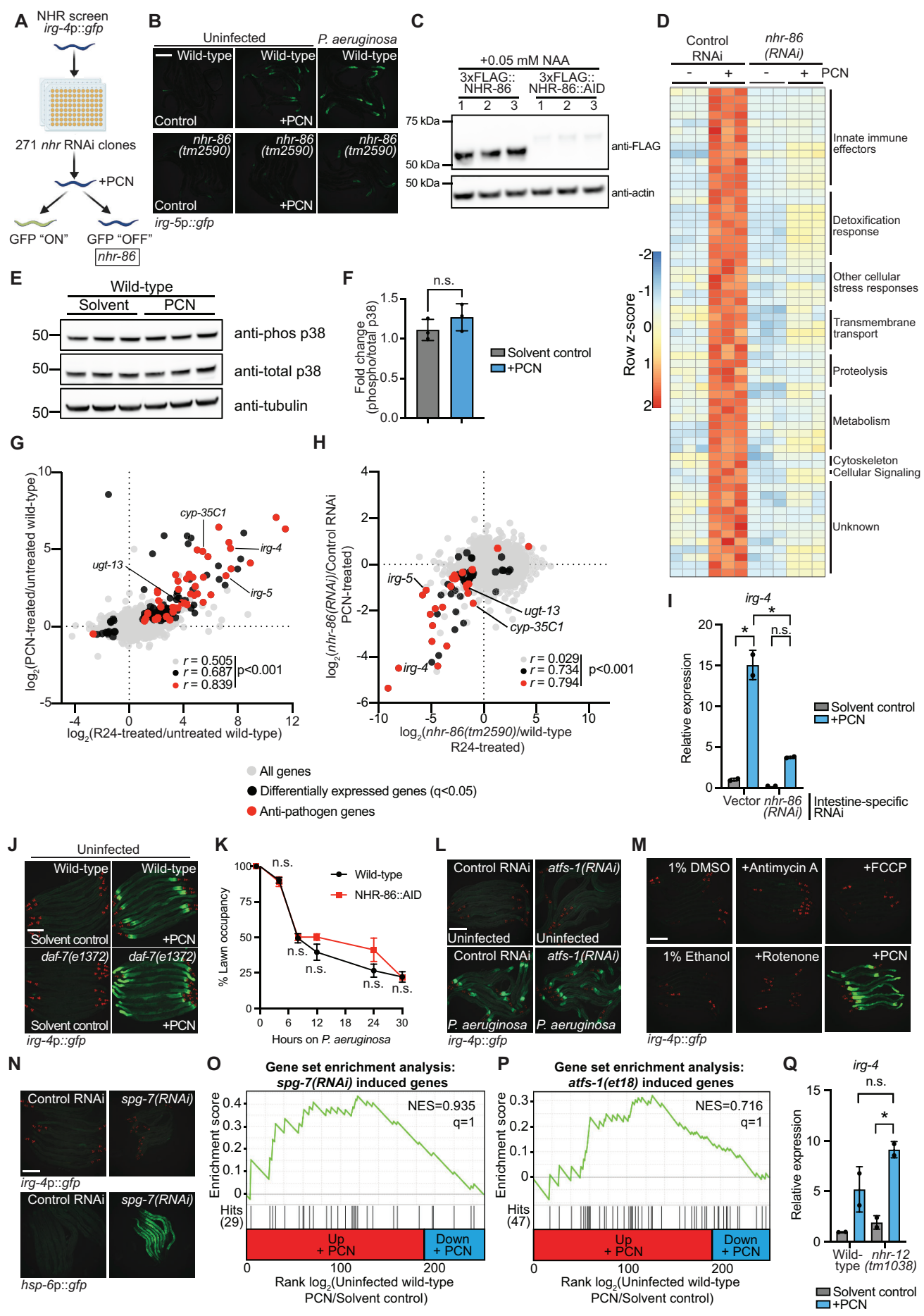
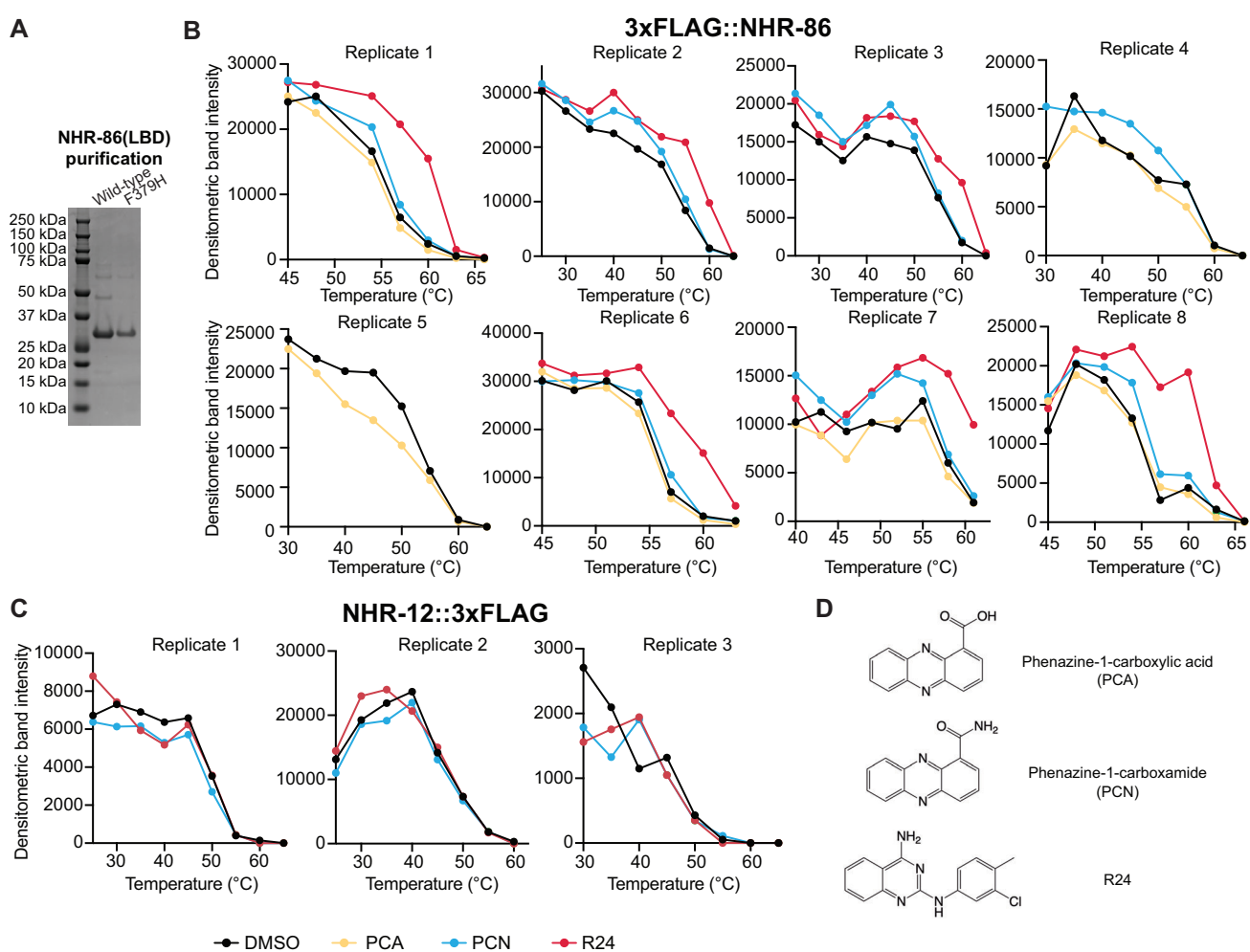


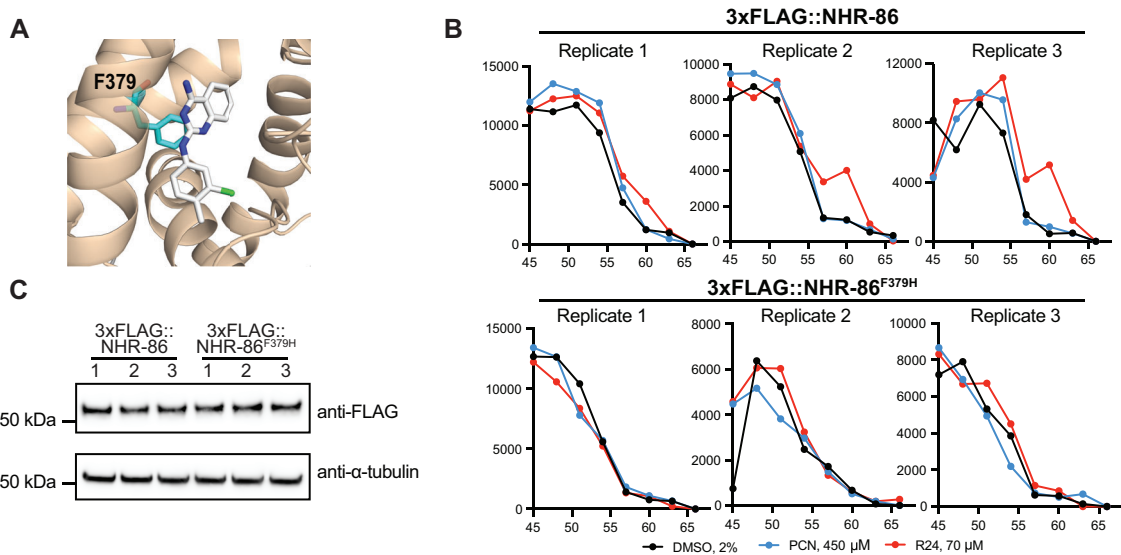
**Figure S1. The pathogen-derived metabolite phenazine-1-carboxamide (PCN) activates anti-pathogen defenses in the *C. elegans* intestine, related to Figure 1. (A)** A schematic of the primary screen of 17 *P. aeruginosa* virulence-related transcription factors using *C. elegans irg-4p::gfp* transcriptional immune reporter animals. Hits are indicated: *rhIR*, *pqsR*, and *lasR*. **(B)** Representative images of *C. elegans irg-4p::gfp* transcriptional reporter animals either uninfected or infected with the indicated *P. aeruginosa* strains. **(C)** Quantification of *irg-4p::gfp* transcriptional