

HGGA, Volume 4

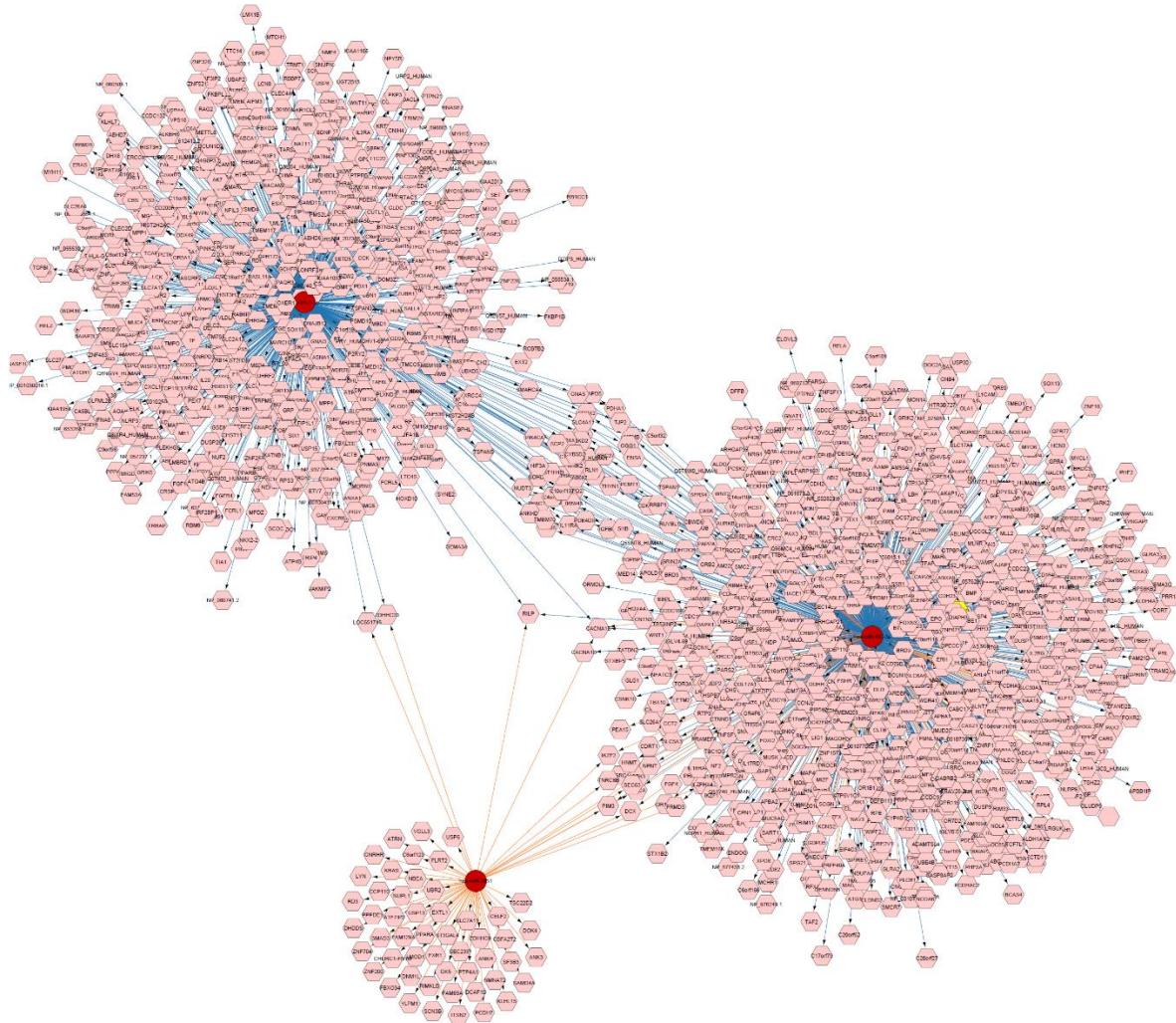
Supplemental information

**Artificial intelligence-driven pan-cancer analysis
reveals miRNA signatures for cancer stage prediction**

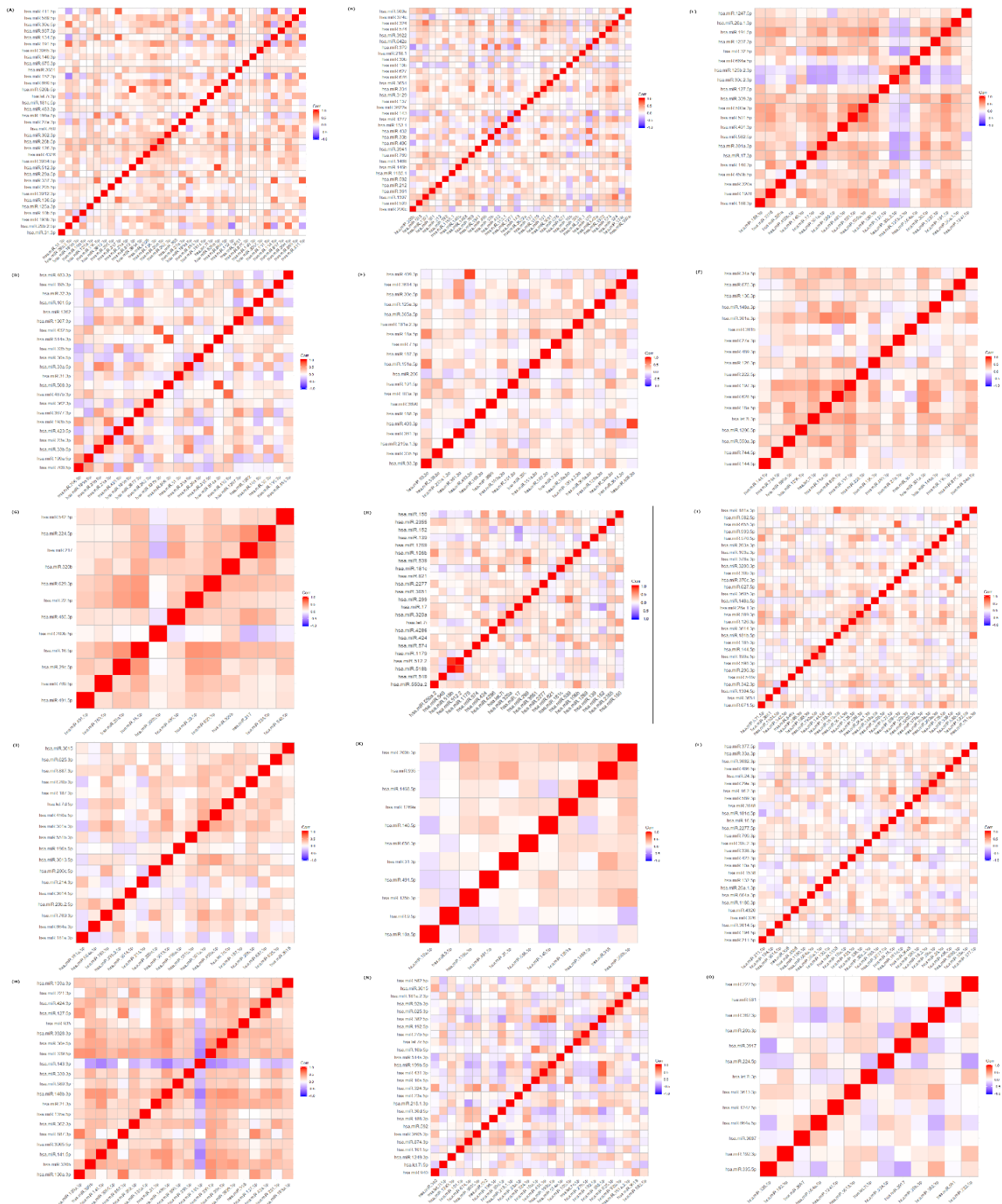
Srinivasulu Yerukala Sathipati, Ming-Ju Tsai, Sanjay K. Shukla, and Shinn-Ying Ho

