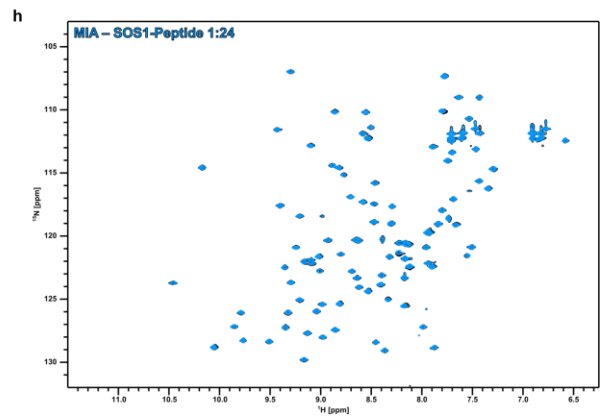
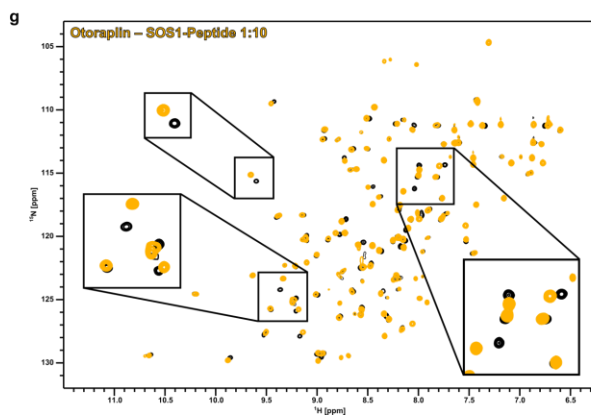
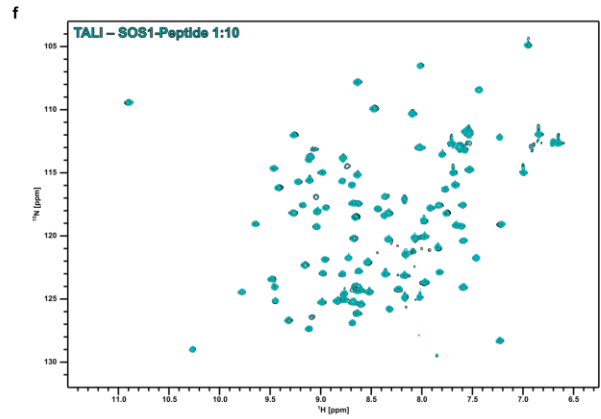
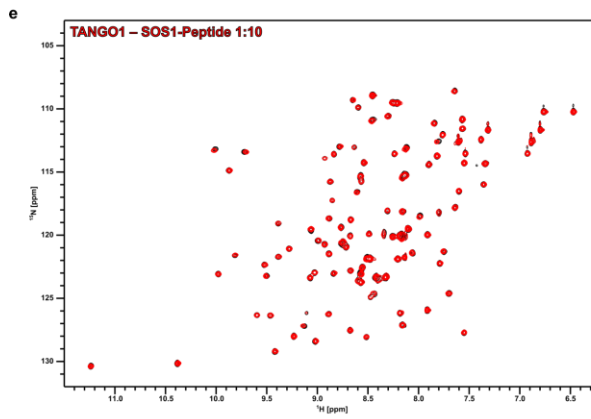
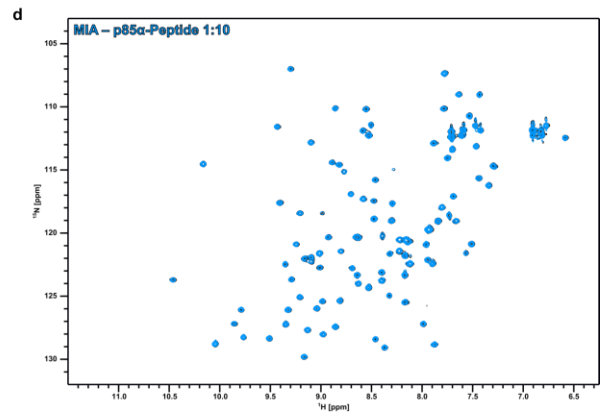
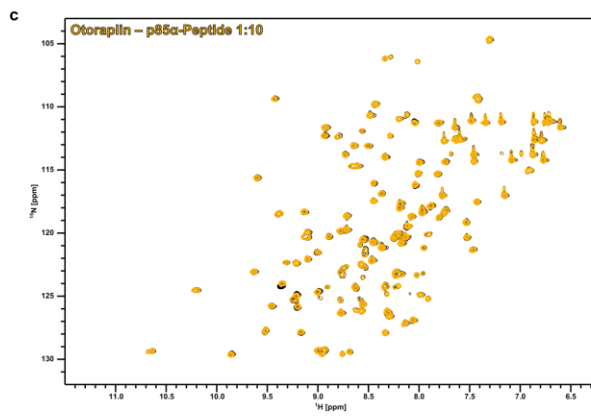
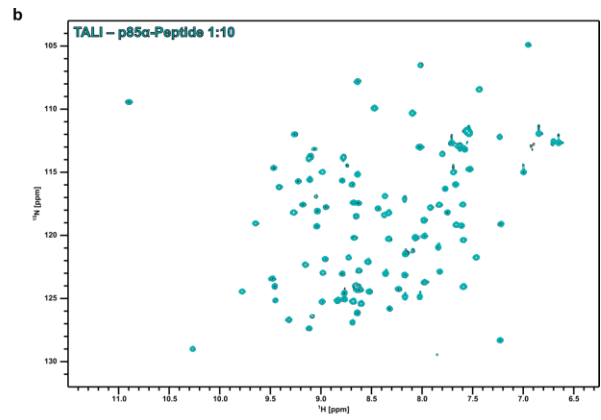
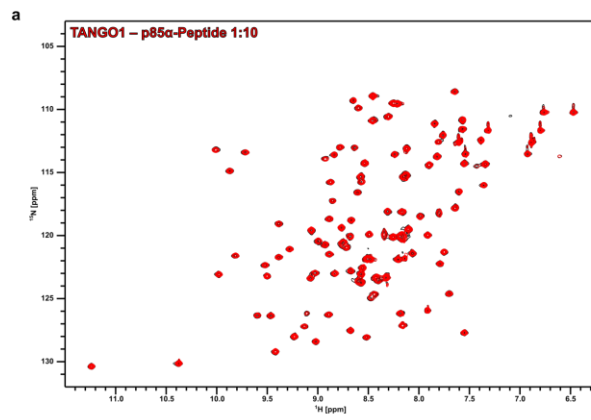


