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Supplemental Information

Genome editing with natural and engineered CjCas9 orthologs

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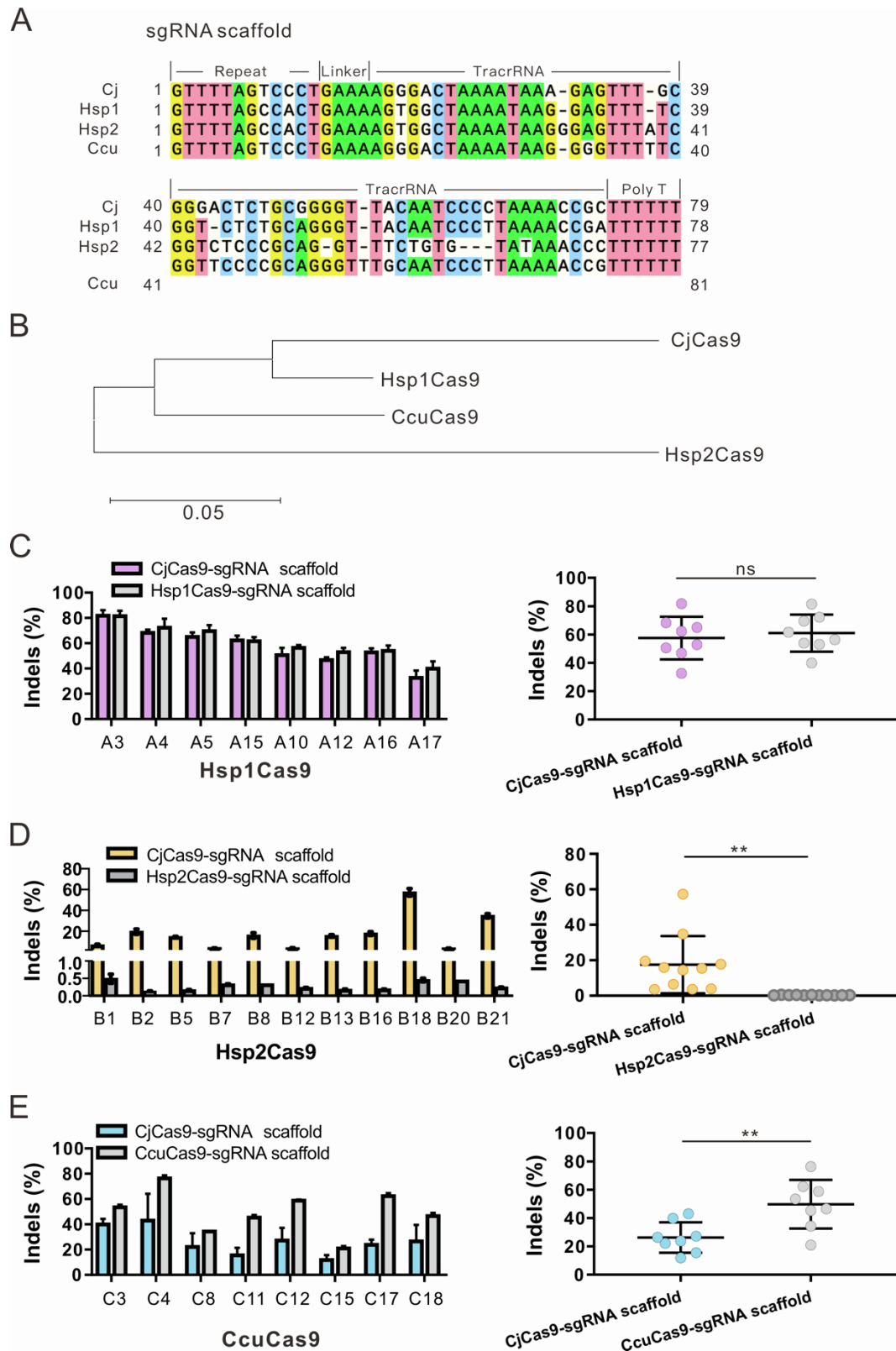


Figure S1. CjCas9 orthologs enable genome editing guided by their sgRNA scaffolds

(A) Comparison of the sgRNA scaffold sequences for CjCas9 orthologs. (B) The phylogenetic tree of the four sgRNA scaffolds. (C) The editing efficiencies of

Hsp1Cas9 with the CjCas9-sgRNA scaffold or Hsp1Cas9-sgRNA scaffold in HEK293T cells. (D) The editing efficiencies of Hsp2Cas9 with the CjCas9-sgRNA scaffold or Hsp2Cas9-sgRNA scaffold in HEK293T cells. (E) The editing efficiencies of CcuCas9 with the CjCas9-sgRNA scaffold or CcuCas9-sgRNA scaffold in HEK293T cells. Cells were treated with puromycin. The data represent the mean \pm SD; n=3. Student's t-test, ** $p < 0.01$. Indel efficiencies were determined by targeted deep sequencing.

