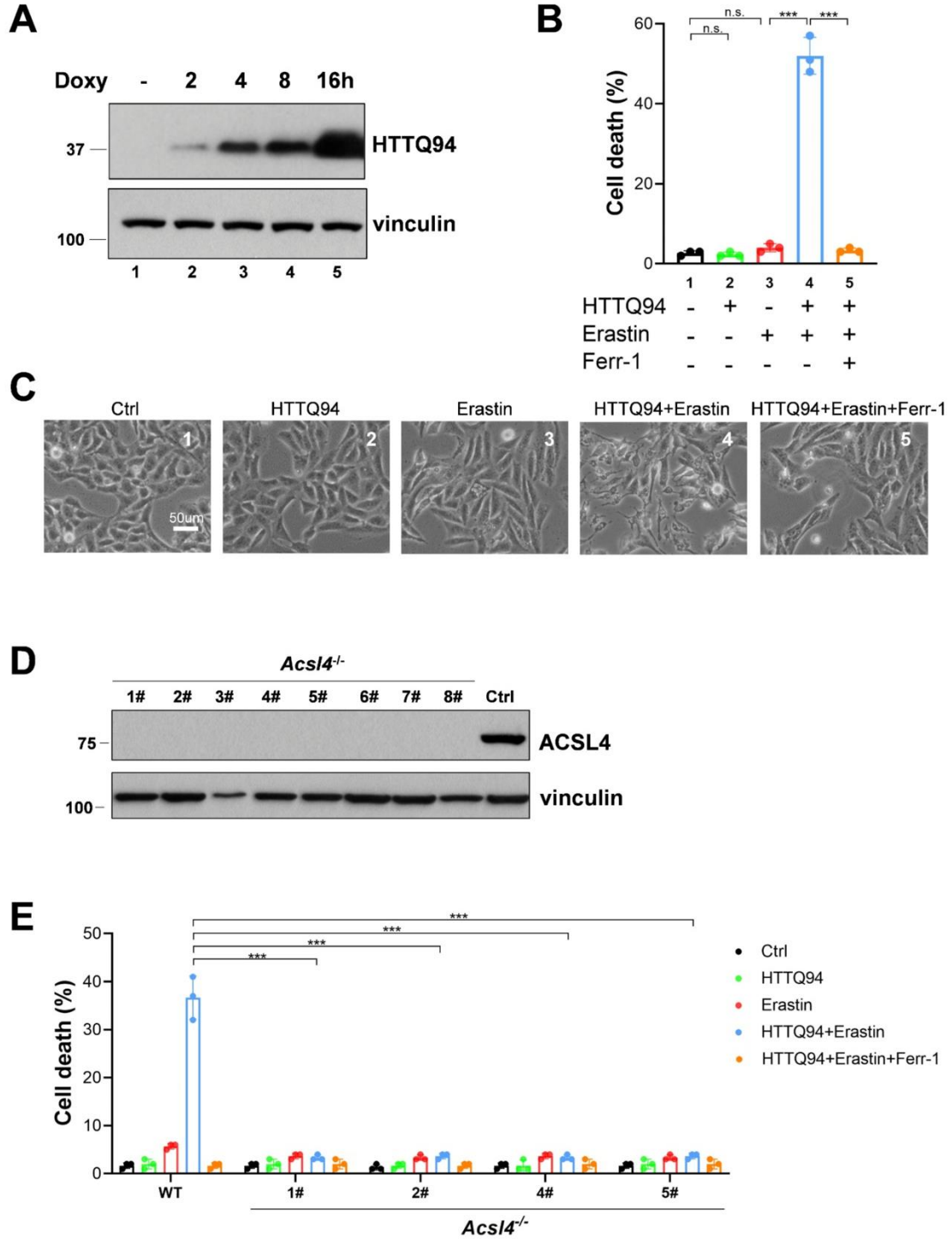


Supplemental Figures and legends

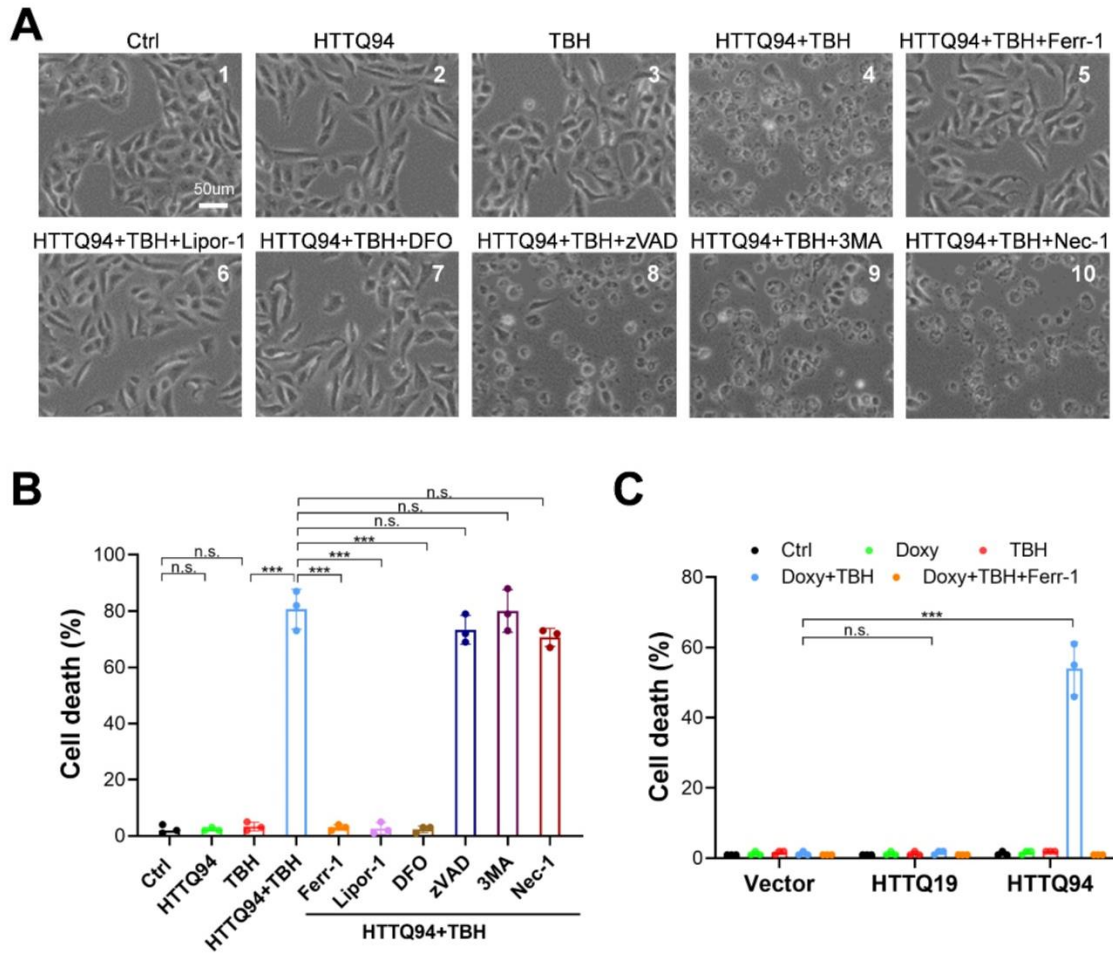
Song et al. Supplemental Figure 1



Supplemental Figure 1. The effect of HTTQ94 on ACSL4-dependent ferroptotic responses.

