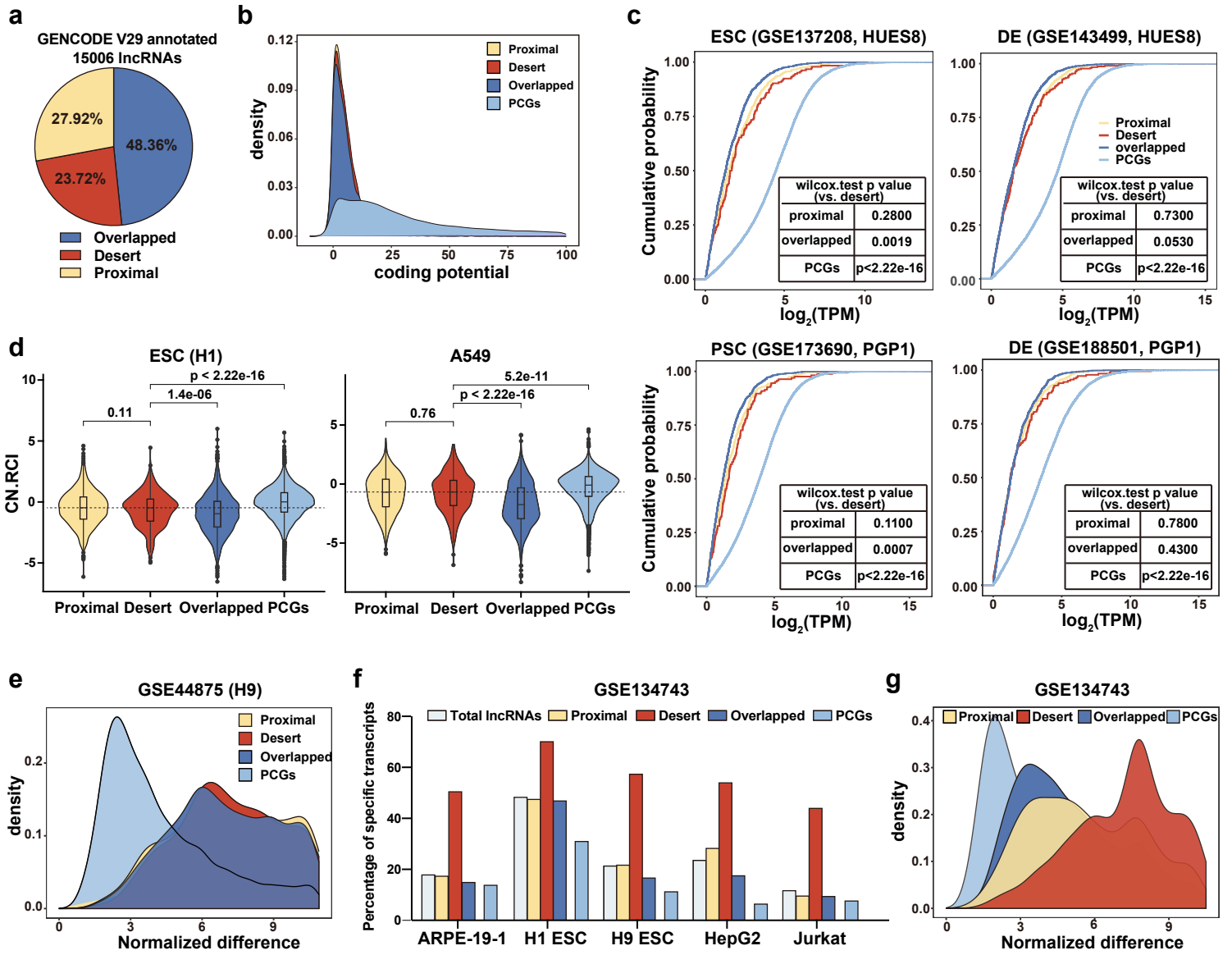


Figure S1



h

Software	Sequence	RNA size	ORF size	Ficket Score	Hexamer Score	Coding Probability	Coding Label	Website
CPAT	HIDEN	1045	237	0.4923	-0.0005937	0.01704	noncoding	http://lilab.research.bcm.edu/cpat/

Software	Sequence	RNA size	ORF size	Ficket Score	ORF integrity	Coding Probability	Coding Label	Website
CPC2	HIDEN	1045	79	0.3653	complete	0.08962	noncoding	http://cpc2.gao-lab.org/

Figure S1. Desert lncRNAs are highly expressed during human endoderm differentiation.

(a) The percentage of overlapped, proximal, and desert lncRNAs in GENCODE V29 annotated 15006 lncRNAs.

(b) The predicted coding potential of lncRNAs and PCGs.

(c) The expression level of differentially expressed lncRNA subsets and PCGs from transcriptome analysis of PSCs and DE cells. The p value between desert lncRNAs and other subsets were listed in the chart.

(d) The subcellular localization of lncRNAs and PCGs, calculated by “relative concentration index” (RCI) based on the transcriptome analysis following cellular compartment separation in H1 ESC and human lung cancer cell A549.

(e) The cell expression specificity of lncRNAs and PCGs in ESCs, DE, pancreatic endocrine (PP), pancreatic alpha and beta cells indicated by Normalized Difference.