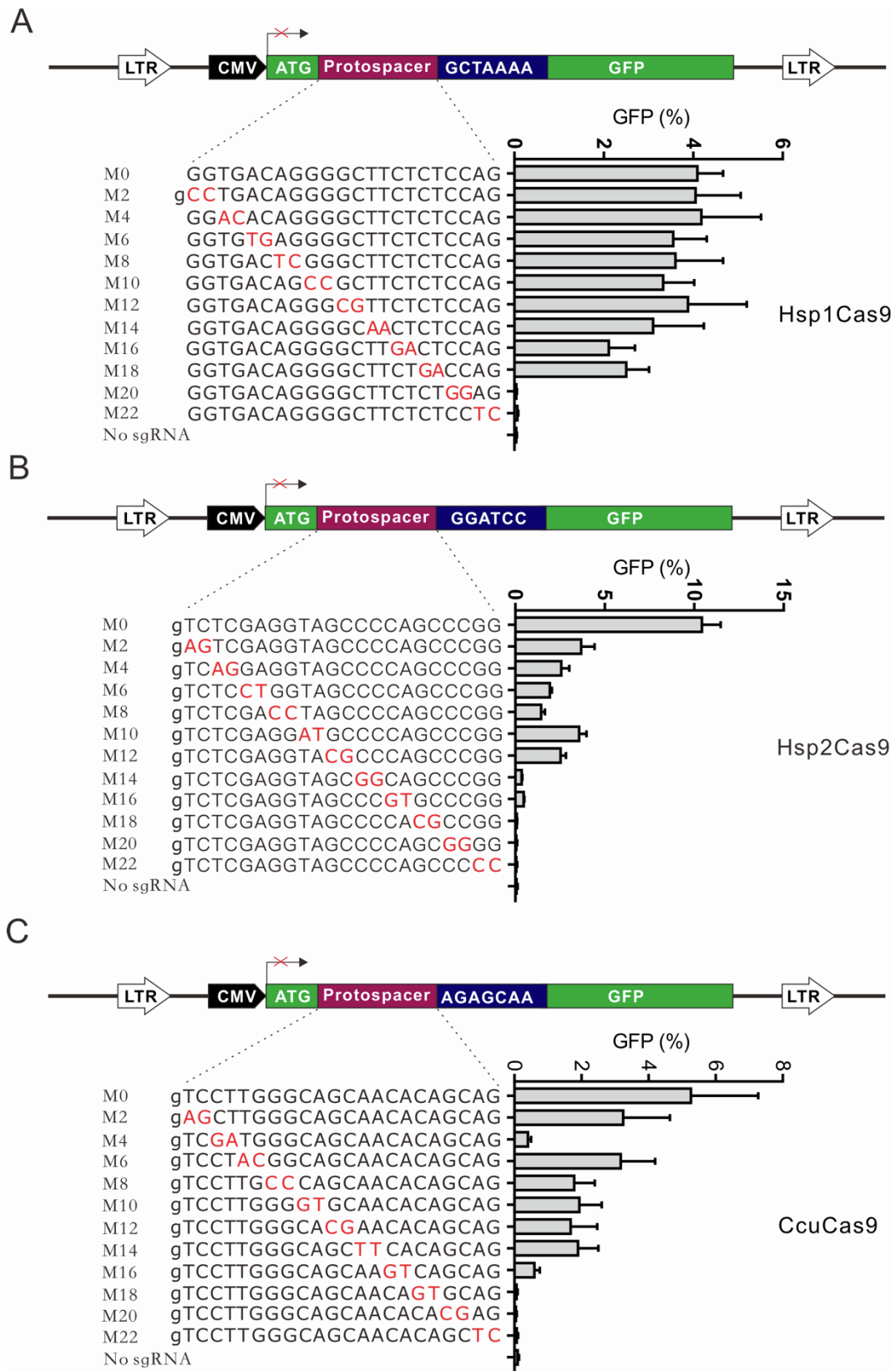


Figure S2. Analysis of specificity for (A) Hsp1Cas9, (B) Hsp2Cas9, and (C) CcuCas9 by the GFP-activation assay. Schematics of the GFP-activation reporters are shown on the top. A target sequence with the corresponding PAM is

inserted between ATG and GFP coding sequence, disrupting GFP expression. Cas9s were transfected with sgRNAs with CjCas9-sgRNA scaffold. Genome editing induced GFP expression. A panel of sgRNAs with dinucleotide mutations (red bases) is shown below. An additional G at the 5' terminal is added for U6 promoter transcription. The sgRNA activities were measured by the proportion of GFP-positive cells. The data represent the mean \pm SD; n=3.

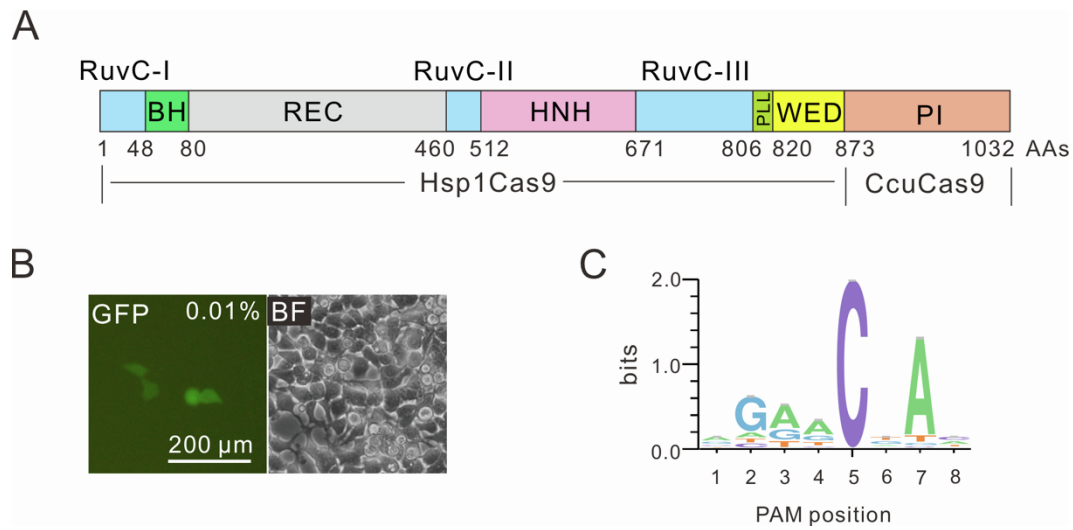


Figure S3. Generation of chimeric Hsp1-CcuCas9

(A) Schematic diagram of chimeric Hsp1-CcuCas9 nuclease. The Hsp1Cas9 PI domain was replaced with the CcuCas9 PI domain. (B) GFP-activation assay revealed that Hsp1-CcuCas9 induced GFP expression. The proportion of GFP-positive cells is shown. BF, bright field; GFP, green fluorescent protein. (C) WebLogo for Hsp1-CcuCas9 is generated based on deep sequencing data.

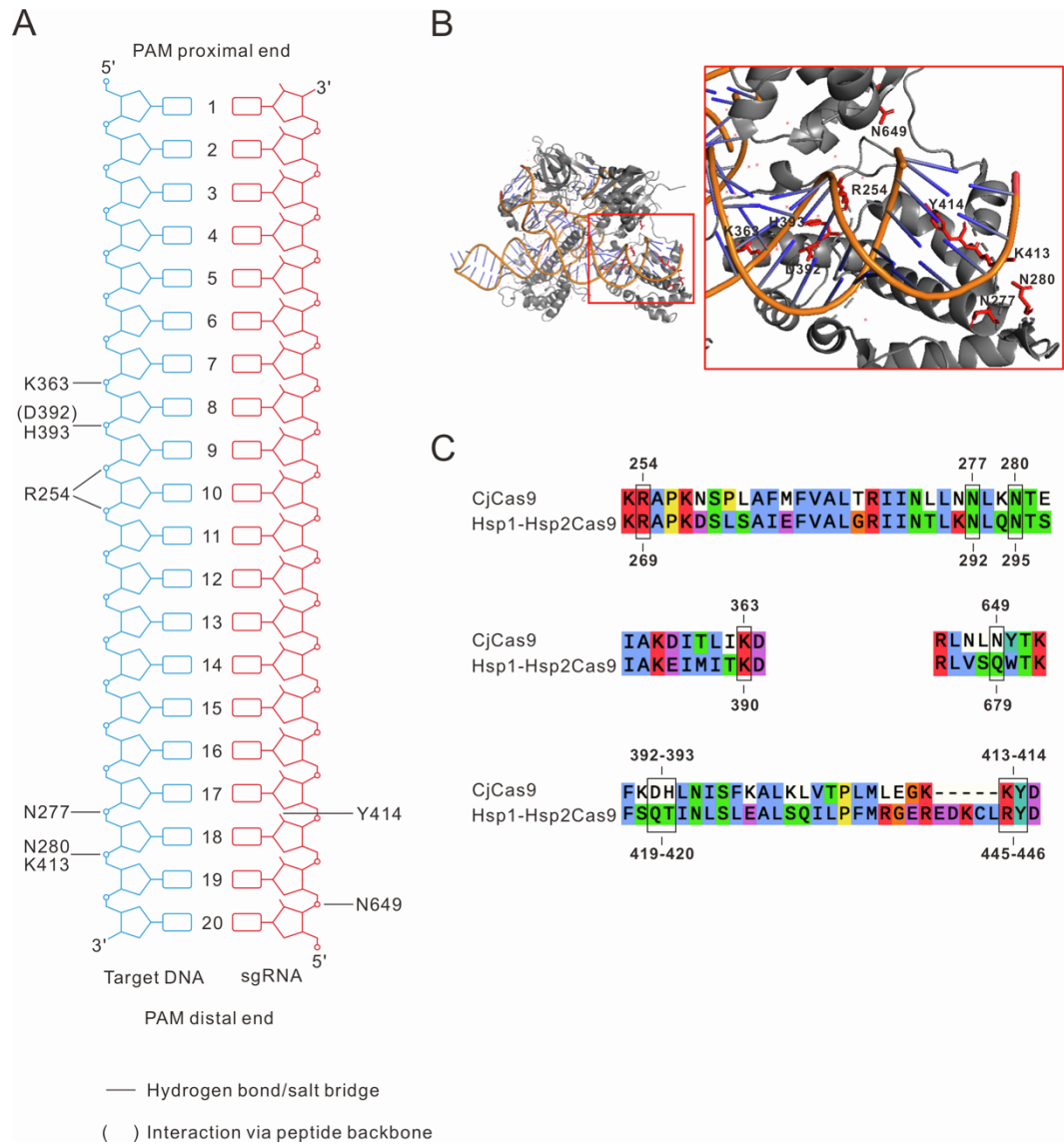


Figure S4. Engineering of Hsp1-Hsp2Cas9 variants based on CjCas9 crystal structure