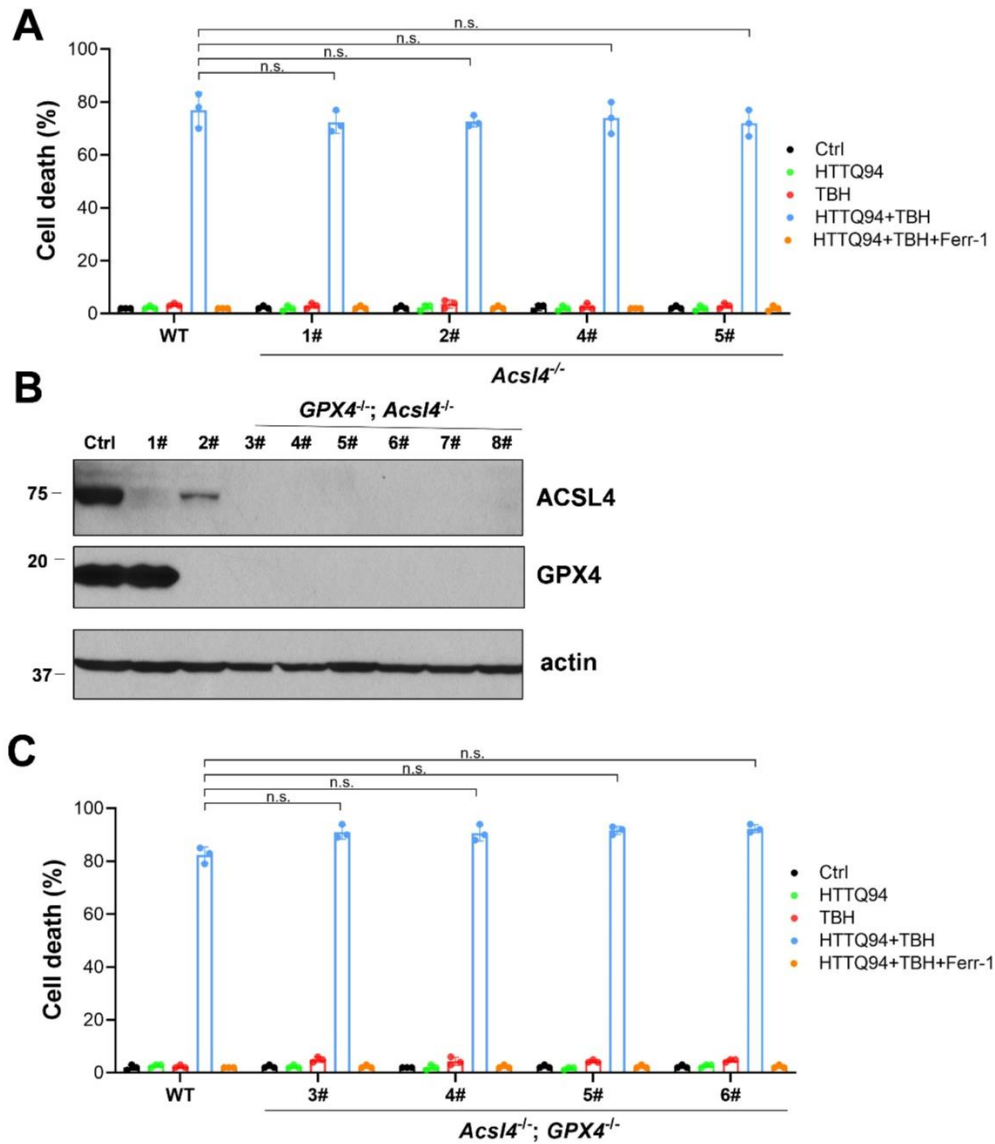
(A) Western blot analysis of HTTQ94 in HTTQ94 Tet-on SK-N-BE(2)C cells treated with doxycycline (0.5µg/ml; Tet) for 2, 4, 8 and 16h. (B) HTTQ94 Tet-on SK-N-BE(2)C cells pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with Erastin (40µM) for 32h with/without Ferr-1 (2µM). (C) Representative phase-contrast images of cell death from, related to panel B. (D) Western blot analysis for ACSL4 in different *Acs14* crispr knockout clones from the HTTQ94 Tet-on SK-N-BE(2)C cell line. (E) Cell death assay. The HTTQ94 Tet-on SK-N-BE(2)C control crispr and four independent *Acs14* crispr cell clones pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with erastin (40µM) for 32h with/without Ferr-1 (2µM). Cell death were calculated from three replicates; Data shown in (B) and (E) are means ± SD. P-values were derived from two-tailed unpaired t-test. (***) P≤0.001, (**) P≤0.01, (*) P≤0.05, (n.s.) P > 0.05.

