

1 **SUPPLEMENTARY MATERIAL TO**

2

3 **Development of a compounded propofol nanoemulsion using multiple non-**
4 **invasive process analytical technologies**

5

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24 **Supplementary material**

25 **Eq. 2** is derived as follows. One starts with an IVLE with a number N of lipid droplets
26 of size d_0 , giving a total lipid volume:

27
$$V_{lip} = N \cdot \frac{\pi d_0^3}{6}$$

28 **Eq. S1**

29 When these droplets swell to a size d by absorption of a propofol volume V_p , volume
30 conservation gives the following relation:

31
$$V_{lip} + V_p = N \cdot \frac{\pi d_0^3}{6} + V_p = N \cdot \frac{\pi d^3}{6} \rightarrow 1 + \frac{6V_p}{\pi N d_0^3} = \left(\frac{d}{d_0}\right)^3$$

32 **Eq. S2**

33 The propofol volume is linked to the total volume V_{tot} by $V_p = \phi_p \cdot V_{tot}$ with ϕ_p the
34 propofol volume fraction. Further we have:

35
$$V_{tot} = V_{IVLE} + V_p = V_{IVLE} + \phi_p \cdot (V_{IVLE} + V_p) \simeq V_{IVLE}(1 + \phi_p)$$

36 **Eq. S3**

37

38
39 Where a small term $\phi_p V_p$ has been ignored. With these relations, and the definition for
40 the initial lipid volume fraction $\phi_0 = V_{lip}/V_{IVLE}$, **Eq. S2** becomes:

41
$$\frac{d}{d_0} = \sqrt[3]{1 + \frac{\phi_p V_{tot}}{V_{lip}}} \simeq \sqrt[3]{1 + \frac{\phi_p (1 + \phi_p)}{\phi_0}} \simeq 1 + \frac{\phi_p}{3 \cdot \phi_0}$$

42 **Eq. S4**

43 In the last step Taylor expansion has been used, which is valid for small values of ϕ_p .

44 To show that this last step from the cubic root to the approximation holds, both

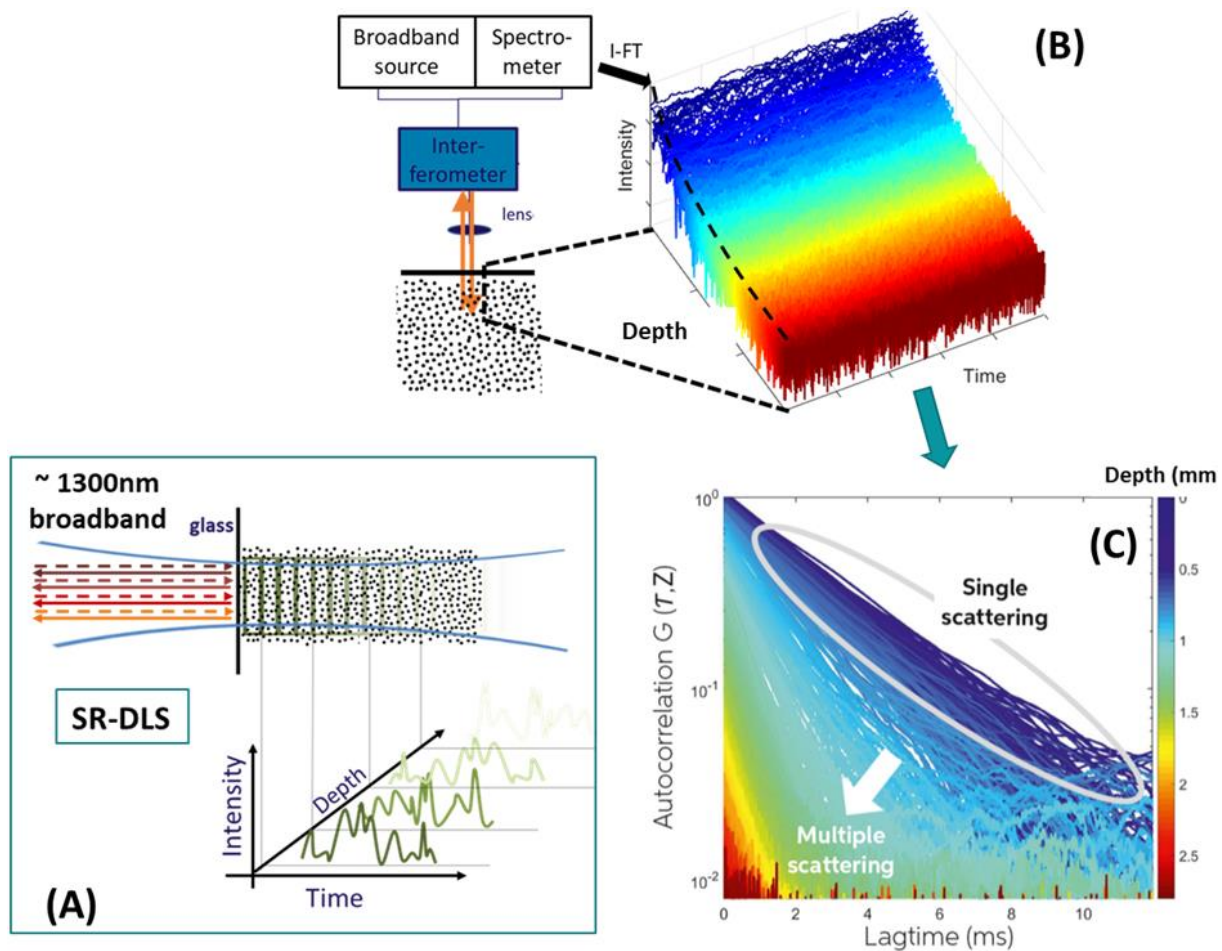
45 equations were simulated. The results are shown in the **Figure S8**. As can be seen
46 from the figure, the approximation holds well in the range used in this study.