(f-g) The cell expression specificity of lncRNAs and PCGs in ESCs and other cell lines such as ARPE-19-1 (human retinal pigment epithelia cell line), HepG2 (liver hepatocellular cancer cell line), Jurkat (immortalized human T lymphocyte cell line), calculated by specificity score (f) or Normalized Difference (g). The GEO accession number was indicated.

(h) The protein-coding potential of *HIDEN* was predicted on CPAT and CPC2 online tools.

Figure S2

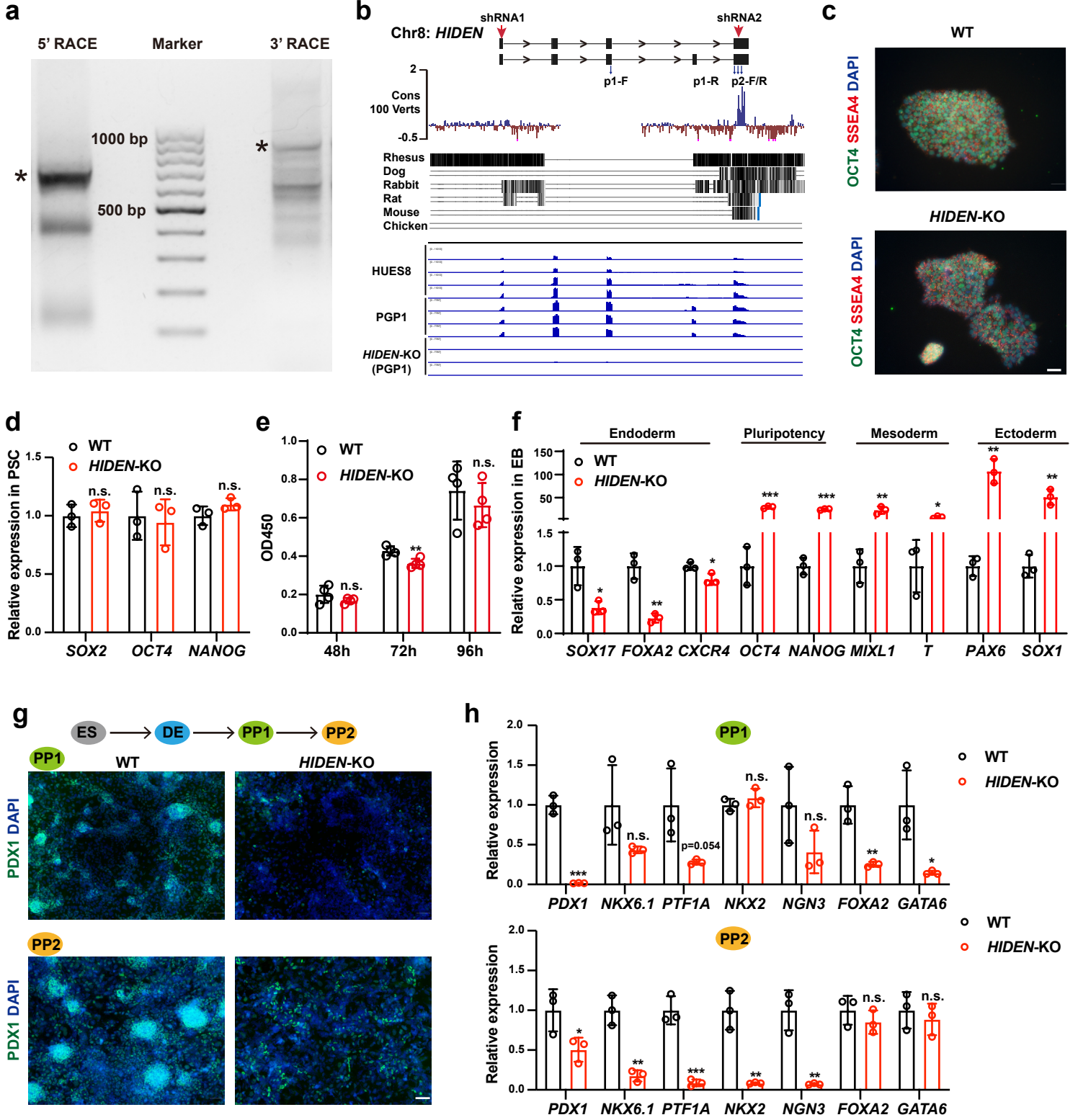


Figure S2. *HIDEN* knockout is not essential for PSC pluripotency but required for endoderm lineage differentiation.

(a) The 5' and 3' ends of the *HIDEN* transcript were defined by RACE-PCR. “**” indicated that the RACE fragment and the lower bands were proved to be non-specific amplification through DNA sequencing.

(b) Top: the full length of *HIDEN* was cloned from the cDNA of HUES8 DE cells. PhyloP value of Vertebrates basewise conservation on *HIDEN* genomic region was shown. *HIDEN* shRNA target sites and qPCR primers were indicated by red arrows and blue arrows respectively. Bottom: the RNA-seq coverage plot of *HIDEN* in HUES8 DE cells, PGP1 DE cells and *HIDEN*-KO DE cells.

(c) Immunofluorescent staining of pluripotency markers (OCT4, SSEA4) in wildtype and *HIDEN* knockout PSCs. Scale bar = 50 μ m.

(d) qPCR analysis of representative pluripotency genes in wildtype and *HIDEN* knockout PSCs (n=3).