Song et al. Supplemental Figure 2



Supplemental Figure 2. HTTQ94 mediated ferroptotic responses upon ROS stress. (A) Representative phase-contrast images of cell death from the HTTQ94 tet-on SK-N-BE(2)C cells. HTTQ94 Tet-on SK-N-BE(2)C cells pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (350µM) for 24h in the presence or absence of the ferroptosis inhibitors (ferrostatin-1, Ferr1, 2µM; liproxstatin-1, Lipor1, 2µM and DFO, 100µM), apoptosis inhibitor (Z-VADFMK, zVAD, 10µM), autophagy inhibitor (3-methyladenine, 3MA, 2mM) or necroptosis inhibitor (necrostatin-1, Nec1, 10µM). (B) Quantification of cell death, related to panel A; (C) Cell death assay. Control, HTTQ94 and HTTQ19 fragment Tet-on H1299 cells pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (50µM) for 24h with/without Ferr-1 (2µM). Cell death were calculated from three replicates; Data shown in (B) and (C) are means ± SD. P-values were derived from two-tailed unpaired t-test. (***) P≤0.001, (**) P≤0.01, (*) P≤0.05, (n.s.) P > 0.05.

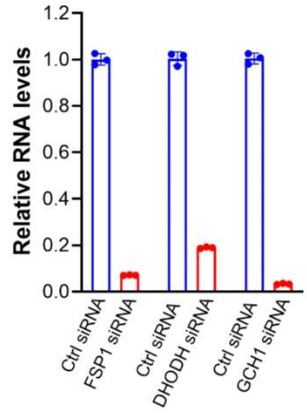
Song et al. Supplemental Figure 3



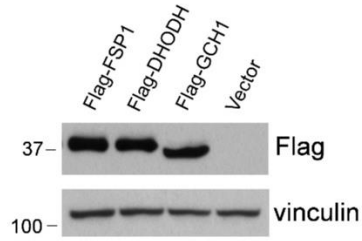
Supplemental Figure 3. HTTQ94 mediated ferroptosis upon ROS stress is independent of GPX4. (A) Cell death assay. HTTQ94 Tet-on SK-N-BE(2)C control crisper and four independent *Acs14* crisper clones pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (350µM) for 24h with/without Ferr-1 (2µM). (B) Western blot analysis of ACSL4 and GPX4 in different *Acs14*/*GPX4* double crisper subclones from the HTTQ94 Tet-on SK-N-BE(2)C cell line. (C) Cell death assay. HTTQ94 Tet-on SK-N-BE(2)C control crisper and four independent *Acs14*/*GPX4* double crisper clones pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (350µM) for 24h with/without Ferr-1 (2µM). Cell death were calculated from three replicates; Data shown in (A) and (C) are means ± SD. P-values were derived from two-tailed unpaired t-test. (n.s.) P > 0.05.