reporter intensity in animals either uninfected or infected with the indicated *P. aeruginosa* strains. Sample size (*n*) for each strain is indicated. Each data point indicates one animal. Box and whisker plots represent the median with minimum, second quartile, third quartile, and maximum indicated for each condition. \*equals  $p < 0.05$  (one-way ANOVA with Dunnett's multiple comparisons test). **(D)** A schematic of the secondary screen of 152 *P. aeruginosa* mutant strains, each with a mutation in a gene dependent on the transcription factor *rhIR* for full expression. Hits are indicated: *phzA2*, *phzB2*, and *phzH*. **(E-G)** Images of the *C. elegans* transcriptional reporters *irg-4p::gfp* **(E)**, *irg-5p::gfp* **(F)**, and *cyp-35C1p::gfp* **(G)** animals either uninfected or infected with the indicated *P. aeruginosa* strains. **(H-L)** Images of *cyp-35C1p::gfp*, *irg-4p::gfp*, and *irg-5p::gfp* transcriptional reporter expression in animals with indicated conditions and as described in Figure 1. **(M)** Quantification of phenazines in *P. aeruginosa* wild-type and *phzH::Tn* replicates by LC-MS/MS. Data are the mean with errors bars giving SEM of biological replicates ( $n=3$ ). \*equals  $p < 0.05$  (two-way ANOVA with Šidák's multiple comparisons test). See also Table S3 for the LC-MS/MS phenazine retention times. Scale bars in all images equal 200  $\mu\text{m}$ . Source data for this figure is in Table S3.



