

**SPARC in cancer-associated fibroblasts is an independent poor prognostic factor in non-metastatic triple-negative breast cancer and exhibits pro-tumor activity**

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## Supplementary Material and Methods

### RT-qPCR

RNA was isolated using the RNeasy® Plus Mini Kit (Qiagen, Hilden, Germany) and 1 µg of total RNA was reverse transcribed using the SuperScript™ III reverse transcriptase kit (Invitrogen). Real-time PCR was performed using SYBR® Premix Ex Taq™ (Tli RNaseH Plus) (Takara Clontech) on a Light Cycler 480 SYBR Green I master and a Light Cycler 480 apparatus (both from Roche Diagnostics, Indianapolis, IN) with the following primers: human CD206 forward: 5'-GGGTTGCTATCACTCTCTATG-3'; human CD206 reverse: 5'-TTTCTTGTCTGTTGCCGTAGTT-3'; human GAPDH forward: 5'-GAAGGTCGGAGTCAACGGATT-3'; human GAPDH reverse: 5'-TGACGGTGCCATGGAATTTG-3'. *CD206* expression was normalized to *GAPDH* expression.

Clinical and tumor characteristics	Univariate analysis HR 95% CI N = 148	Multivariate analysis HR 95% CI N = 148
<b>Age</b>	N=148	
< 55 years	1	
≥ 55 years	2.21 [1.03-4.75]	
	<b>P = 0.027</b>	
<b>Tumor size</b>	N=148	
T1	1	1
T2	3.04 [1.17-7.93]	1.99 [0.74-5.30]
T3/T4	6.73 [2.34-19.38]	3.66 [1.22-11.0]
	<b>P &lt; 0.001</b>	<b>P = 0.050</b>
<b>Nodal status</b>	N=148	
N-	1	1
N+	2.47 [1.36-4.47]	2.38 [1.25-4.55]
	<b>P = 0.002</b>	<b>P = 0.008</b>
<b>Histological grade (SBR)</b>	N=146	
1-2	1	
3	1.07 [0.45-2.53]	
	<b>P = 0.883</b>	
<b>Histology</b>	N=145	
Ductal	1	
Lobular	0.70 [0.22-2.29]	
Other	1.17 [0.42-3.29]	
	<b>P = 0.777</b>	
<b>Adjuvant chemotherapy</b>	N=148	
No	1	1
Yes	0.36 [0.20-0.64]	0.33 [0.18-0.60]
	<b>P = 0.006</b>	<b>P &lt; 0.001</b>

<b>Basal-like phenotype</b>	N=147
Yes	1
No	1.16 [0.64-2.08]
	<i>P</i> = 0.624
<b>SPARC expression in tumor cells</b>	N=132
Negative	1
Positive	0.75 [0.40-1.41]
	<i>P</i> = 0.363
<b>SPARC expression in CAFs</b>	N=126
Negative	1
Positive	1.92 [0.59-6.23]
	<i>P</i> = 0.235
<b>SPARC expression in TAMs</b>	N=118
Negative	1
Positive	0.59 [0.29-1.16]
	<b><i>P</i> = 0.139</b>
<b>SPARC expression in endothelial cells</b>	N=109
Negative	1
Positive	0.73 [0.35-1.51]
	<i>P</i> = 0.401
<b>SPARC expression in TILs</b>	N=82
Negative	1
Positive	1.10 [0.32-3.75]
	<i>P</i> = 0.882
<b>TIL density</b>	N=142
[0-1]	1
>1	0.94 [0.50-1.75]
	<i>P</i> = 0.839
<b>PD-L1 expression in tumor cells</b>	N=136
< 1%	1
≥ 1%	0.91 [0.49-1.70]
	<i>P</i> = 0.770

<b>PD-L1 expression in TILs</b>	N=134
0	1
]0-50[	1.90 [0.66-5.45]
≥ 50	1.10 [0.33-3.66]
	<i>P</i> = 0.238
<b>PD1 expression in TILs</b>	N=140
0	1
]0-50[	0.94 [0.40-2.25]
≥ 50	0.73 [0.22-2.38]
	<i>P</i> = 0.832
<b>Fibrosis</b>	N=137
≤ 50%	1
> 50%	1.10 [0.60-2.02]
	<i>P</i> = 0.746
<b>TAMs (inflammation)</b>	N=143
0/1	1
2	1.11[0.51-2.43]
3	0.67 [0.31-1.45]
	<i>P</i> = 0.285