(A) Schematic depicting interactions of CjCas9 residues with the target DNA-sgRNA duplex, based on PDB accession 5X2H and 5X2G¹. (B) The crystal structure of the CjCas9 interacting with the target DNA-sgRNA duplex (SMTL ID: 5x2g.1.A). The nine residues that form hydrogen bonds at the target DNA-sgRNA interface are shown. (C) Alignment of CjCas9 and Hsp1-Hsp2Cas9 protein sequences. Black boxes indicate equivalent residues of CjCas9 and Hsp1-Hsp2Cas9 interacting with the target DNA-sgRNA duplex. The positions of the residues are shown.

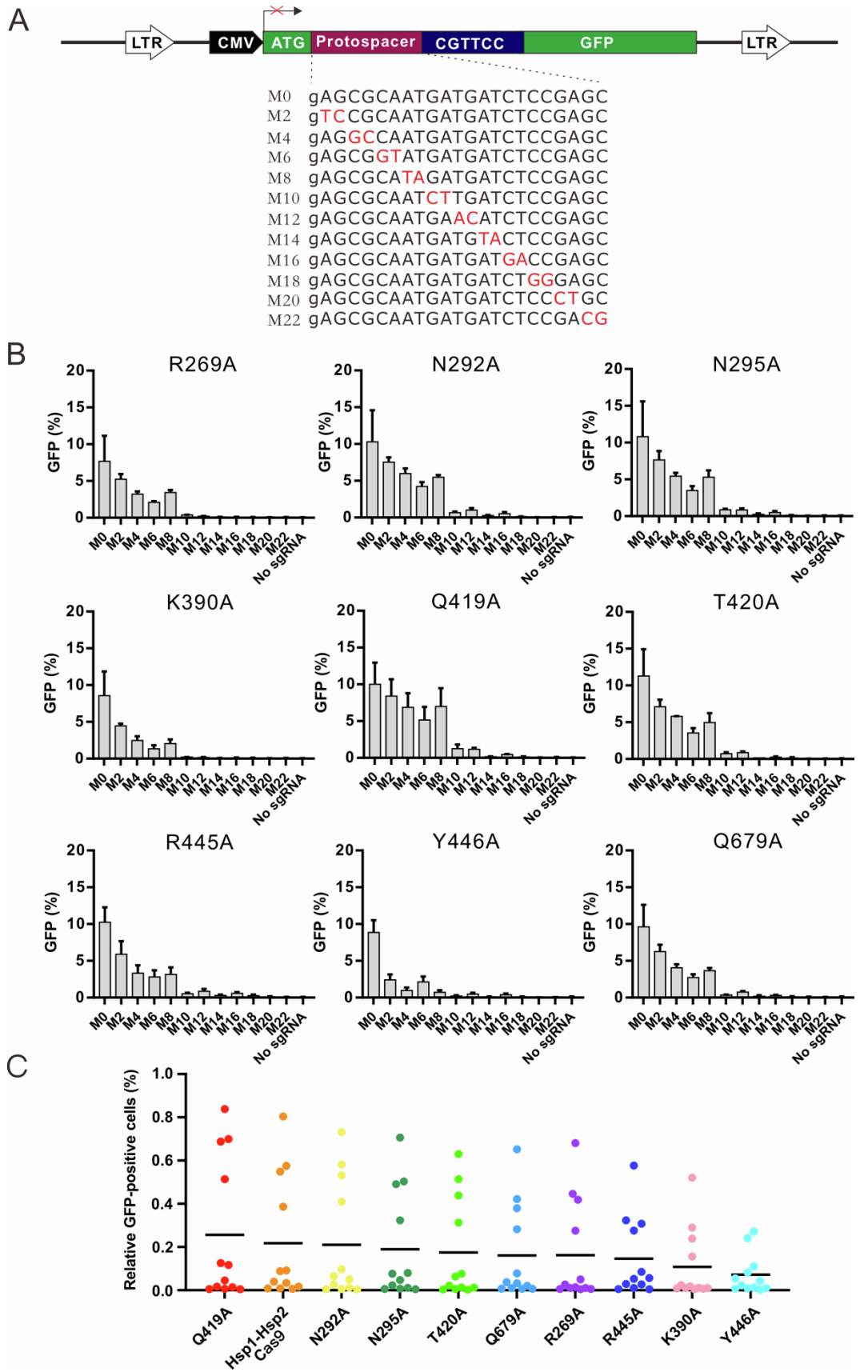


Figure S5. Specificity analysis of Hsp1-Hsp2Cas9 variants

(A) Schematic of the GFP-activation assay showing the specificity of the nucleases

in HEK293T cells on the top. Cas9s were transfected with sgRNAs with CjCas9-sgRNA scaffold. A panel of sgRNAs with dinucleotide mutations (red bases) is shown below. An additional G is added at the 5' end of sgRNA for U6 promoter transcription. (B) Specificity of single-mutation Hsp1-Hsp2Cas9 variants analyzed by the GFP-activation assay. The activity of each sgRNA of the Hsp1-Hsp2Cas9 variants was measured by the proportion of GFP-positive cells. The data represent the mean \pm SD; n=3. (C) Quantification of off-target editing efficiency based on GFP-activation assay. The off-target editing efficiency is normalized by on-target editing efficiency.