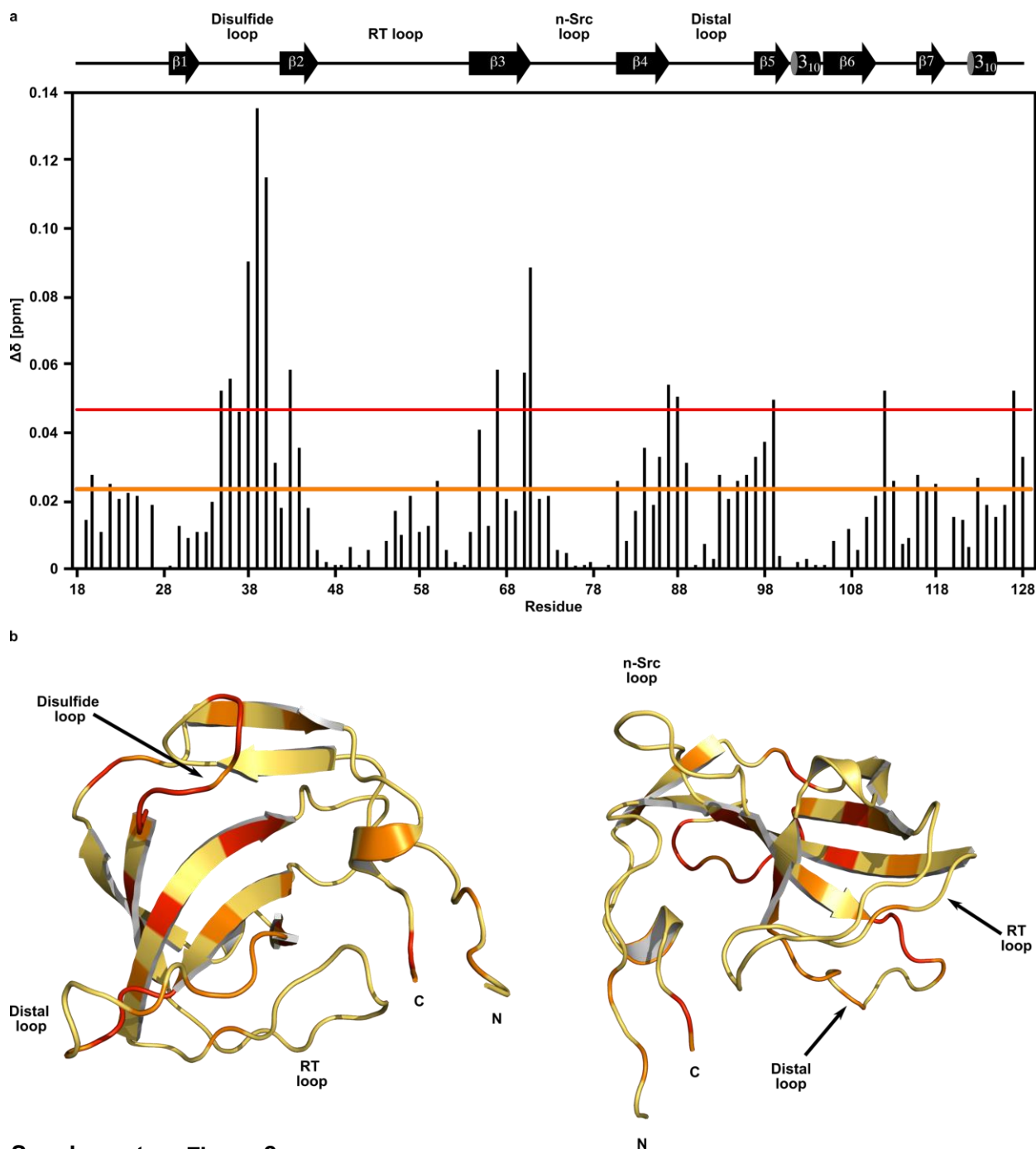
SUPPLEMENTAL INFORMATION



Supplementary Figure 1

¹H¹⁵N-HSQC titration spectra of class I and II PPII ligands with homologous cargo-recognition domains encoded by the *mia* gene family.

a-d, Titration spectra of MOTH domains from TANGO1 (red), TALI (cyan), Otoraplin (yellow), and MIA (blue), respectively, with an exemplary class I PPII ligand. The peptide was derived from residues 91 to 104 of the phosphatidylinositol-3-kinase regulatory subunit alpha (p85 α) known to interact with the SH3 domain of human tyrosine-kinase Fyn. ¹ Reference spectra for all domains are shown in black. **e-h**, Analogous titration spectra with a class II PPII ligand. The peptide sequence corresponds to residues 1149 to 1158 of the guanidine exchange factor Son-of-Sevenless 1 (SOS1) of the Ras protein, reported to bind to the N-terminal SH3 domain of the growth factor receptor-bound protein 2 (Grb2). ² A selection of significant CSPs for the Otoraplin/SOS1 titration is highlighted in **g**.