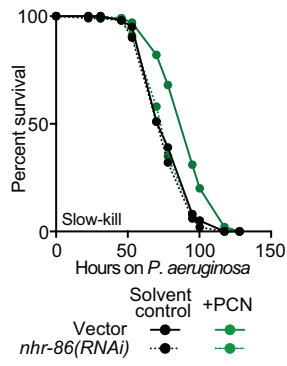
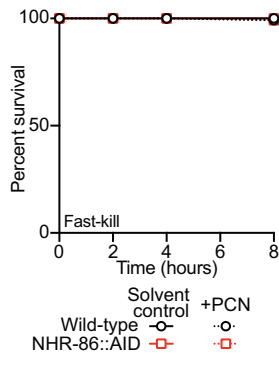
**Figure S2. The anti-pathogen transcriptional program induced by PCN requires the *C. elegans* nuclear hormone receptor *nhr-86*, related to Figure 2. (A)** A schematic of the screen of 271 *nhr* RNAi clones using *C. elegans* *irg-4p::gfp* transcriptional immune reporter animals. The lone hit is indicated: *nhr-86*. **(B)** Images of *irg-5p::gfp* transcriptional reporter expression with indicated genotypes and conditions, as described in Fig. 2C. Scale bars in all images equal 200  $\mu$ m. **(C)** Immunoblot analysis of lysates from biological replicates of 3xFLAG::NHR-86 and 3xFLAG::NHR-86::AID animals expressing the TIR1 transgene in all somatic tissues treated with 50  $\mu$ M NAA ( $n=3$ ). Lysates were probed with anti-FLAG and anti-actin antibodies. The expected size of 3xFLAG::NHR-86 and 3xFLAG::NHR-86::AID is 49.8 kDa and 54.6 kDa, respectively. **(D)** A heat map of the 63 genes that are induced in *C. elegans* during PCN exposure in an *nhr-86*-dependent manner ( $q<0.05$  RNA-seq analysis, see Materials and Methods). Gene expression in each condition was scaled by calculating a row z-score for each gene. See also Table S1C. **(E)** Immunoblot analysis of lysates from wild-type animals exposed to solvent control (1% DMSO) or PCN (25  $\mu$ g/mL) ( $n=3$ ). Lysates were probed using antibodies that recognize the doubly phosphorylated TGY motif of PMK-1 (phos-PMK-1), total PMK-1 protein (total PMK-1) and tubulin ( $\alpha$ -tubulin). PMK-1 is a 43.9 kDa protein and tubulin is a 50 kDa protein. **(F)** The band intensities of three biological replicates of the Western blot shown in Fig. S2E were quantified. Error bars represent SEM. n.s.=not significant (unpaired t-test,  $n=3$ ). **(G)** An mRNA-seq experiment as described in Fig. 1H, except the genes differentially expressed in wild-type animals exposed to R24 in the absence of infection is compared to the genes induced by PCN. See also Table S1D. **(H)** Data from an mRNA-seq experiment as described in Fig. 2L, except the genes that require *nhr-86* for their expression during PCN and R24 treatment are compared. See also Table S1E. **(I)** qRT-PCR analysis of *irg-4* using a strain engineered to perform RNAi only in the intestine. Animals of the indicated genotypes were exposed to solvent control or 25  $\mu$ g/mL PCN ( $n=2$ ). Data are the mean of replicates with error bars giving SEM. \*equals  $p<0.05$  (two-way ANOVA with Tukey's multiple comparisons test). **(J)** Images of *C. elegans* wild-type and *daf-7(e1372)* animals expressing *irg-4p::gfp* exposed to the indicated conditions. **(K)** Occupancy of wildtype and NHR-86::AID animals on a lawn of *P. aeruginosa* was measured over time. Data points are the average of three replicates and error bars represent SEM. n.s.