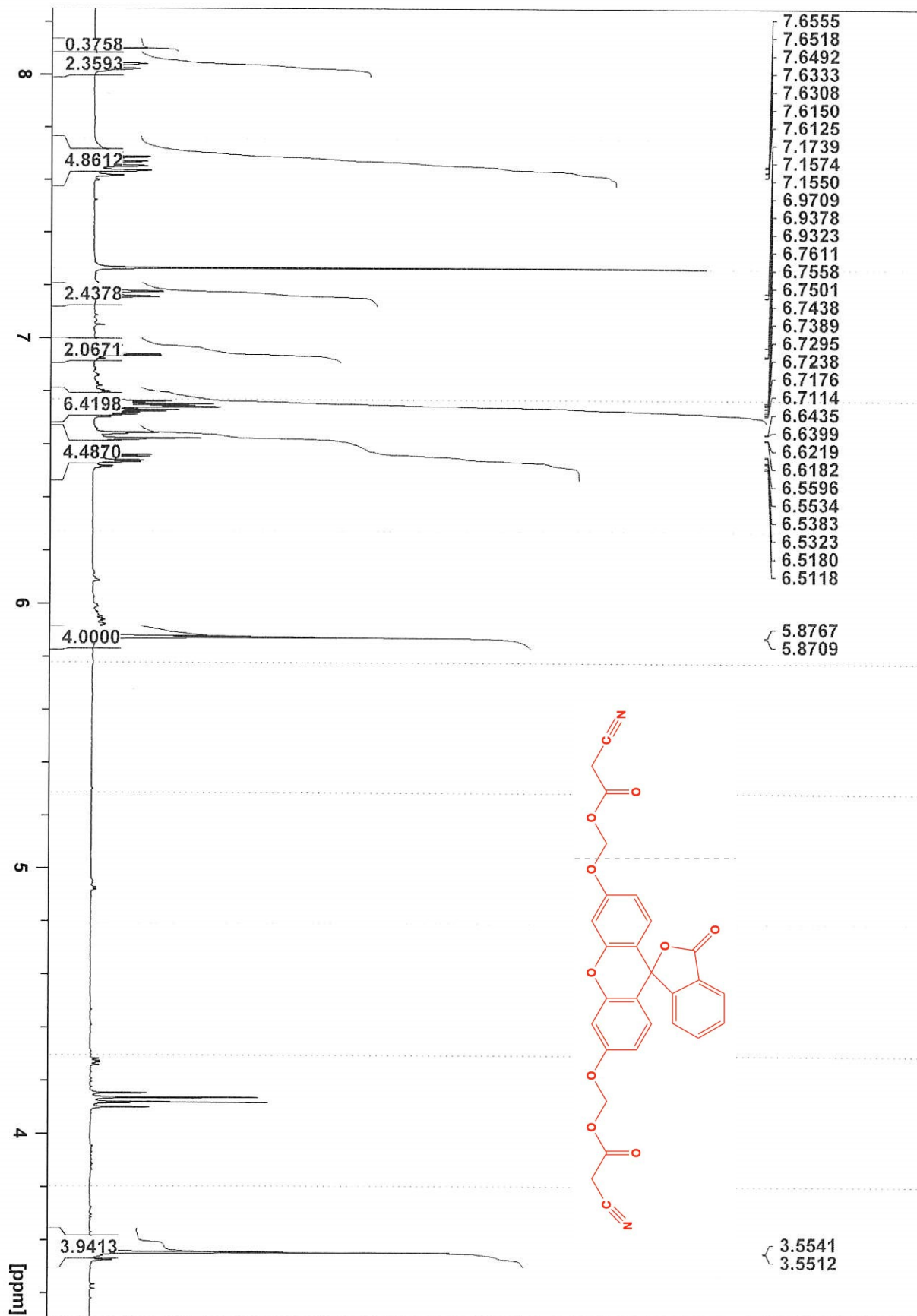
7.25e3



Data from sample

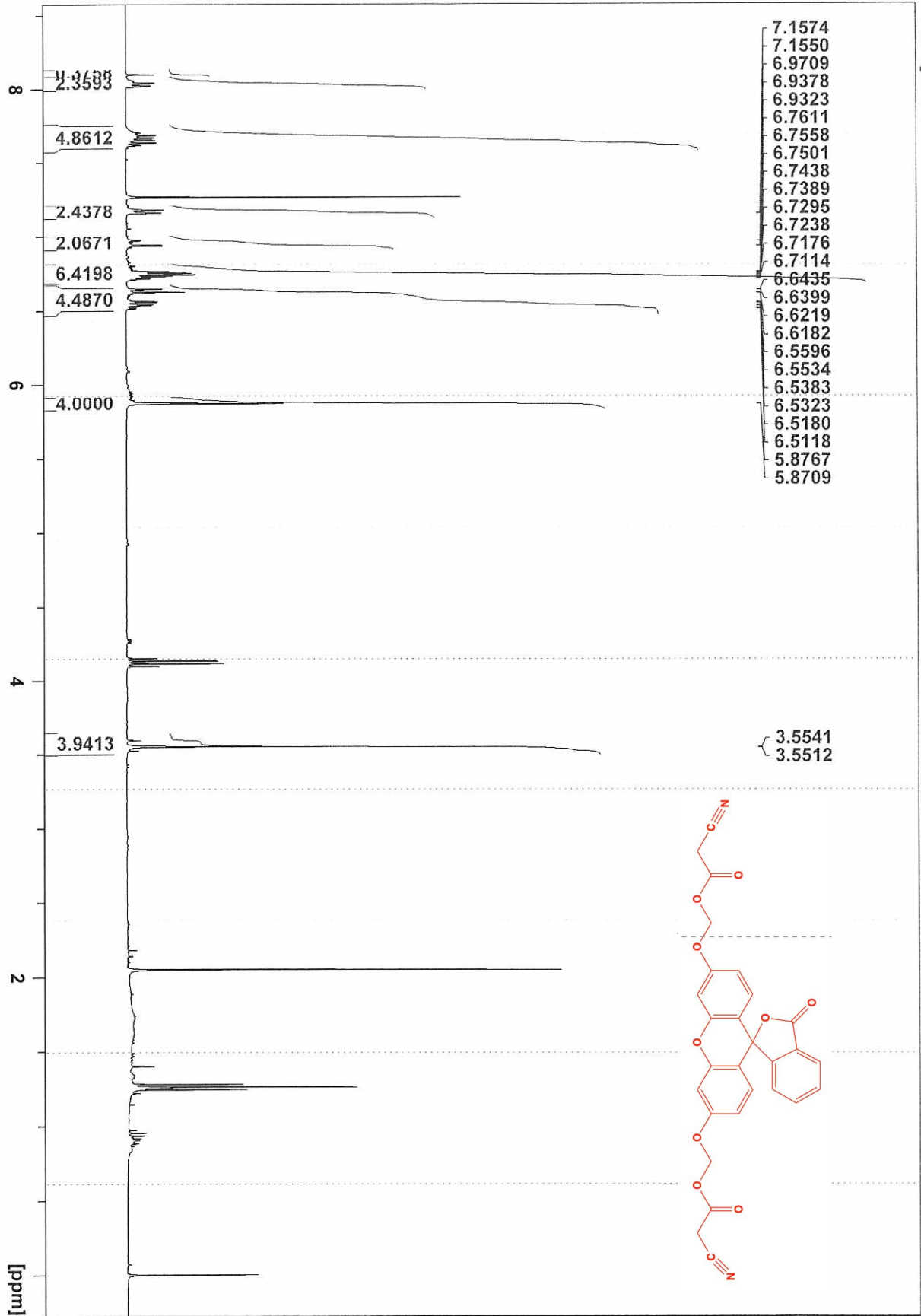
Figure S13

Figure S14



"JJE-cyanoacetic acid ester 28 min in cdcl3" 10 1 C:\Bruker\Topspin3.2\data\routine\nmr

Figure S15



"JJE-cyanocetic acid ester 28 min in cdcl3" 10 1 C:\Bruker\TopSpin3.2\data\routine\nmr

Figure S16

JJE-cyanoacetic acid ester 28 min in cdcl3

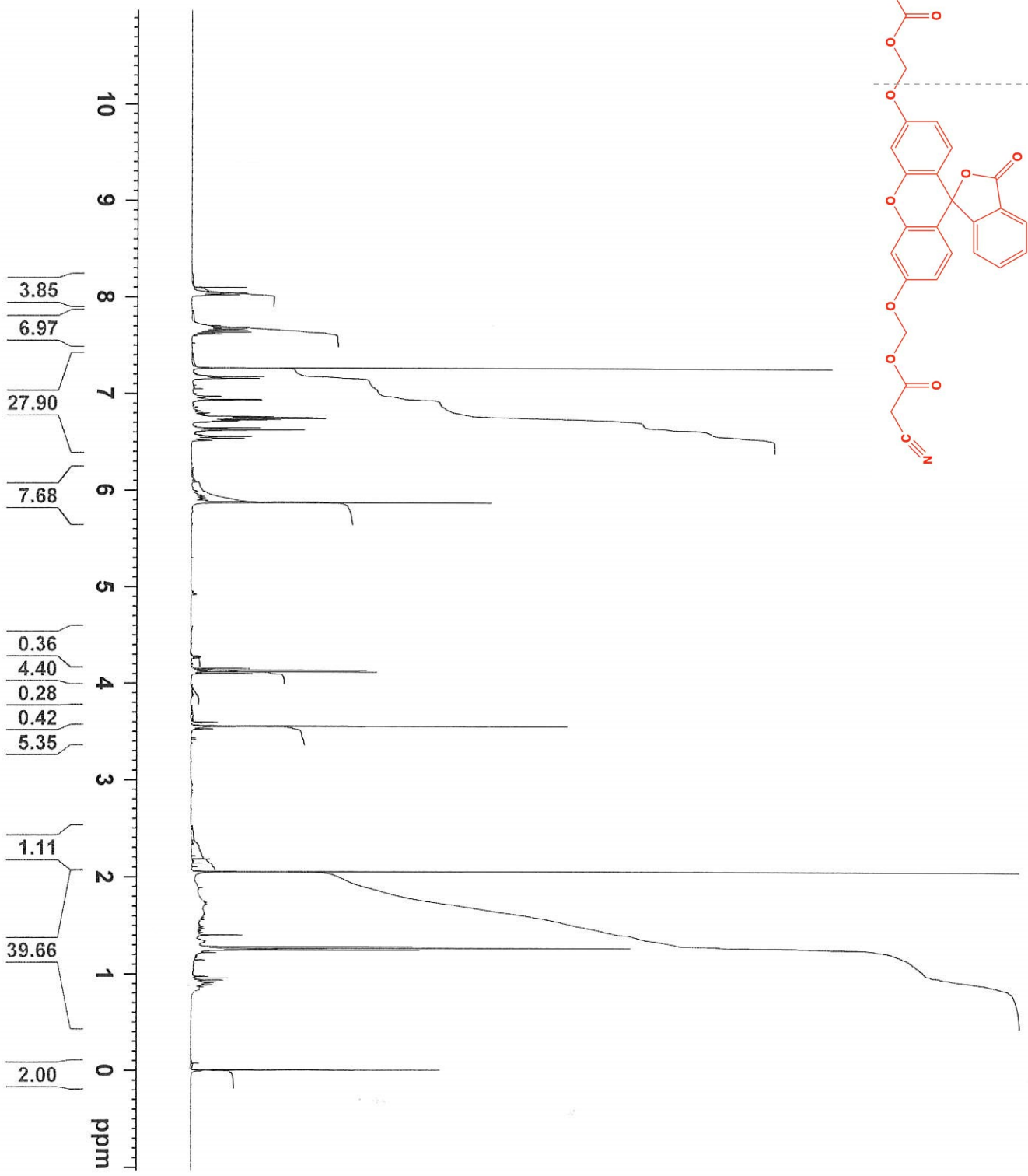
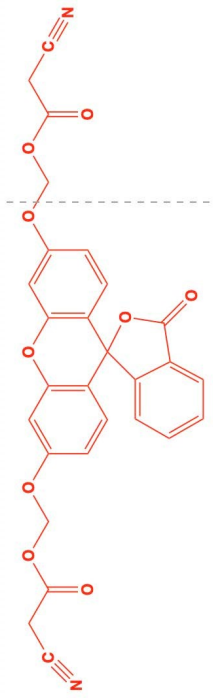
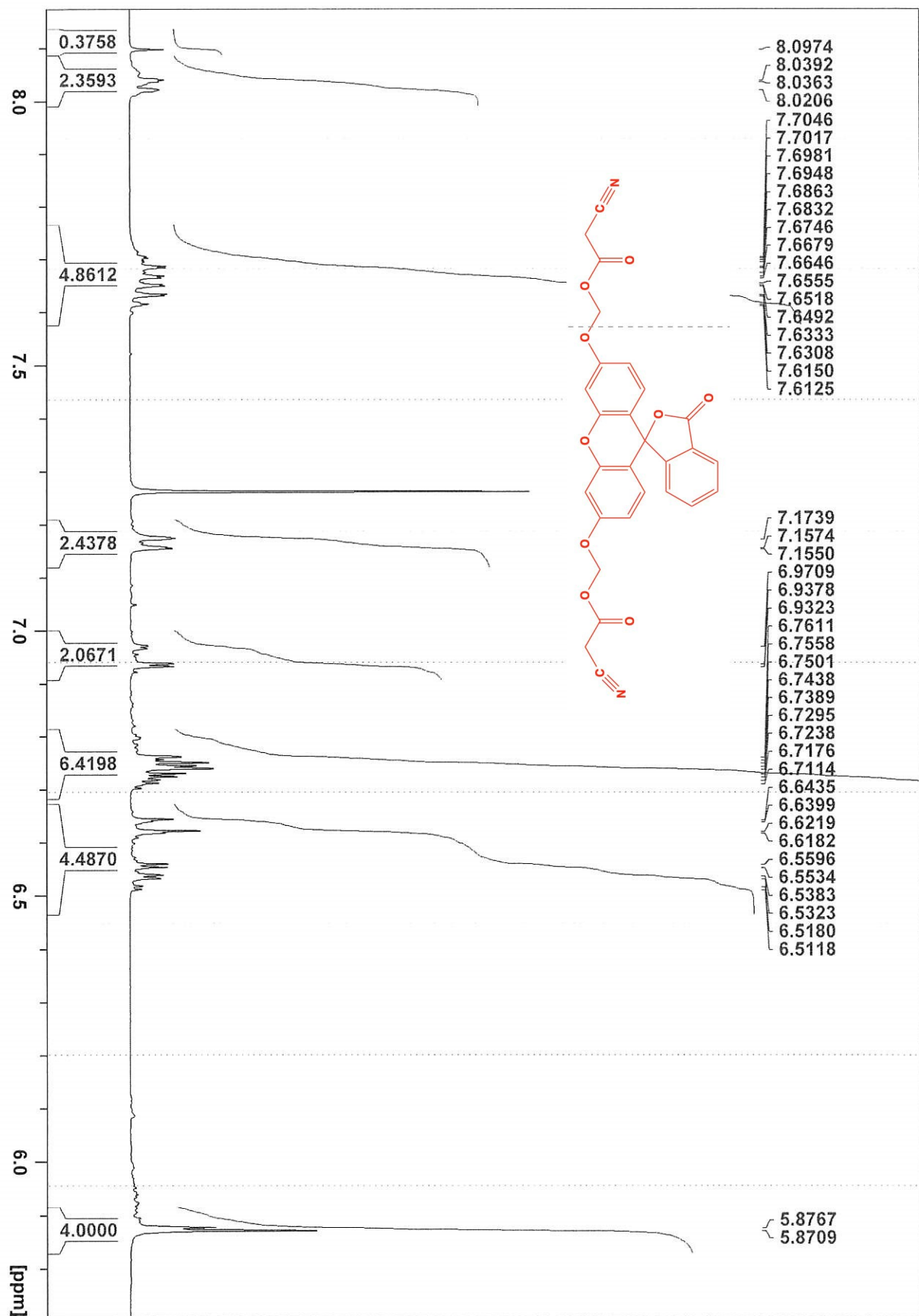
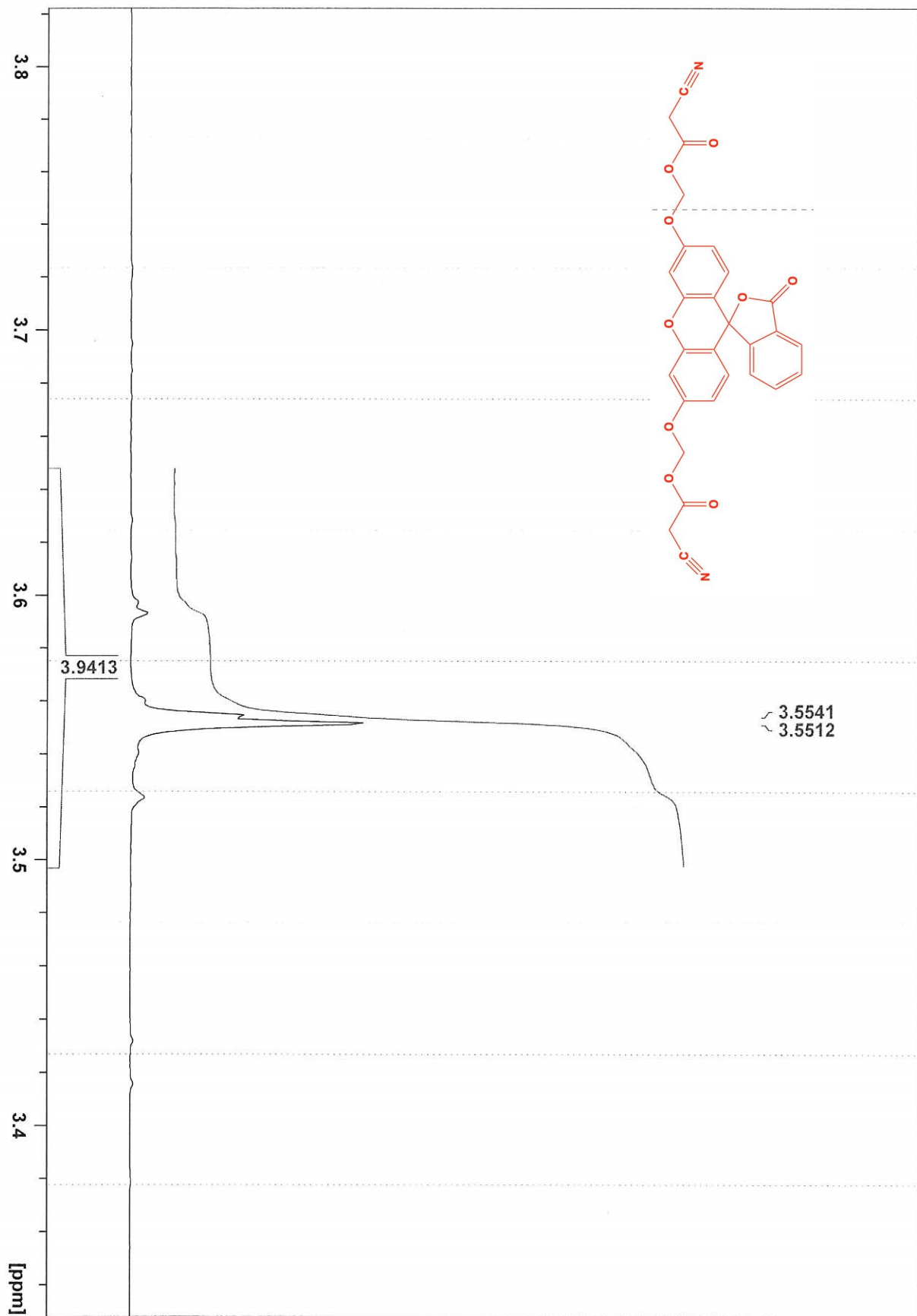


Figure S17



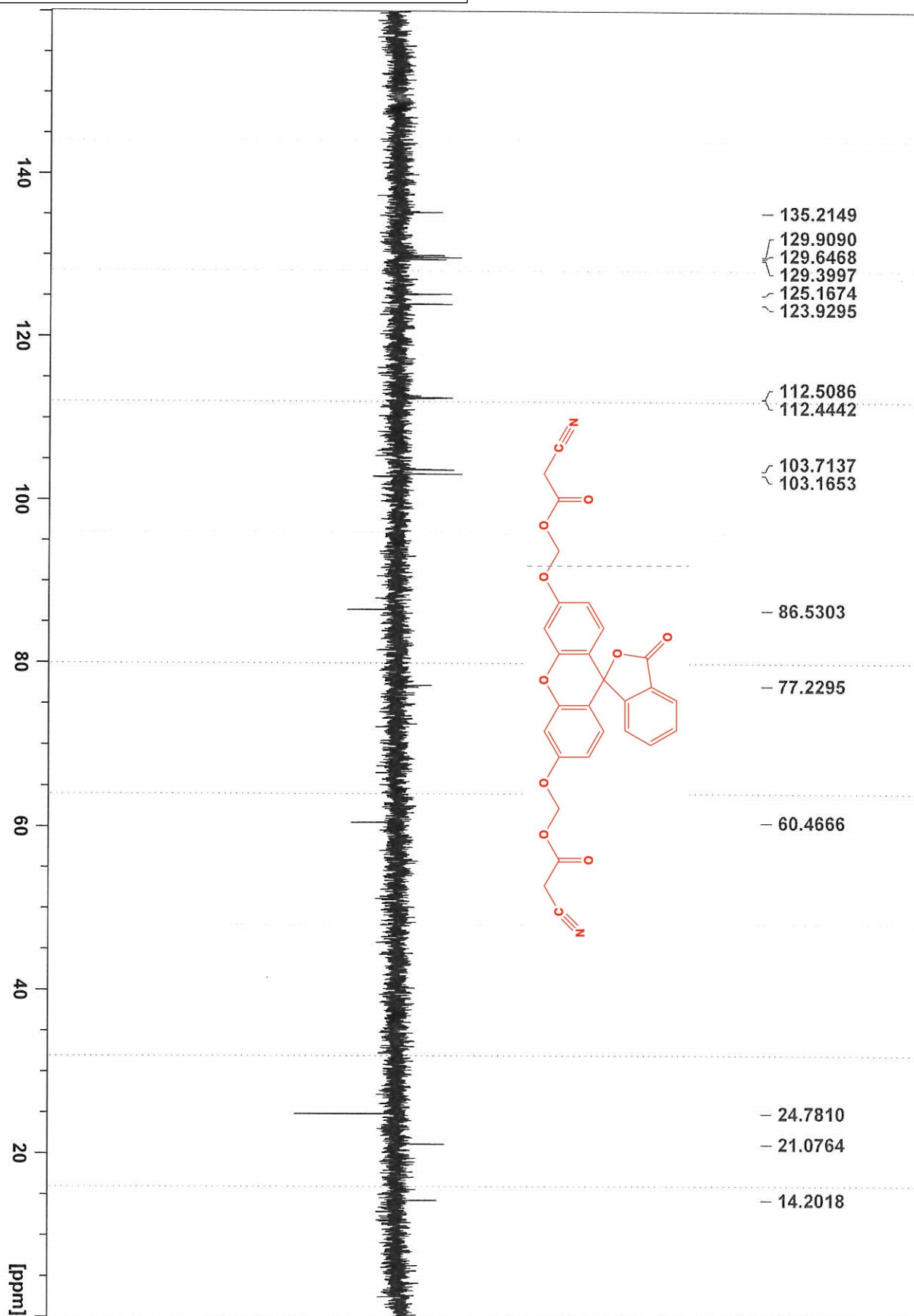
"JJE-cyanocetic acid ester 28 min in cdcl3" 10 1 C:\Bruker\Topspin3.2\data\routine\mmr

Figure S18



"JDE-cyanoacetic acid ester 28 min in cdcl3" 10 1 C:\Bruker\Topspin3.2\data\routine\nmr

Figure S19



"JJE-cyanoacetic acid ester 28 min in cdcl3" 11 1 C:\Bruker\TopSpin3.2\data\routine\mmr

Figure S20

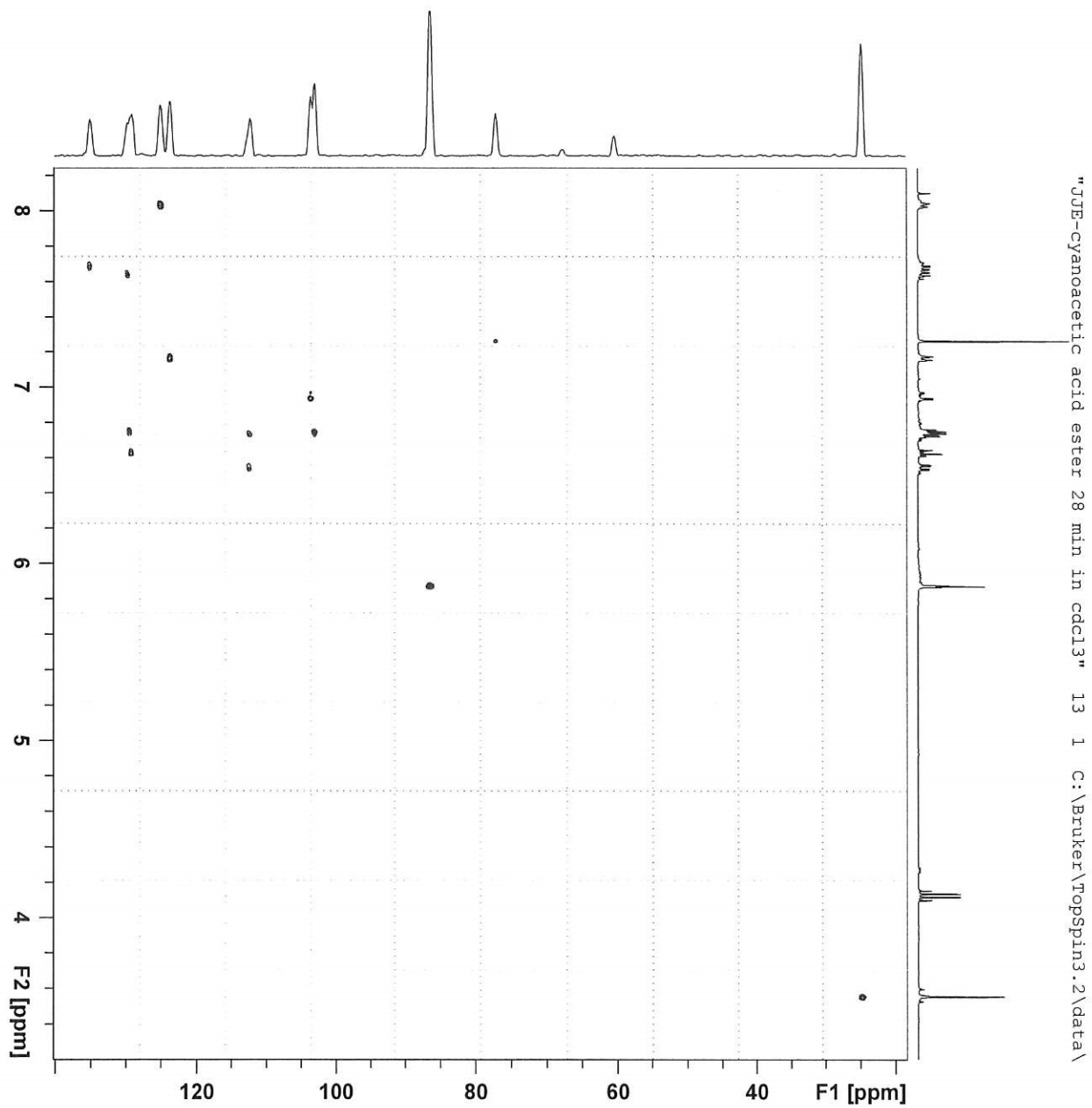
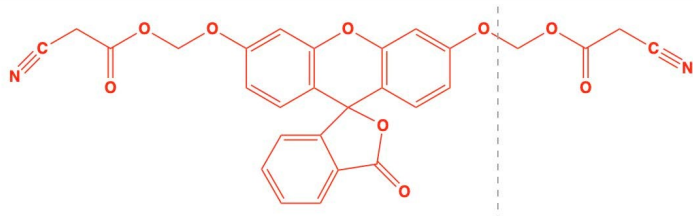
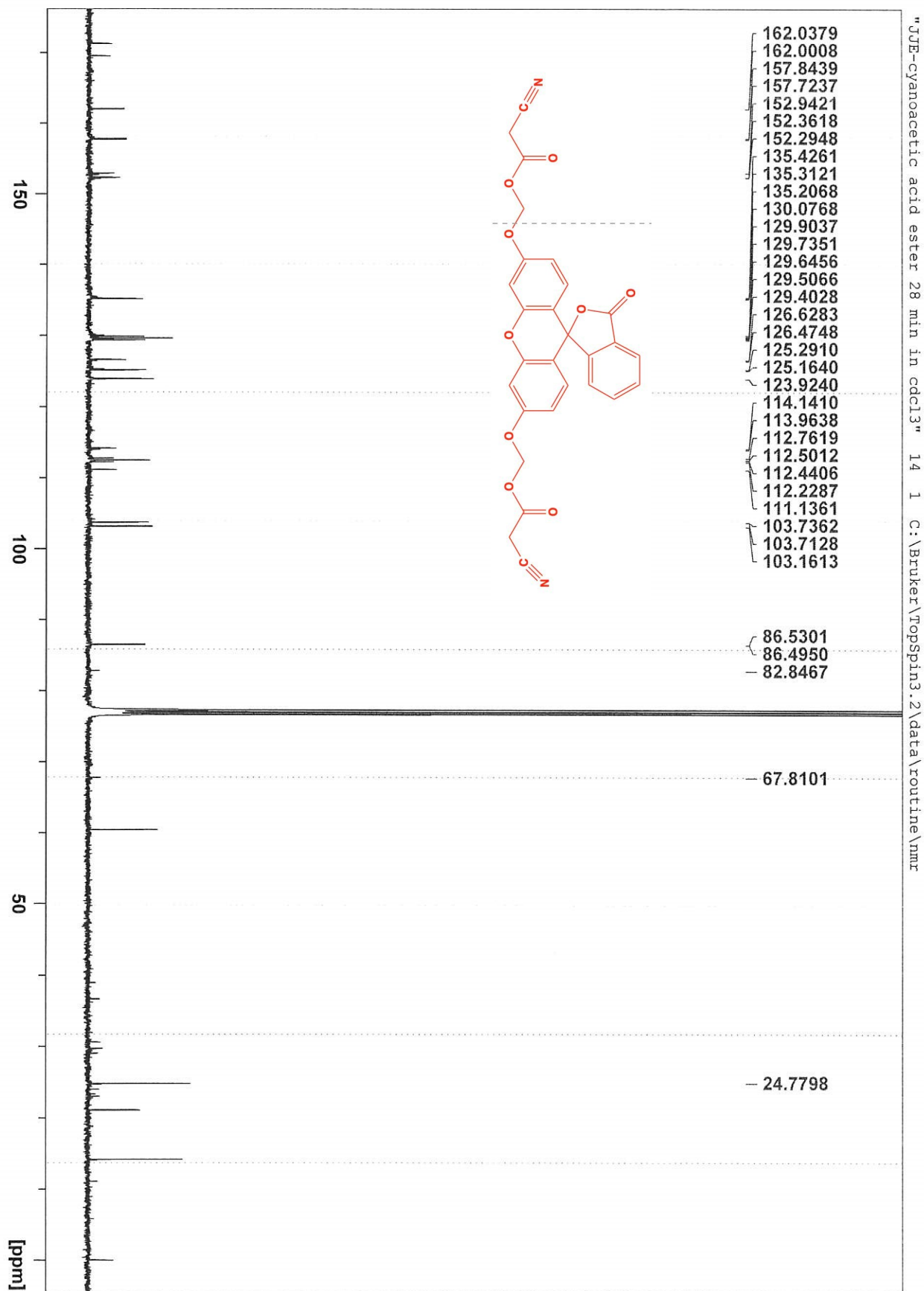
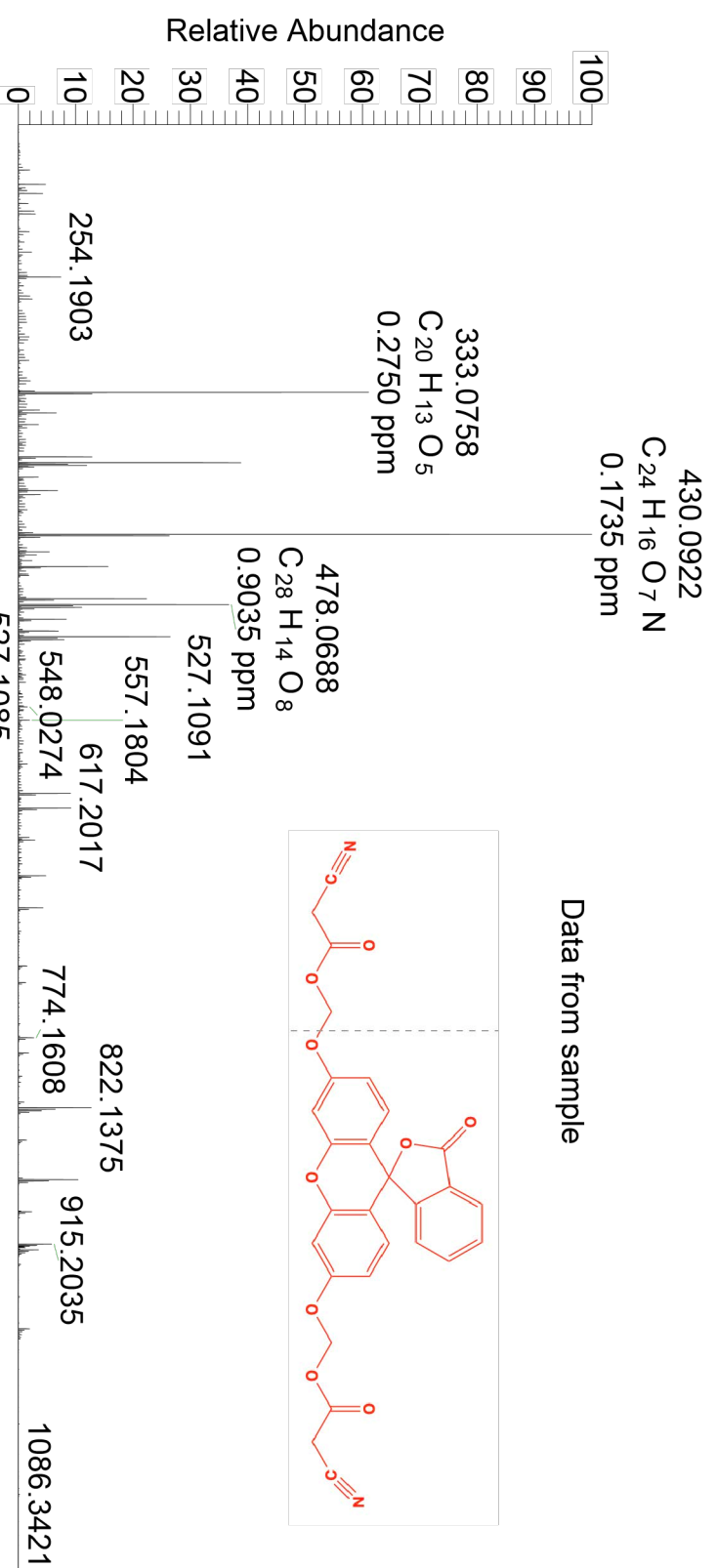


Figure S21



NL:
4.78E6
BU-4#26-59 RT: 0.81-0.47
AV: 34 T: FTMS + pESI Full
ms [150.00-2000.00]



Theoretical for MH⁺

NL:
7.17E5
C₂₈ H₁₈ N₂ O₉ H:
C₂₈ H₁₉ N₂ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Figure S22