**Table S1. Univariate and multivariate Cox proportional hazard models to identify prognostic factors of overall survival (OS) in TNBC**

SBR: Scarff-Bloom-Richardson; CAFs: cancer-associated fibroblasts; TAMs: tumor-associated macrophages; TILs: tumor-infiltrating lymphocytes  
HR = hazard ratio; CI = confidence interval; p values in bold, statistically significant.

Clinical and tumor characteristics	SPARC expression in CAFs Negative (N=15)	SPARC expression in CAFs Positive (N=111)	<i>P</i> value
<b>Age</b>			0.018
< 55 years	1 (6.7%)	43 (38.7%)	
≥ 55 years	14 (93.3%)	68 (61.3%)	
<b>Tumor size</b>			0.334
T1	6 (40.0%)	36 (32.4%)	
T2	9 (60.0%)	59 (53.2%)	
T3/T4	0	16 (14.4%)	
<b>Nodal status</b>			0.863
N-	9 (60.0%)	64 (57.7%)	
N+	6 (40.0%)	47 (42.3%)	
<b>Histological grade (SBR)</b>			0.130
1-2	3 (20.0%)	8 (7.3%)	
3	12 (80.0%)	101 (92.7%)	
<b>Histology</b>			0.080
Ductal	11 (73.3%)	95 (88.0%)	
Lobular	3 (20.0%)	5 (4.6%)	
Other	1 (6.7%)	8 (7.4%)	
<b>Adjuvant chemotherapy</b>			0.186
No	7 (46.7%)	33 (29.7%)	

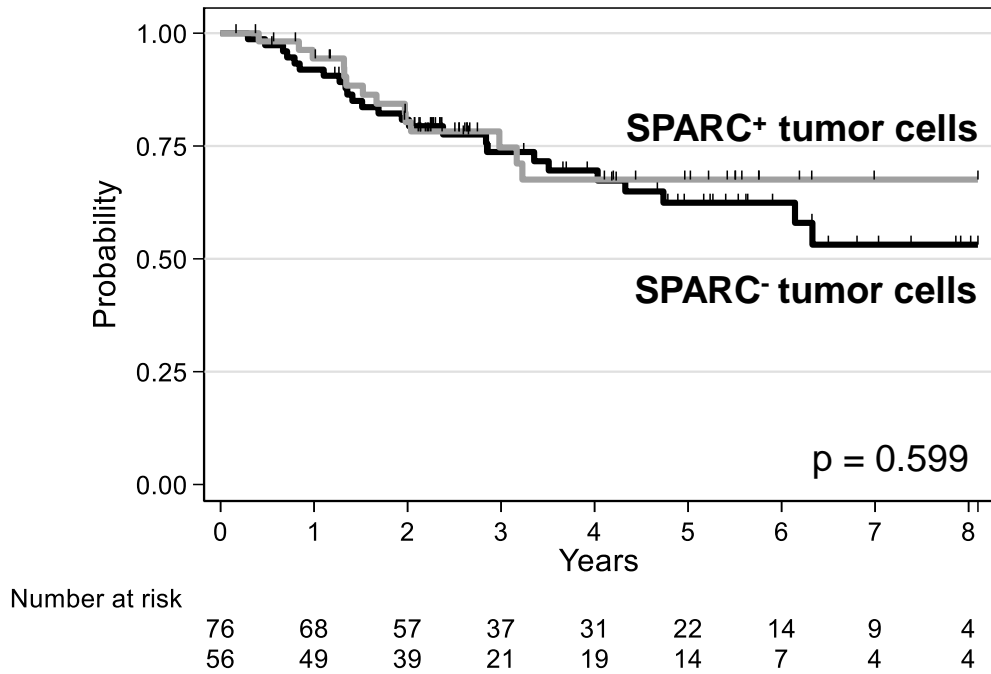
Yes	8 (53.3%)	78 (70.3%)	
<b>Basal-like phenotype</b>			<b>0.678</b>
≤ 10%	9 (60.0%)	72 (65.4%)	
Basal	6 (40.0%)	38 (34.6%)	
<b>SPARC expression in tumor cells</b>			<b>0.603</b>
Negative	8 (53.3%)	67 (60.4%)	
Positive	7 (46.7%)	44 (39.6%)	
<b>SPARC expression in TAMs</b>			<b>0.007</b>
Negative	7 (58.3%)	20 (19.4%)	
Positive	5 (41.7%)	83 (80.6%)	
<b>SPARC expression in endothelial cells</b>			<b>0.026</b>
Negative	7 (50.0%)	20 (22.0%)	
Positive	7 (50.0%)	71 (78.0%)	
<b>SPARC expression in TILs</b>			<b>1.000</b>
Negative	8 (100.0%)	65 (89.0%)	
Positive	0	8 (11.0%)	
<b>TIL density</b>			<b>0.127</b>
[0-1]	6 (42.9%)	26 (23.9%)	
> 1	8 (57.1%)	83 (76.1%)	
<b>PD-L1 expression in tumor cells</b>			<b>0.109</b>
< 1%	7 (53.9%)	31 (28.4%)	
≥ 1%	6 (46.1%)	78 (71.6%)	