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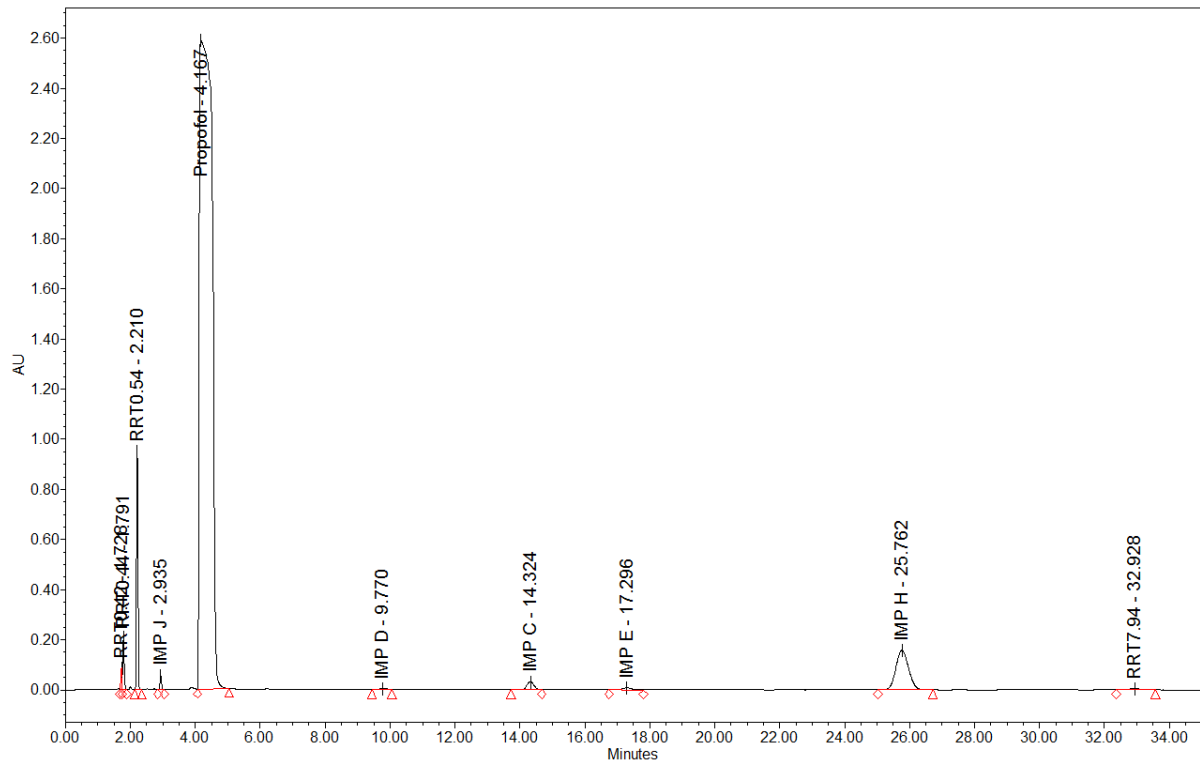
48 **Figure S1** Measurement configuration for simultaneous non-invasive DR-NIRS and