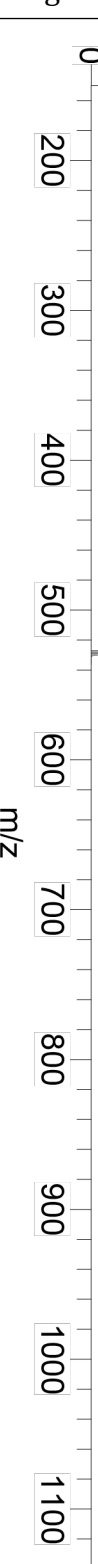


Figure S23

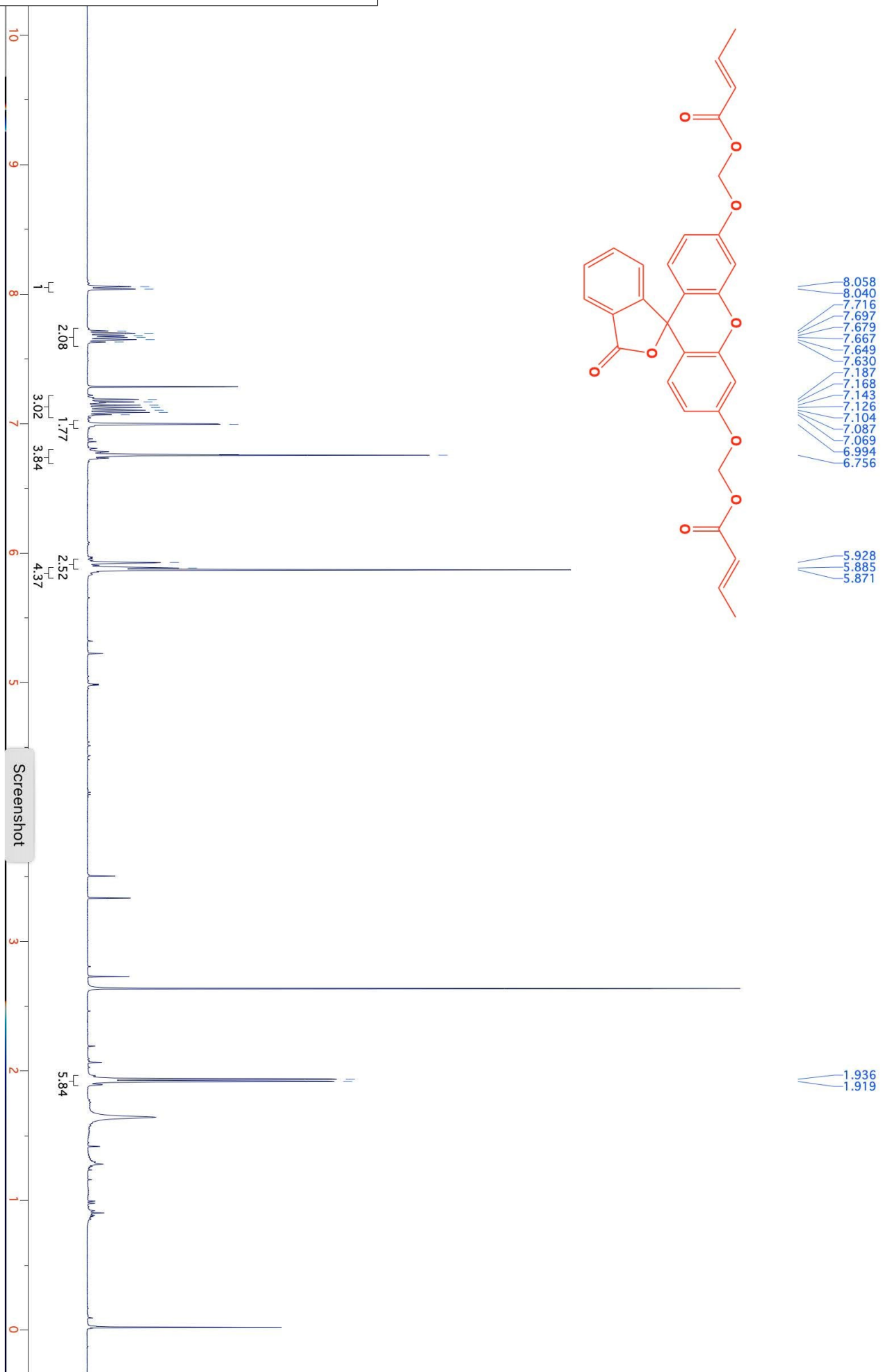


Figure S24

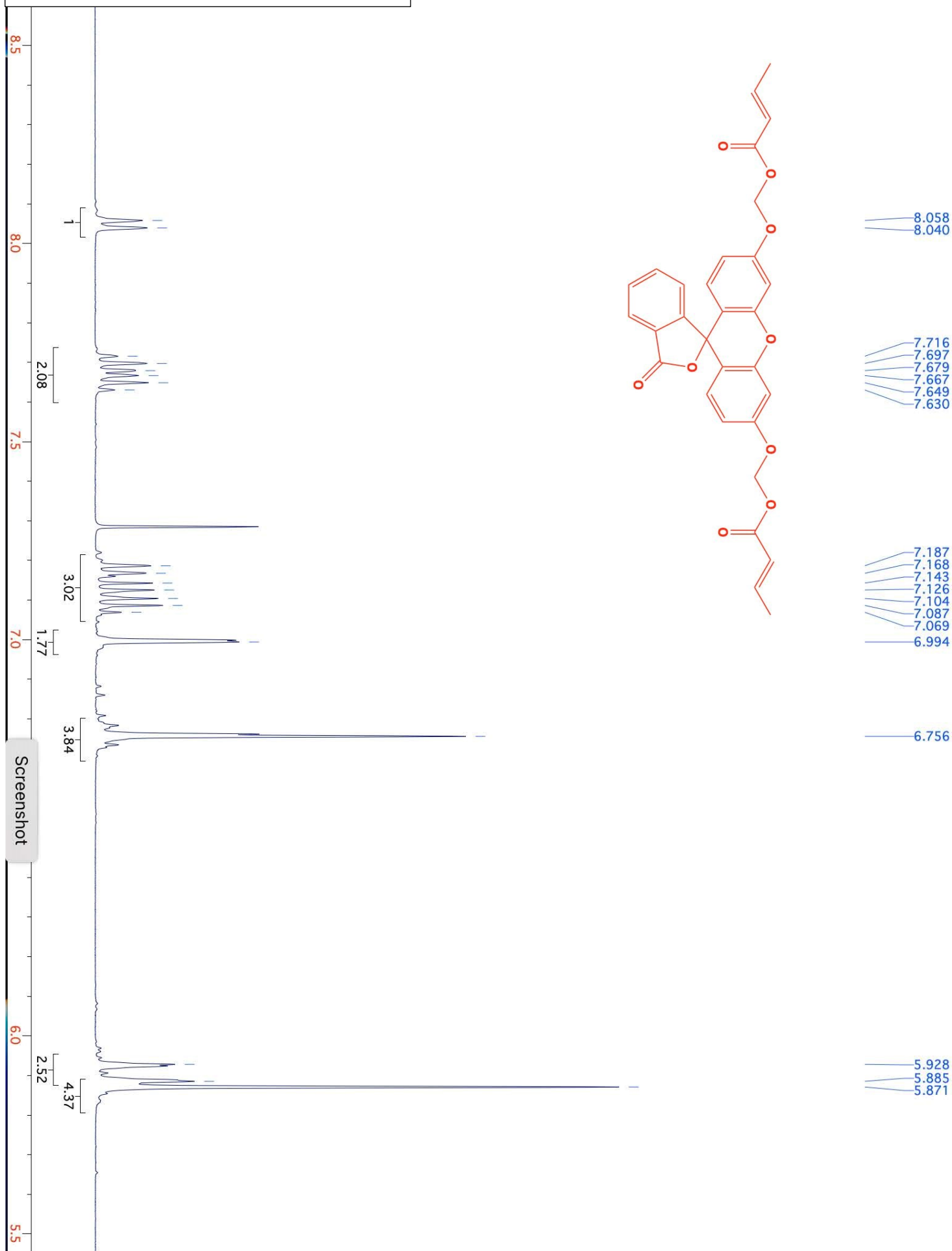


Figure S25

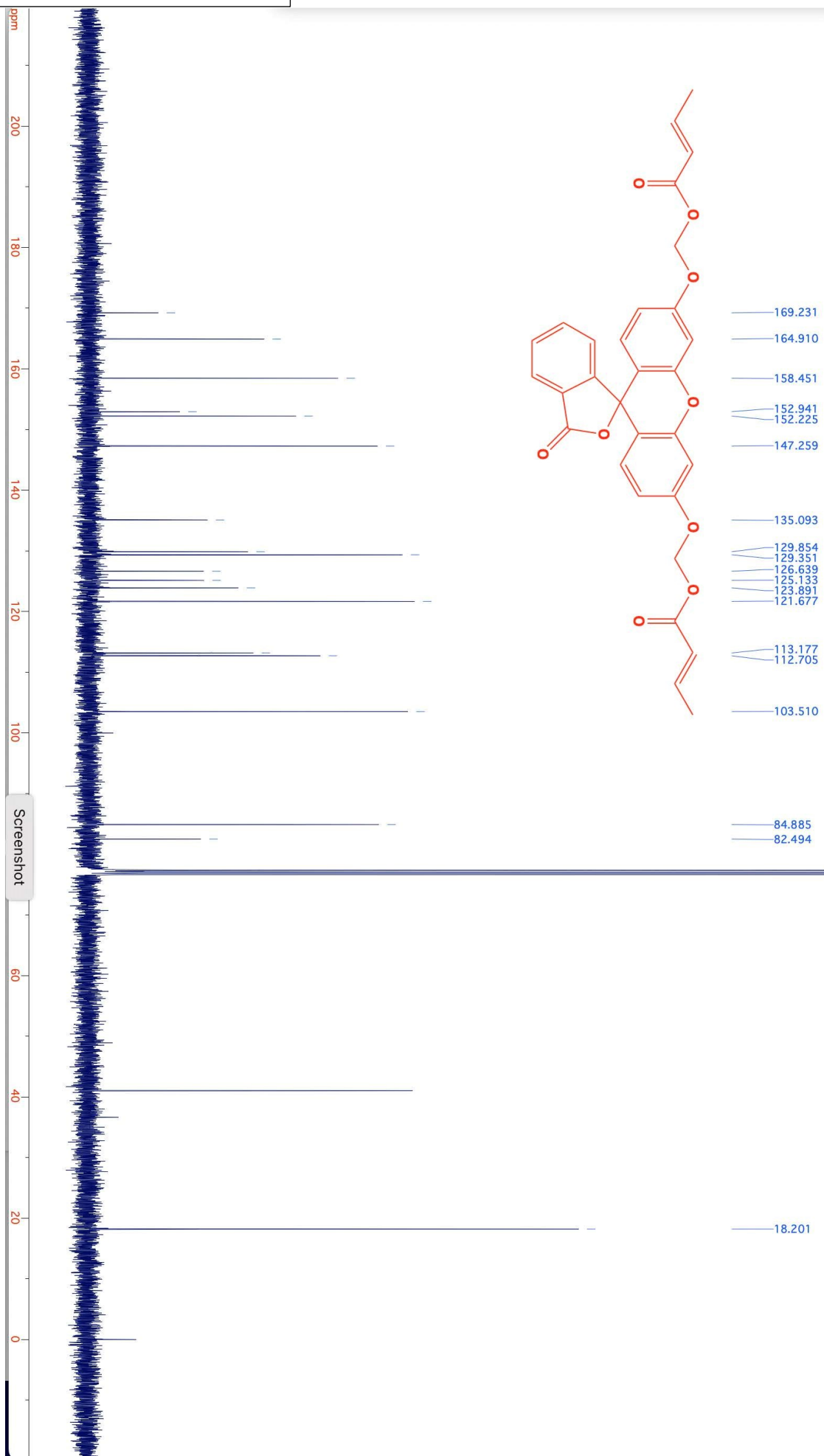


Figure S26

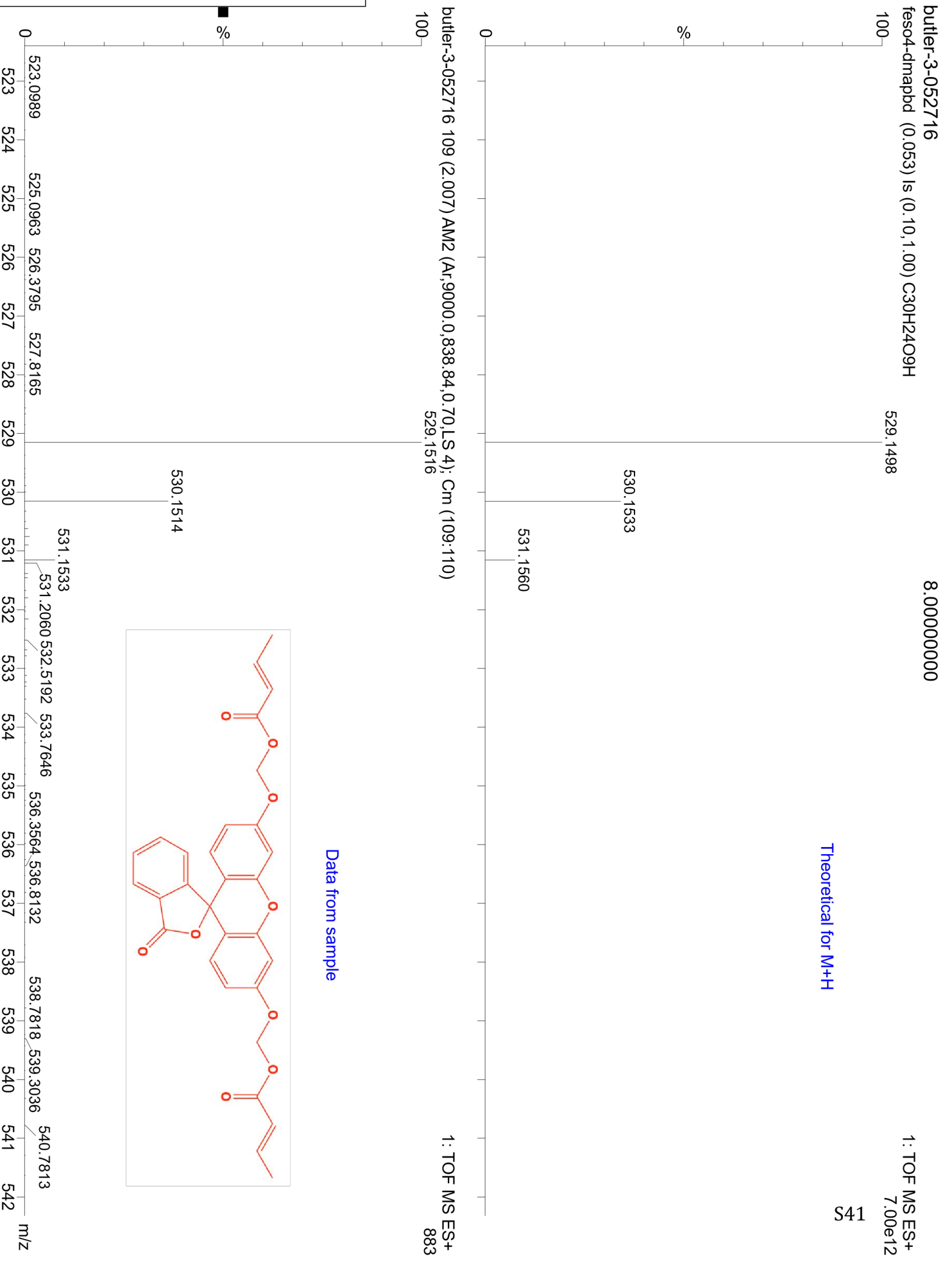


Figure S27

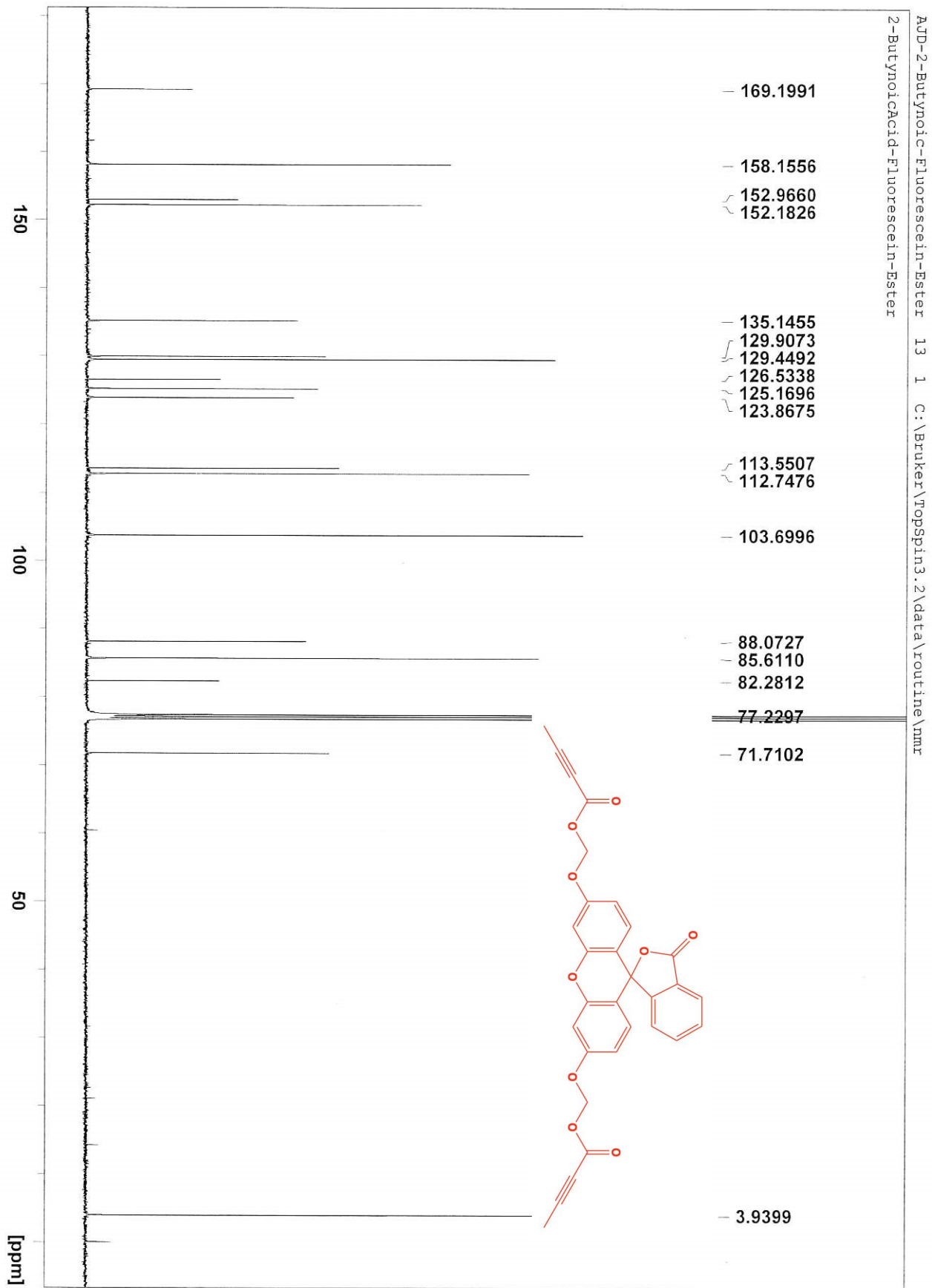


Figure S28

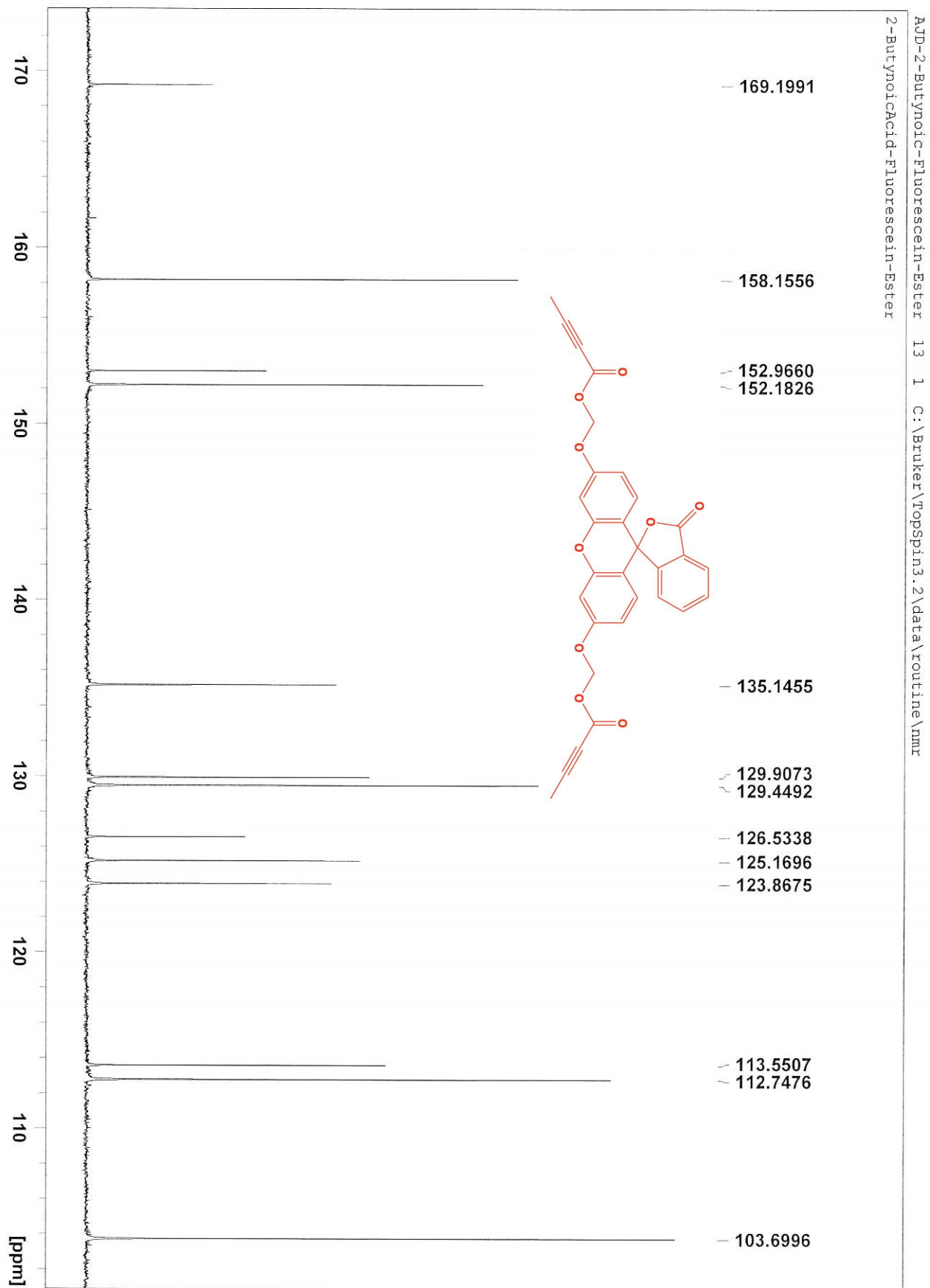


Figure S29

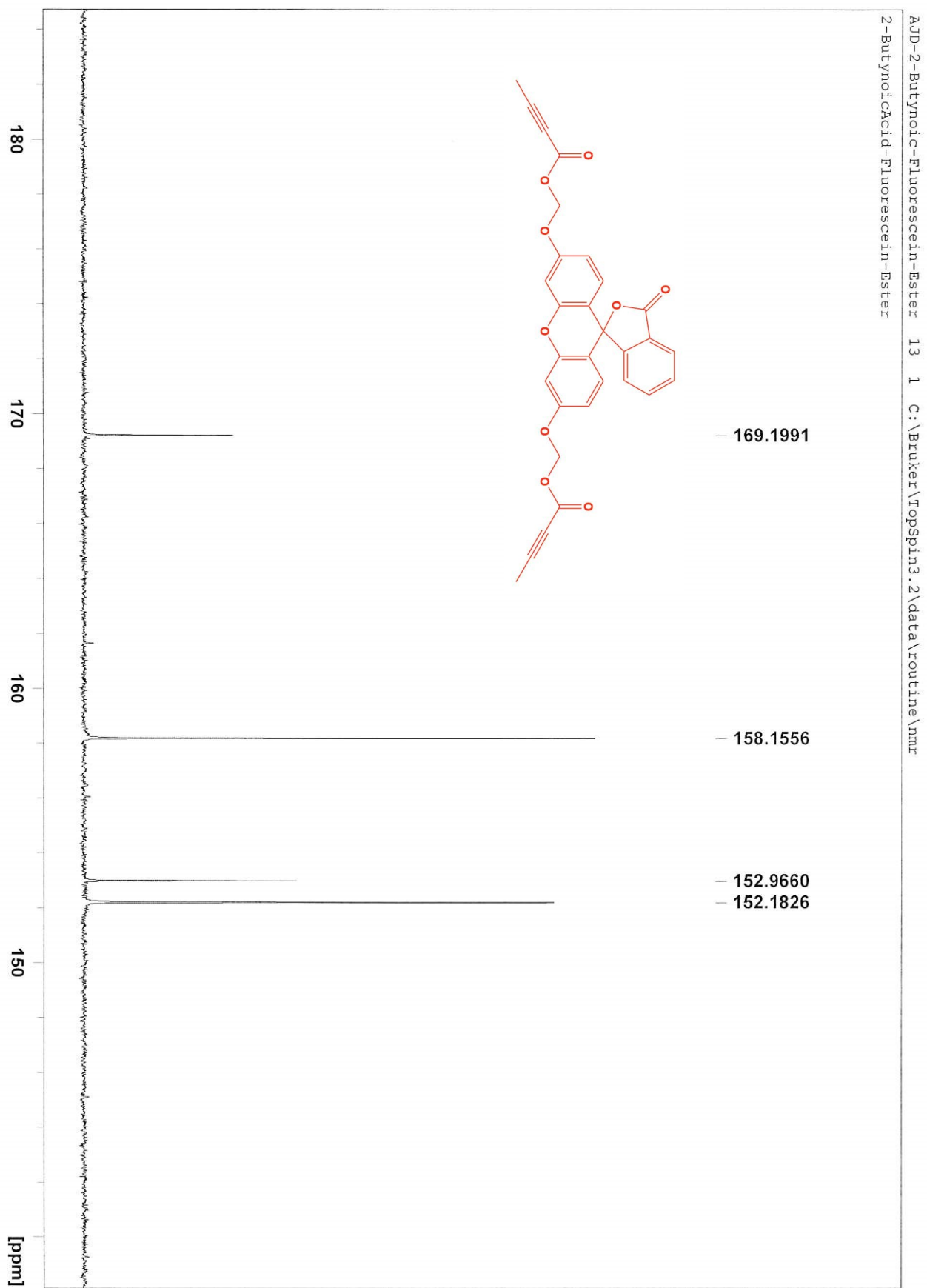


Figure S30

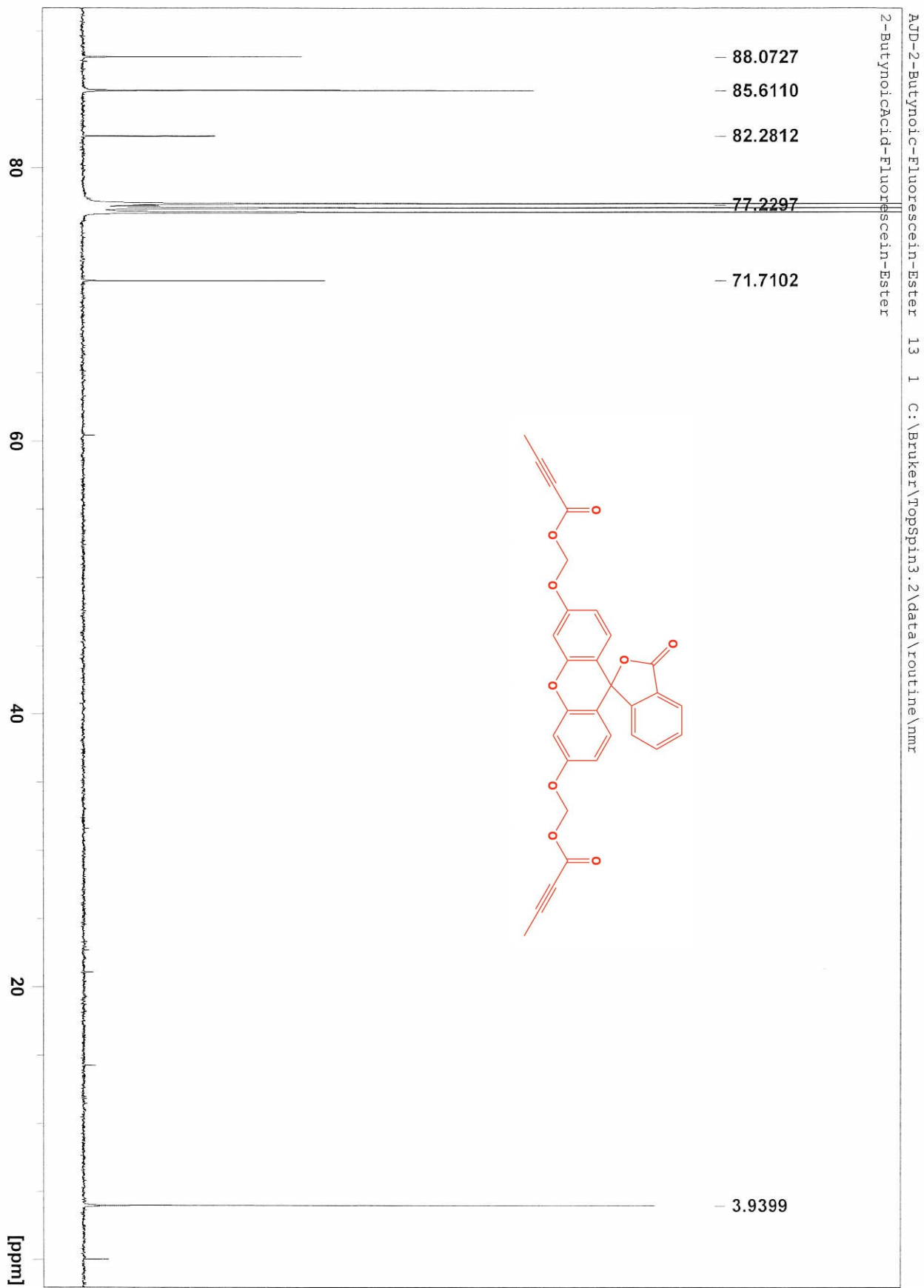


Figure S31

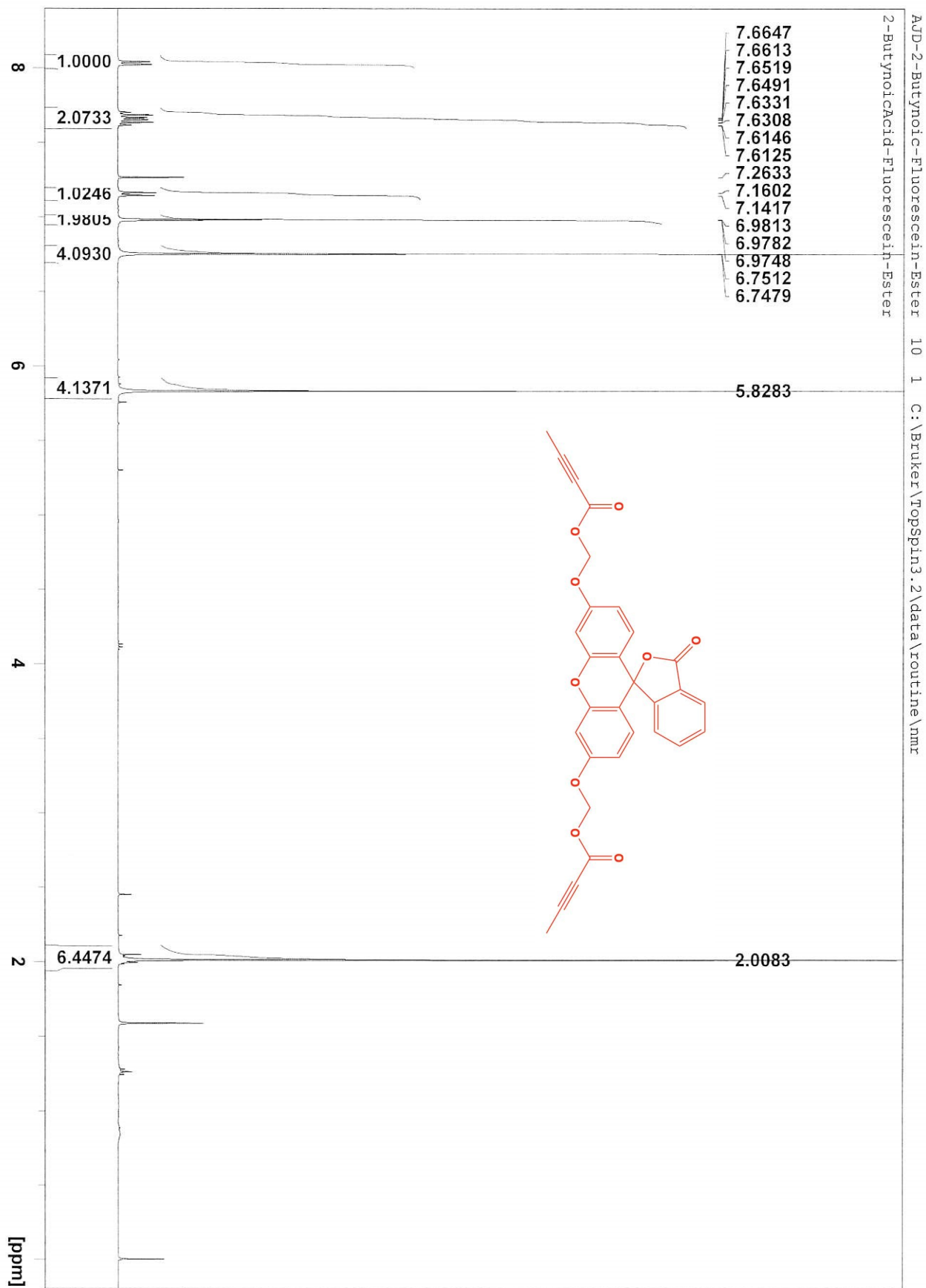


Figure S32

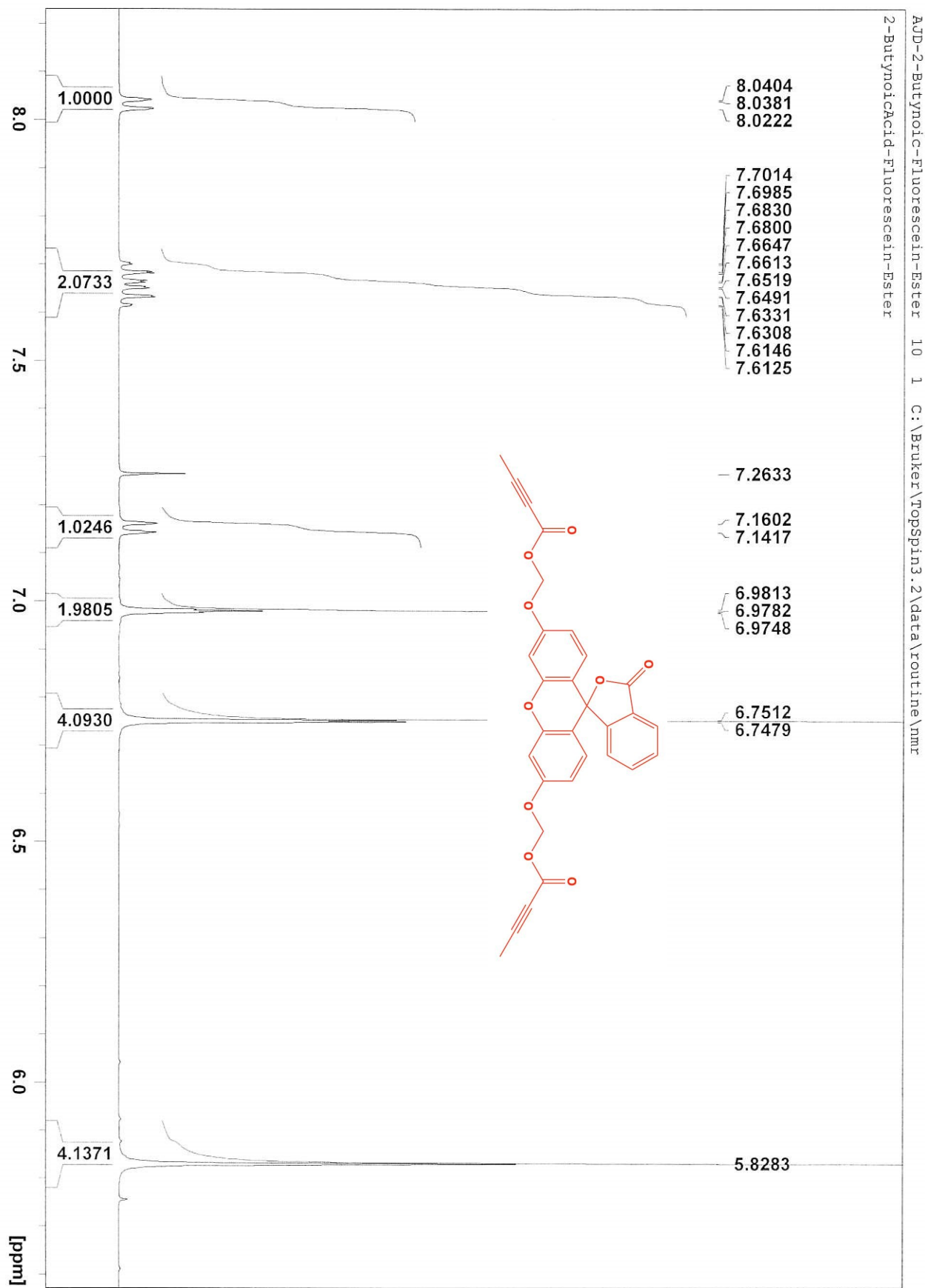
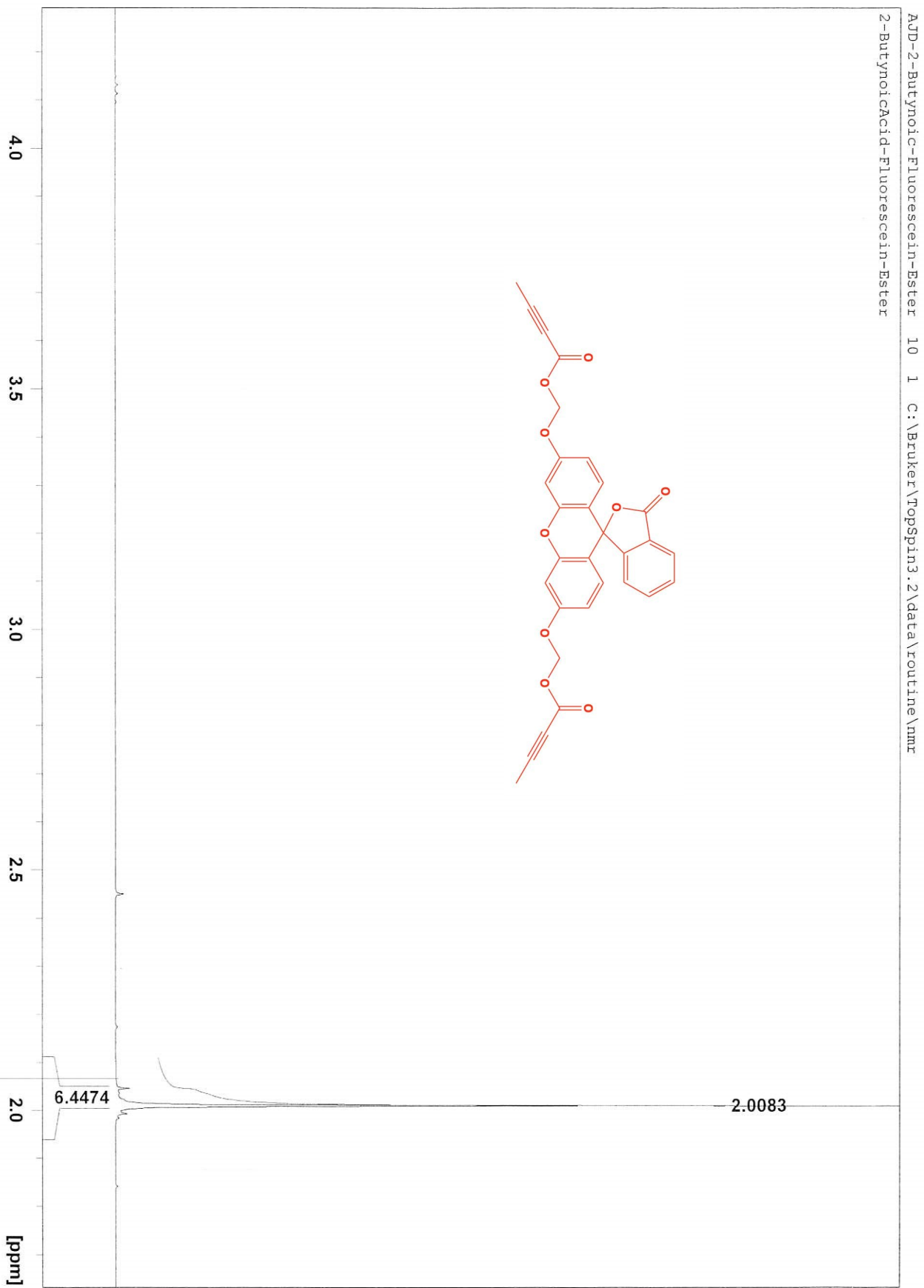


Figure S33



2-ButynoicAcid-Fluorescein-Ester

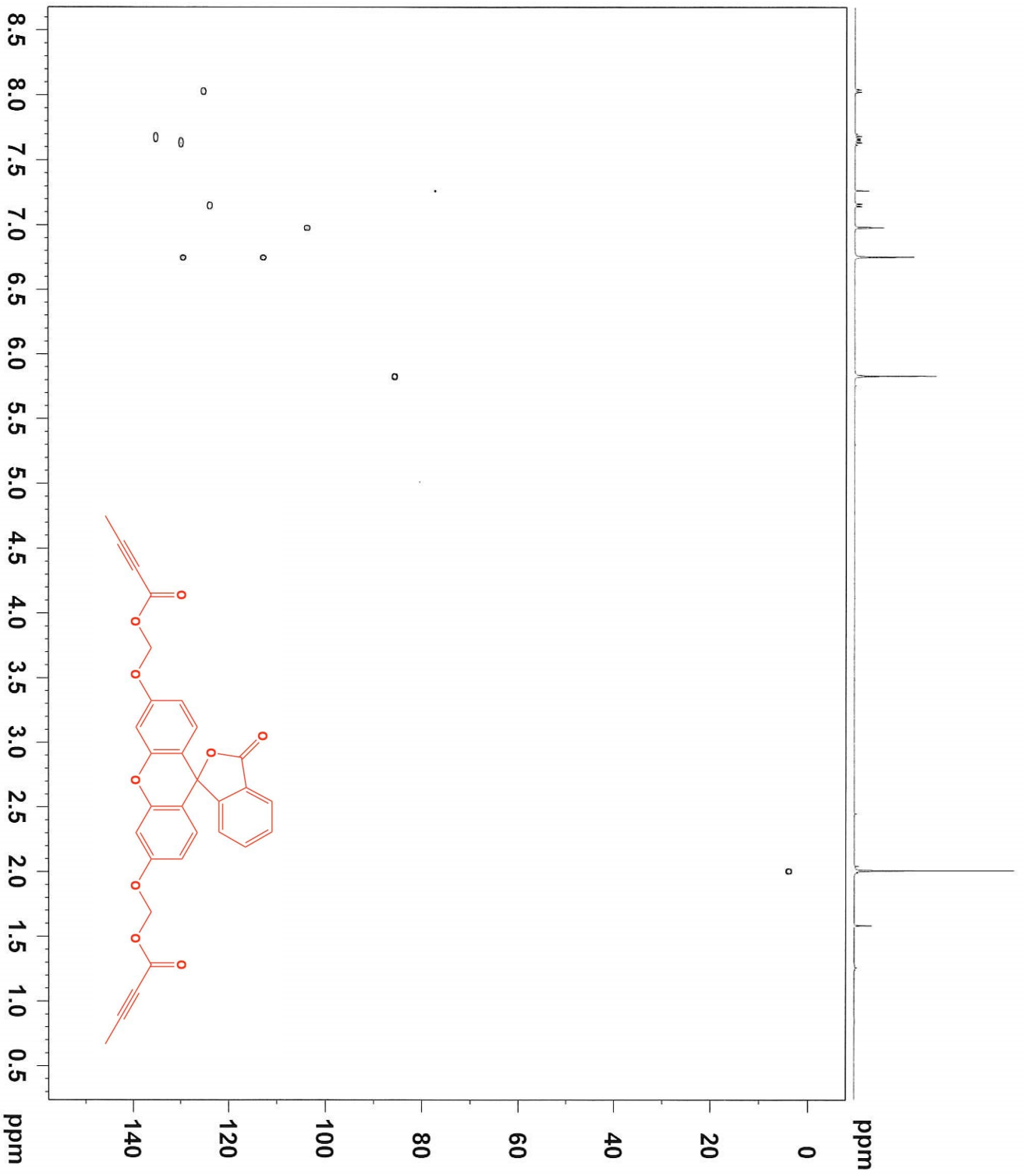
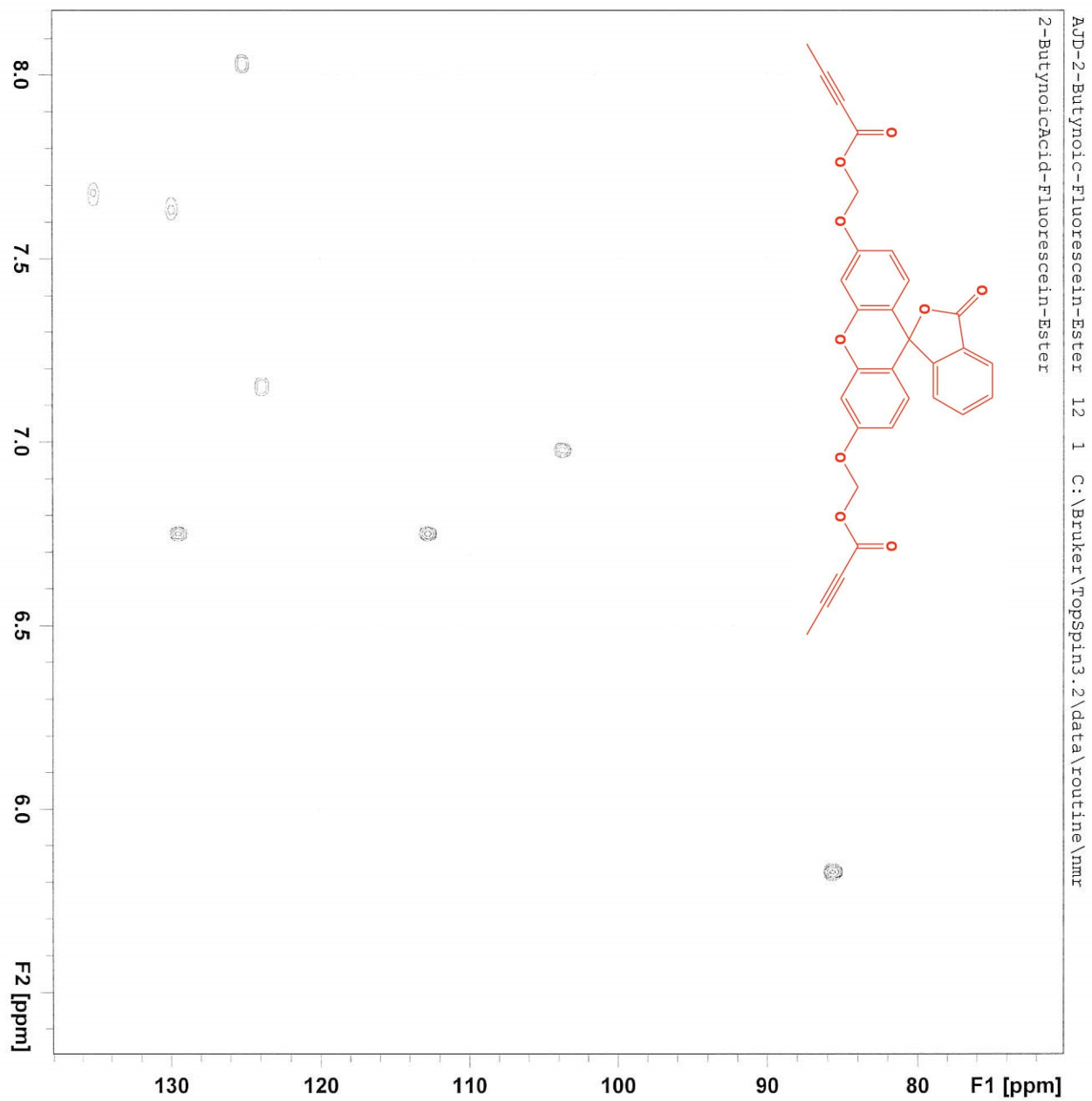


Figure S34

Figure S35



2-ButynoicAcid-Fluorescein-Ester

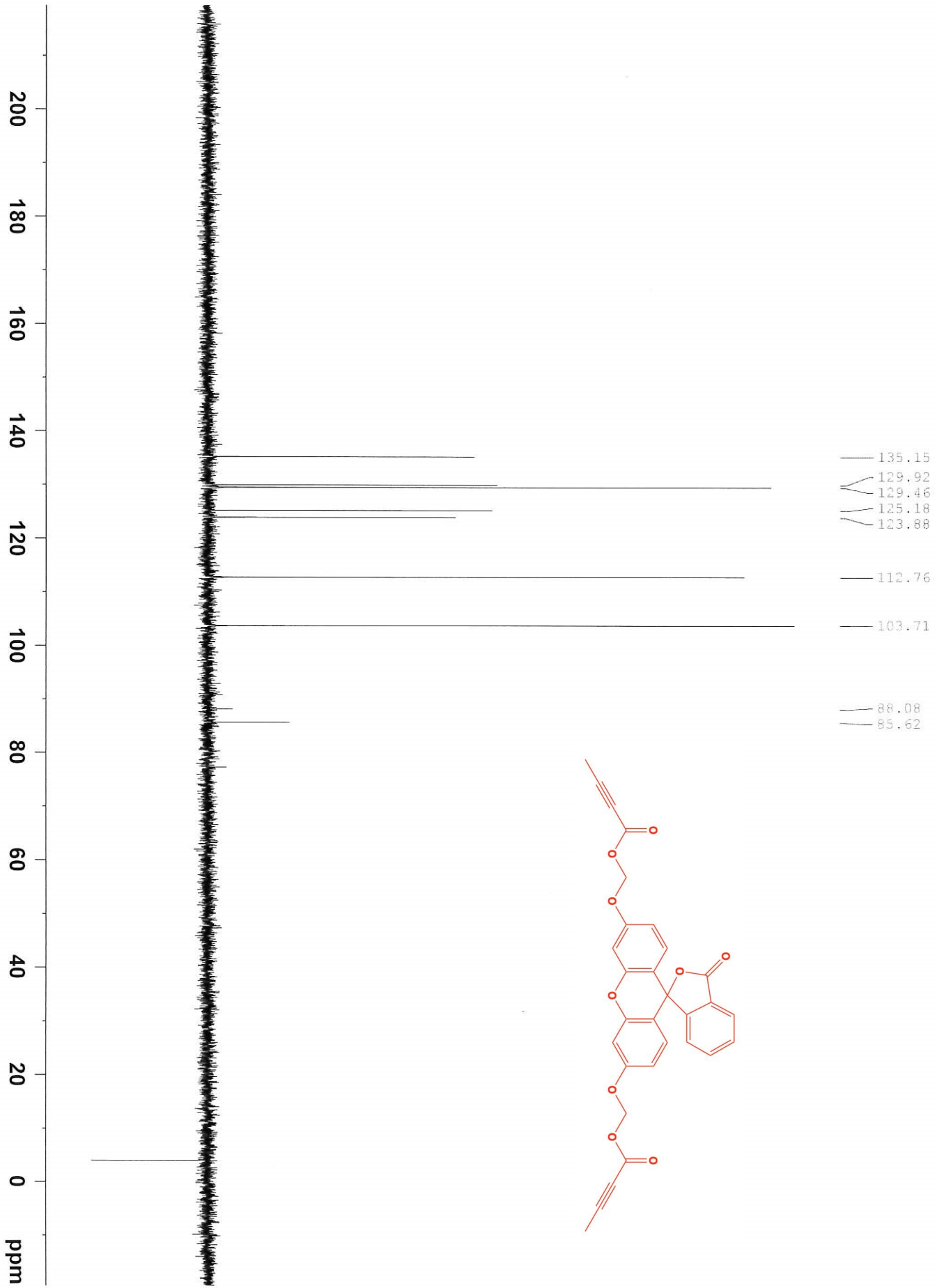
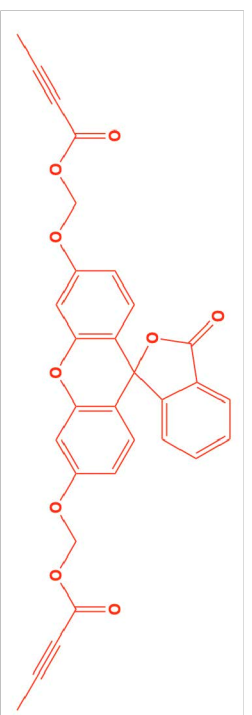
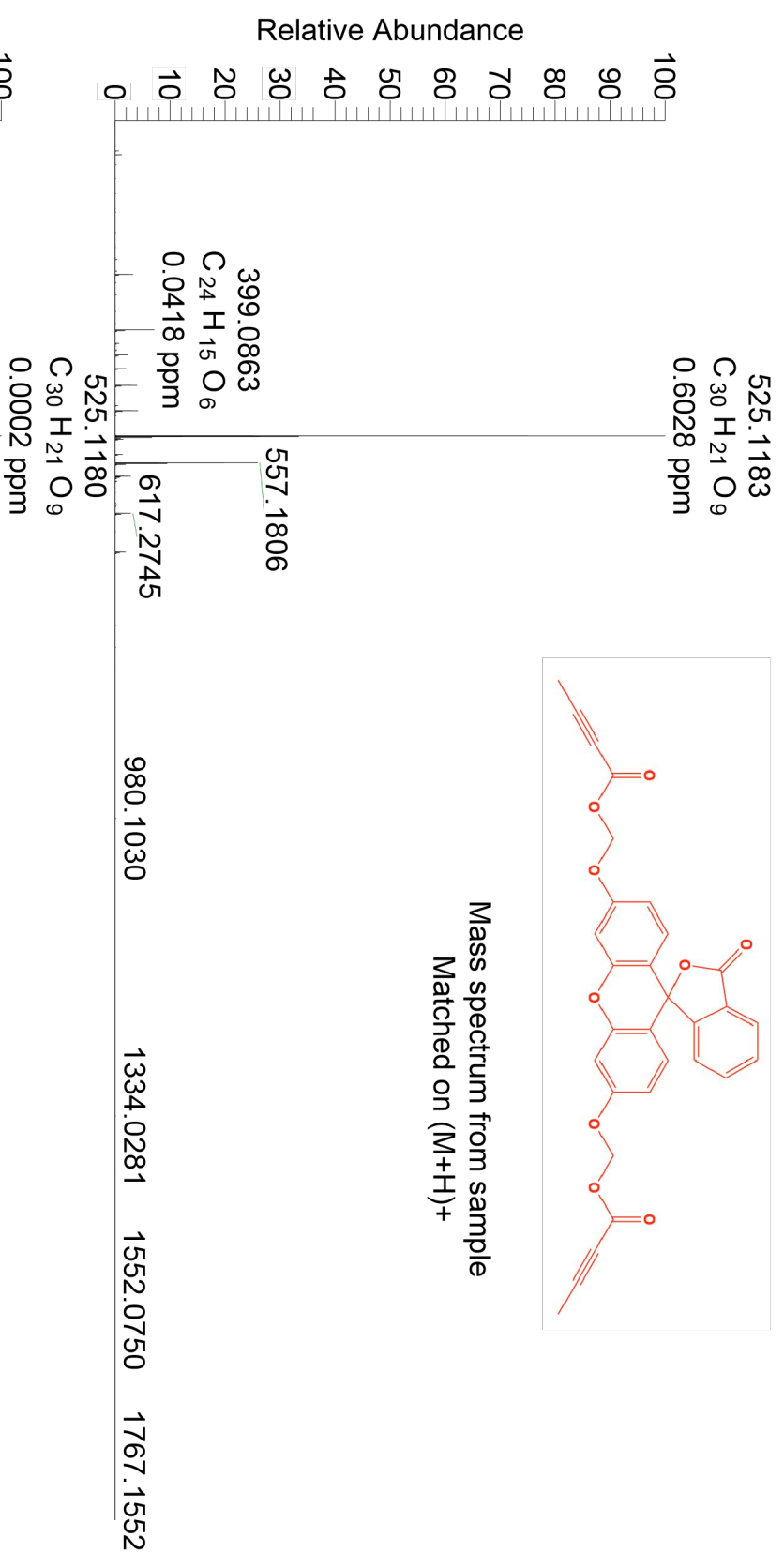


Figure S36



Mass spectrum from sample
Matched on (M+H)+

NL:
3.61E7
JE-1#19-21 RT: 0.170.19
AV: 3 T: FTMS + p APPI
corona Full ms
[150.00-1800.00]



NL:
7.07E5

C₃₀H₂₀O₉H:
C₃₀H₂₁O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Theoretical mass spectrum for (M+H)+

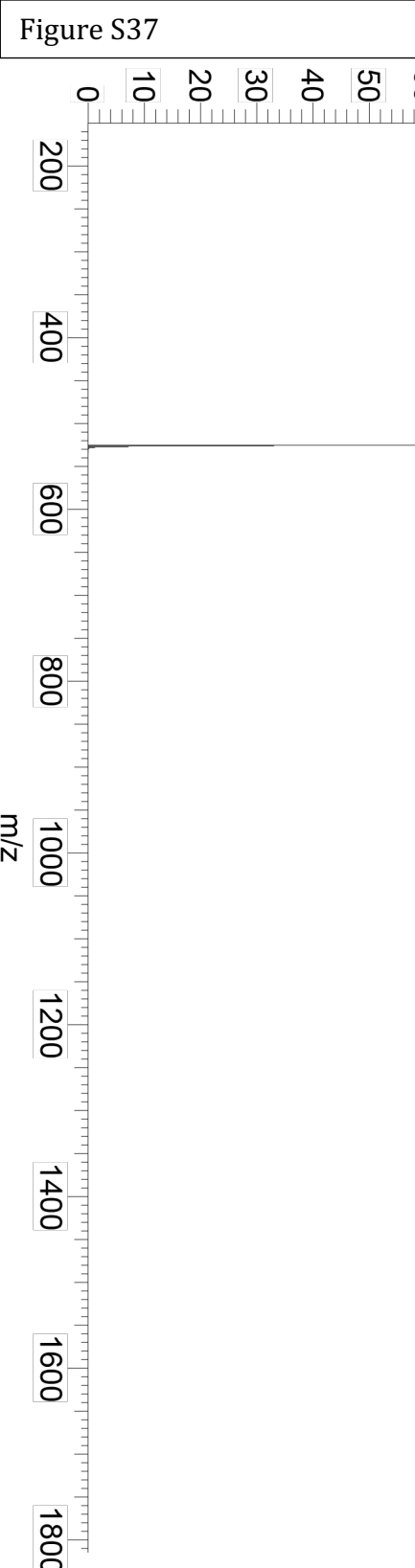
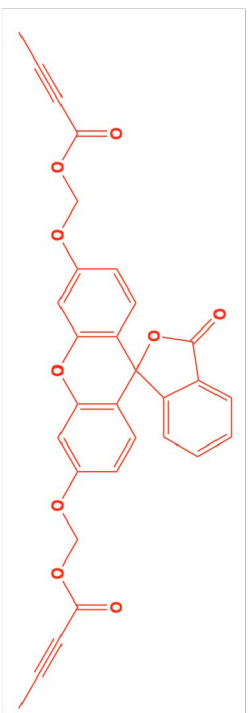
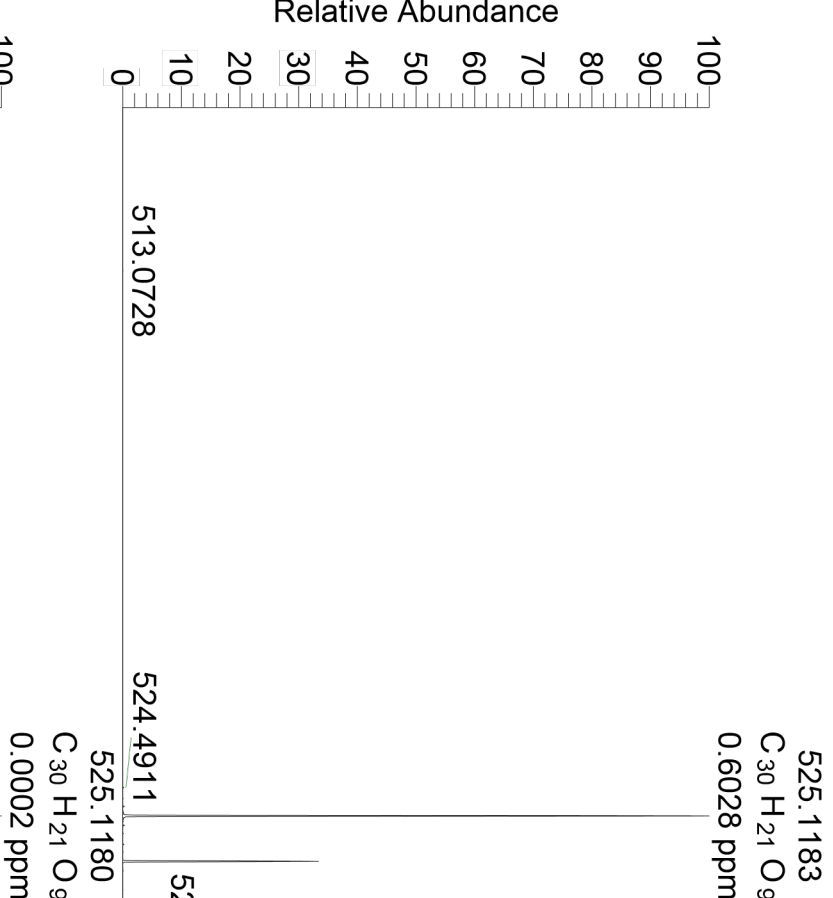


Figure S37



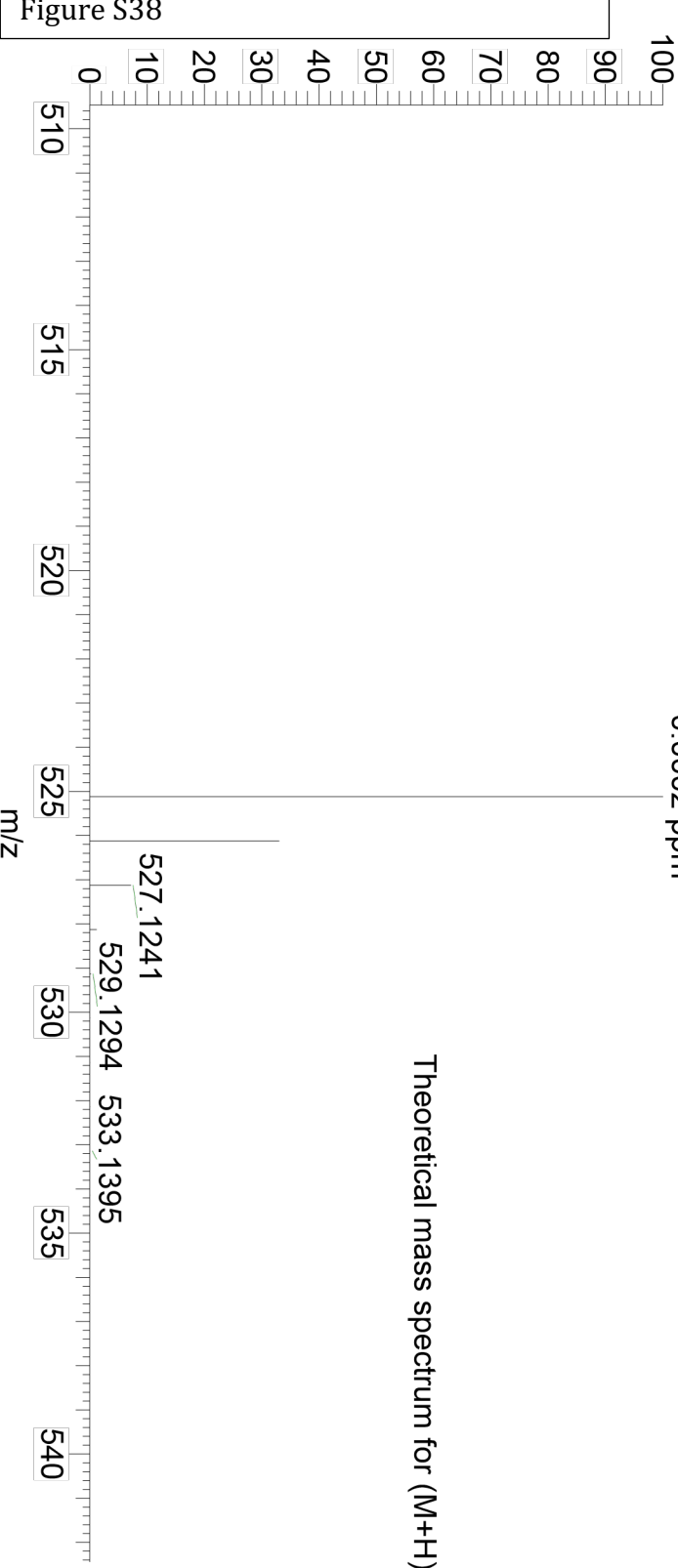
NL:
3.61E7
JE-1#19-21 RT: 0.17-0.19
AV: 3 T: FTMS + p APPI
corona Full ms
[150.00-1800.00]



525.1183
C₃₀ H₂₁ O₉
0.6028 ppm

525.1180
C₃₀ H₂₁ O₉
0.0002 ppm

Theoretical mass spectrum for (M+H)⁺



NL:
7.07E5
C₃₀ H₂₀ O₉ H:
C₃₀ H₂₁ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Figure S38

Figure S39

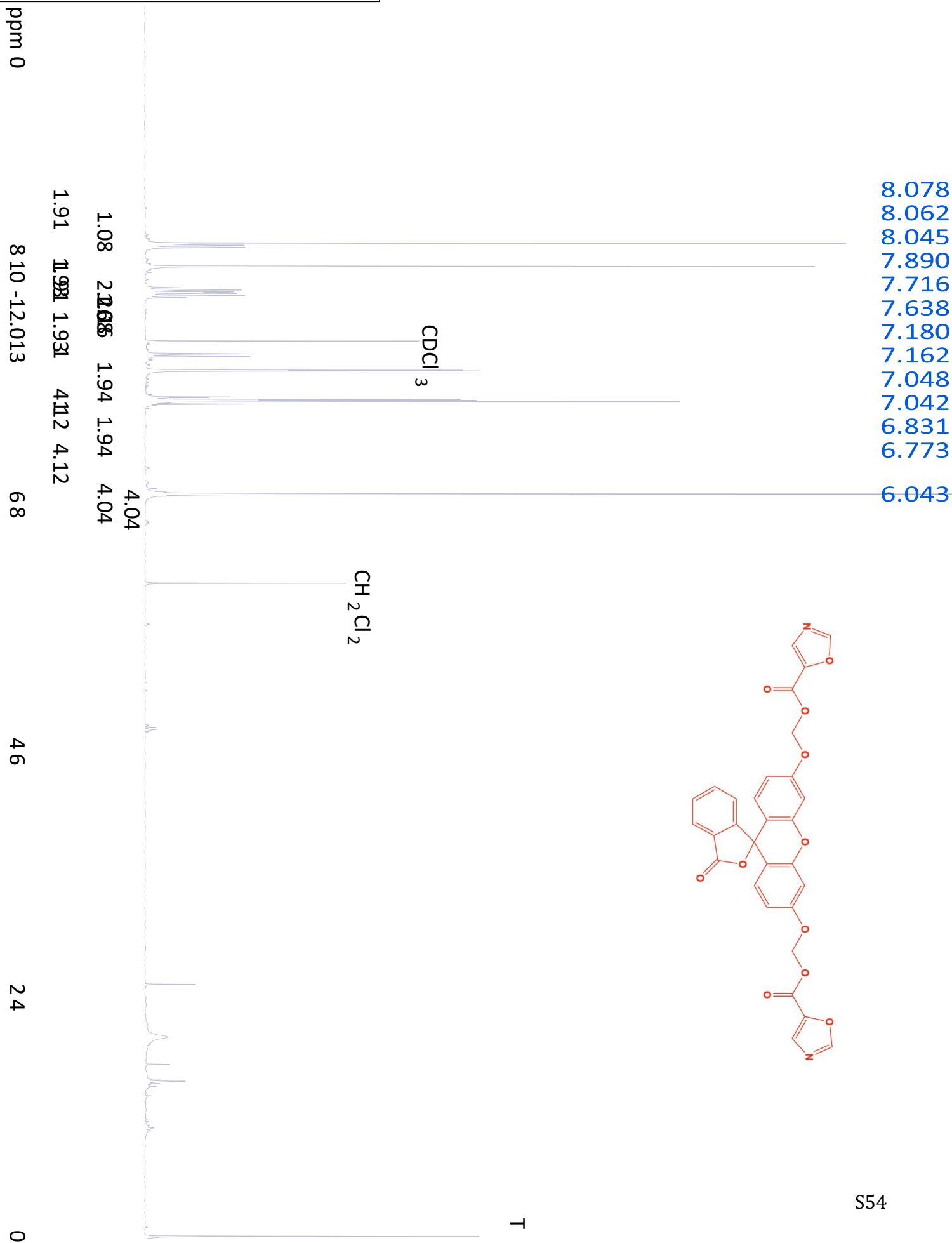


Figure S40

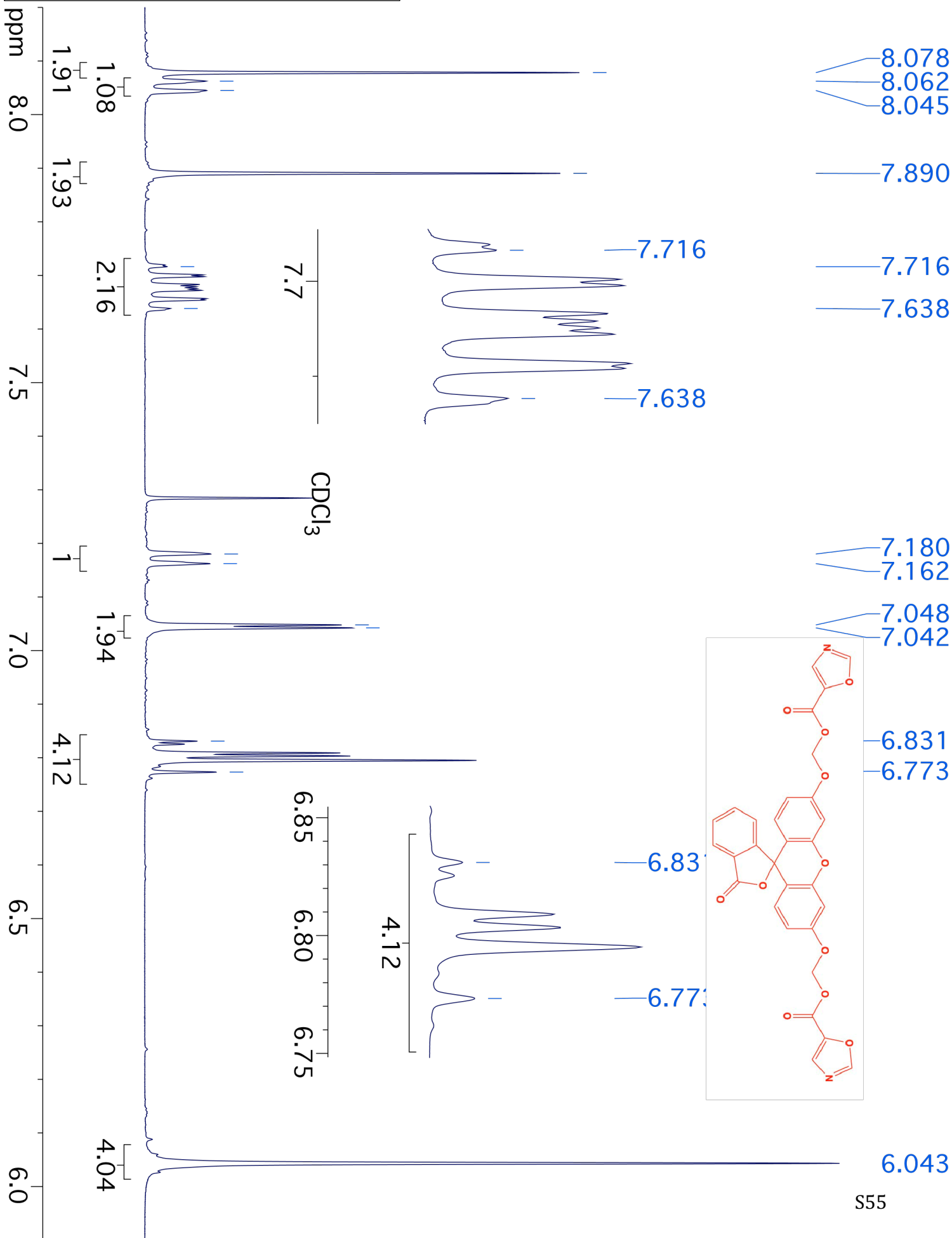


Figure S41

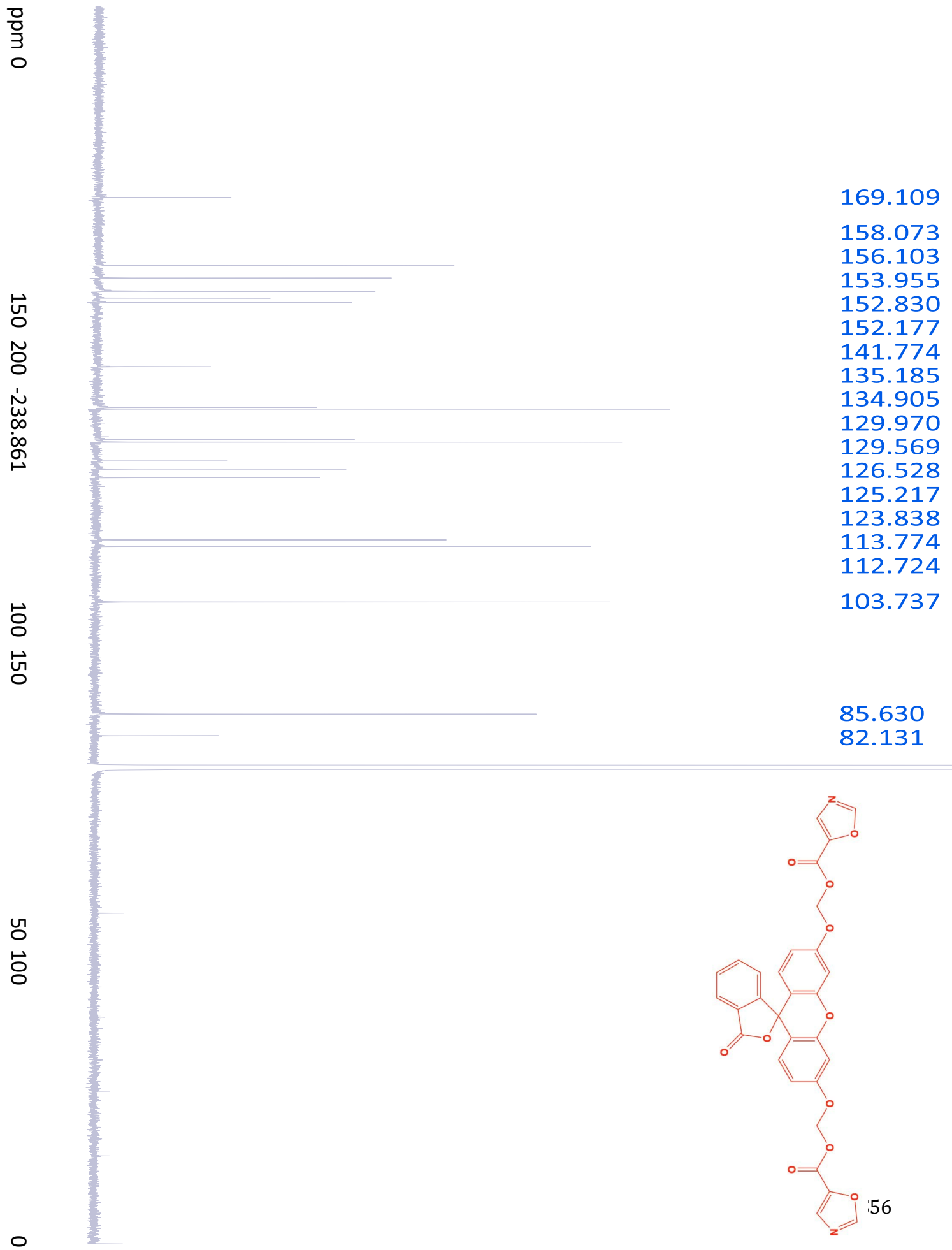


Figure S42

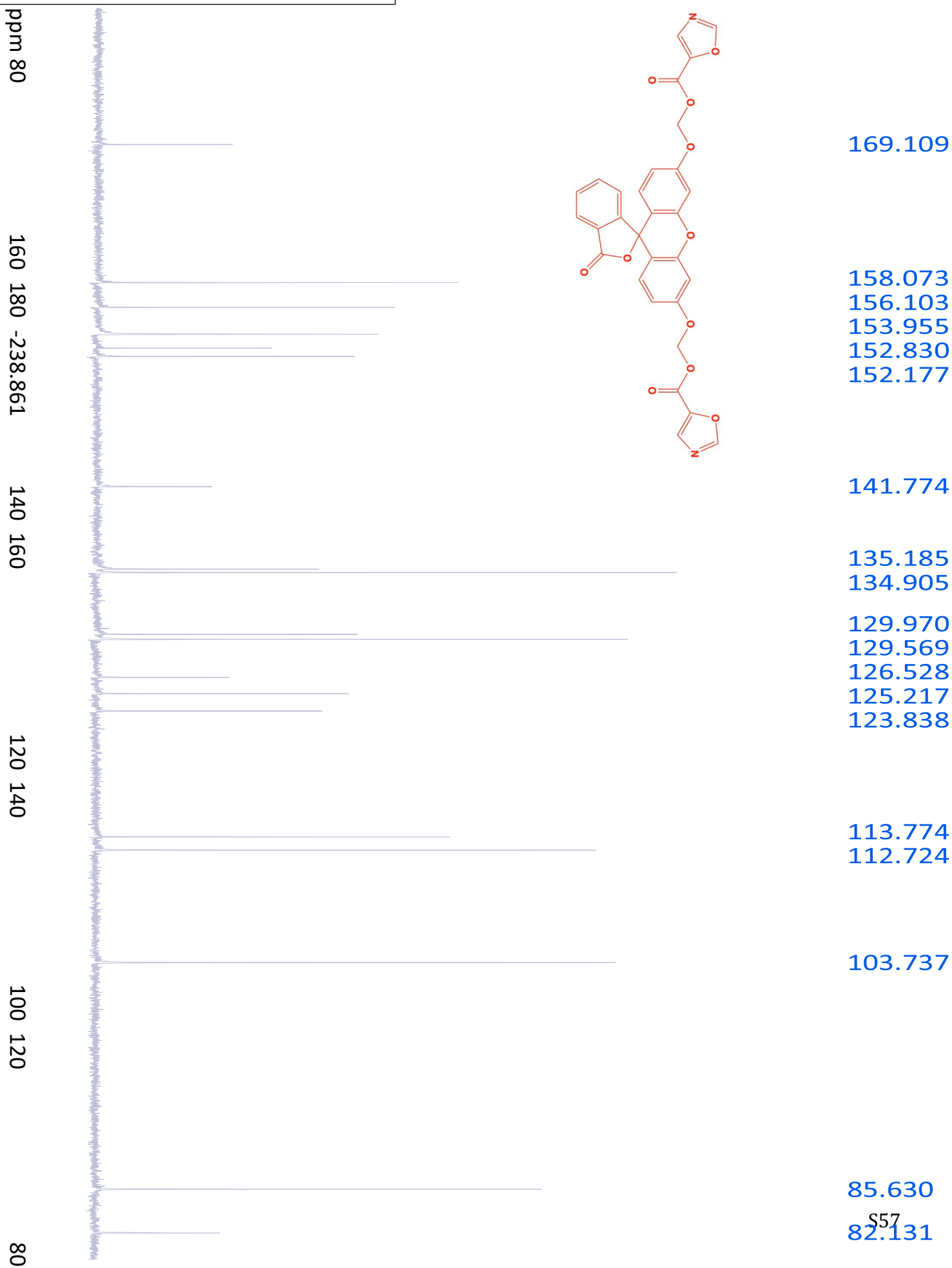


Figure S43

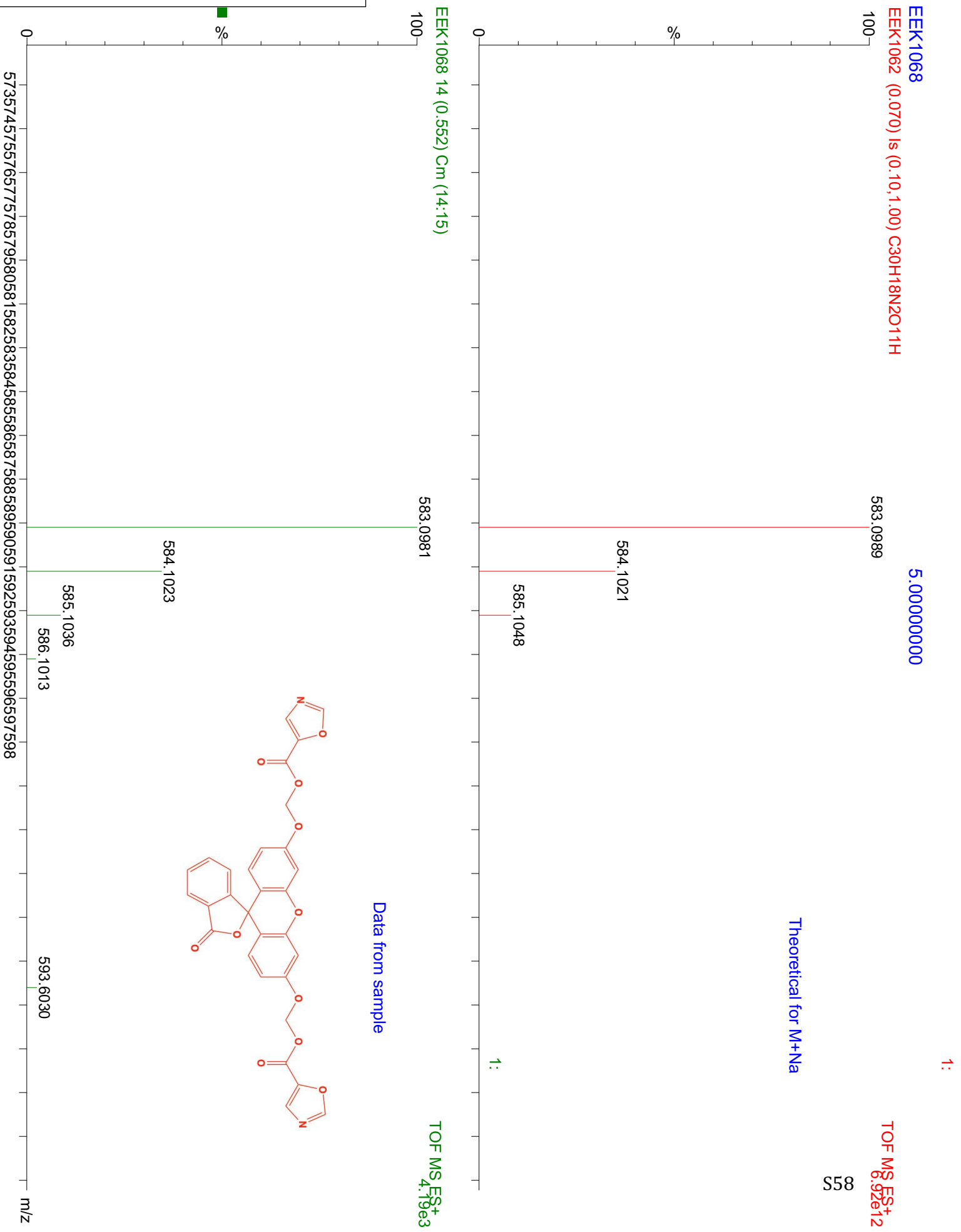


Figure S44

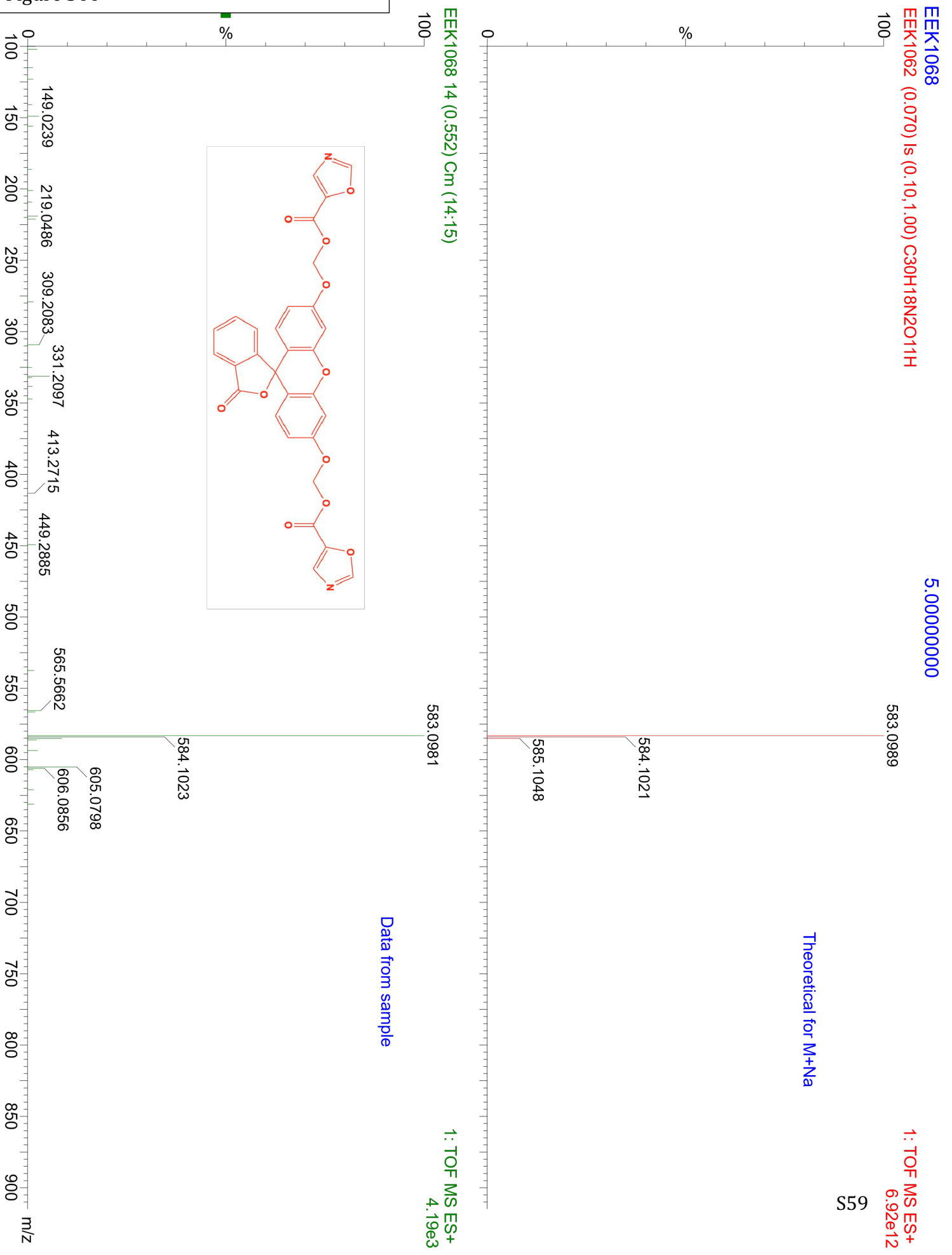


Figure S45

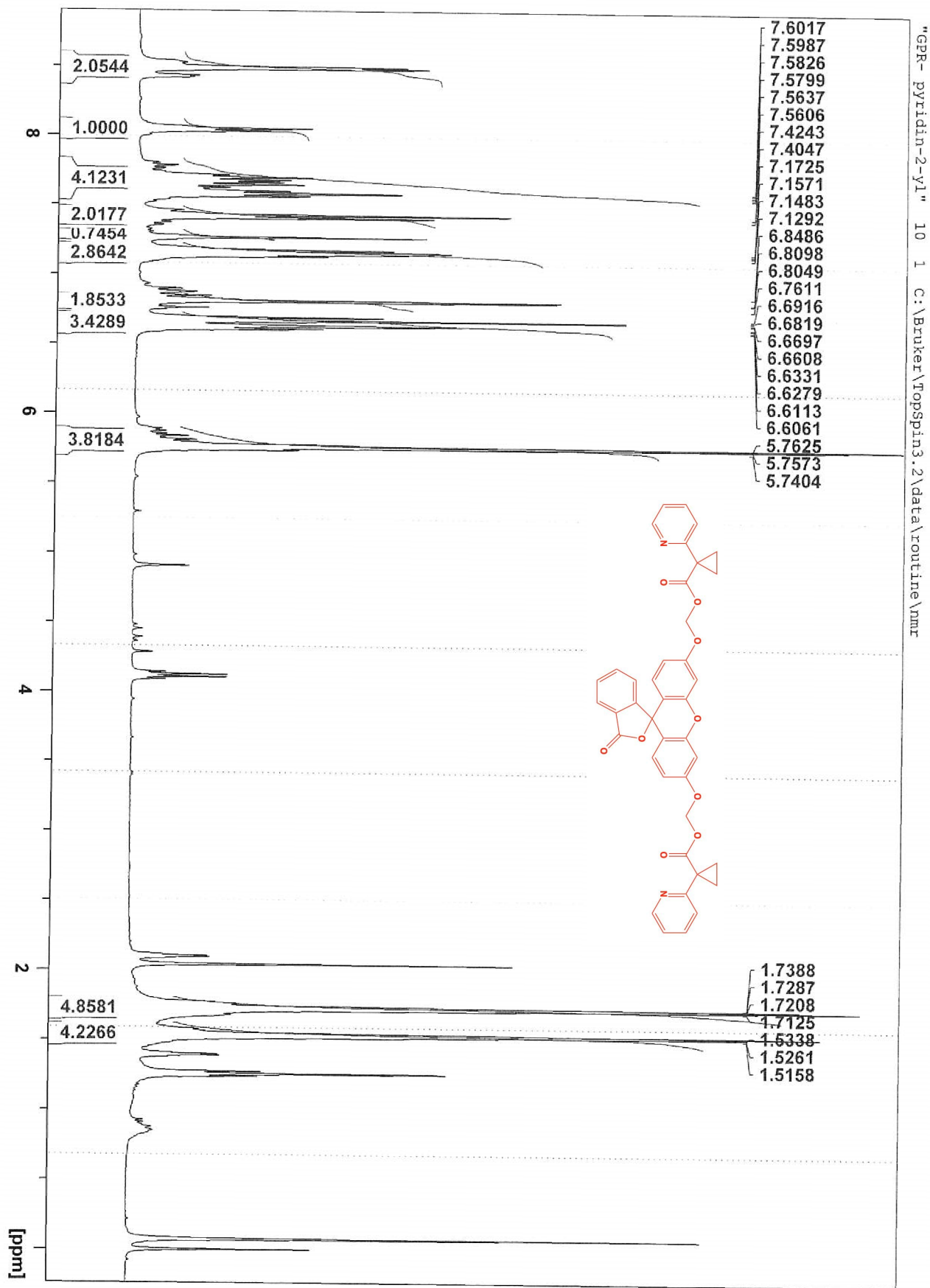


Figure S46

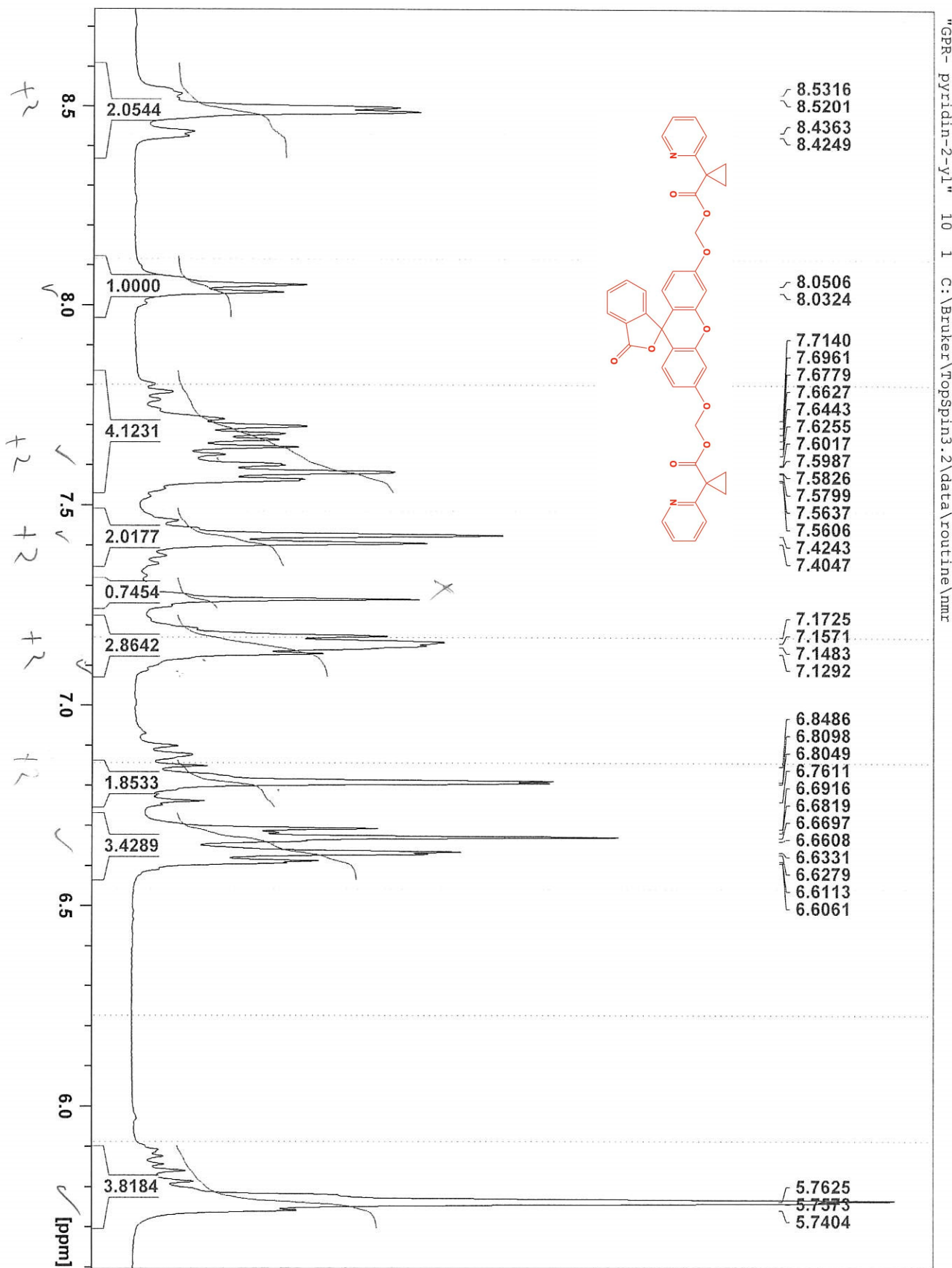
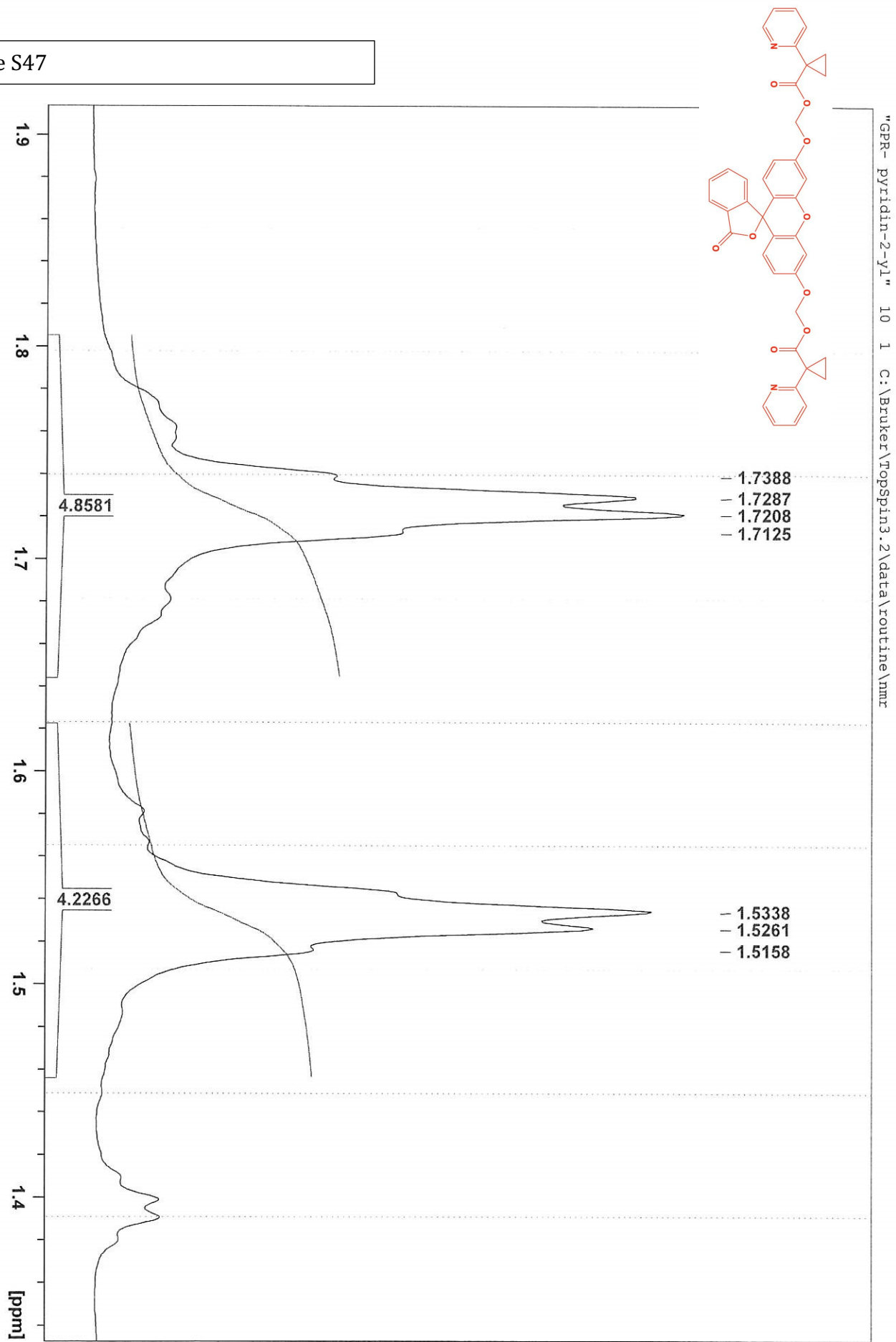