(e) Cell proliferation detected by CCK kit at different time points in wildtype and *HIDEN* knockout PSCs (n=4).

(f) RNA expression of representative endoderm genes, pluripotency genes, mesoderm genes and ectoderm genes were examined in wildtype or *HIDEN*-KO EB (n=3).

(g-h) The immunofluorescence staining results of pancreatic marker PDX1 (g) and RNA expression levels of pancreatic transcription factors (h) in wildtype and *HIDEN* knockout cells at PP1 (pancreatic progenitors 1) and PP2 (pancreatic progenitors 2) stage (n=3). Scale bar = 100 μ m.

Figure S3

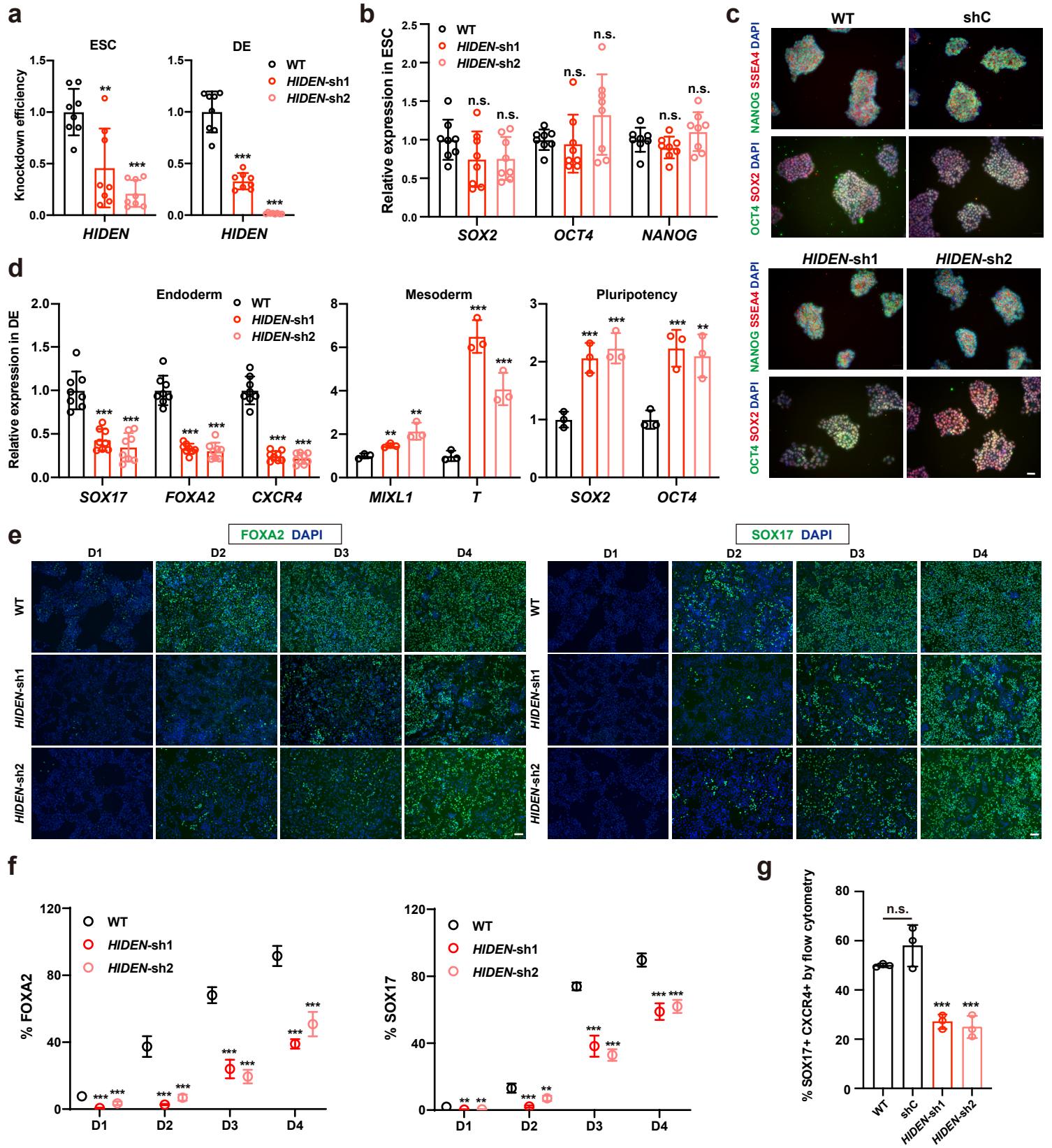


Figure S3. *HIDEN* knockdown exhibits decreased endoderm differentiation.

(a) *HIDEN* KD efficiencies were examined by RT-qPCR in wildtype and *HIDEN*-KD ESCs and DE cells (n=8).

(b) The expression levels of pluripotency genes were examined in wildtype or *HIDEN* knockdown ESCs (n=8).

(c) Immunofluorescence staining results of pluripotency genes (OCT4, SOX2, NANOG, SSEA4) in wildtype, shRNA control (shC) or *HIDEN* knockdown ESCs. Scale bar = 50 μ m.