<b>PD-L1 expression in TILs</b>			<b>0.049</b>
0	4 (30.8%)	10 (9.2%)	
]0-50[	7 (53.8%)	61 (56.0%)	
≥ 50	2 (15.4%)	38 (34.8%)	
<b>PD1 expression in TILs</b>			<b>0.415</b>
0	2 (15.4%)	11 (10.2%)	
]0-50[	8 (61.5%)	83 (76.9%)	
≥ 50	3 (23.1%)	14 (12.9%)	
<b>Fibrosis</b>			<b>0.028</b>
≤ 50%	3 (20.0%)	56 (51.4%)	
> 50%	12 (80.0%)	53 (48.6%)	
<b>TAMs (inflammation)</b>			<b>0.349</b>
0/1	4 (28.6%)	15 (13.8%)	
2	3 (21.4%)	32 (29.4%)	
3	7 (50.0%)	62 (56.9%)	

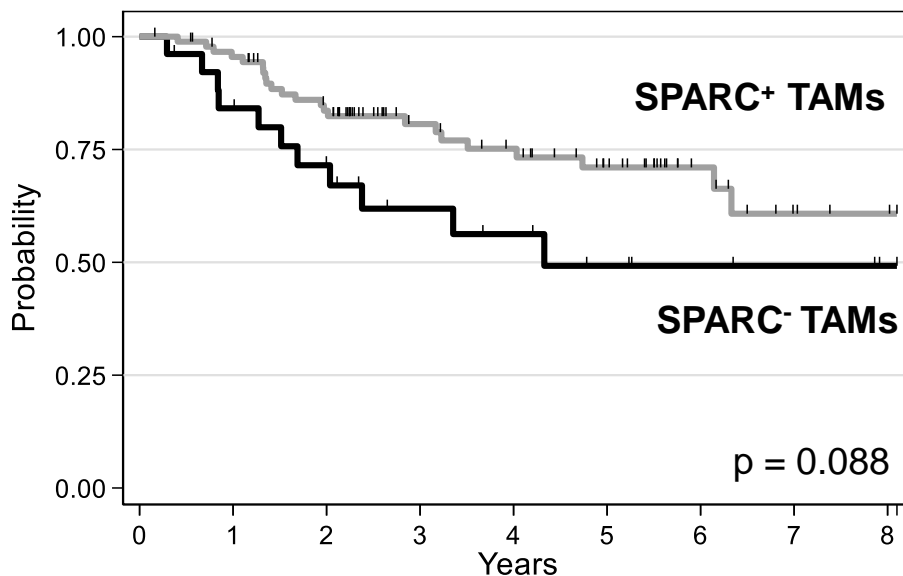
**Table S2. Clinicopathological characteristics in function of SPARC expression (SPARC<sup>+</sup> and SPARC<sup>-</sup>) in CAFs**

SBR: Scarff-Bloom-Richardson; CAFs: cancer-associated fibroblasts; TAMs: tumor-associated macrophages; TILs: tumor-infiltrating lymphocytes.