Figure S47



1.9
1.8
1.7
1.6
1.5
1.4
[ppm]

4.8581

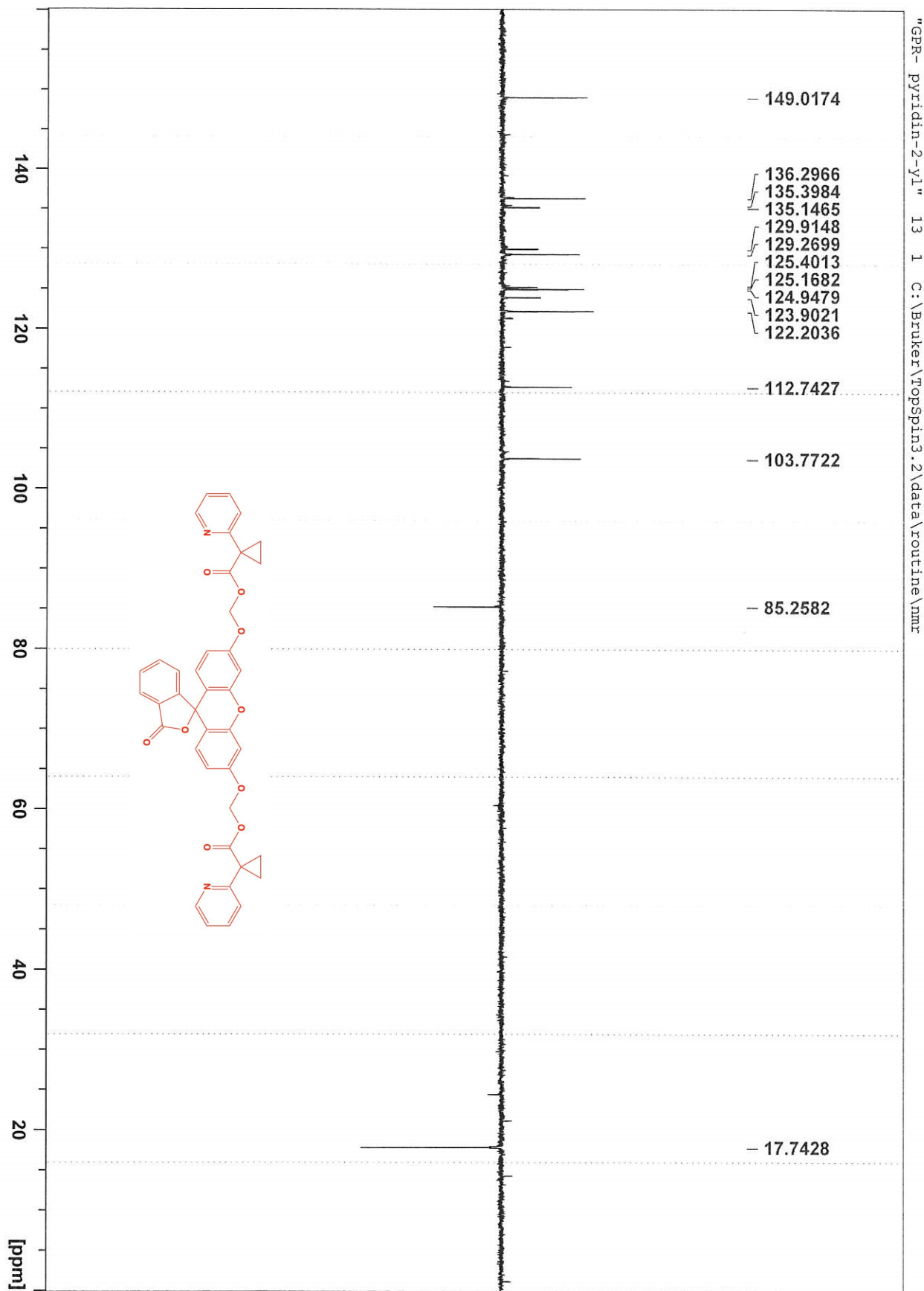
4.2266

- 1.7388
- 1.7287
- 1.7208
- 1.7125

- 1.5338
- 1.5261
- 1.5158

"GPR- pyridin-2-yl" 10 1 C:\Bruker\TopSpin3.2\data\routine\nmr

Figure S48



"GPR- pyridin-2-yl" 13 1 C:\Bruker\TopSpin3.2\data\routine\nmr

Figure S49

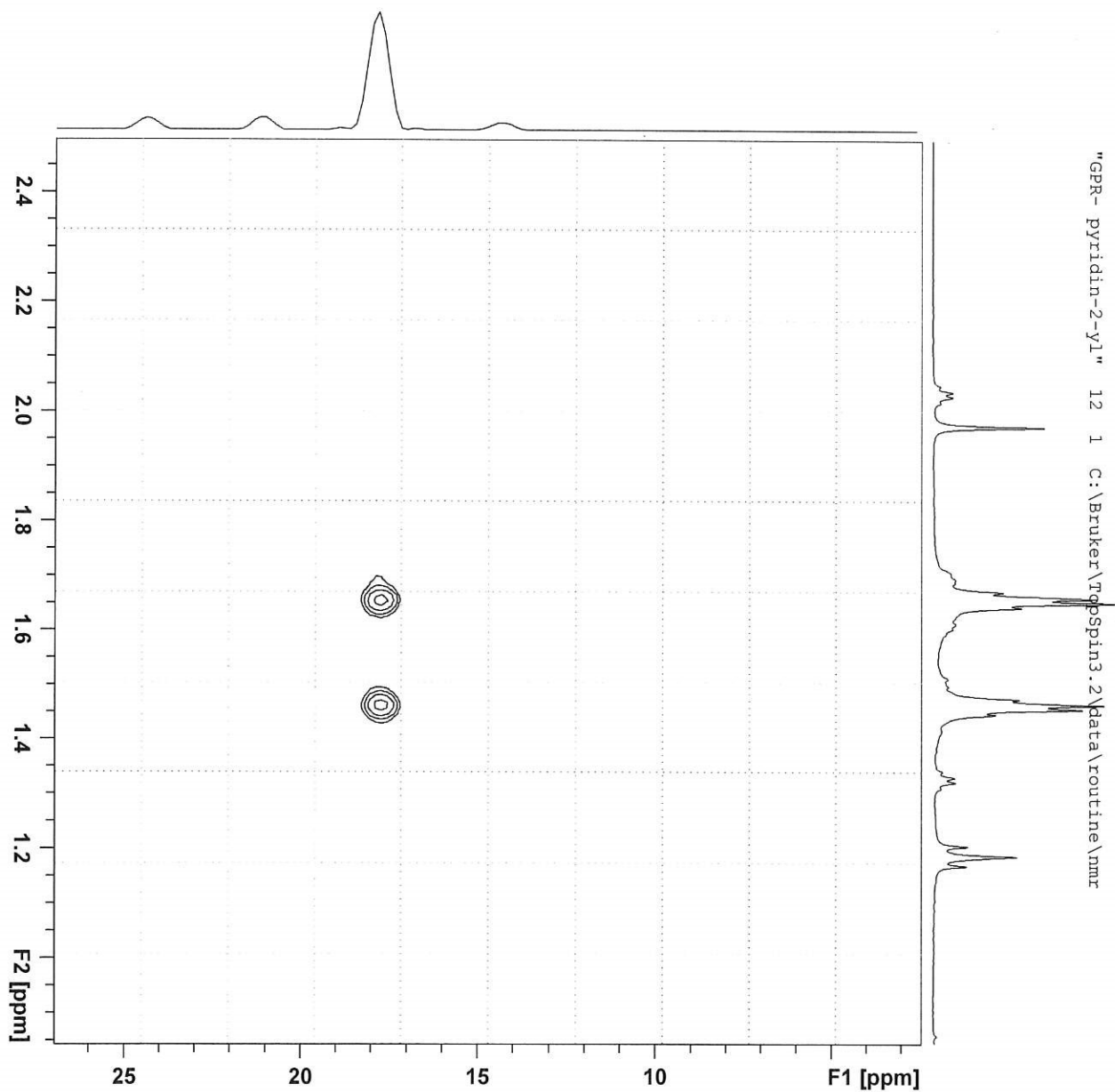
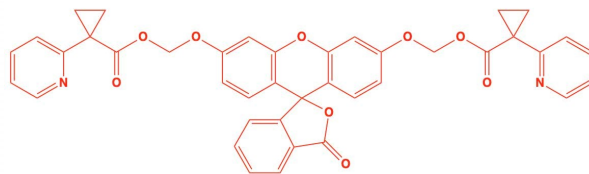


Figure S50

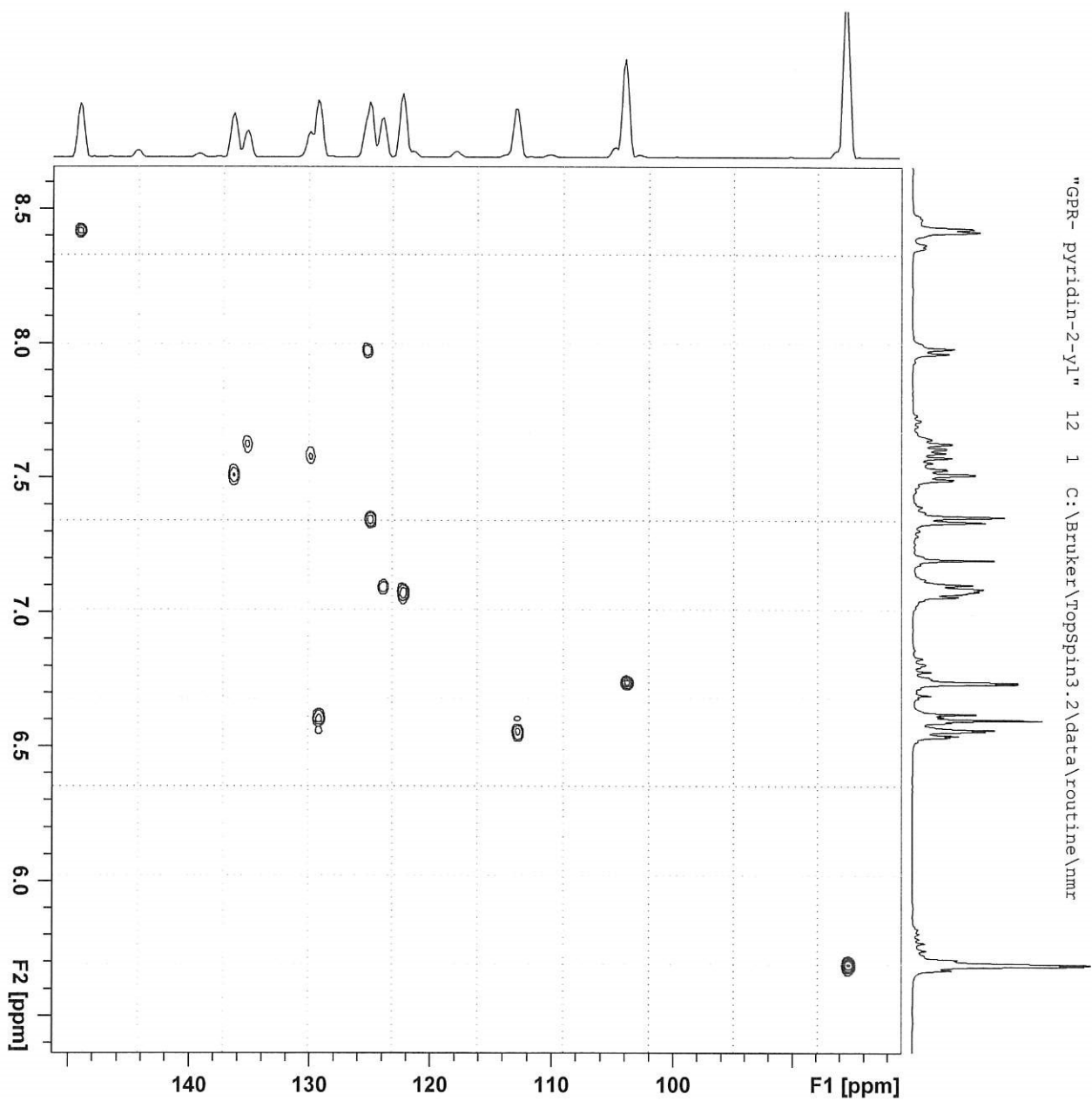
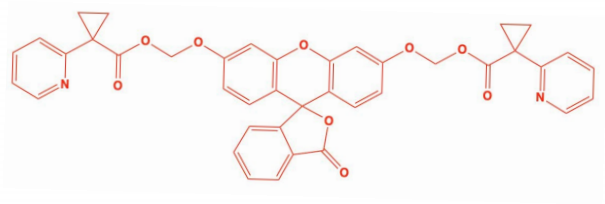


Figure S51

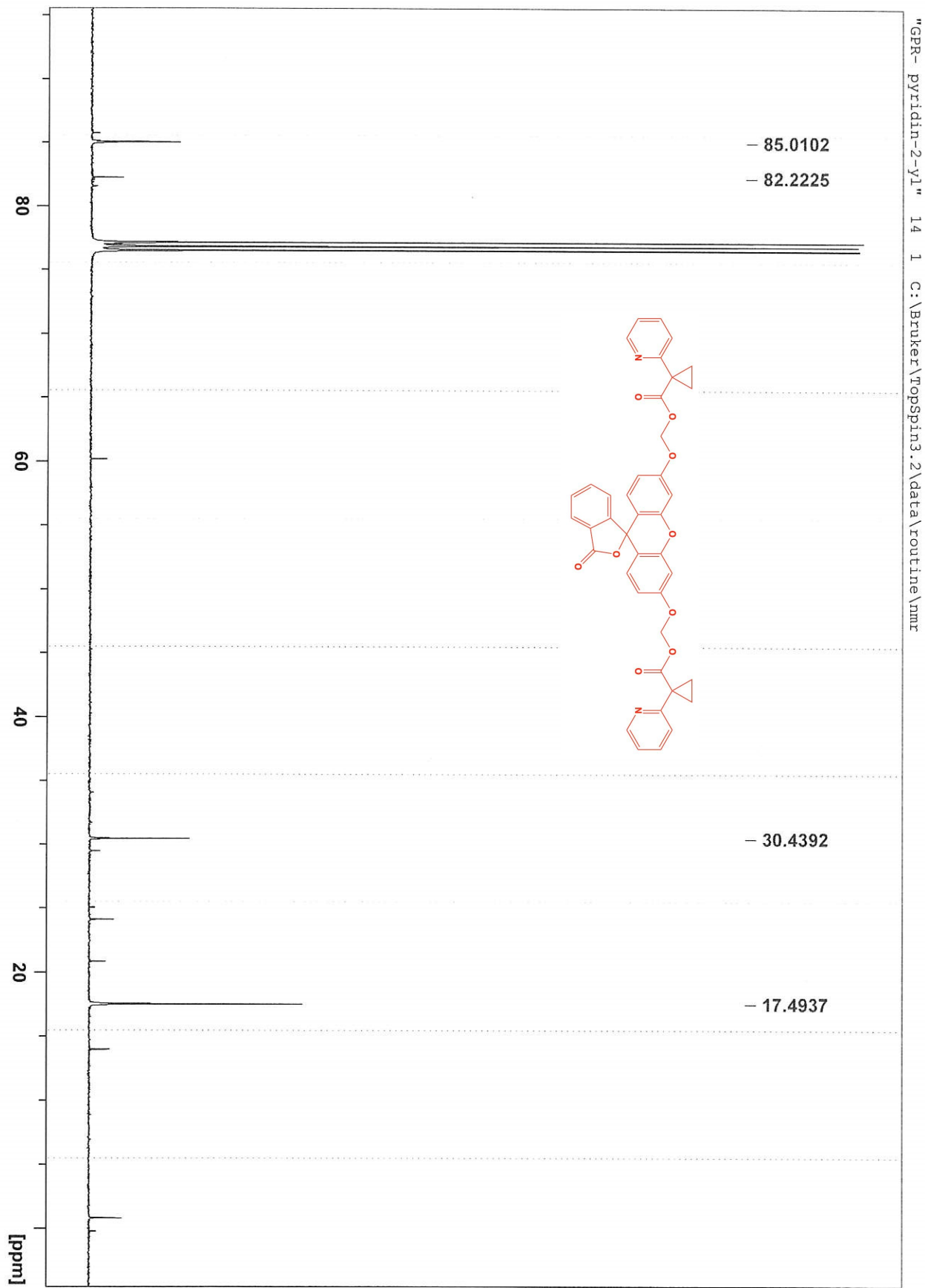


Figure S52

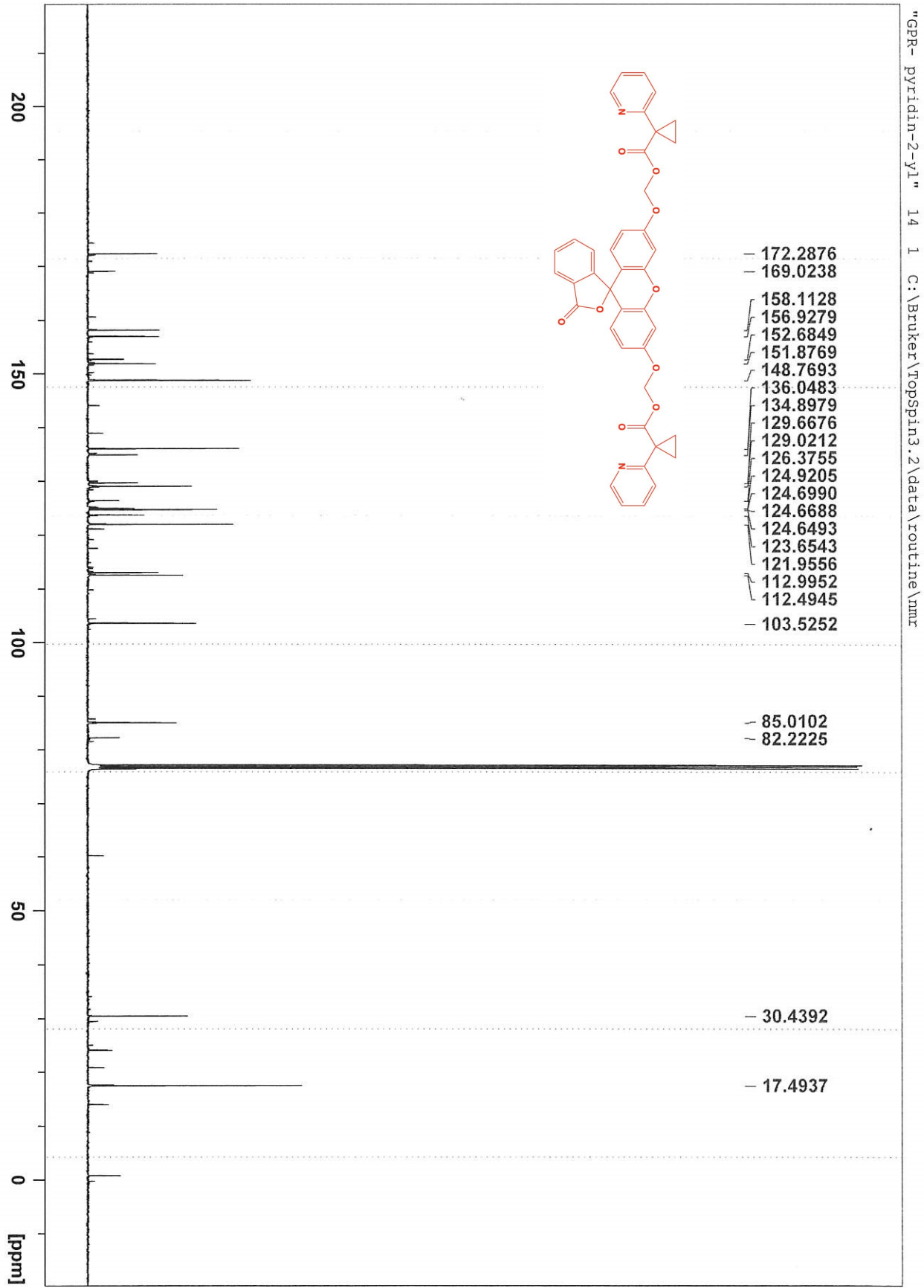
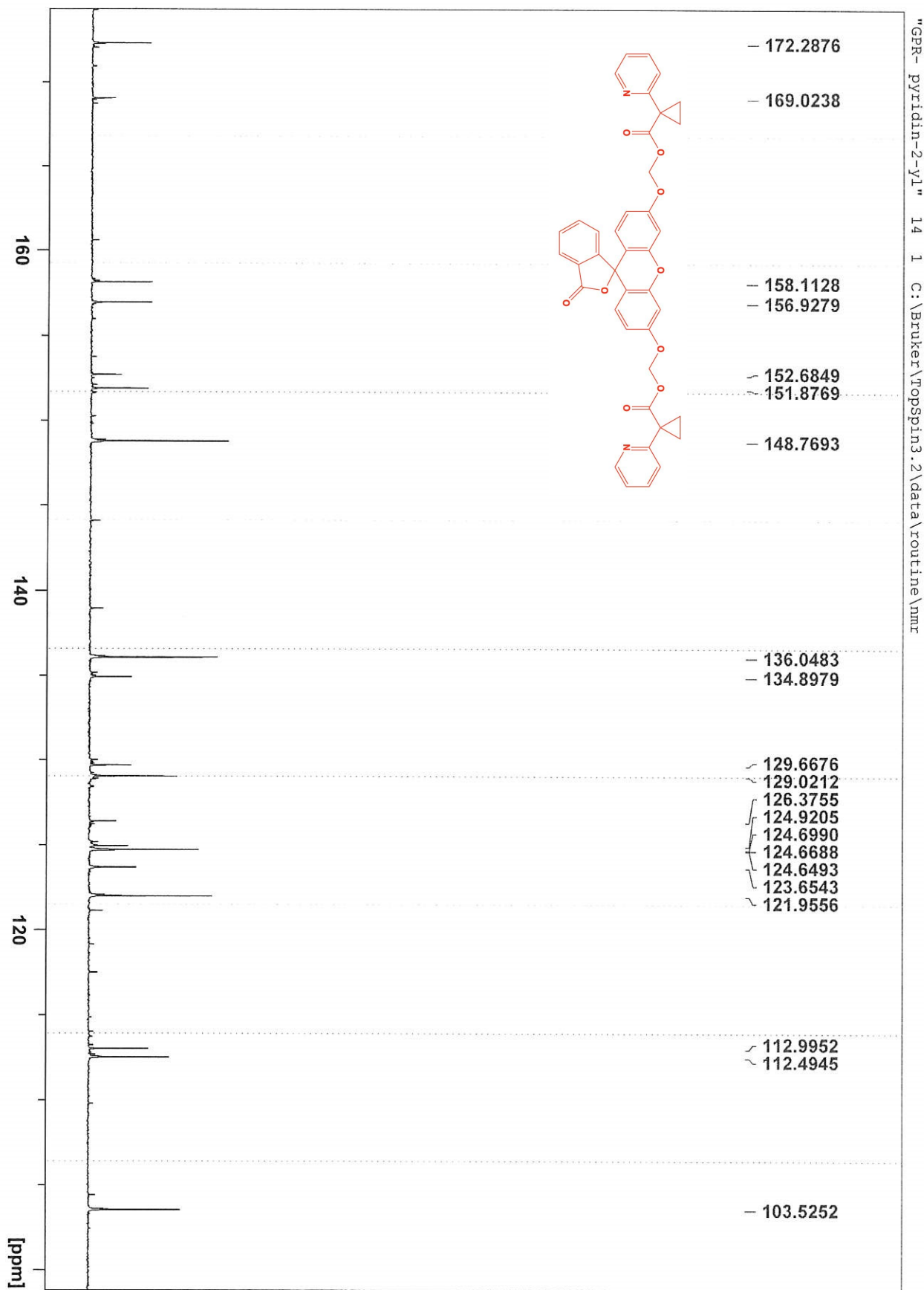
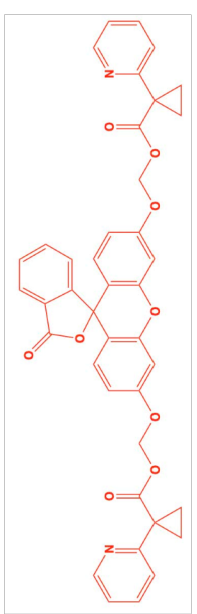


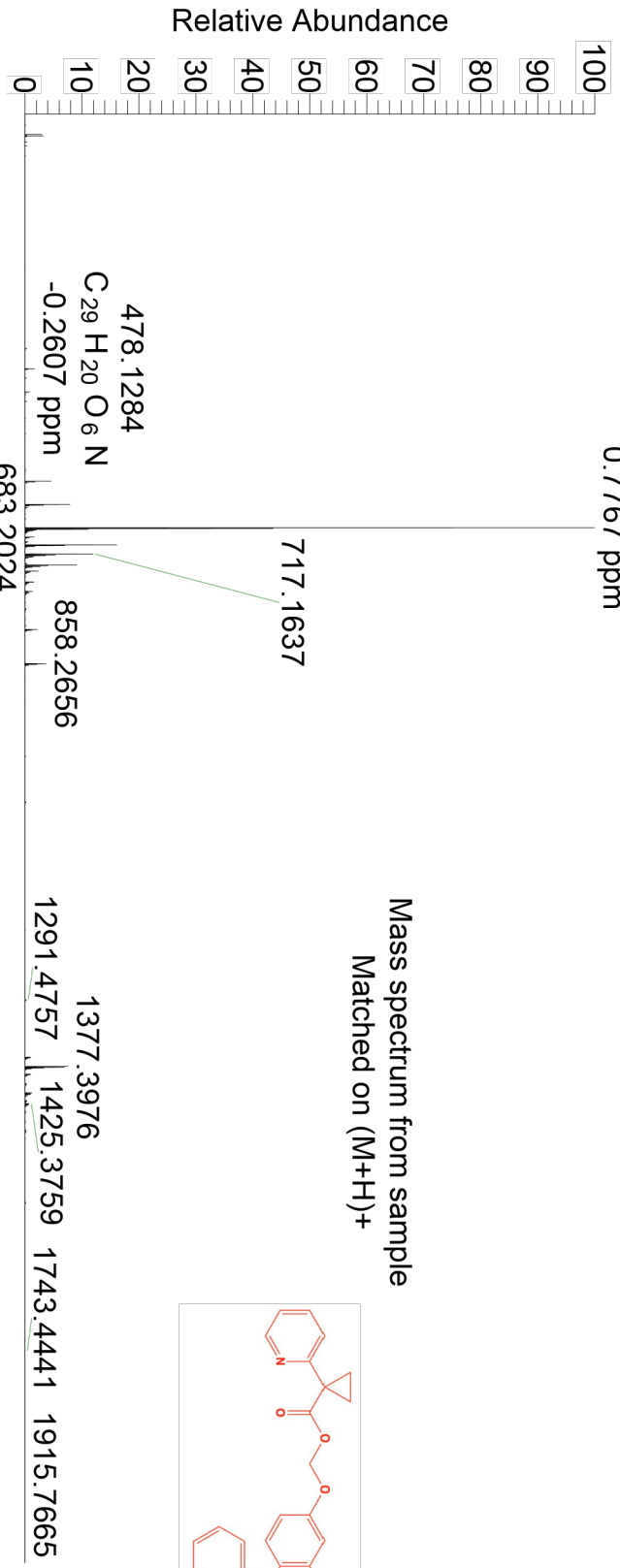
Figure S53



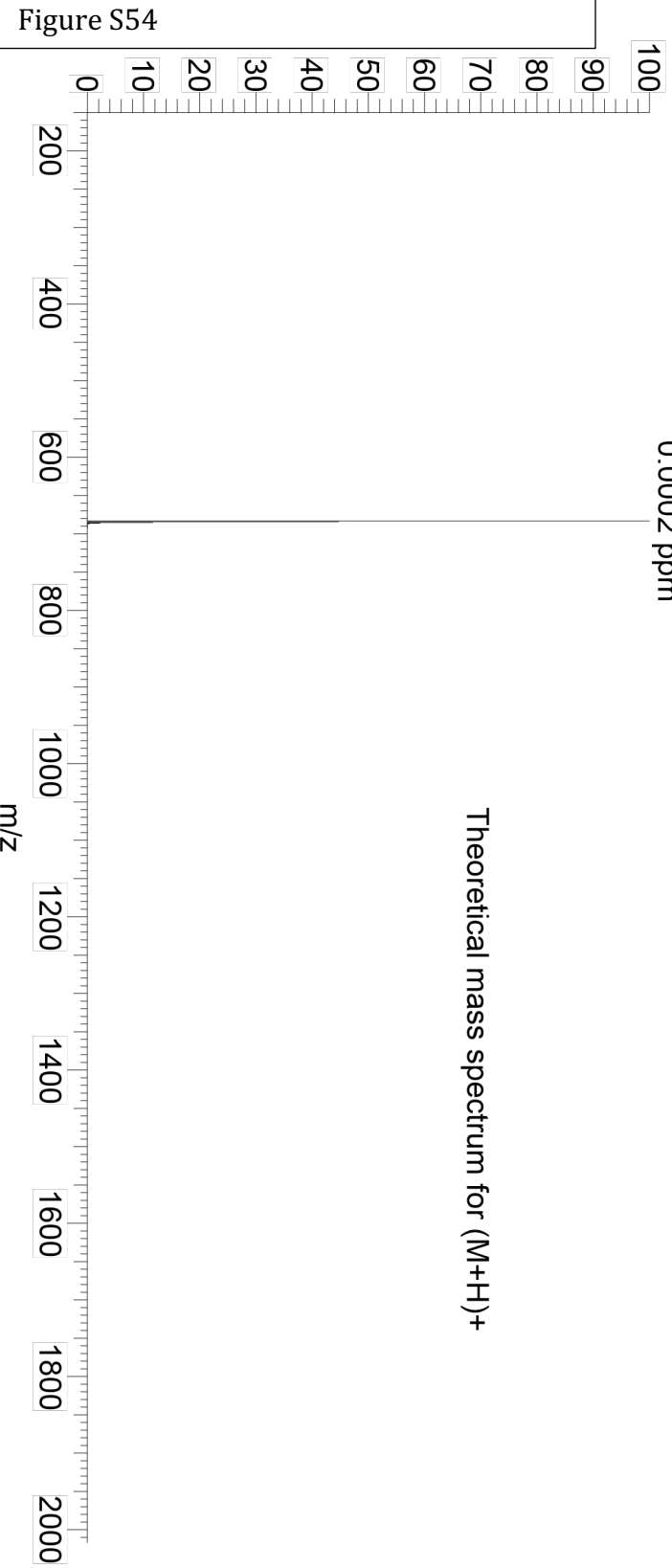
NL:
 6.08E7
 BU-2#9-20 RT: 0.15-0.34
 AV: 12 T: FTMS + PESI Full
 ms [150.00-2000.00]



Mass spectrum from sample
 Matched on (M+H)⁺



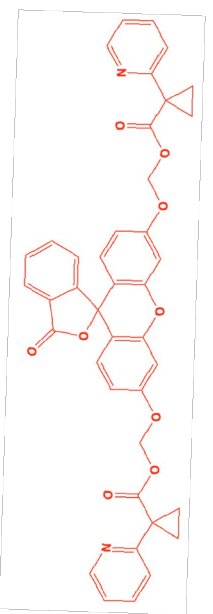
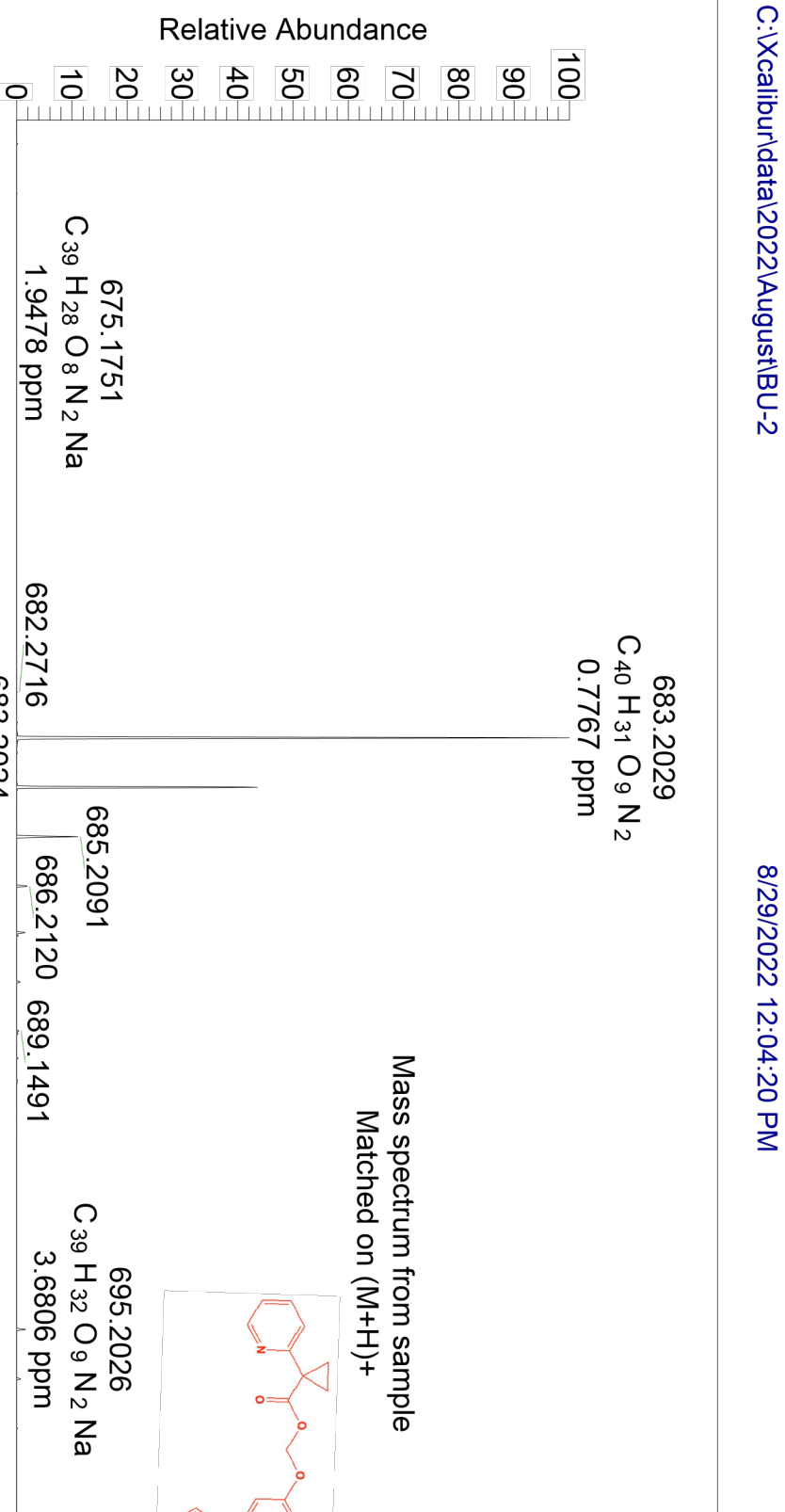
Theoretical mass spectrum for (M+H)⁺



NL:
 6.29E5
 C₄₀H₃₀N₂O₉H:
 C₄₀H₃₁N₂O₉
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res .Pwr. @FWHM

Figure S54

NL:
6.08E7
BU-2#9-20 RT: 0.19-0.34
AV: 12 T: FTMS + PESI Full
ms [150.00-2000.00]



NL:
6.29E5

C₄₀H₃₀N₂O₉H:
C₄₀H₃₁N₂O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr. @FWHM

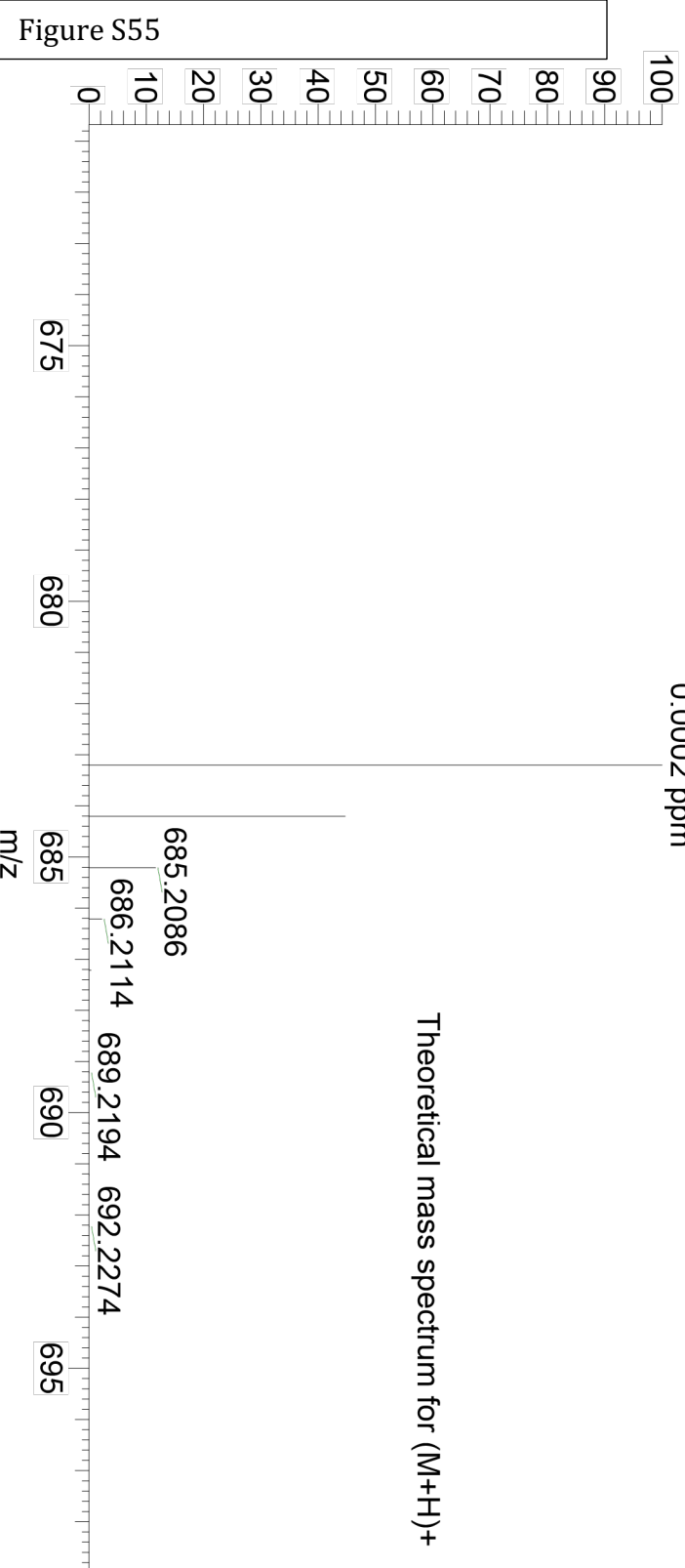


Figure S55

Figure S56

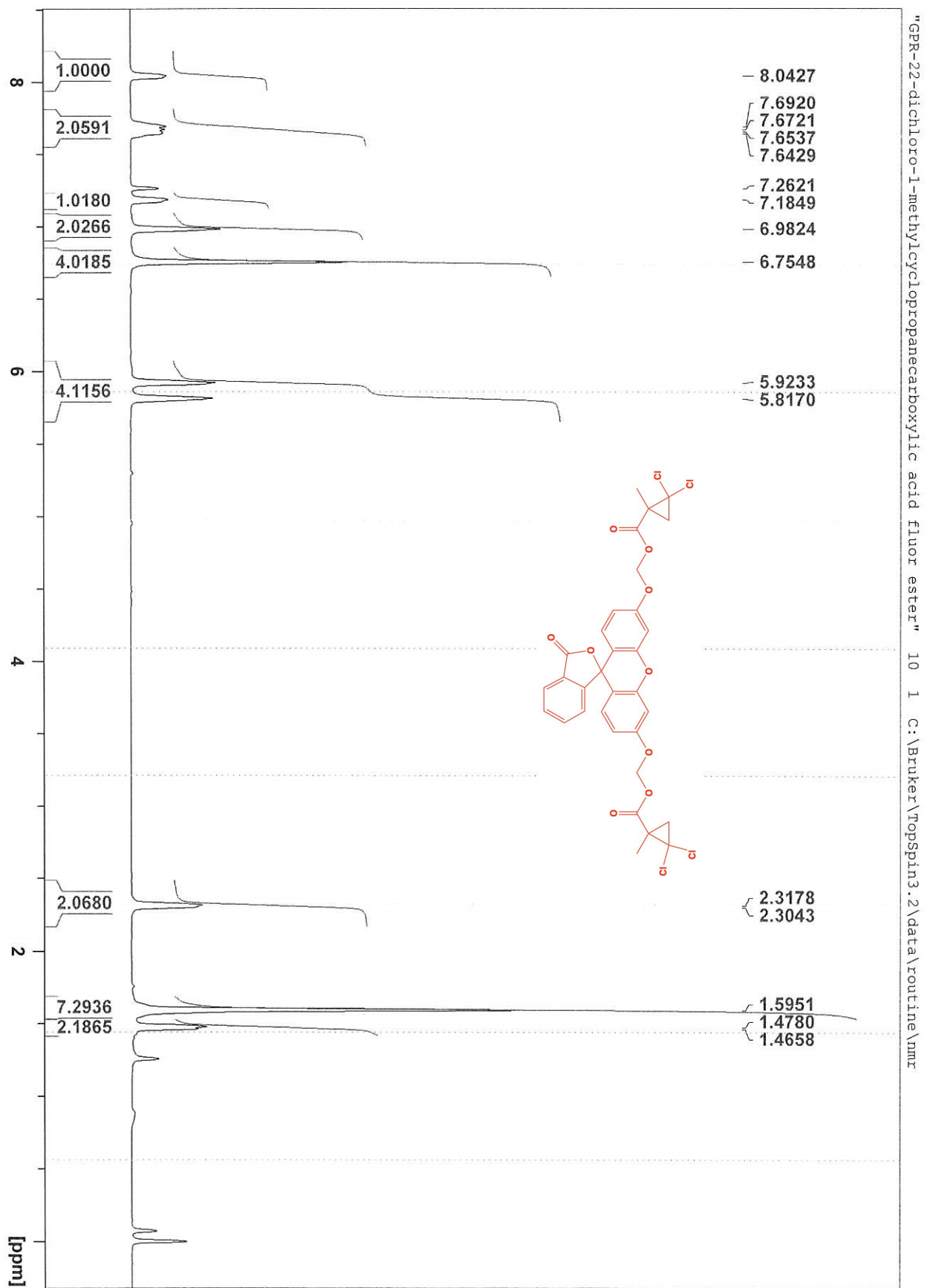


Figure S57

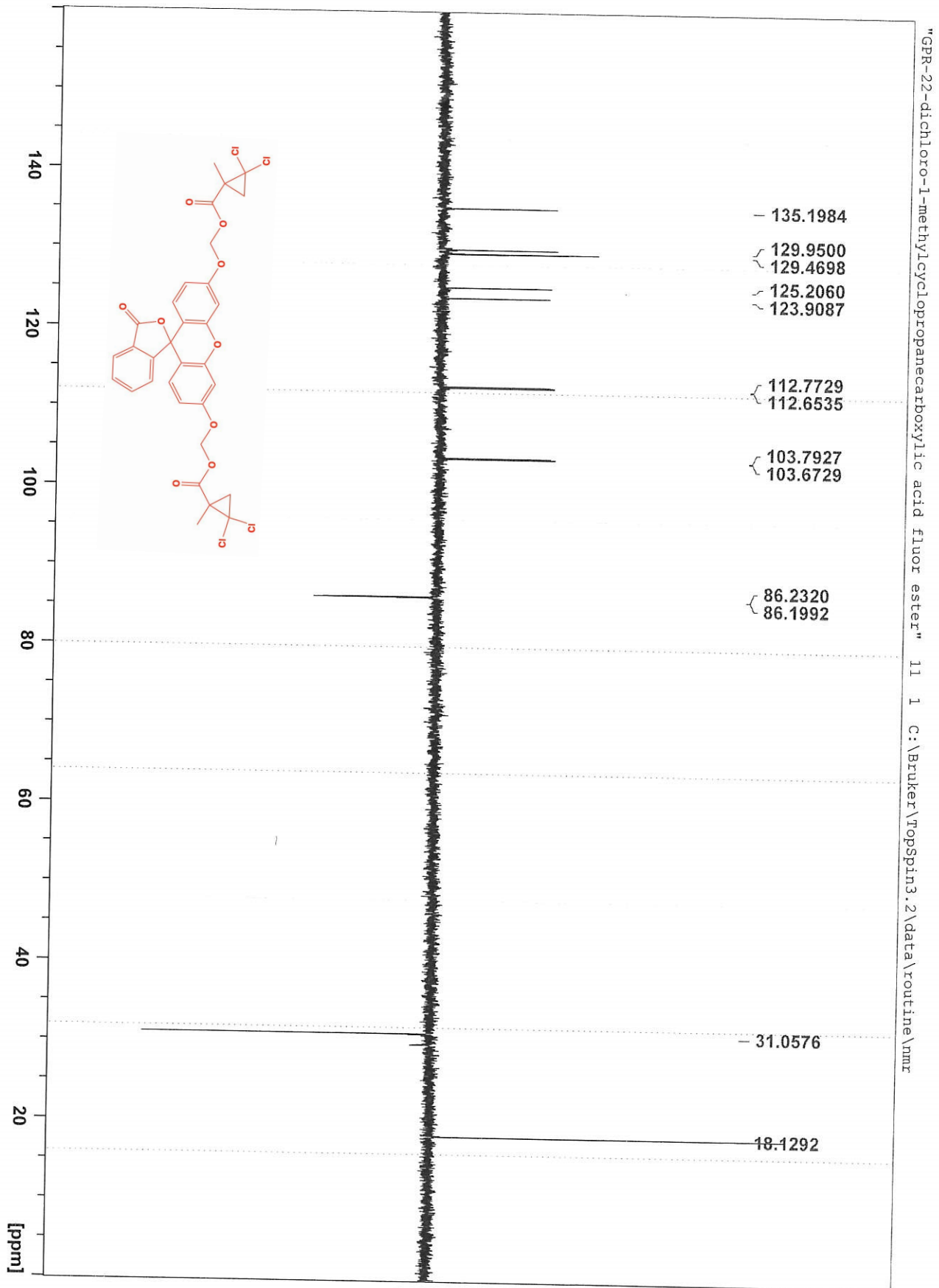


Figure S58

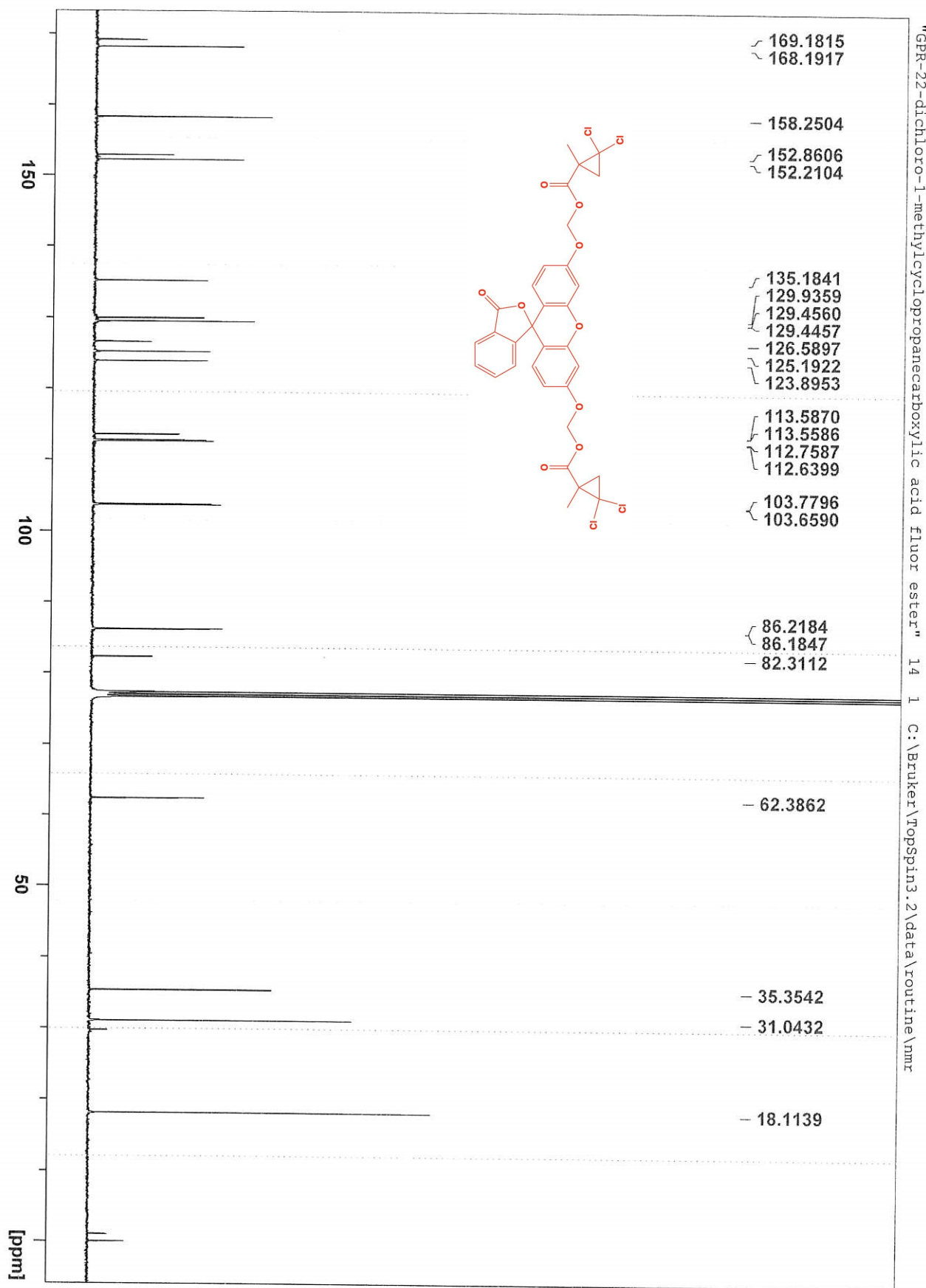


Figure S59

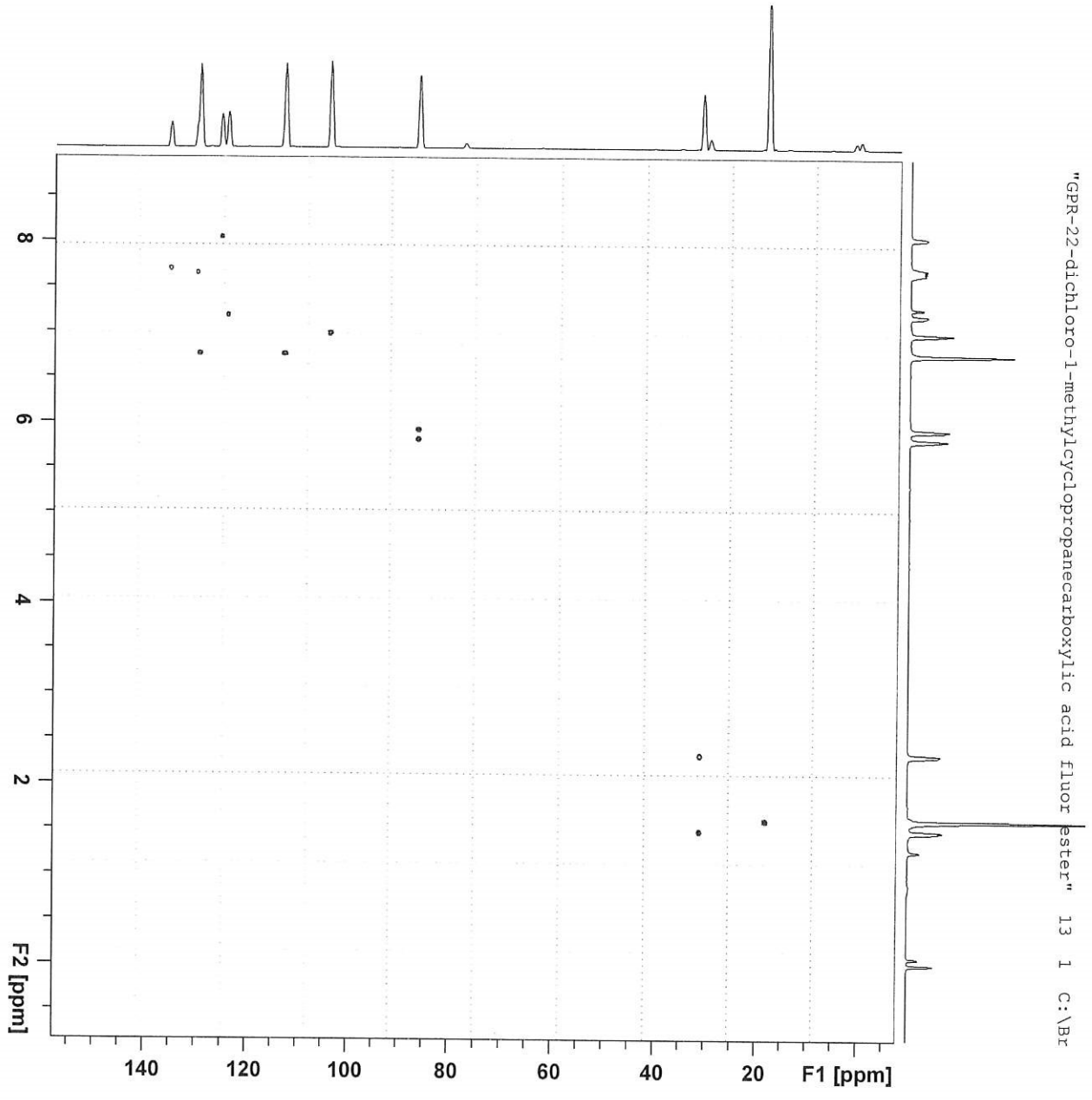
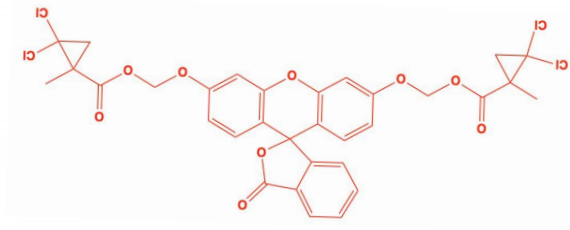
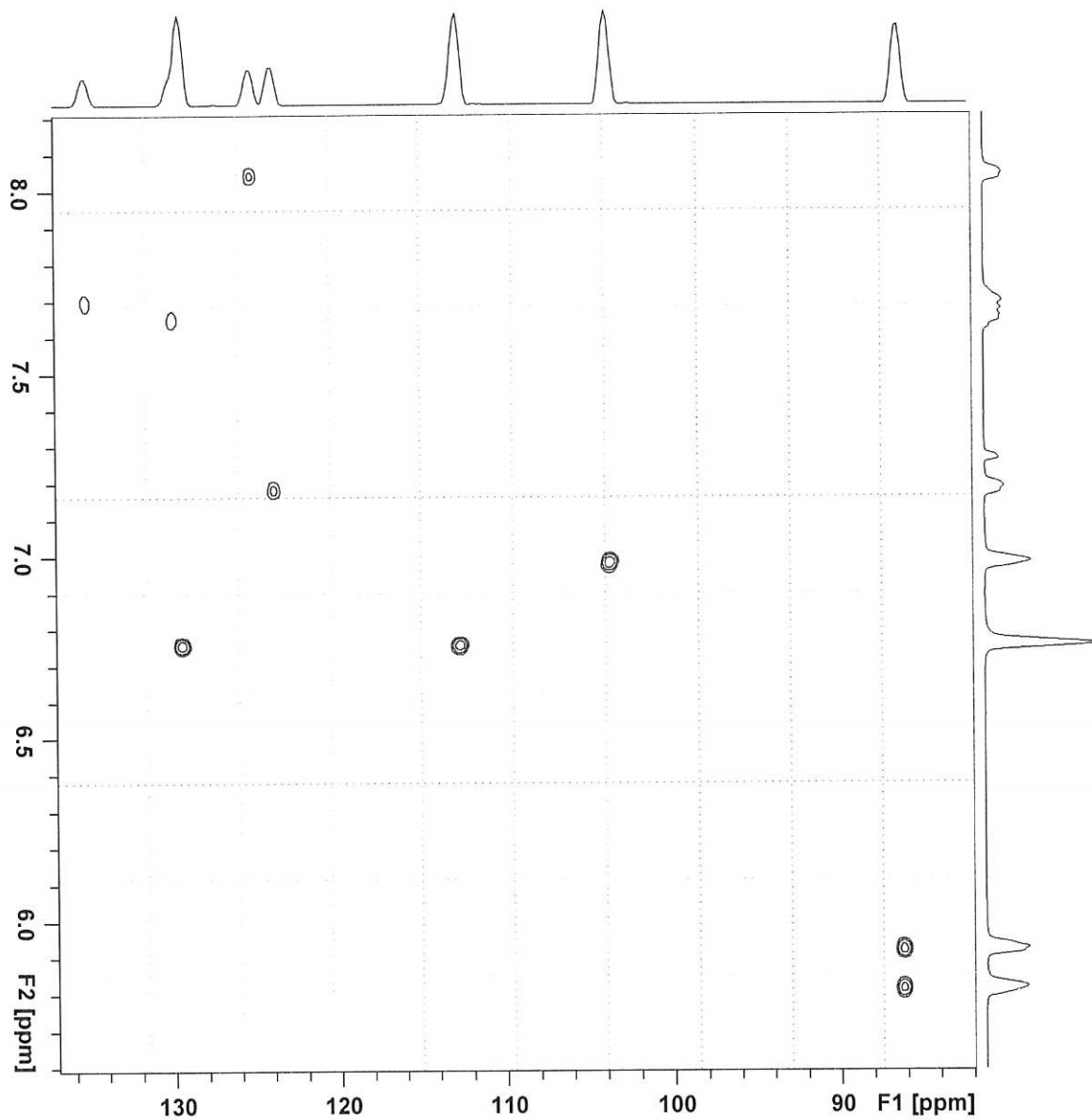
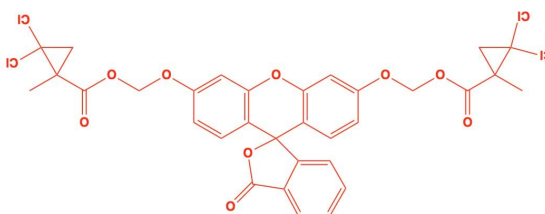
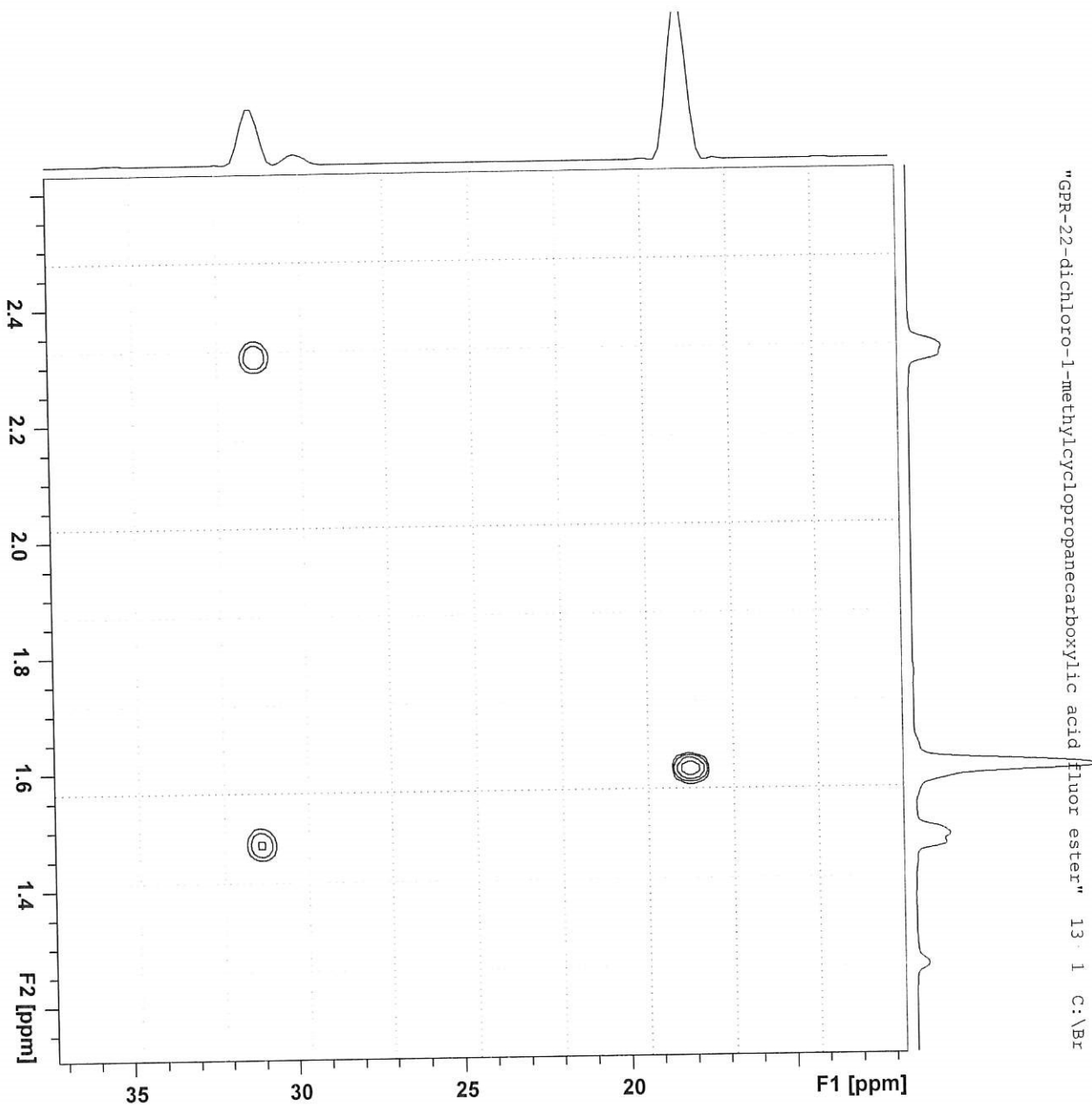