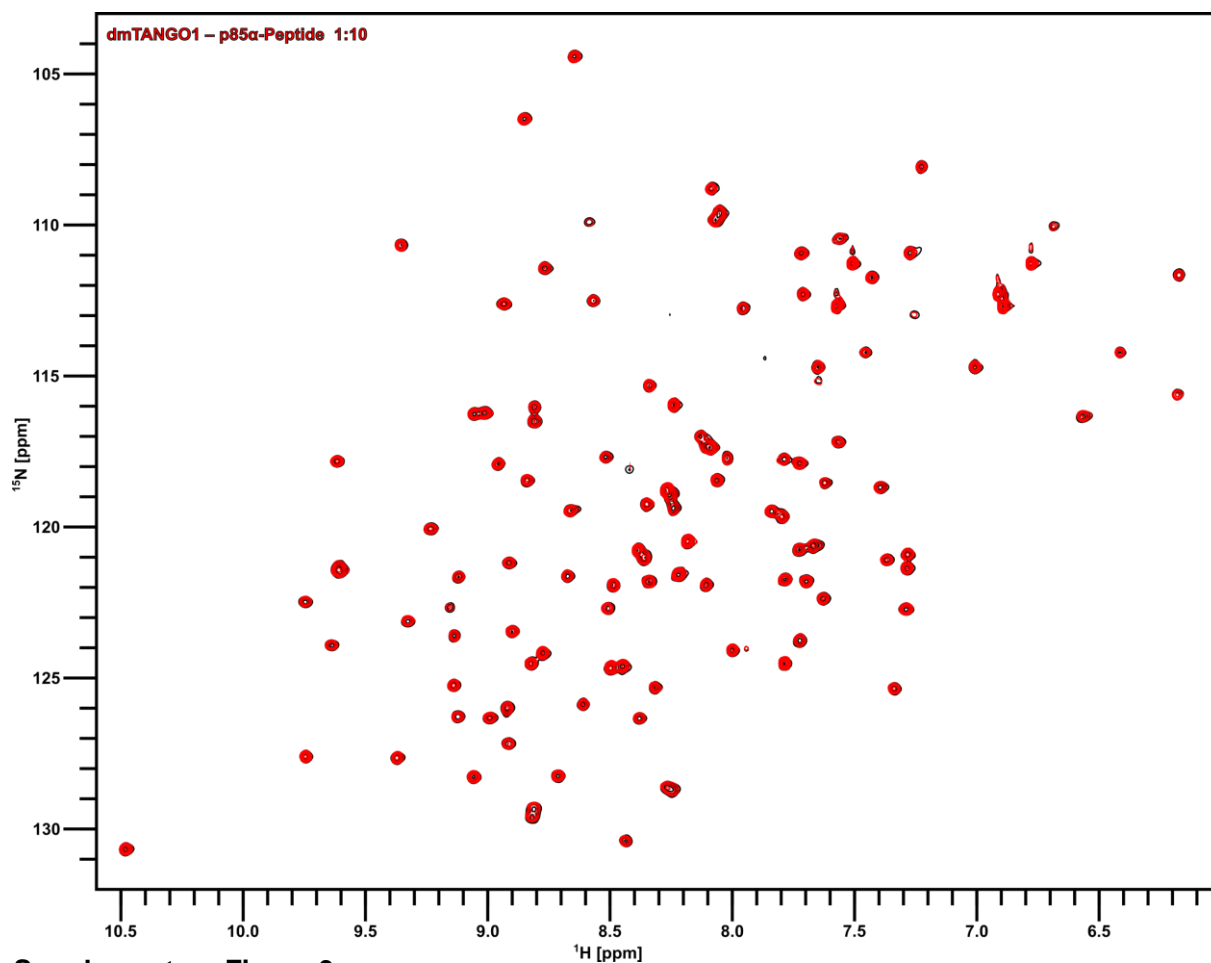


Supplementary Figure 2

CSP analysis of Otoraplin's interaction with a class II PPII ligand.

a, Chemical shift differences between a reference spectrum and a tenfold molar excess of peptide plotted against the amino acid sequence. Source data are provided as a Source Data file. Orange and red line indicate the single and double standard deviation (SD), respectively, based on the average shift difference for all residues. **b**, Chemical shift differences exceeding the single (orange) or double (red) SD projected onto predicted structure of Otoraplin by AlphaFold (AF-Q9NRC9-F1