**Figure S1. Relapse-free survival in function of the SPARC expression status in TNBC cancer cells.**

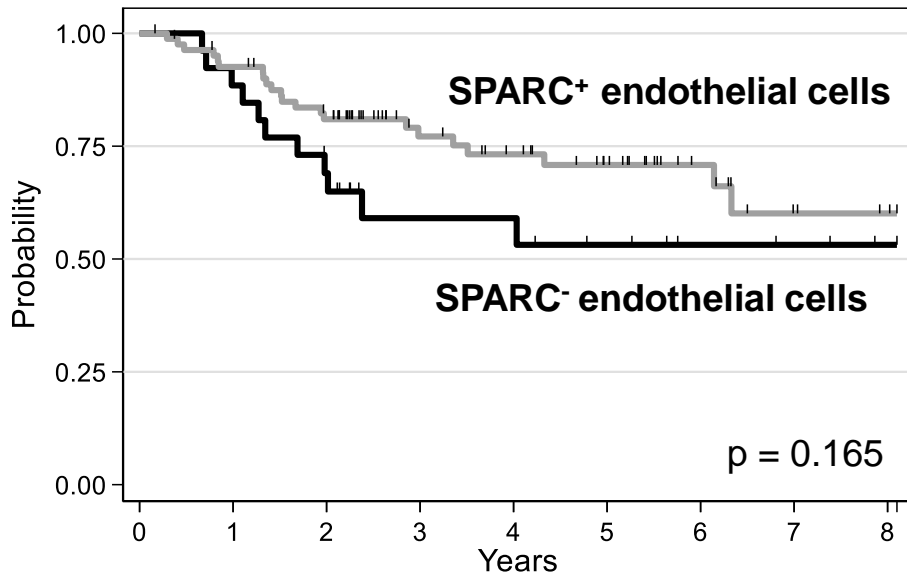
Patients with TNBC were divided in two subgroups according to SPARC expression in tumor cells: SPARC<sup>+</sup> and SPARC<sup>-</sup>.



Number at risk		0	1	2	3	4	5	6	7	8
SPARC+		27	20	16	11	9	6	4	3	1
SPARC-		91	84	69	45	39	28	15	8	5

**Figure S2. Relapse-free survival in function of SPARC status in TAMs.**

Patients with TNBC were divided in two subgroups according to SPARC expression in TAMs: SPARC+ and SPARC-.