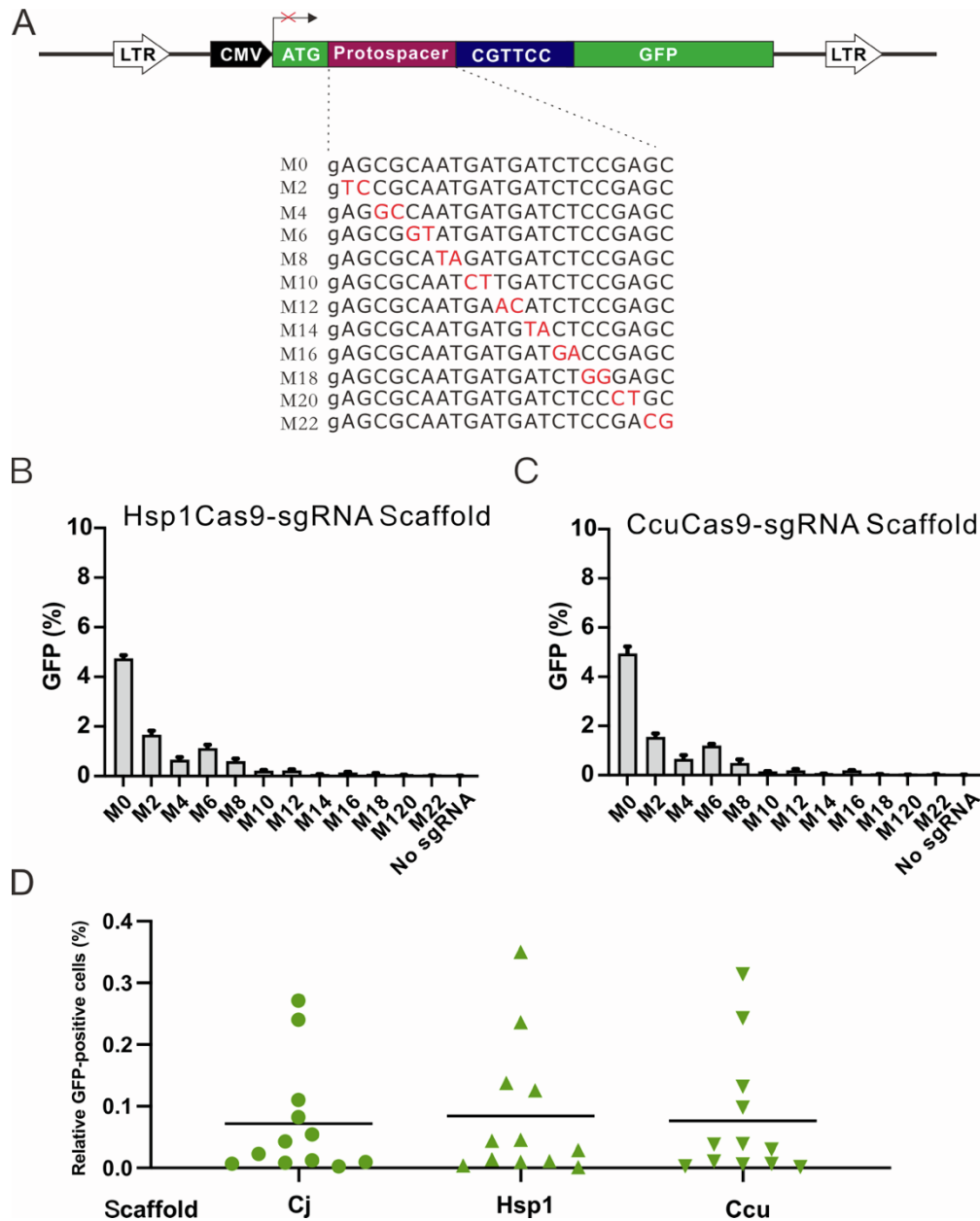


Figure S6. Specificity analysis of Hsp1-Hsp2Cas9-Y with different scaffolds

(A) Schematic of the GFP-activation assay showing the specificity of the nucleases in HEK293T cells on the top. A panel of sgRNAs with dinucleotide mutations (red bases) is shown below. An additional G is added at the 5' end of sgRNA for U6 promoter transcription. (B) Specificity of Hsp1-Hsp2Cas9-Y analyzed by the GFP-activation assay with sgRNAs with Hsp1Cas9-sgRNA scaffold. The data represent the mean \pm SD; n=3. (C) Specificity of Hsp1-Hsp2Cas9-Y analyzed by the GFP-activation assay with sgRNAs with CcuCas9-sgRNA scaffold. The data represent the mean \pm SD; n=3. (D) Quantification of off-target editing efficiency based on GFP-activation assay. The off-target editing efficiency is normalized by on-target editing efficiency.

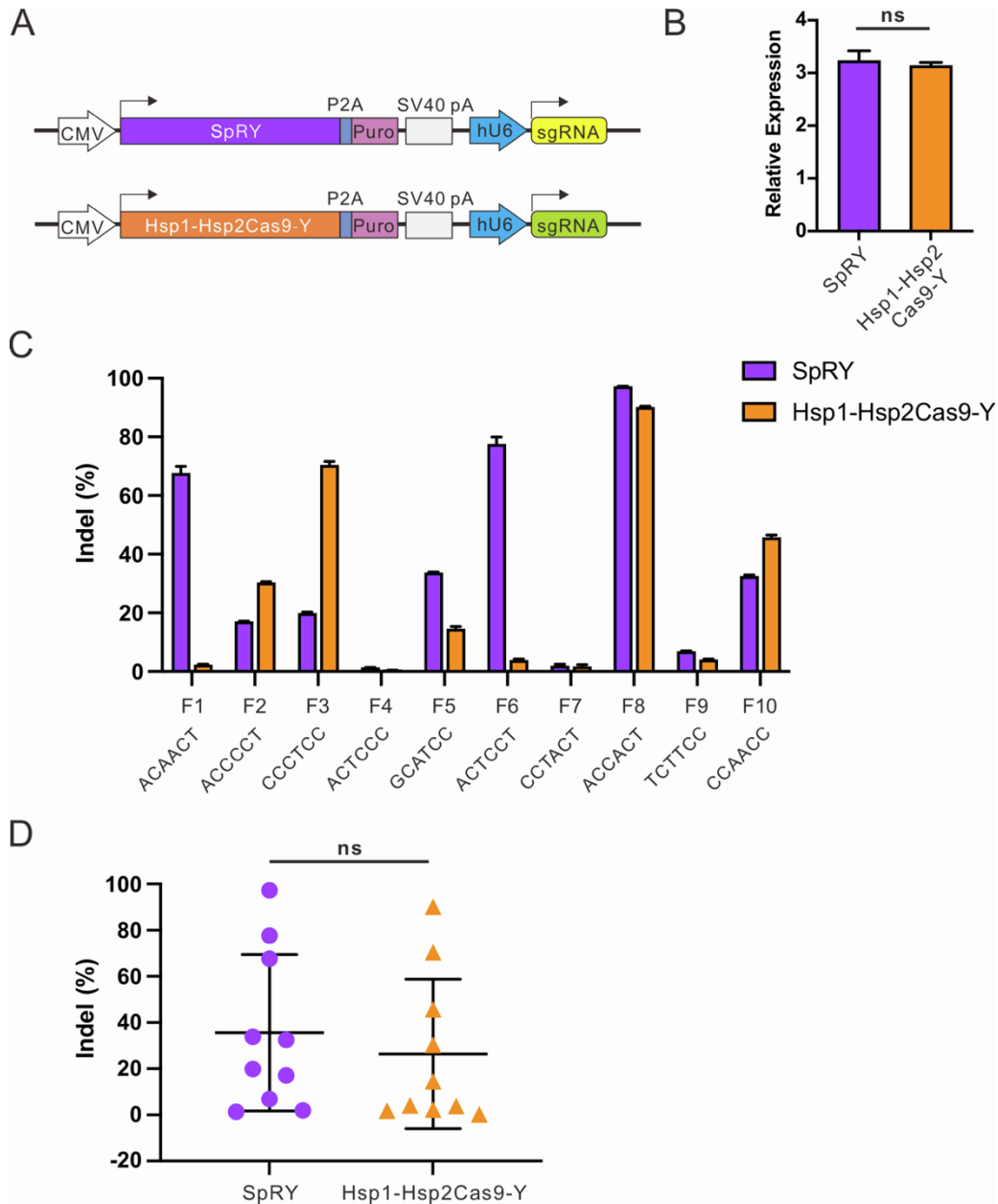


Figure S8. Comparison of genome editing activity between SpRY and Hsp1-Hsp2Cas9-Y

(A) Schematic of the Cas9 nuclease and sgRNA-expressing plasmid constructs. pA, polyA; Puro, puromycin resistant gene; hU6, human U6 promoter. SpRY was transfected with sgRNAs with the SpCas9-sgRNA scaffold. Hsp1-Hsp2Cas9-Y was transfected with sgRNAs with the CjCas9-sgRNA scaffold. (B) Expression levels of SpRY and Hsp1-Hsp2Cas9-Y relative to *GAPDH* were measured by RT-qPCR. (C) Comparison of SpRY and Hsp1-Hsp2Cas9-Y genome editing activity at ten endogenous sites with the N₄CY PAM in HEK293T cells. Cells were treated with puromycin. Indel efficiencies were determined by targeted deep sequencing. The data represent the mean \pm SD; n=3. (D) Quantification of the editing efficiencies of SpRY and Hsp1-Hsp2Cas9-Y. The data represent the mean \pm SD.