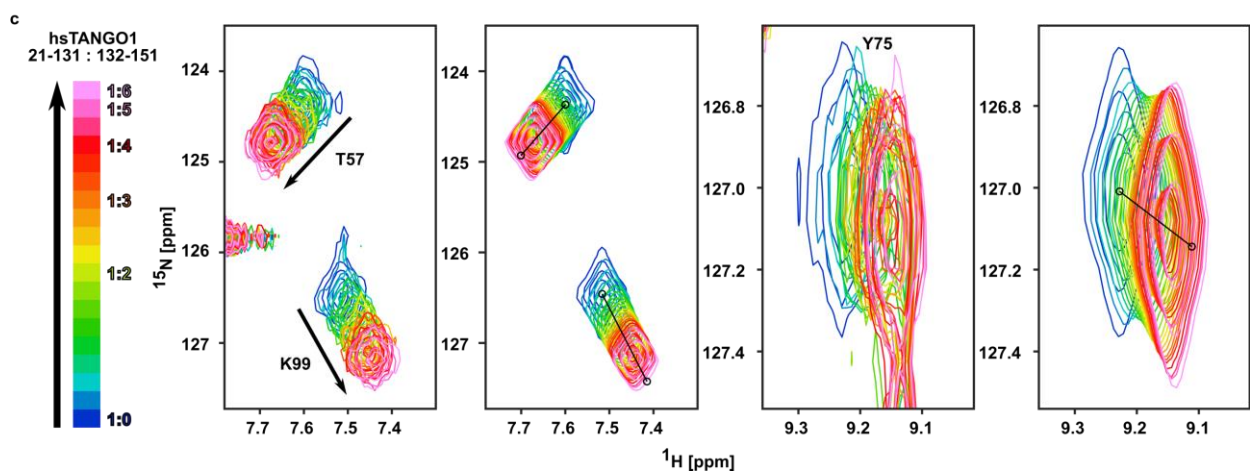
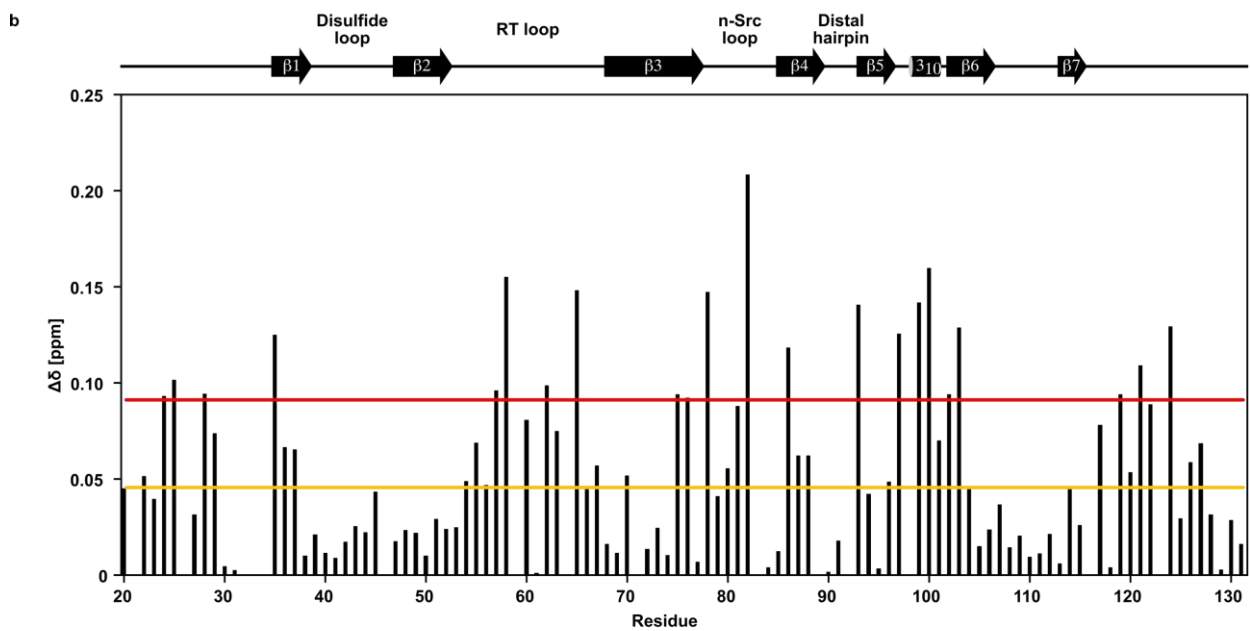
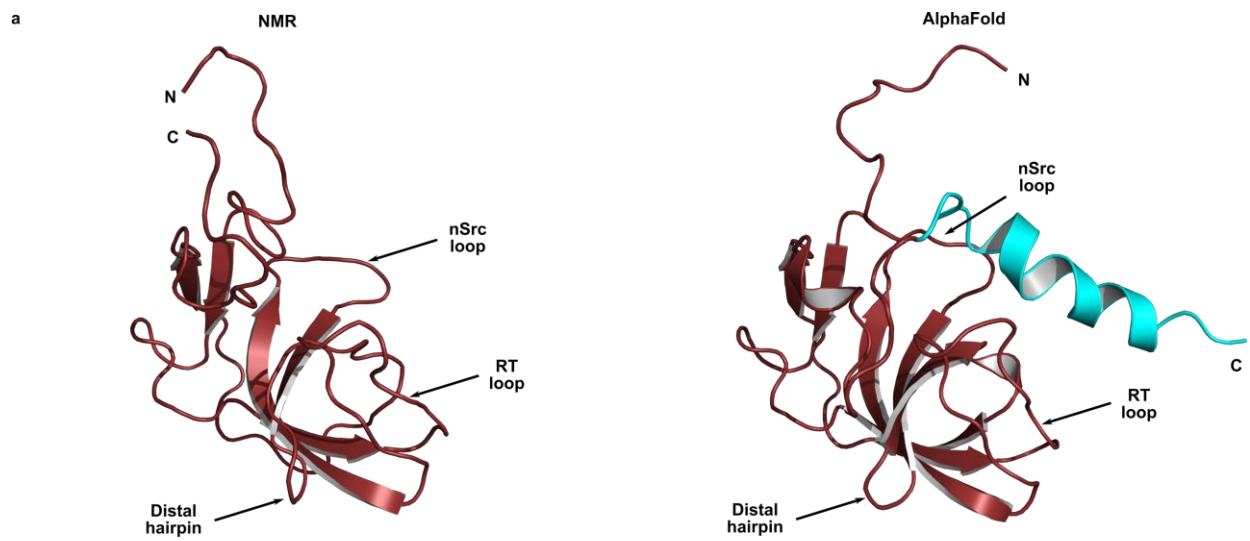
[<https://alphafold.ebi.ac.uk/entry/Q9NRC9>]. Accessed via <https://alphafold.ebi.ac.uk/>). Otoraplin exhibited a very weak interaction with class II ligands, and CSP mapping revealed the interaction site to be located opposite to the canonical SH3 binding groove between the RT and nSrc loop. Moreover, the interaction appears to be mainly mediated by electrostatic interactions between arginine side chains of the peptide and negative charges of the disulfide and distal loop of Otoraplin.