Song et al. Supplemental Figure 4

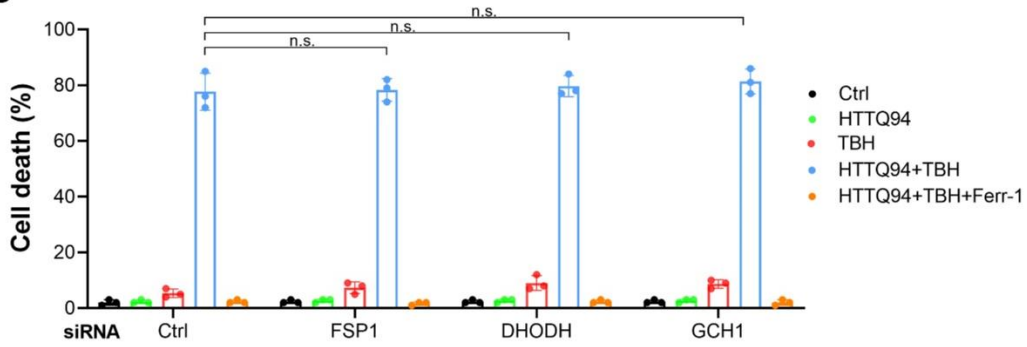
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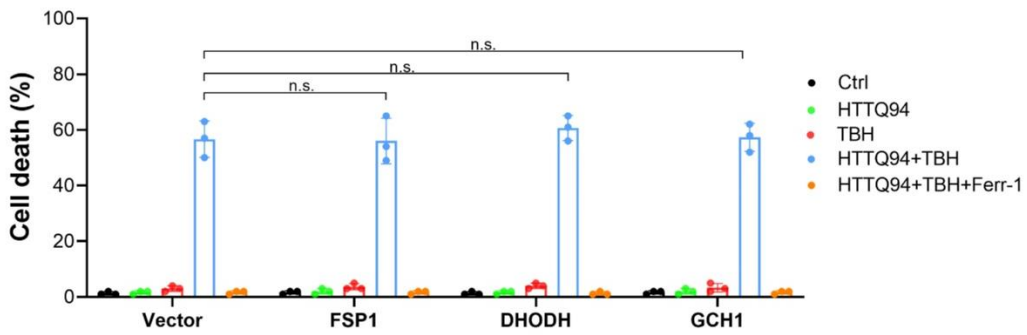
B