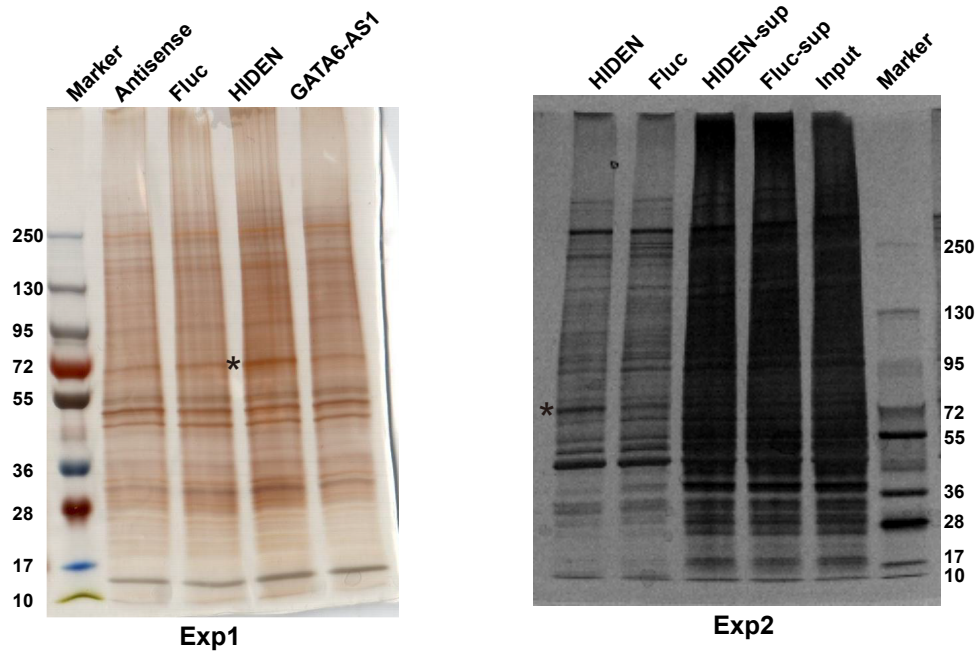
(d) RNA expression levels of marker genes, including endoderm genes (n=8), mesoderm genes (n=3), and pluripotency genes (n=3), were examined in wildtype or *HIDEN* knockdown DE cells.

(e-f) The immunofluorescence staining results of endoderm markers FOXA2 and SOX17 in wildtype and *HIDEN* knockdown DE cells during four days of endoderm differentiation. Scale bar = 100 μ m. Quantitative results were shown (f) (n=5).

(g) The statistical results of flow cytometric analysis of the SOX17 and CXCR4 positive cells in wildtype, shRNA control and *HIDEN* knockdown DE cells (n=3).