Figure S60

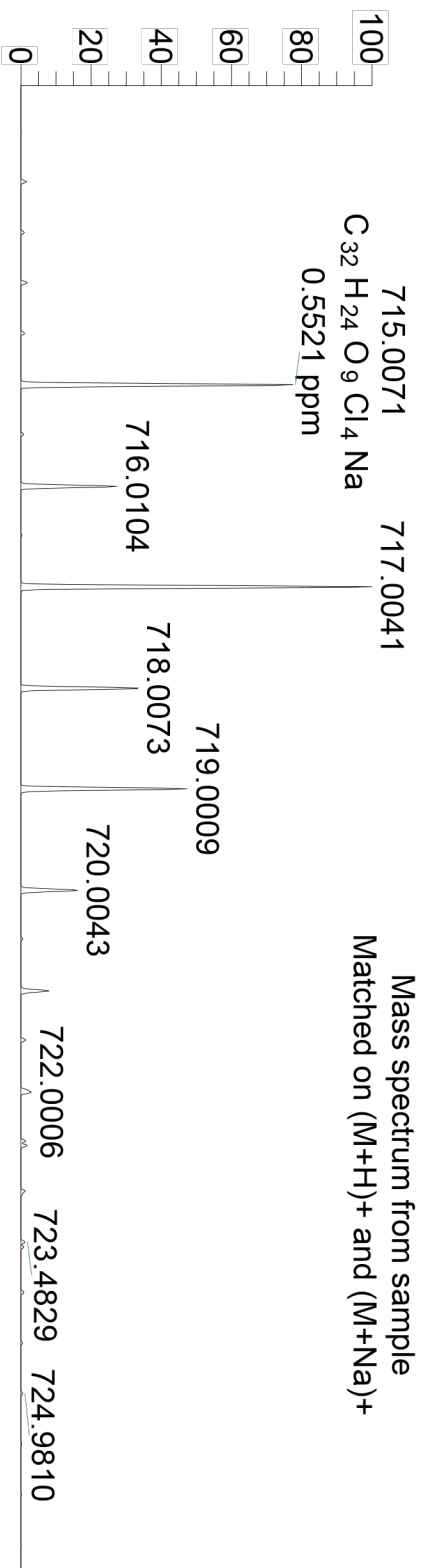


"GPR-22-dichloro-1-methylcyclopropanecarboxylic acid fluor ester" 13 C: \Br

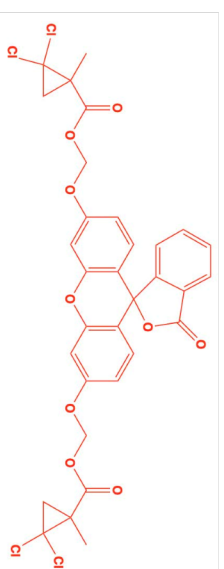
Figure S61



"GPR-22-dichloro-1-methylcyclopropanecarboxylic acid fluor ester" 13 C: Br

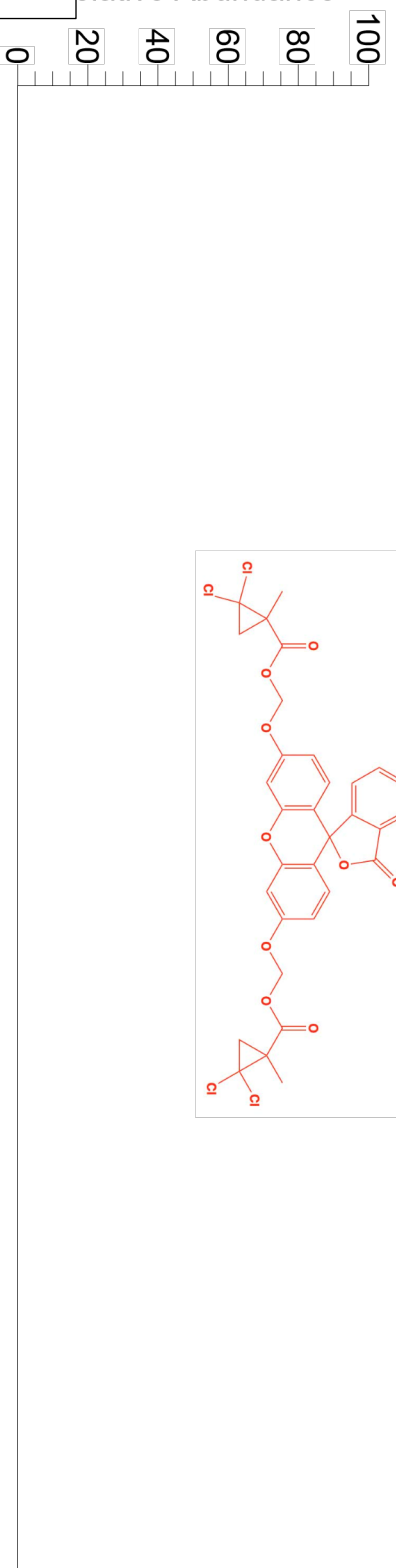


NL:
2.64E6
BU-6#11-18 RT: 0.18-0.30
AV: 8 T: FTMS + p ESI Full
ms [150.00-2000.00]



NL:
0
C₃₂ H₂₄ Cl₄ O₉ H:
C₃₂ H₂₅ Cl₄ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

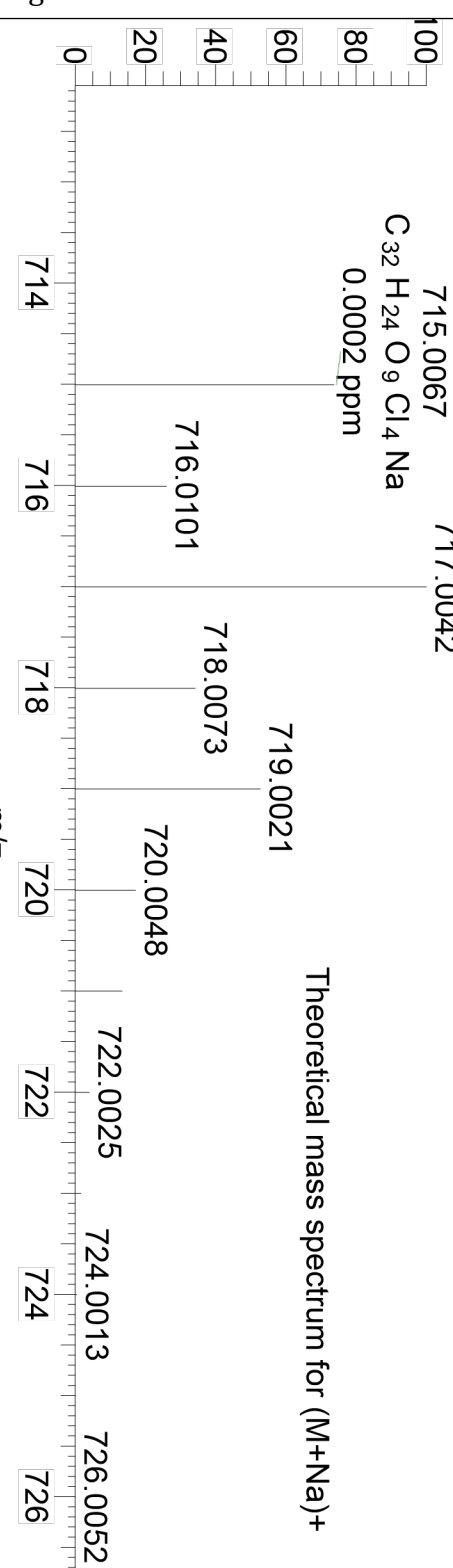
relative Abundance



NL:
3.09E5

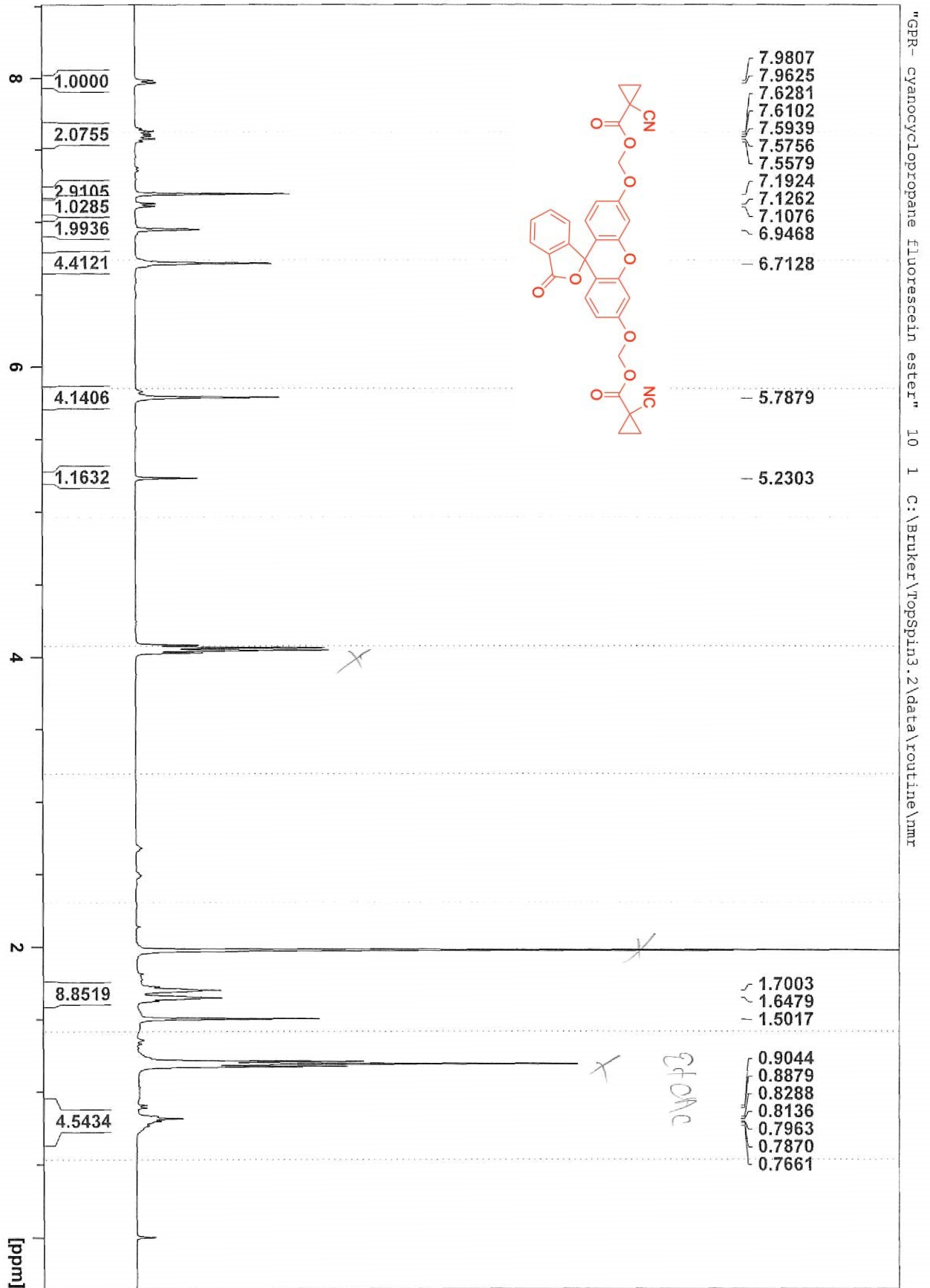
C₃₂ H₂₄ Cl₄ O₉ Na:
C₃₂ H₂₄ Cl₄ O₉ Na¹
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Figure S62



m/z

Figure S63



GPR- cyanocyclopropane Fluorescein ester

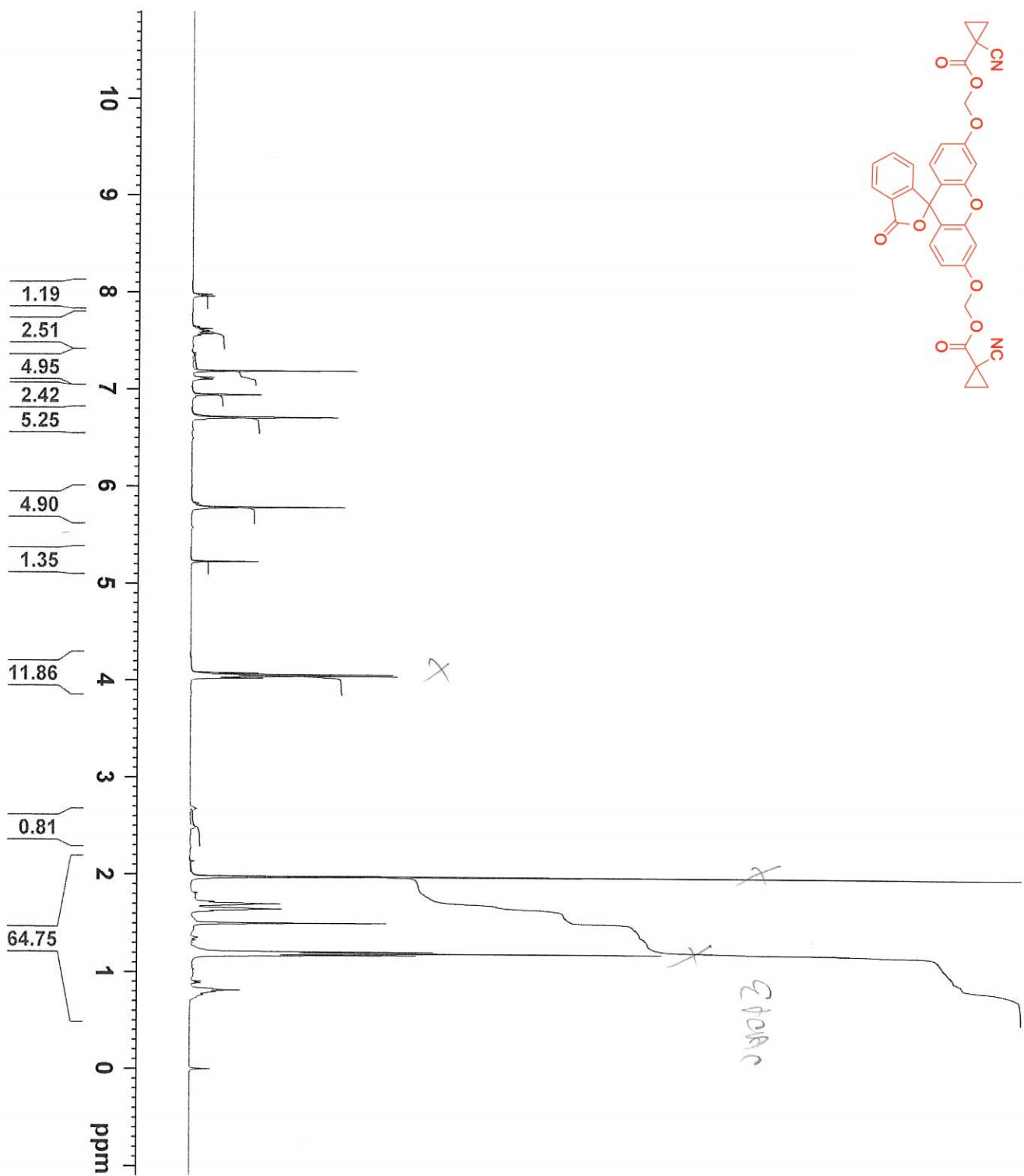
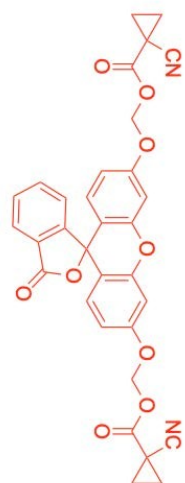


Figure S64

Figure S65

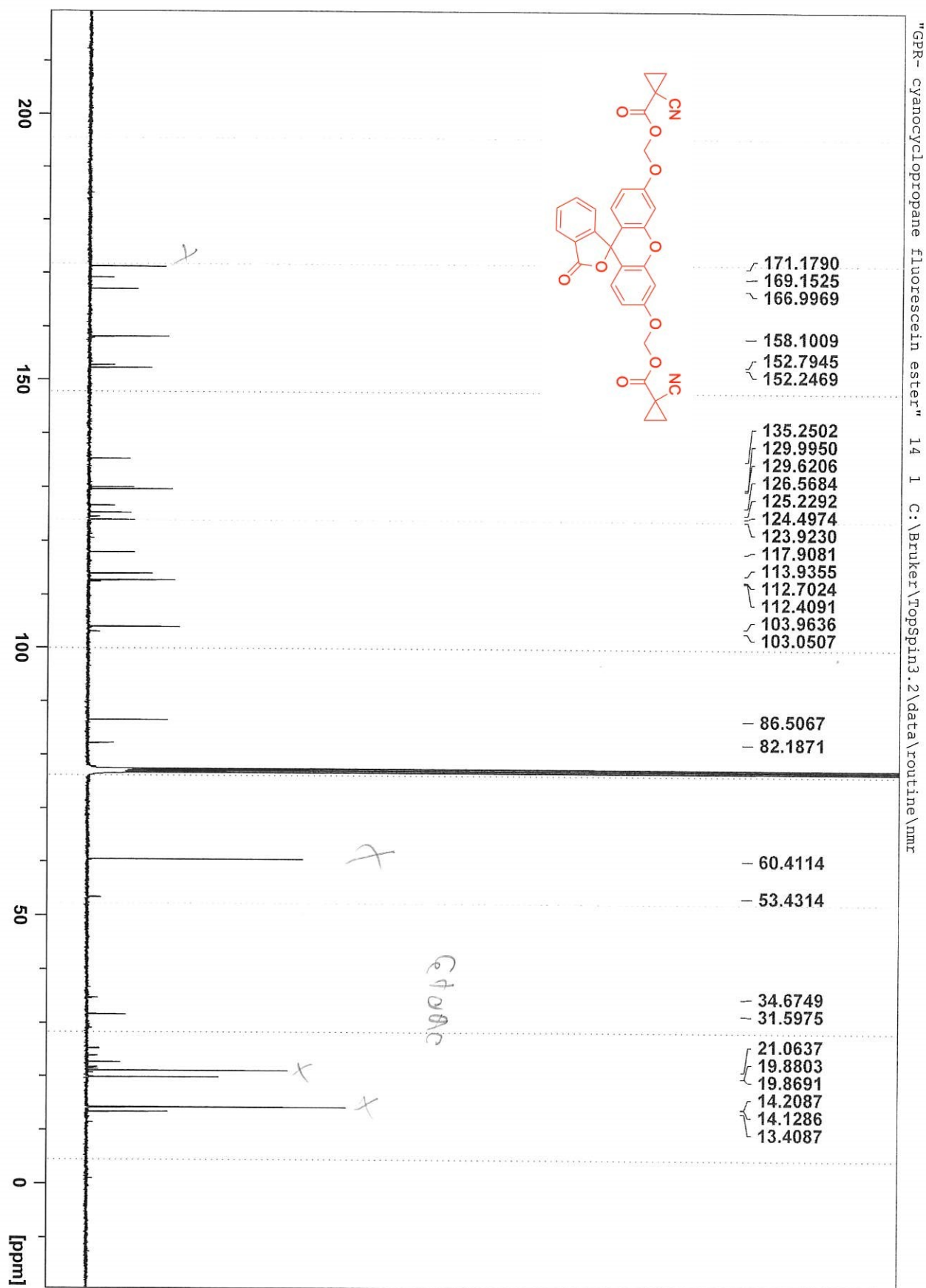


Figure S66

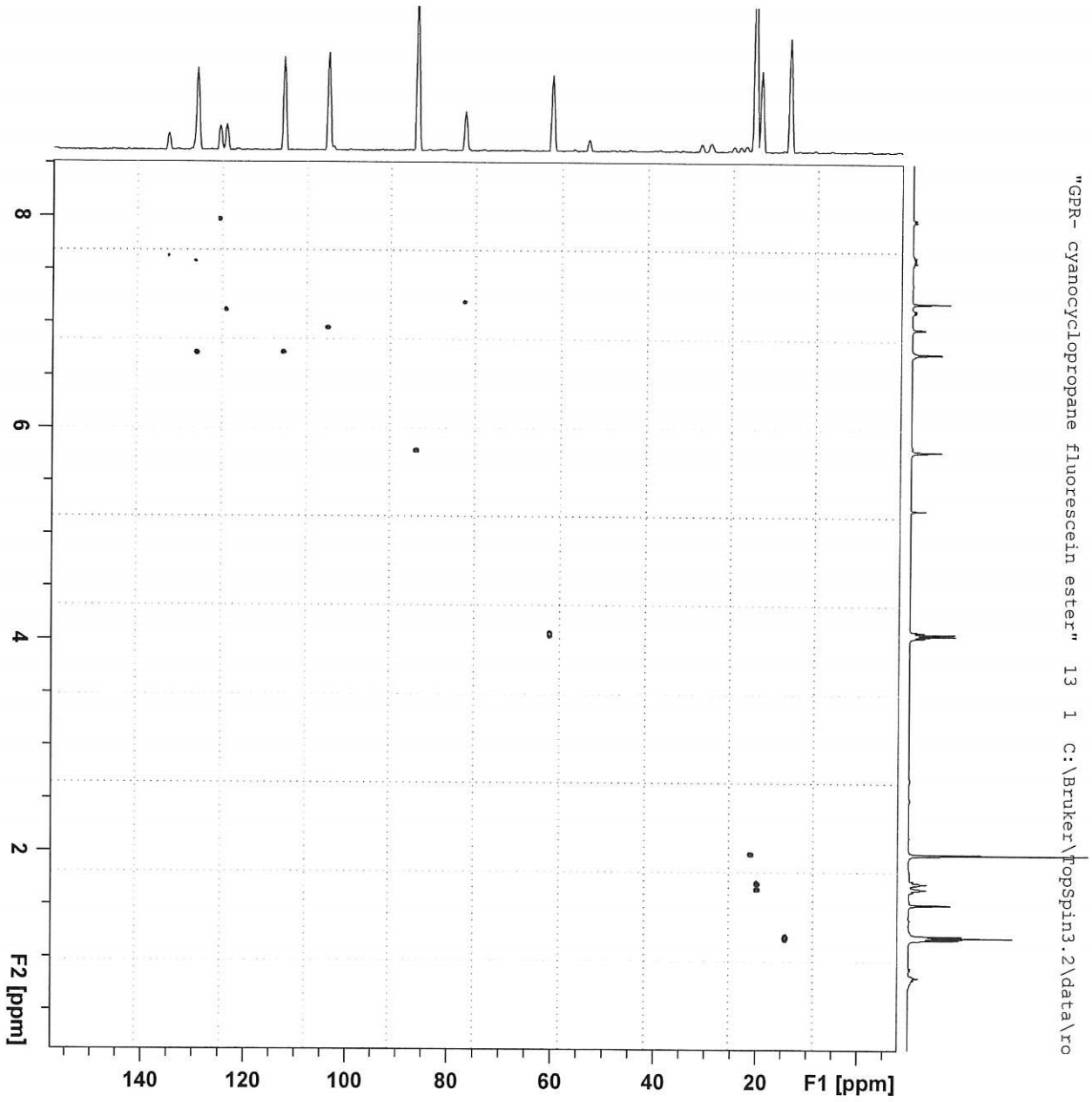
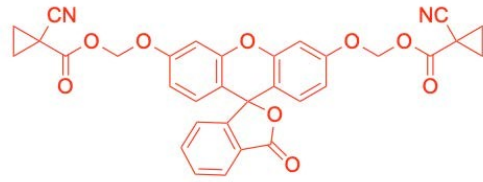
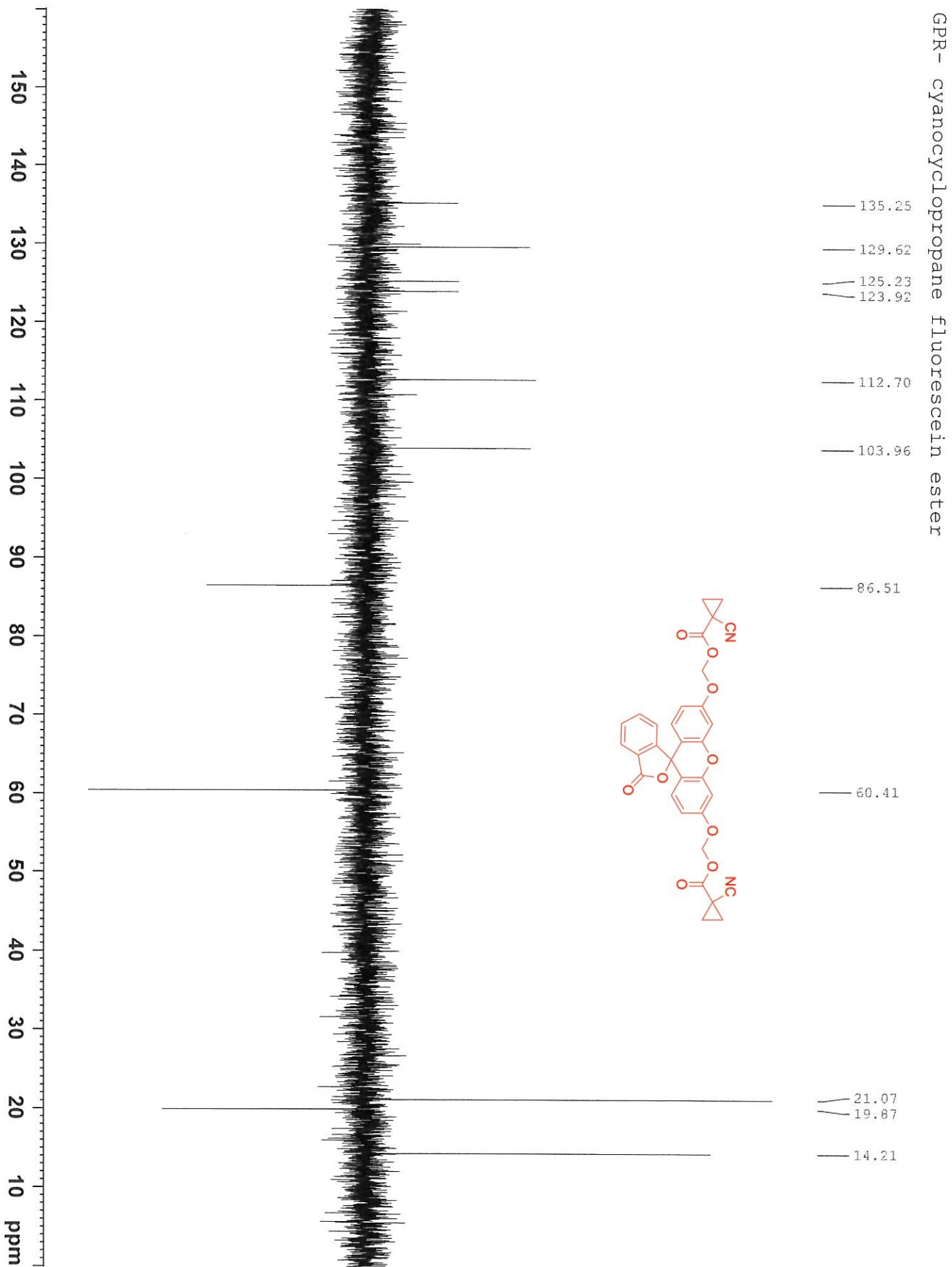
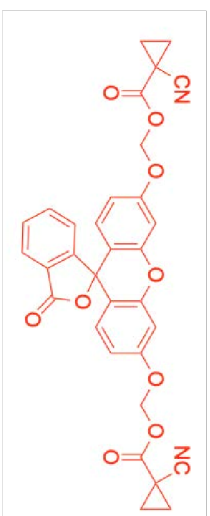
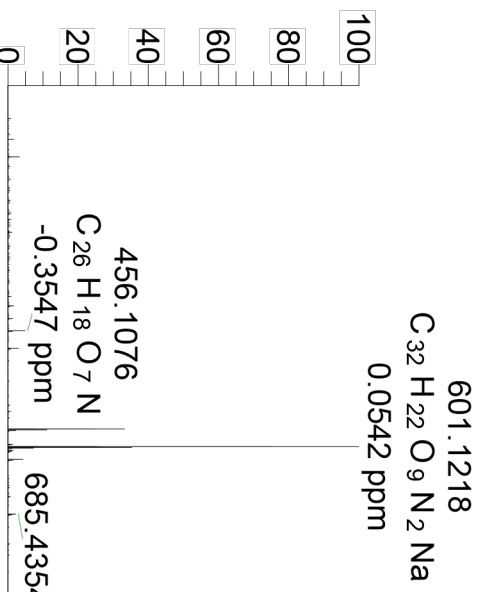


Figure S67

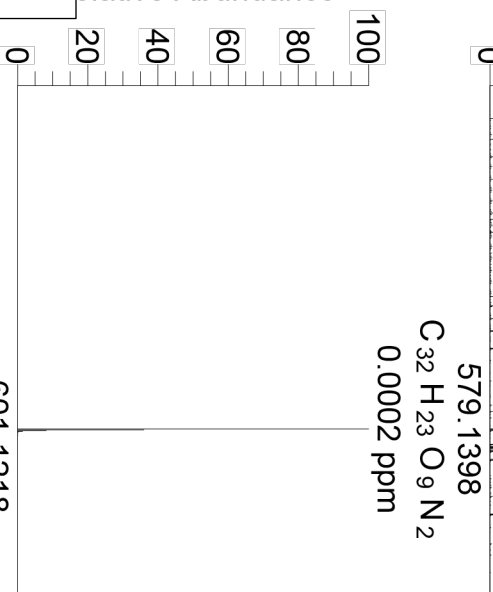




NL:
1.23E7
BU-10#10-15 RT: 0016-0.25
AV: 6 T: FTMS + p ESI Full
ms [150.00-2000.00]

NL:
6.86E5

C₃₂ H₂₂ N₂ O₉ H:
C₃₂ H₂₃ N₂ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM



Theoretical mass spectrum for (M+H)+

NL:
6.87E5

C₃₂ H₂₂ N₂ O₉ Na:
C₃₂ H₂₂ N₂ O₉ Na¹
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+Na)+

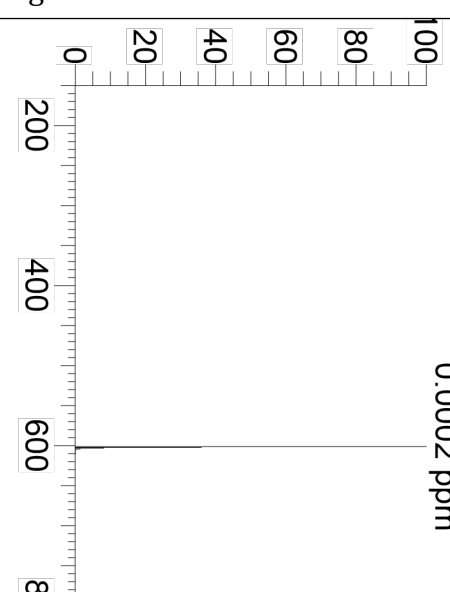


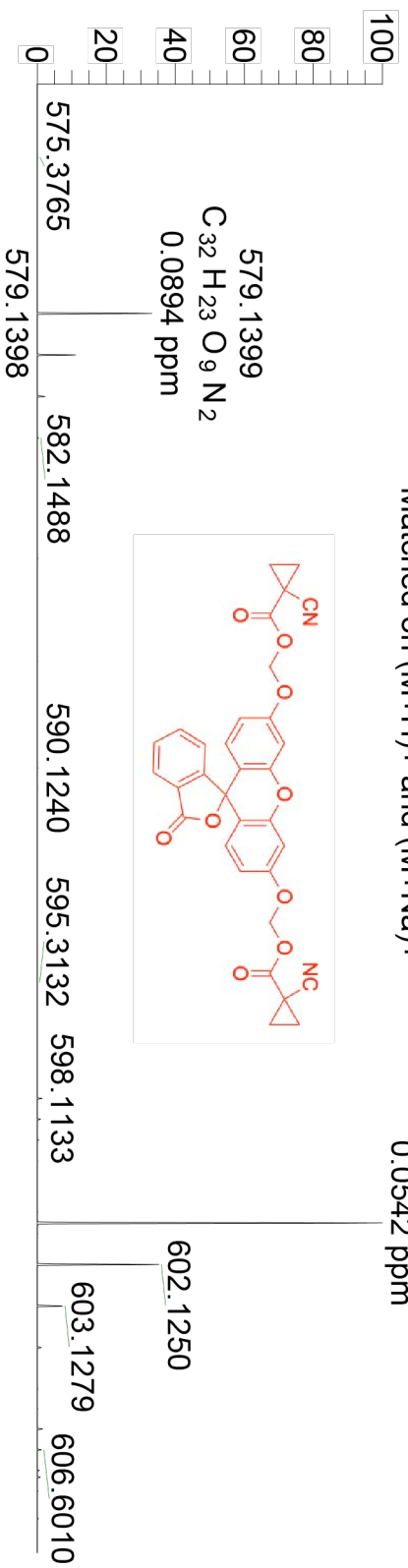
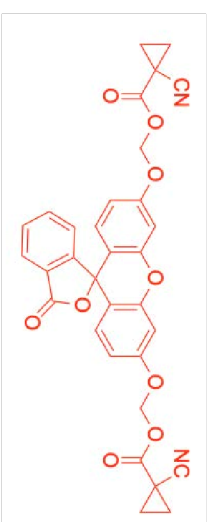
Figure S68

m/z

Mass spectrum from sample
Matched on (M+H)⁺ and (M+Na)⁺

601.1218
C₃₂H₂₂O₉N₂Na
0.0542 ppm

NL:
1.23E7
BU-10#10-15 RT: 0:16-0:25
AV: 6 T: FTMS + p ESI Full
ms [150.00-2000.00]



NL:
6.86E5

C₃₂H₂₂N₂O₉H:
C₃₂H₂₃N₂O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+H)⁺



NL:
6.87E5

C₃₂H₂₂N₂O₉Na:
C₃₂H₂₂N₂O₉Na¹
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+Na)⁺

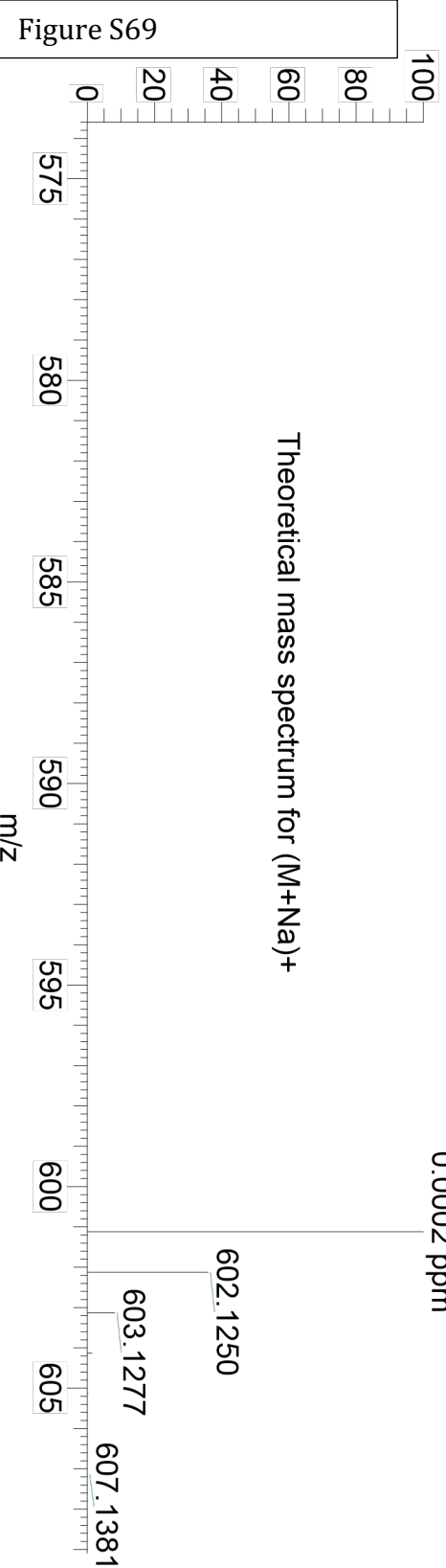


Figure S70

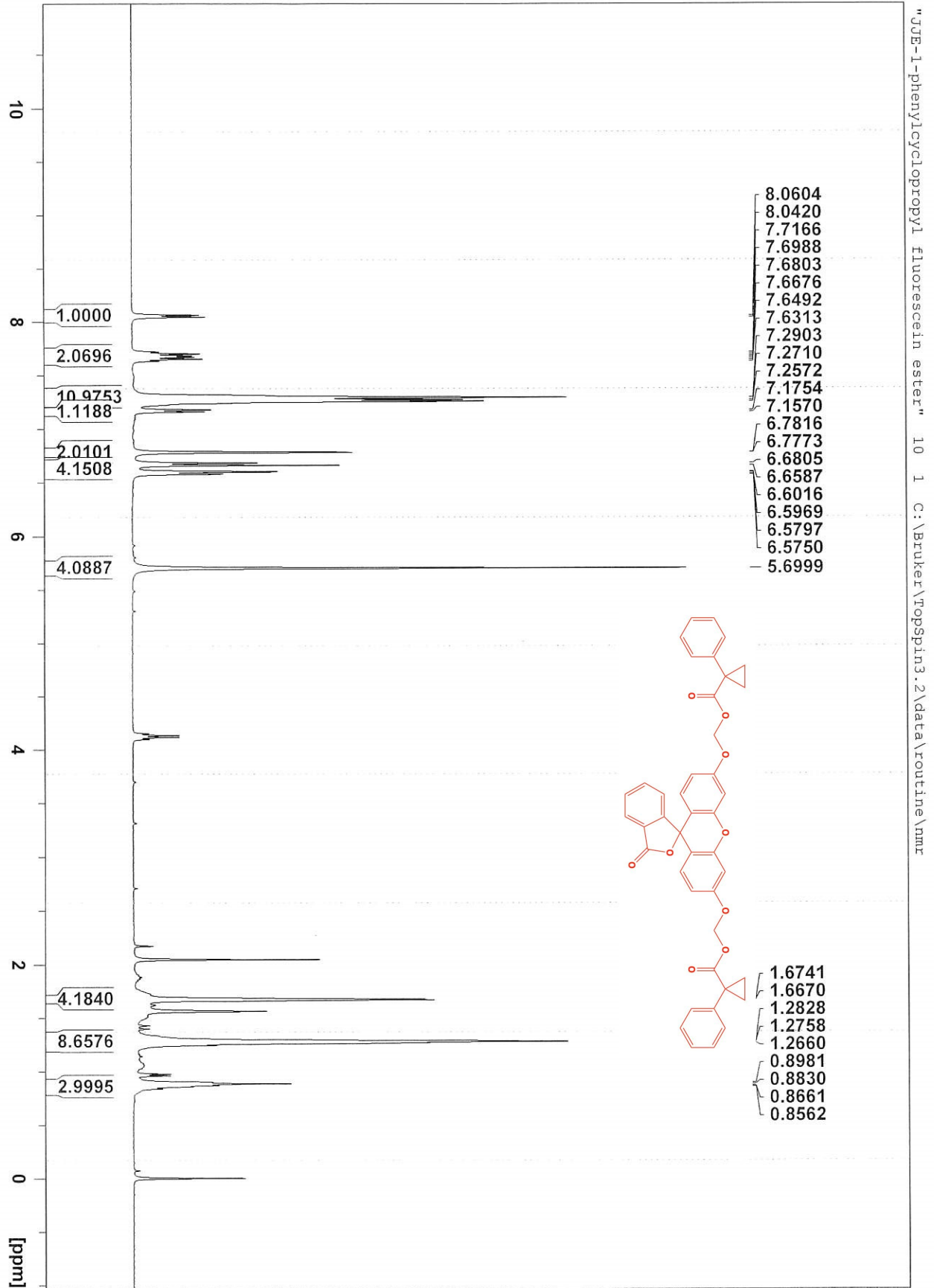


Figure S71

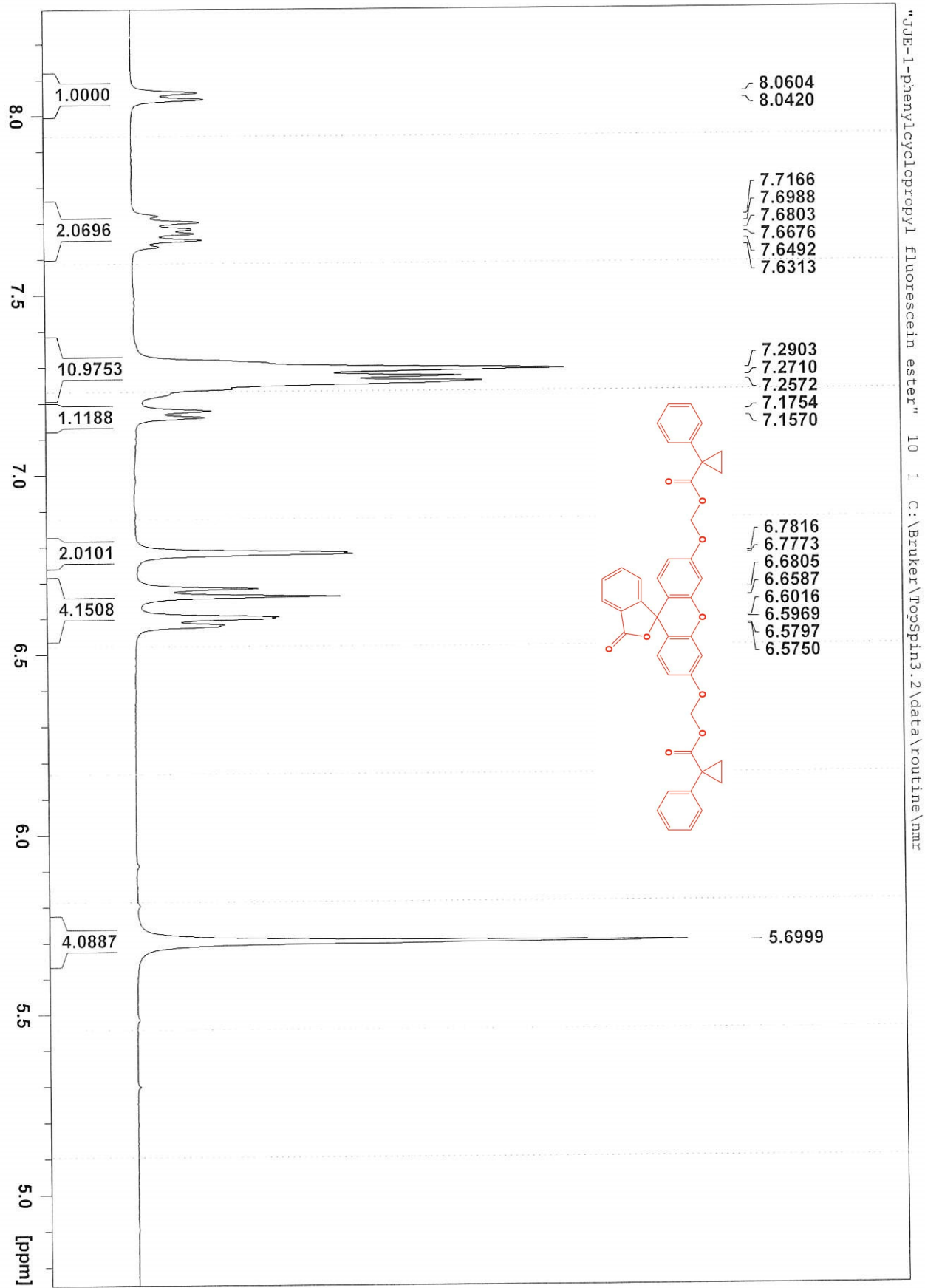


Figure S72

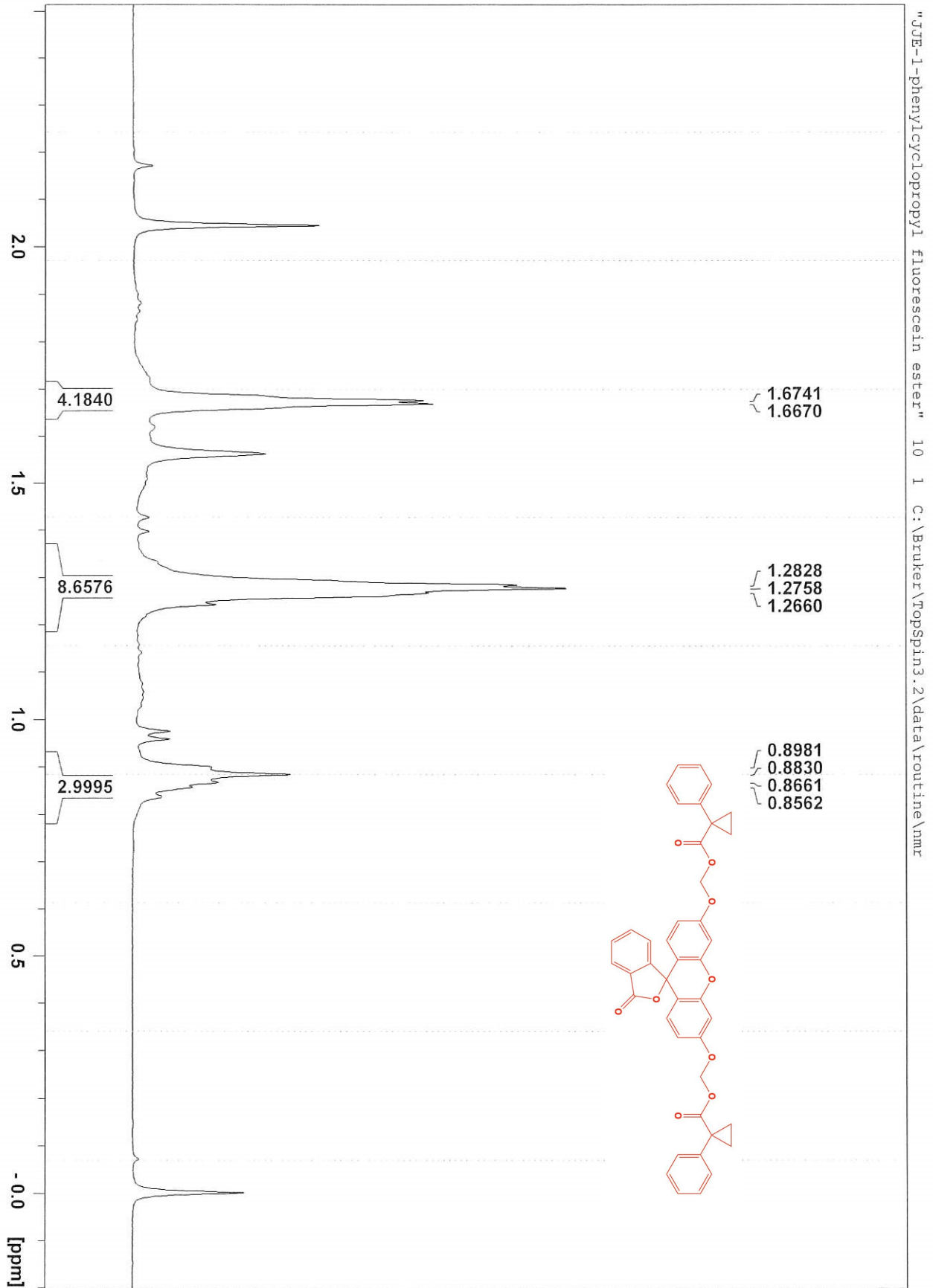


Figure S73

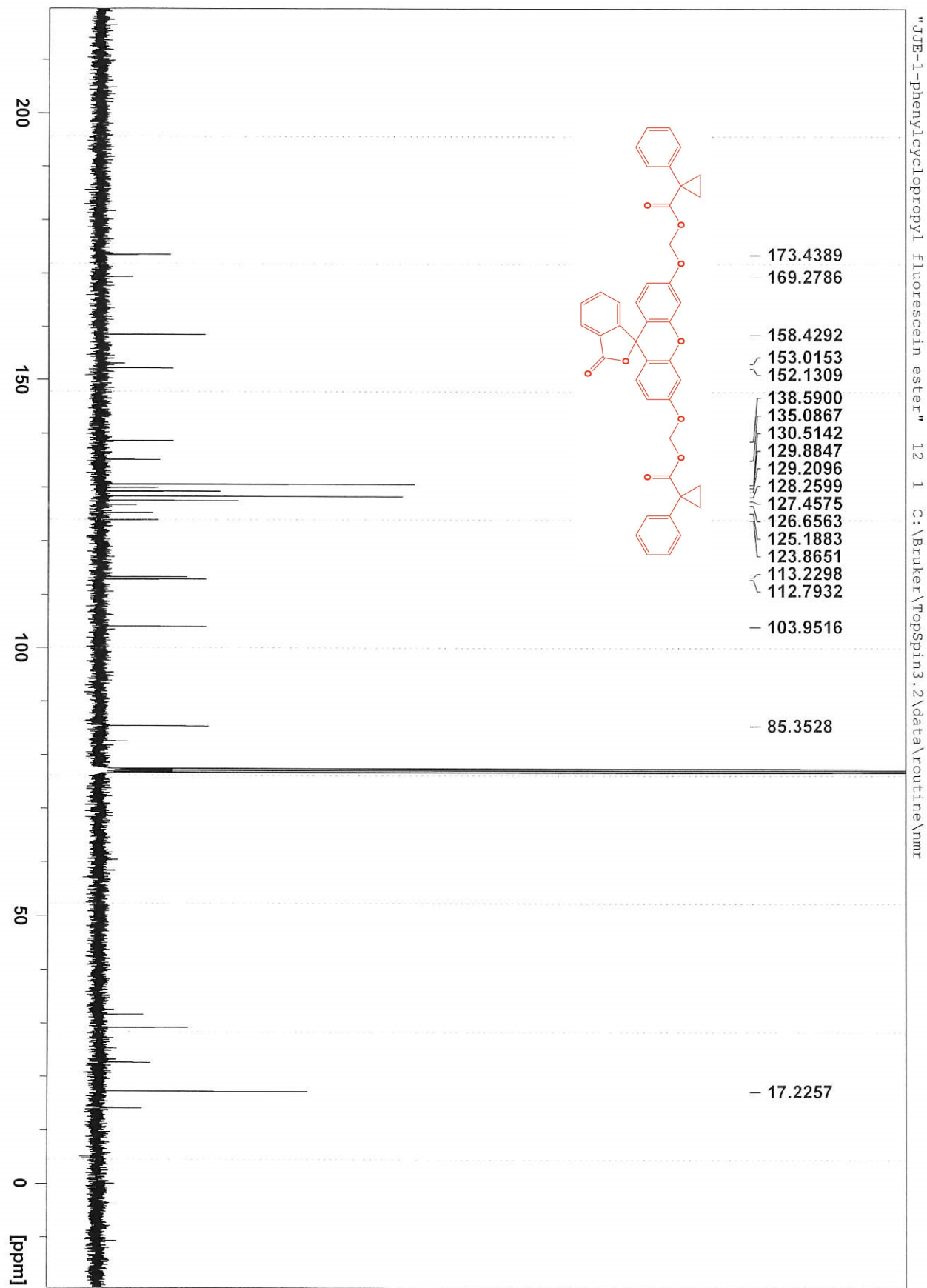


Figure S74

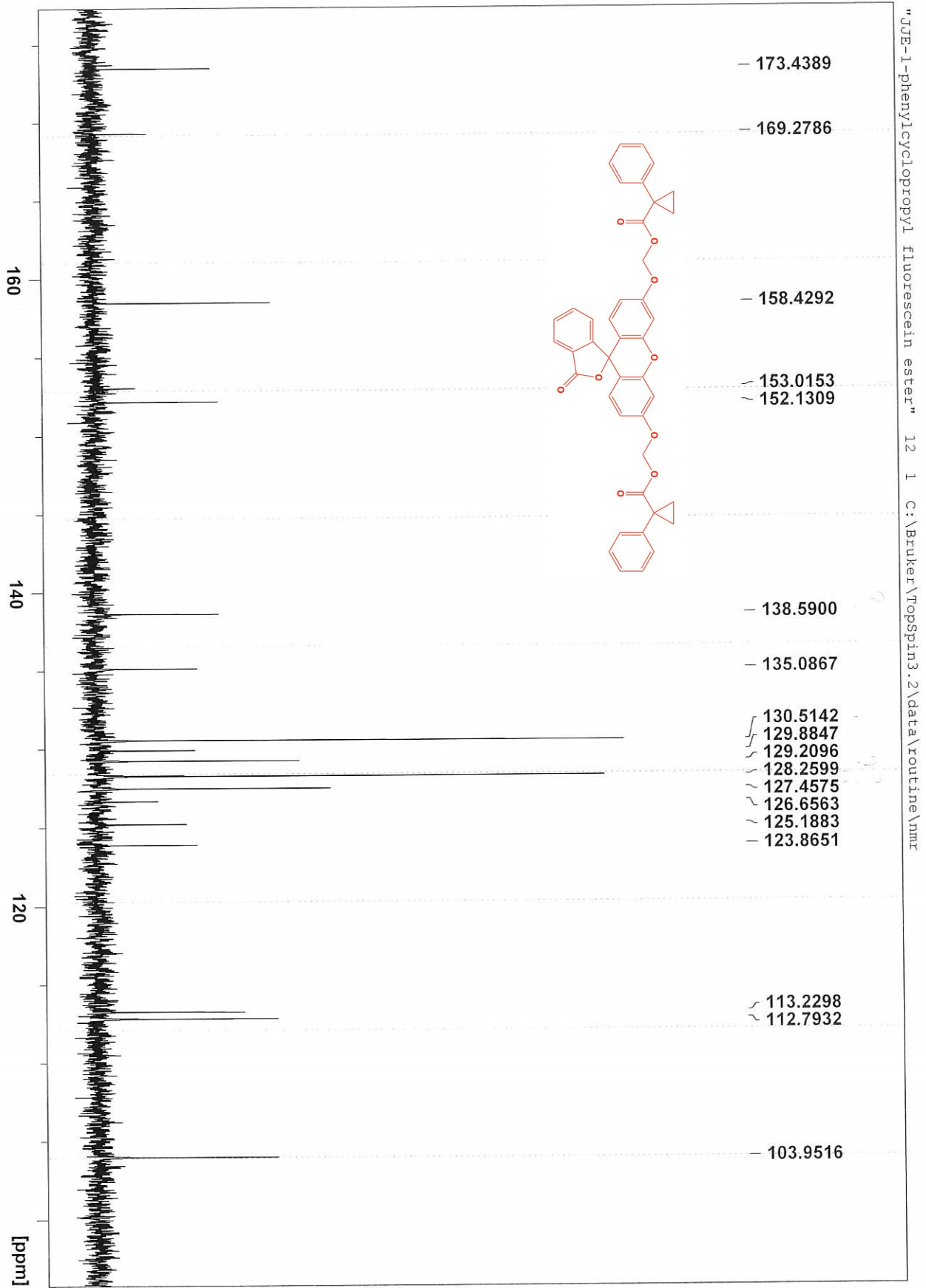


Figure S75

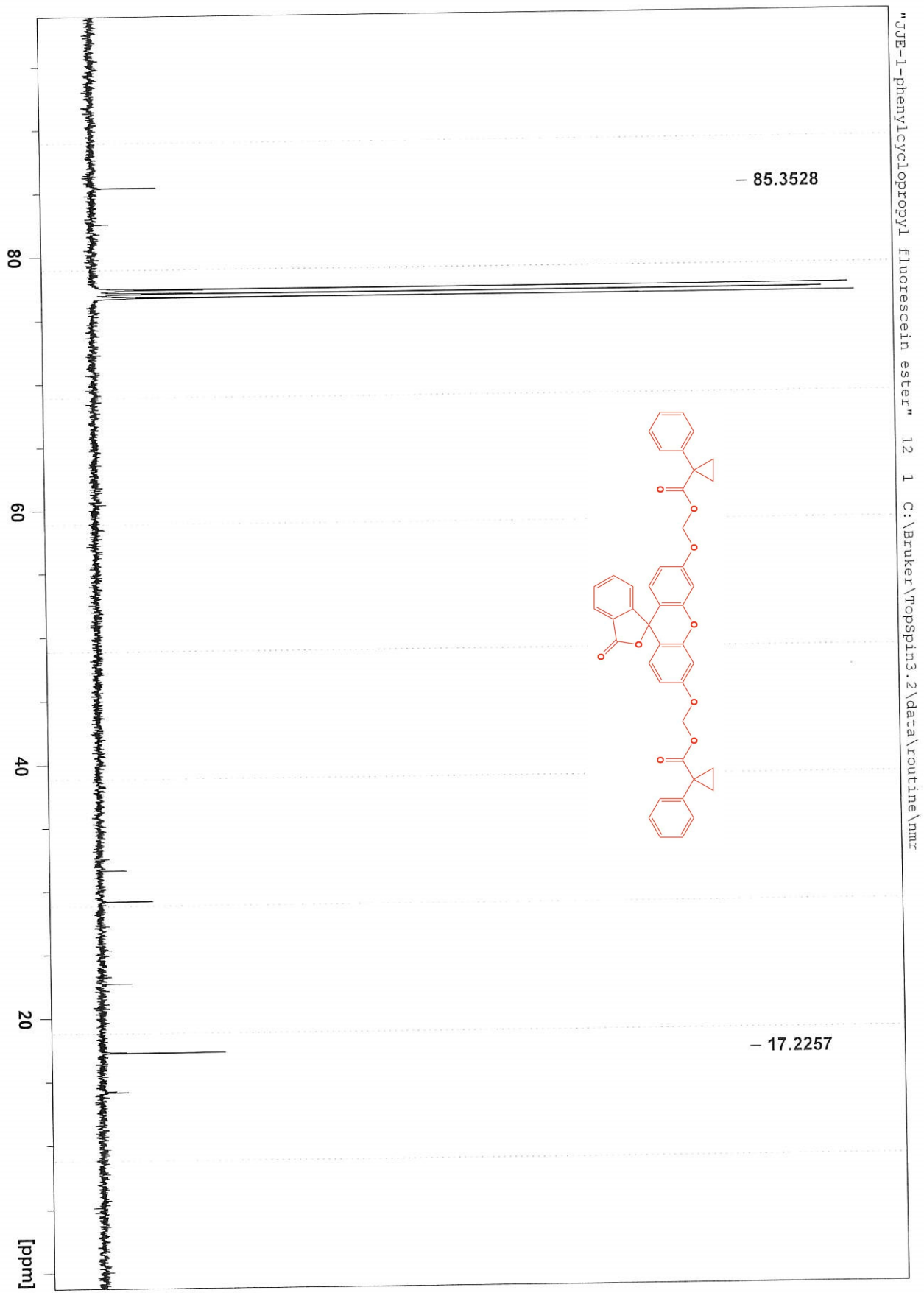


Figure S76

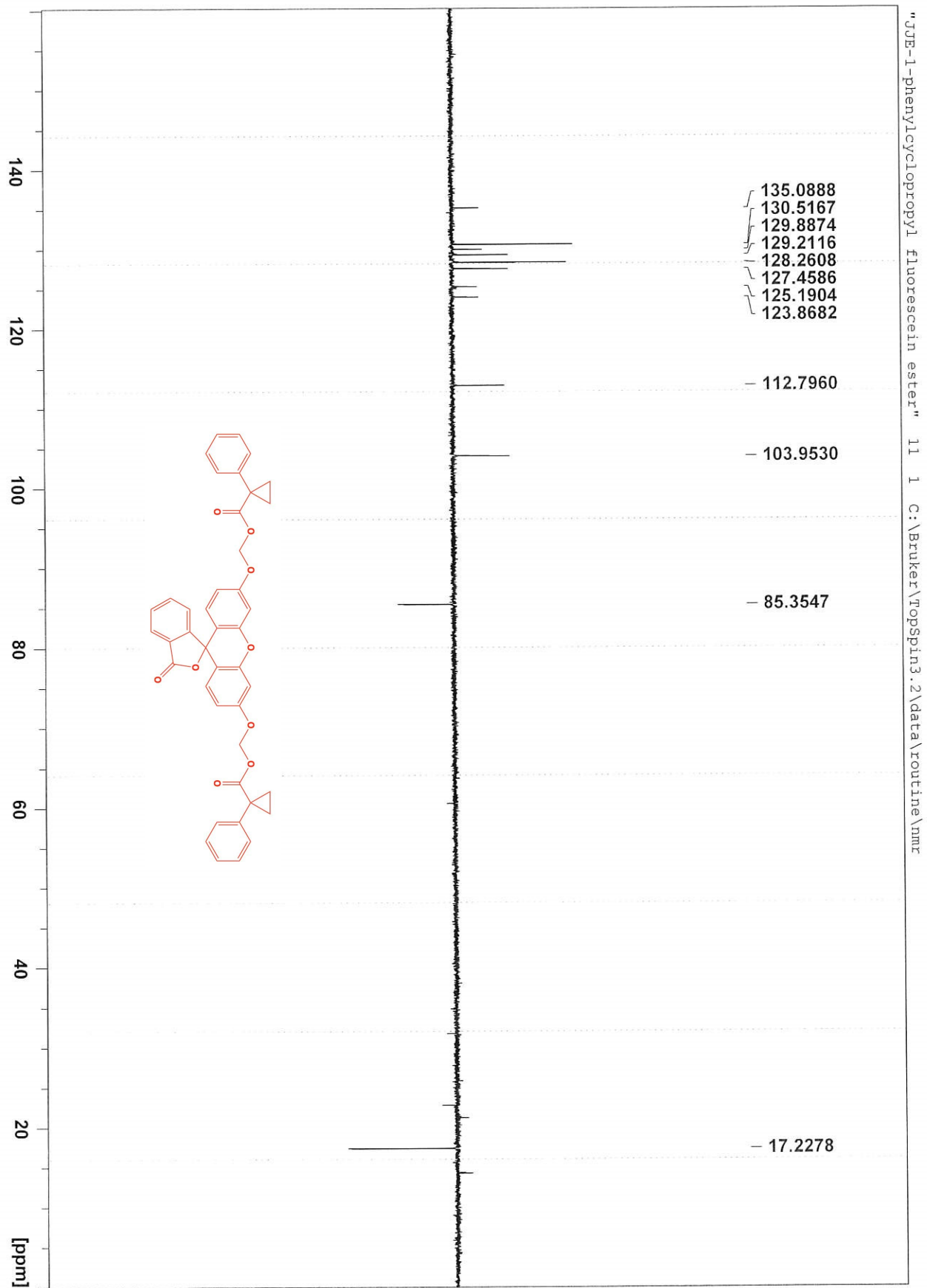


Figure S77

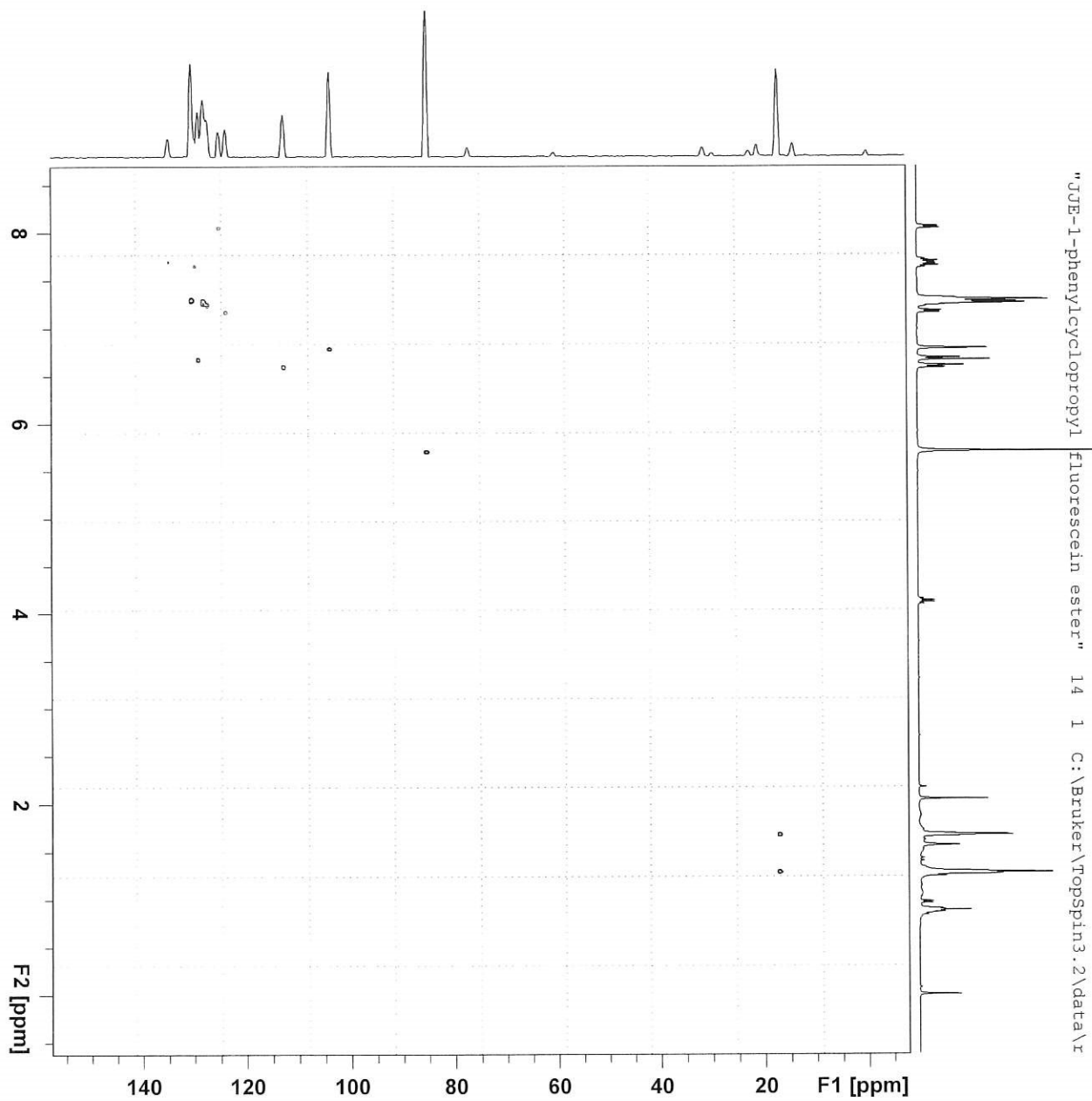
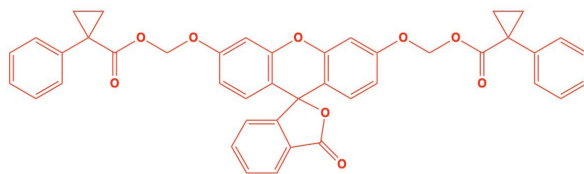