49 SR-DLS measurements.

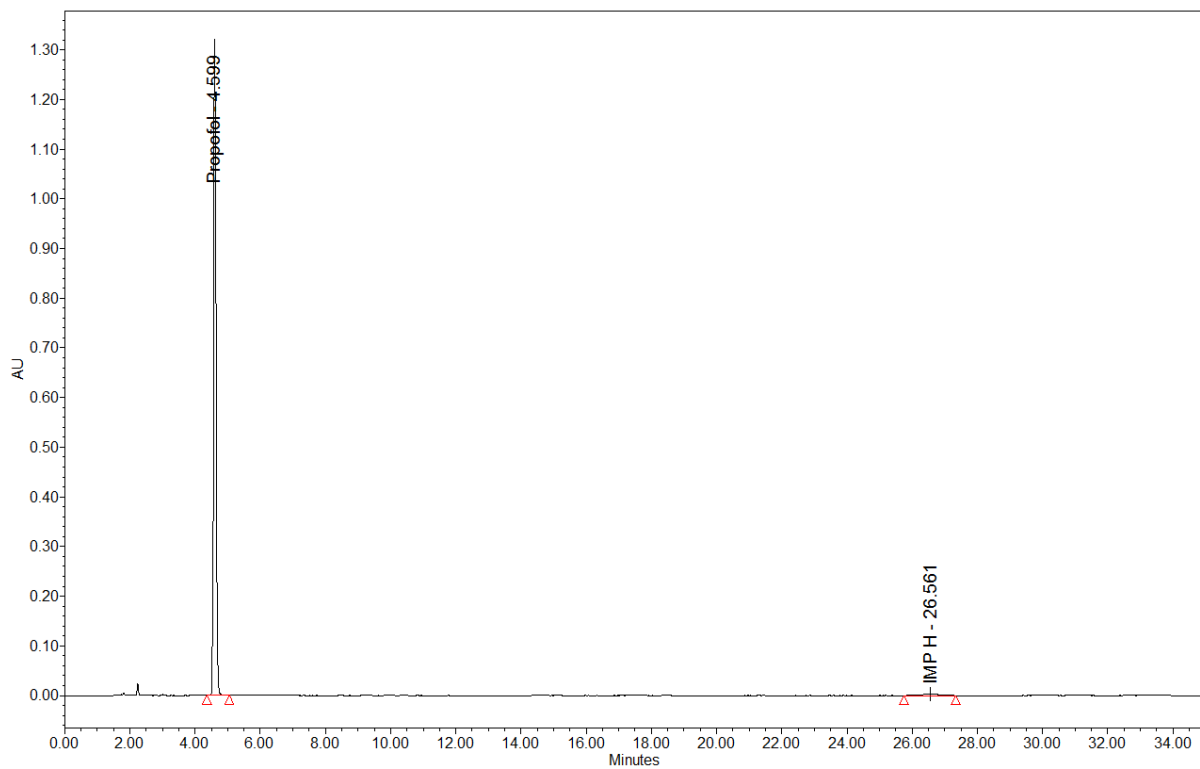


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51 **Figure S2** A) Basic sketch of SR-DLS. B) Spatial Resolution via Fourier
 52 Transformation of broadband interferograms. C) Depth resolved Correlation functions
 53 allow spatial filtering of multiple scattered light.