Supplementary information

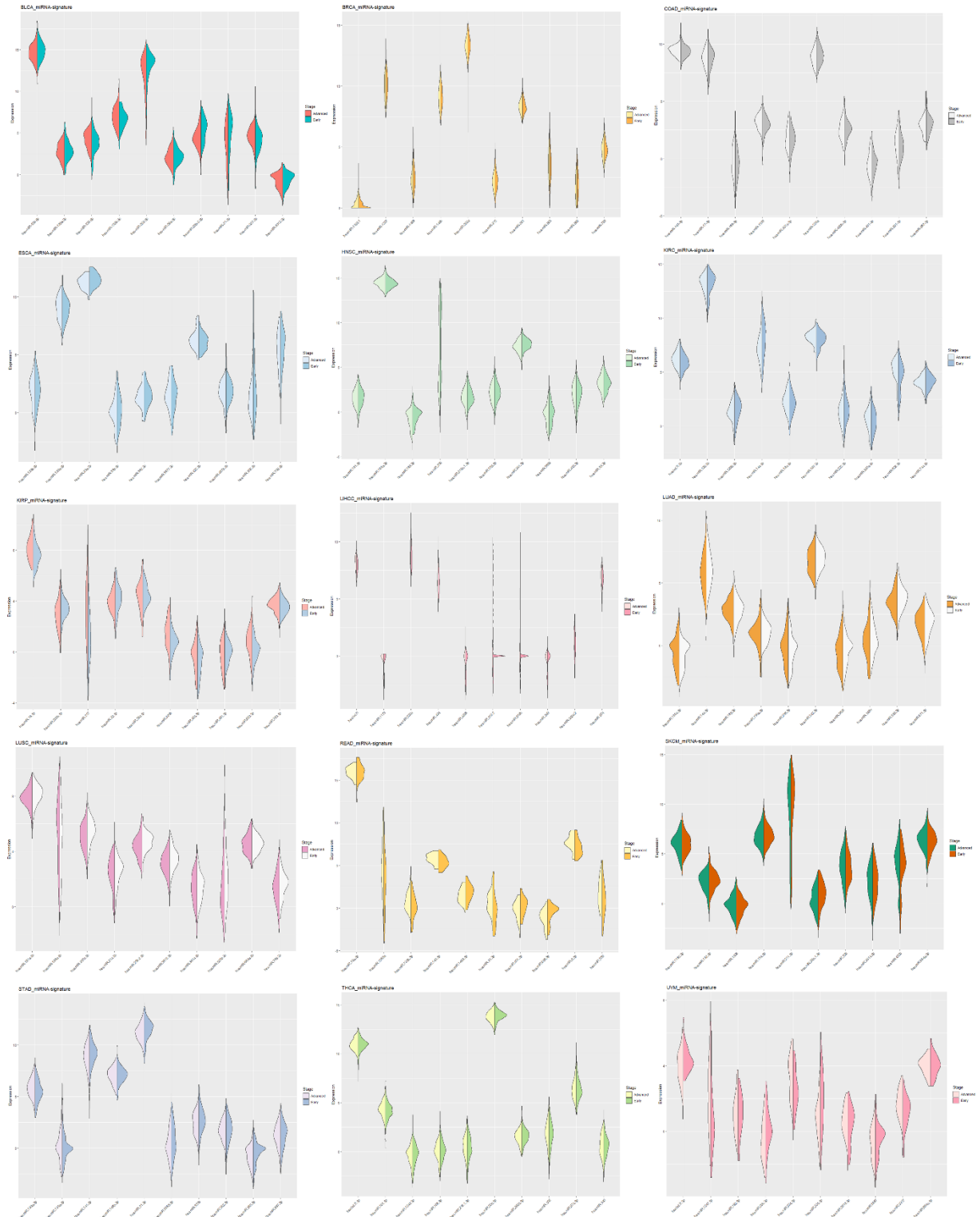
Supplementary Figures



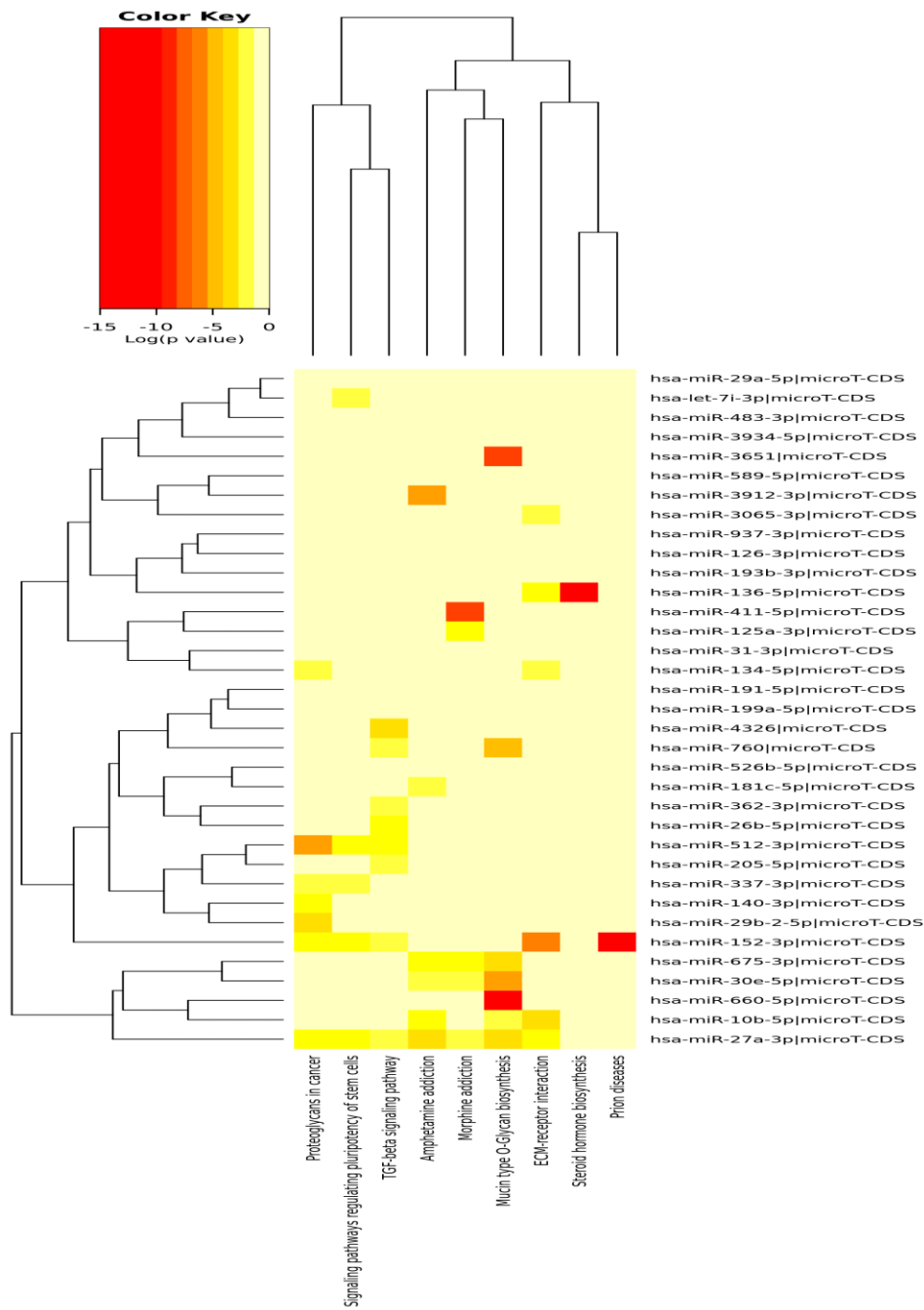
Supplementary Figure S1. Three miRNAs and corresponding target genes predicted using MicroCosm, miRTar base, and TargetScan. In this network microRNAs and target genes are defined as red circles and pink rounded hexagons respectively. The predicted microRNA-Target interactions are visualized in blue and in orange color.



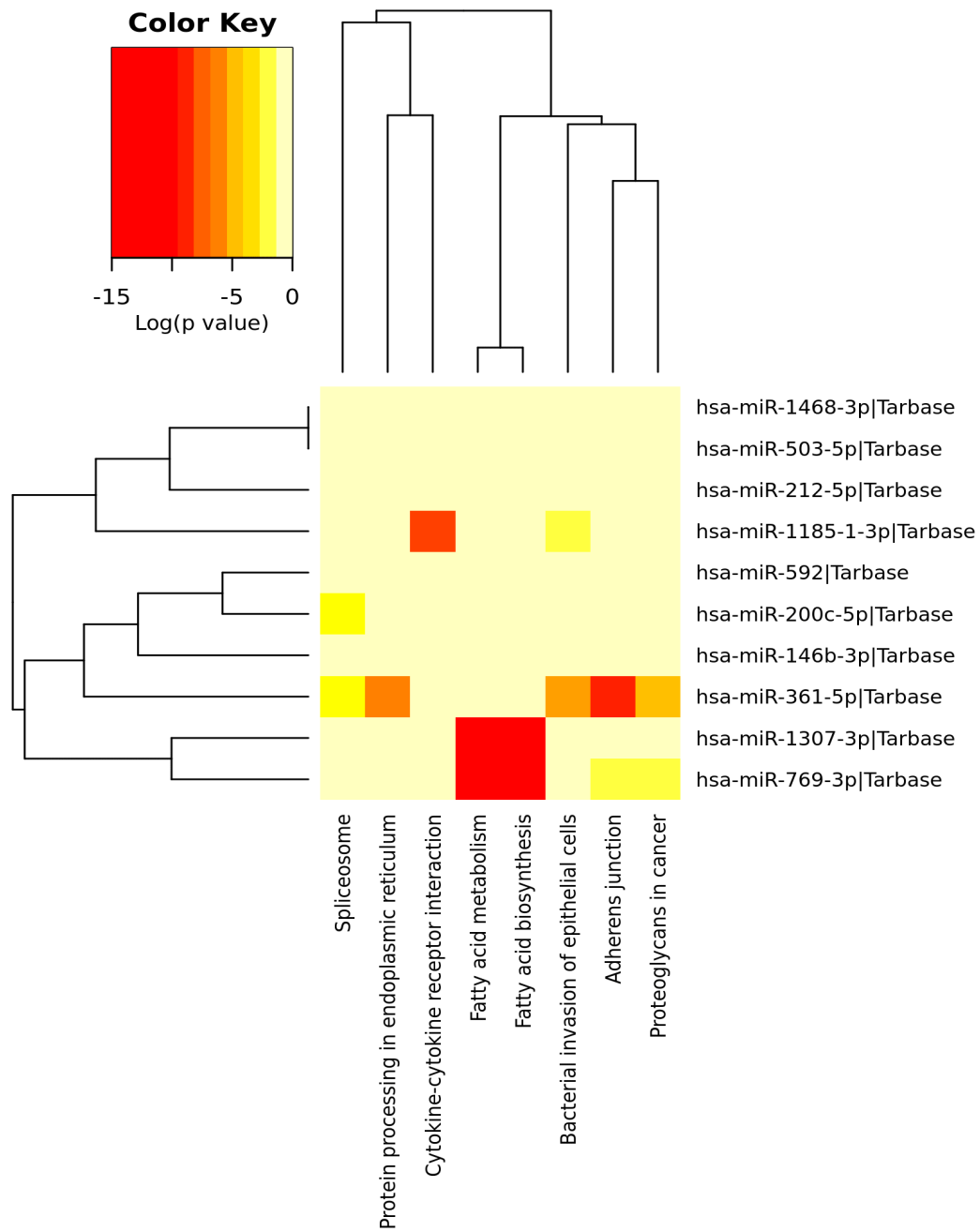
Supplementary Figure S2. Co-expression analysis of miRNA signatures across 15 cancers. (A) BLCA, (B) BRCA, (C) COAD, (D) ESCA, (E) HNSC, (F) KIRC, (G) KIRP, (H) LIHCC, (I) LUAD, (J) LUSC, (K) READ, (L) SKCM, (M) STAD, (N) THCA, and (O) UVM.



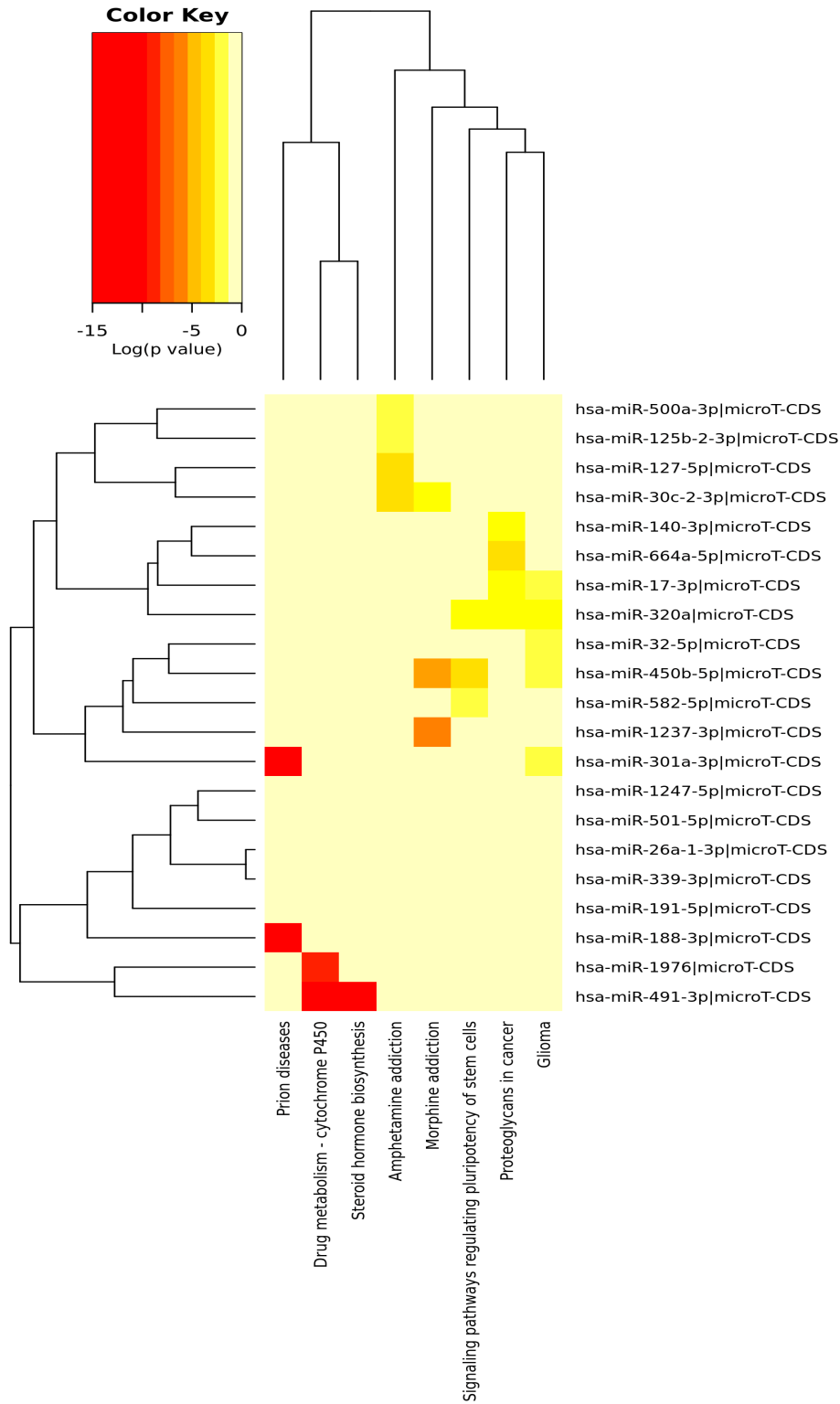
Supplementary Figure S3. Expression difference analysis of the miRNA signatures across 15 cancers. Relative expression differences of the miRNA signatures between early and advanced stages across cancers.