Figure S78

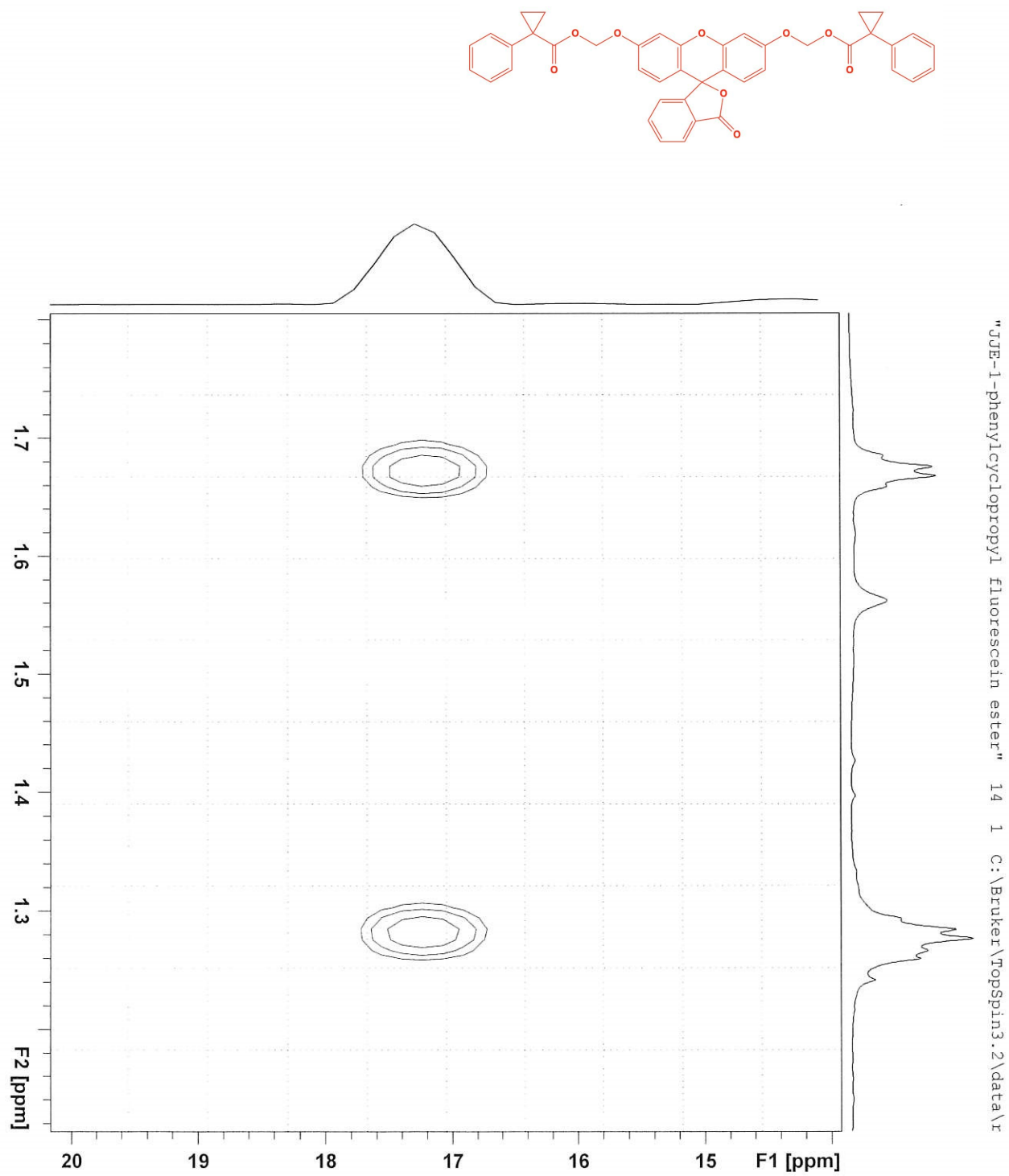
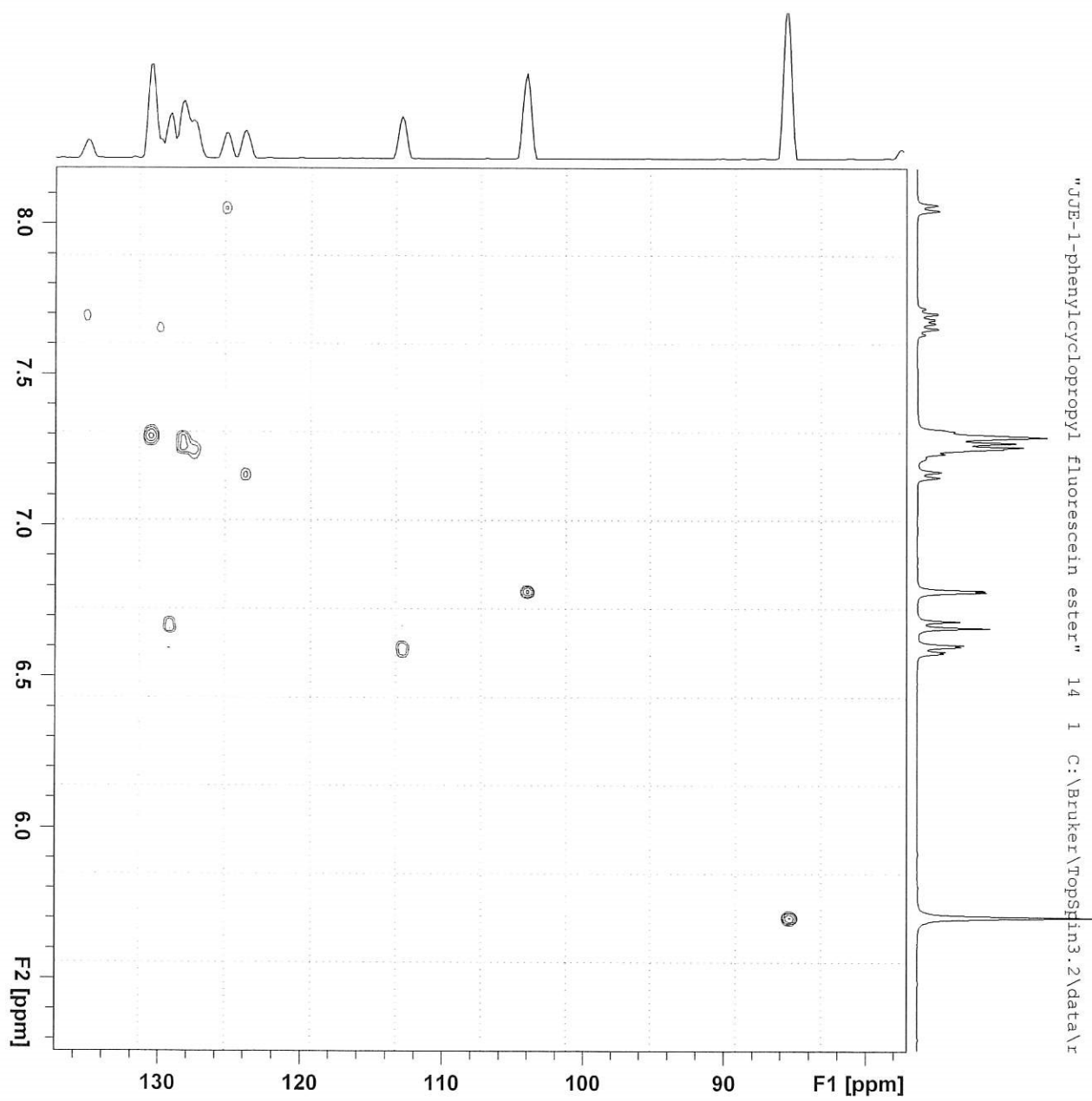
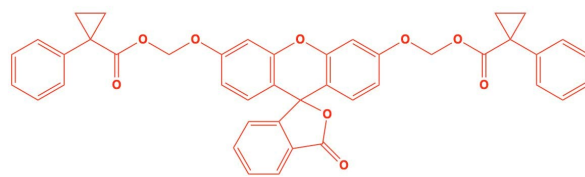
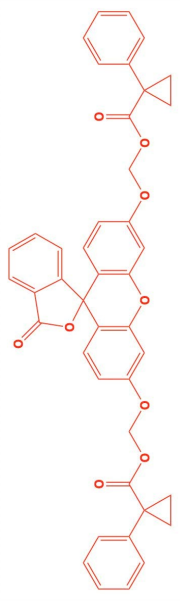
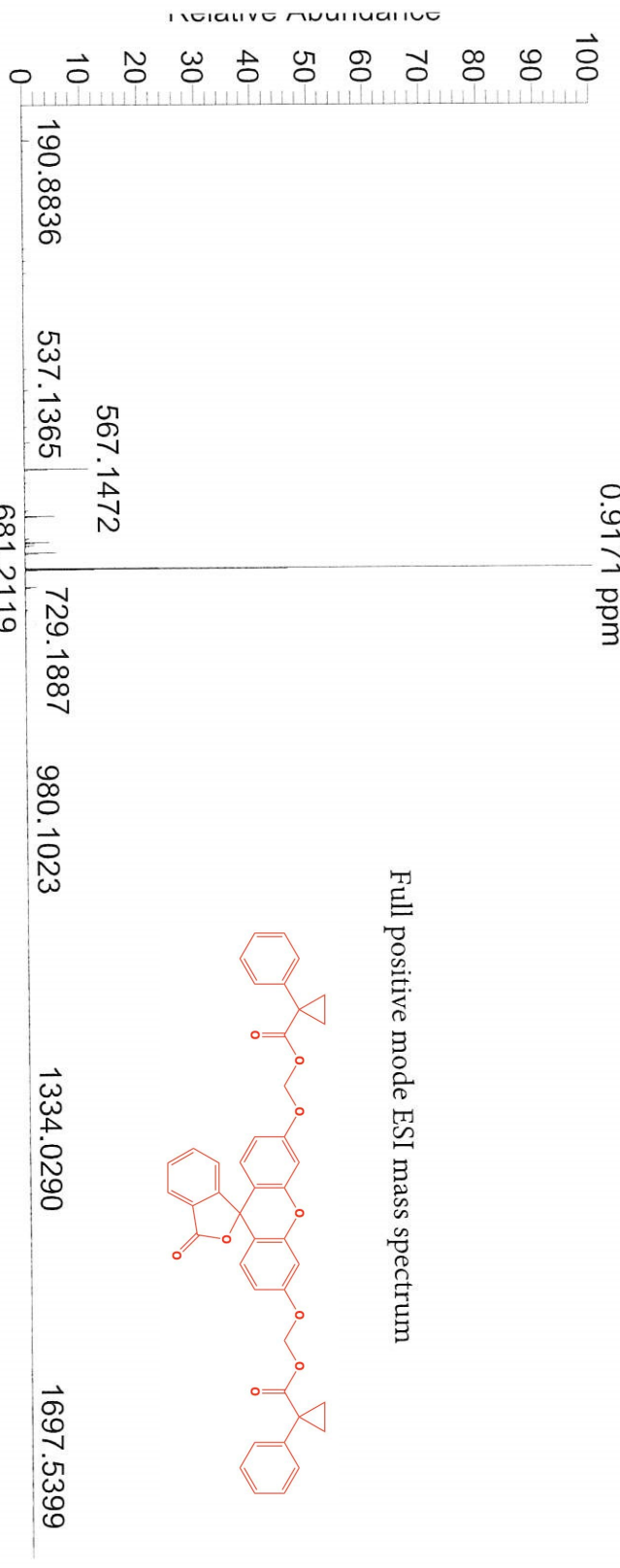


Figure S79

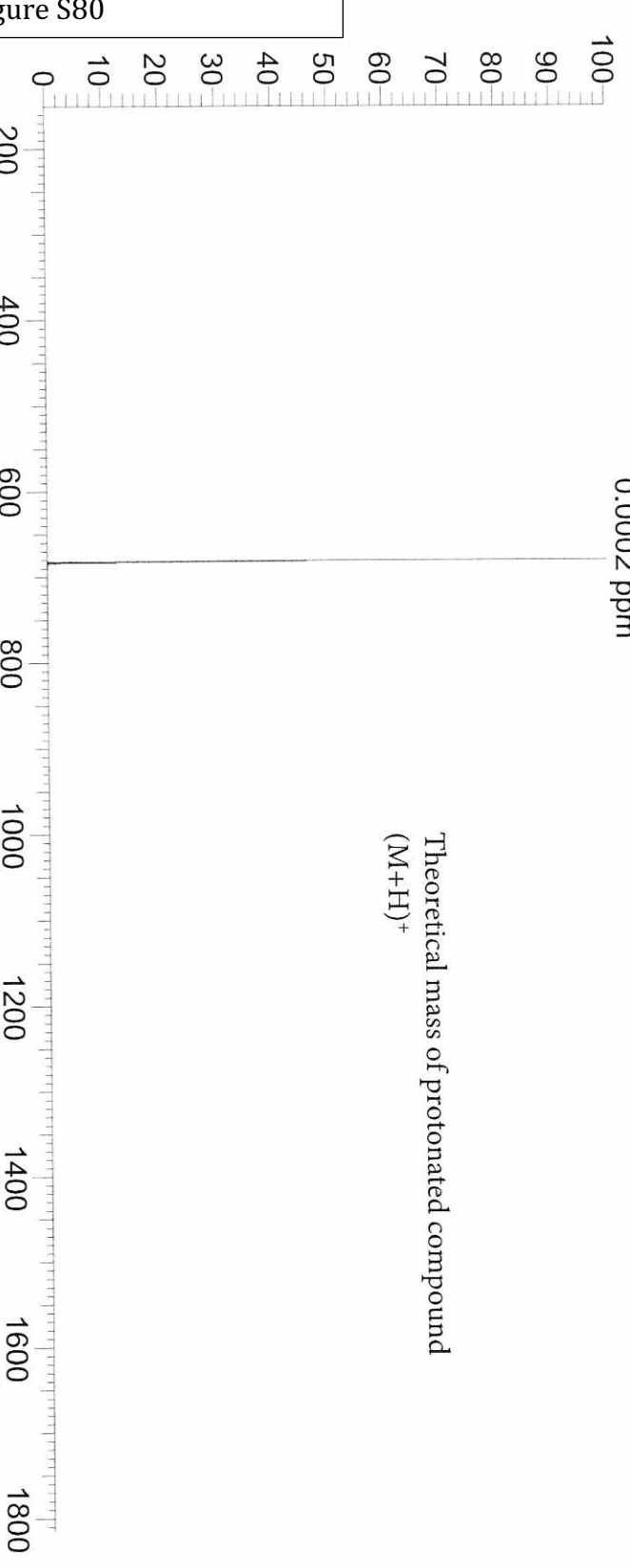


1-phenylcyclopropane carbonylic acid

NL:
6.36E7
JE-10#24-30 RT: 0.22-0.26
AV: 7 T: FTMS + p APPI
corona Full ms
[150.00-1800.00]



Theoretical mass of protonated compound
(M+H)⁺



NL:
6.20E5
C₄₂H₃₂O₉ H:
C₄₂H₃₃O₉
c (gss, s/p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Figure S80

NL:
 6.36E7
 JE-10#24-30 RT: 092-0.26
 AV: 7 T: FTMS + p APPI
 corona Full ms
 [150.00-1800.00]



Observed mass of protonated compound
 (M+H)⁺

681.2125
 C₄₂ H₃₃ O₉
 0.9171 ppm

675.3541

663.4540

684.2215

681.2119
 C₄₂ H₃₃ O₉
 0.0002 ppm

703.1939

729.1887

750.3921

761.1532

771.2594

Theoretical mass of protonated compound
 (M+H)⁺

NL:
 6.20E5
 C₄₂ H₃₂ O₉ H:
 C₄₂ H₃₃ O₉
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res .Pwr . @FWHM

Figure S81

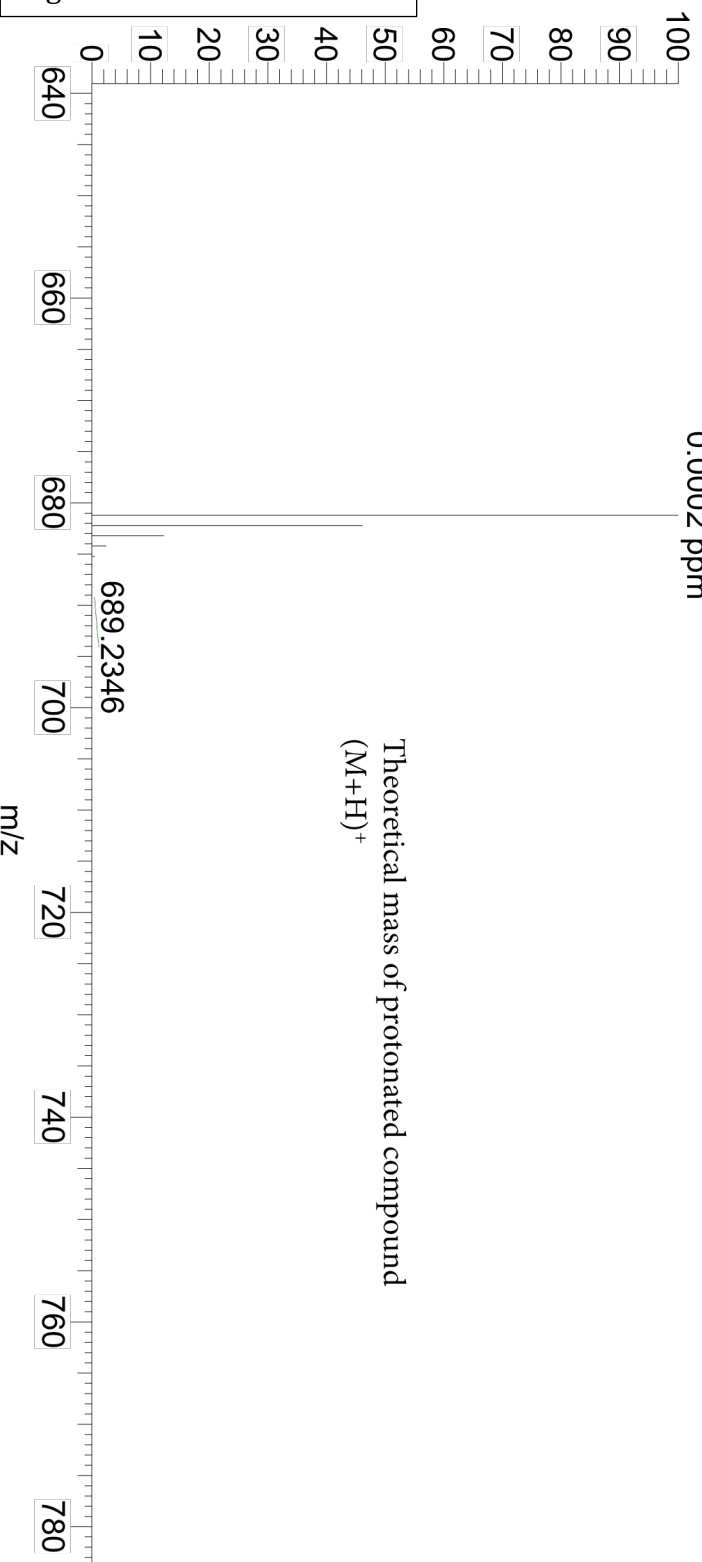


Figure S82

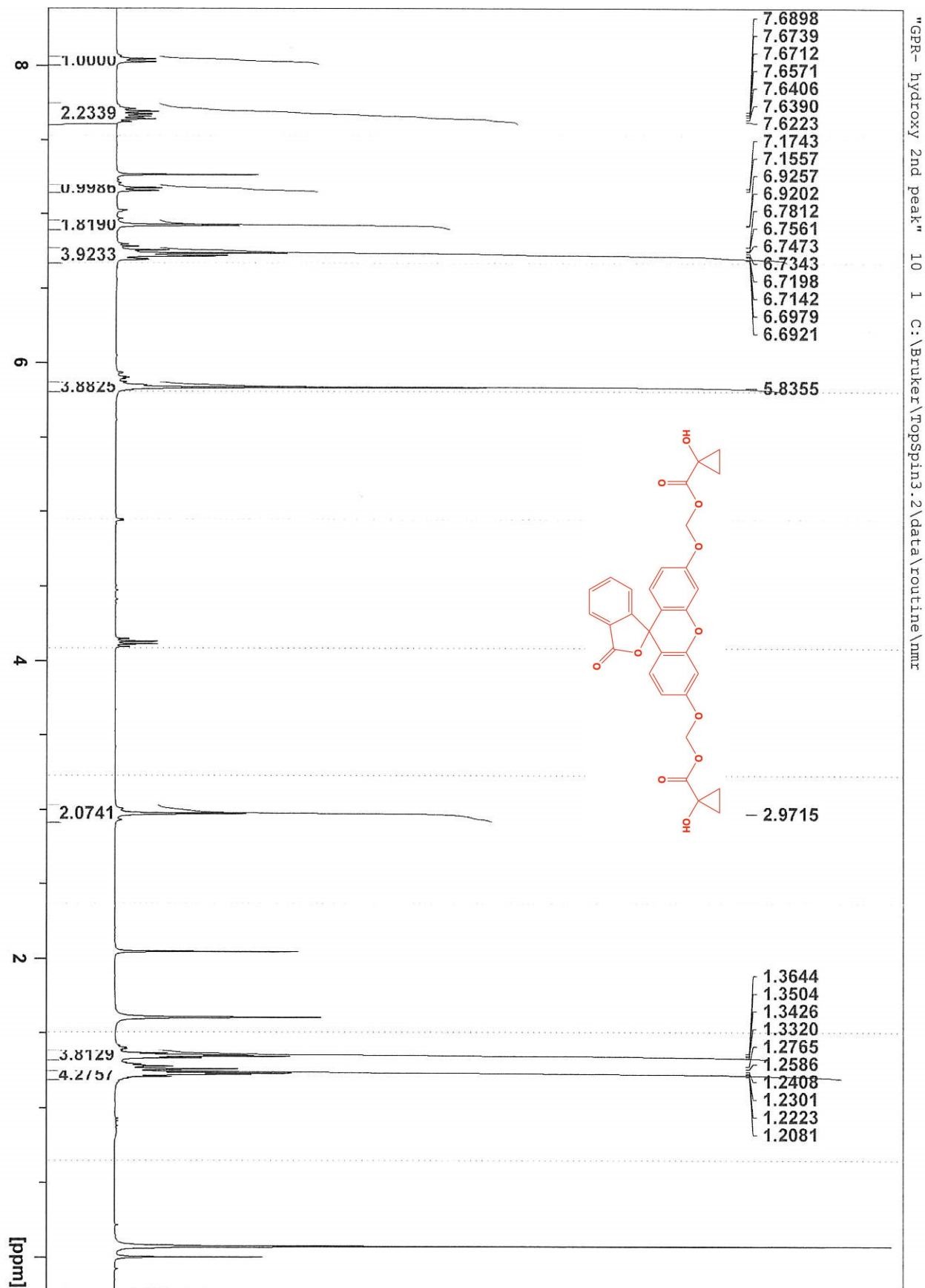


Figure S83

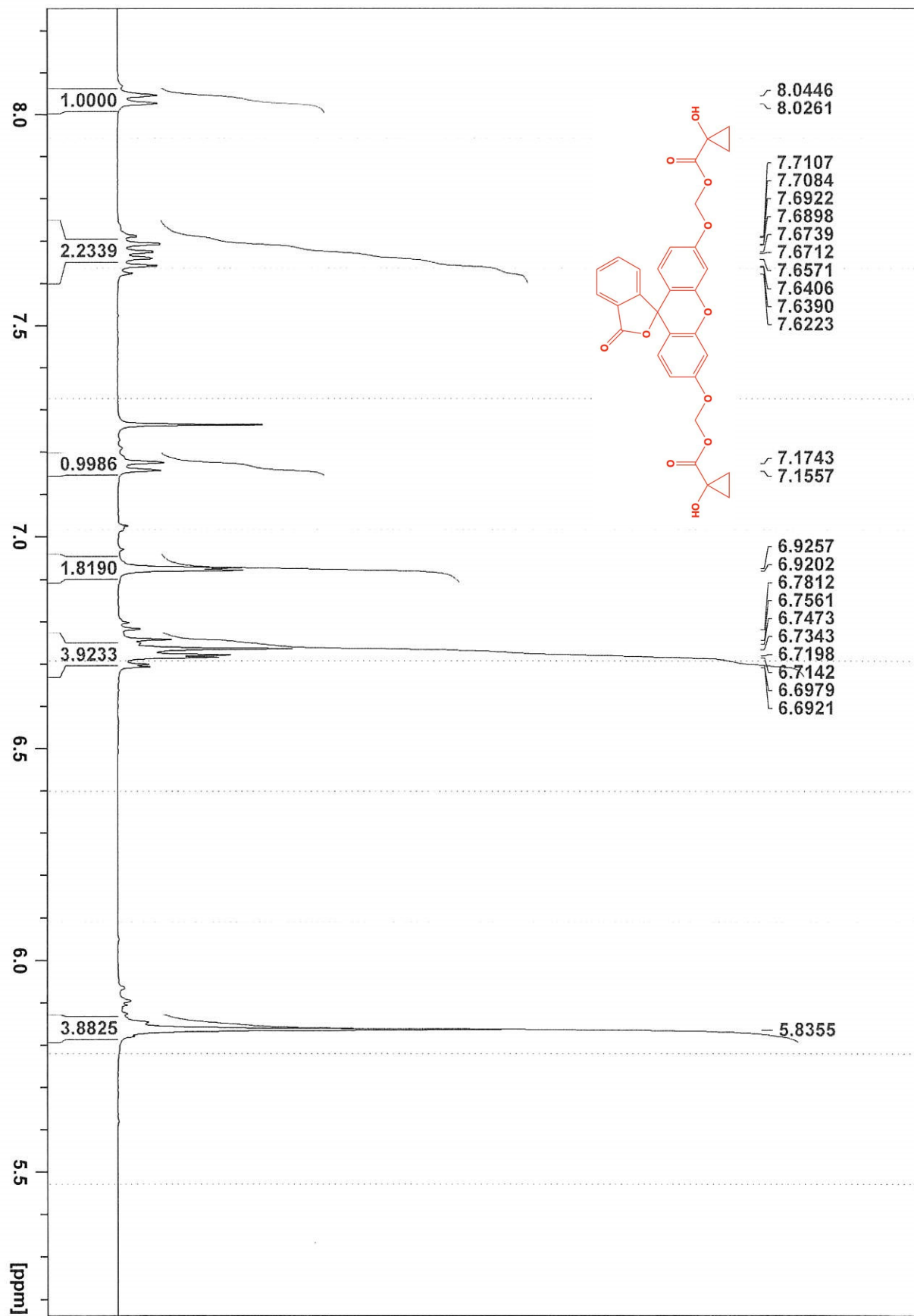


Figure S84

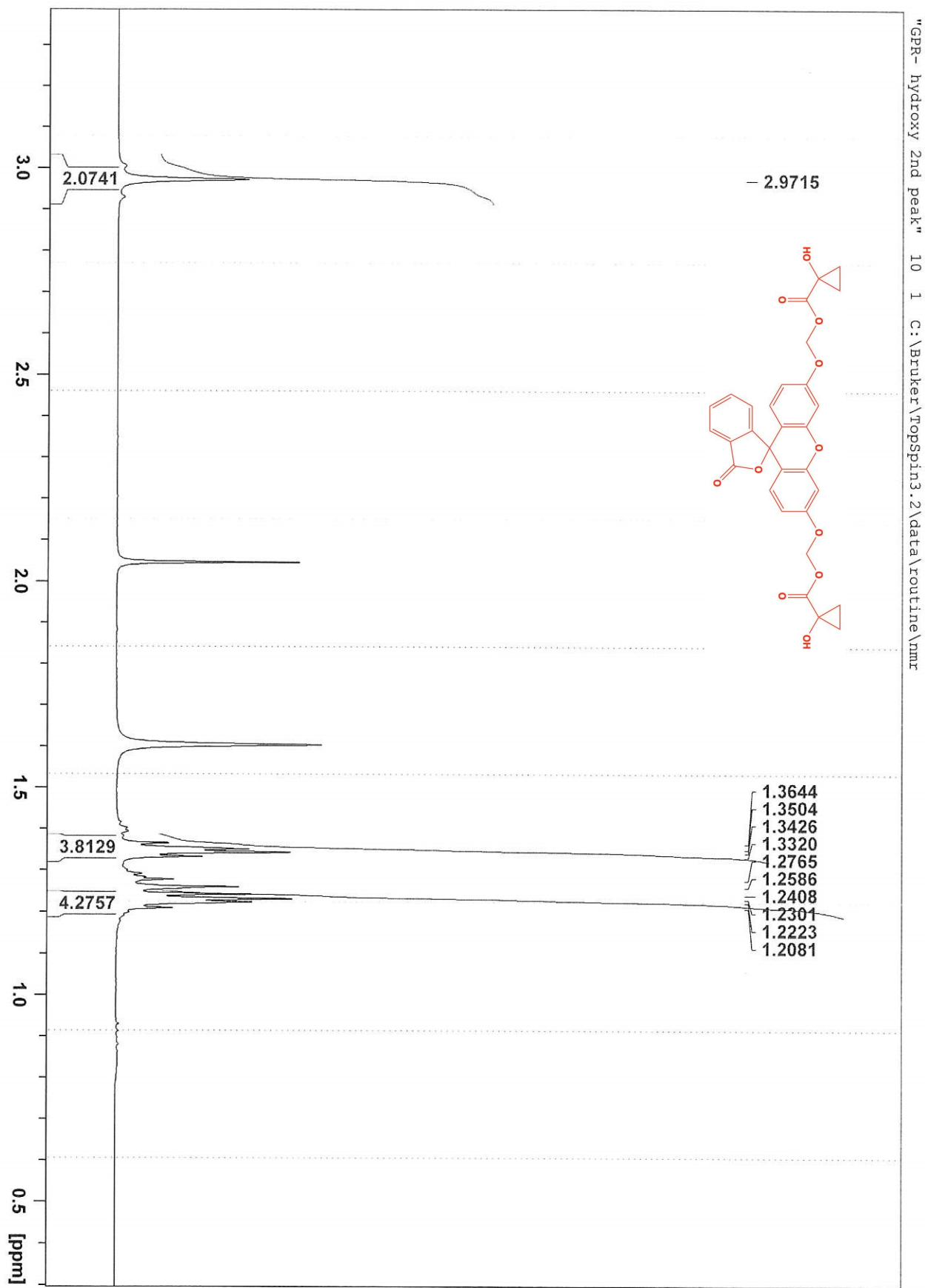


Figure S85

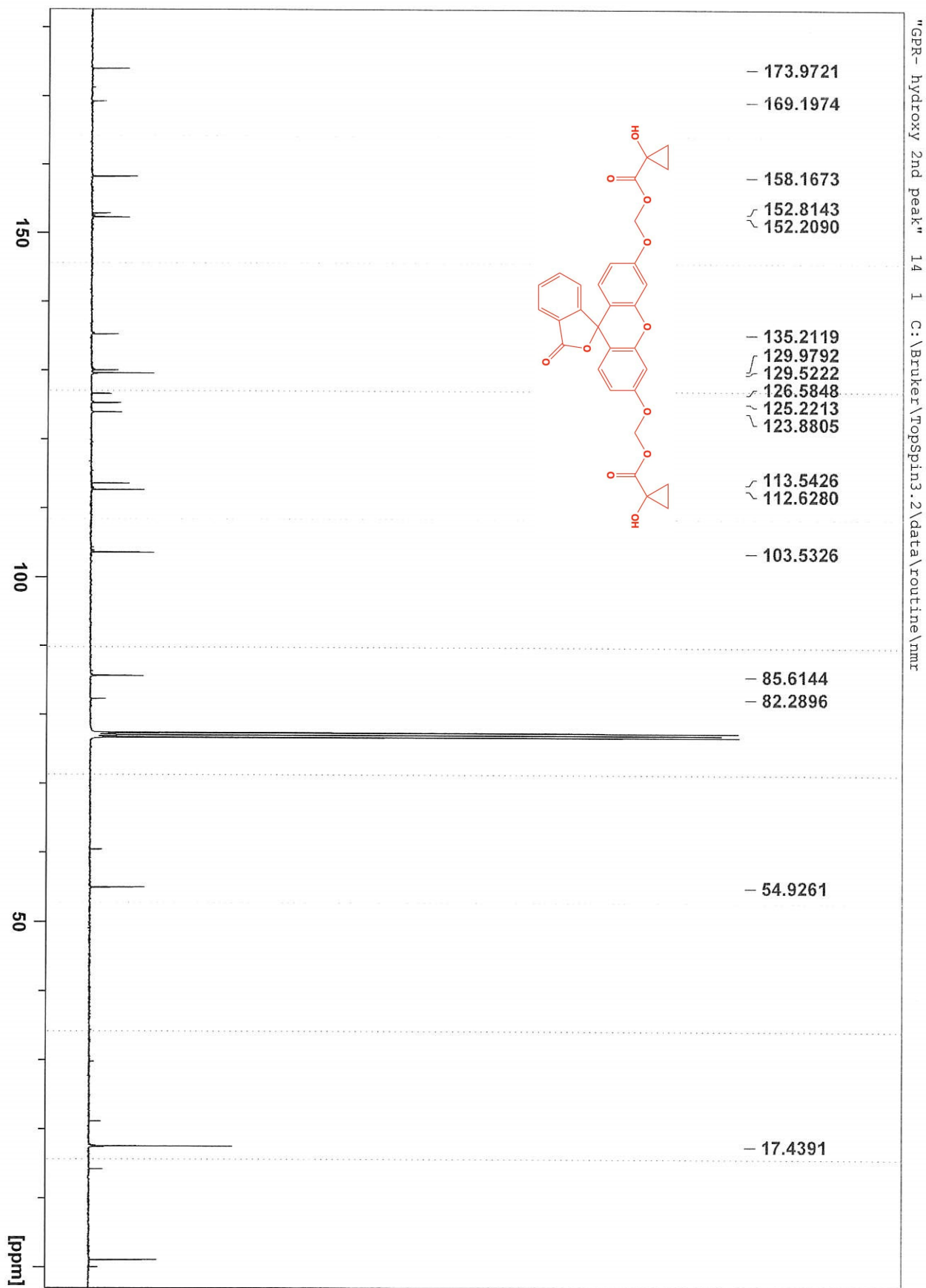


Figure S86

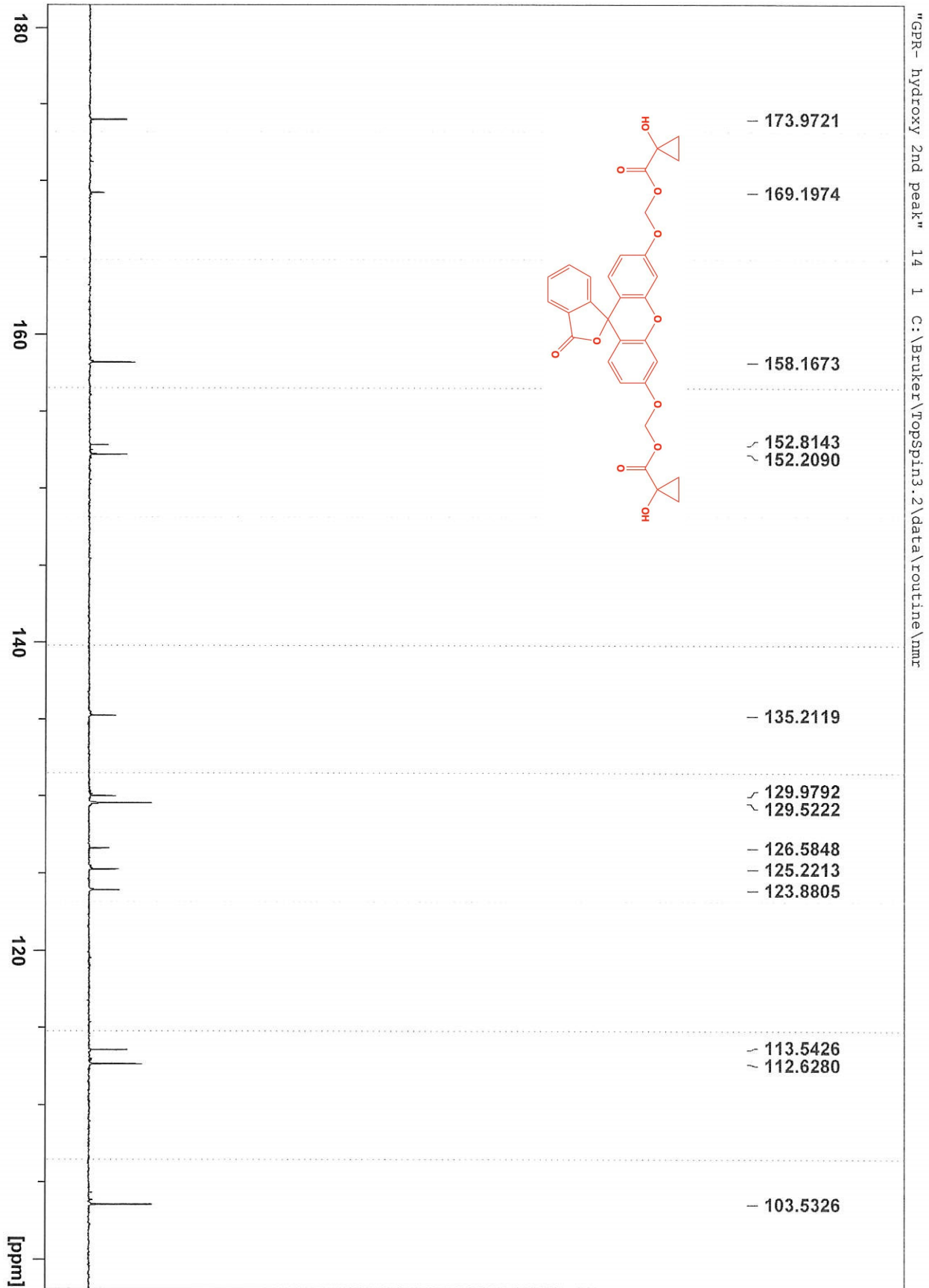


Figure S87

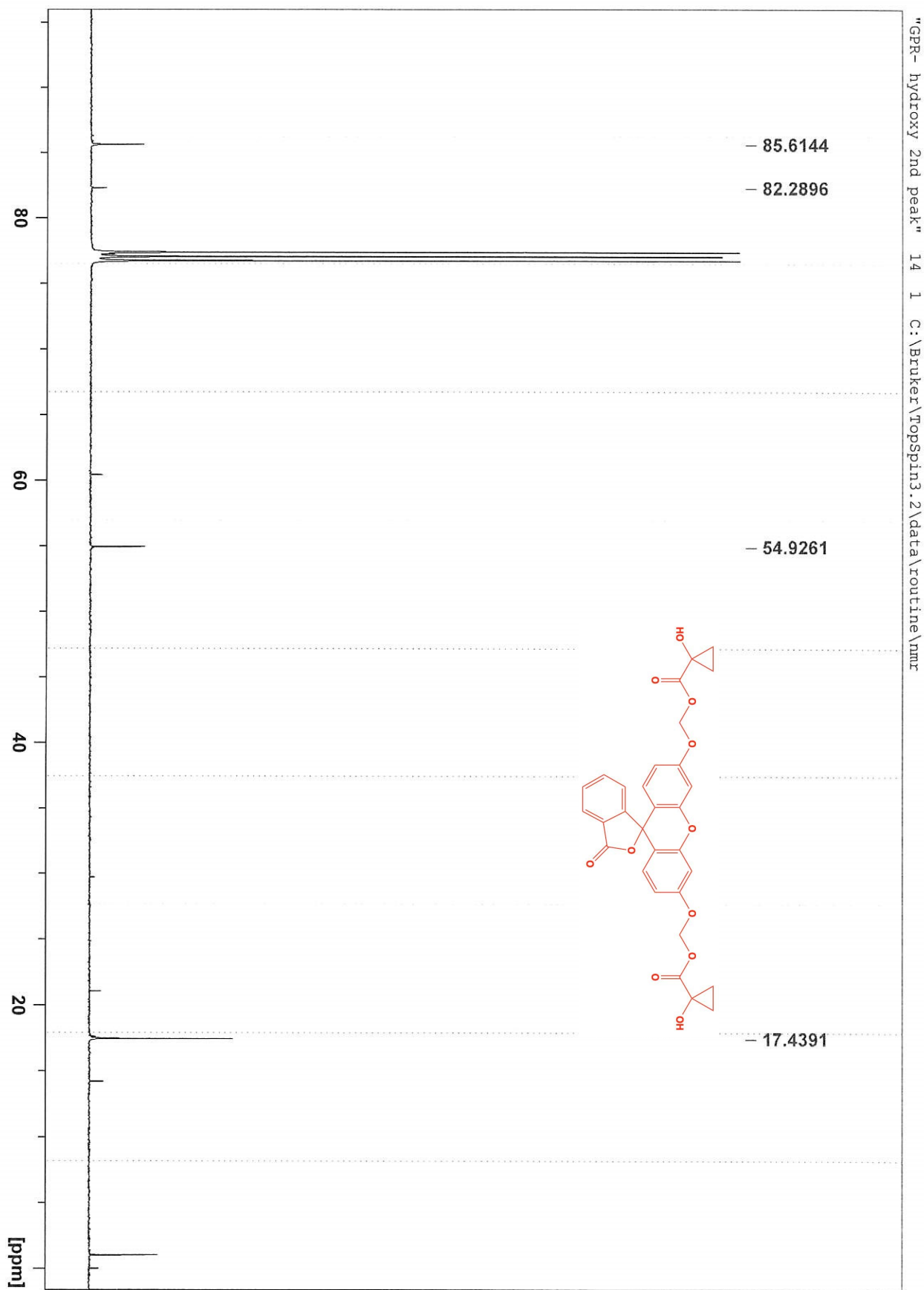


Figure S88

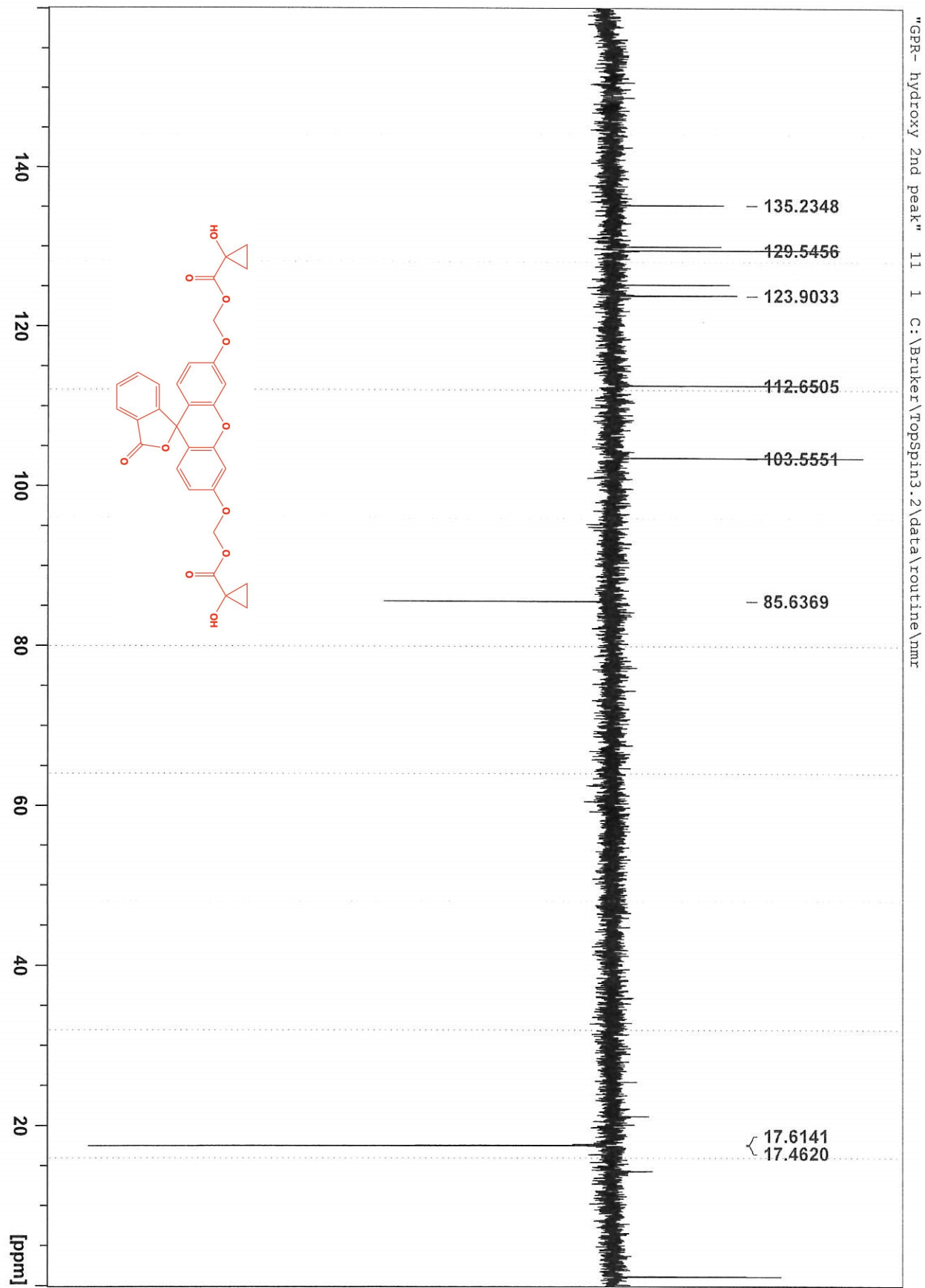
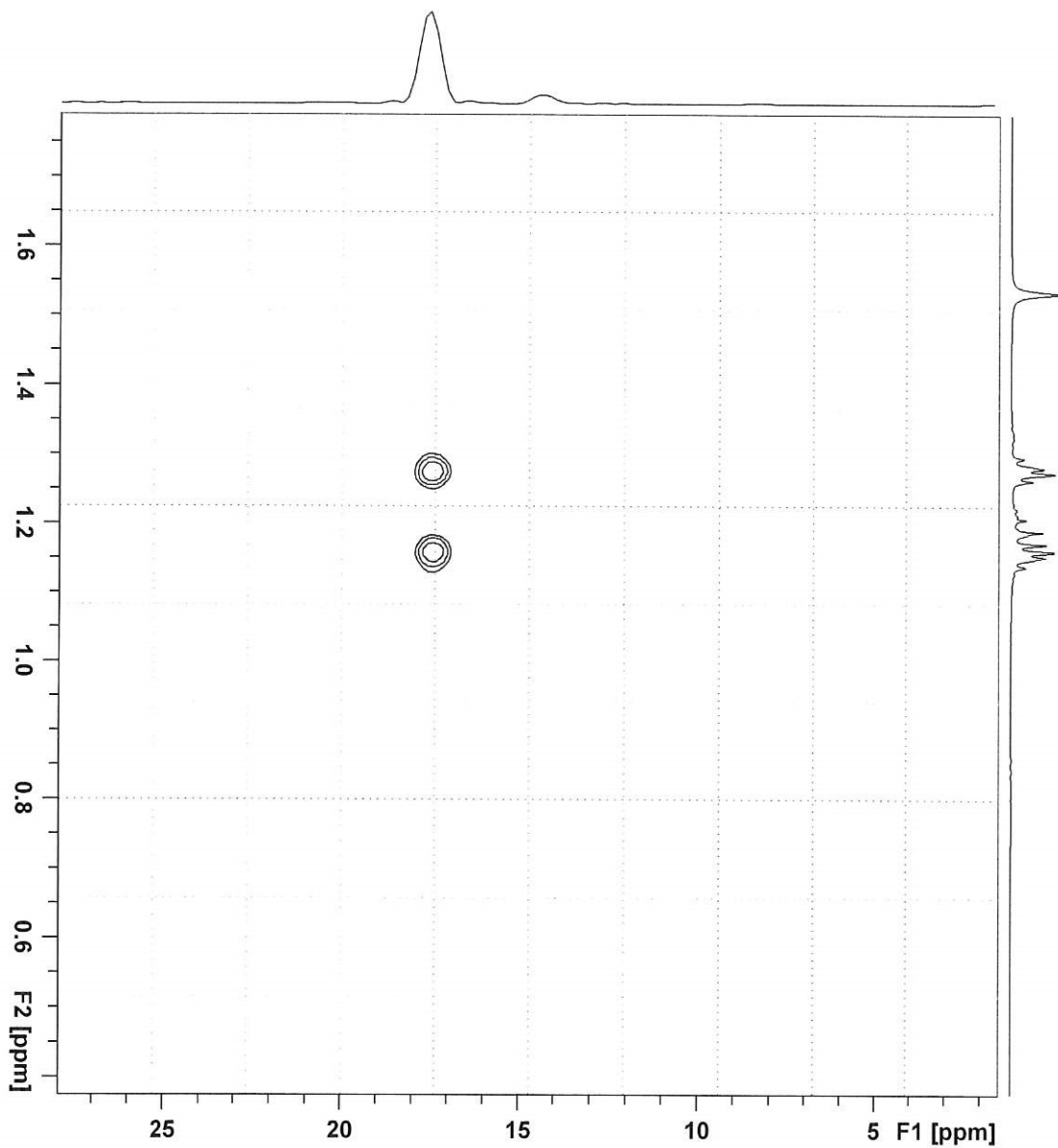
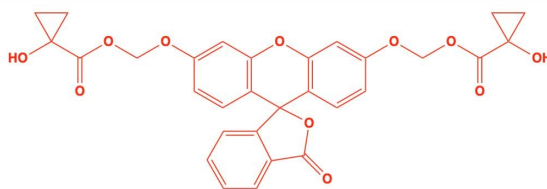


Figure S89



"GPR- hydroxy 2nd peak" 13 1 C:\Bruker\TopSpin3.2\data\routine\nmr

Figure S90

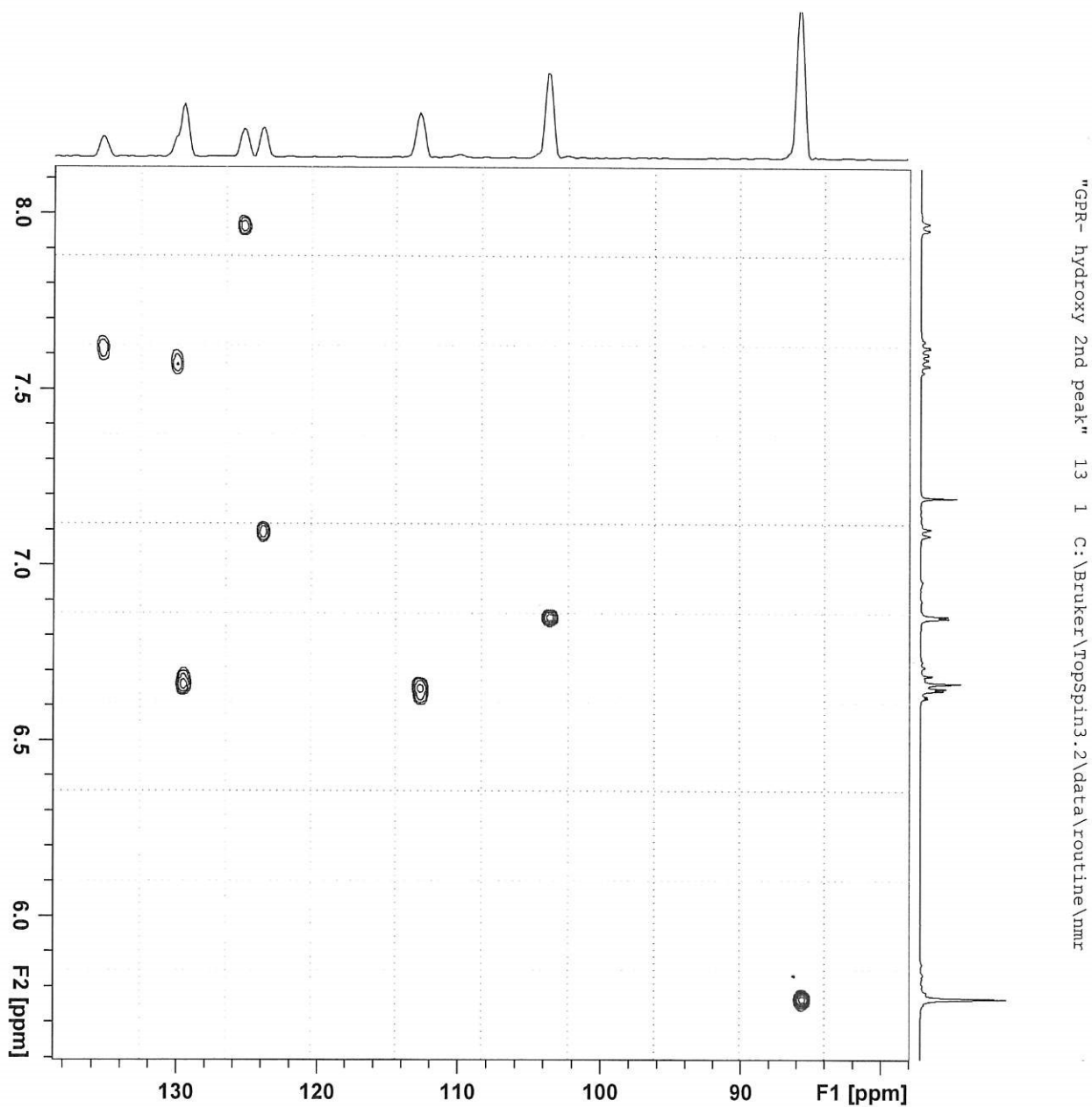
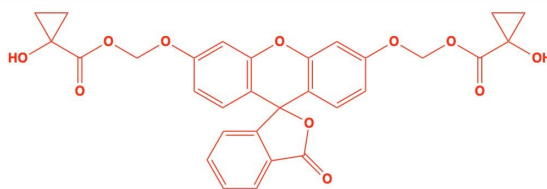


Figure S91

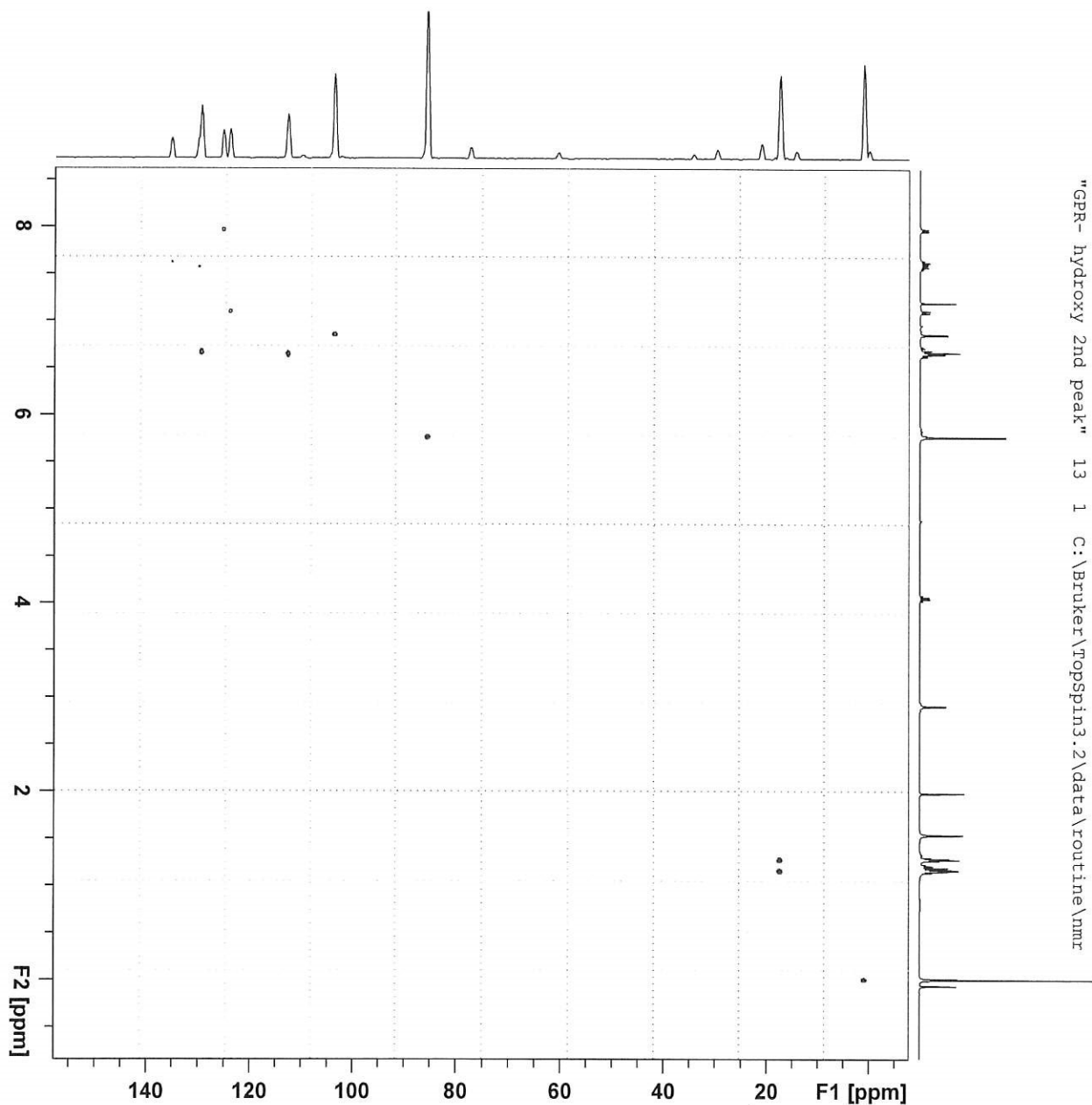
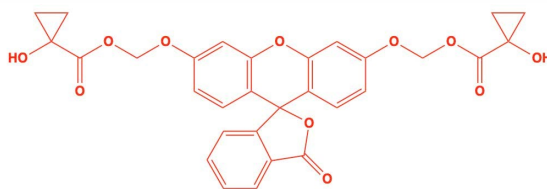
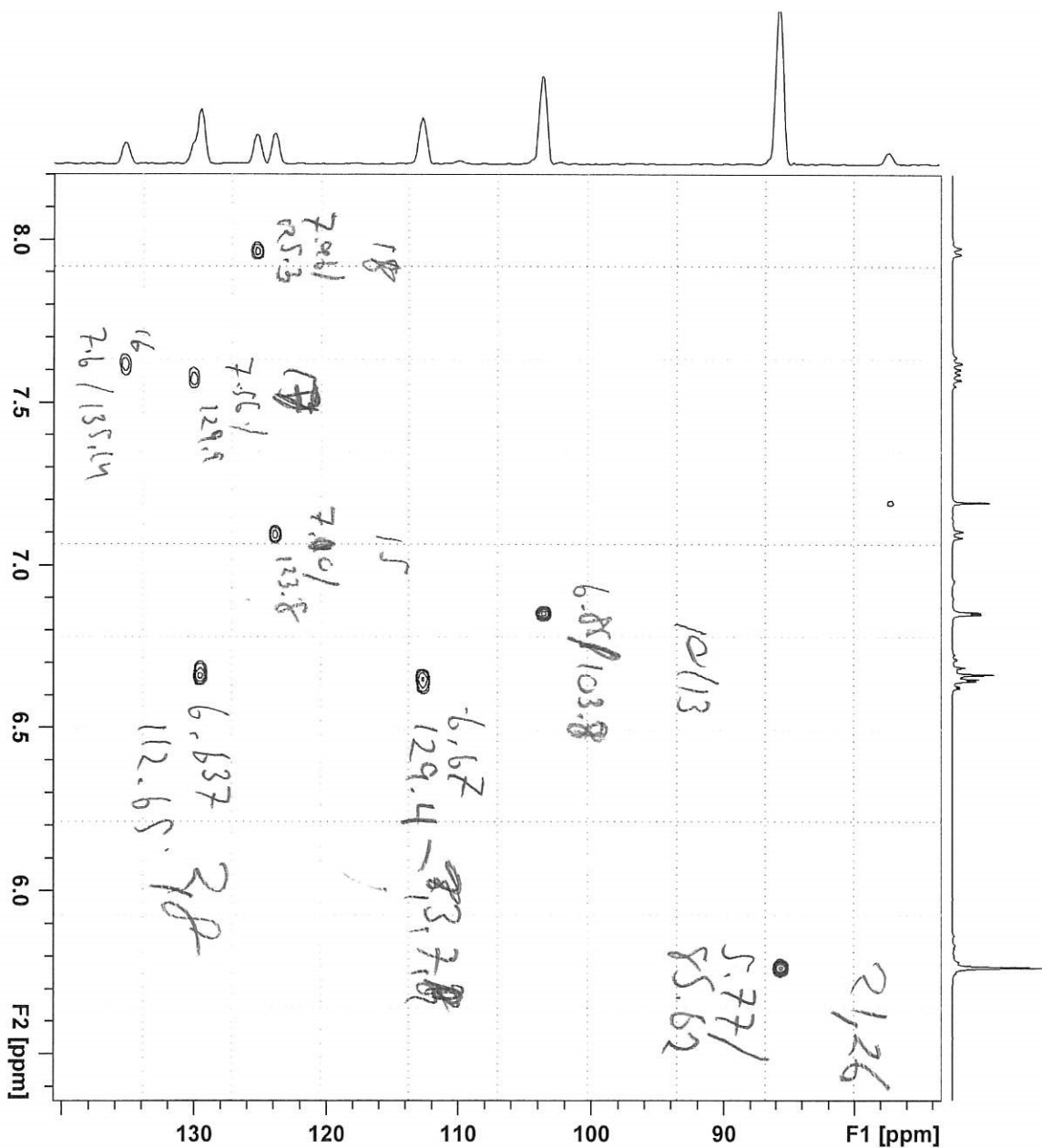
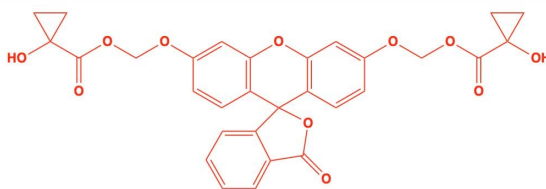
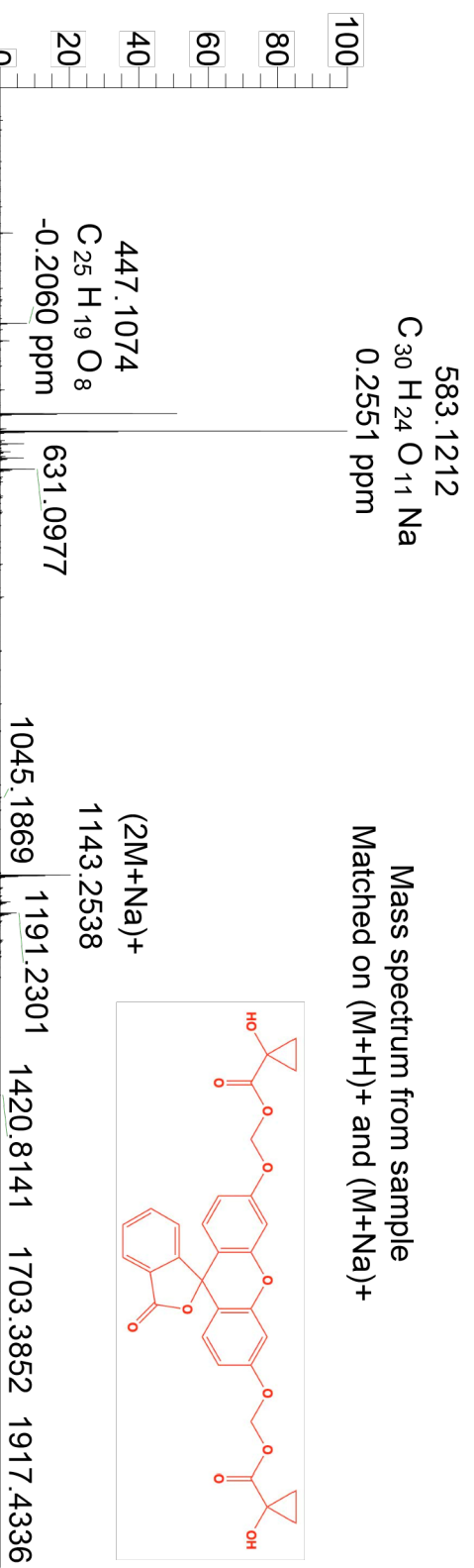


Figure S92



"GPR- hydroxy 2nd peak" 13 1 C:\Bruker\TopSpin3.2\data\routine\nmr



NL:
1.44E7
BU-4#9-18 RT: 0.180.29
AV: 10 T: FTMS + PFSI Full
ms [150.00-2000.00]

NL:
7.03E5

C₃₀H₂₄O₁₁H:
C₃₀H₂₅O₁₁
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr .@FWHM

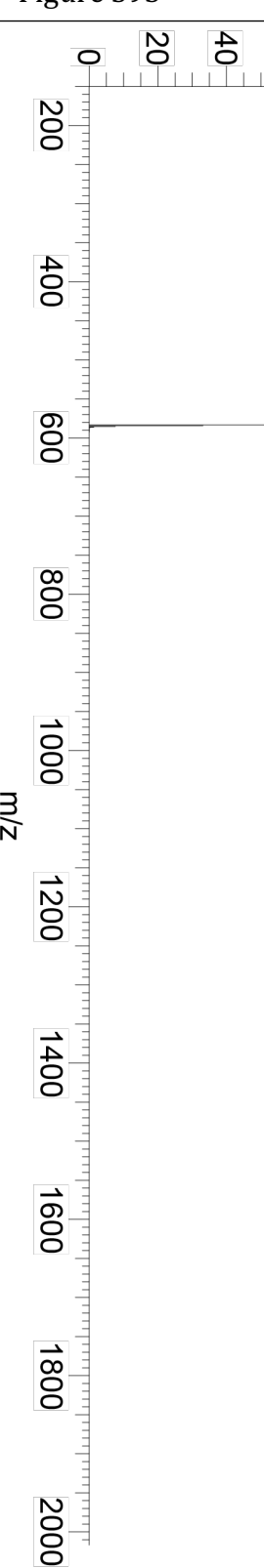


NL:
7.03E5

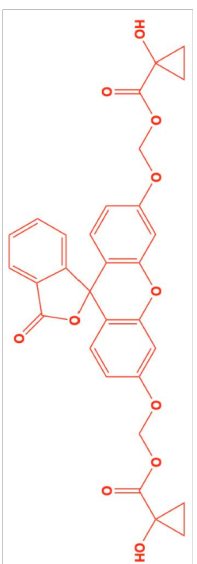
C₃₀H₂₄O₁₁Na:
C₃₀H₂₄O₁₁Na¹
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr .@FWHM

Theoretical mass spectrum for (M+Na)⁺

Figure S93



Mass spectrum from sample
Matched on (M+H)+ and (M+Na)+



NL:
1.44E7
BU-4#9-18 RT: 0.15-0.29
AV: 10 T: FTMS + PFSI Full
ms [150.00-2000.00]

NL:
7.03E5

C₃₀ H₂₄ O₁₁ H:
C₃₀ H₂₅ O₁₁
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM