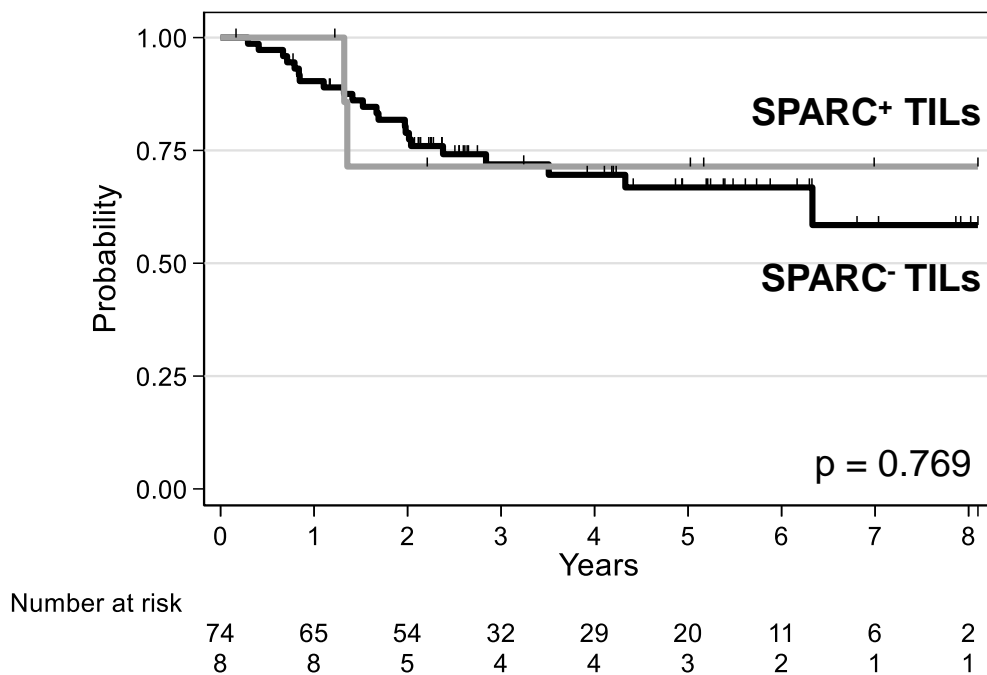


Number at risk

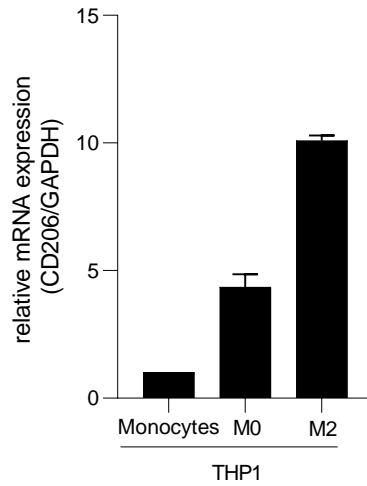
27	23	17	10	10	7	4	3	1
82	74	62	40	34	25	15	8	5

**Figure S3. Relapse-free survival according in function of SPARC expression status in endothelial cells.**

Patients with TNBC were divided in two subgroups according to SPARC expression in endothelial cells within the tumor microenvironment: SPARC<sup>+</sup> and SPARC<sup>-</sup>.



**Figure S4. Relapse-free survival according to SPARC expression status in TILs.** Patients with TNBC were divided in two subgroups according to SPARC expression in TILs: SPARC<sup>+</sup> and SPARC<sup>-</sup>.



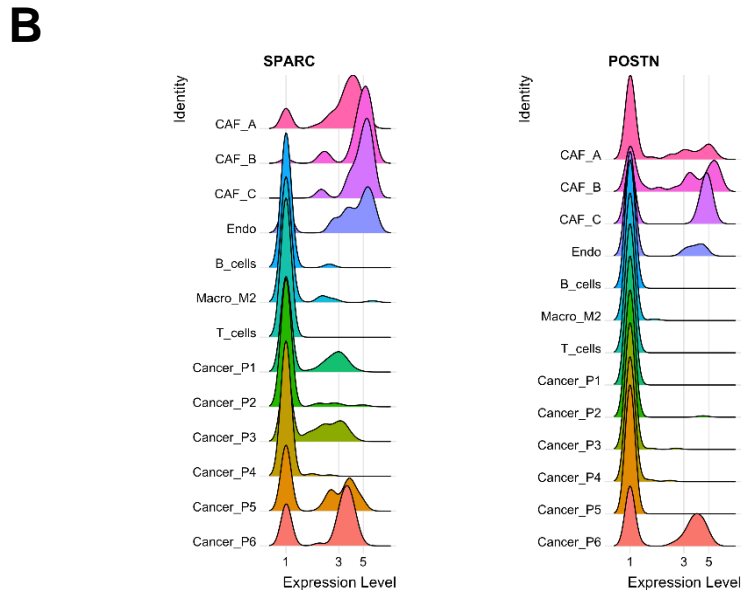
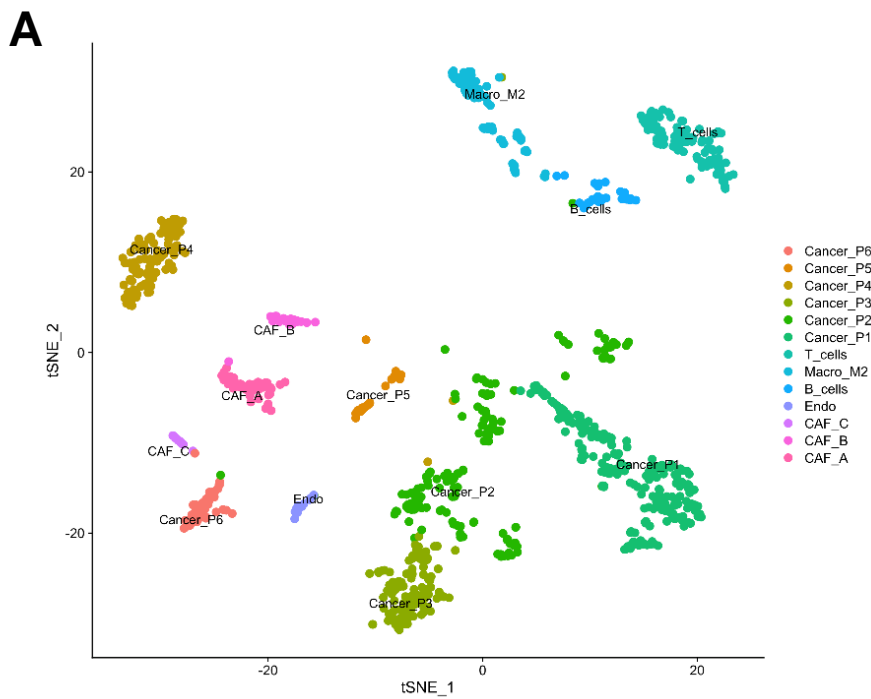
**Figure S5. THP1 monocyte differentiation into M2 macrophages.**