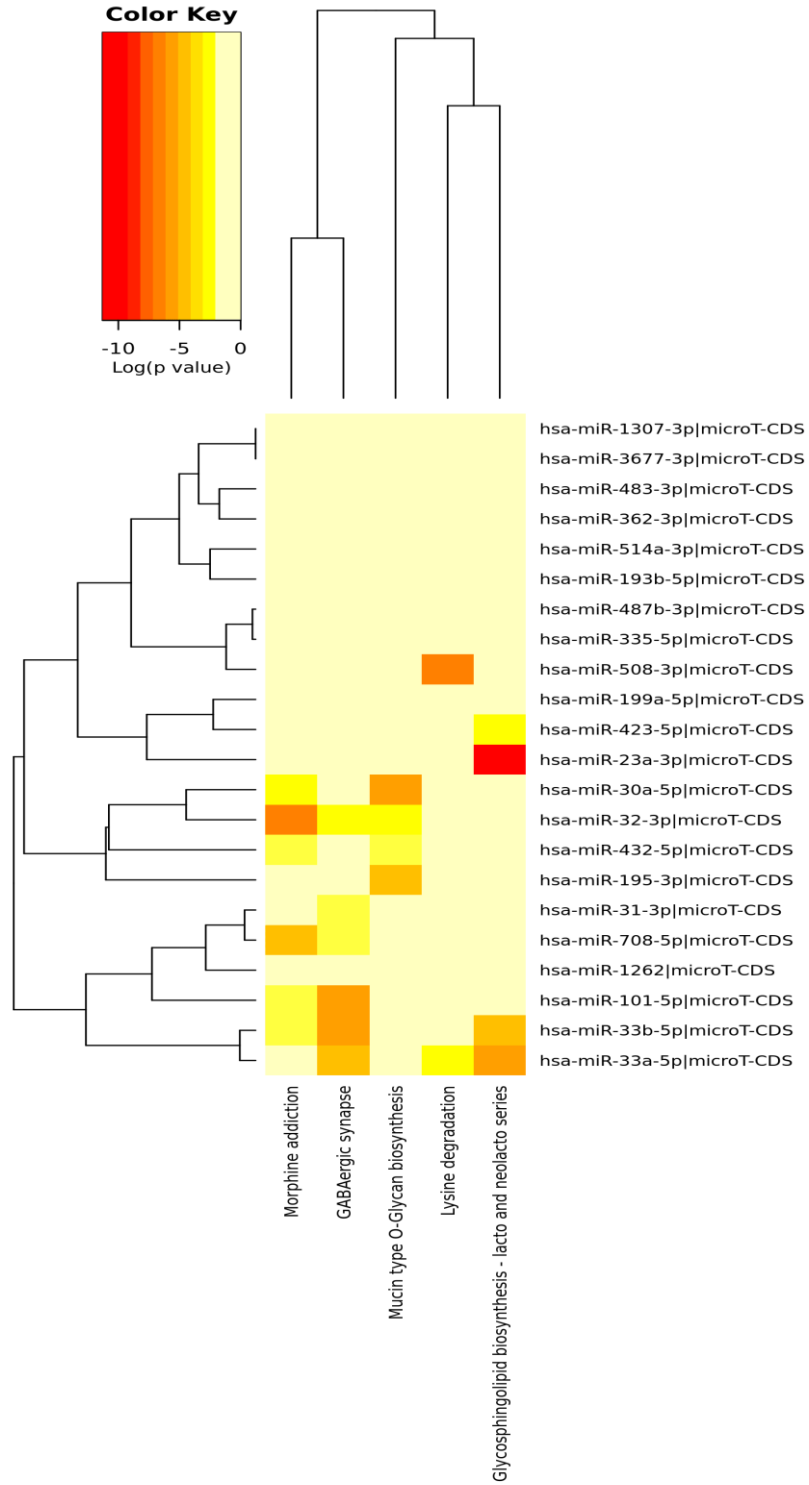
Supplementary Figure S4.1. KEGG pathway enrichment analysis of miRNA signatures in BLCA



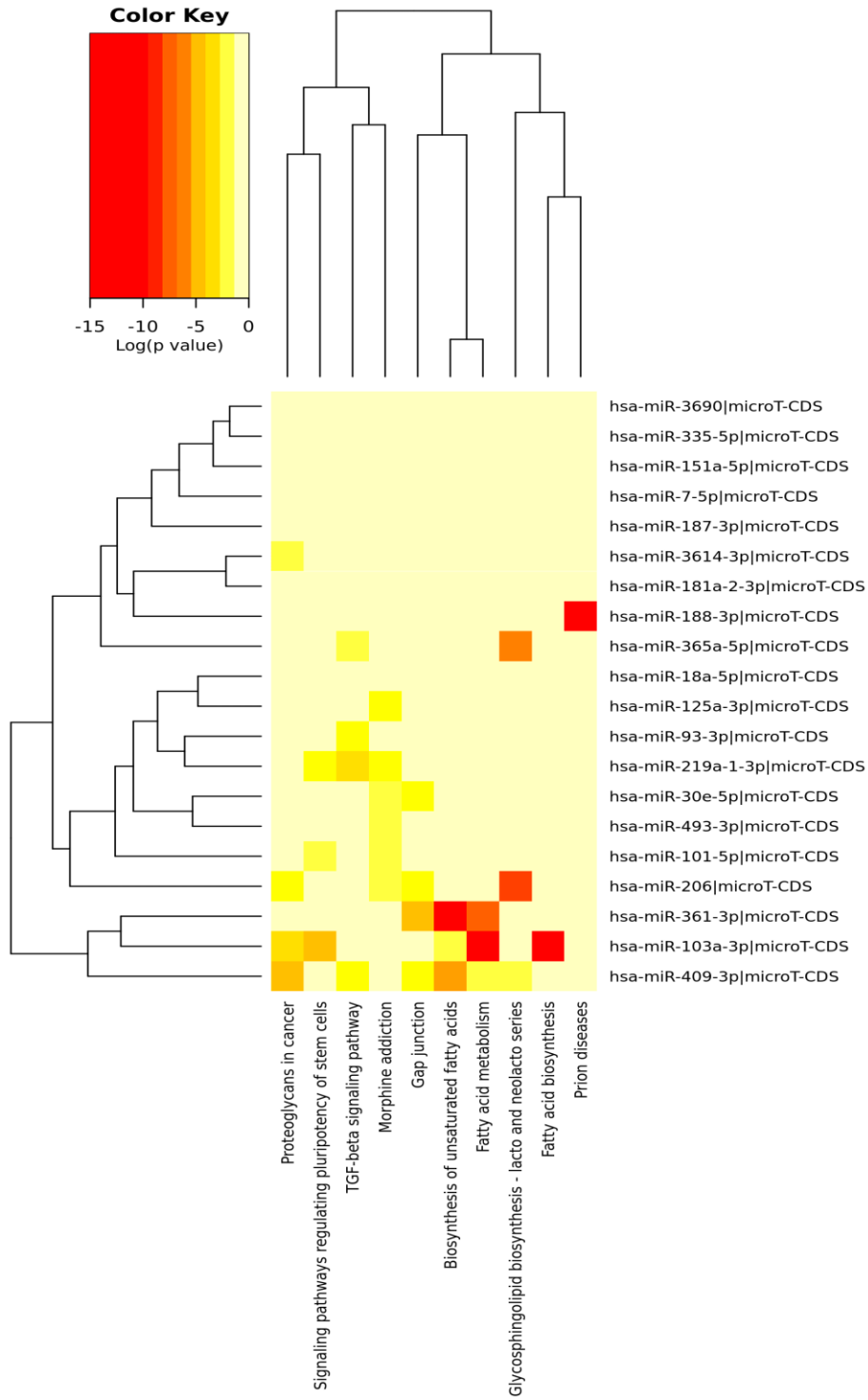
Supplementary Figure S4.2. KEGG pathway enrichment analysis of miRNA signatures in BRCA



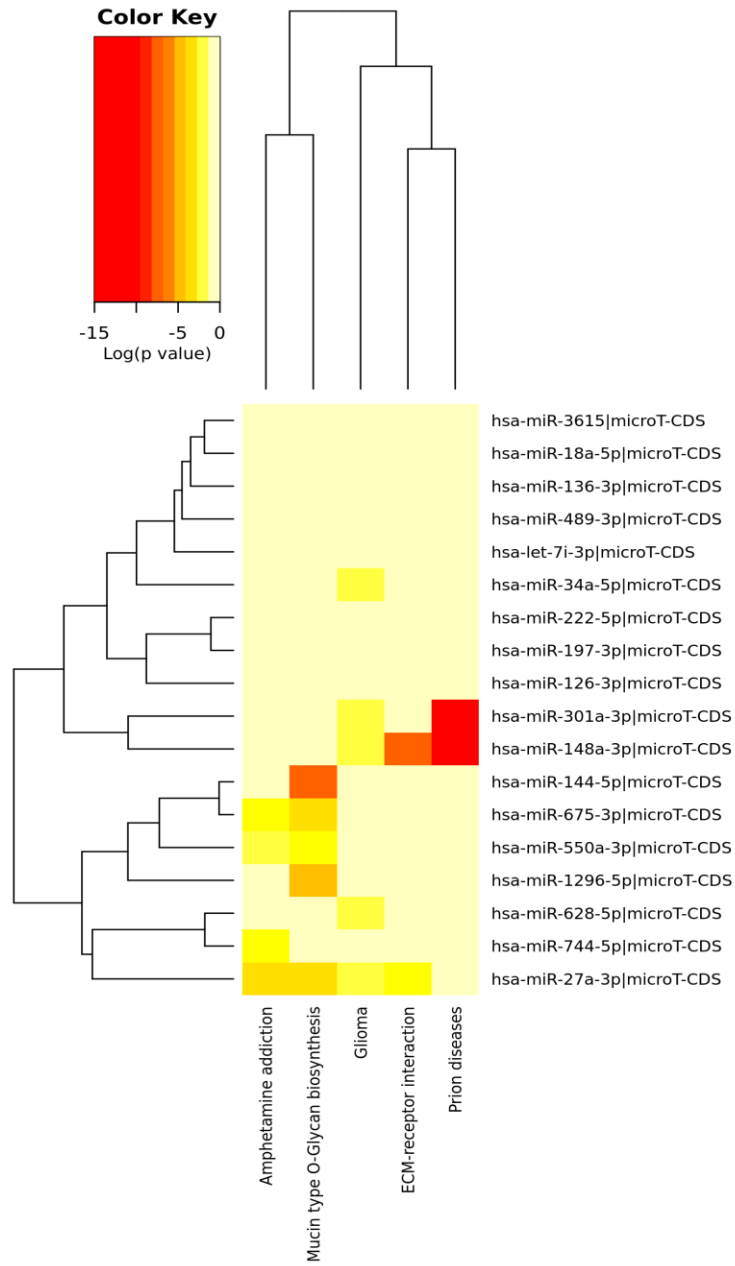
Supplementary Figure S4.3. KEGG pathway enrichment analysis of miRNA signatures in COAD