NL:
7.03E5

C₃₀ H₂₄ O₁₁ Na:
C₃₀ H₂₄ O₁₁ Na₁
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+Na)+

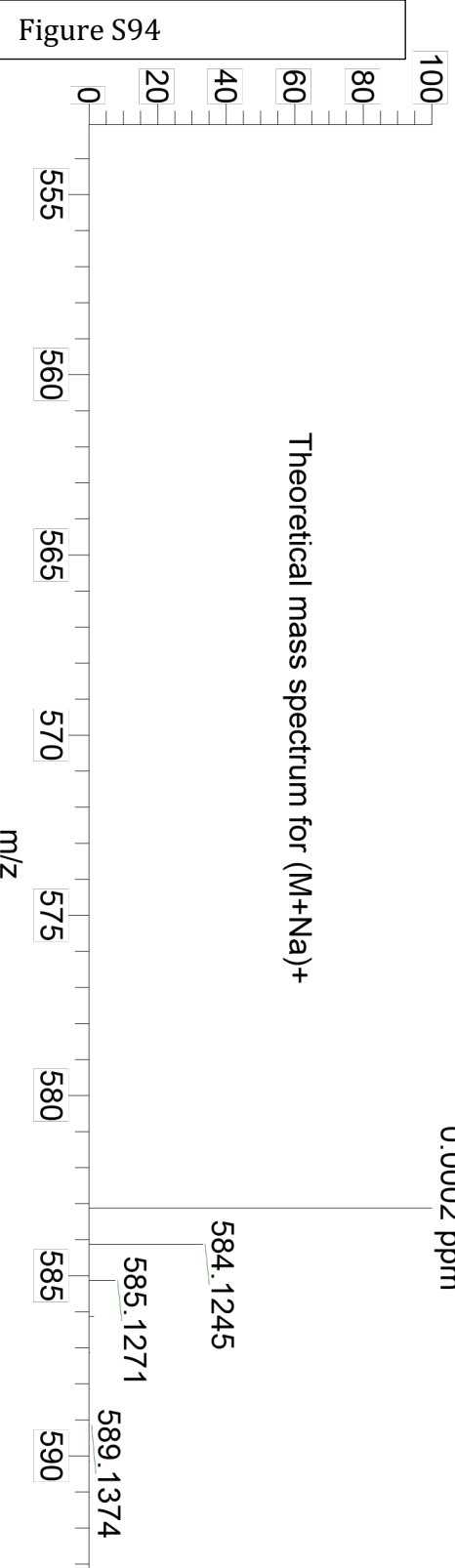
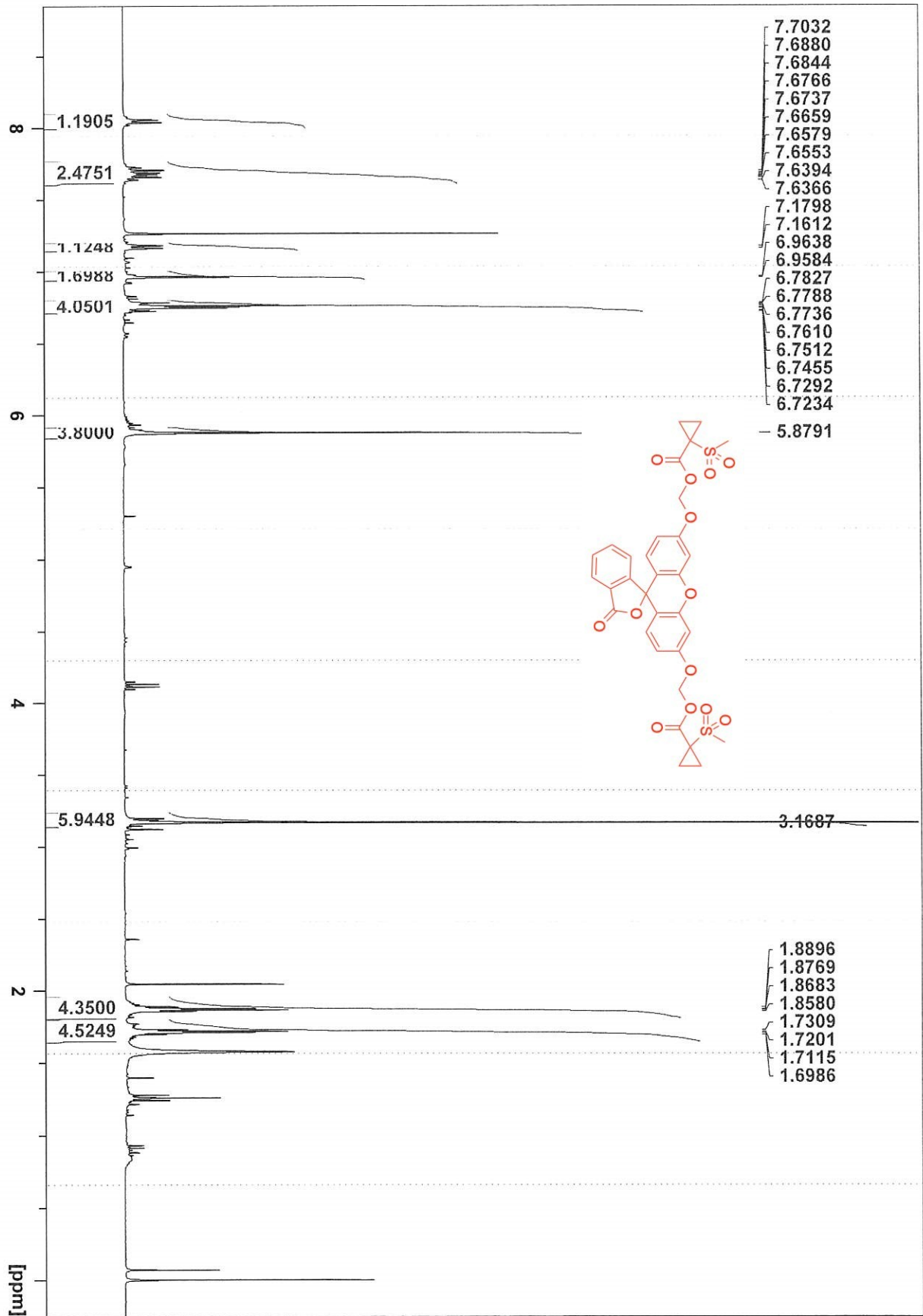


Figure S94

Figure S95



"GPR- 3 - methanesulFonyl" 10 1 C:\Bruker\TopSpin3.2\data\routine\mmr

[ppm]

8

6

4

2

1.1905

2.4751

1.1248

1.6988

4.0501

3.8000

5.9448

4.3500

4.5249

7.7032

7.6880

7.6844

7.6766

7.6737

7.6659

7.6579

7.6553

7.6394

7.6366

7.1798

7.1612

6.9638

6.9584

6.7827

6.7788

6.7736

6.7610

6.7512

6.7455

6.7292

6.7234

3.1687

1.8896

1.8769

1.8683

1.8580

1.7309

1.7201

1.7115

1.6986

Figure S96

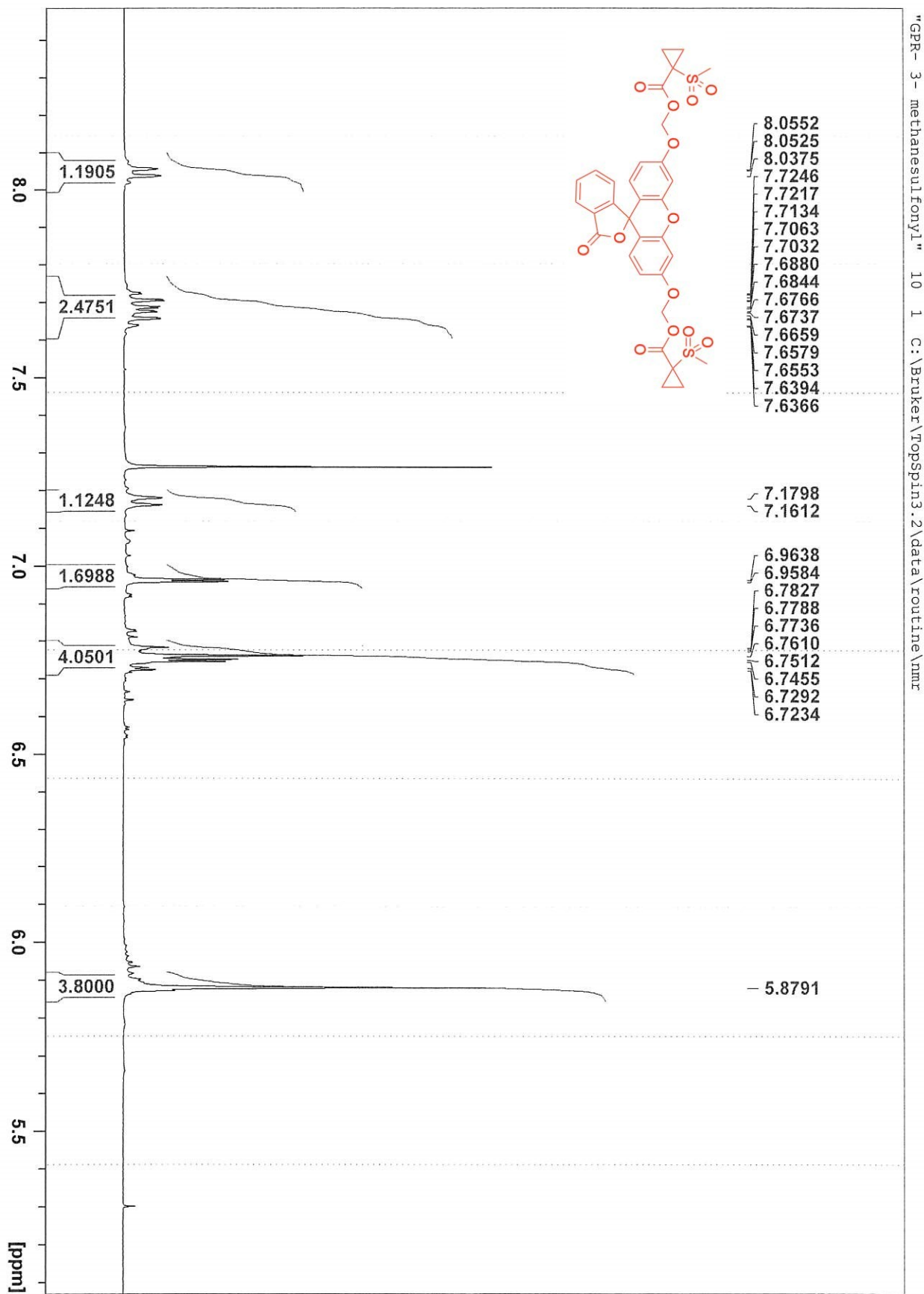


Figure S97

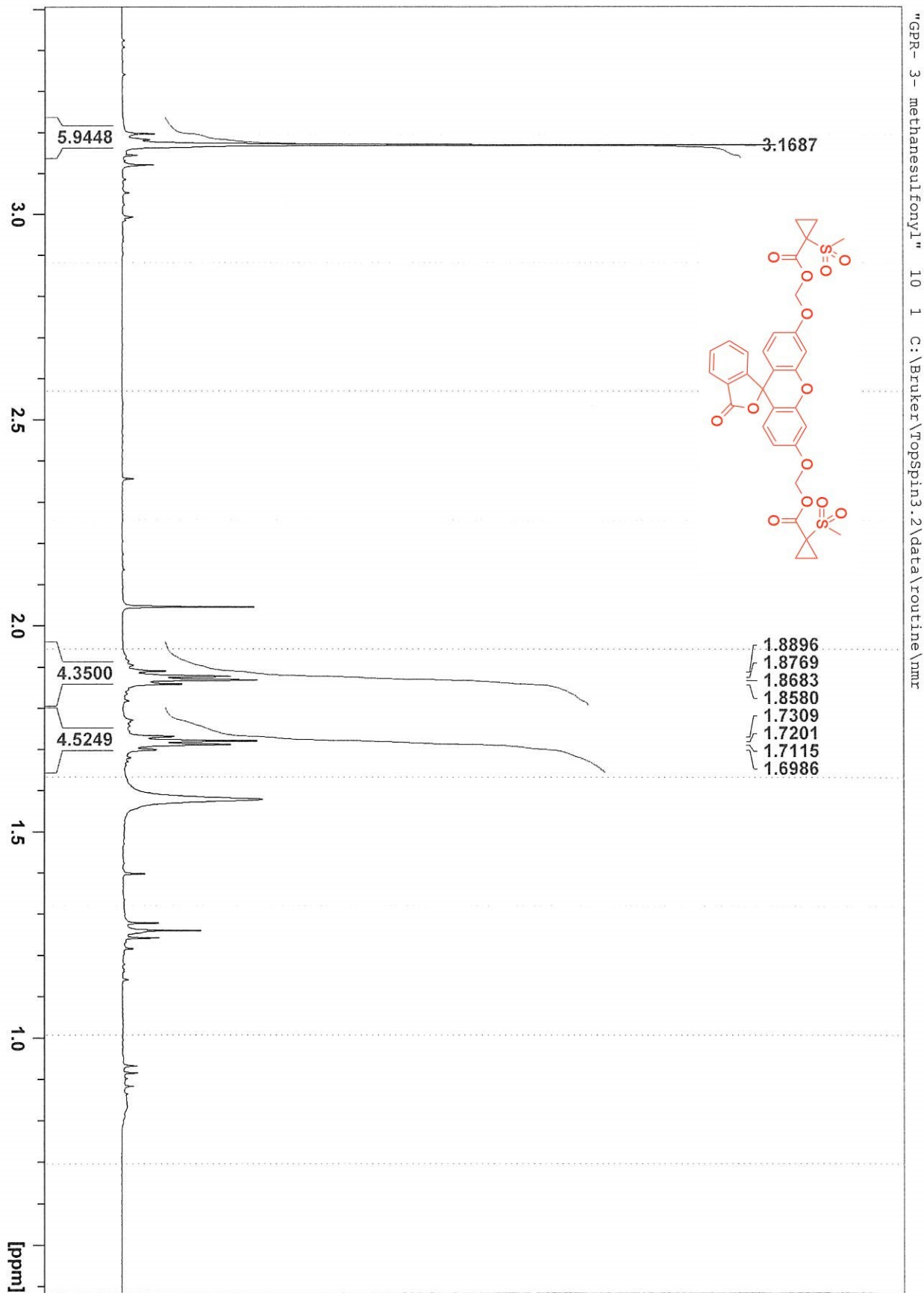


Figure S98

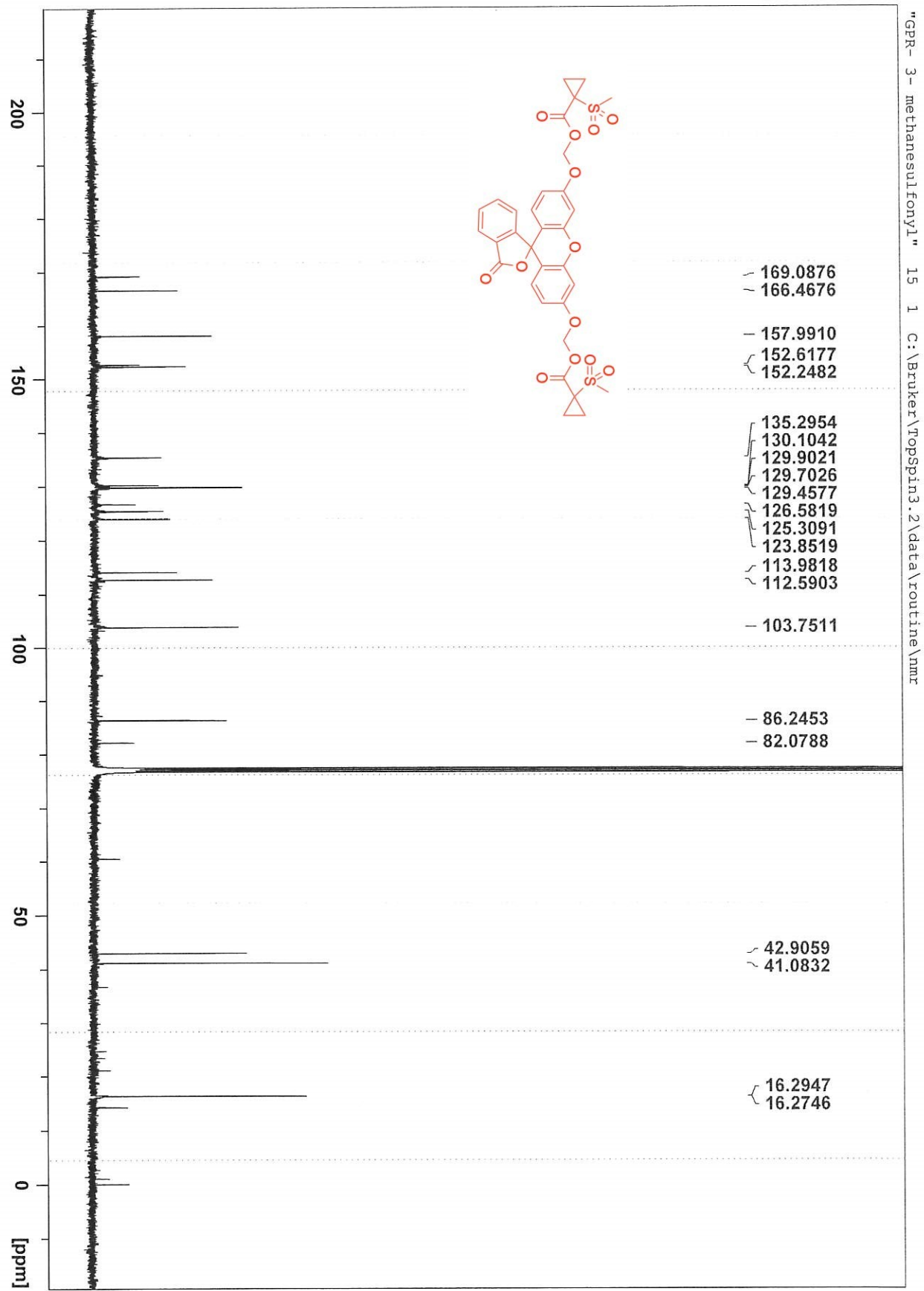


Figure S99

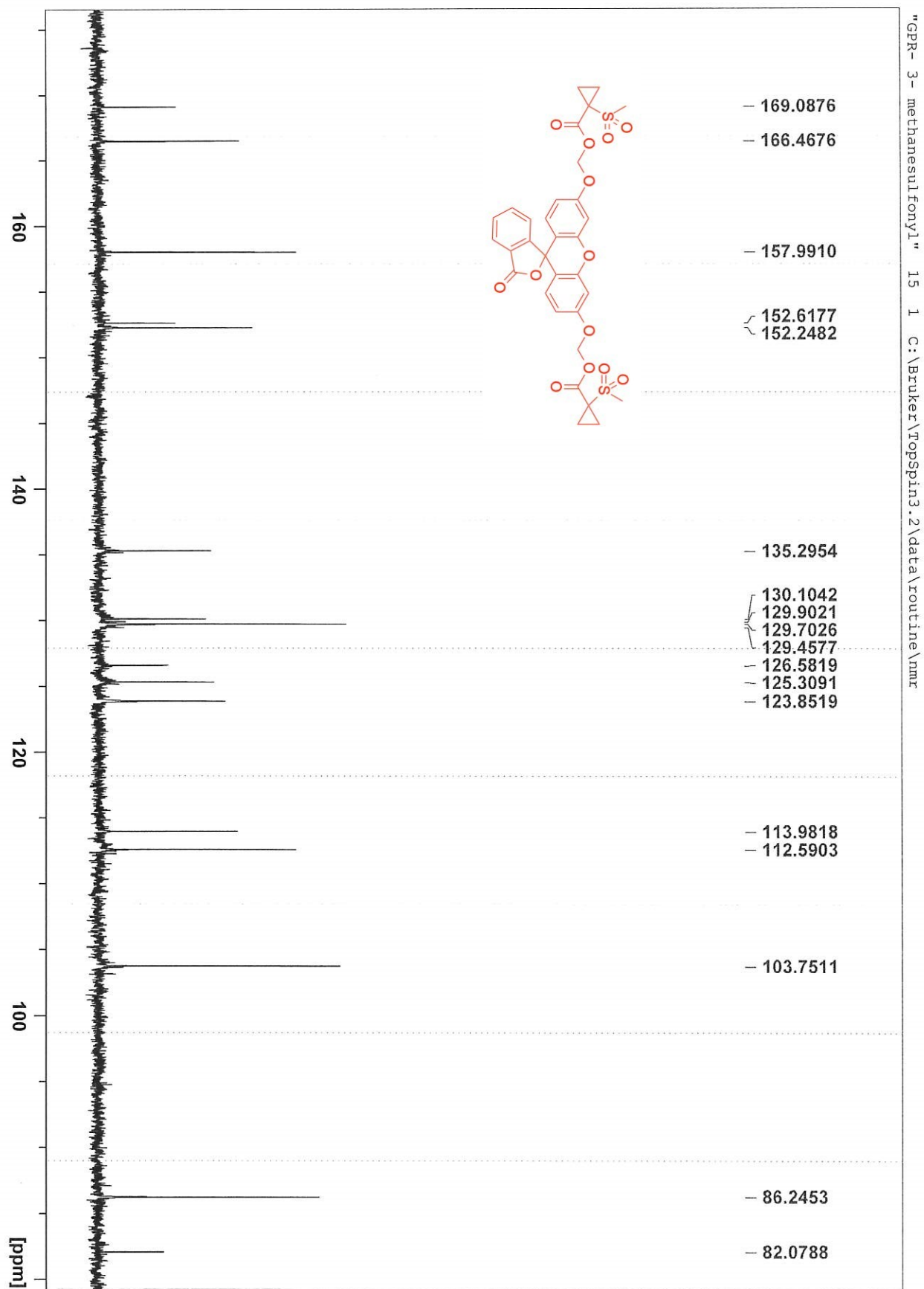


Figure S100

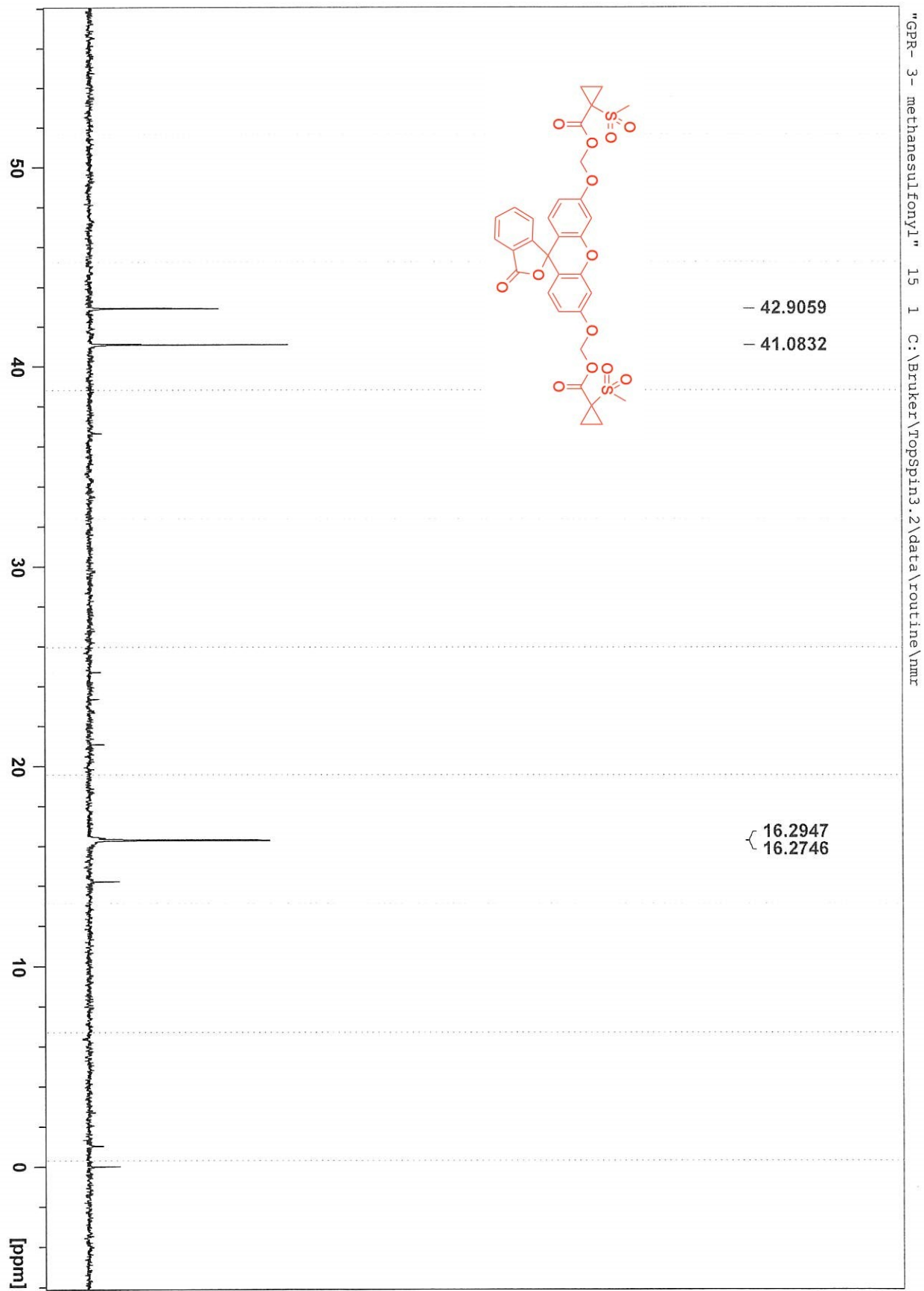
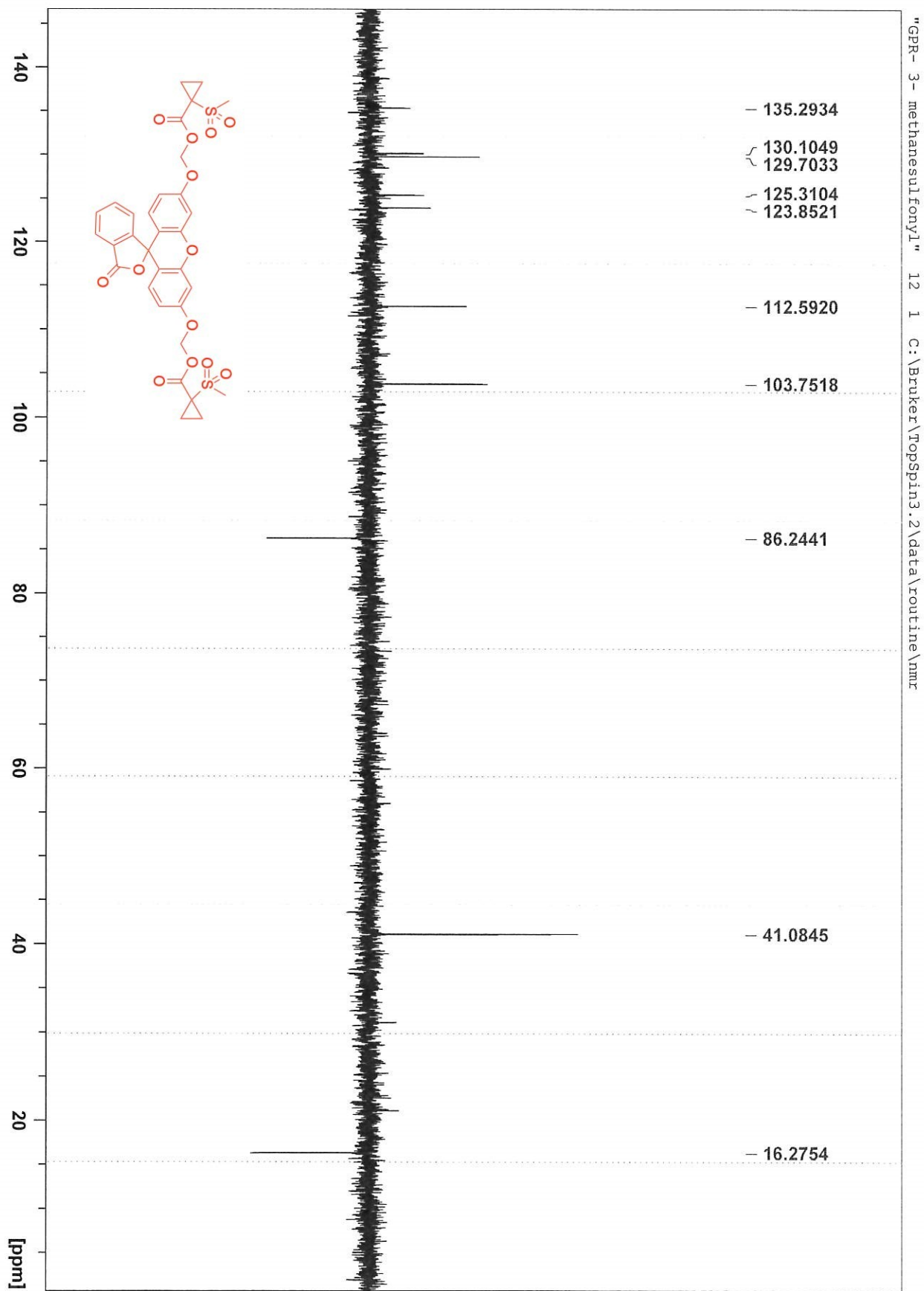


Figure S101



GPR- 3- methanesulfonylfonyl

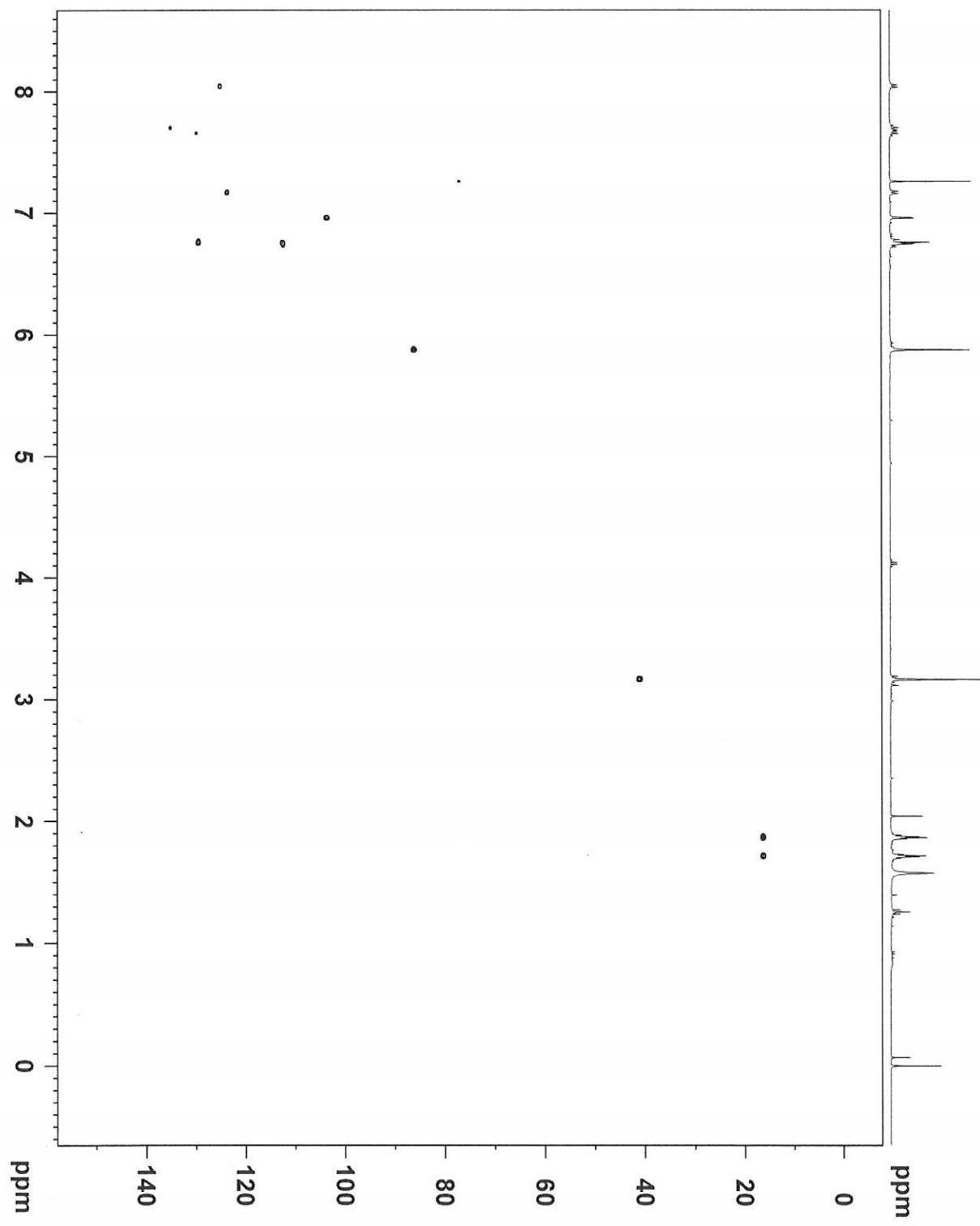
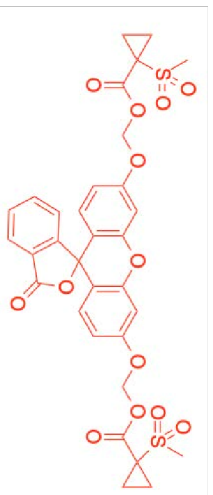
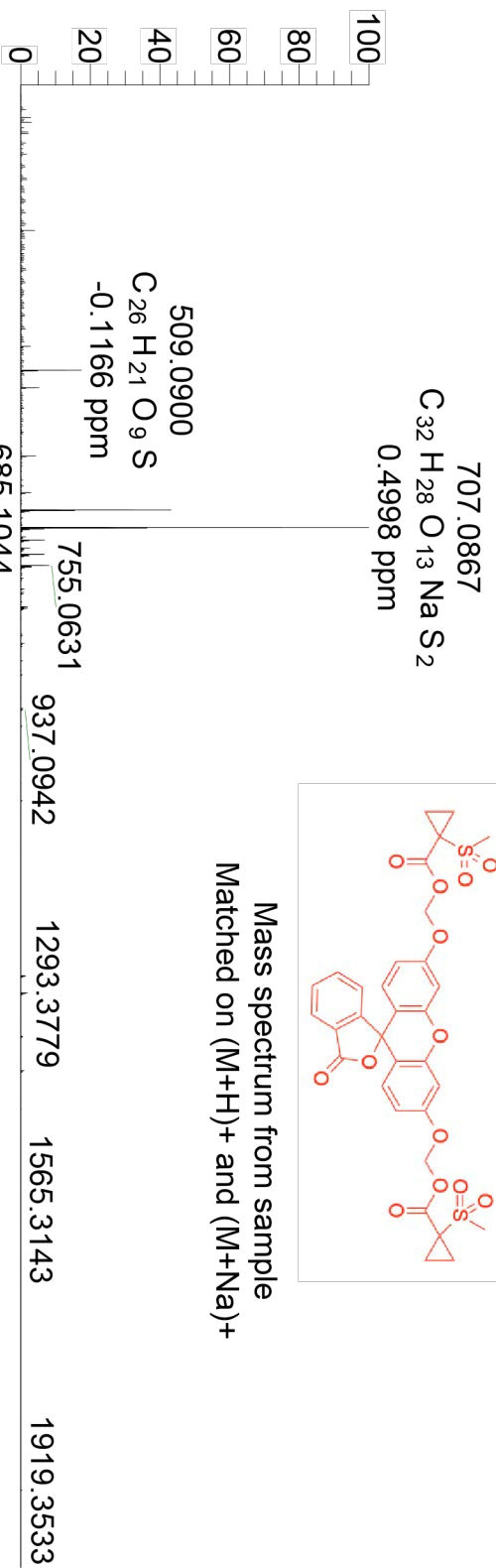


Figure S102



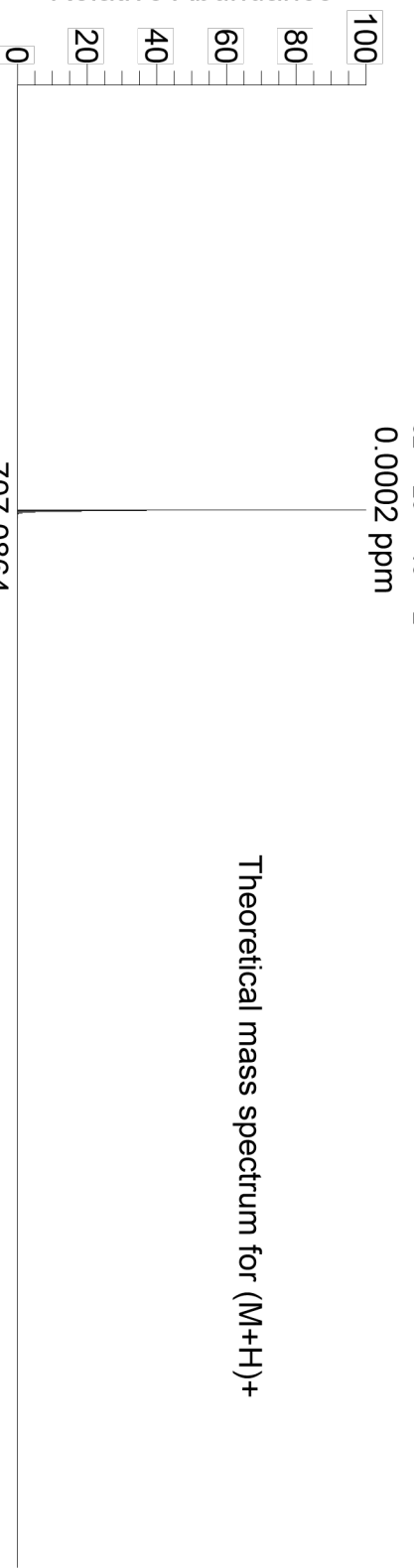
Mass spectrum from sample
Matched on (M+H)⁺ and (M+Na)⁺



NL:
9.26E6
BU-3#19-49 RT: 0.95-0.39
AV: 31 SB: 122 0.08-0.11
1.06-1.98 T: FTMS + p ESI
Full ms [150.00-2000.00]

NL:
6.17E5

C₃₂ H₂₈ O₁₃ S₂ H:
C₃₂ H₂₉ O₁₃ S₂
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

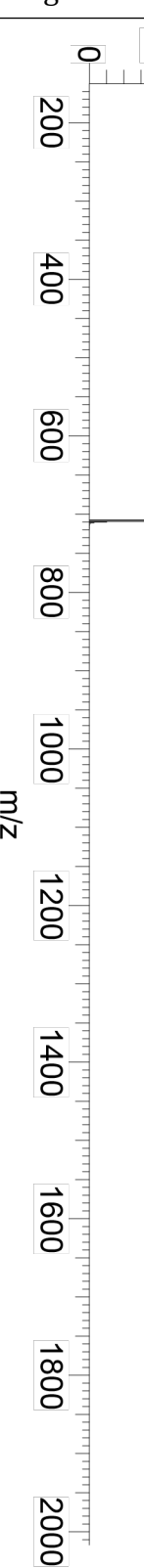


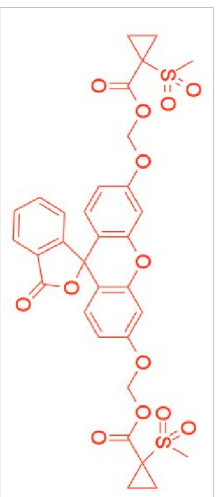
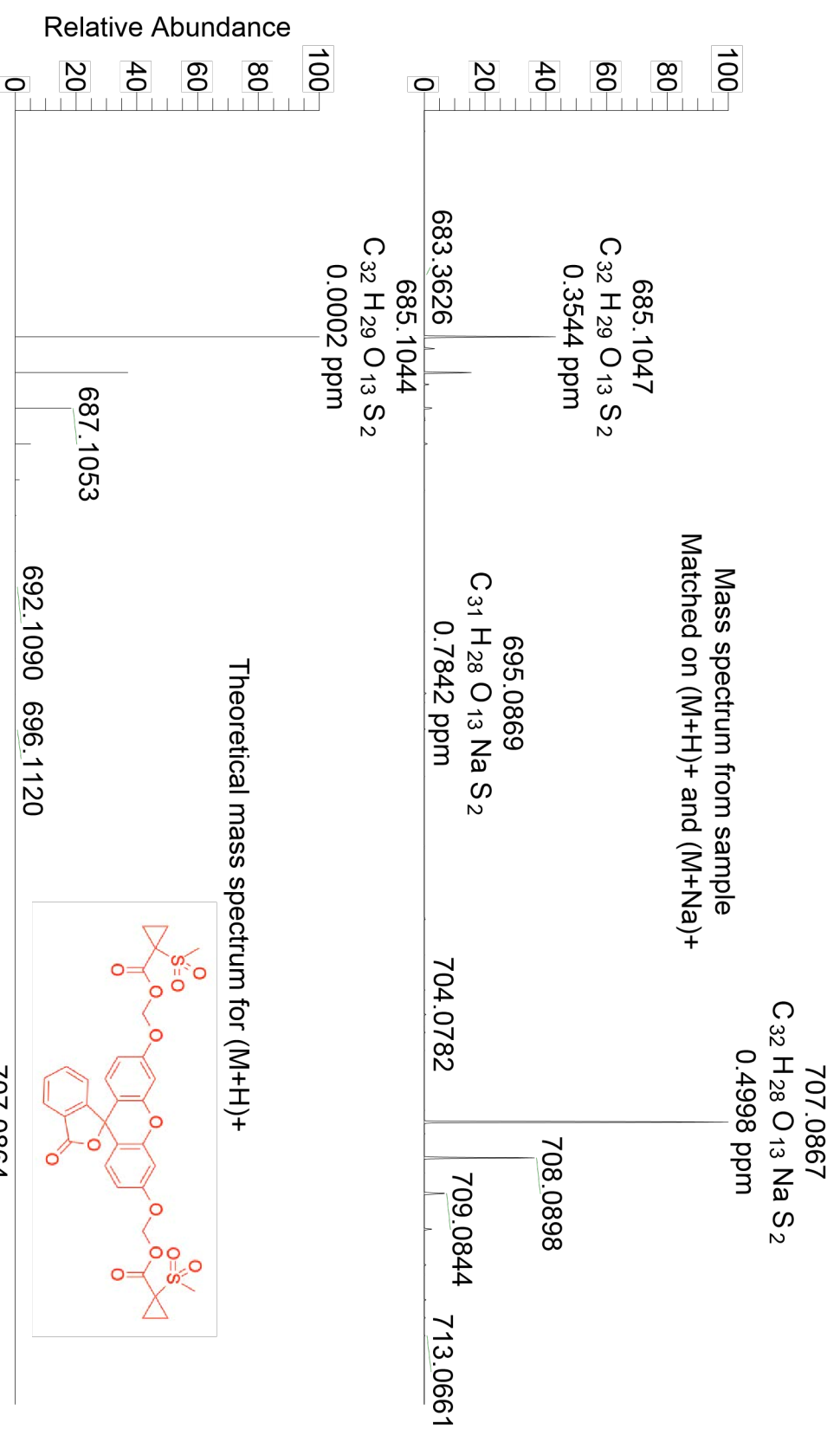
Theoretical mass spectrum for (M+H)⁺

NL:
6.17E5

C₃₂ H₂₈ O₁₃ S₂ Na:
C₃₂ H₂₈ O₁₃ S₂ Na₁
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+Na)⁺





NL:
9.26E6
BU-3#19-49 RT: 0.95-0.39
AV: 31 SB: 122 0.06-0.11
1.06-1.98 T: FTMS + p ESI
Full ms [150.00-2000.00]

NL:
6.17E5
C₃₂ H₂₈ O₁₃ S₂ H:
C₃₂ H₂₉ O₁₃ S₂
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

NL:
6.17E5
C₃₂ H₂₈ O₁₃ S₂ Na:
C₃₂ H₂₈ O₁₃ S₂ Na₁
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res. Pwr. @FWHM

Figure S105

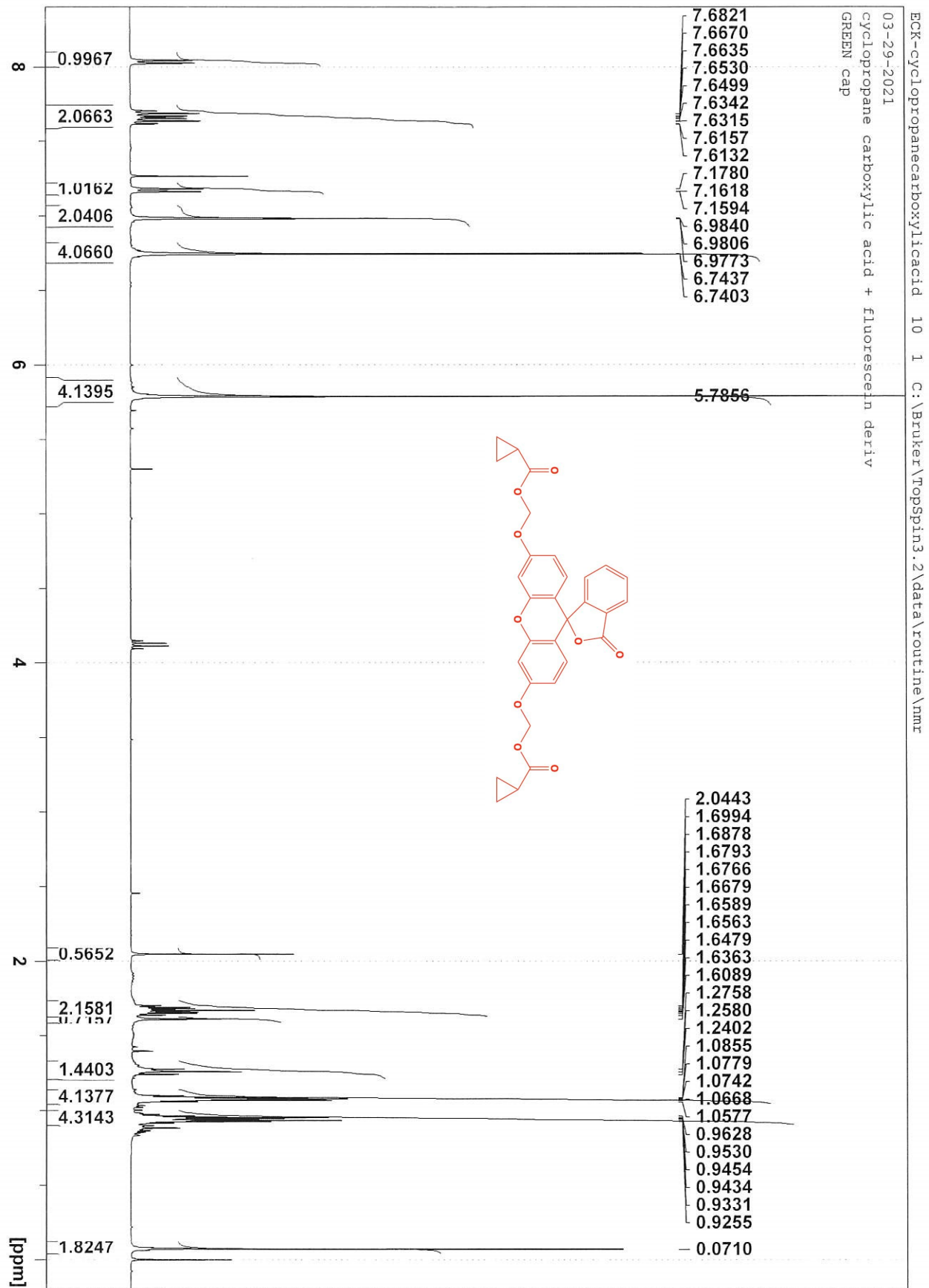


Figure S106

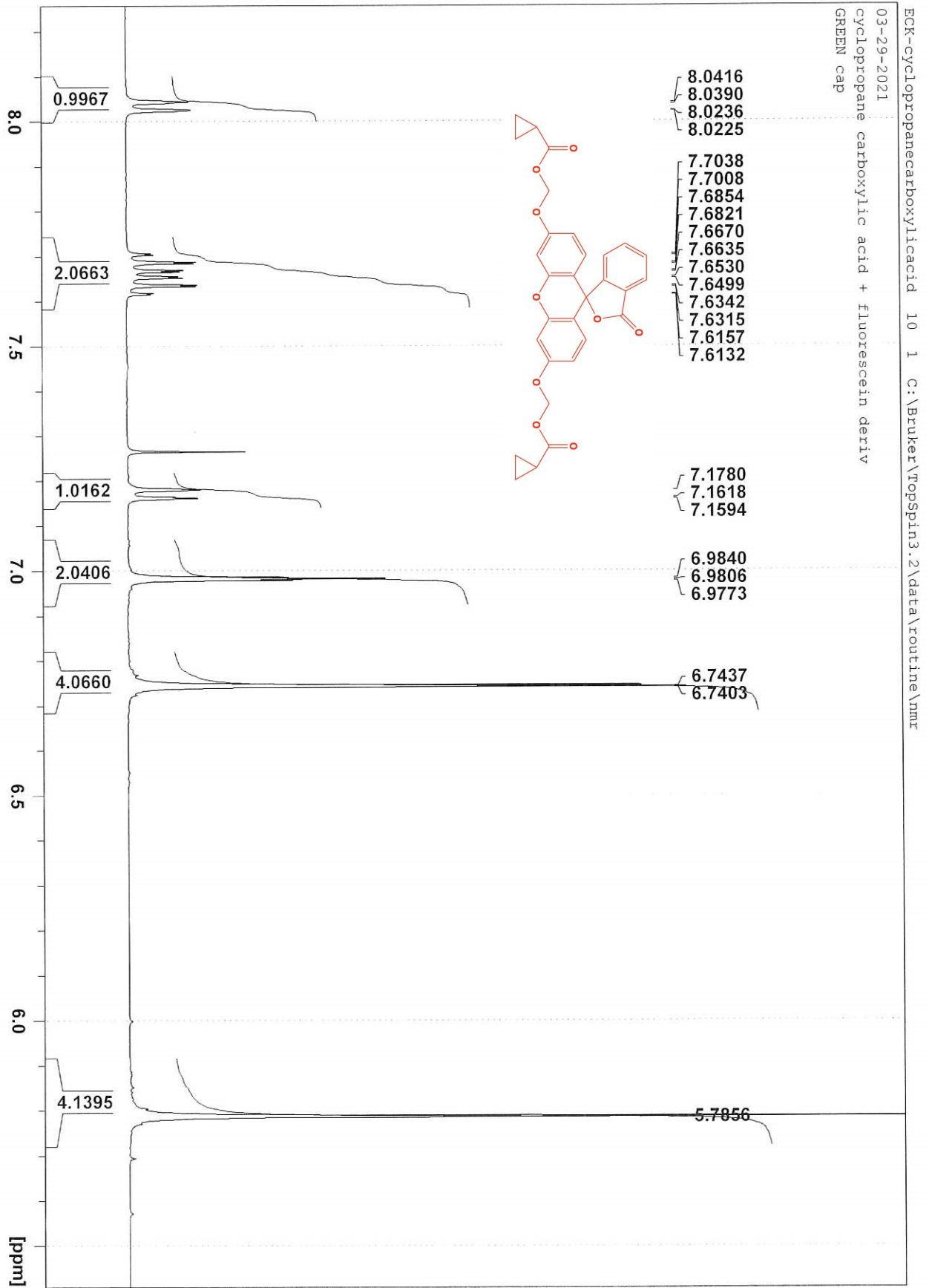


Figure S107

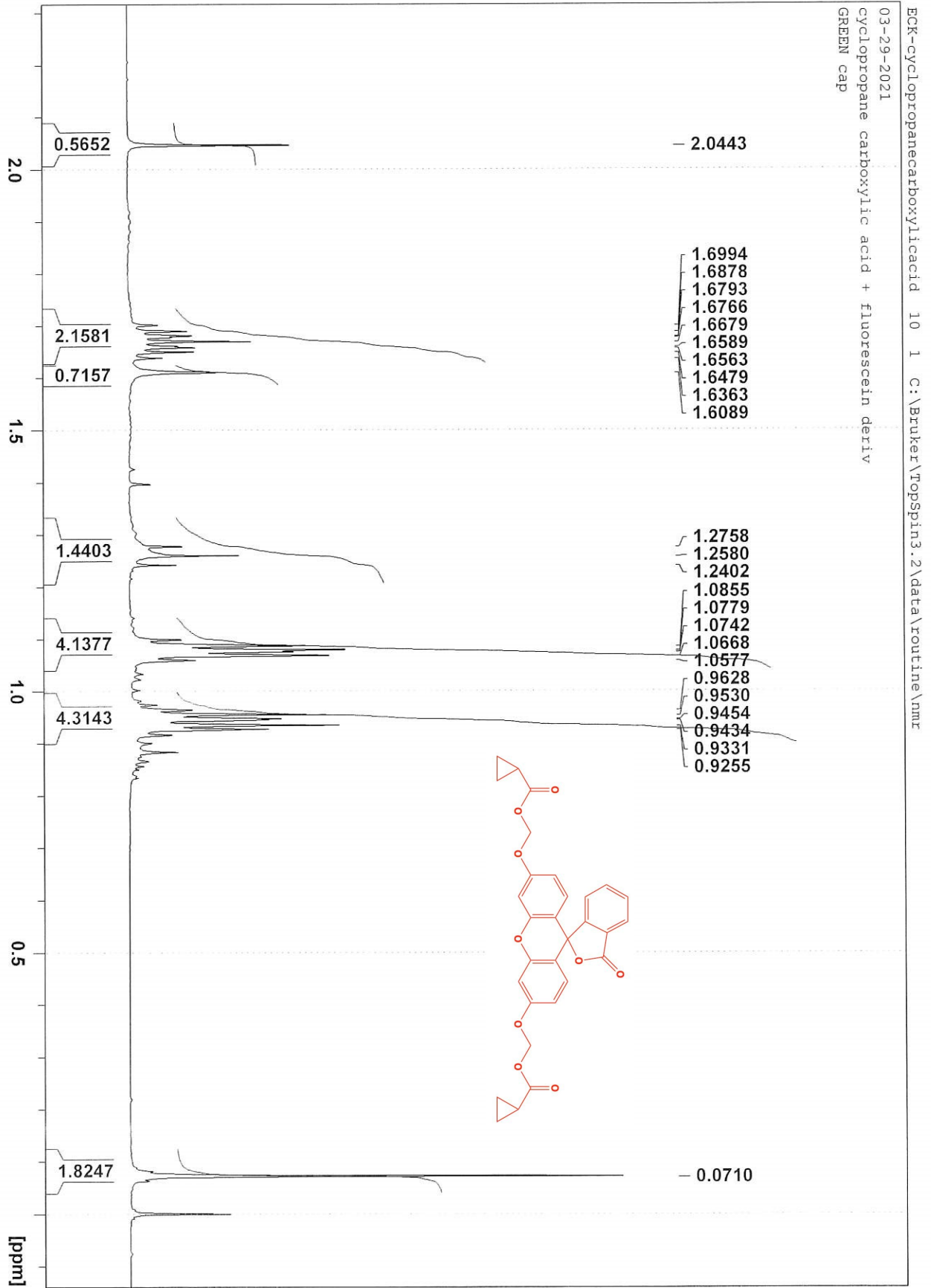


Figure S108

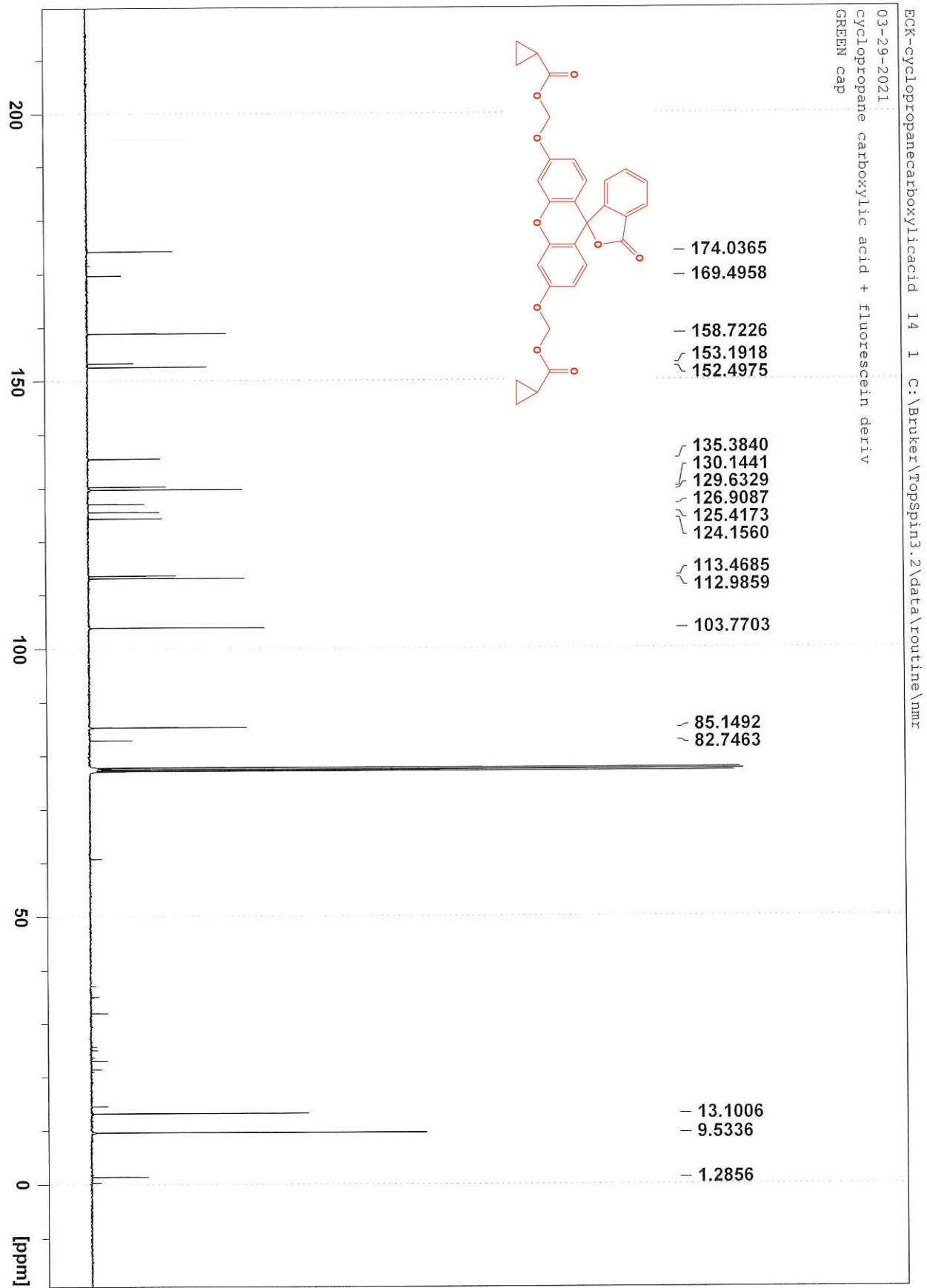


Figure S109

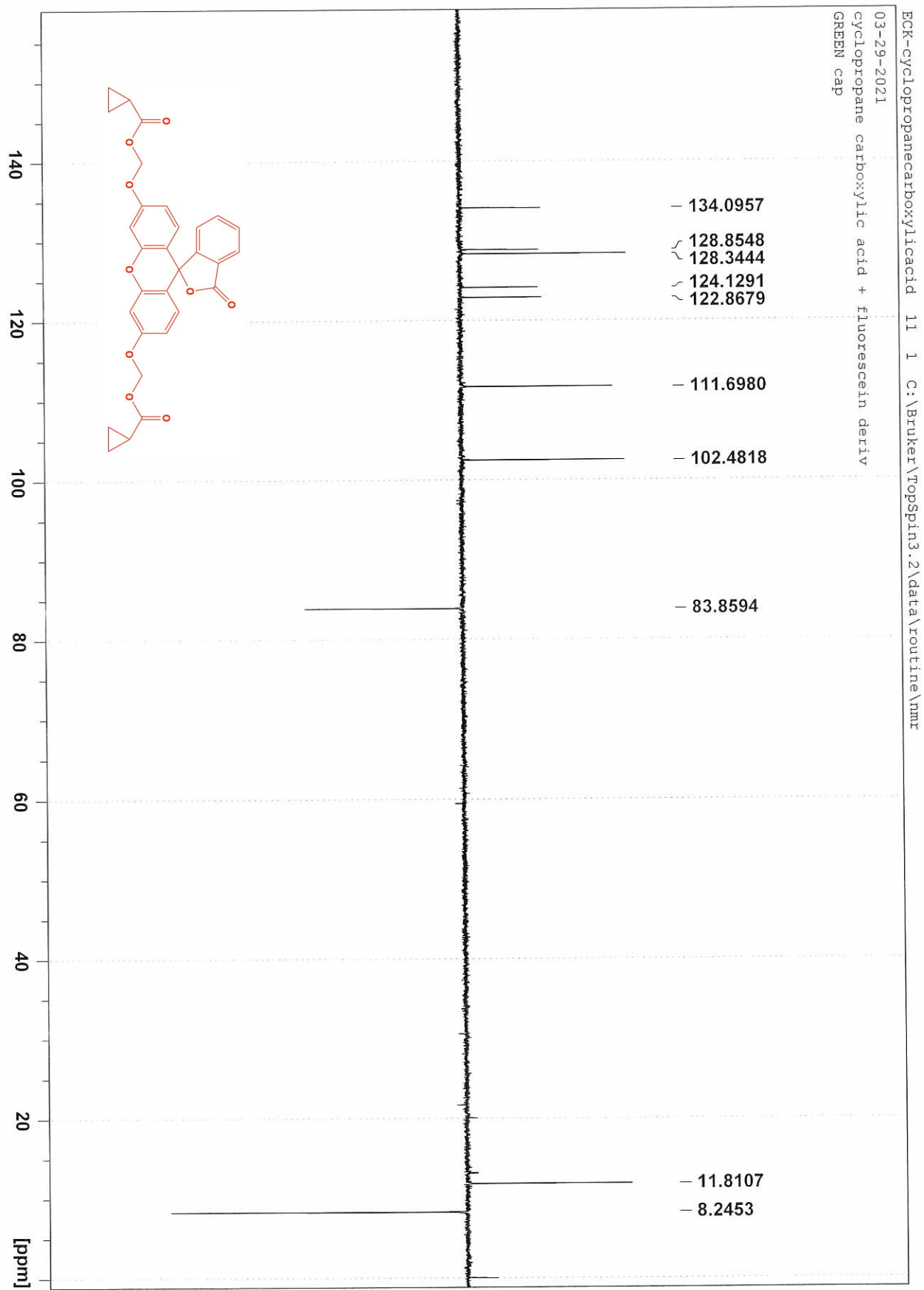


Figure S110

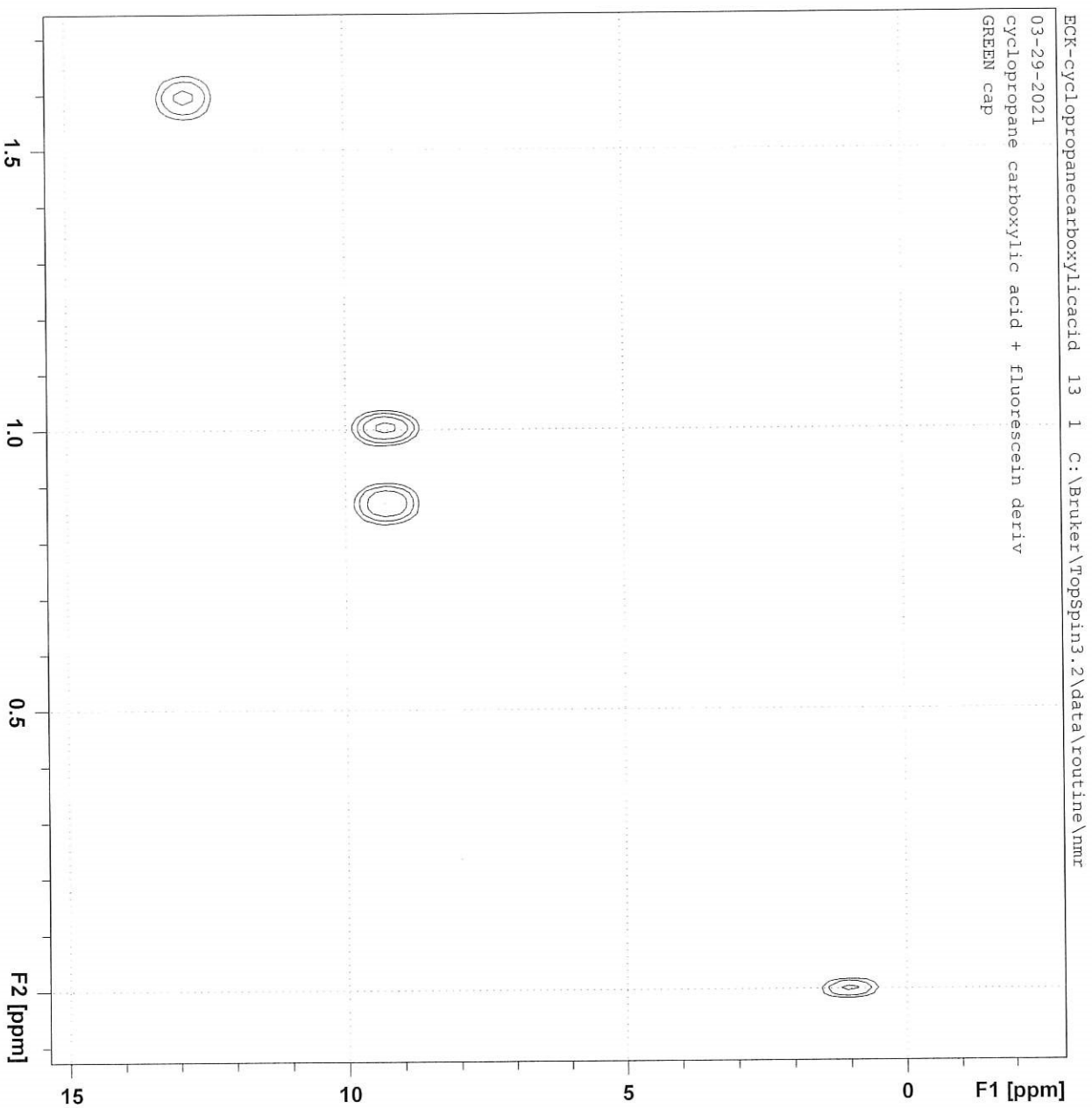
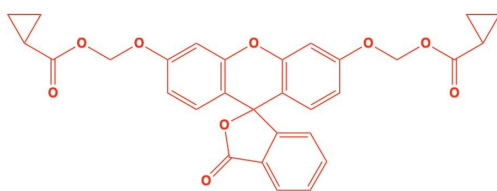
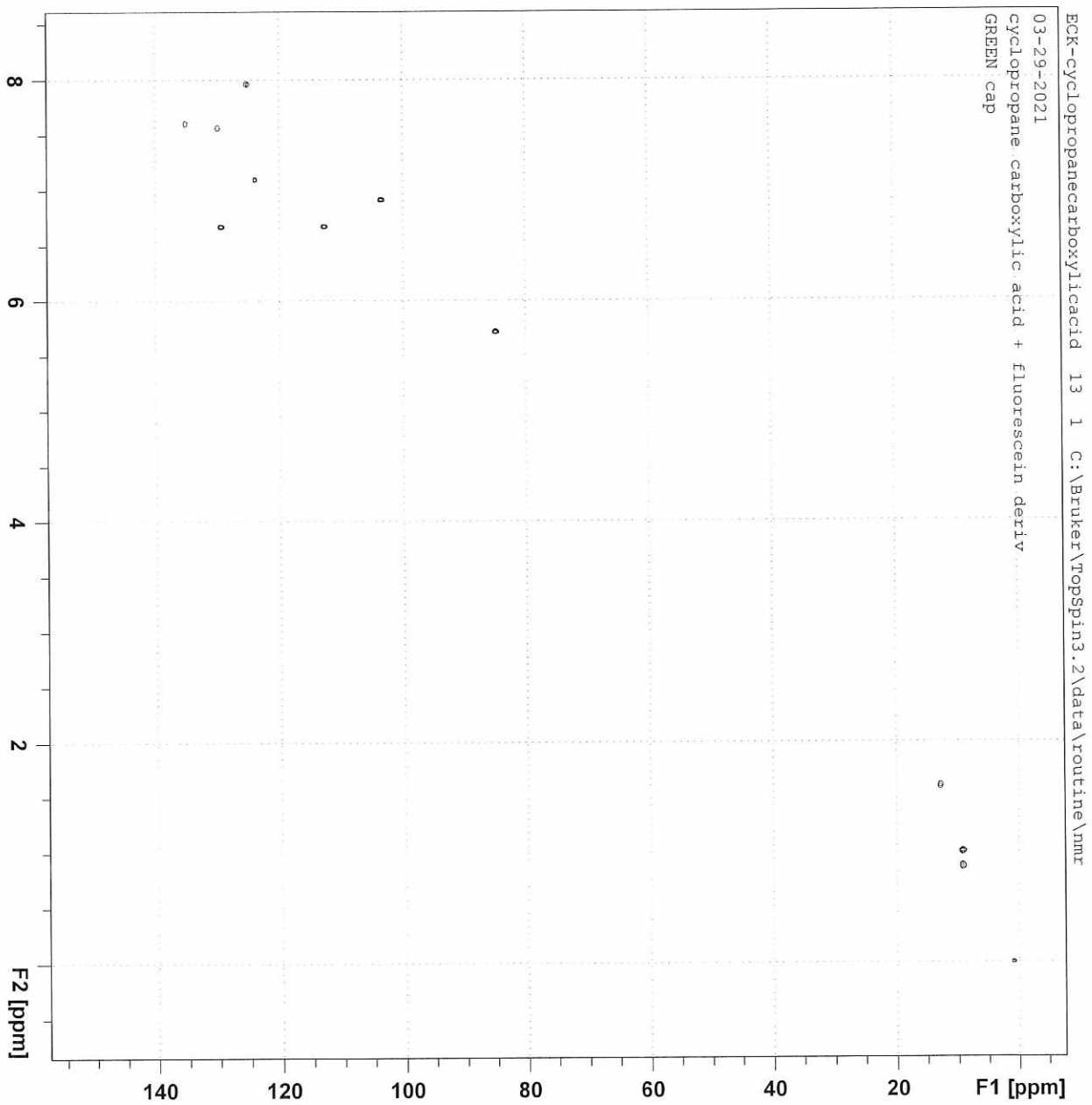
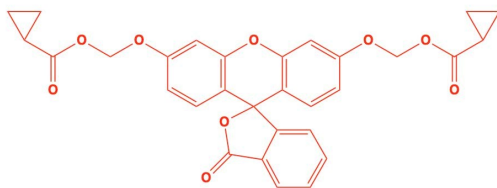