Supplementary Figure 3

$^1\text{H}^{15}\text{N}$ -HSQC of dmTANGO1(30-139) and titration spectrum of PPII class I peptide.

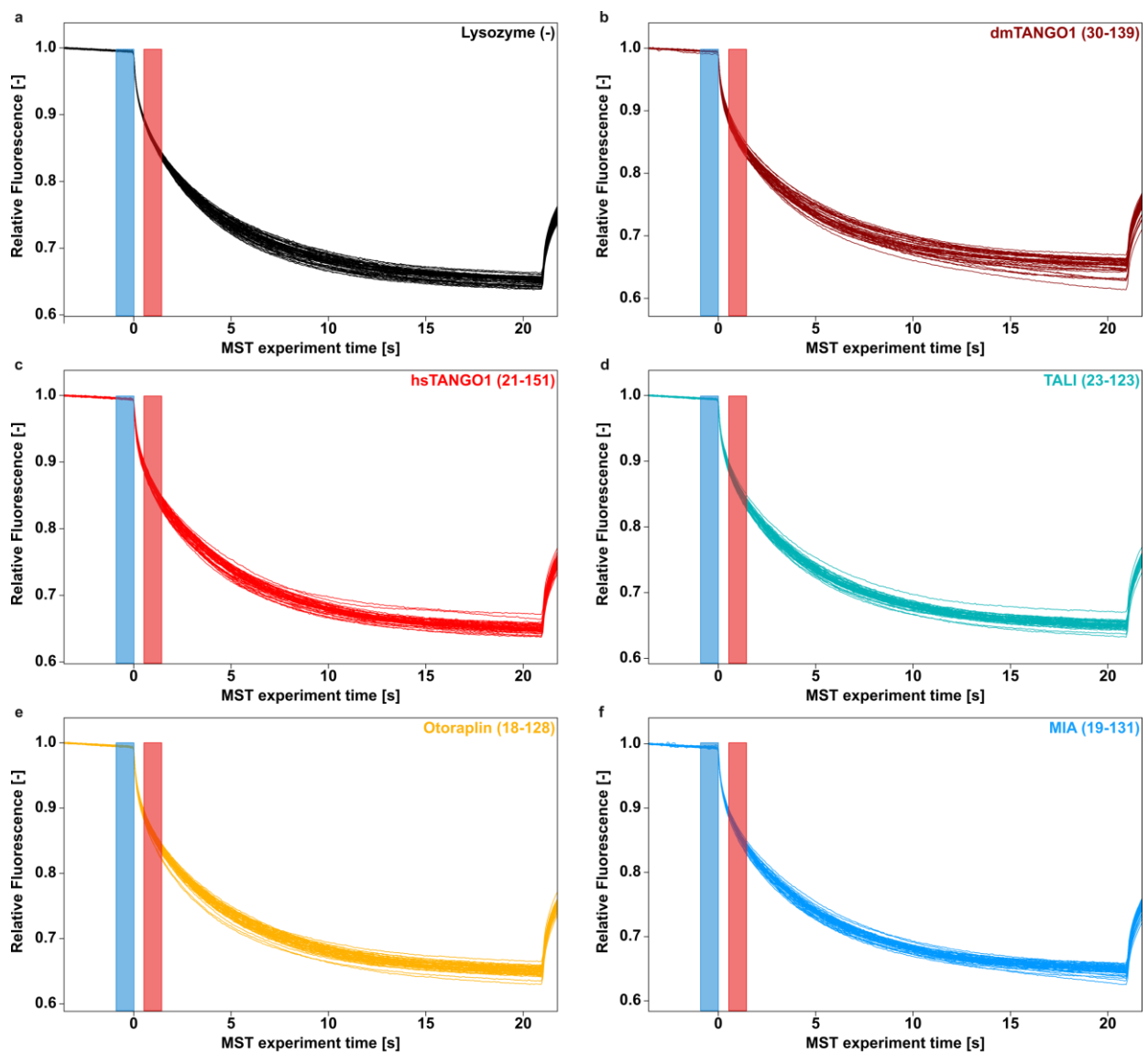
Titration spectra of TANGO1's cargo-recognition domain from *D. melanogaster* with an exemplary class I PPII ligand. The peptide was derived from residues 91 to 104 of the phosphatidylinositol-3-kinase regulatory subunit alpha (p85 α) known to interact with the SH3 domain of human tyrosine-kinase Fyn. ¹ Reference spectrum is shown in black.