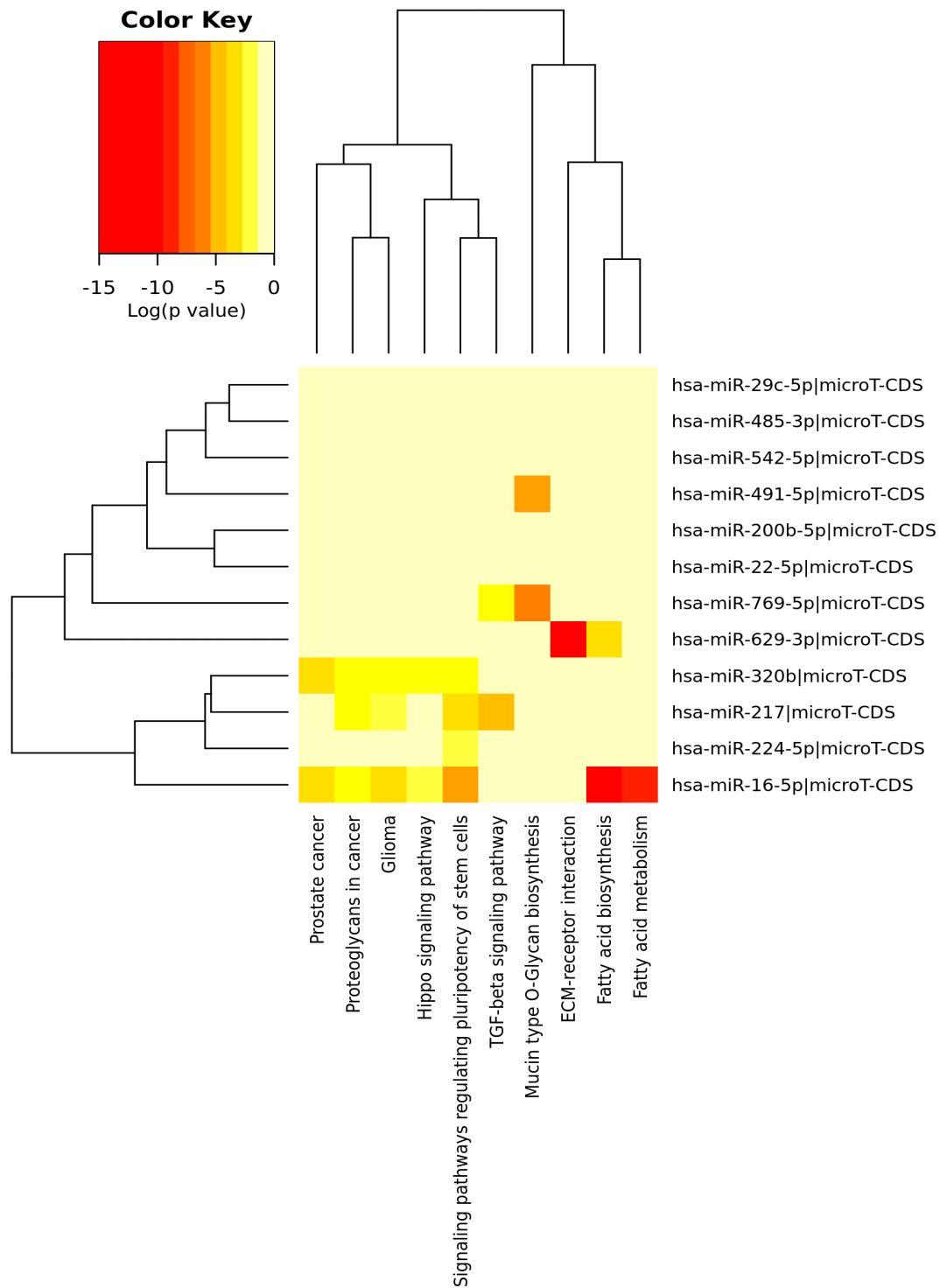
Supplementary Figure S4.4. KEGG pathway enrichment analysis of miRNA signatures in ESCA



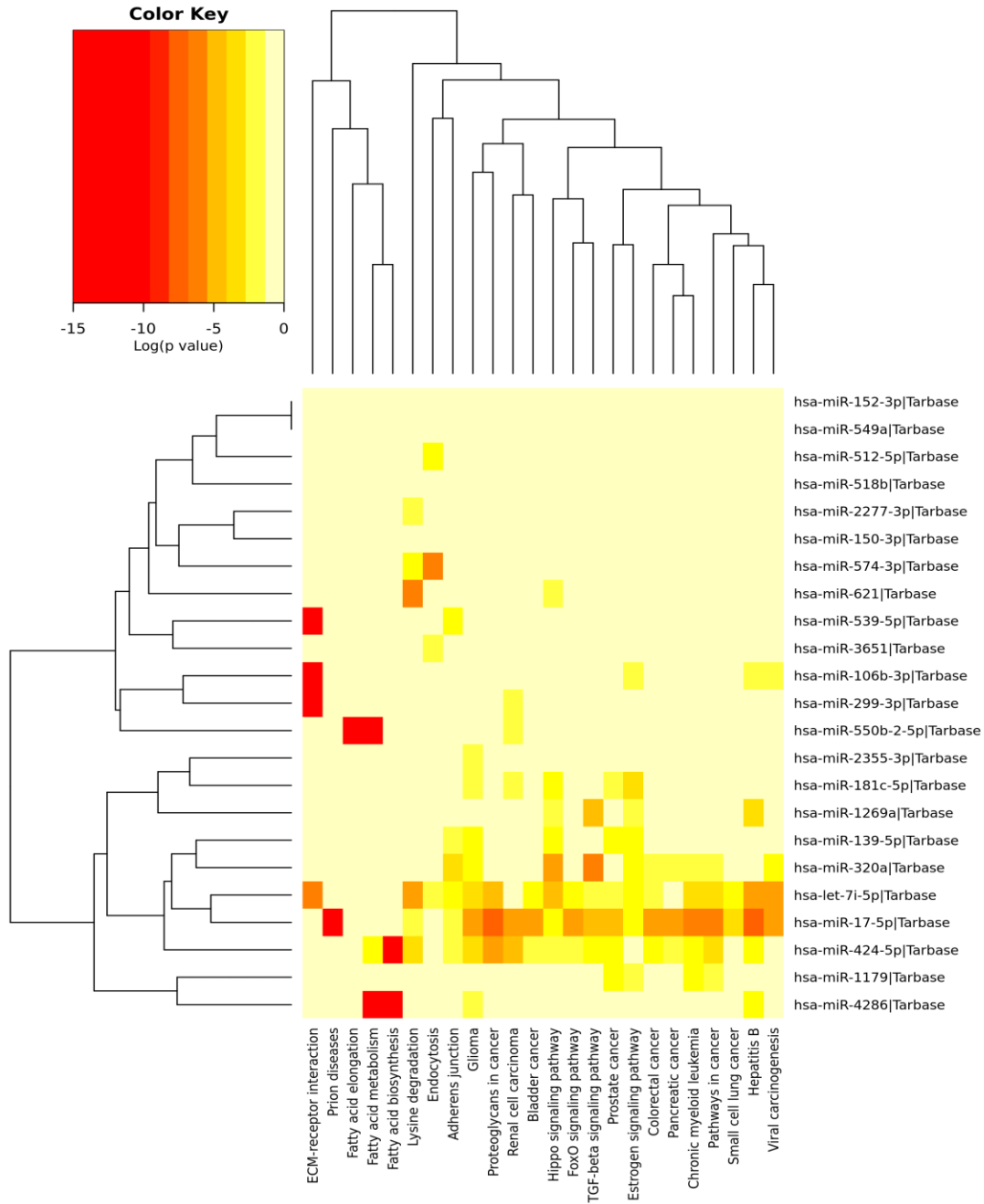
Supplementary Figure S4.5. KEGG pathway enrichment analysis of miRNA signatures in HNSC



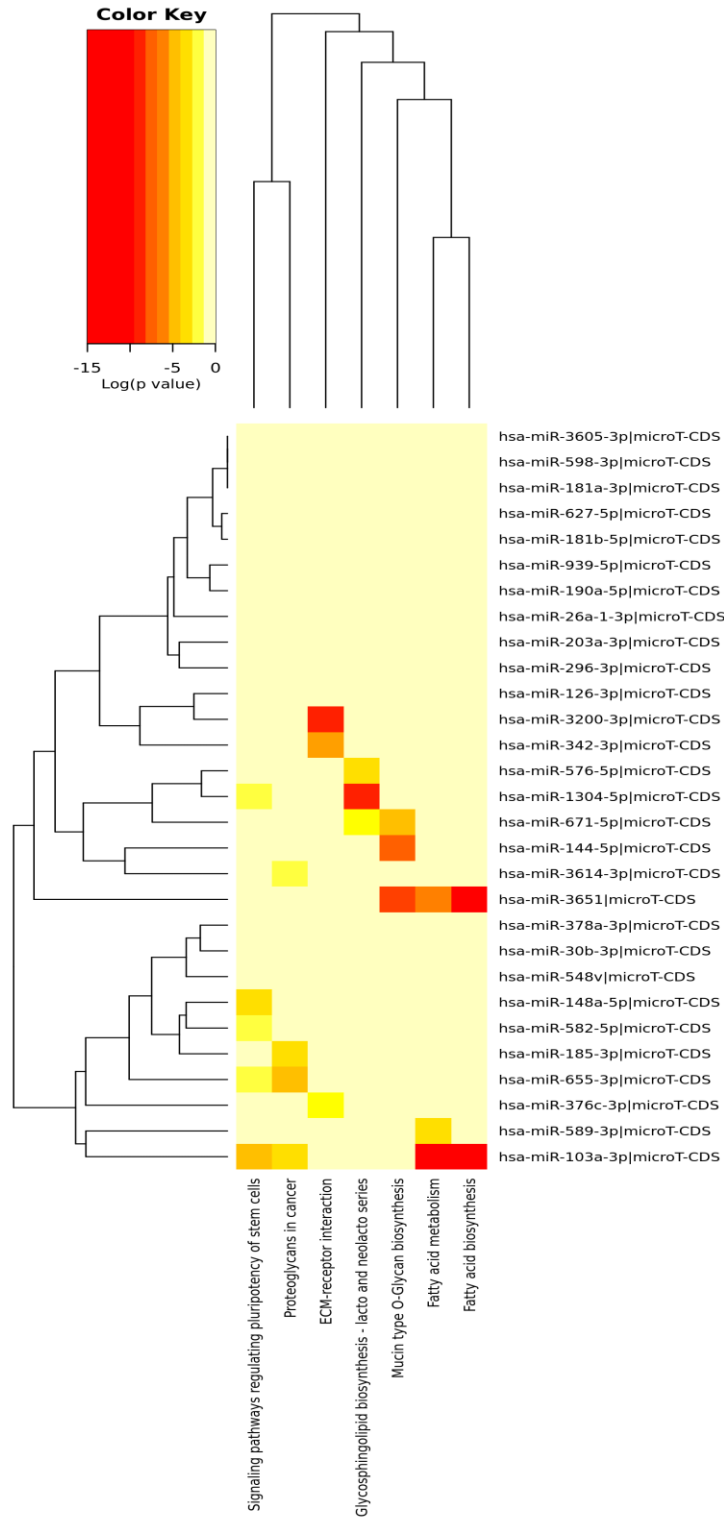
Supplementary Figure S4.6. KEGG pathway enrichment analysis of miRNA signatures in KIRC