mRNA expression of the M2 macrophage marker CD206 was quantified by RT-qPCR in THP1 monocytes, M0-, and M2-polarized THP1 macrophages. Data were normalized to GAPDH expression level. Results are expressed as mean  $\pm$  SD (n = 3).

<b>SPARC</b>	<b>myCAFs</b>	<b>iCAFs</b>	<b>imPVL</b>	<b>dPVL</b>	<b>Myoepithelial</b>	<b>Endothelial</b>
<b>myCAFs</b>	-	2.39e-63		1.07e-25	4.26e-59	3.19e-33
<b>iCAFs</b>	2.39e-63	-	2.22e-15	1.01e-51		5.48e-120
<b>imPVL</b>		2.22e-15	-		8.09e-22	
<b>dPVL</b>	1.07e-25	1.01e-51		-	9.12e-6	
<b>Myoepithelial</b>	4.26e-59		8.09e-22	9.12e-6	-	3.78e-24
<b>Endothelial</b>	3.19e-33	5.48e-120			3.78e-24	-

**Figure S6. Differential *SPARC* gene expression in different cell populations within TNBC samples by single-cell RNA-seq data analysis.**

The previously published single-cell RNA-seq dataset PRJEB35405 included five patients with TNBC<sup>9</sup>. P values were calculated with the Wilcoxon Rank Sum test and adjusted using the Bonferroni correction based on the total number of genes.