C

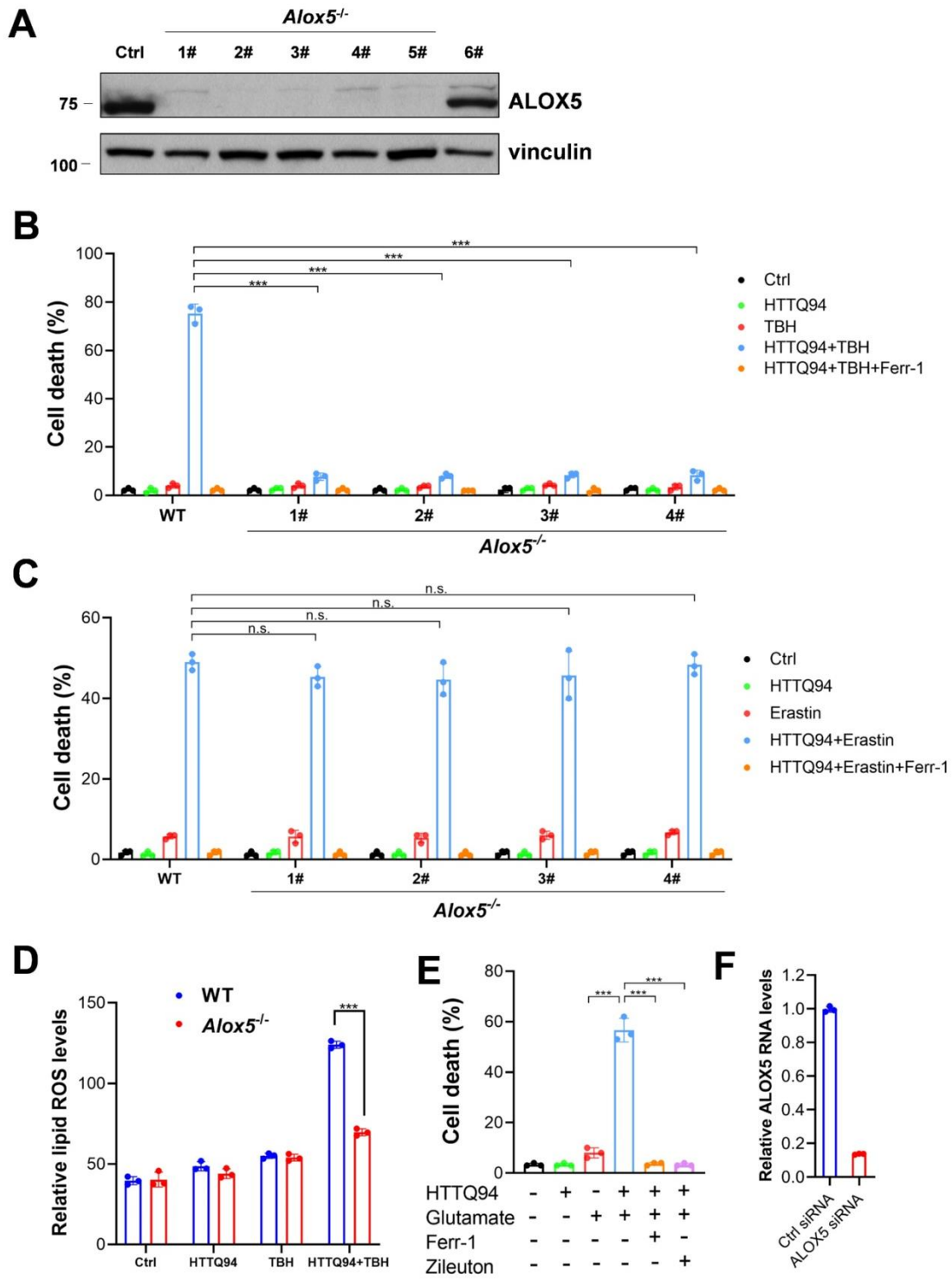


D



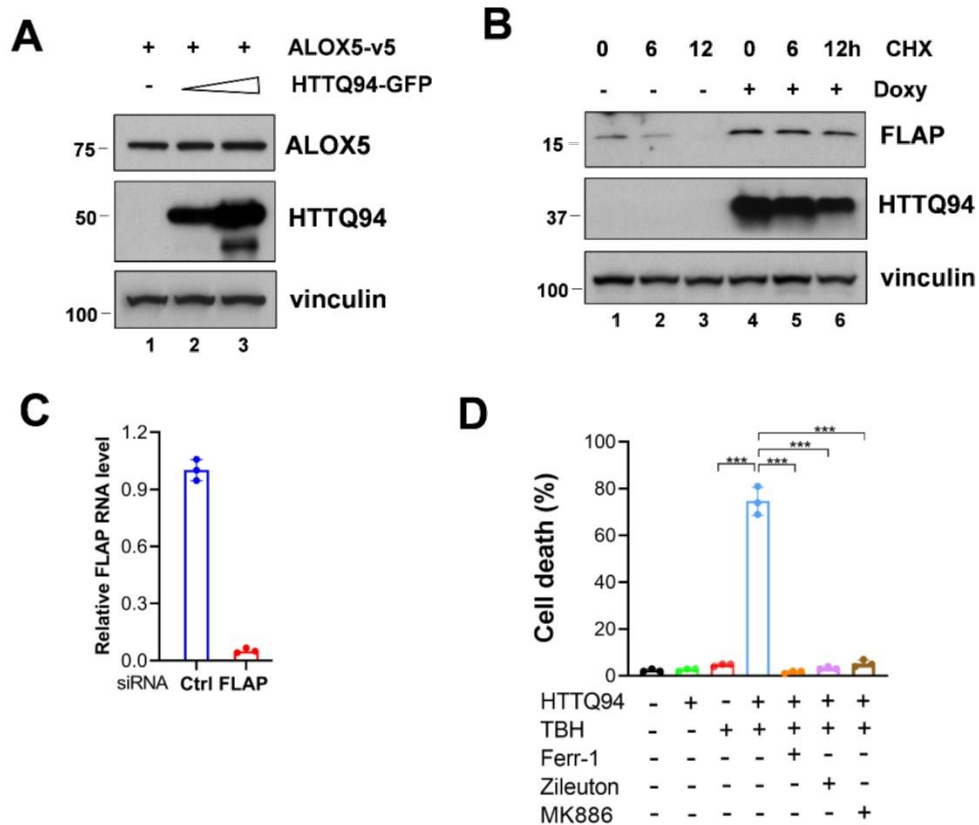
Supplemental Figure 4. HTTQ94 mediated ferroptosis upon ROS stress is independent of FSP1, DHODH and GCH1. (A) Q-PCR analysis of the knockdown efficiency of FSP1, GCH1 and DHODH in HTTQ94 Tet-on SK-N-BE(2)C cells transfected with control, FSP1, GCH1 or DHODH siRNA. (B) Western blot analysis of FSP1, GCH1 or DHODH in HTTQ94 Tet-on H1299 cells transfected with either Flag-FSP1, Flag-DHODH, Flag-GCH1 expressing plasmid or an empty vector. (C) Cell death assays for SK-N-BE(2)C knockdown cells. HTTQ94 Tet-on SK-N-BE(2)C cells transfected with control (ctrl), FSP1, GCH1 or DHODH siRNA were pre-incubated with doxycycline (0.5µg/ml) for 16h, then treated with TBH (350µM) for additional 24h. (D) Cell death assays for the overexpression cells. HTTQ94 Tet-on H1299 cells transfected with either Flag-FSP1, Flag-DHODH, Flag-GCH1 expressing plasmid or an empty vector were pre-incubated with doxycycline (0.5µg/ml) for 16h, then treated with TBH (50µM) for additional 24h. Cell death were calculated from three replicates; Data shown in (C) and (D) are means \pm SD. P-values were derived from two-tailed unpaired t-test. (n.s.) $P > 0.05$.