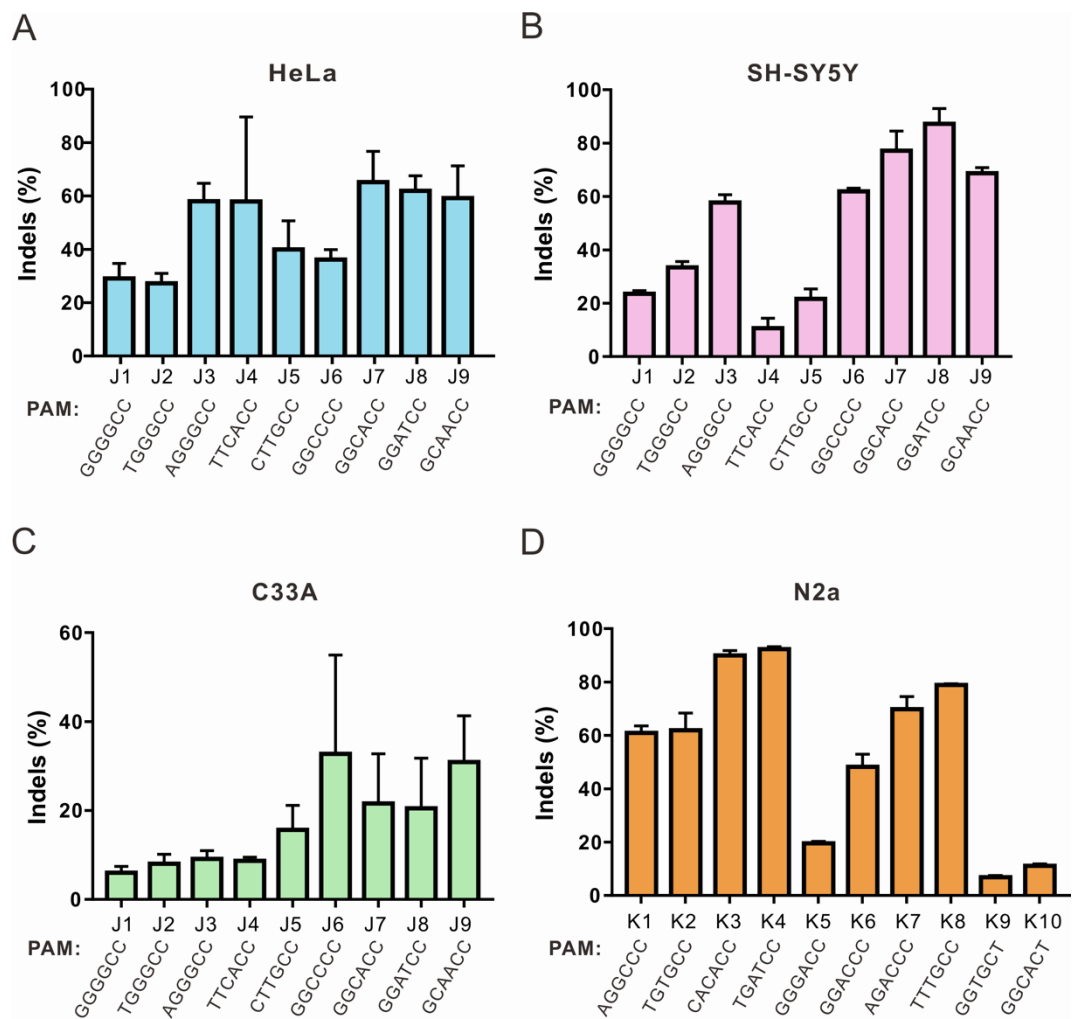


Figure S9. Hsp1-Hsp2Cas9-Y enables genome editing in diverse cell types
Hsp1-Hsp2Cas9-Y enables genome editing in diverse cell types, including (A) HeLa, (B) SH-SY5Y, (C) C33A, and (D) N2a cells. The PAM sequences are shown below. Hsp1-Hsp2Cas9-Y was transfected with sgRNAs with the CjCas9-sgRNA scaffold. Cells were treated with puromycin. Indel efficiencies were determined by targeted deep sequencing. The data represent the mean \pm SD; n=3.