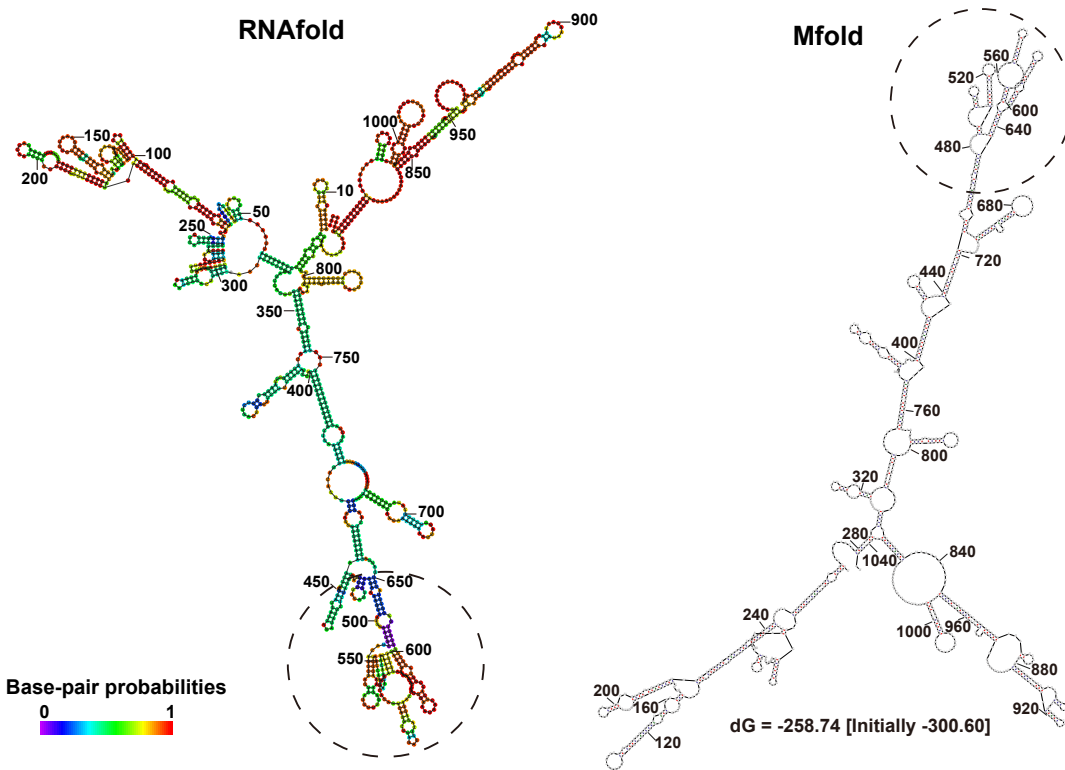
Figure S4

a

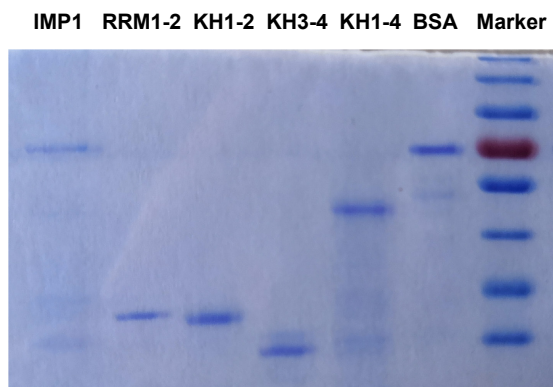


b

Predicted *HIDEN* structure



c



d

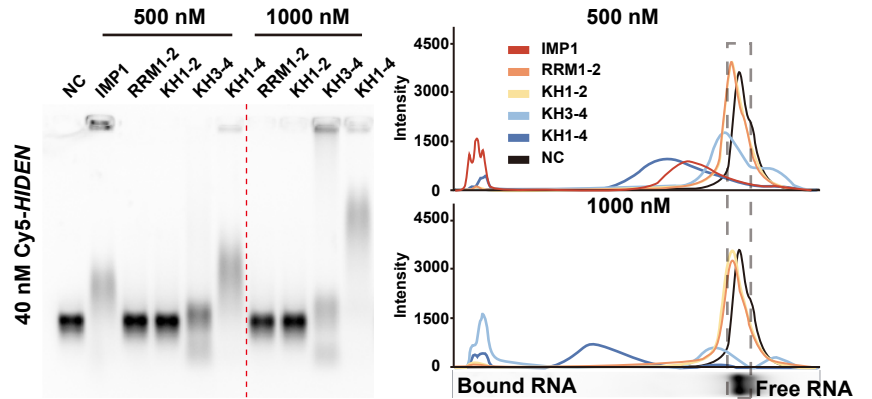


Figure S4. *HIDEN* physically interacts with IMP1.

(a) The silver staining results of RNA pulldown in DE cells. Antisense *HIDEN*, *Luciferase (Fluc)* and unrelated lncRNA *GATA6-AS1* were used as negative controls. “*” indicated the specific band around 72 kDa in *HIDEN*-IP extracts. Two independent experiments were shown.

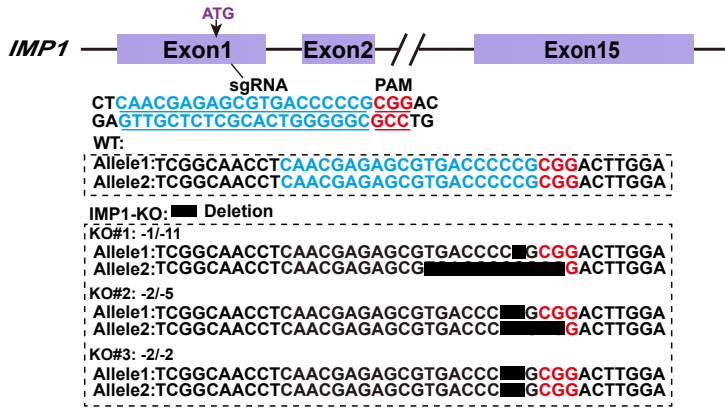
(b) The predicted RNA secondary structure of *HIDEN* using RNAfold and Mfold.

(c) Coomassie brilliant blue staining results of purified His-tagged IMP1 and IMP1 truncations expressed in *E. coli*. BSA was used to normalize protein concentration.

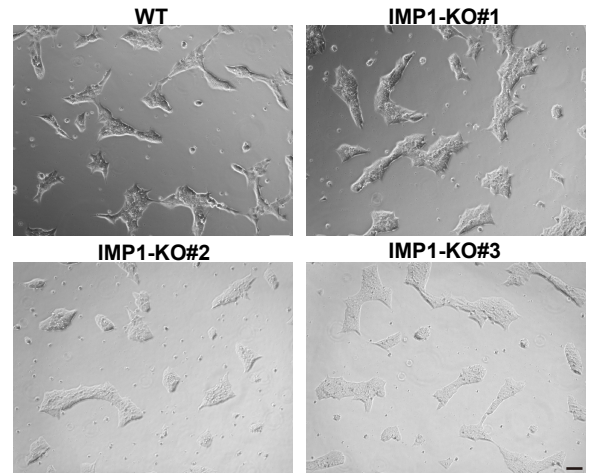
(d) EMSA results of *in vitro* transcribed Cy5-labeled *HIDEN* transcript (40 nM) and IMP1 truncated proteins at different concentrations.