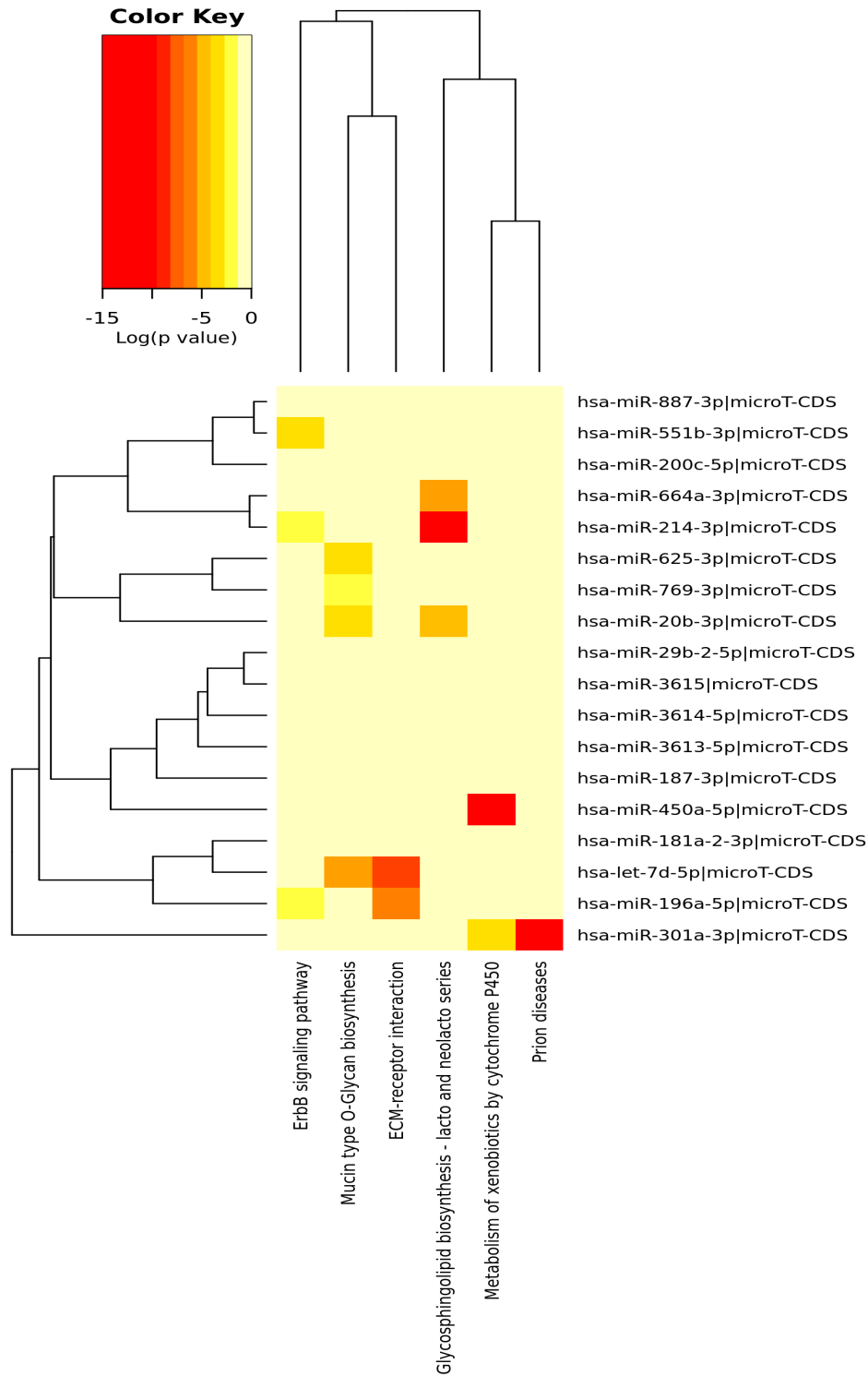
Supplementary Figure S4.7. KEGG pathway enrichment analysis of miRNA signatures in KIRP



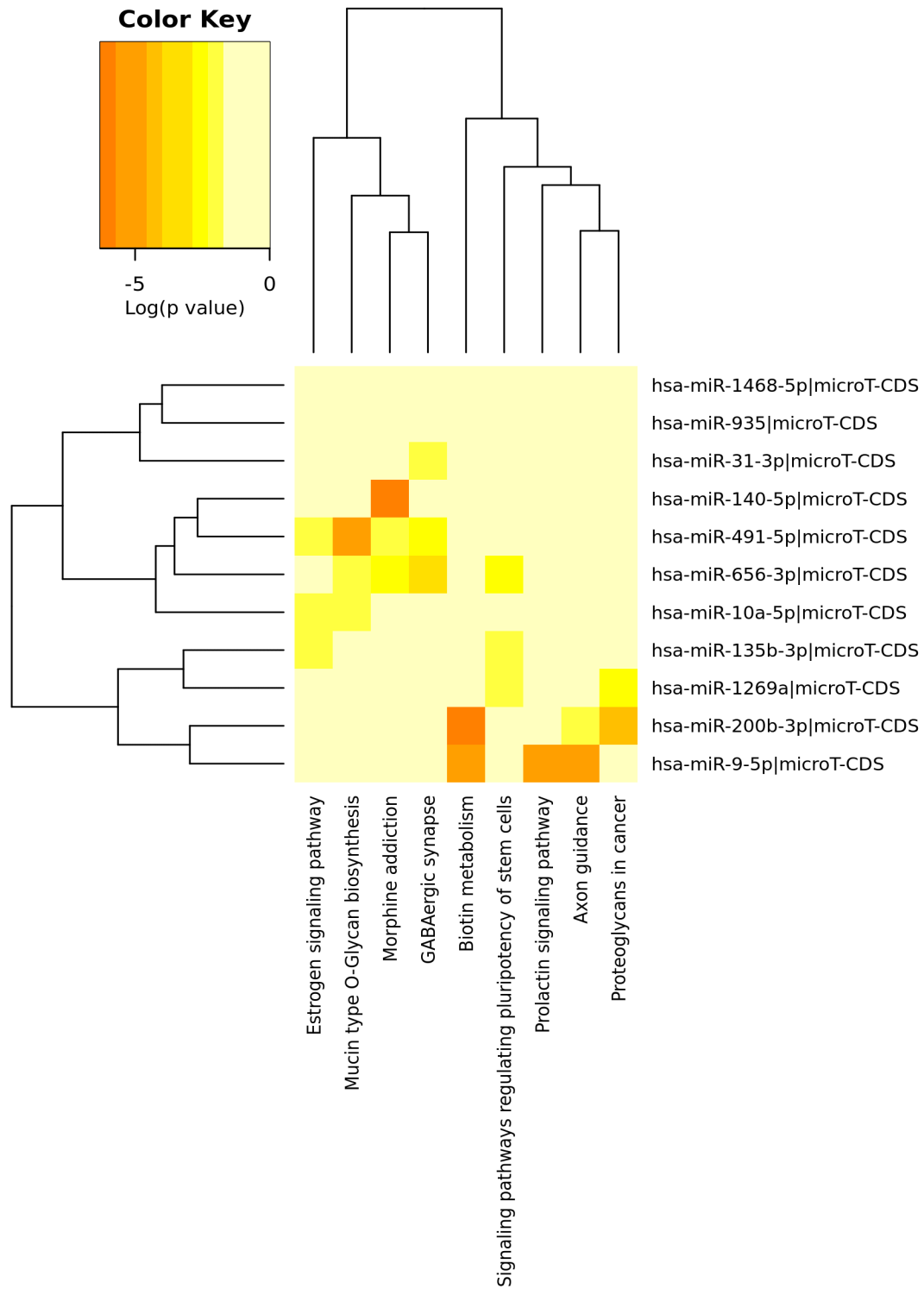
Supplementary Figure S4.8. KEGG pathway enrichment analysis of miRNA signatures in LIHC



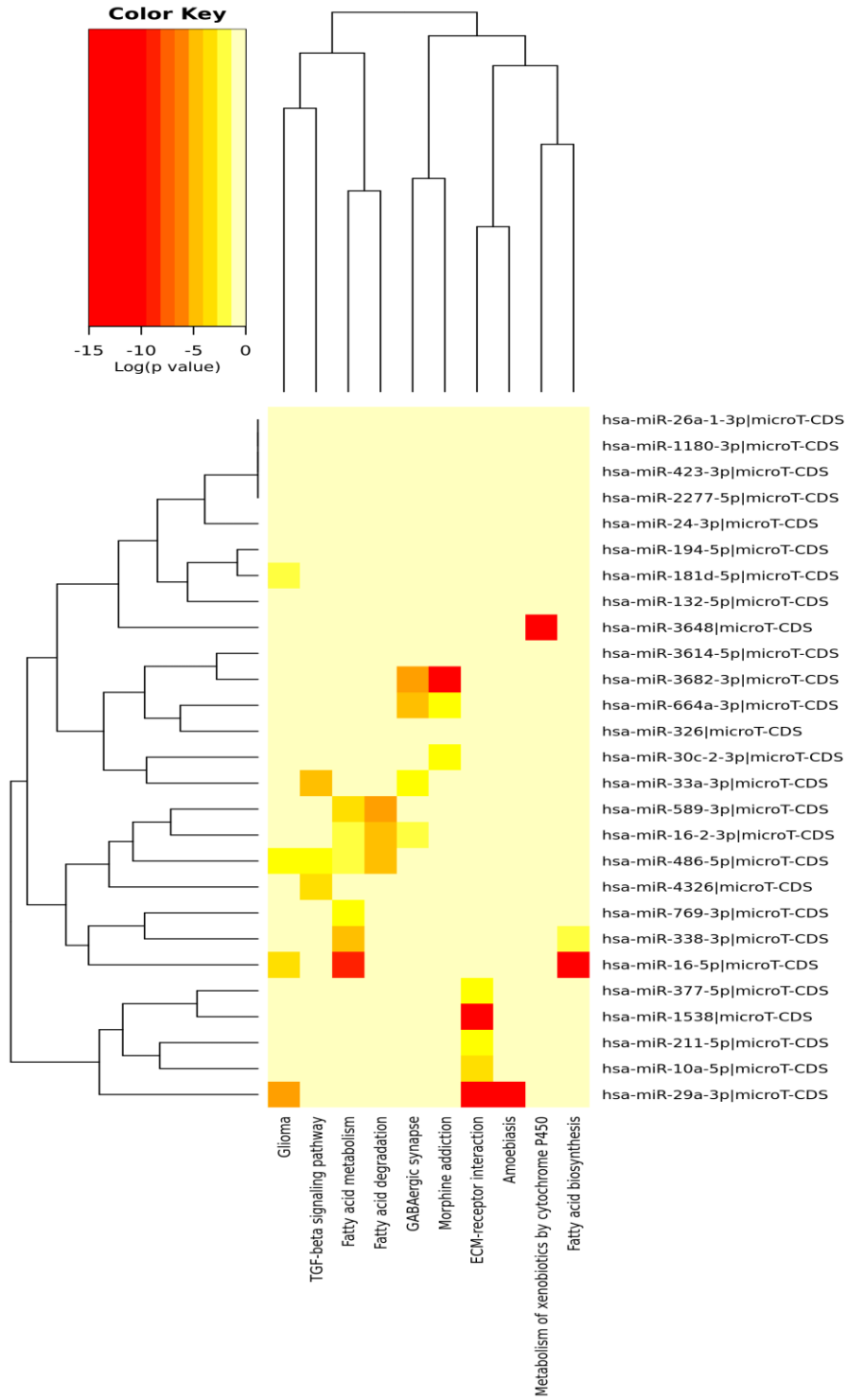
Supplementary Figure S4.9. KEGG pathway enrichment analysis of miRNA signatures in LUAD