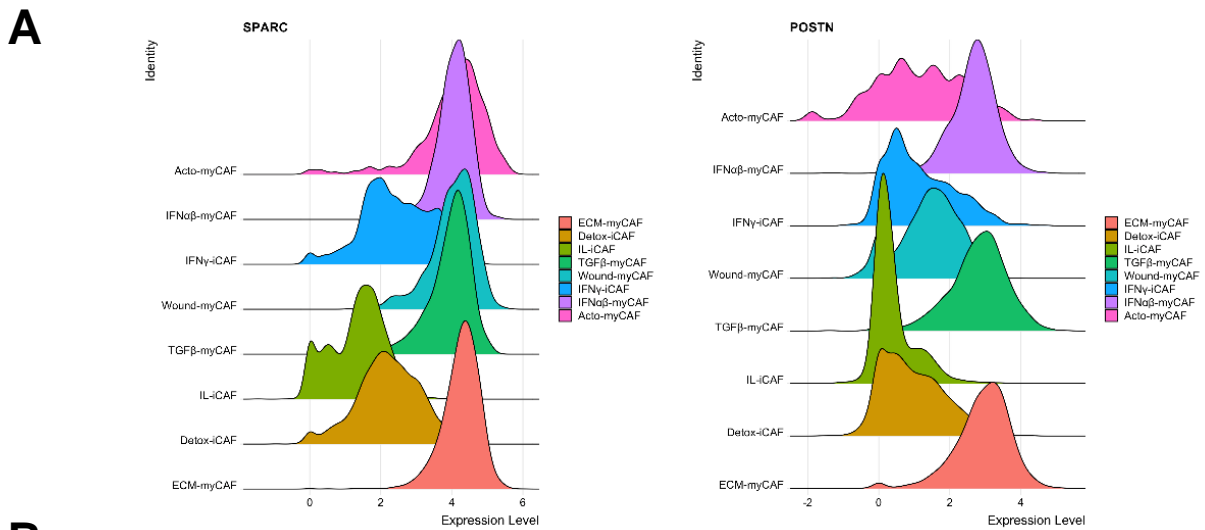


**Figure S7. Expression of *SPARC* and *POSTN* mRNAs in TNBC by single-cell RNA-seq analysis.**

**(A) Cell populations.** Thirteen cell populations were identified by single-cell RNA-seq analysis of the previously published GSE118390 dataset (n=6 TNBC samples), according to<sup>10</sup>.

**(B) *SPARC* and *POSTN* mRNA expression.** *SPARC* and *POSTN* mRNA relative expressions in each of the 13 cell populations identified by single-cell RNA-seq analysis according to<sup>10</sup>. Individual cell populations were annotated as published in the original single-cell RNA-seq study<sup>10</sup> except for TNBC cells that are labelled Cancer\_P1, \_P2, \_P3, \_P4, \_P5, \_P6 (patient 1 to 6, respectively).