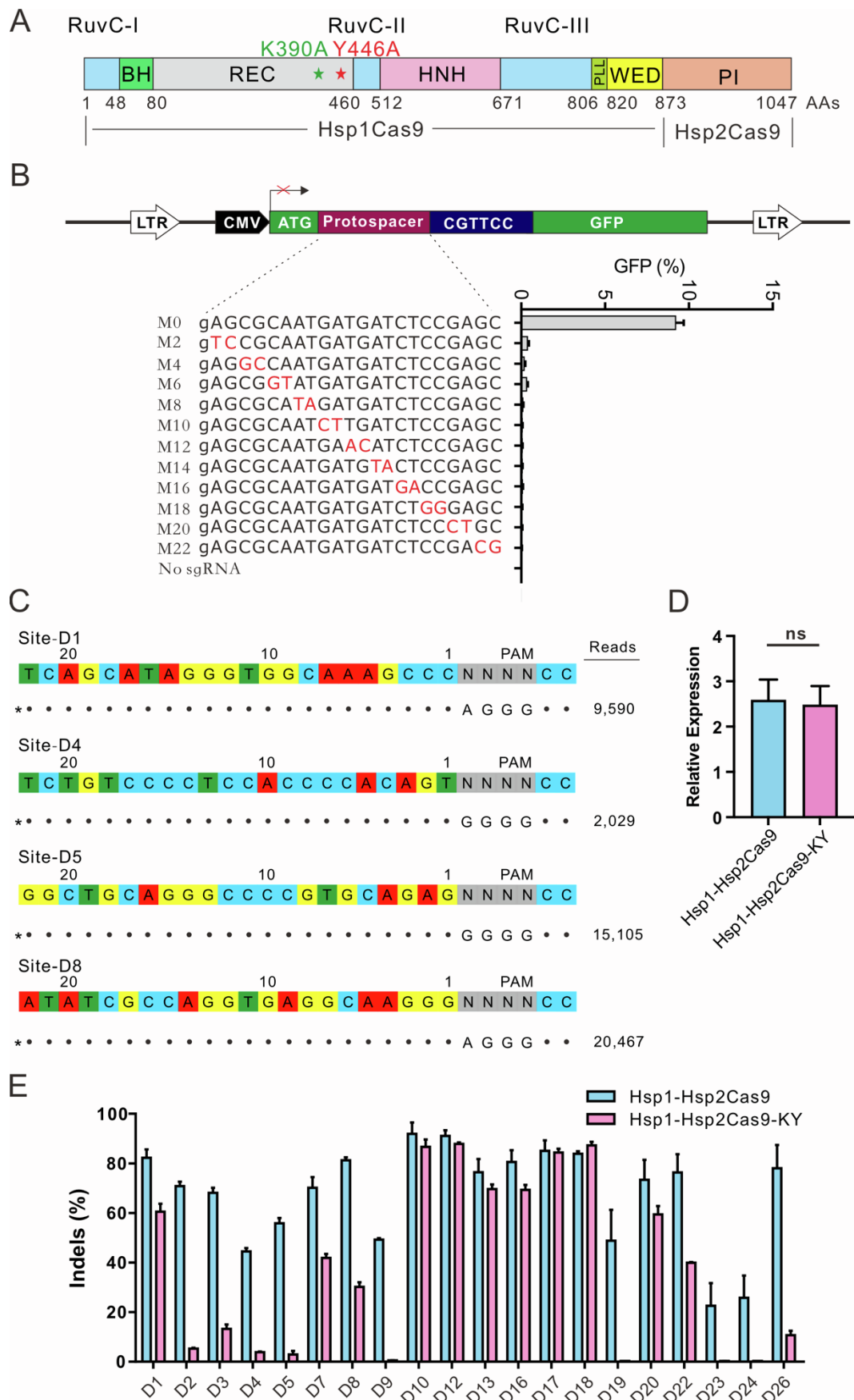


Figure S10. Characterization of Hsp1-Hsp2Cas9-KY for Genome editing

(A) Schematic diagram of chimeric Hsp1-Hsp2Cas9-KY nuclease. It contains K390A and Y446A mutations. (B) The specificity of Hsp1-Hsp2Cas9-KY is evaluated by the GFP-activation assay. Hsp1-Hsp2Cas9-KY was transfected with sgRNAs with the CjCas9-sgRNA scaffold. A panel of sgRNAs with dinucleotide mutations (red bases) is shown below. An additional G at the 5' terminal is added for U6 promoter transcription. The data represent the mean \pm SD; n=3. (C) Genome-wide specificities of Hsp1-Hsp2Cas9-KY determined by GUIDE-seq. Hsp1-Hsp2Cas9-KY was transfected with sgRNAs with the CjCas9-sgRNA scaffold. The on-target site is marked with "*". Read counts are listed on the right. (D) Expression levels of Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KY relative to *GAPDH* were measured by RT-qPCR. (E) Comparison of Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KY genome editing activity at 20 endogenous sites with the N₄CC PAM in HEK293T cells. Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KY were transfected with sgRNAs with the CjCas9-sgRNA scaffold. Cells were treated with puromycin. The data represent the mean \pm SD; n=3.

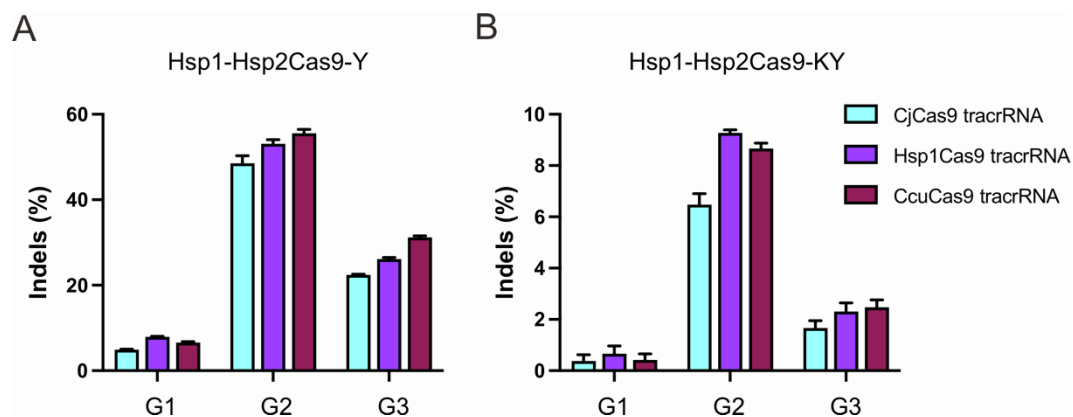


Figure S11. Test of genome editing activity of Hsp1-Hsp2Cas9-Y and Hsp1-Hsp2Cas9-KY with different sgRNA scaffolds

(A) Genome editing ability of Hsp1-Hsp2Cas9-Y with different scaffolds measured by targeted deep sequencing. Cells were treated with puromycin. The data represent the mean \pm SD; n=3. (B) Genome editing ability of Hsp1-Hsp2Cas9-KY with different scaffolds measured by targeted deep sequencing. The data represent the mean \pm SD; n=3.

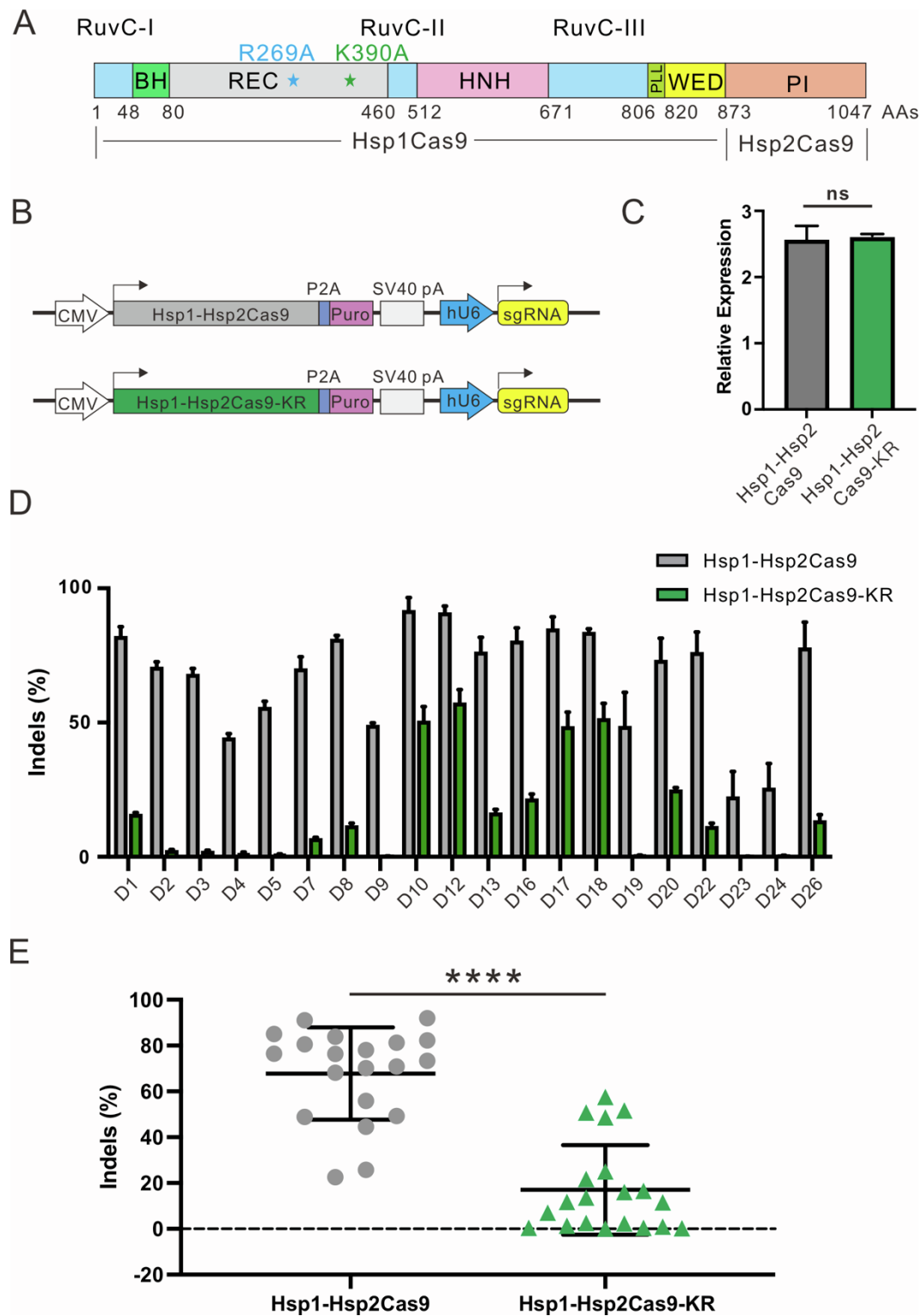


Figure S12. Comparison of genome editing activity between Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KR

(A) Schematic diagram of chimeric Hsp1-Hsp2Cas9-KR nuclease. It contains K390A and R269A mutations. (B) Schematic of plasmid constructs of the Cas9 nuclease and sgRNA with CjCas9-sgRNA scaffold. pA, polyA; Puro, puromycin resistant gene; hU6, human U6 promoter. Both Cas9 were transfected with sgRNAs

with the CjCas9-sgRNA scaffold. (C) Expression levels of Hsp1-Hsp2Cas9-Y and Hsp1-Hsp2Cas9-KY relative to *GAPDH* were measured by RT-qPCR. (D) Comparison of Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KR genome editing activity at 20 endogenous sites with the N₄CC PAM in HEK293T cells. Cells were treated with puromycin. The data represent the mean \pm SD; n=3. (E) Quantification of the editing efficiencies of Hsp1-Hsp2Cas9 and Hsp1-Hsp2Cas9-KY. Indel efficiencies were determined by targeted deep sequencing. The data represent the mean \pm SD.