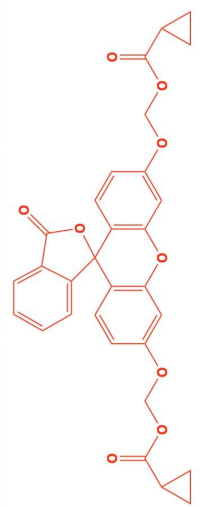
Figure S111



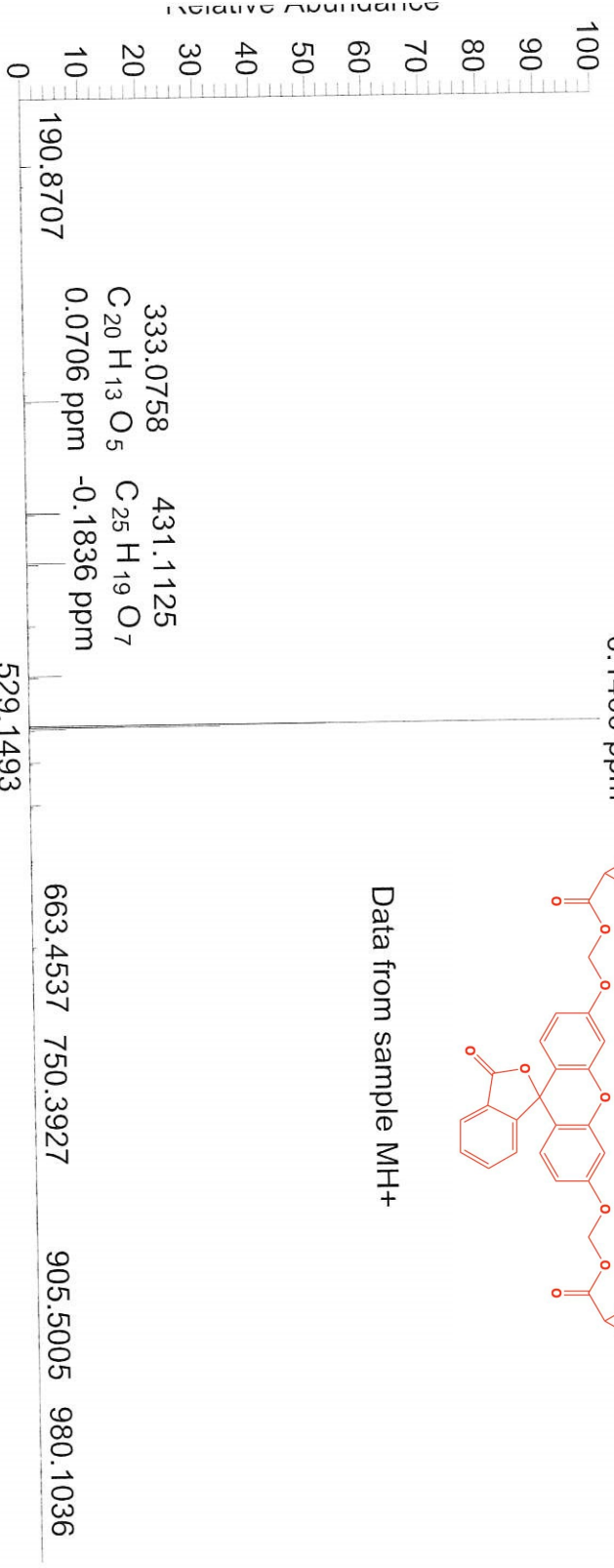
Cyclopropene Carboxylic acid

(of 1)

NL:
 3.45E8
 JE-7#26-29 RT: 0.23-0.26
 AV: 4 T: FTMS + p APCI
 corona Full ms
 [150.00-1800.00]



Data from sample MH+



NL:
 7.06E5

C₃₀ H₂₄ O₉ H:
 C₃₀ H₂₅ O₉
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res .Pwr . @FWHM

Theoretical for MH+

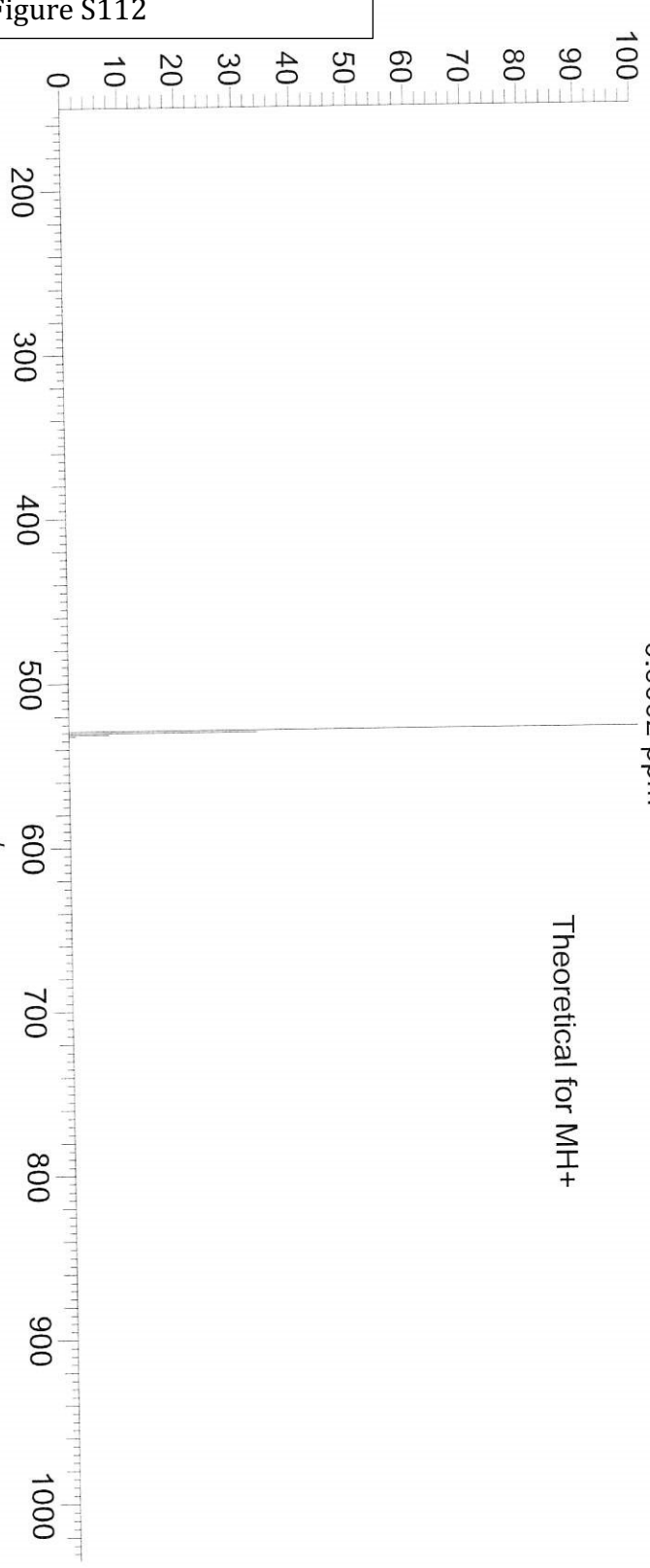
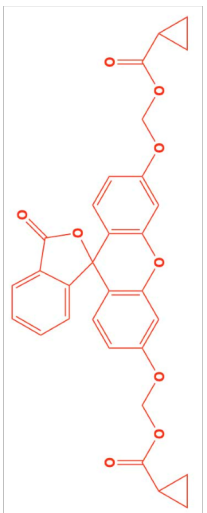
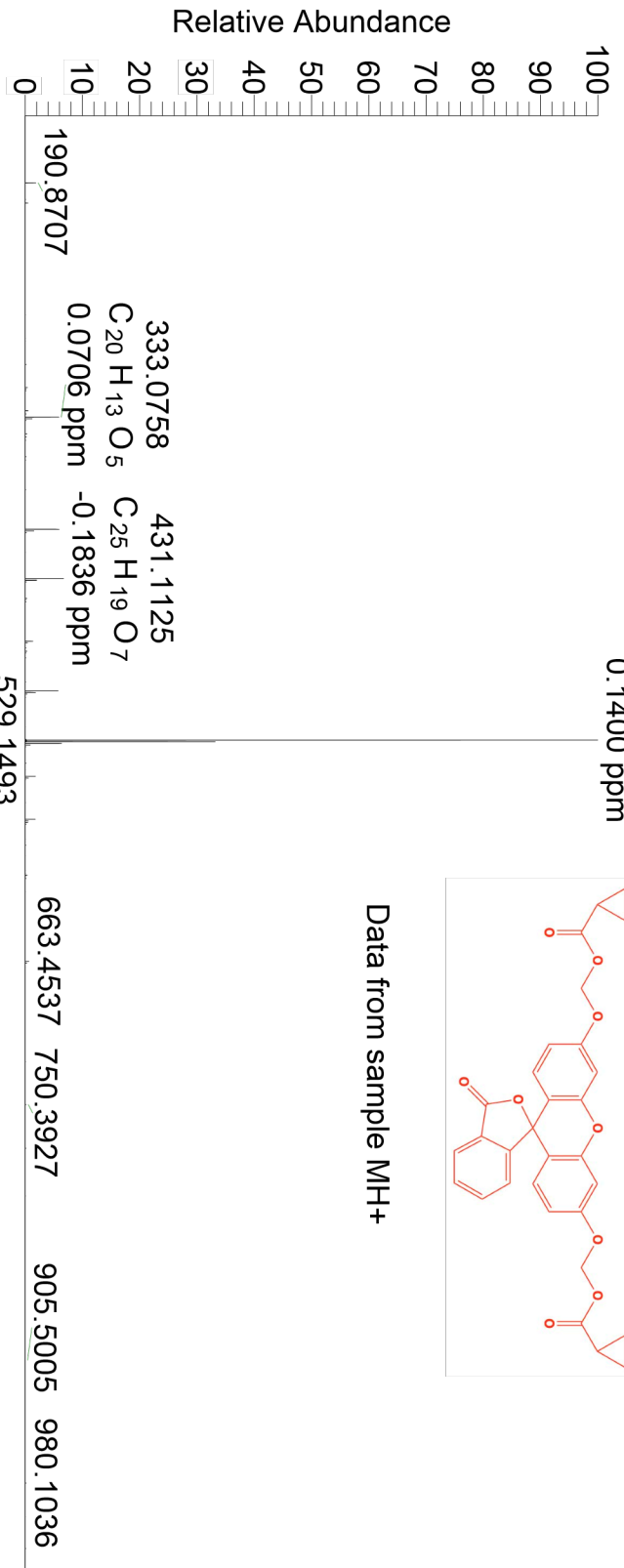


Figure S112



NL:
3.45E8
JE-7#26-29 RT: 0.29-0.26
AV: 4 T: FTMS + p AP CI
corona Full ms
[150.00-1800.00]

Data from sample MH+



NL:
7.06E5
C₃₀H₂₄O₉H:
C₃₀H₂₅O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Theoretical for MH+

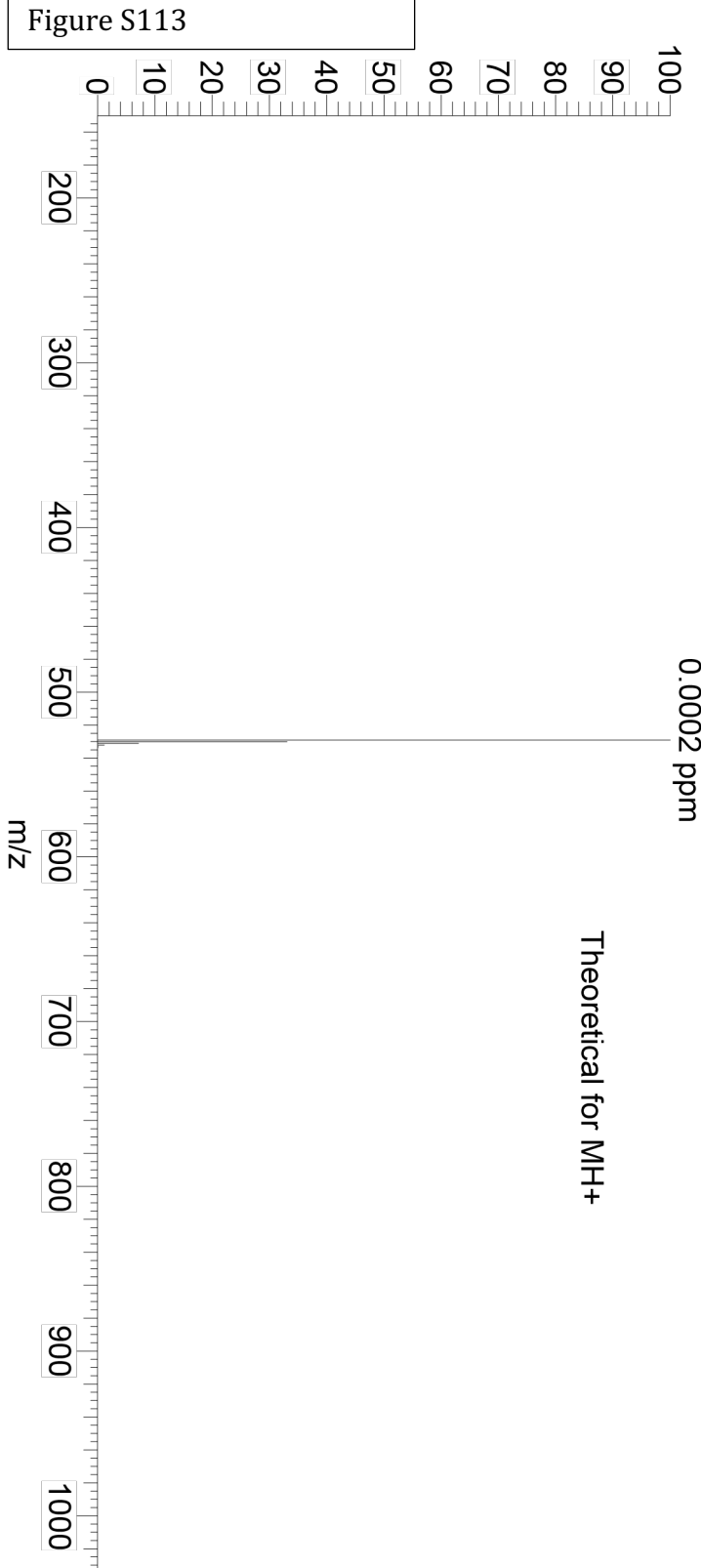


Figure S113

Figure S114

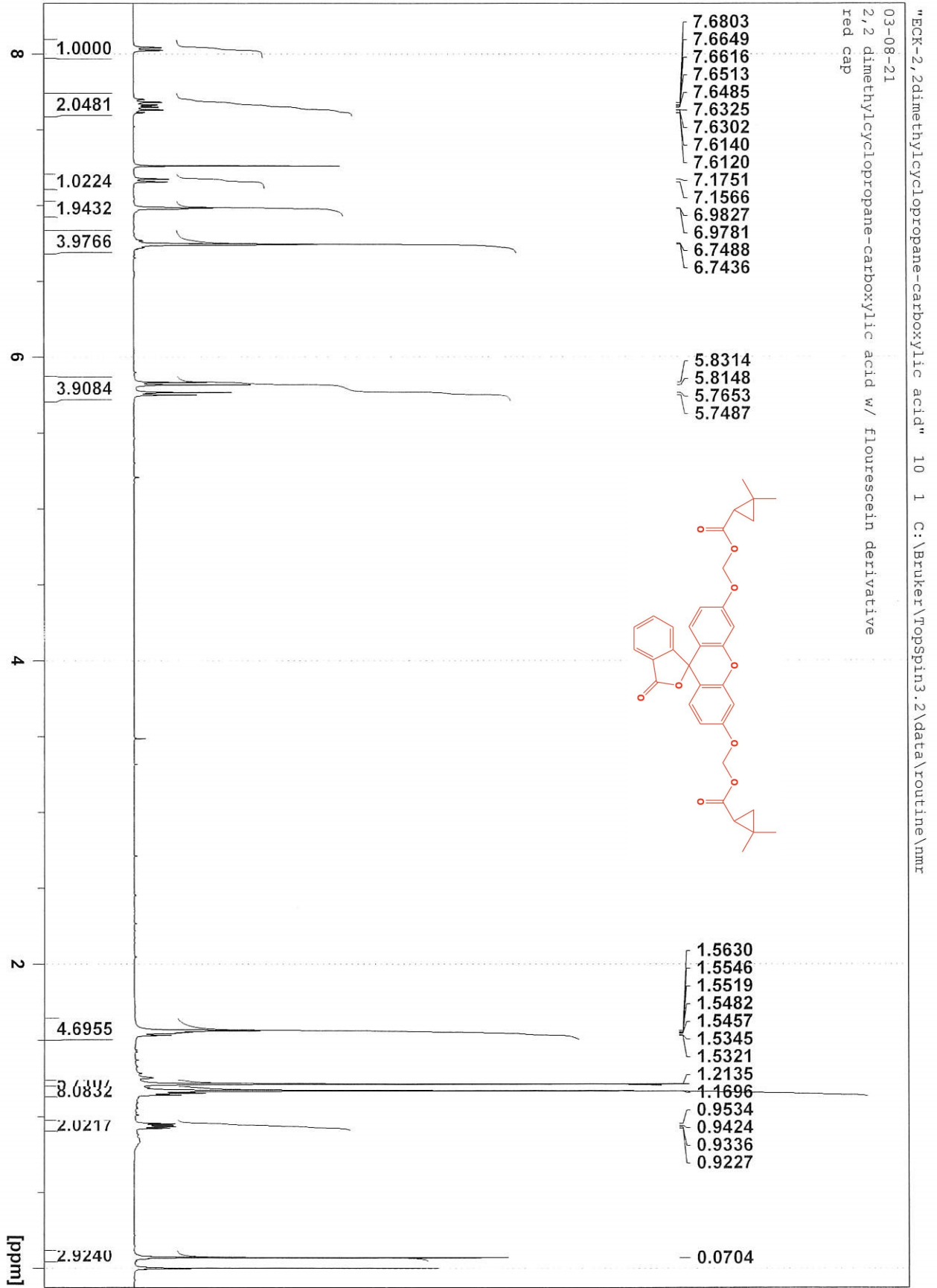


Figure S115

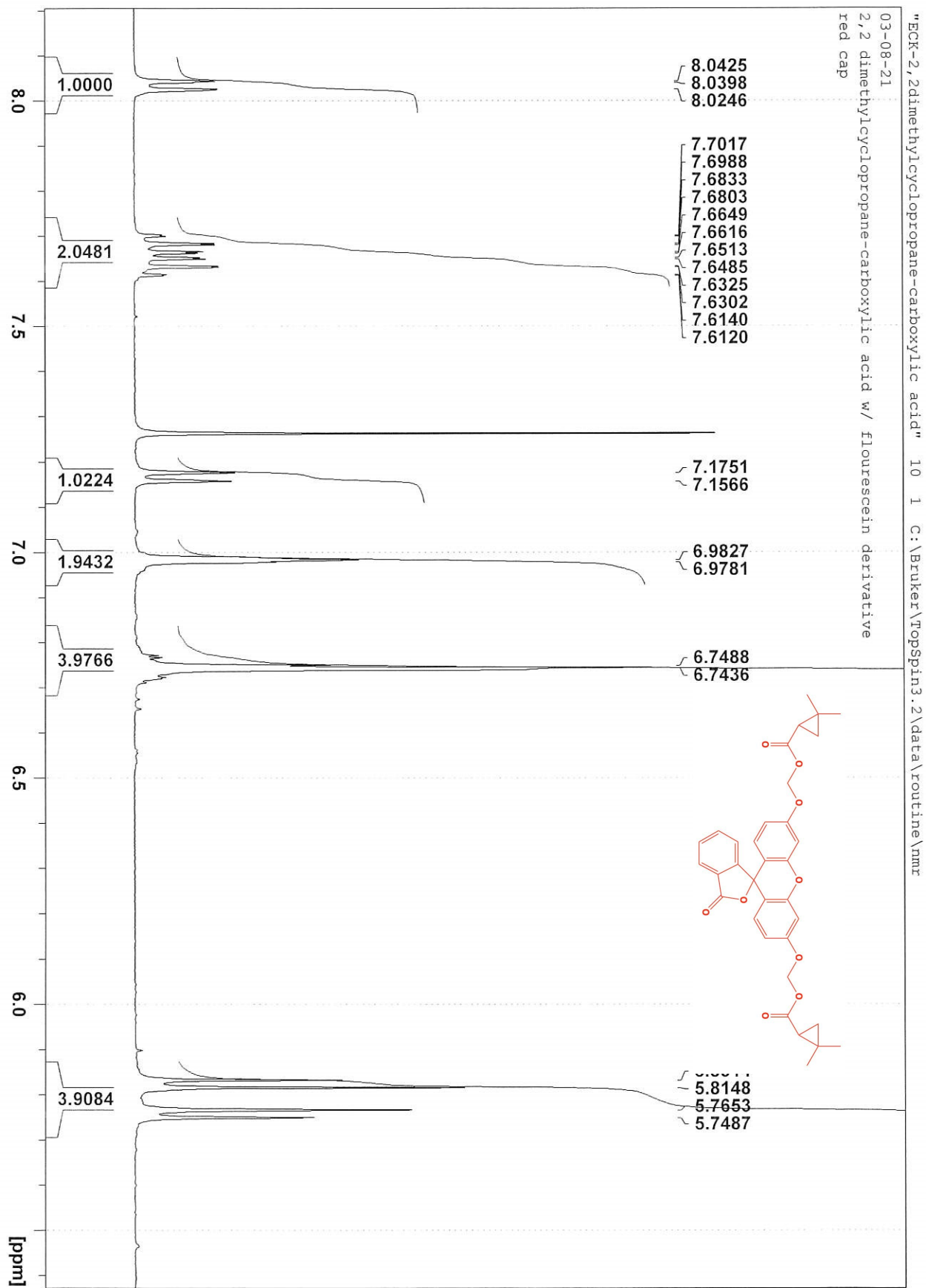


Figure S116

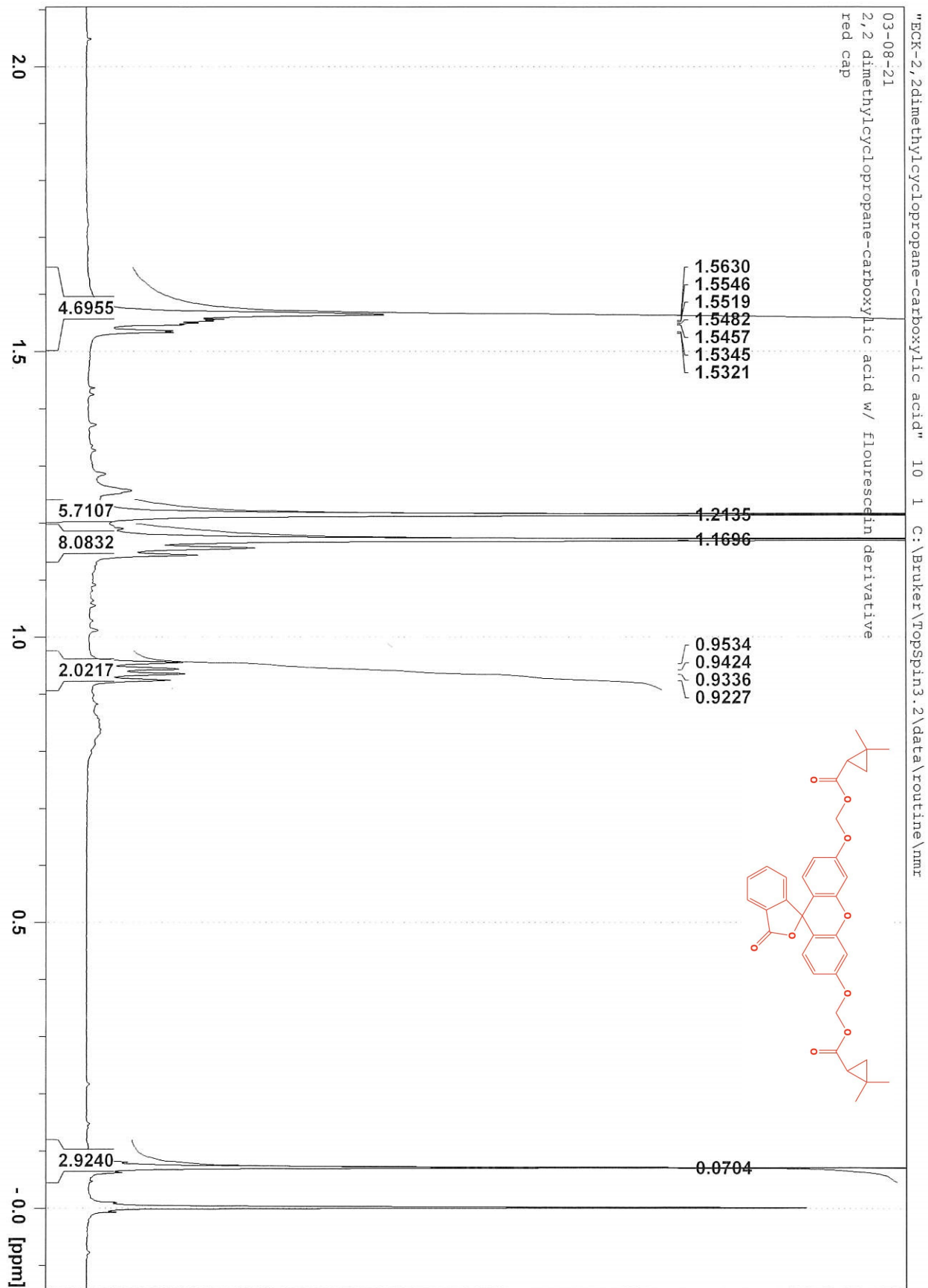


Figure S117

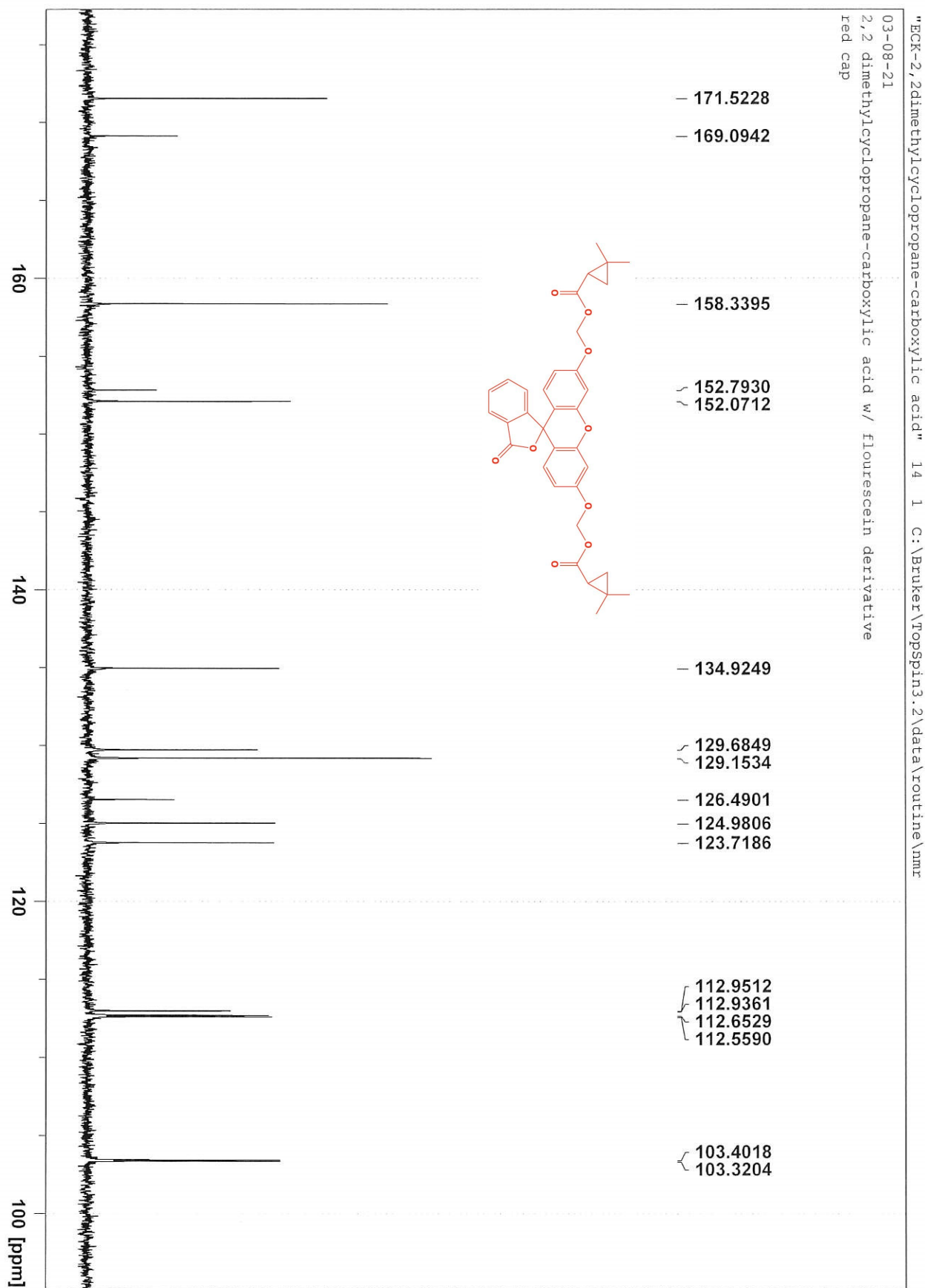


Figure S118

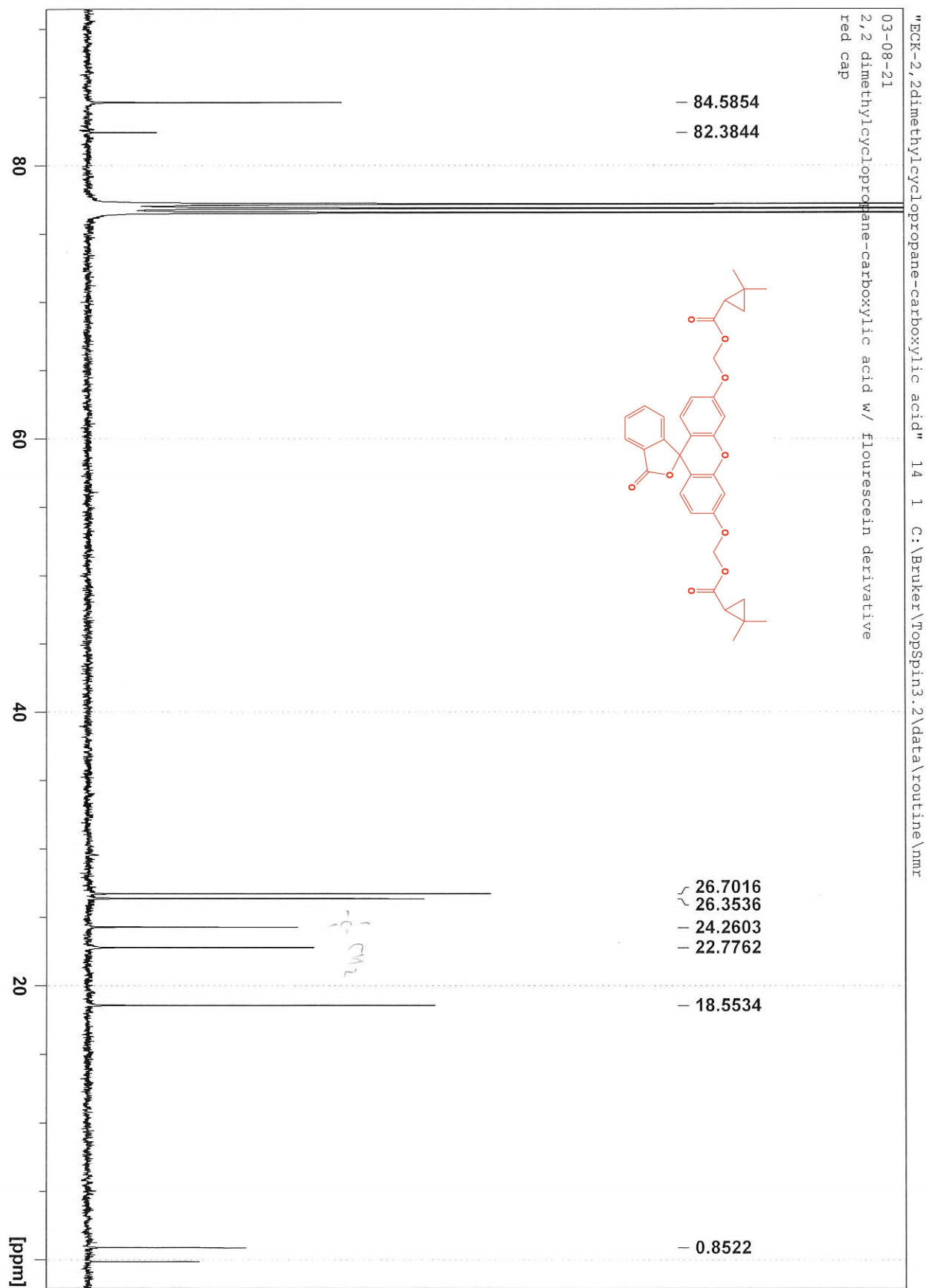


Figure S119

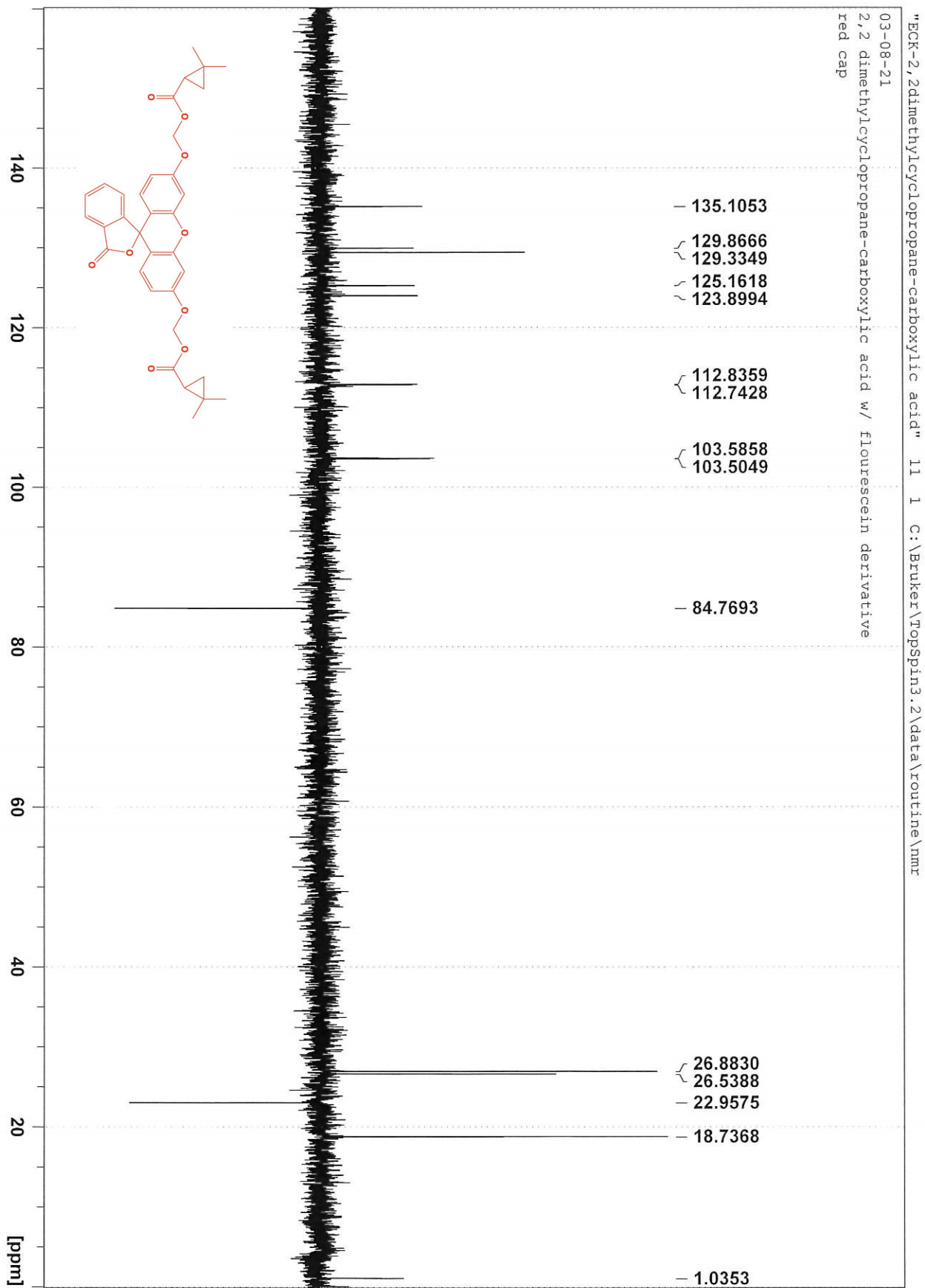


Figure S120

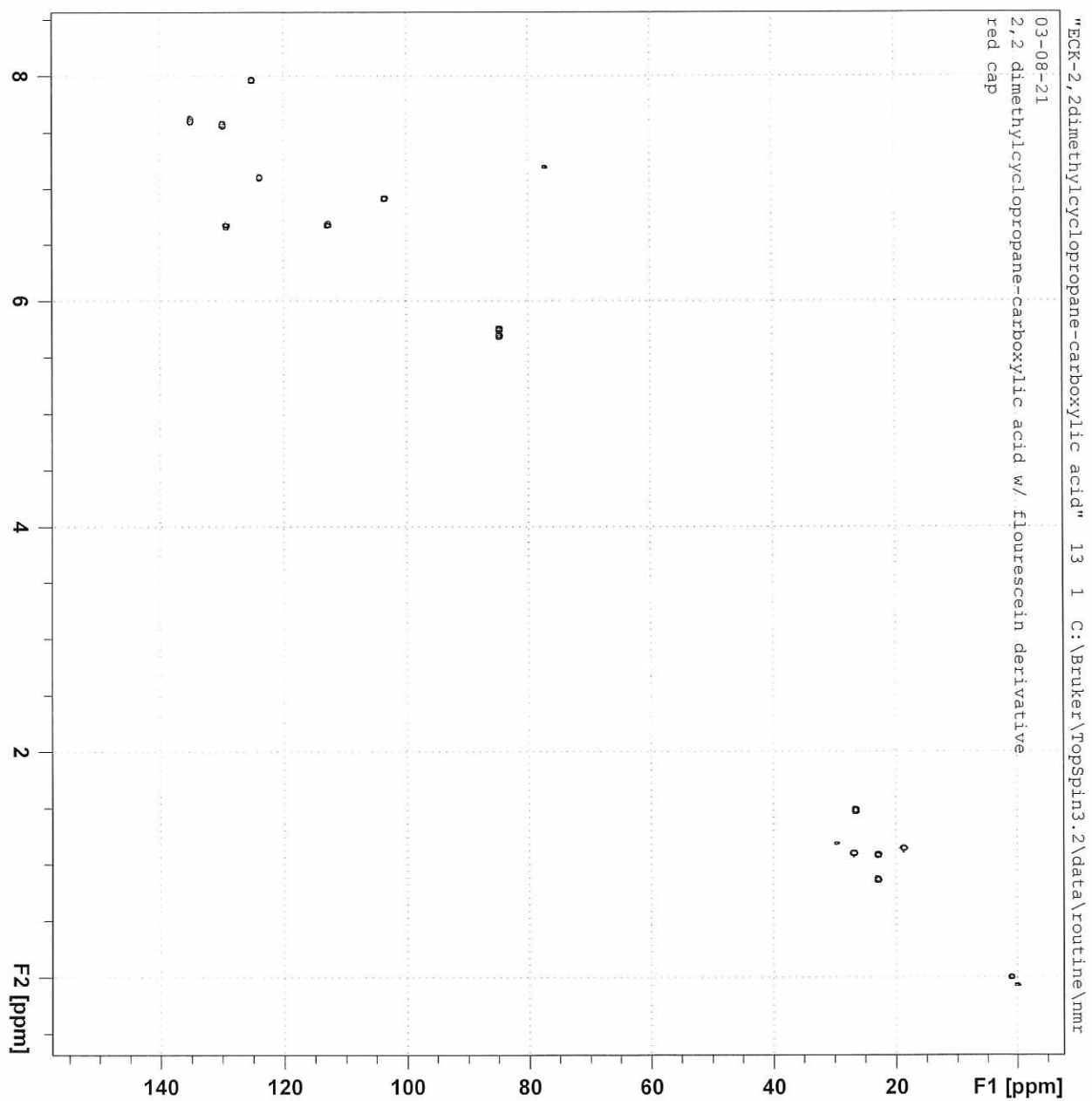
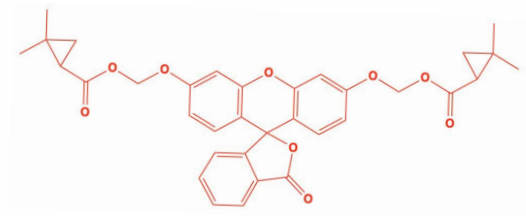


Figure S121

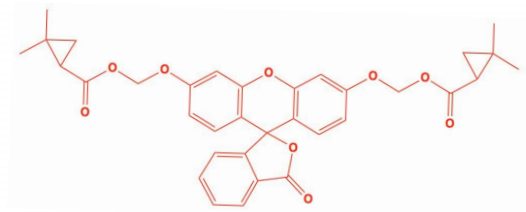
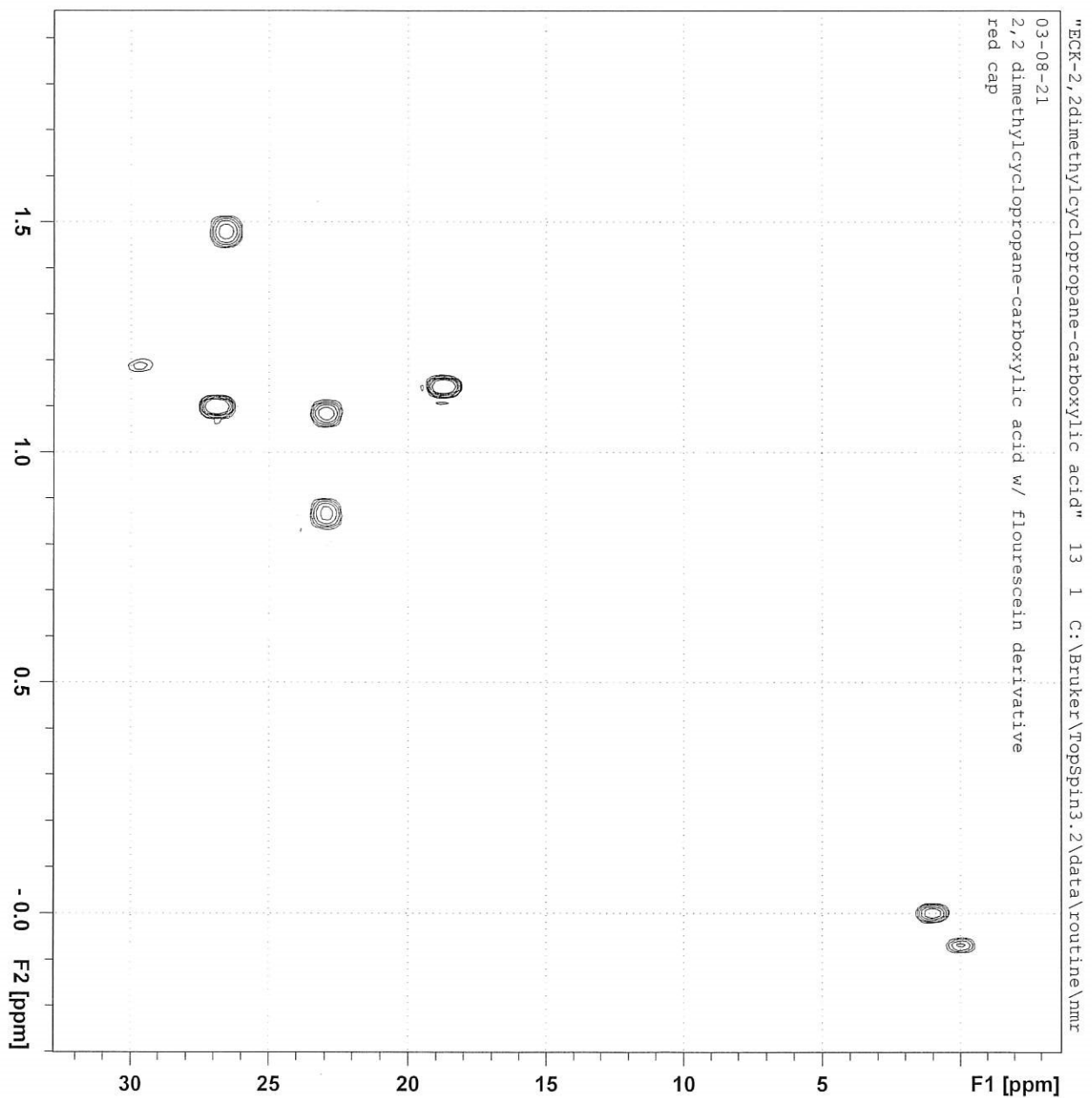
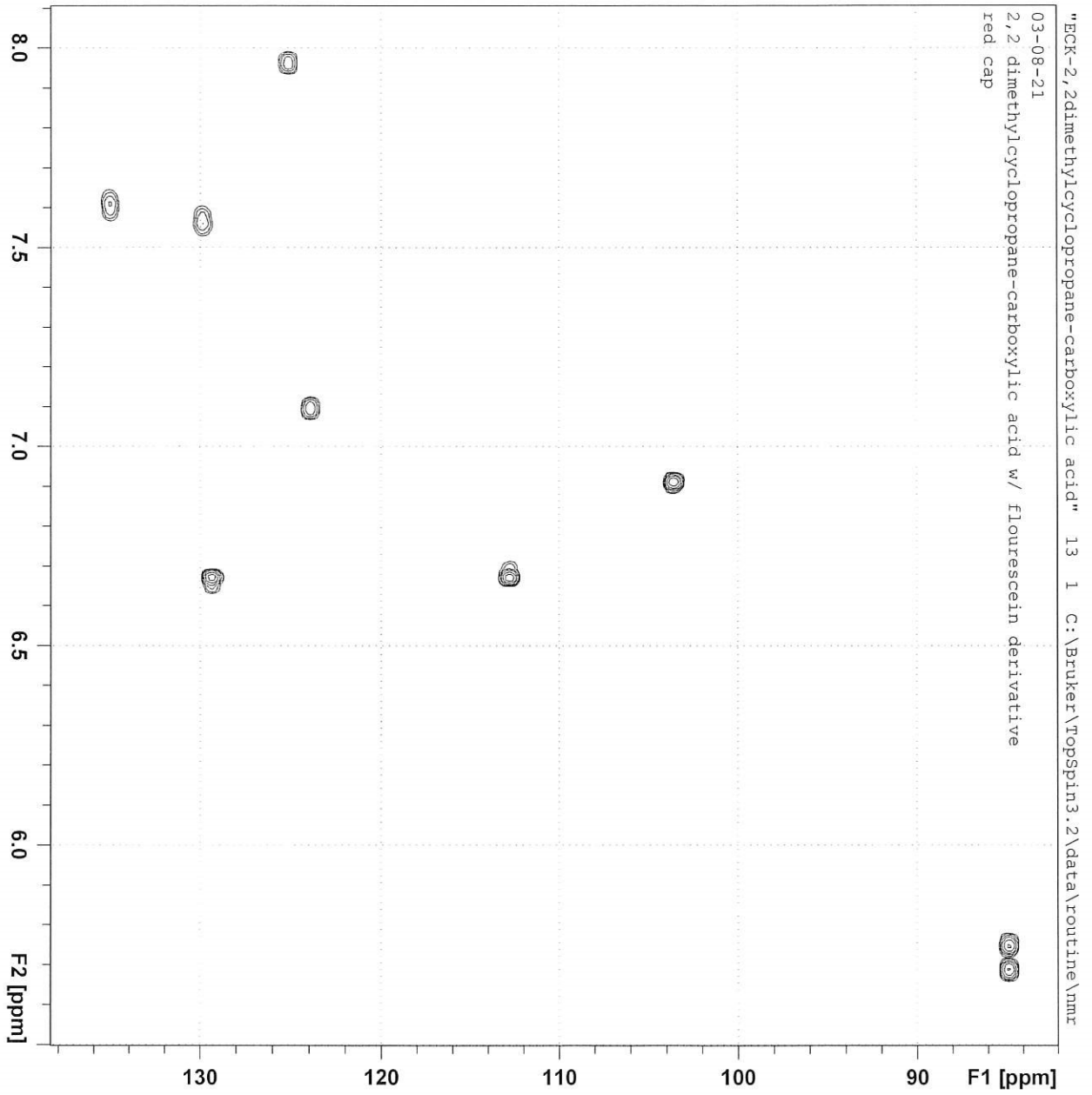
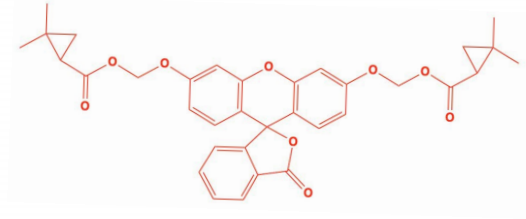
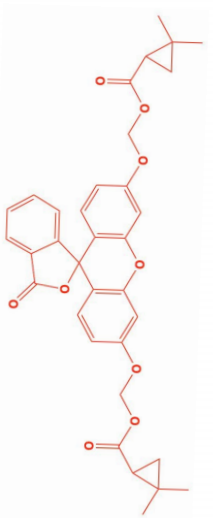


Figure S122

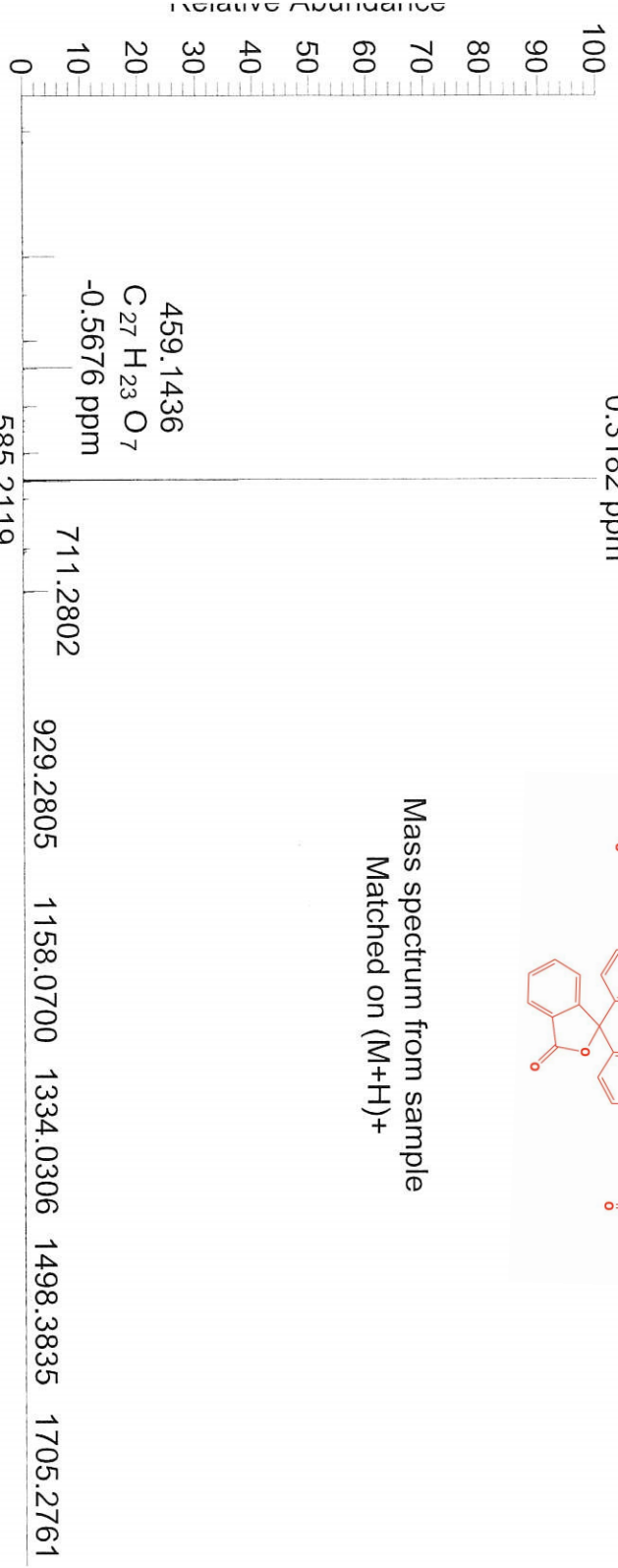


2,2-dimethylcyclopropane carboxylic acid



NL:
4.12E7
JE-4#18-21 RT: 0.16-0.19
AV: 4 T: FTMS + p APCI
corona Full ms
[150.00-1800.00]

Mass spectrum from sample
Matched on (M+H)+



NL:
6.76E5

C₃₄ H₃₂ O₉ H:
C₃₄ H₃₃ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Theoretical mass spectrum for (M+H)+

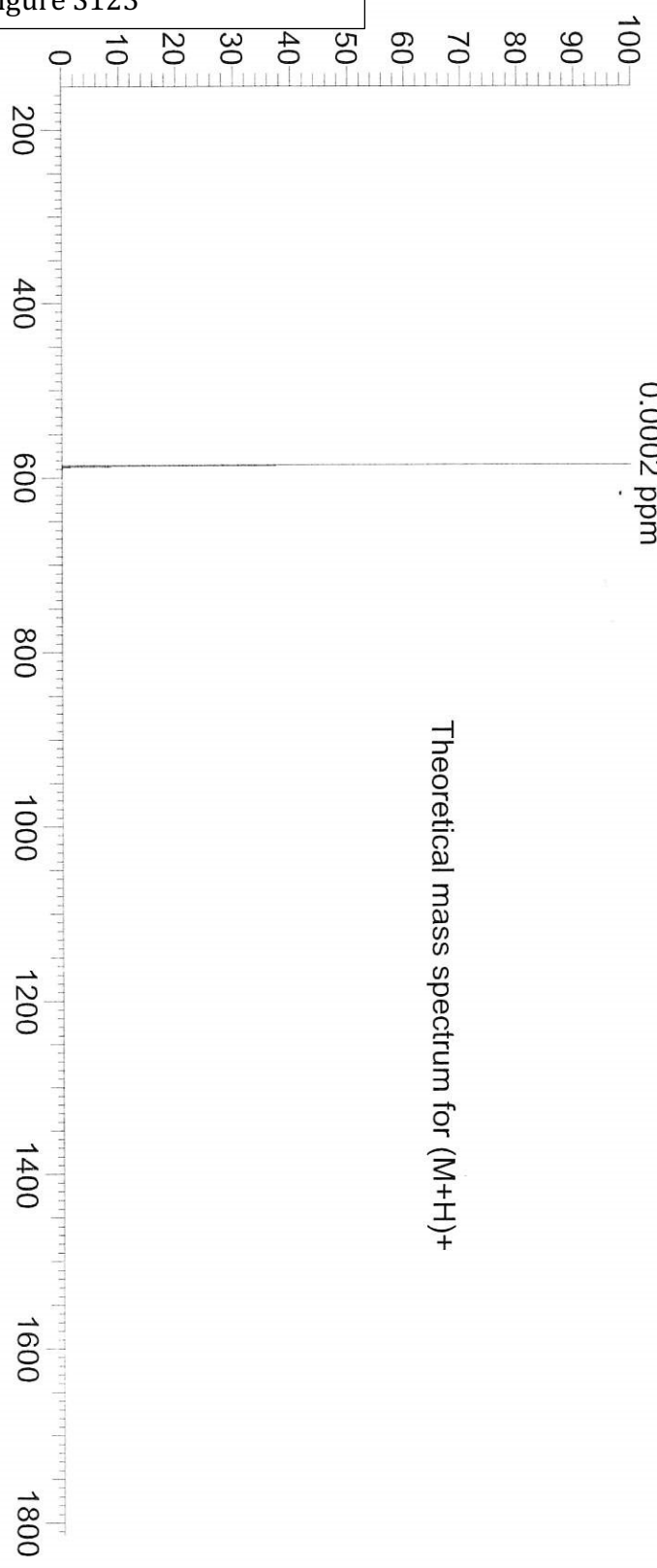


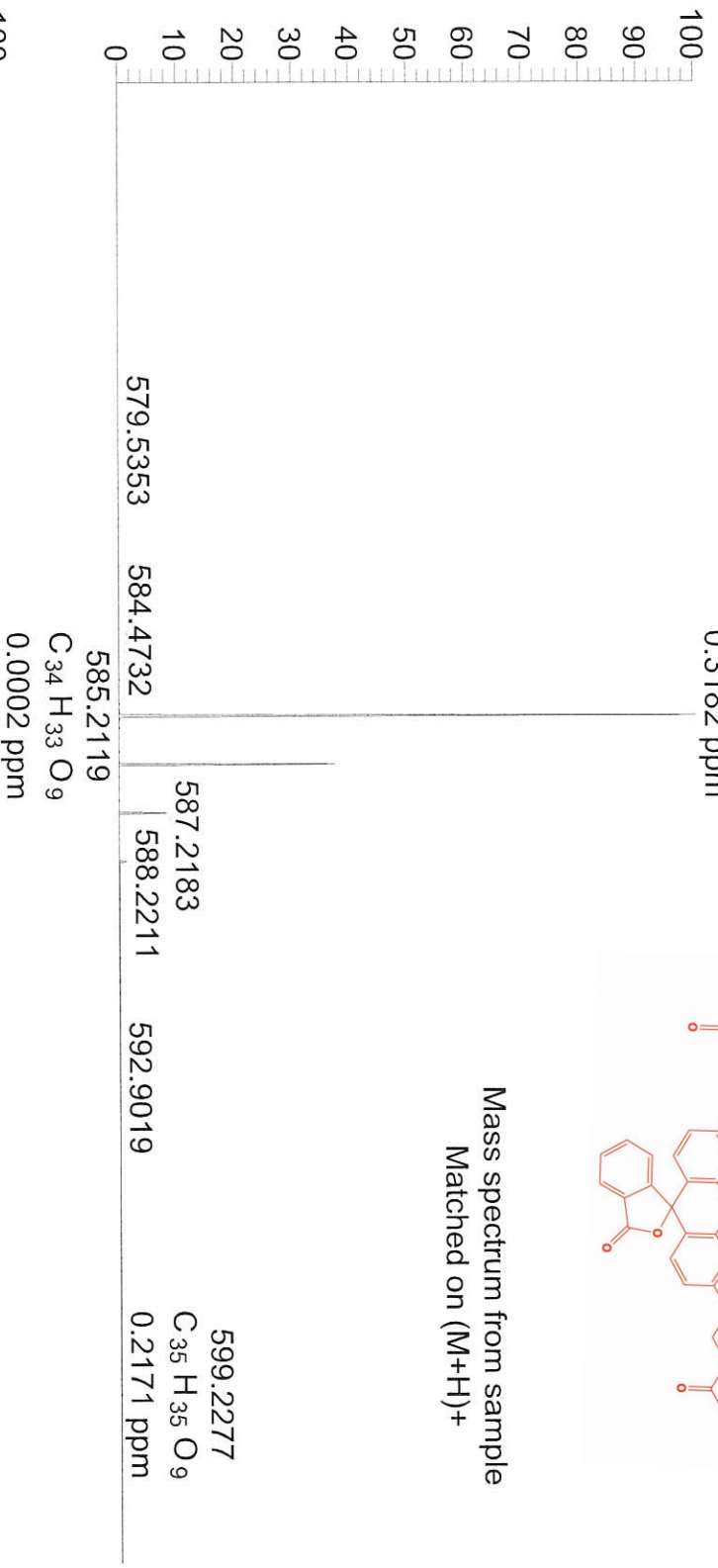
Figure S123

2,2-dimethylcyclopropane Carboxylic acid



NL:
4.12E7
JE-4#18-21 RT: 0.16-0.19
AV: 4 T: FTMS + p APCI
corona Full ms
[150.00-1800.00]