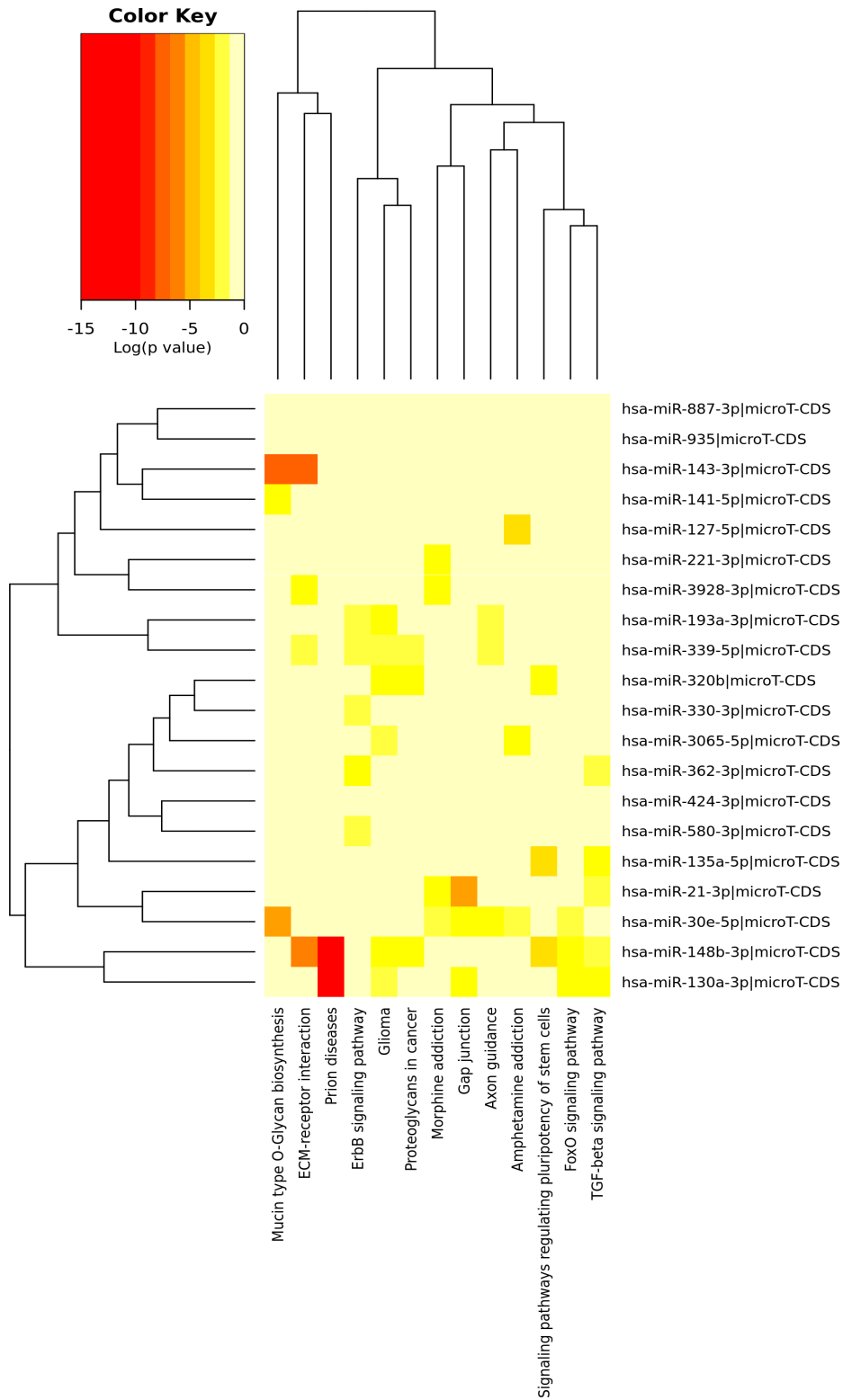
Supplementary Figure S4.10. KEGG pathway enrichment analysis of miRNA signatures in LUSC



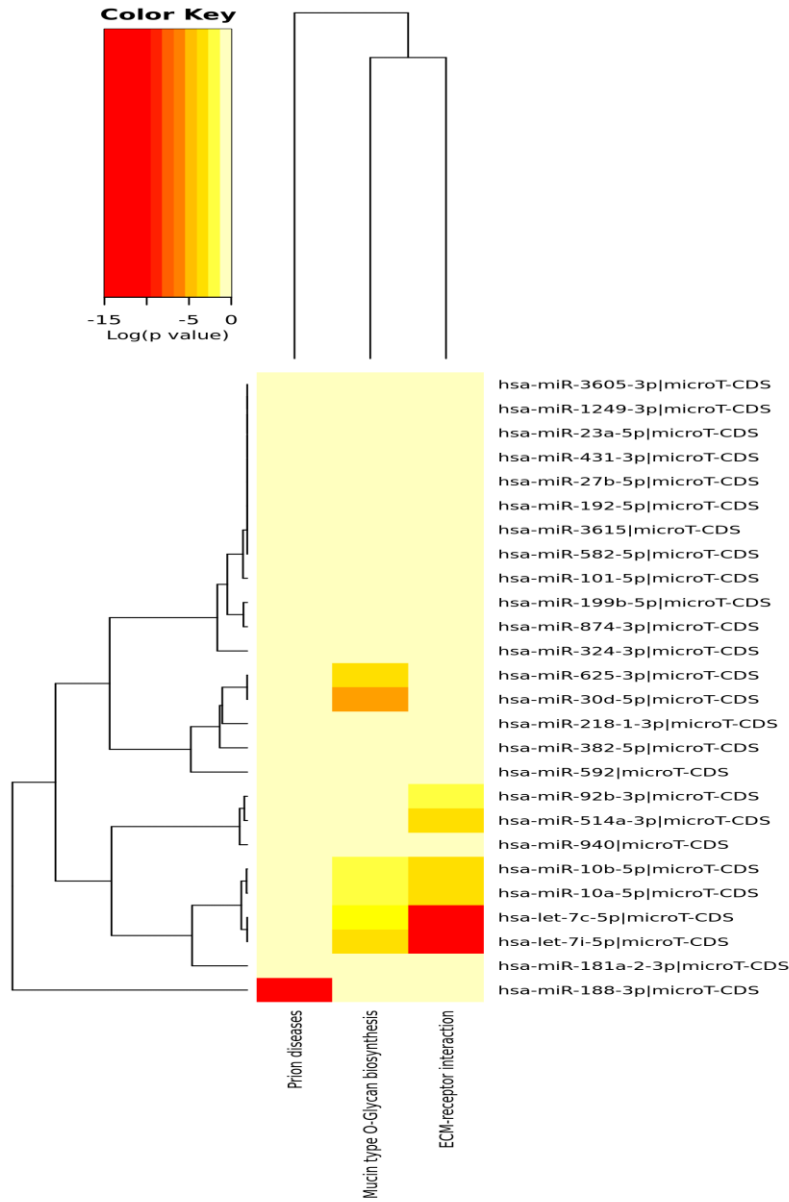
Supplementary Figure S4.11. KEGG pathway enrichment analysis of miRNA signatures in READ



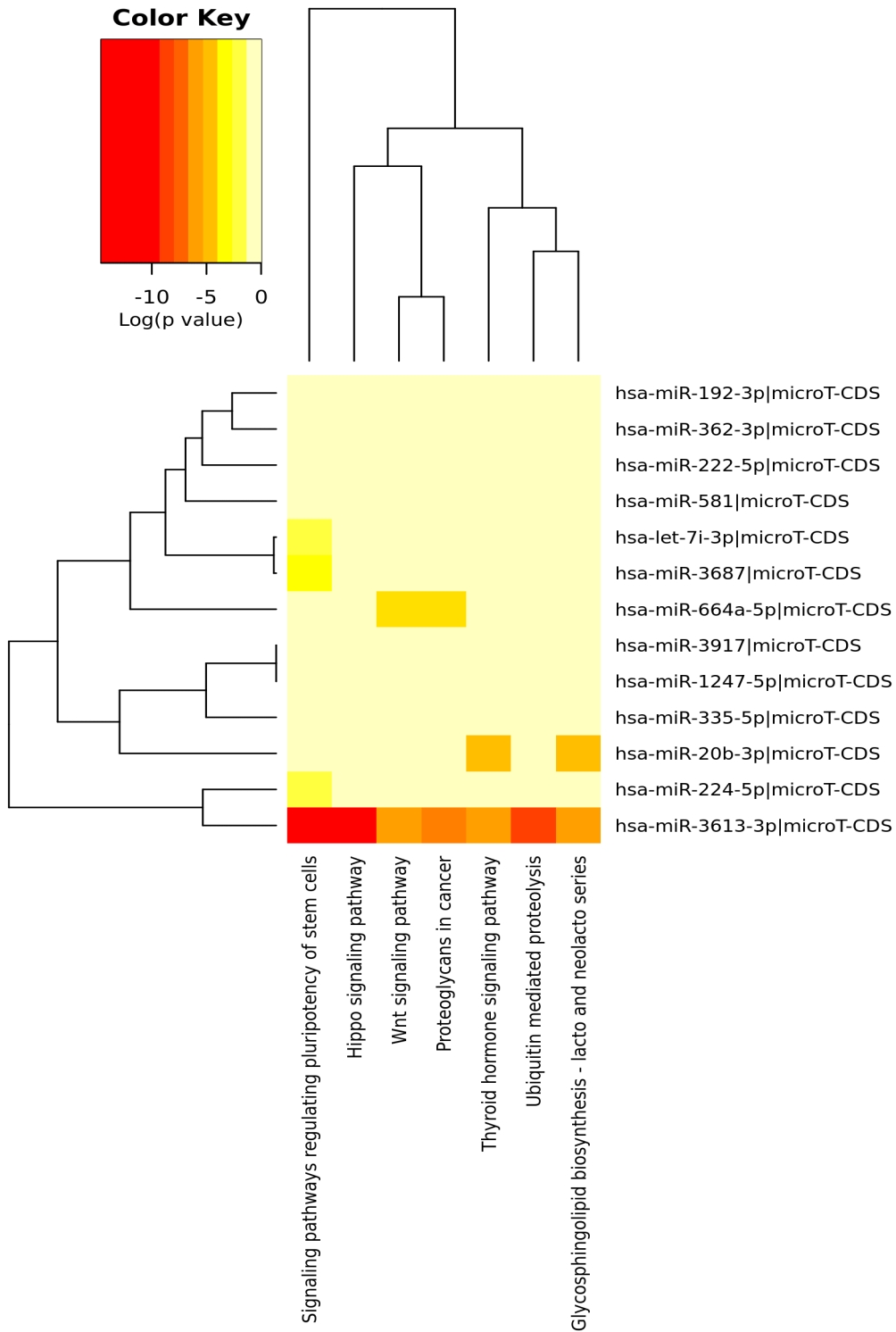
Supplementary Figure S4.12. KEGG pathway enrichment analysis of miRNA signatures in SKCM



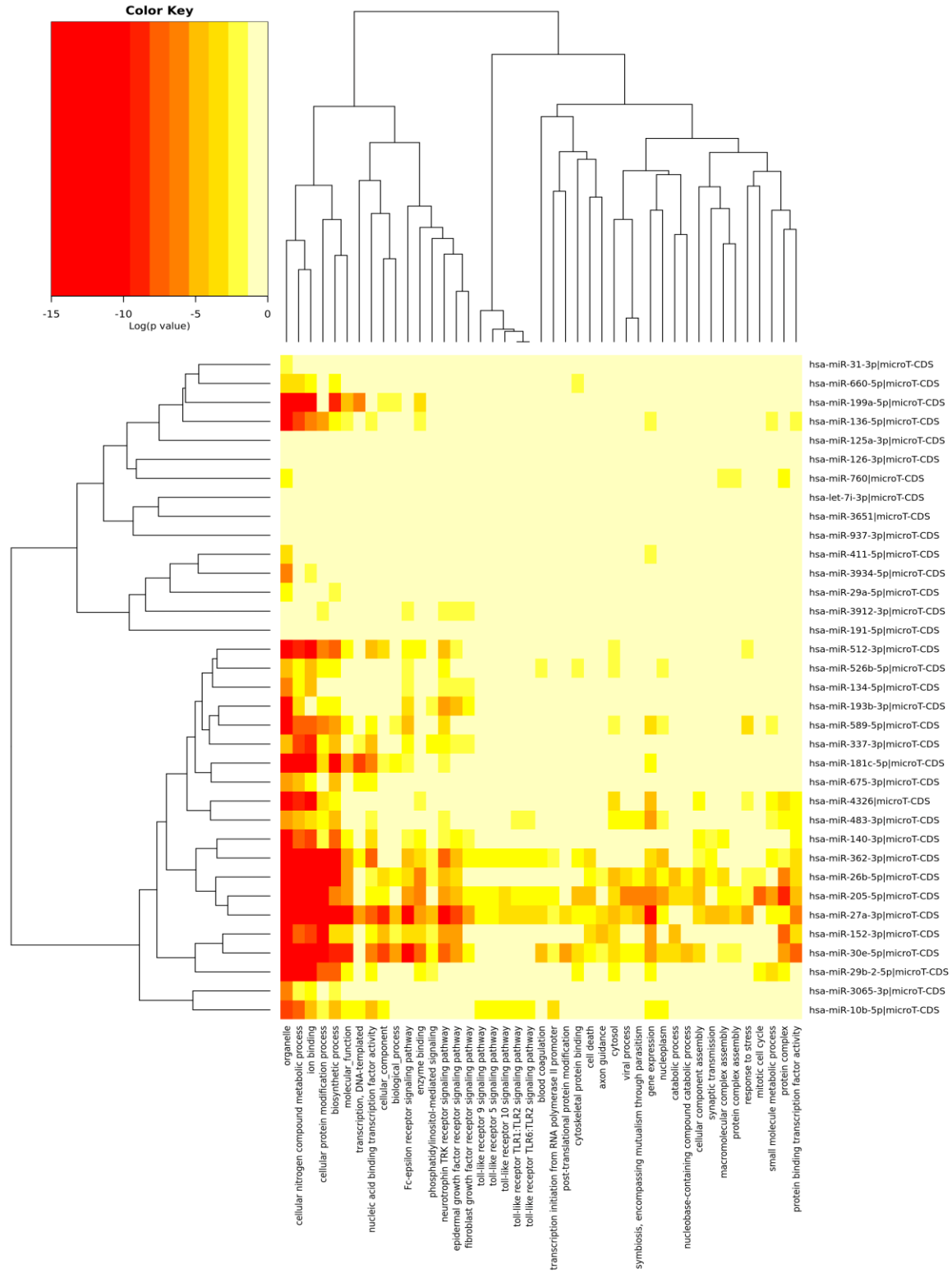
Supplementary Figure S4.13. KEGG pathway enrichment analysis of miRNA signatures in STAD