=not significant for comparison at each time point (two-way ANOVA with Šidák's multiple comparisons test). **(L)** Images of *C. elegans* *irg-4p::gfp* reporter treated with *atfs-1(RNAi)* and exposed to PCN. **(M)** Images of *C. elegans* *irg-4p::gfp* animals exposed to either the solvent controls (1% DMSO or 1% ethanol), the indicated mitochondrial toxins, or PCN. FCCP (trifluoromethoxy carbonyl cyanide phenylhydrazone). **(N)** Images of *C. elegans* *irg-4p::gfp* or *hsp-6p::gfp* reporters treated with *spg-7(RNAi)*. The induction of *hsp-6p::gfp* in *spg-7(RNAi)* animals is the control for this experiment. **(O and P)** Gene set enrichment analysis (GSEA) of genes induced in *spg-7(RNAi)* treated animals **(O)** or in the *atfs-1(et18)* gain-of-function allele **(P)** in the RNA-seq of wild-type *C. elegans* exposed to PCN. In **(O)** and **(P)**, fold change in the expression of the significantly differentially expressed genes ( $q<0.05$ ) in uninfected animals exposed to PCN in the absence of infection are ranked from higher expression (red) to lower expression (blue). Normalized enrichment score (NES) and q-value are indicated. Genes induced by either condition and found in the PCN transcriptional profile are indicated by hit number in the left margin and black lines. **(Q)** qRT-PCR analysis of *irg-4* in animals of the indicated genotypes, exposed to solvent control or PCN ( $n=2$ ). Data are the mean of biological replicates with error bars giving SEM. \*equals  $p<0.05$  (two-way ANOVA with Tukey's multiple comparisons test). Scale bars in all images equal 200  $\mu$ m. Source data for this figure is in Table S3.



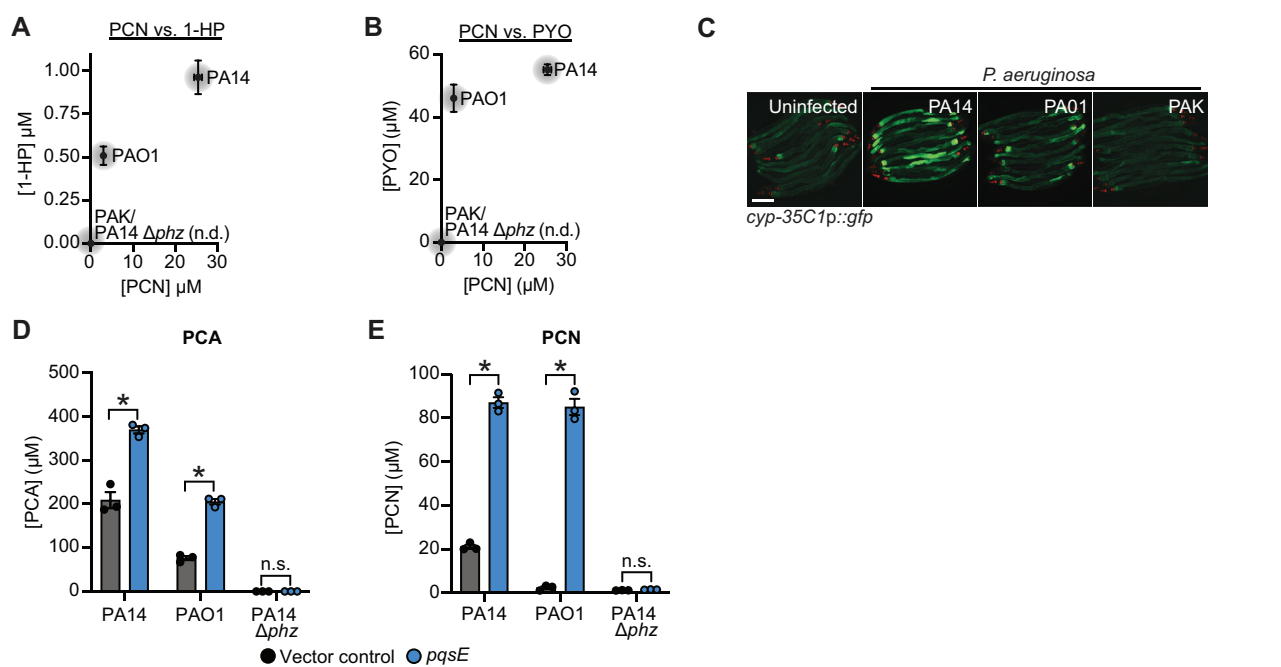
**Figure S3. The bacterial metabolite PCN and synthetic immunostimulatory molecule R24 bind to the ligand-binding domain of NHR-86, related to Figure 3. (A)** SDS-PAGE analysis of purified NHR(LBD) and NHR(LBD)<sup>F379H</sup>, expected molecular weight of both proteins is 32.3 kDa. **(B)** Quantification of 3xFLAG::NHR-86 immunoblot band intensities for each treatment condition and temperature for all replicates. **(C)** Quantification of NHR-12::3xFLAG immunoblot band intensities for each treatment condition and temperature for each replicate. **(D)** Chemical structures of PCA, PCN, and R24. Source data for this figure is in Table S3.