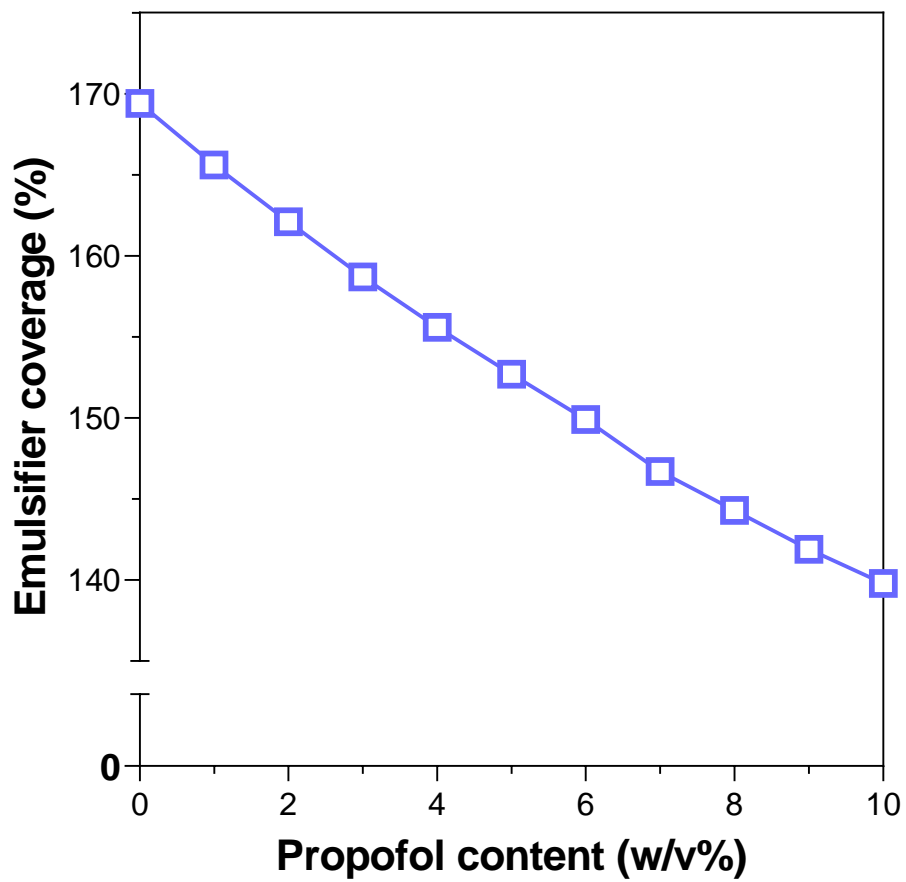


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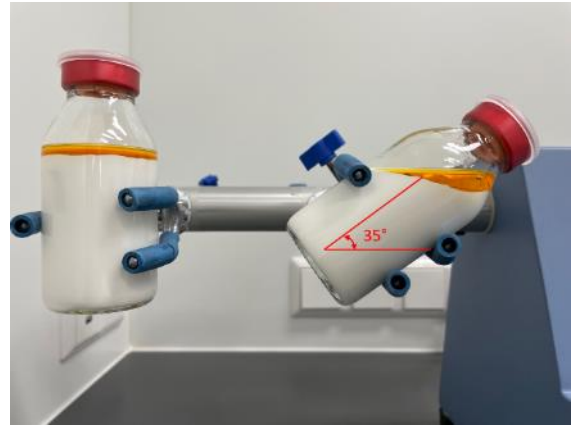
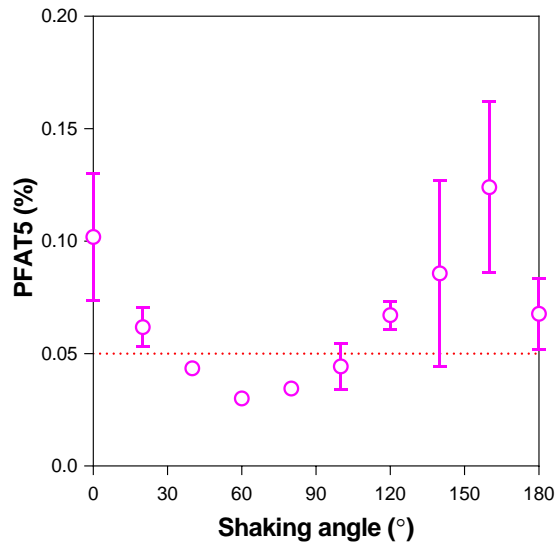
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56 **Figure S3** Typical HPLC chromatograms of the propofol drug substance obtained with
 57 the normal phase method for related substances (**upper panel**) and assay (**lower**
 58 **panel**). When known, specified impurities have been labelled, otherwise the relative
 59 retention time (RRT) is given.