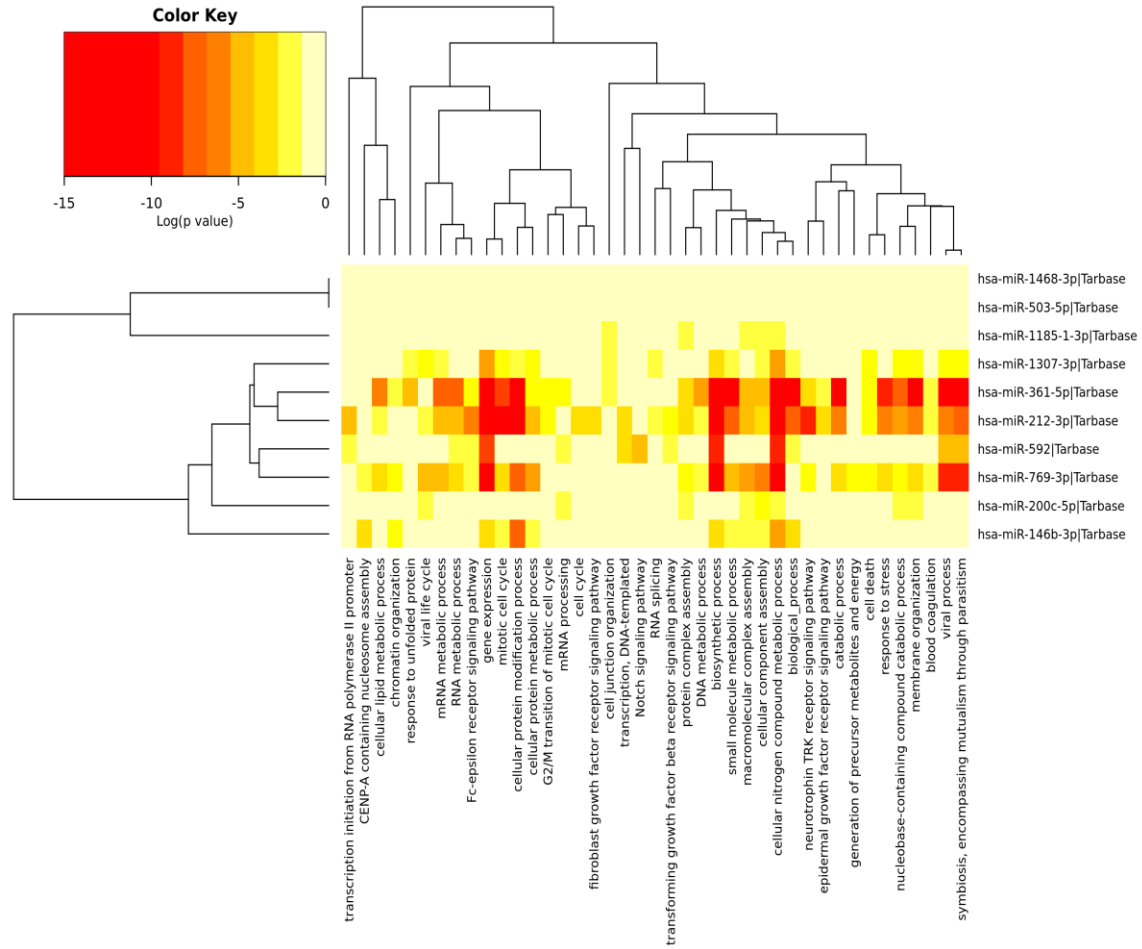
Supplementary Figure S4.14. KEGG pathway enrichment analysis of miRNA signatures in THCA



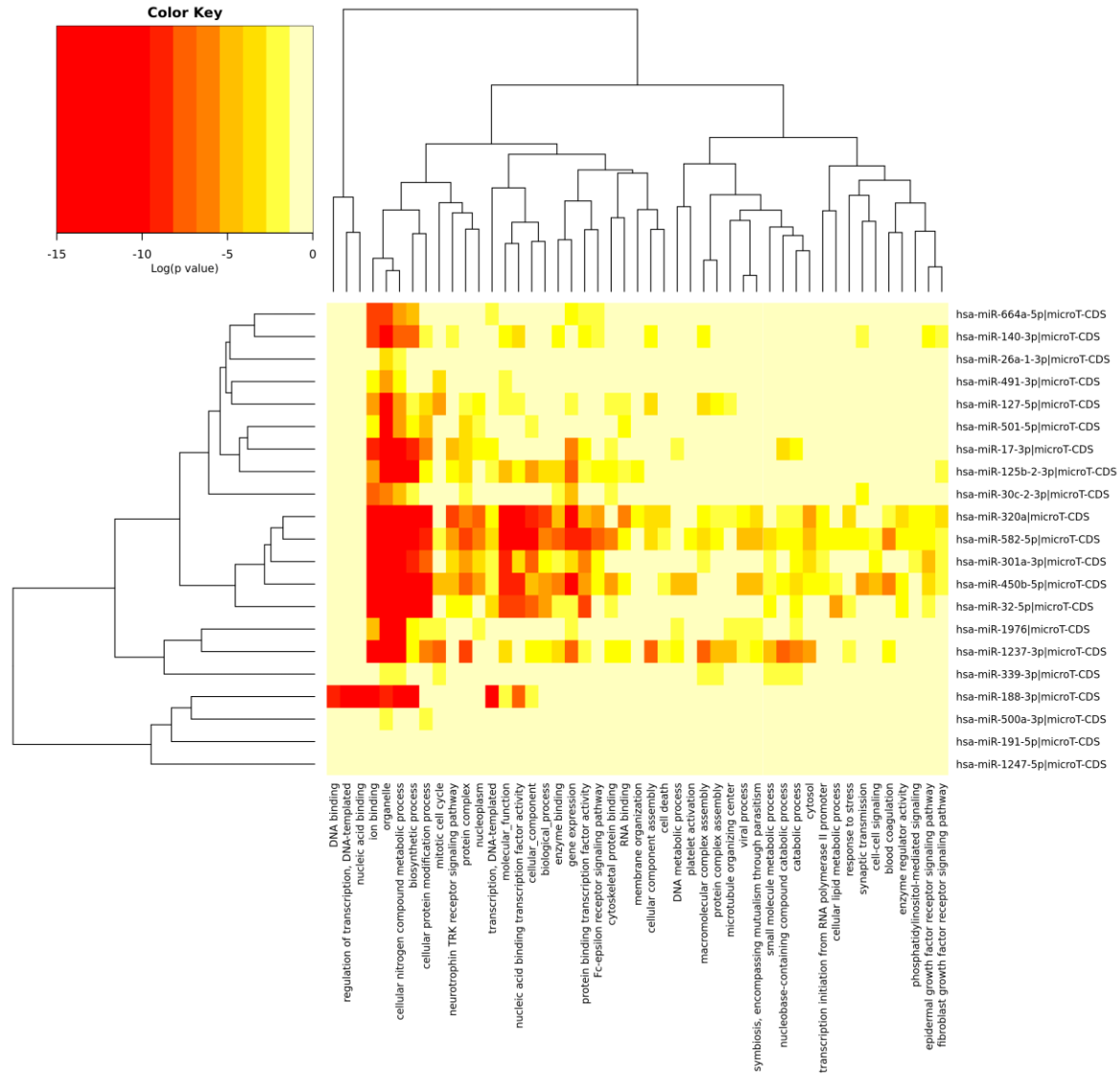
Supplementary Figure S4.15. KEGG pathway enrichment analysis of miRNA signatures in UVM



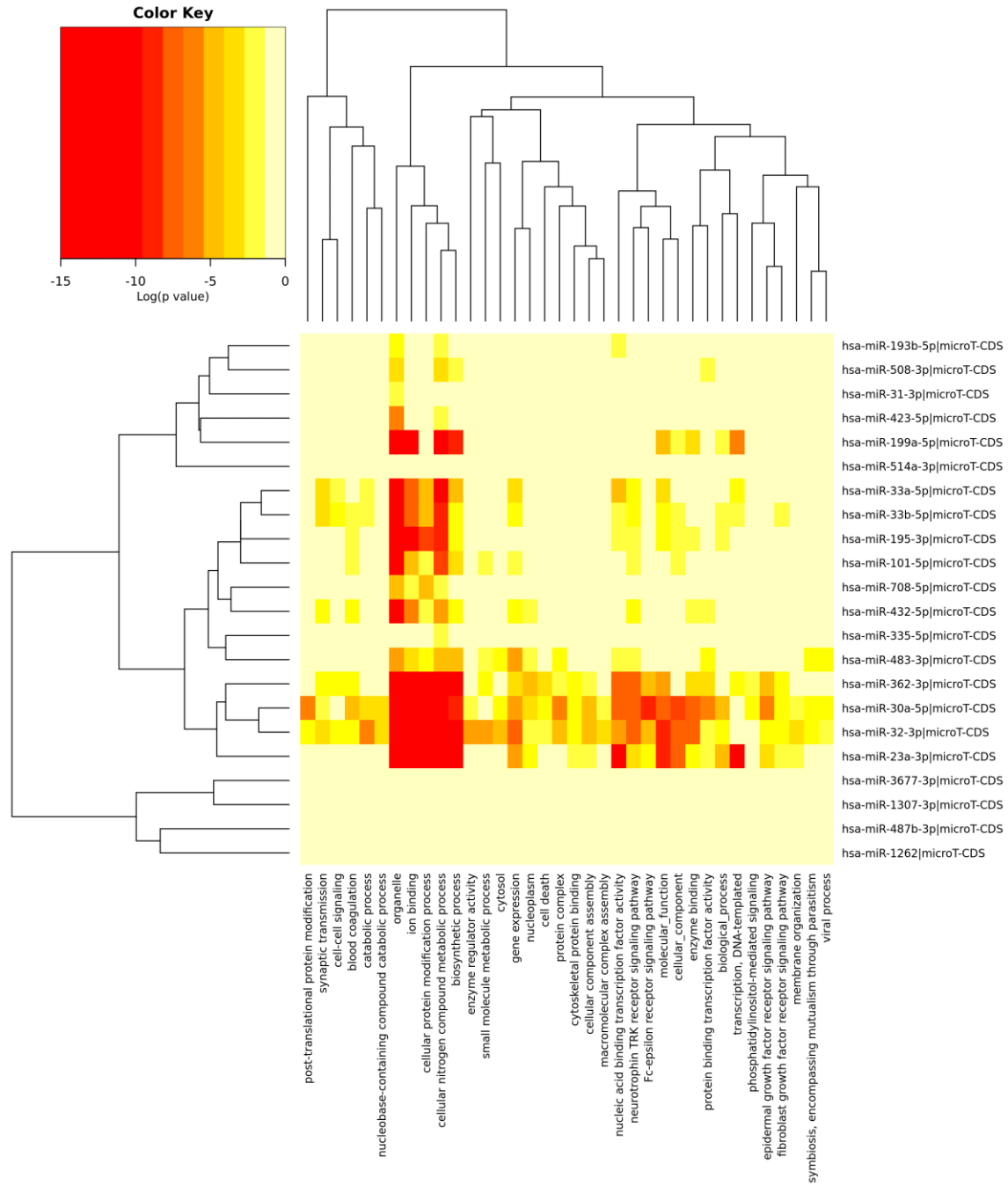
Supplementary Figure S5.1. GO category enrichment analysis of miRNA signatures in BLCA