Supplementary Figure 4

Comparison of TANGO1's MOTH domain's structure from NMR spectroscopy and AlphaFold's structure prediction.

a, Structures of TANGO1's MOTH domain from structure determination by solution NMR spectroscopy and prediction by AlphaFold (AF-Q5JRA6-F1 [<https://alphafold.ebi.ac.uk/entry/Q5JRA6>]) display an RMSD for the structured region excluding the C-terminal helix of the prediction (cyan) of 0.84 Å. Most differences can be observed for non-secondary structure elements, predominantly for the nSrc loop and region between β -strands six and seven. These exhibit dynamic movements on the pico- to nanosecond timescale according to the hetNOE (Figure 2). **b**, Chemical shift differences observed for the human TANGO1 (21-131) upon titration of a peptide corresponding to residues 132-151 (displayed in **a** in cyan) with a sixfold molar excess. Source data are provided as a Source Data file. Single and double SD based on the average shift difference for all residues are indicated by orange and red line, respectively. **c** Exemplary excerpts of two-dimensional lineshape analysis of the interaction between human TANGO1(21-131) and a peptide corresponding to residues 132 to 151 of human TANGO1 using TITAN. Real spectra are displayed in respective left panels, re-calculated spectra based on fitted parameters are shown in the right panels.



Supplementary Figure 5

MST traces of various *mia* gene members with labeled type IV collagen

Lysozyme served as a negative control. MST-off times used for analysis are indicated with blue, on times by red rectangles. Traces of capillaries displaying aggregation of adsorption were excluded for final analysis and are not shown. Source data are provided as a Source Data file.

Supplementary Table 1.

Statistics for all conformational restraints used in the calculation and geometric quality statistics for the final NMR ensemble of the 20 lowest energy structures for TANGO1's cargo-recognition domain. Deviations \pm standard deviations of this ensemble from averaged coordinates are summarized in the RMSD.

Conformational restraints	
Distance restraints	
Intraresidual ($i = j$)	815
Sequential ($ i - j = 1$)	314
Short-range ($2 < i - j < 3$)	69
Medium-range ($4 < i - j < 5$)	27
Long-range ($ i - j \geq 5$)	404
Ambiguous	175
Dihedral restraints (Φ/Ψ)	195
Other structural restraints	
Oxidized cysteines	4
<i>cis</i> -prolines	1
Total number of restraints	2004
Structure quality	
Average RMSD of secondary structures	[Å]
Backbone	0.29 ± 0.05
Heavy atoms	0.64 ± 0.08
Ramachandran statistics	[%residues]
Core regions	80.1
Allowed regions	18.3
Generous regions	0.7
Disallowed regions	1.0

Supplementary Table 2.