Figure S5

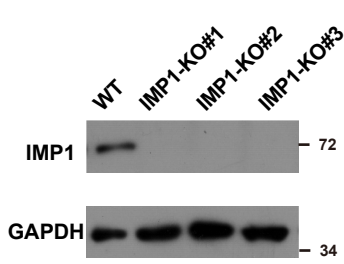
a



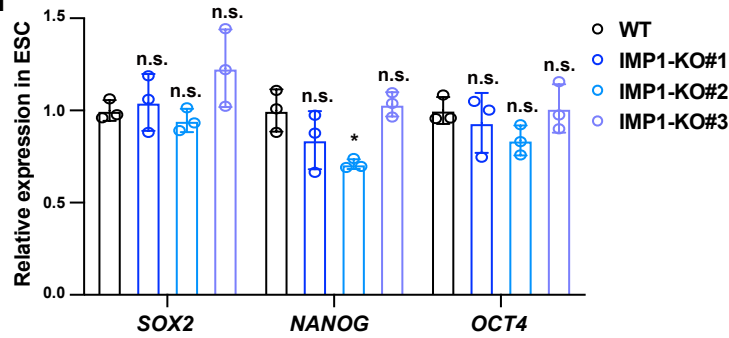
b



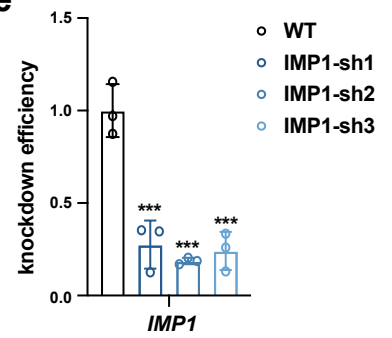
c



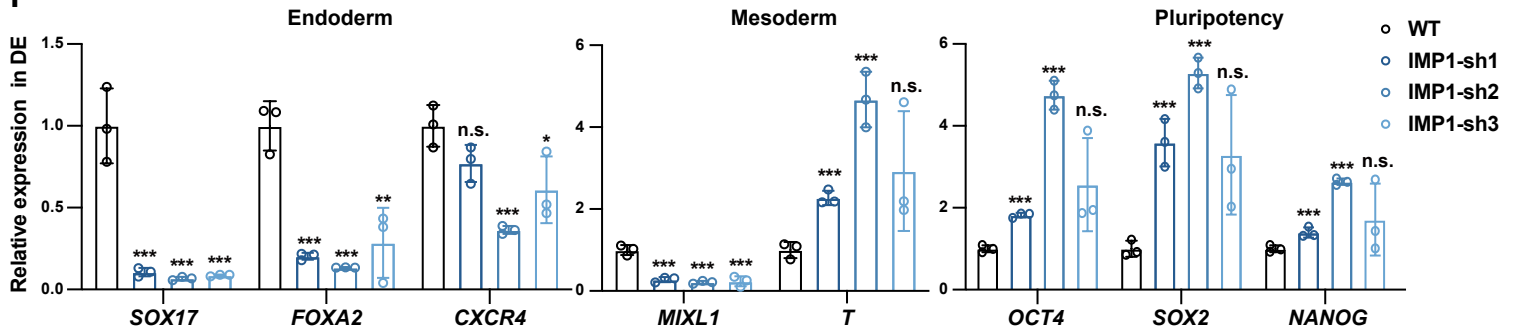
d



e



f



g

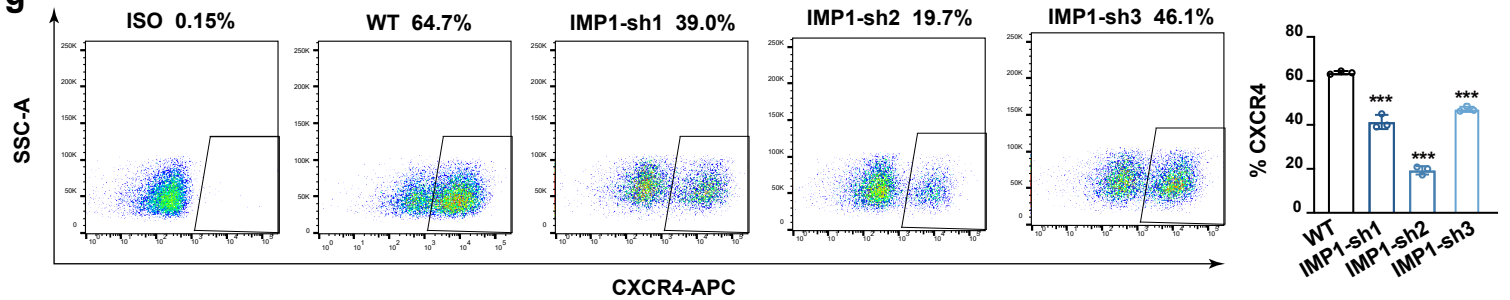


Figure S5. IMP1 is required for DE differentiation.

(a) The genotypes of the three IMP1 knockout H9 ESC lines. sgRNA target sequence and PAM sequence were indicated in blue and red respectively. Deleted nucleotides were marked in black squares.

(b) The bright field image of IMP1 knockout ESCs. Scale bar=200 μ m.

(c) IMP1 knockout detected by immune blot.

(d) The pluripotency gene expression in wildtype and IMP1 knockout ESCs (n=3).

(e) The knockdown efficiency of IMP1 in three IMP1-KD ESC cell lines (n=3).

(f) RT-qPCR analysis of representative endoderm genes, mesoderm genes and pluripotency genes in wildtype and IMP1-knockdown DE cells (n=3).

(g) Flow cytometric analysis of CXCR4⁺ cells in wildtype and IMP1-knockdown DE cells (n=3).