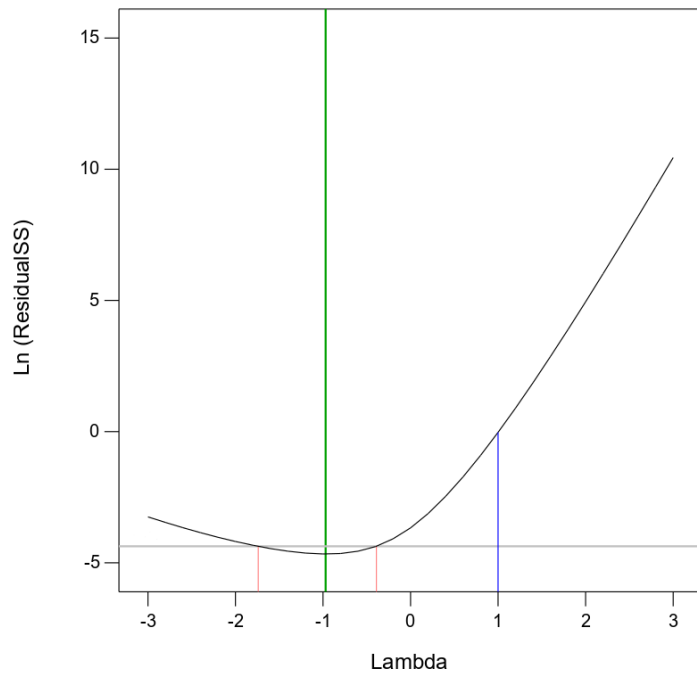


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61 **Figure S4** Theoretical lipid droplet total specific surface area covered by
62 phosphatidylcholine.

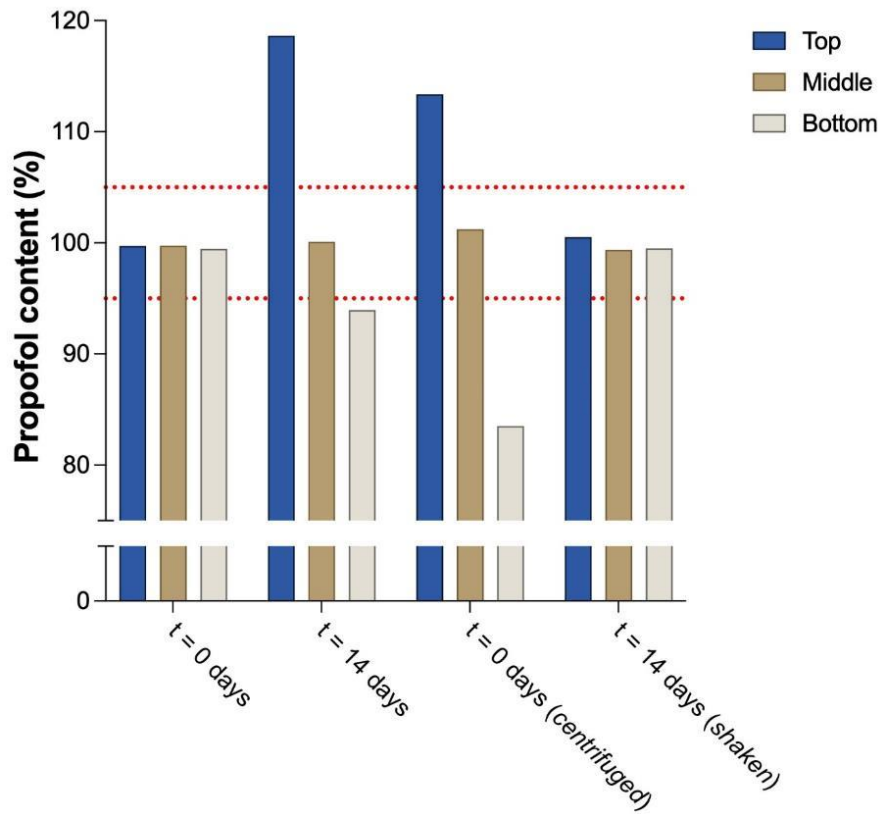


63 **Figure S5** Effect of shaking angle ($^{\circ}$) on the PFAT5 after 5 minutes for a 6% w/v
 64 remotely loaded propofol emulsion in IVLE. Symbols depict the mean and standard
 65 deviation ($n=3$), if no error bars are shown the value is smaller than the symbol value.