REFERENCES

1. Yamada, M., Watanabe, Y., Gootenberg, J.S., Hirano, H., Ran, F.A., Nakane, T., Ishitani, R., Zhang, F., Nishimasu, H., and Nureki, O. (2017). Crystal Structure of the Minimal Cas9 from *Campylobacter jejuni* Reveals the Molecular Diversity in the CRISPR-Cas9 Systems. *Mol Cell* *65*, 1109-1121 e1103. [10.1016/j.molcel.2017.02.007](https://doi.org/10.1016/j.molcel.2017.02.007).

Table S1. Summary of the type II-A and type II-C Cas9 orthologs

Type	Cas9	PAM	Size (aa)	Activity	Specificity	Reference
Type II-A	SpCas9	NGG	1368	High	Low	Jinek et al. (2012)
Type II-A	SaCas9	NNGRRT	1053	High	Medium	Ran et al. (2015)
Type II-A	ScCas9	NNG	1380	Medium	Low	Chatterjee et al. (2018)
Type II-A	SlugCas9	NNGG	1054	High	Medium	Hu et al. (2021)
Type II-A	SauriCas9	NNGG	1061	High	Medium	Hu et al. (2020)
Type II-A	SchCas9	NNGR	1054	Medium	High	Wang et al. (2022)
Type II-A	St1Cas9	NNRGAA	1121	Medium	Medium	Agudelo et al. (2020)
Type II-C	NmeCas9	NNNNGATT	1082	Medium	High	Amrani et al. (2018)
Type II-C	Nme2Cas9	NNNNCC	1082	Medium	High	Edraki et al. (2019)
Type II-C	CjCas9	NNNNRYAC	984	Medium	High	Kim et al. (2017)
Type II-C	Nsp2Cas9	NNNNCC	1067	Medium	High	Wei et al. (2022)
Type II-C	BlatCas9	NNNNCNAA	1092	Medium	High	Gao et al. (2020)

The following tables are submitted in Excel format

Table S2. DNA sequences of plasmids were used in this study

Table S3. The human codon-optimized Cas9 sequences

Table S5. Primers used in this study

Table S6. All data in this study

Table S7. The information of raw sequencing data

Table S4. Target sites used in this study

	Name	Gene	Target Seq	PAM	Description
Figure3B	A1	VEGFA	ggtgacaaggggctctctccag	GCTAAAA	Endogenous target site of human
	A2	GRIN2B	gtgacaaggggaataaagcca	CCTGAAA	Endogenous target site of human
	A3	VEGFA	ccgattcaagtgggaaatggca	AGCAAAA	Endogenous target site of human
	A4	VEGFA	gggcaaggggttgtaactgag	GGTAAAA	Endogenous target site of human
	A5	VEGFA	tcagttcgaggaaggggaaggg	GGCAAAA	Endogenous target site of human
	A6	EMX1	gaggaacctaatacaattgtg	ACTAAAA	Endogenous target site of human
	A7	GRIN2B	cagtattcagtgtcgaagaagg	TCAAGAA	Endogenous target site of human
	A8	EMX1	aaagactgagagaacatgagg	ACAGAAA	Endogenous target site of human
	A9	EMX1	caagagaataggctctaagaag	AGTAAAA	Endogenous target site of human
	A10	GRIN2B	cagtcagctctgtgtgtgagg	GGGAAAA	Endogenous target site of human
	A11	GRIN2B	tttggaagaaaaggccaaga	TATGAAA	Endogenous target site of human
	A12	GRIN2B	aggatgggtaagatggcactg	AGGAAAA	Endogenous target site of human
	A13	VEGFA	actgagggtaaaagactggg	GTAGGAA	Endogenous target site of human
	A14	VEGFA	ggctgggagcctcgaatgcaag	GAGGAAA	Endogenous target site of human
	A15	VEGFA	tggggctgtctgggaactggg	CTGGGAA	Endogenous target site of human
	A16	GRIN2B	gaggtgacaaggggggaagga	GTGGGAA	Endogenous target site of human
	A17	GRIN2B	ggtgacagcagcaatgagaatg	TATAGAA	Endogenous target site of human
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	B4	GRIN2B	aaagggagtgatggaatgaga	GGATCC	Endogenous target site of human
	B5	AAVS1	aggaactgactgggtcagcag	GCTGCC	Endogenous target site of human
	B6	GRIN2B	aaagggagtgatggaatgaga	GATCCC	Endogenous target site of human
	B7	AAVS1	aaagcctgagcgcctctctggg	CTTGCC	Endogenous target site of human
	B8	AAVS1	tggtgcaagcggcgaagaaggag	TGCTCC	Endogenous target site of human
	B9	GRIN2B	aatatcaagccaactcaaatag	ACTACC	Endogenous target site of human
	B10	GRIN2B	gtagcttctctctctccaagg	TCTGCC	Endogenous target site of human
	B11	VEGFA	atcttaagtgtatgctctgtgg	ACTTCC	Endogenous target site of human
	B12	VEGFA	ccgggagcctccaactcctctg	GGCCCC	Endogenous target site of human
	B13	VEGFA	aggggggtgcccaggaaggaag	GGCACC	Endogenous target site of human
	B14	VEGFA	atgtctatcagcgcagctactg	CCATCC	Endogenous target site of human
	B15	VEGFA	gagagaaggggctctcagcag	GCATCC	Endogenous target site of human
	B16	VEGFA	tctcgaaggaagcccaagccgg	GGATCC	Endogenous target site of human
	B17	VEGFA	tggaaaggaagcaatgctctctg	GTCTCC	Endogenous target site of human
	B18	VEGFA	tggaggtagagcagcaaggaag	GGCTCC	Endogenous target site of human
	B19	VEGFA	ccgaagcctcgggaagcagcgg	CCGGCC	Endogenous target site of human
	B20	GRIN2B	tttaacatgagagaacaatactg	GCATCC	Endogenous target site of human
	B21	EMX1	ctccaagcctgggcgatacaggg	AGATCC	Endogenous target site of human
	B22	VEGFA	tagcagcagccctgtccactgg	CTTTCC	Endogenous target site of human
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	C4	AAVS1	agggaggaagatgcccggaga	GGACCCA	Endogenous target site of human
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	C6	VEGFA	tgggacactgaggaagcaag	AGAGCAA	Endogenous target site of human
	C7	VEGFA	gggagcaagcaagcagactcca	ACAACCA	Endogenous target site of human
	C8	AAVS1	tgaagaatggtgctgctactggtg	TTCACCA	Endogenous target site of human
	C9	VEGFA	ctgttccaagaatgttaacccc	CTCCCTA	Endogenous target site of human
	C10	VEGFA	agggcggcgtgtgcaagcaag	TGCTCCA	Endogenous target site of human
	C11	VEGFA	tctccaagcctcaactctgccc	AGTGCTA	Endogenous target site of human
	C12	VEGFA	tctcgaaggaagcccaagccgg	GGATCCA	Endogenous target site of human
	C13	VEGFA	gcaagcaaggaagcctccaatg	CACCCAA	Endogenous target site of human
	C14	VEGFA	aggggacggaagattcaatccc	CTTCCAA	Endogenous target site of human
	C15	VEGFA	caaacctggaagcaagtgcca	CCACCAA	Endogenous target site of human
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	C17	VEGFA	gtgagaagtgagaagagaagca	CGGGCCA	Endogenous target site of human
	C18	TIMM8B	atgtatgtccaaggtcatgtag	CCAGCAA	Endogenous target site of human
	C19	VEGFA	agaataagcccaagcctctctg	ACTGCTA	Endogenous target site of human
	C20	VEGFA	ctggaaagagcaagaaagaaag	GCAACAA	Endogenous target site of human
	C21	TIMM8B	gtgggaggaagaggggtgctcgg	ATGACCA	Endogenous target site of human
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C23	LINCO1588	ttggaagtgtttgtagagga	GGGACAA	Endogenous target site of human	
C24	VEGFA	ccaaggaagatgagaagccaagga	AGGACCA	Endogenous target site of human	
C25	VEGFA	ggagggggagaagggacagag	AGGGCAA	Endogenous target site of human	
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	D5	AAVS1	ggctgcaagggcccgtgcaag	GGGGCC	Endogenous target site of human
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	D7	AAVS1	cttctccaagcctgcaatgccc	TGGGCC	Endogenous target site of human
	D8	AAVS1	atatcgccaagtgaggcaaggg	AGGGCC	Endogenous target site of human
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	D12	AAVS1	tgagaatggtgctgctactggtg	TTCACC	Endogenous target site of human
	D13	AAVS1	aaagcctgagcgcctcctctggg	CTTGCC	Endogenous target site of human
	D14	AAVS1	gttccctttctctctctctct	GGGGCC	Endogenous target site of human
	D15	AAVS1	atgctgtcctgagatgaggacata	GGGGCC	Endogenous target site of human