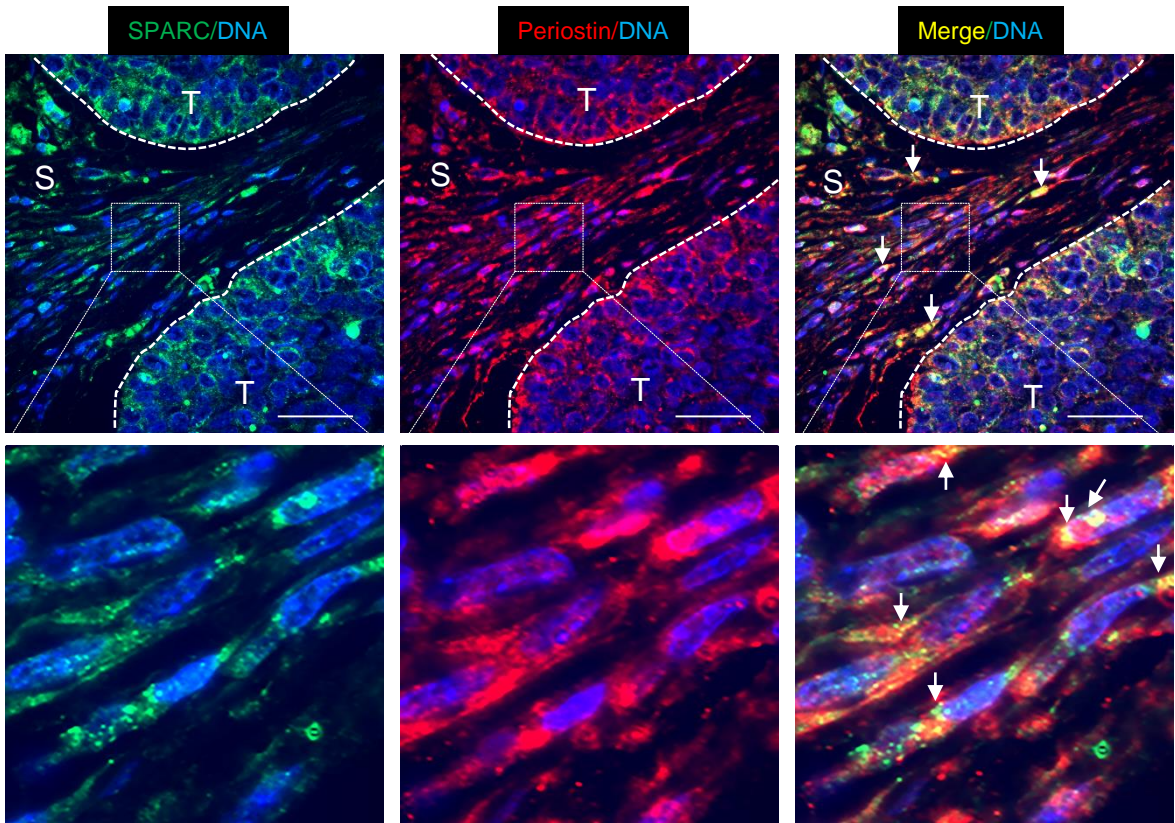
**B**

SPARC	Acto-myCAF	IFN $\alpha\beta$ -myCAF	IFN $\gamma$ -iCAF	Wound-myCAF	TGF $\beta$ -myCAF	IL-iCAF	Detox-iCAF	ECM-myCAF
Acto-myCAF	-		2.84e-76			1.47e-139	6.54e-116	
IFN $\alpha\beta$ -myCAF		-	4.88e-173			0	0	
IFN $\gamma$ -iCAF	2.84e-76	4.88e-173	-	1.26e-192	6.87e-222	5.86e-153		0
Wound-myCAF			1.26e-192	-		0	0	
TGF $\beta$ -myCAF			6.87e-222		-	0	0	
IL-iCAF	1.47e-139	0	5.86e-153	0	0	-	0	0
Detox-iCAF	6.54e-116	0		0	0	0	-	0
ECM-myCAF			0			0	0	-

**Figure S8. Expression of *SPARC* and *POSTN* mRNAs in CAF-S1 clusters in breast cancer by single-cell RNA-seq analysis.**

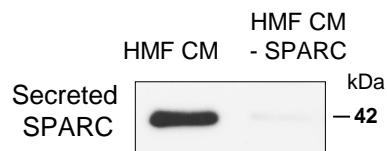
**(A)** Eight CAF-S1 clusters were identified in the previously published single-cell RNA-seq dataset EGAS00001004030 ( $n = 8$  primary breast cancer samples) according to <sup>11</sup>. *SPARC* and *POSTN* mRNA relative expression in the eight CAF-S1 clusters, according to <sup>11</sup>, is shown. Individual CAF-S1 clusters were annotated as published in the original single-cell RNA-seq study <sup>11</sup>. According to <sup>11</sup>, myCAFs were identified in the following five clusters: ECM-myCAF: associated with extracellular matrix (ECM) remodeling, cell-substrate adhesion, and collagen formation; TGF $\beta$ -myCAF: associated with TGF $\beta$  signaling pathway and matrisome; Wound-myCAF: associated with assembly of collagen fibrils and wound healing; IFN $\alpha\beta$ -myCAF: associated with IFN $\alpha/\beta$  signaling; Acto-myCAF: associated with the actomyosin complex. iCAF were identified in the following three clusters: Detox-iCAF: associated with the detoxification and inflammatory responses; IL-iCAF: associated with the response to growth factors, TNF signaling, and interleukin (IL) pathway; IFN $\gamma$ -iCAF: associated with the response to IFN $\gamma$  and cytokine-mediated signaling pathways.

**(B)** Differential *SPARC* gene expression analysis in the indicated cell populations; p values were calculated with the Wilcoxon Rank Sum test and adjusted using the Bonferroni correction based on the total number of genes.



**Figure S9. Co-localization of SPARC with periostin in TNBC PDX.**

PDX B1995 tumor sections were co-incubated with an anti-SPARC polyclonal antibody (15274-1-AP) (green) and an anti-periostin monoclonal antibody (Proteintech) (red). Nuclei were stained with Hoechst 33342 (blue). Top panels: SPARC, periostin, and merge. Bottom panels: higher magnification of the boxed areas. Arrows indicate SPARC and periostin co-localization. Scale bar, 10  $\mu$ m.



**Figure S10. Immunodepletion of SPARC secreted in the conditioned medium from HMFs.** SPARC was immunodepleted or not from HMF CM by immunoprecipitation. SPARC immunodepletion in HMF CM (HMF CM - SPARC) was confirmed by western blot analysis with an anti-SPARC monoclonal antibody (clone AON-5031).