**Figure S4. The phenylalanine at position 379 of the NHR-86 is required for the binding of PCN and R24, related to Figure 4. (A)** An *in silico* model of R24 bound to the identified binding pocket in the NHR-86(LBD). The interaction of phenylalanine 379 (F379) (cyan) and R24 (white) is shown. **(B)** Quantification of 3xFLAG::NHR-86 and 3xFLAG::NHR-86<sup>F379H</sup> immunoblot band intensities for each treatment condition and temperature for all replicates. **(C)** Immunoblot analysis of lysates from biological replicates of 3xFLAG::NHR-86 and 3xFLAG::NHR-86<sup>F379H</sup> animals ( $n=3$ ). Lysates were probed with anti-FLAG and anti- $\alpha$ -tubulin antibodies. The expected size of 3xFLAG::NHR-86 is 49.8 kDa. Source data for this figure is in Table S3.



**Figure S5. The bacterial metabolite PCN is a pattern of pathogenesis sensed by *C. elegans* NHR-86 to activate innate immunity, related to Figure 5. (A) A phenazine toxicity assay (also called the “fast kill” assay) with *P. aeruginosa* and *C. elegans* of the indicated genotypes either treated with solvent control or PCN. (B) *C. elegans*-*P. aeruginosa* infection assay (also called the “slow kill” assay) with the indicated strains and conditions. Sample sizes, mean lifespan, and p-values for all replicates are shown in Table S2. Source data for this figure is in Table S3.**



**Figure S6. *C. elegans* NHR-86 senses PCN as a marker of pathogen virulence to activate protective anti-pathogen defenses, related to Figure 6.** (A-B) HPLC-UV spectroscopy was used to quantify the individual phenazines in the indicated *P. aeruginosa* strains. 1-HP (A) and PYO (B) were compared to PCN production in biological replicates of the indicated *P. aeruginosa* strains ( $n=3$ ). Phenazines not detected (n.d.). (C) Images of *C. elegans* *cyp-35C1p::gfp* animals either uninfected or infected with the indicated *P. aeruginosa* genotypes. Scale bar in all images equals 200  $\mu\text{m}$ . (D and E). Quantification of phenazines PCA (D) and PCN (E) in indicated *P. aeruginosa* strains by HPLC-UV spectroscopy from biological replicates ( $n=3$ ). \*equals  $p < 0.05$  (two-way ANOVA with Tukey's multiple comparisons test). See also Table S3 for the HPLC-UV and LC-MS/MS phenazine quantification retention times and abundance. Source data for this figure is in Table S3.