	D16	VEGFA	ccgggacccctcca ctctcctg	GGCCCC	Endogenous target site of human	
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	E3	AAVS1	gtggcta a a gcca gggga ga cgg	GGTACT	Endogenous target site of human	
	E4	AAVS1	aga a ga cta gctga gctctcgg	ACCCCT	Endogenous target site of human	
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	E9	EMX1	tgcggtga ca ga gca a gtgctg	GGGGCT	Endogenous target site of human	
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	sg3	B4GALNT2	gga tgccta cga cccgcgtga c	CTCCCC	Target site of BeGALNT2 exon3	
Figure6D	sg4	B4GALNT2	a cccgcgtga cctcccgcga gt	GAACCT	Target site of BeGALNT2 exon3	
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	sg3	CMAH	ta a cccgtggga tta ca ga a ccc	AGATCT	Target site of CMAH exon3	
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	H3	EMX1	ggga cta ca ggcata cca cc	ACACCT	Endogenous target site of human	
	H4	EMX1	tgtcta gggggcctgta gga a c	CCCTCC	Endogenous target site of human	
	H5	EMX1	a gcca ttttcta a ta tga tgg	GCATCC	Endogenous target site of human	
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	H8	EMX1	tgtcta gcctca tgtgtctgc	TCACCT	Endogenous target site of human	
	H9	EMX1	tggcctggggcgta gga ggc	CAAACC	Endogenous target site of human	
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	H12	AAVS1	cca ctga gca cta a a ggcctgg	CCGGCC	Endogenous target site of human	
	H13	EMX1	tca gtgtcca a ta a a gttca a	ACTCCT	Endogenous target site of human	
	H14	EMX1	ata a ga tctctgtttcccttc	ACTCCC	Endogenous target site of human	
	Figure S9A-C	F1	EMX1	ga a ca ca tga ggcata ca ggt	ACAACCT	Endogenous target site of human
		F2	EMX1	agtgtcta gggggcctgta gga	ACCCTT	Endogenous target site of human
		F3	EMX1	tgtcta gggggcctgta gga a c	CCCTCC	Endogenous target site of human
		F4	EMX1	ata a ga tctctgtttcccttc	ACTCCC	Endogenous target site of human
		F5	EMX1	a gcca ttttcta a ta tga tgg	GCATCC	Endogenous target site of human
F6		EMX1	ca cca ca ta tta tca a a a g	ACTCCT	Endogenous target site of human	
F7		EMX1	a gca gta a ta tta tca a a a g	CCTACT	Endogenous target site of human	
F8		EMX1	ca a a a ta a ttggcca gggctc	ACCACT	Endogenous target site of human	
F9		EMX1	toga cctcctgggctoga ga ga	TCTTCC	Endogenous target site of human	
F10		EMX1	tggcctggggcgta gga ggc	CAAACC	Endogenous target site of human	
Figure S9D	J1	AAVS1	gtgaa ctgga gttgta ca gacct	GGGGCC	Endogenous target site of human	
	J2	AAVS1	cttcctcca cctgca ta gcc	TGGGCC	Endogenous target site of human	
	J3	AAVS1	ata tgcga ggtga gga a ggg	AGGGCC	Endogenous target site of human	
	J4	AAVS1	tga ga a tgggtgctccta ggtg	TTCACC	Endogenous target site of human	
	J5	AAVS1	a a gctcga gccctctcctggg	CTTGCC	Endogenous target site of human	
	J6	VEGFA	ccgggacccctcca ctctcctg	GGCCCC	Endogenous target site of human	
	J7	VEGFA	aggggggtgccga gga ccga a g	GGCACC	Endogenous target site of human	
	J8	VEGFA	tctcga ggta gcccc gcccgg	GGATCCA	Endogenous target site of human	
	J9	VEGFA	ctgtccctcctga gccca tgg	GCAACC	Endogenous target site of human	
	K1	mEMX1	gggtgggaa ggtga gcta a gca g	AGGGCC	Endogenous target site of Mouse	
Figure S9D	K2	mEMX1	ta gga tggttctgcccgggga	TGTGCC	Endogenous target site of Mouse	
	K3	mEMX1	ccggctctga cgggtga cccgg	CACACC	Endogenous target site of Mouse	
	K4	mTh	gtgtcttggga ga ga gcccca	TGATCC	Endogenous target site of Mouse	
	K5	mTh	ga gtagga cttga gga a gccaca	GGGACC	Endogenous target site of Mouse	
	K6	mTh	a ctagga cgttcta ga a ccca	GGACCC	Endogenous target site of Mouse	
	K7	mRNF2	aggcca gcttga a cta cata g	AGACCC	Endogenous target site of Mouse	
	K8	mRNF2	aa gaa gccca a gga tga a gctg	TTTGCC	Endogenous target site of Mouse	
	K9	mEMX1	gatccggga ccttga a ga ca g	GGTGCT	Endogenous target site of Mouse	
	K10	mTh	atgca gcta a ga a gta tga a gg	GGCACT	Endogenous target site of Mouse	
	Figure S9D	G1	AAVS1	cca cca a cggcca cggta tca g	CGCCCT	Endogenous target site of human
G2		AAVS1	cccctcttga ggcctgca tc	ATCACC	Endogenous target site of human	
G3		AAVS1	cca ctga gca cta a a ggcctgg	CCGGCC	Endogenous target site of human	