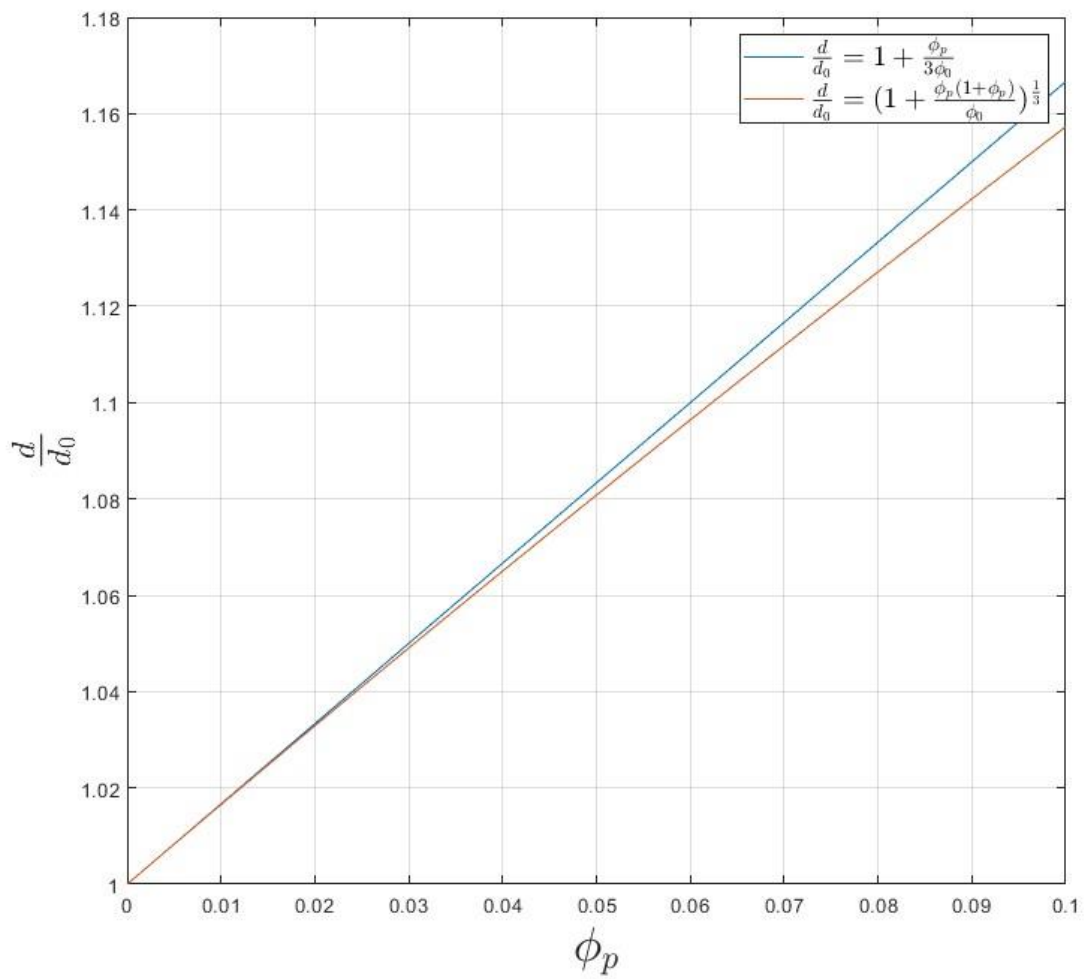
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67 **Figure S6** Box-Cox plot for transformations. The current power (no transformation) is
68 indicated by the blue line ($\lambda = 1$) The minimum model residual is indicated by the green
69 line and the 95%-confidence interval with both red lines.



70

71 **Figure S7** Propofol concentrations determined by HPLC withdrawn from three position
 72 within the glass container (top, middle and bottom). Centrifuged samples were
 73 exposed to 1450 x g for 30 minutes at room temperature. Results are expressed as
 74 the percentage of the theoretical concentration (4% w/v). Dashed lines at 105-95%
 75 indicated upper and lower specification limits as per Ph Eur.



76

77 **Figure S8** Simulation of the initial and final approximations of d/d_0 for ϕ_p ranging from

78 0 to 0.1.

79 **Table S1** Overview of registered parenteral lipid emulsions in the Netherlands (anatomical therapeutic chemical code B05BA02).