Song et al. Supplemental Figure 5



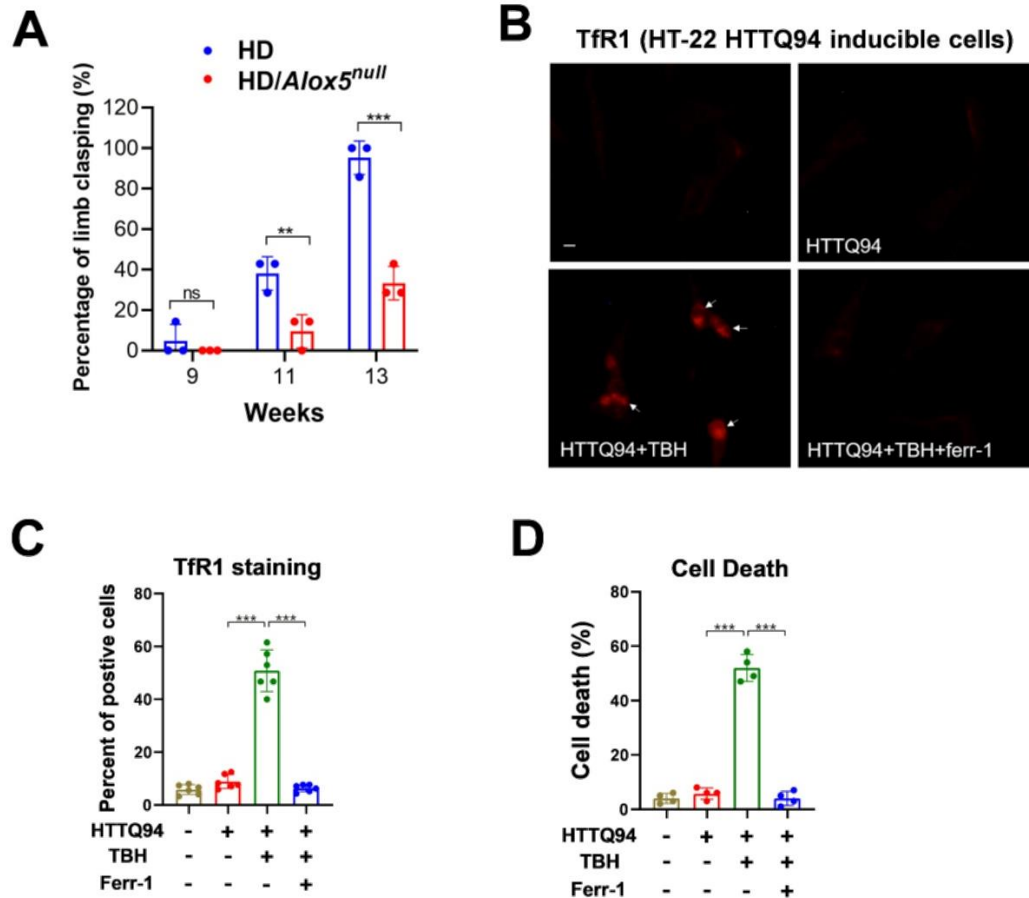
Supplemental Figure 5. Inactivation of ALOX5 abolishes HTTQ94 mediated ferroptotic responses upon ROS-induced stress and glutamate. (A) Western blot analysis of ALOX5 in different *Alox5* crispr subclones from the HTTQ94 Tet-on SK-N-BE(2)C cell line. (B) Cell death assay. The HTTQ94 Tet-on SK-N-BE(2)C control crispr and four independent *Alox5* crispr cell lines pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (350µM) for 24h with/without Ferr-1 (2µM). (C) Cell death assay. The HTTQ94 Tet-on SK-N-BE(2)C control crispr and four independent *Alox5* crispr clones pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with Erastin (40µM) for 32h with/without Ferr-1. (D) Quantification of lipid ROS levels from three replicates, related to main Figure 4F. (E) Cell death assay. The HTTQ94 Tet-on HT-22 cells were treated with doxycycline (0.5µg/ml) and glutamate (10mM) in the presence or absence of Ferr-1(2µM) or Zileuton (10µM) for 20h. (F) Q-PCR analysis of knockdown efficiency of ALOX5 in HTTQ94 Tet-on HT-22 cells, related to main Figure 4G. Data shown in (B), (C), (D) and (E) are means ± SD. P-values were derived from two-tailed unpaired t-test. (***) P≤0.001, (**) P≤0.01, (*) P≤0.05, (n.s.) P > 0.05.