Figure S6

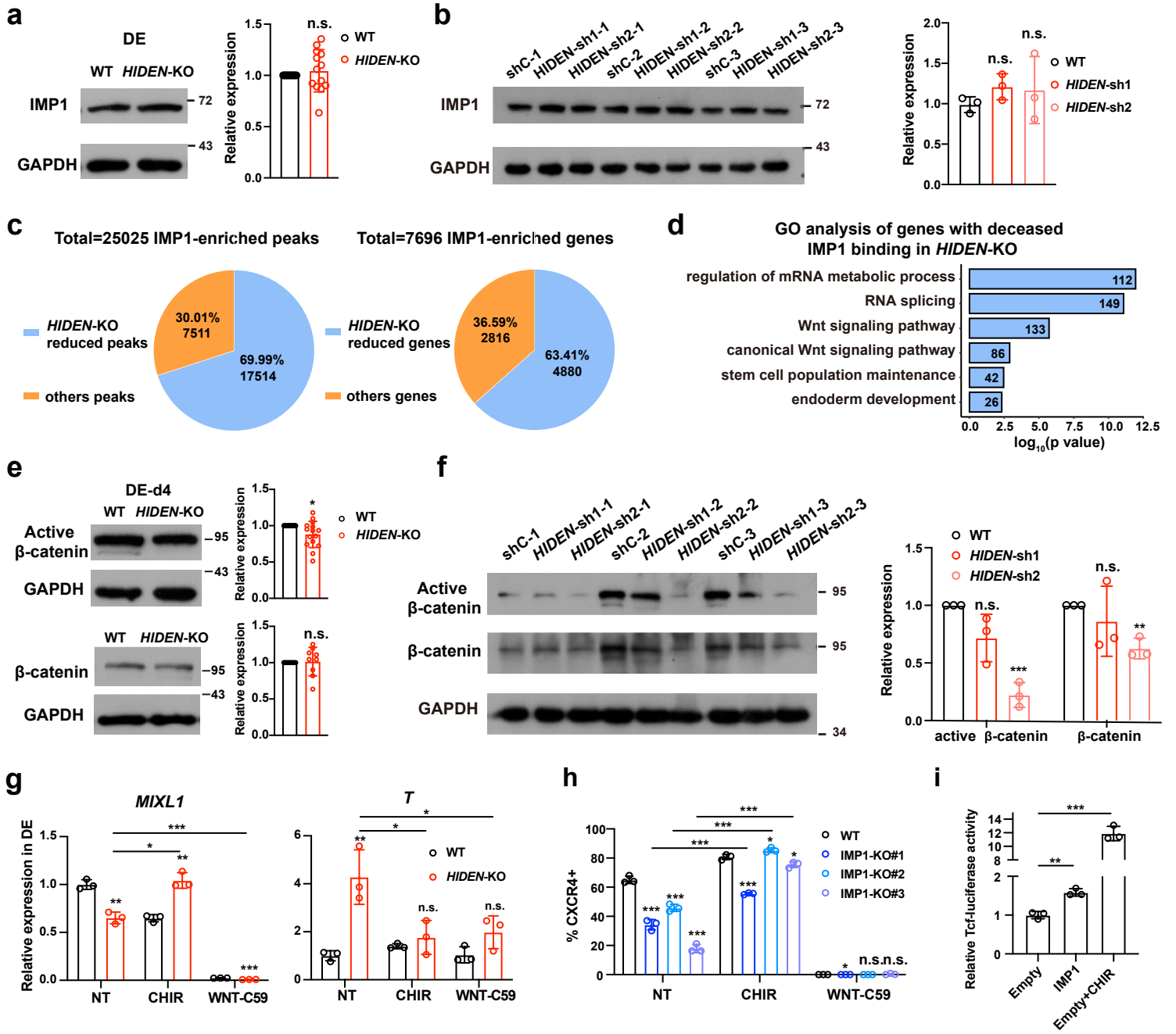


Figure S6. WNT pathway is regulated by *HIDEN*/IMP1.

- (a) The protein level of IMP1 in wildtype and *HIDEN* knockout DE cells (n=13).
- (b) The protein level of IMP1 in wildtype and *HIDEN* knockdown DE cells (n=3).
- (c) IMP1-bound peaks or genes decreased by *HIDEN*-KO, determined by RIP-seq in wildtype and *HIDEN*-KO DE cells were shown.
- (d) The GO analysis of genes with decreased IMP1 binding in *HIDEN*-KO cells.
- (e) The protein level of active β -catenin (n=15) and total β -catenin (n=9) in wildtype and *HIDEN* knockout DE cells.
- (f) The protein level of active β -catenin and total β -catenin in wildtype and *HIDEN* knockdown DE cells (n=3).
- (g) RT-qPCR analysis of mesoderm genes (*MIXL1*, *T*) in wildtype or *HIDEN*-knockout DE cells treated with WNT signaling activator (CHIR at 1 μ M) or inhibitor (Wnt-C59 at 1 μ M) (n=3).
- (h) Flow cytometry analysis of CXCR4⁺ cells in wildtype or IMP1-knockout DE cells treated with WNT signaling activator (CHIR at 1 μ M) or inhibitor (Wnt-C59 at 1 μ M) (n=3).
- (i) The TCF-luciferase activity in 293T transfected with IMP1 or empty vector (n=3). Cells treated with 1 μ M CHIR were used as positive controls.