Parameters extracted from microscale thermophoresis dilution series

Protein	K_D [μM]	K_D confidence [μM]	n	Signal to noise
Lysozyme	n.d.	n.d.	4	n.d.
dmTANGO1(30-139)	6.9	3.2	3	7.54
hsTANGO1(21-151)	3.3	1.2	3	10.4
TALI(23-123)	11.4	4.8	3	11.8
Otoraplin(18-128)	9.9	2.9	3	17.1
MIA(19-131)	n.d.	n.d.	3	n.d.

Supplementary Table 3.

Pulse programs used for NMR spectroscopy.

Pulse programs and parameters such as number of scans (NS), amount of recorded data points using non-uniform sampling (NUS), sweep width (SW), and points recorded in the time domain (TD) used for NMR spectroscopy.

Pulse program	F1		F2		F3		F4		Ref		
	NS	NUS	SW [ppm]	TD	SW [ppm]	TD	SW [ppm]	TD			
zggpw5	8	-	15.9	32768	-	-	-	-	-	3	
hsqcfpf3gpplhwg	8	-	29.4	256	15.9	2048	-	-	-	4-7	
hsqcctetgppsp	32	-	80.0	256	13.0	1024	-	-	-	8	
hncagpwg3d	16	-	32.0	128	29.4	48	15.9	2048	-	9-11	
hncacbpgwg3d	128	30%	80	128	29.4	40	15.9	2048	-	12,13	
hncogpwg3d	16	-	9.0	128	29.4	40	15.9	2048	-	9-11	
hncacogpwg3d	64	30%	9.0	128	29.4	40	15.9	2048	-	11,14	
cbcaconhgp3d	64	30%	80.0	128	29.4	40	15.9	2048	-	13,15	
hncocacbpgwg3d	16	25%	80.0	128	30.0	40	15.9	2048	-	16	
hccconhgpwg3d3	64	25%	80.0	128	35.0	40	15.9	2048	-	17-22	
noesyhsqcf3gpwg3d	32	-	14.0	128	29.4	40	15.9	2048	-	23	
hnhagp3d	32	-	29.4	40	14.0	128	15.9	2048	-	24,25	
hccconhgpwg3d2	64	25%	15.9	128	35.0	40	15.9	2048	-	17-22	
noesyhsqcetgp3d	32	-	14.0	128	80.0	64	14.0	2048	-	26	
hcchcogp3d	16	-	14.0	128	80.0	64	14.0	2048	-	27	
hcchdigp3d	16	-	14.0	128	80.0	64	14.0	2048	-	27	
hsqcnoesyhsqccngp4d	8	30%	80.0	32	10.0	64	35.0	32	15.9	2048	28
hsqcnoesyhsqcccgp4d	8	-	80.0	32	10.0	64	80.0	32	15.9	2048	13,23,29,30
hsqcnoef3gpsi3d	32	-	35.0	2	29.4	256	15.9	2048	-	-	23,31

Supplementary Table 4.

Parameters used for calculations with ARIA 2.3.1 for the structure determination of TANGO1's cargo-recognition domain.

General settings	
Assignment frequency window	[ppm]
¹ H	0.04
¹³ C/ ¹⁵ N	0.5
Step 1 – Assignment and unrefined structure calculation	
Violation tolerance [Å]	
Iteration 0	5.0
Iteration 1	4.0
Iteration 2	3.0
Iteration 3 – 5	1.0
Iteration 6 – 8	0.5
Number of conformers	calculated/analyzed
Iteration 0 – 4	800/20
Iteration 5 – 6	200/20
Iteration 7 – 8	200/7
Structures with no violations in iteration 8	33
Simulated-annealing	
Distance restraint potential	log-harmonic
#steps high-temperature	20,000
#steps cooling 1	10,000
#steps cooling 2	8,000
Step 2 – Structure refinement	
Violation tolerance [Å]	
Iteration 0	0.5
Number of conformers	calculated/analyzed
Iteration 0	6,000/50
Refined structures	200
Structures with no violations after refinement	76
Simulated-annealing	
Distance restraint potential	flat-bottom
#steps high-temperature	10,000
#steps cooling 1	20,000
#steps cooling 2	16,000

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