Mass spectrum from sample
Matched on (M+H)⁺



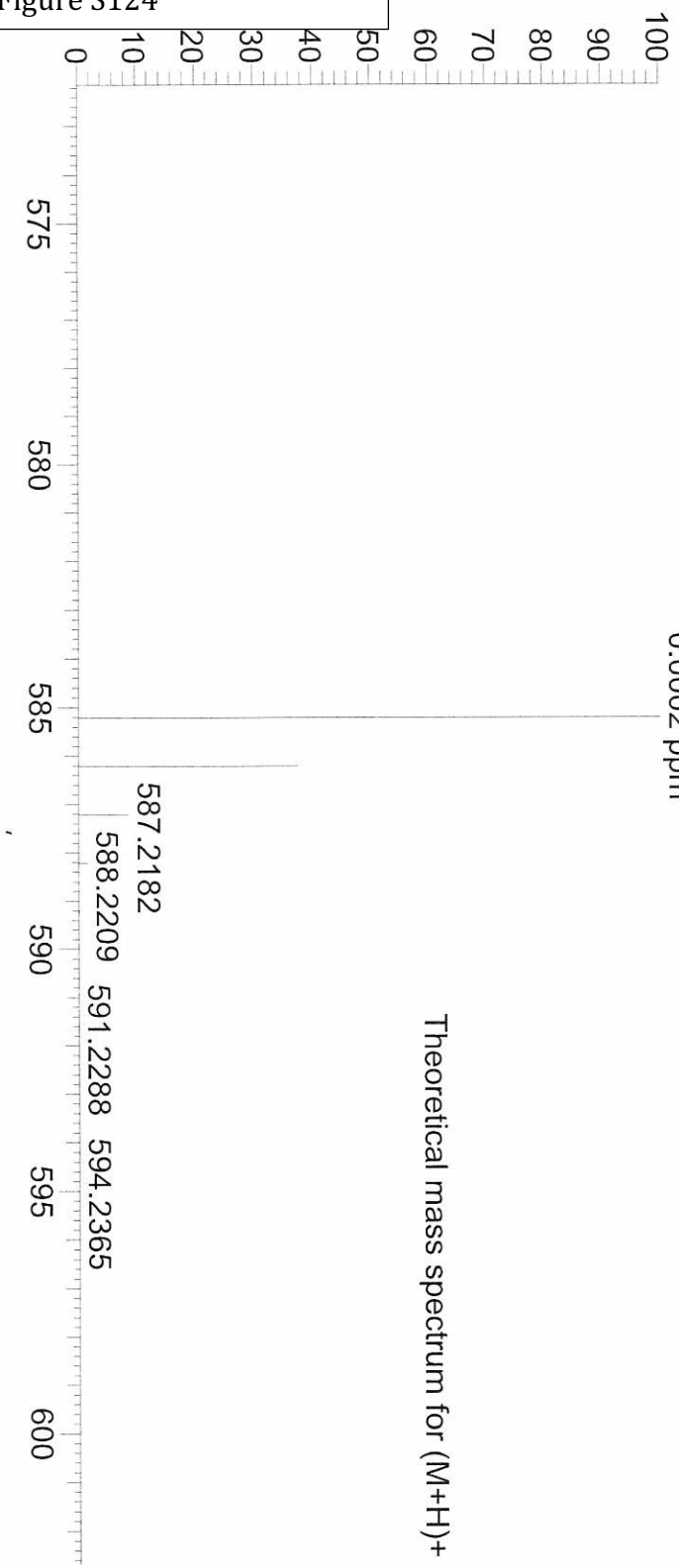
585.2119
C₃₄ H₃₃ O₉
0.0002 ppm

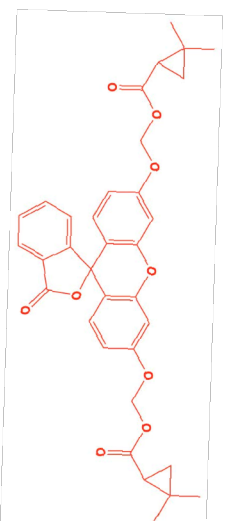
599.2277
C₃₅ H₃₅ O₉
0.2171 ppm

Theoretical mass spectrum for (M+H)⁺

NL:
6.76E5
C₃₄ H₃₂ O₉ H:
C₃₄ H₃₃ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

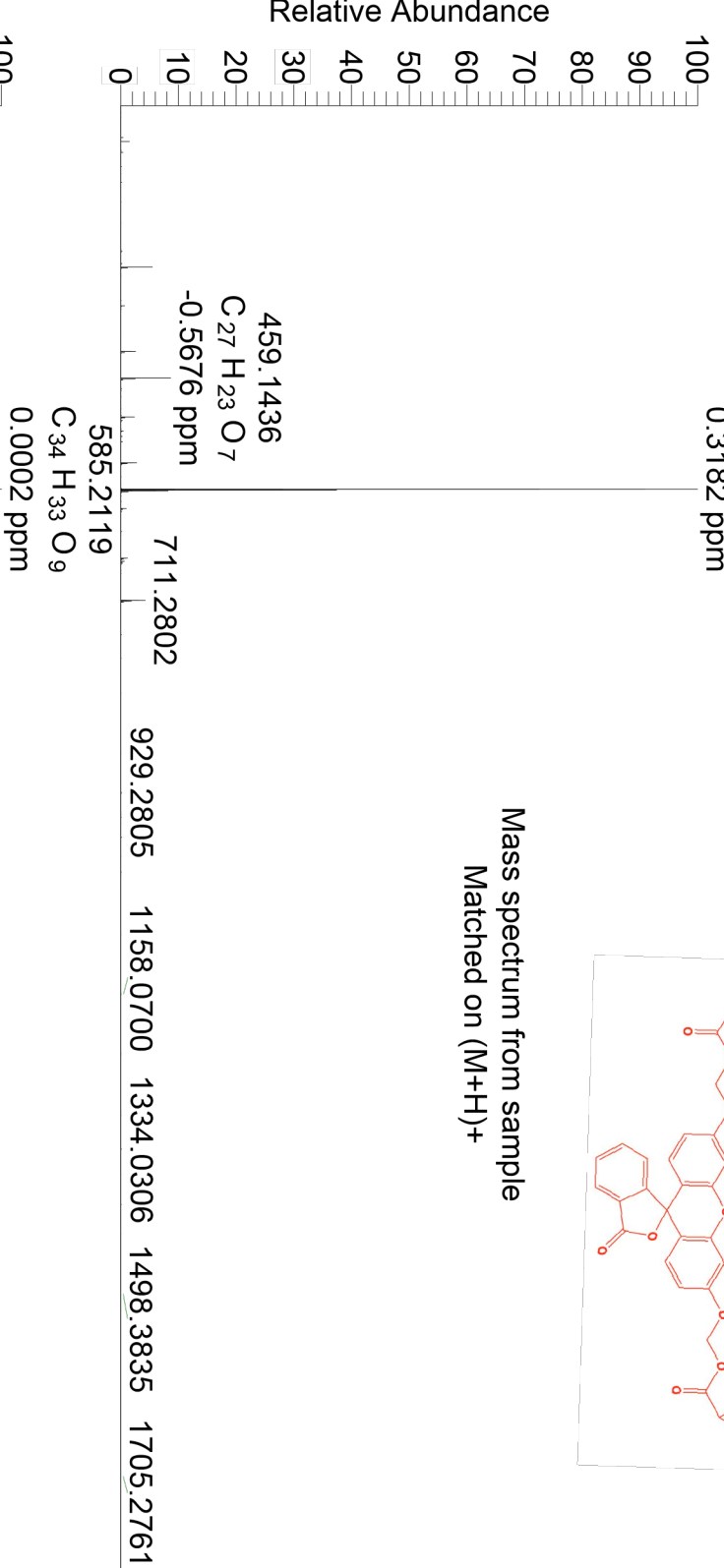
Figure S124





NL:
 4.12E7
 JE-4#18-21 RT: 0.18-0.19
 AV: 4 T: FTMS + p AP CI
 corona Full ms
 [150.00-1800.00]

Mass spectrum from sample
 Matched on (M+H)⁺



NL:
 6.76E5

C₃₄ H₃₂ O₉ H:
 C₃₄ H₃₃ O₉
 c (gss, s /p:40)(Val) Chrg 1
 R: 20000 Res. Pwr. @FWHM

Theoretical mass spectrum for (M+H)⁺

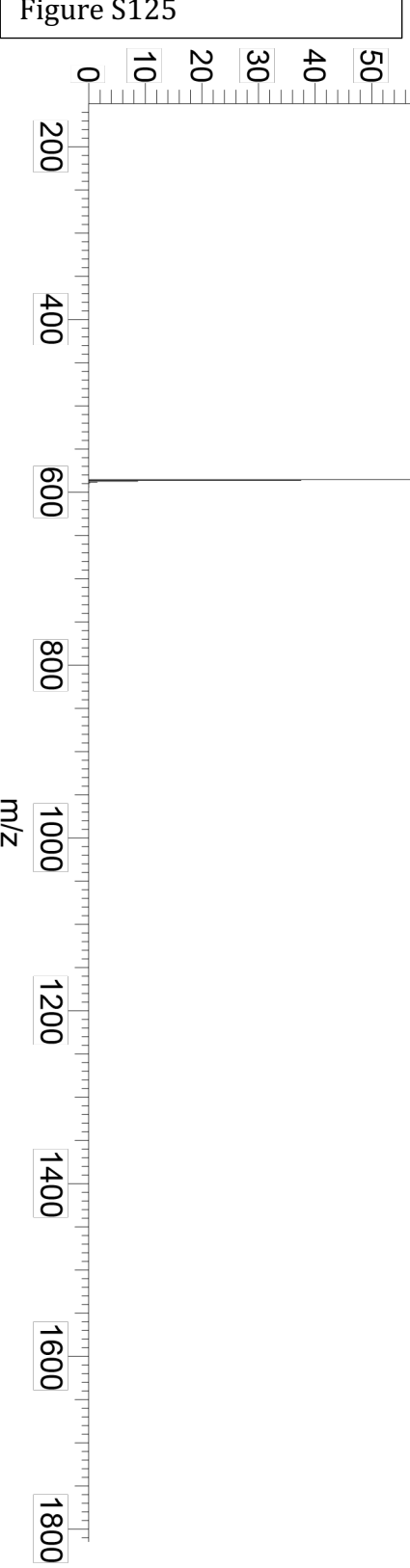
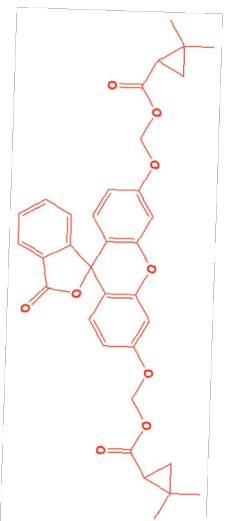
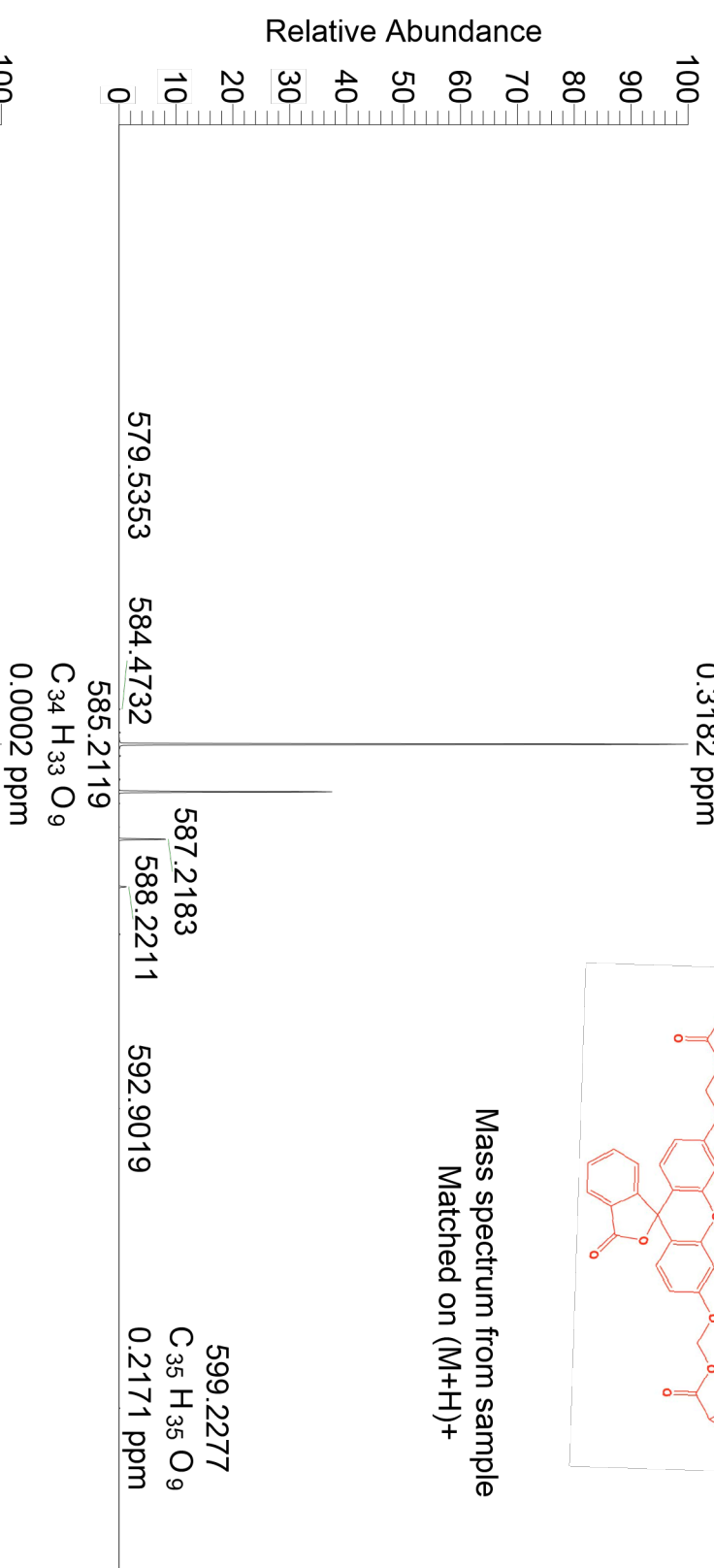


Figure S125



NL:
4.12E7
JE-4#18-21 RT: 0.16-0.19
AV: 4 T: FTMS + p APPI
corona Full ms
[150.00-1800.00]

Mass spectrum from sample
Matched on (M+H)+



NL:
6.76E5

C₃₄ H₃₂ O₉ H:
C₃₄ H₃₃ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Theoretical mass spectrum for (M+H)+

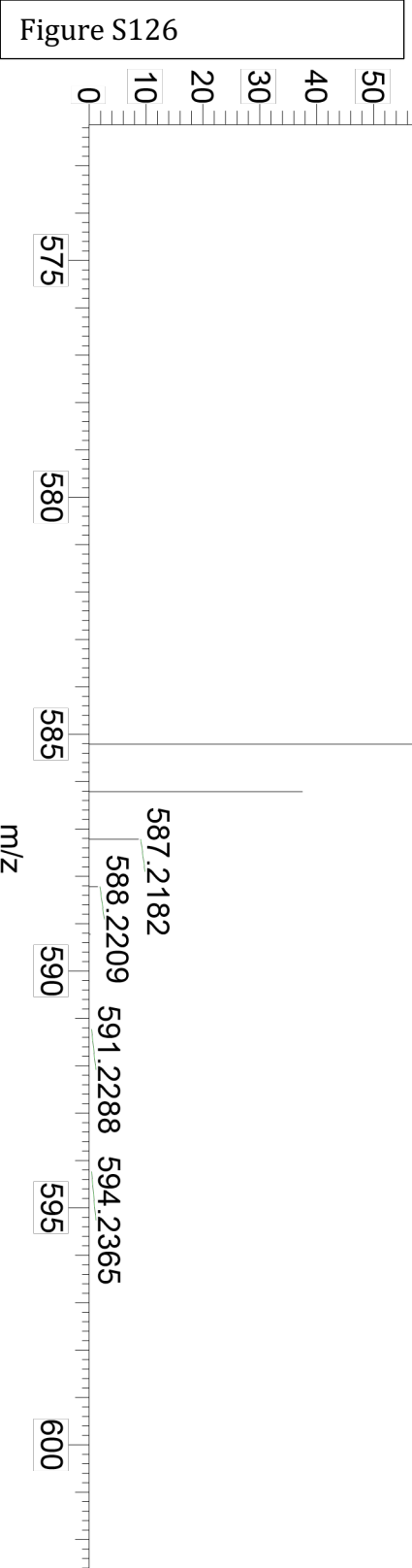


Figure S126

Figure S127

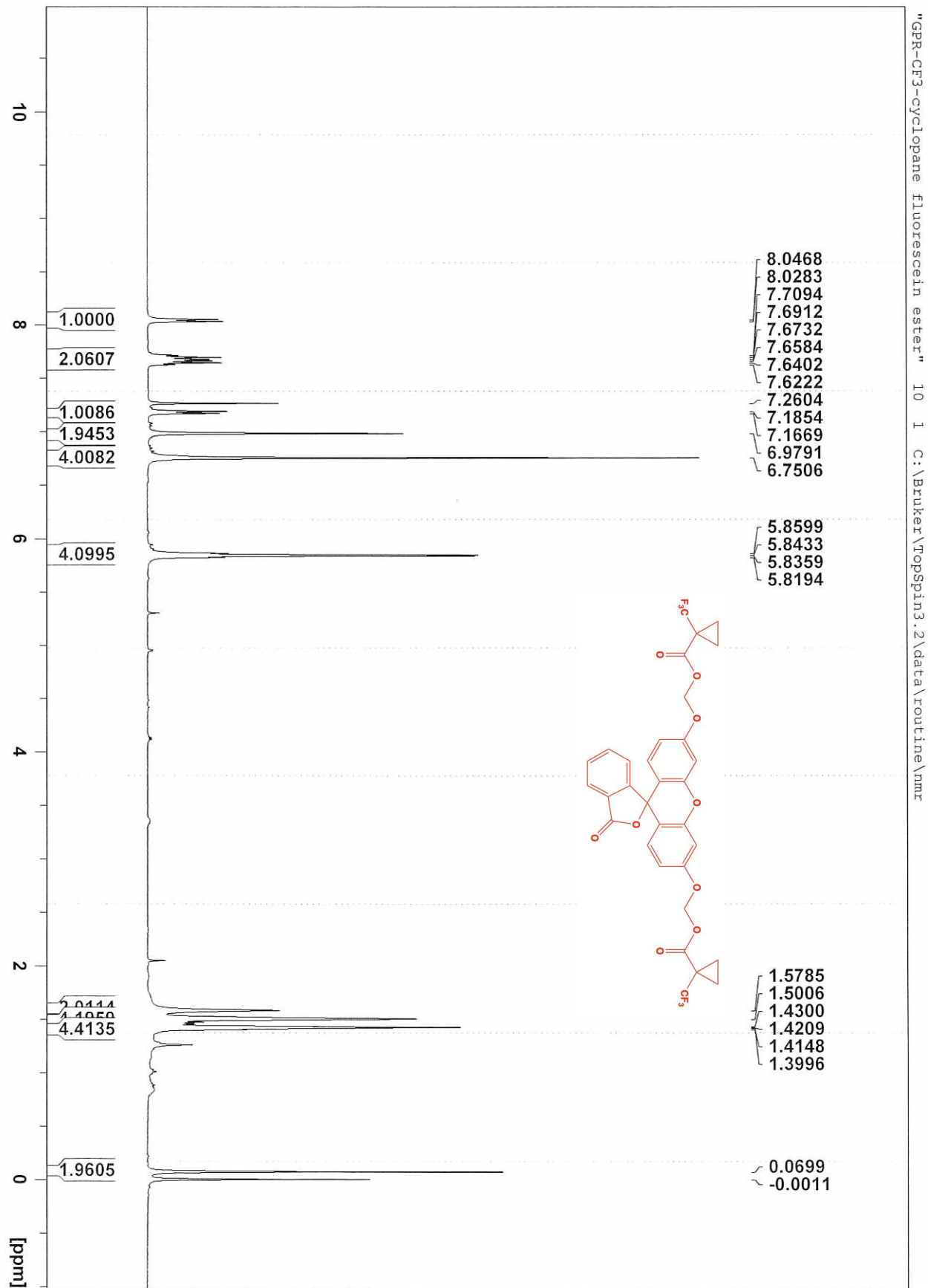


Figure S128

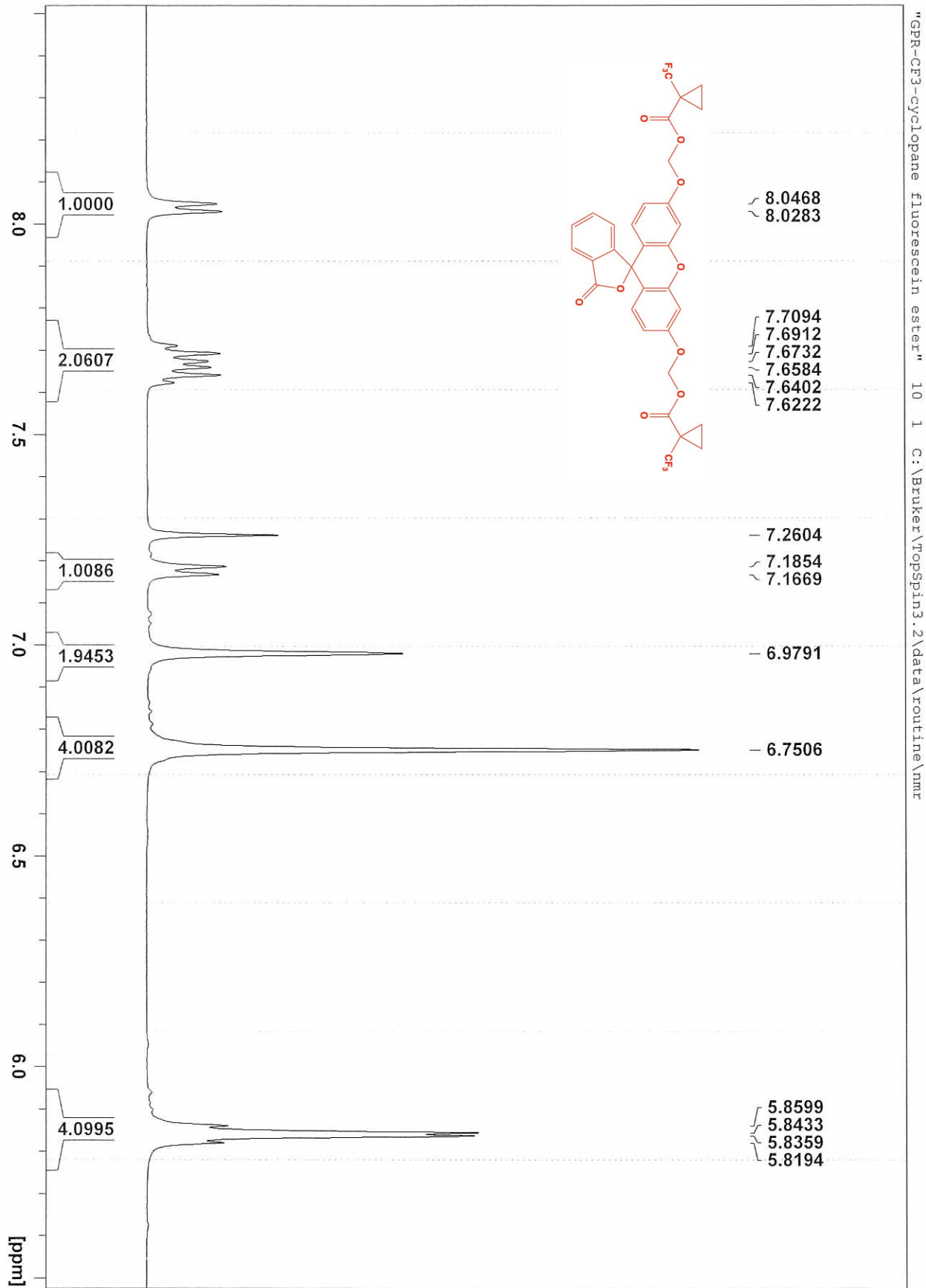


Figure S129

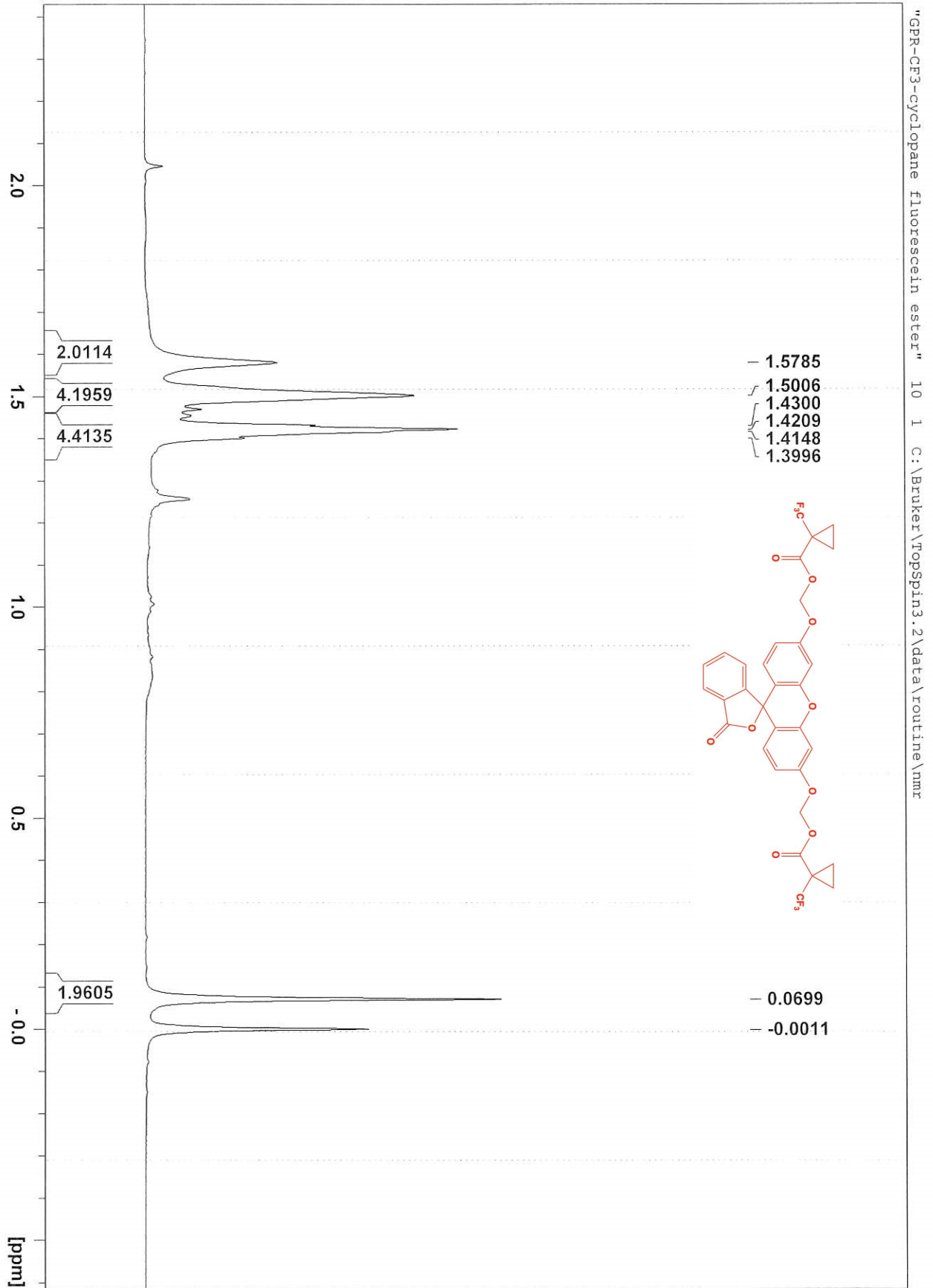


Figure S130

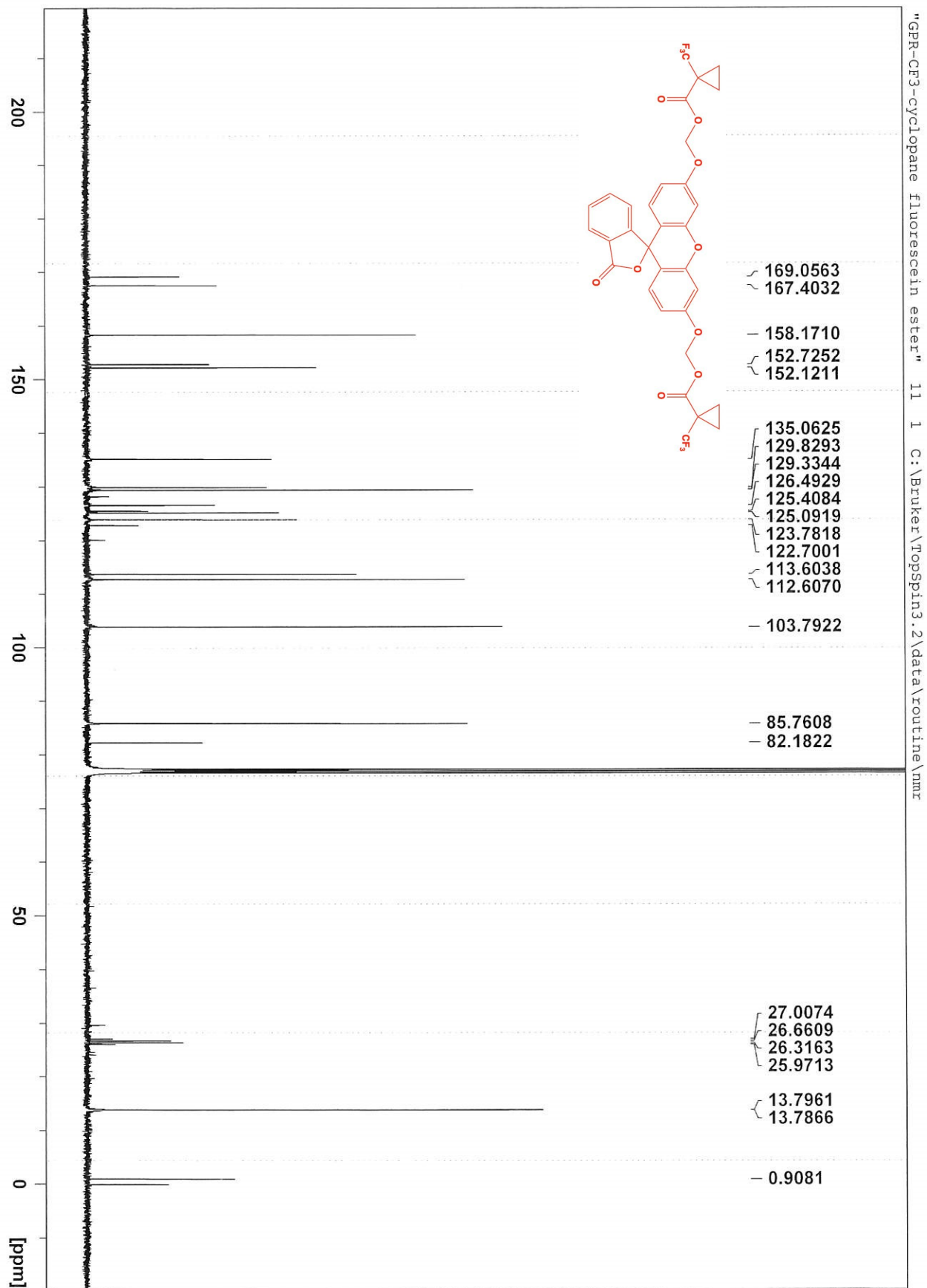


Figure S131

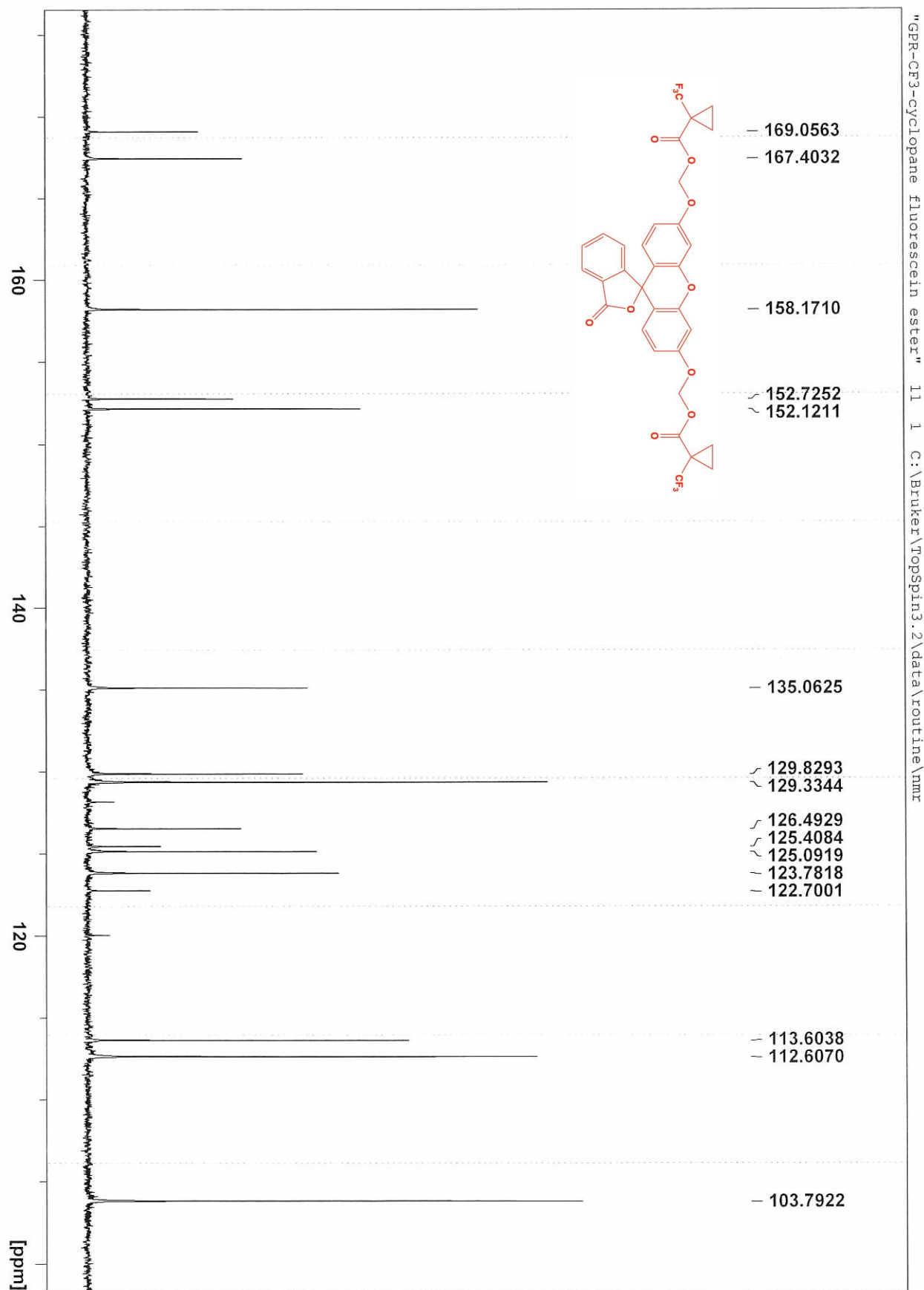


Figure S132

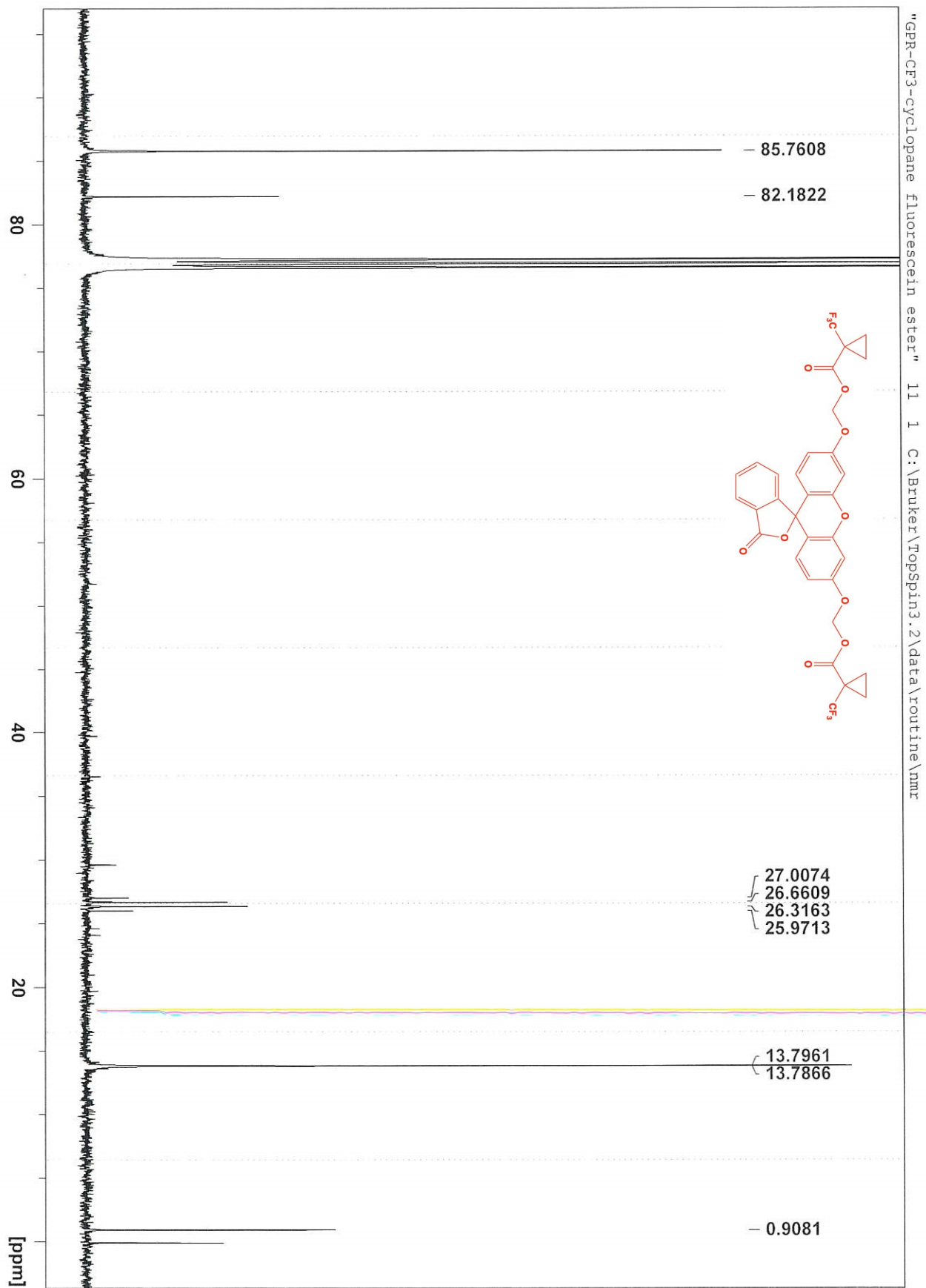
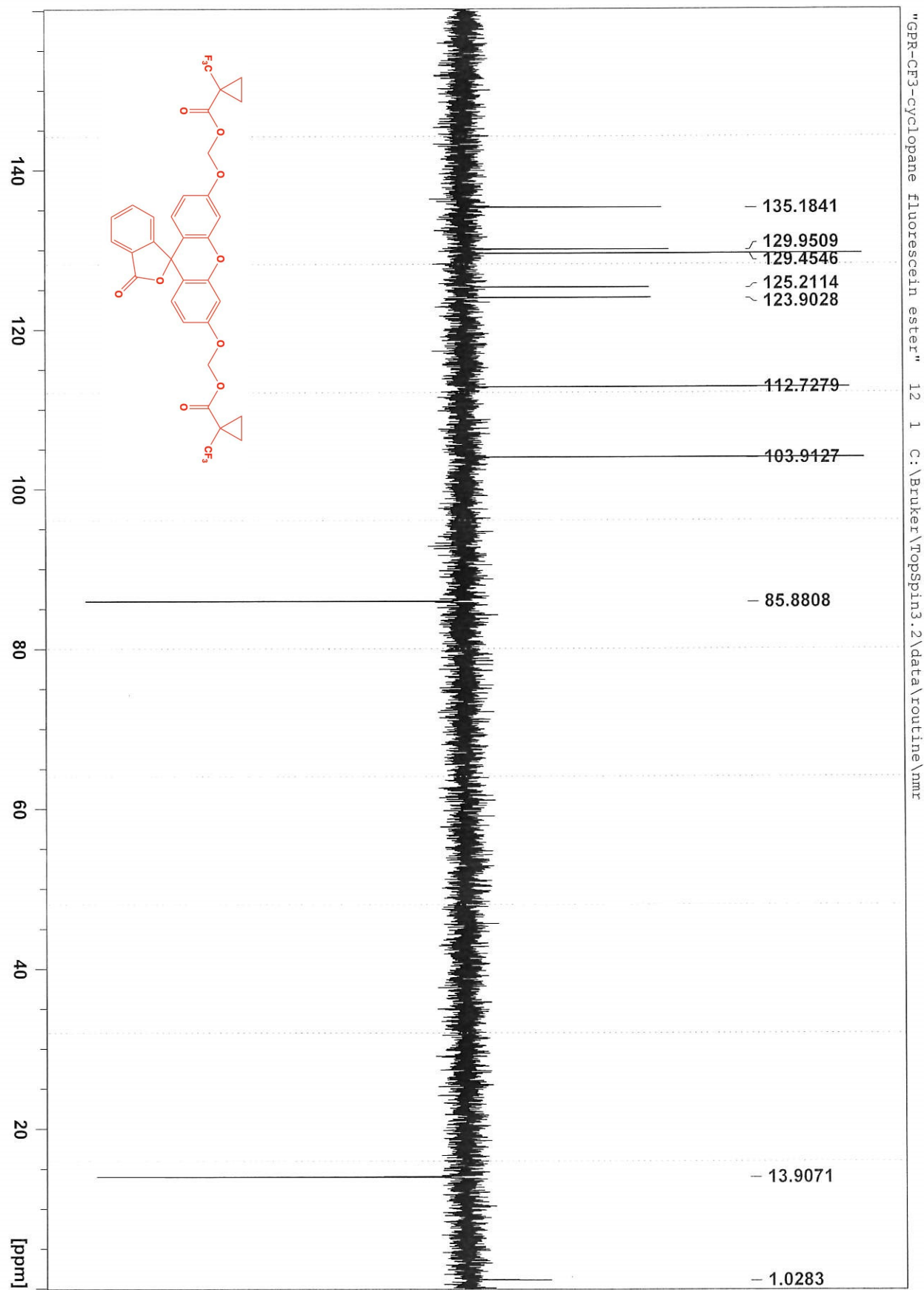


Figure S133



GPR-CF3-cyclopane fluorescein ester

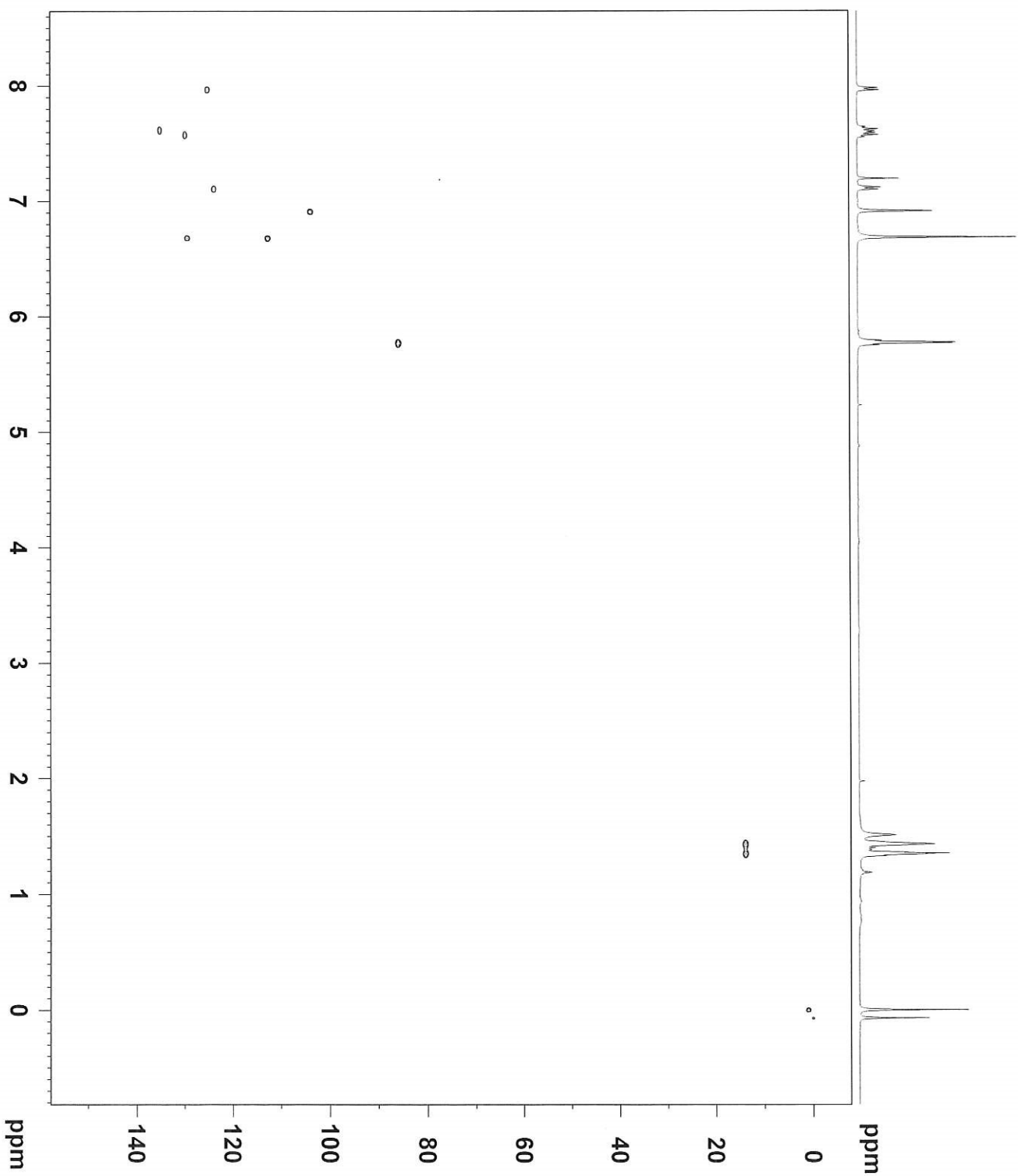
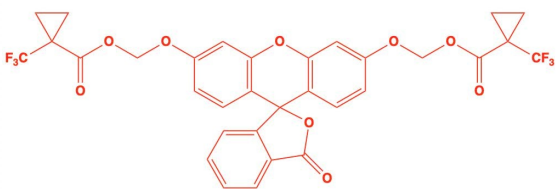


Figure S134

Huber

Figure S135

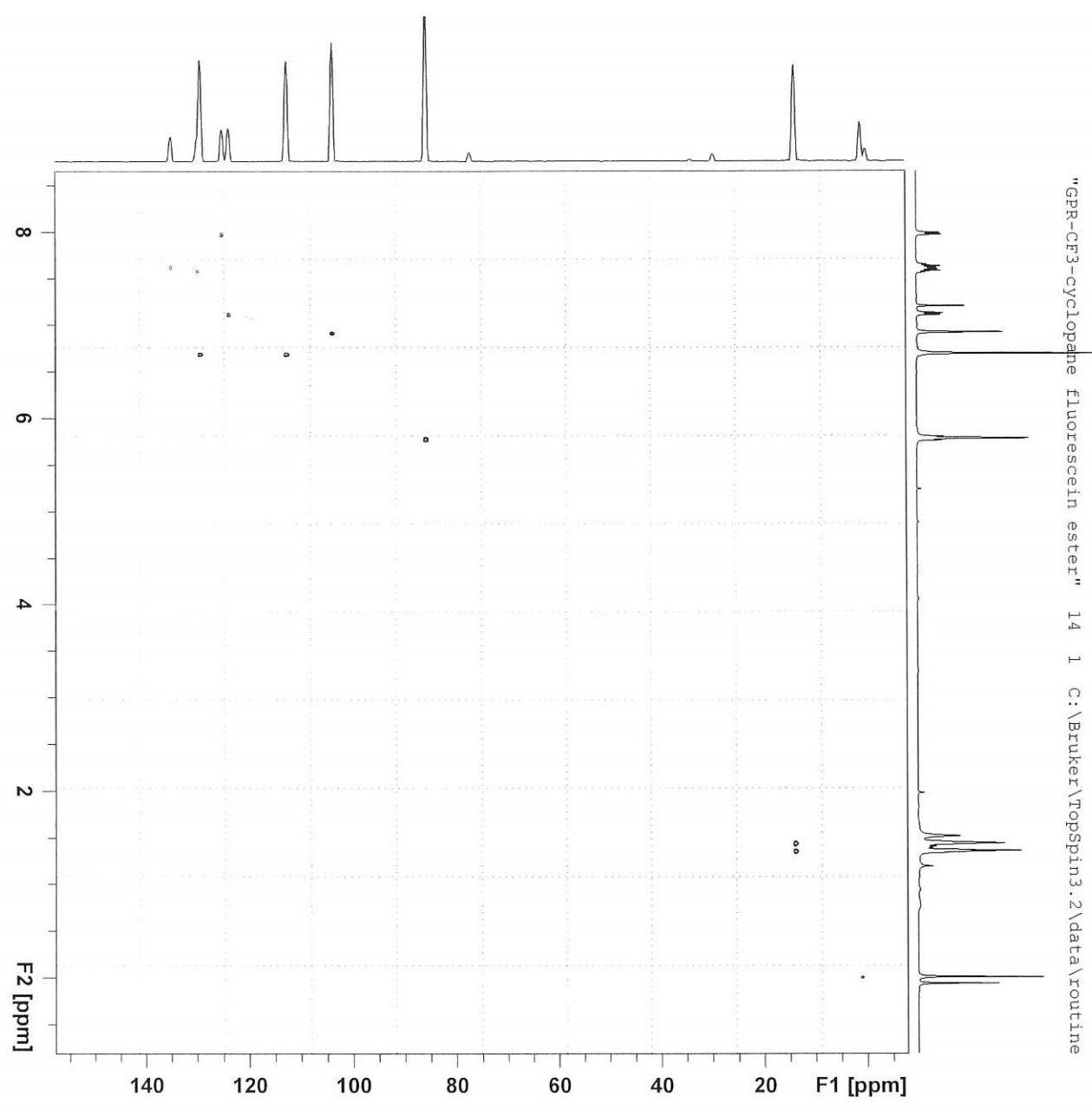
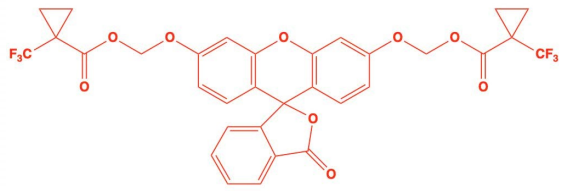


Figure S136

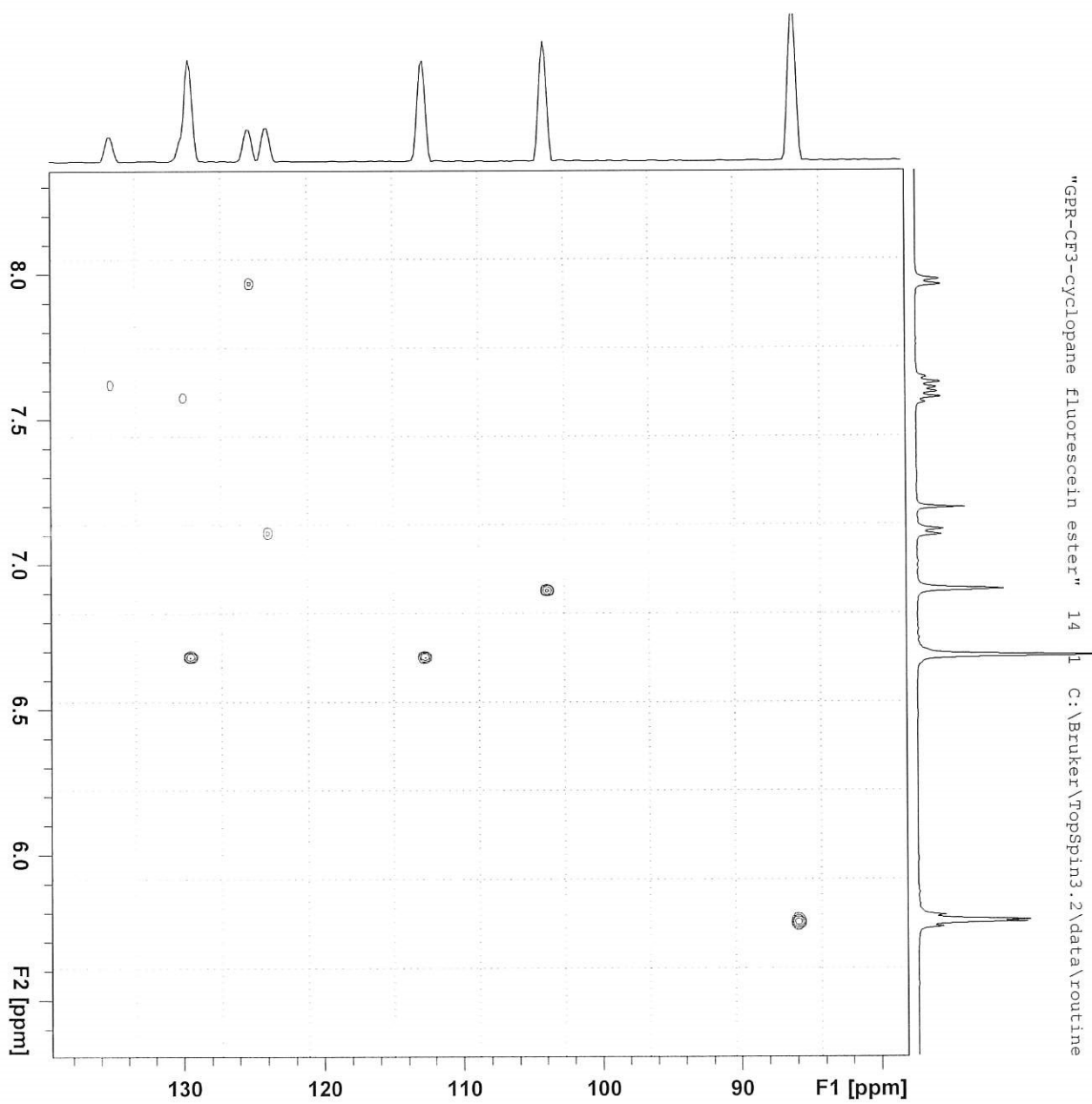
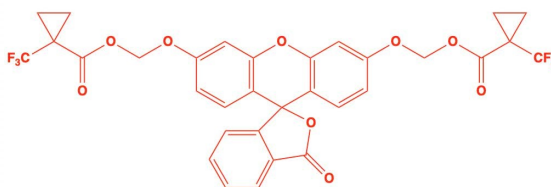
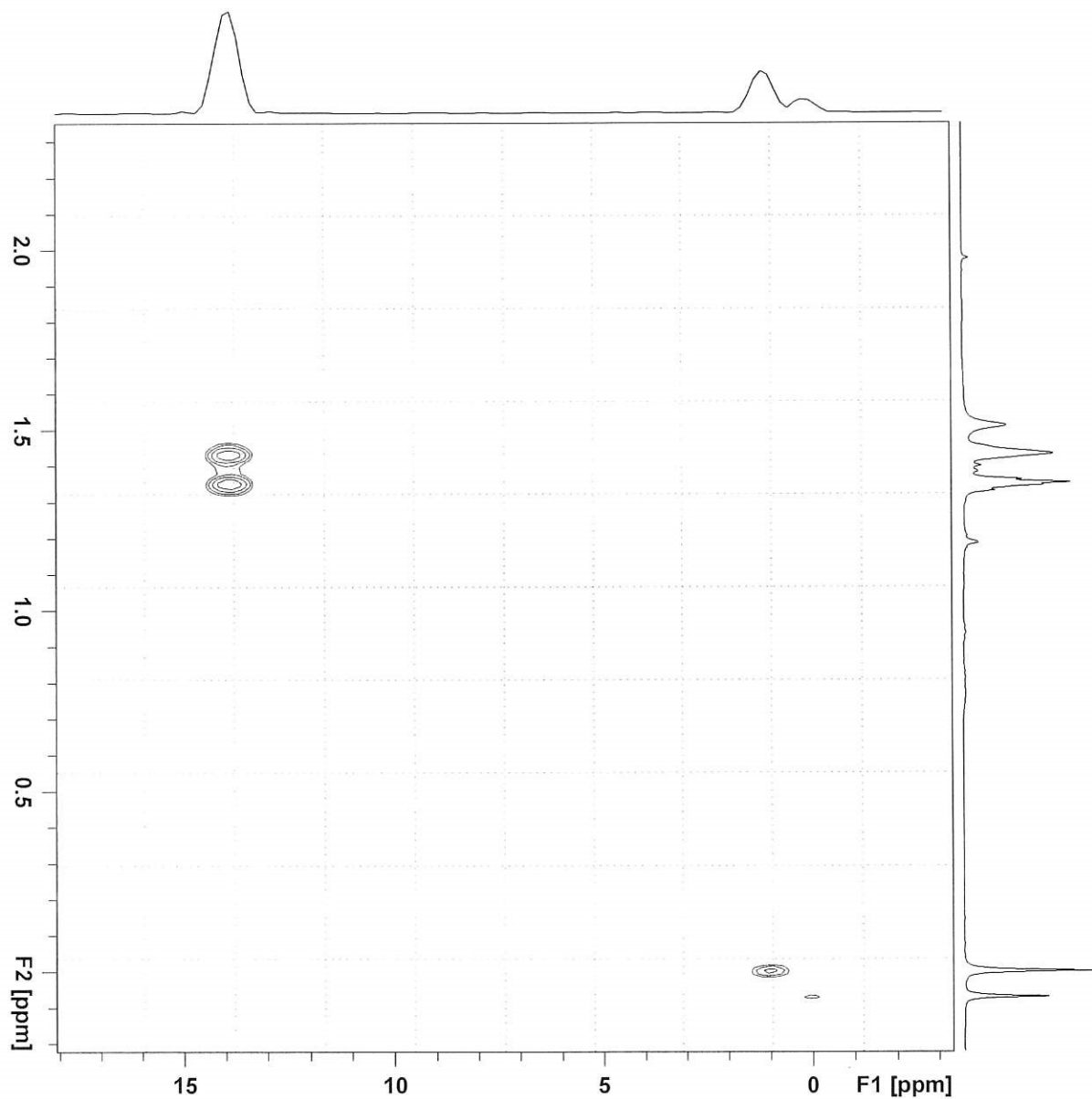
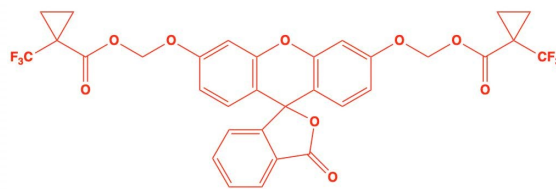
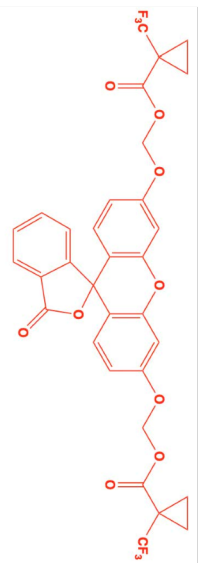


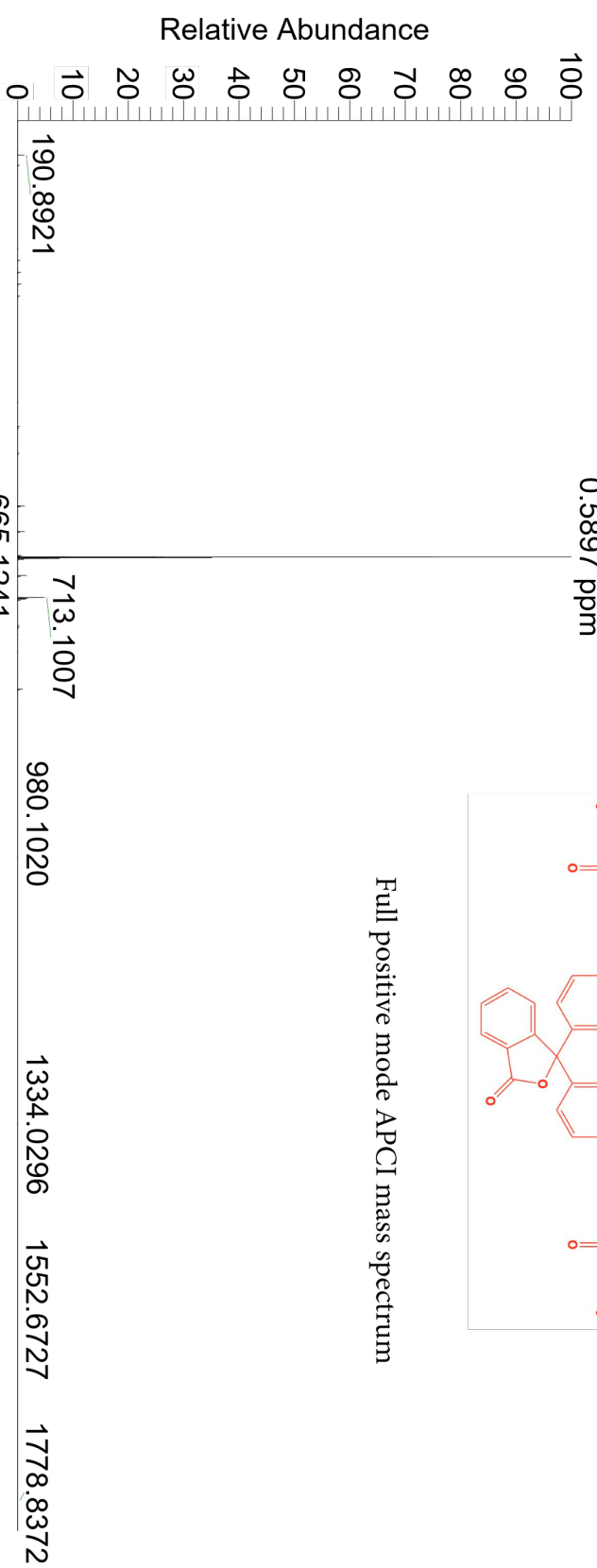
Figure S137





NL:
2.52E8
JE-11#25-32 RT: 0.22-0.28
AV: 8 T: FTMS + p APCI
corona Full ms
[150.00-1800.00]

Full positive mode APCI mass spectrum

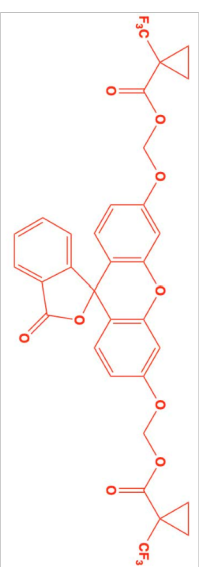


665.1245
C₃₂ H₂₃ O₉ F₆
0.5897 ppm

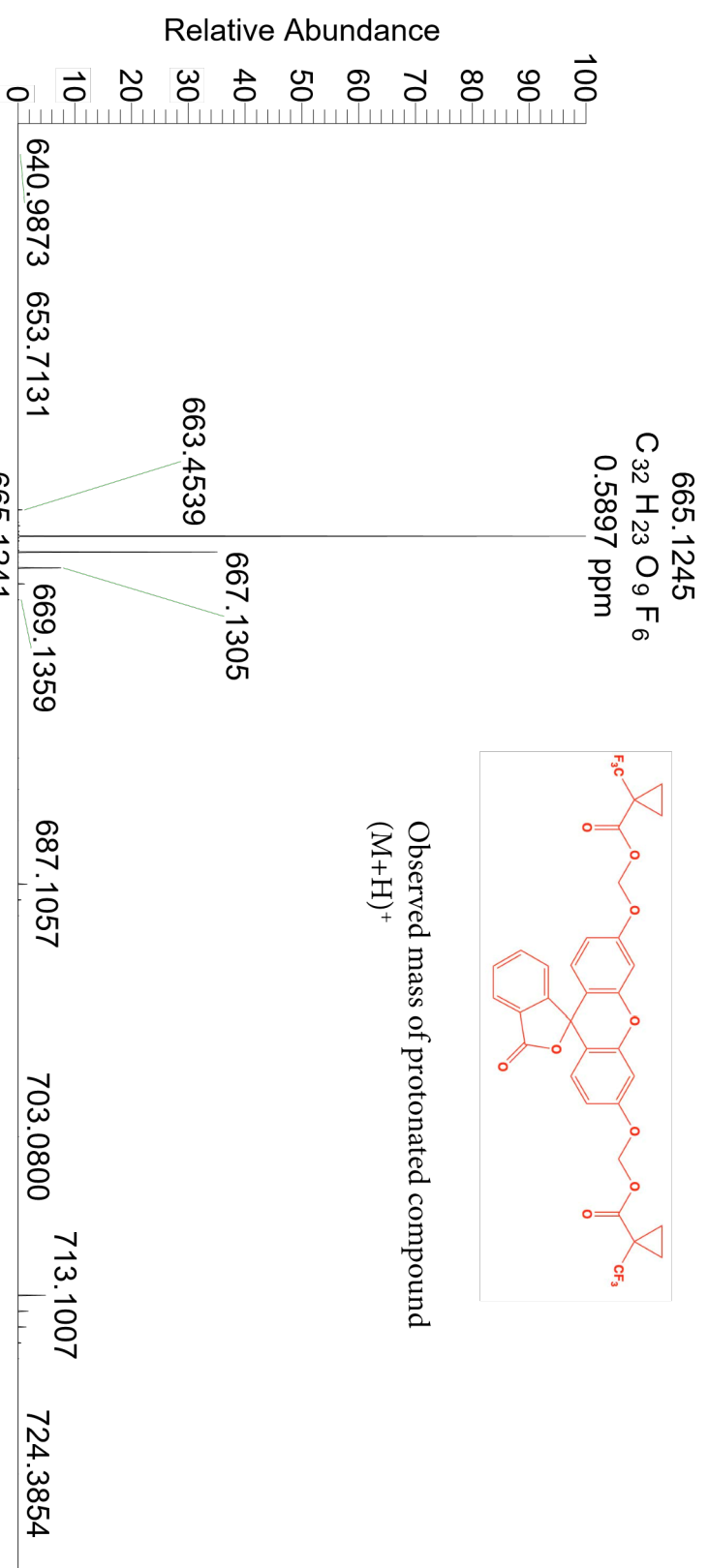
665.1241
C₃₂ H₂₃ O₉ F₆
0.0002 ppm

Theoretical mass of protonated compound
(M+H)⁺

NL:
6.92E5
C₃₂ H₂₂ F₆ O₉ H:
C₃₂ H₂₃ F₆ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM



NL:
2.52E8
JE-11#25-32 RT: 0.22-0.28
AV: 8 T: FTMS + p APCI
corona Full ms
[150.00-1800.00]



NL:
6.92E5

C₃₂ H₂₂ F₆ O₉ H:
C₃₂ H₂₃ F₆ O₉
c (gss, s /p:40)(Val) Chrg 1
R: 20000 Res .Pwr . @FWHM

Theoretical mass of protonated compound
(M+H)⁺