Name	RVG	MAH	MA date (dd-mm-yyyy)	Presentation(s)	Oil phase (per 100 ml)	Excipients	Generation	Ref.
ClinOleic 20%	16863	Baxter	13-10-1997	Infusion bags (100, 250, 350, 500 and 1000 ml)	16 gram OO 4 gram SO	Phospholipids (egg), glycerol, sodium oleate, sodium hydroxide and purified water	3 rd	CBG, 2023a
Intralipid 20%	02608	Fresenius Kabi	06-02-1974	Infusion bags (100, 250, and 500 ml)	20 gram SO	Phospholipids (egg), glycerol, sodium hydroxide and purified water	2 nd	CBG, 2023b
Lipoplus 200 mg/ml	31376	B. Braun Melsungen	05-01-2006	Glass bottles (100, 250, 500 and 1000 ml)	10 gram MCT 8 gram SO 2 gram FO	Phospholipids (egg), glycerol, sodium oleate, ascorbyl palmitate, dl- α -tocopherol, sodium hydroxide and purified water	3 rd	CBG, 2023c
Omegaven-Fresenius	23043	Fresenius Kabi	20-04-1999	Glass bottles (50 and 100 ml)	10 gram FO	Phospholipids (egg), glycerol, sodium oleate, dl- α -tocopherol, sodium hydroxide and purified water	3 rd	CBG, 2023d
SMOFlipid 200 mg/ml	30787	Fresenius Kabi	17-01-2005	Glass bottles (100, 250 and 500 ml) and infusion bags (100, 250, 500 and 1000 ml)	6 gram SO 6 gram MCT 5 gram OO 3 gram FO	Phospholipids (egg), glycerol, sodium oleate, dl- α -tocopherol, sodium hydroxide and purified water	3 rd	CBG, 2023e

80 **Abbreviations:** Ref. = reference; RVG = register verpakte geneesmiddelen (national authorisation number); MA(H) = marketing

81 authorization (holder); OO = olive oil, SO = soybean oil; MCT = medium chain triglycerides and FO = fish oil.

82 **References to Table S1**

83 College ter Beoordeling van Geneesmiddelen (CBG), 2023a. Samenvatting van de
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103 **Table S2** Design table in actual units of each variable including the experimental
 104 results for the volume-weighted percentage of large-diameter droplets (PFAT5).

Run no.	Space type*	Concentration (% w/v)	Angle (°)	Time (min.)	PFAT5 (%)
1	Axial	9.36	60	10	0.070
2	Axial	2.63	60	10	0.036
3	Axial	6	23.36	10	0.058
4	Factorial	8	40	5	1.704
5	Factorial	8	80	5	0.102
6	Axial	6	60	1.59	0.068
7	Center	6	60	10	0.052
8	Center	6	60	10	0.049
9	Axial	6	93.63	10	0.144
10	Factorial	4	40	5	0.061
11	Factorial	4	80	5	0.044
12	Factorial	8	80	15	0.131
13	Factorial	8	40	15	0.086
14	Center	6	60	10	0.044
15	Factorial	4	80	15	0.095
16	Center	6	60	10	0.053
17	Center	6	60	10	0.045
18	Factorial	4	40	15	0.030
19	Center	6	60	10	0.045
20	Axial	6	60	18.41	0.068

105 * Space type indicates the geometric location of each run within the design space.

106 **Table S3** ANOVA table showing the statistical output of the final model used to
 107 describe the evolution of the PFAT5 as a function of the A) propofol concentration, B)
 108 shaking angle and C) time.

Source	Sum of Squares	Df	Mean Square	F-value	p-value	
Model	1057.64	8	132.20	19.88	< 0.0001	significant
A-Concentration	96.40	1	96.40	14.50	0.0026	
B-Angle	60.43	1	60.43	9.09	0.0118	
C-Time	13.98	1	13.98	2.10	0.1750	
AB	60.58	1	60.58	9.11	0.0117	
BC	226.17	1	226.17	34.01	0.0001	
B ²	170.59	1	170.59	25.65	0.0004	
C ²	92.23	1	92.23	13.87	0.0034	
AB ²	21.73	1	21.73	3.27	0.0981	
Residual	73.13	11	6.65			
Lack of Fit	61.17	6	10.20	4.25	0.0668	not significant
Pure Error	11.98	5	2.40			
Cor Total	1130.80	19				

109 **Abbreviations:** Df = degrees of freedom.