Figure S7

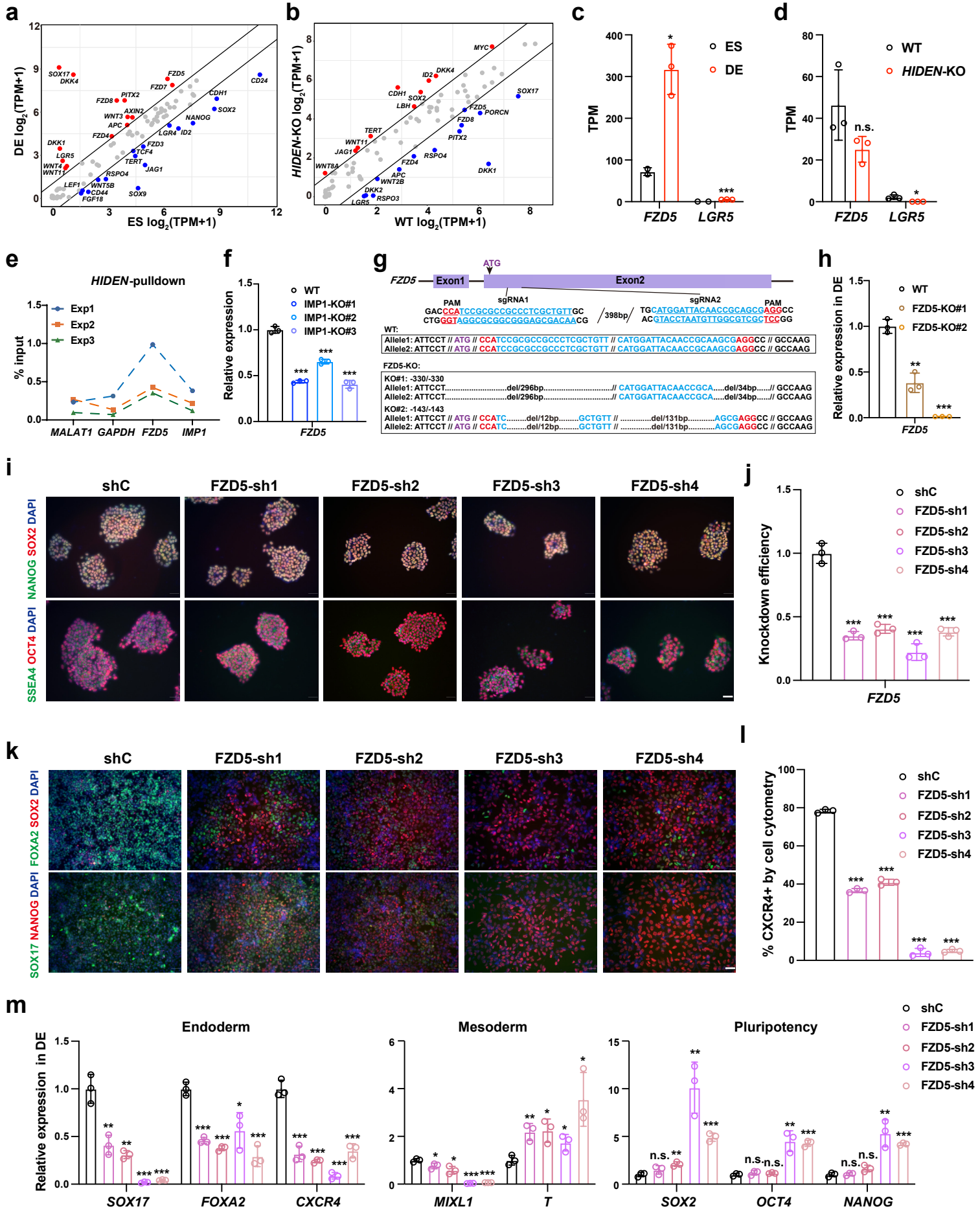


Figure S7. *HIDEN* promotes WNT pathway through *FZD5*.

(a) The plot of WNT-associated gene expression during DE differentiation from human ESCs.

(b) The plot of WNT-associated gene expression upon *HIDEN*-KO in DE cells.

(c-d) The TPM value of *FZD5*, *LGR5* during DE differentiation (c), and in *HIDEN*-KO DE cells compared to wildtype (d) (n=3).

(e) The enrichment of mRNAs or lncRNAs pulled down by *HIDEN* in HUES8 DE cells (normalized by input). Three independent experiments were shown.

(f) The relative expression of *FZD5* in wildtype or *IMP1* knockout DE cells, as shown by RT-qPCR (n=3).

(g) The genotype of *FZD5*-knockout ESC lines. The sgRNA targeting sequences, PAM sequence and deleted nucleotides were indicated.

(h) The knockout efficiency of *FZD5* in wildtype or *FZD5*-knockout DE cells, as shown by RT-qPCR (n=3).

(i) Immunofluorescent staining of pluripotency gene markers (*OCT4*, *SSEA4*, *NANOG*, *SOX2*) in shRNA control or *FZD5* knockdown ESCs. Scale bar=50 μ m.

(j) The determination of knockdown efficiency of *FZD5* (n=3).

(k) Immunofluorescent staining of DE markers (*SOX17*, *FOXA2*) and pluripotency markers (*NANOG*, *SOX2*) in shC and *FZD5*-KD DE cells. Scale bar=50 μ m.

(l) Flow cytometry analysis of *CXCR4*⁺ cells in shC or *FZD5*-KD DE cells (n=3).

(m) RT-qPCR analysis of RNA levels of representative endoderm genes, mesoderm genes and pluripotency genes in wildtype or *FZD5* knockdown DE cells (n=3).