Song et al. Supplemental Figure 6



Supplemental Figure 6. The effect of ALOX5 and/or FLAP on HTTQ94-mediated ferroptosis. (A) Western blot analysis of V5-ALOX5 and HTTQ94 in HEK293 cells transfected with a V5-ALOX5 expressing plasmid and either an empty vector or indicated mHTT-GFP expressing vector. (B) Western Blot analysis for FLAP and HTTQ94 in HTTQ94 Tet-on SK-N-BE(2)C cells pre-incubated with/without doxycycline (0.5µg/ml) for 16h, and then treated with cycloheximide (CHX) at 200µg/ml for the indicated times. (C) Q-PCR analysis of FLAP knockdown efficiency in HTTQ94 Tet-on SK-N-BE(2)C cells transfected with control siRNA or FLAP specific siRNA. (D) Cell death assay. HTTQ94 Tet-on SK-N-BE(2)C cells pre-incubated with doxycycline (0.5µg/ml) for 16h were treated with TBH (400µM) for 24h in the presence or absence of Ferr-1 (2µM), Zileuton (10µM) or MK886 (10µM) for 24h. Data shown in (D) are means ± SD. P-values were derived from two-tailed unpaired t-test. (***) P≤0.001.

Song et al. Supplemental Figure 7



Supplemental Figure 7. TfR1 staining is able to specifically recognize ferroptotic cells induced by HTTQ94 and ROS stress. (A) Limb clasping analysis in HD and HD/*Alox5*^{null} mice (9-, 11- and 13-week-old). $p < 0.01$, $n = 7$. (B-D), TfR1 staining in HT-22 HTTQ94 inducible cells upon TBH treatment. HT-22 HTTQ94 inducible cells were treated 1 μ g/ml doxycycline O/N to induce HTTQ94 expression, followed by treatment with 30 μ M TBH for 6h, and then cells were stained with TfR1 antibody. (B) Representative images of TfR1 staining. (C) Quantification of TfR1 staining. (D) Cell death assay, related to panel B. Data shown in (A) are means \pm SD. Data shown in (C) and (D) are means \pm SEM. P-values were derived from two-tailed unpaired t-test. (***) $P \leq 0.001$, (**) $P \leq 0.01$, (*) $P \leq 0.05$, (n.s.) $P > 0.05$.