Supporting Information

Sphingosine Kinase 2 Inhibitors: Rigid Aliphatic Tail Derivatives Deliver Potent and Selective Analogs

Srinath Pashikanti^{†,‡}, Daniel J. Foster[†], Yugesh Kharel[‡], Anne M. Brown^{§,J}, David R. Bevan,^{§,J} Kevin R. Lynch[‡], Webster L. Santos^{†,L*}

[†]Department of Chemistry, Virginia Tech, Blacksburg, VA 24060, USA ^Virginia Tech Center for Drug Discovery, Virginia Tech, Blacksburg, VA 24060, USA [‡]Department of Pharmacology, University of Virginia, Charlottesville, VA 22908, USA [§]Department of Biochemistry, Virginia Tech, Blacksburg, VA 24060, USA [‡]Department of Biochemistry, Virginia Tech, Blacksburg, VA 24060, USA



Figure S1. Molecular docking of top compounds in hSphK1 (PDB ID: 3VZB). Molecular docking of **23d** (blue, A) and **23e** (pink, B) in hSphK1 position the ligand towards the top of the binding cavity near ATP but are out of range for hydrogen bonding to the oxadiazole ring as observed in SphK2. Binding cavity steric hindrance and lack of inhibitor flexibility in alkyl tail analogs **26d** (cyan, C) and **26a** (blue, D) do not allow the utilization of the side pocket near Phe303. Compound **14c** (green, E) does not utilize the side pocket near Phe303 and is further distorted to position the guanidine group away from Asp178 due to steric hindrance of the trifluoromethyl group with Phe303. Black lines with measurements are shown for distance comparison to SphK2 results and are outside of the range for a hydrogen bond (A,B).

¹H NMR (400 MHz, CD₃OD) **14c**



¹³C NMR (101 MHz, CD₃OD) 14c



¹H NMR (400 MHz, CD₃OD) **14e**



¹³C NMR (126 MHz, CD₃OD) 14e



¹H NMR (500 MHz, CD₃OD) 14i



¹³C NMR (101 MHz, CD₃OD) 14i



¹H NMR (400 MHz, CD₃OD) **23d**



¹³C NMR (101 MHz, CD₃OD) **23d**



UPLC of 14c

An approximately 6 year old sample of compound **14c** (**SLP9101555**) (stored at -20 C in DMSO) was rerun on Waters Acquity H Class UPLC. Conditions: Solvent A: Water (0.1% TFA); solvent B: acetonitrile (0.1% TFA); column: Acquity BEH C18 1.7 μ m 2.1 x 50 mm; method: isocratic 60% A, 40% B from 0-3.50 min then linear gradient from 40-95% B by 5 minutes, return to 40% B by 6 minutes, then hold for 2 minutes at 60% A, 40% B; UV wavelength = 254 nm; flow rate: 0.613 mL/min. These studies indicate 91% purity.



Chiral HPLC analysis: Daicel IC column

An approximately 6 year old sample of compound **14c** (**SLP9101555**) (stored at -20 C in DMSO) was run using a super critical fluid chromatography - Waters SFC. The chiral column used was *Daicel IC*, 250 x 4.6 mm, 5 μ m. Method: flow rate of 3 ml/min; linear gradient from 80% CO₂, 20% (MeOH+0.5% Isopropylamine) to 60% CO₂, 40% (MeOH+0.5% Isopropylamine) from 0-5 min, then hold for 5 min.



Chiral HPLC analysis: Daicel IA column

An approximately 6 year old sample of compound **14c** (**SLP9101555**) (stored at -20 C in DMSO) was run using a super critical fluid chromatography - Waters SFC. The chiral column used was *Daicel IC*, 250 x 4.6 mm, 5 μ m. Method: flow rate of 3 ml/min; linear gradient from 80% CO₂, 20% (MeOH+0.5% Isopropylamine) to 60% CO₂, 40% (MeOH+0.5% Isopropylamine) from 0-5 min, then hold for 5 min.



Using the two different chiral columns suggest that compound 14c is a single enantiomer.