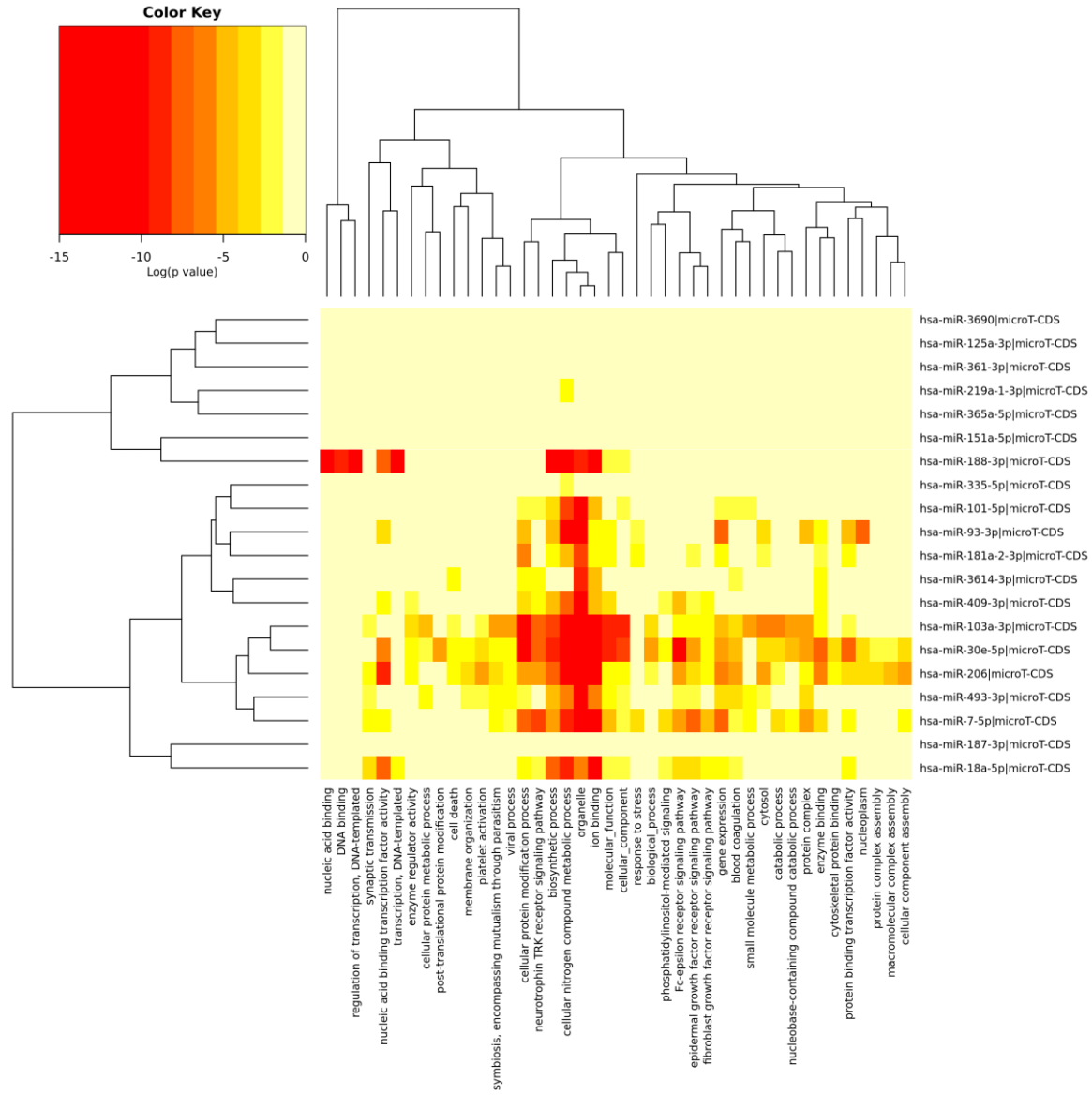
Supplementary Figure S5.2. GO category enrichment analysis of miRNA signatures in BRCA



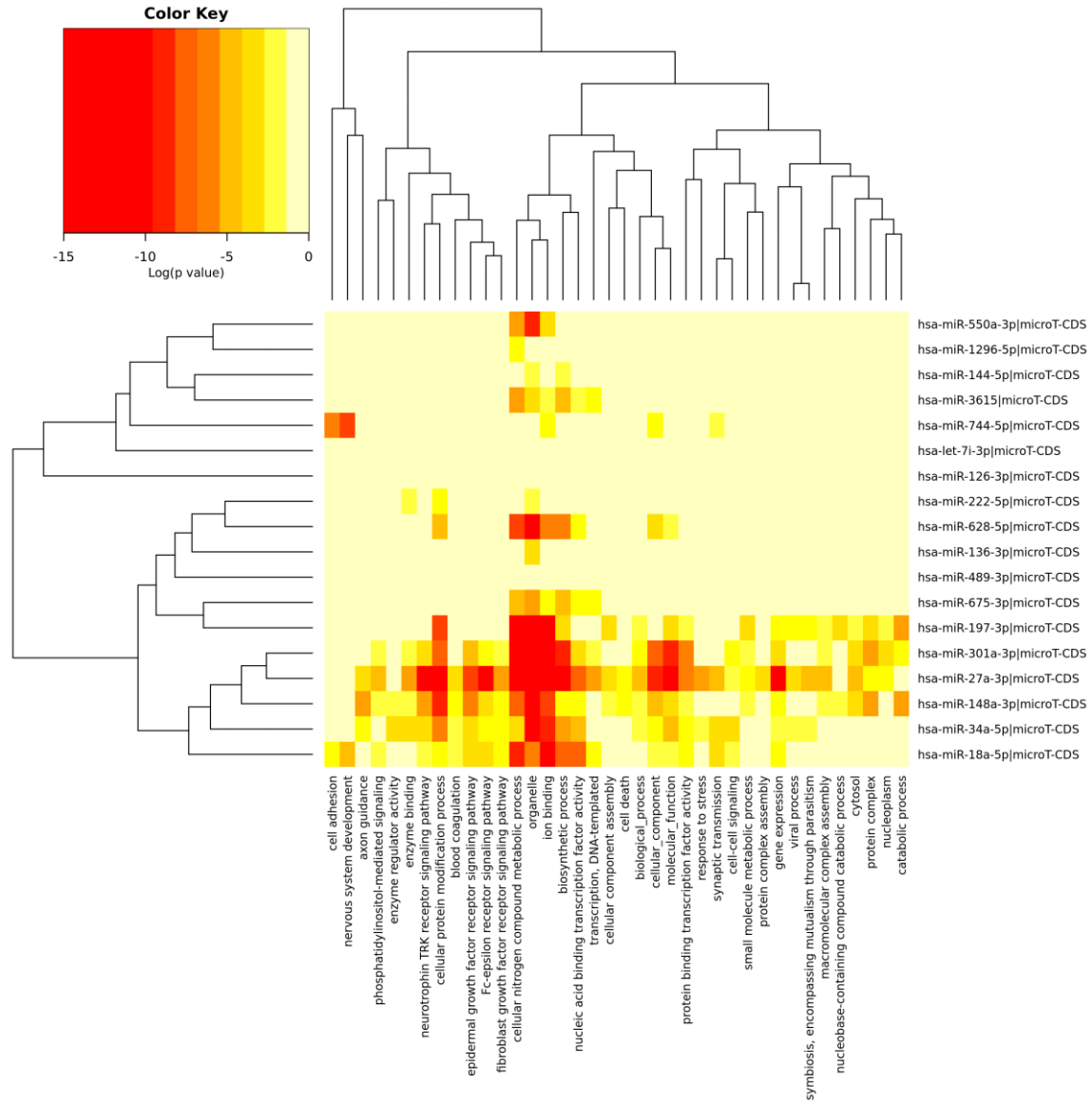
Supplementary Figure S5.3. GO category enrichment analysis of miRNA signatures in COAD



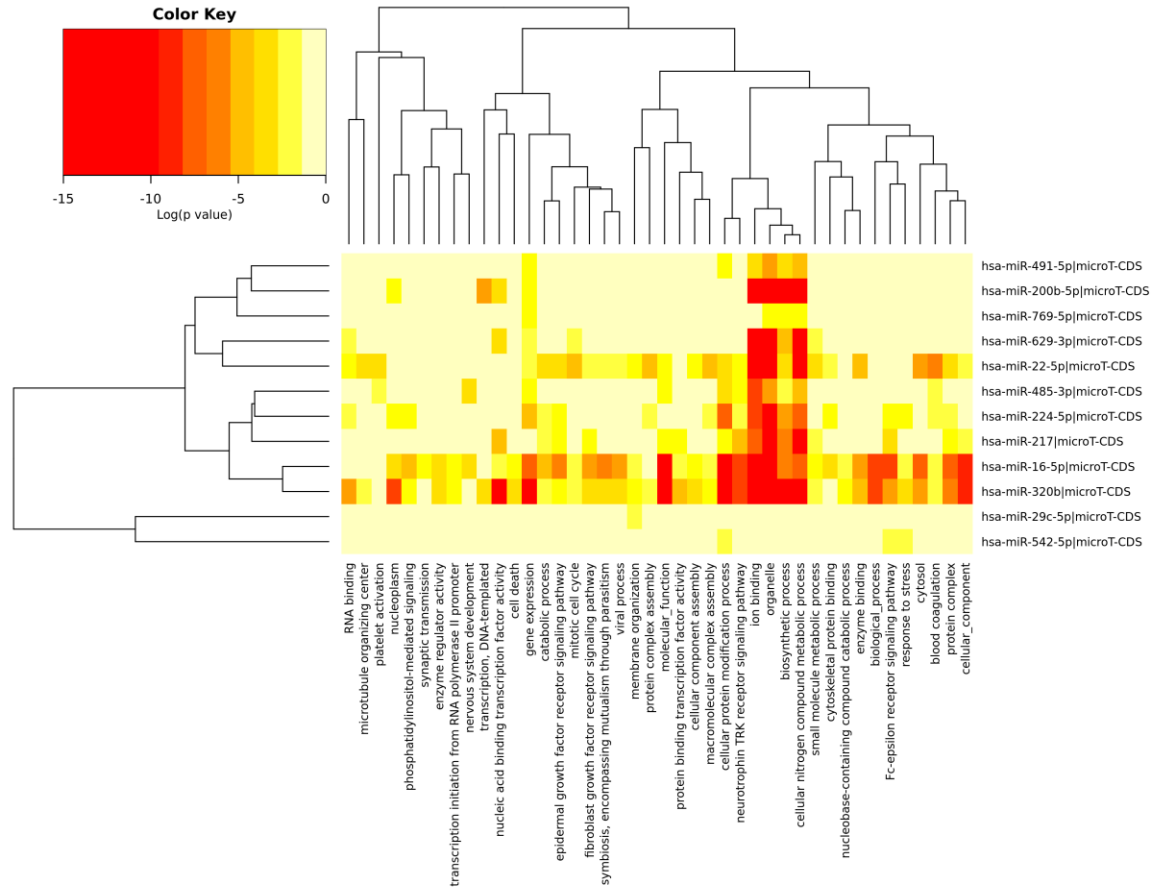
Supplementary Figure S5.4. GO category enrichment analysis of miRNA signatures in ESCA



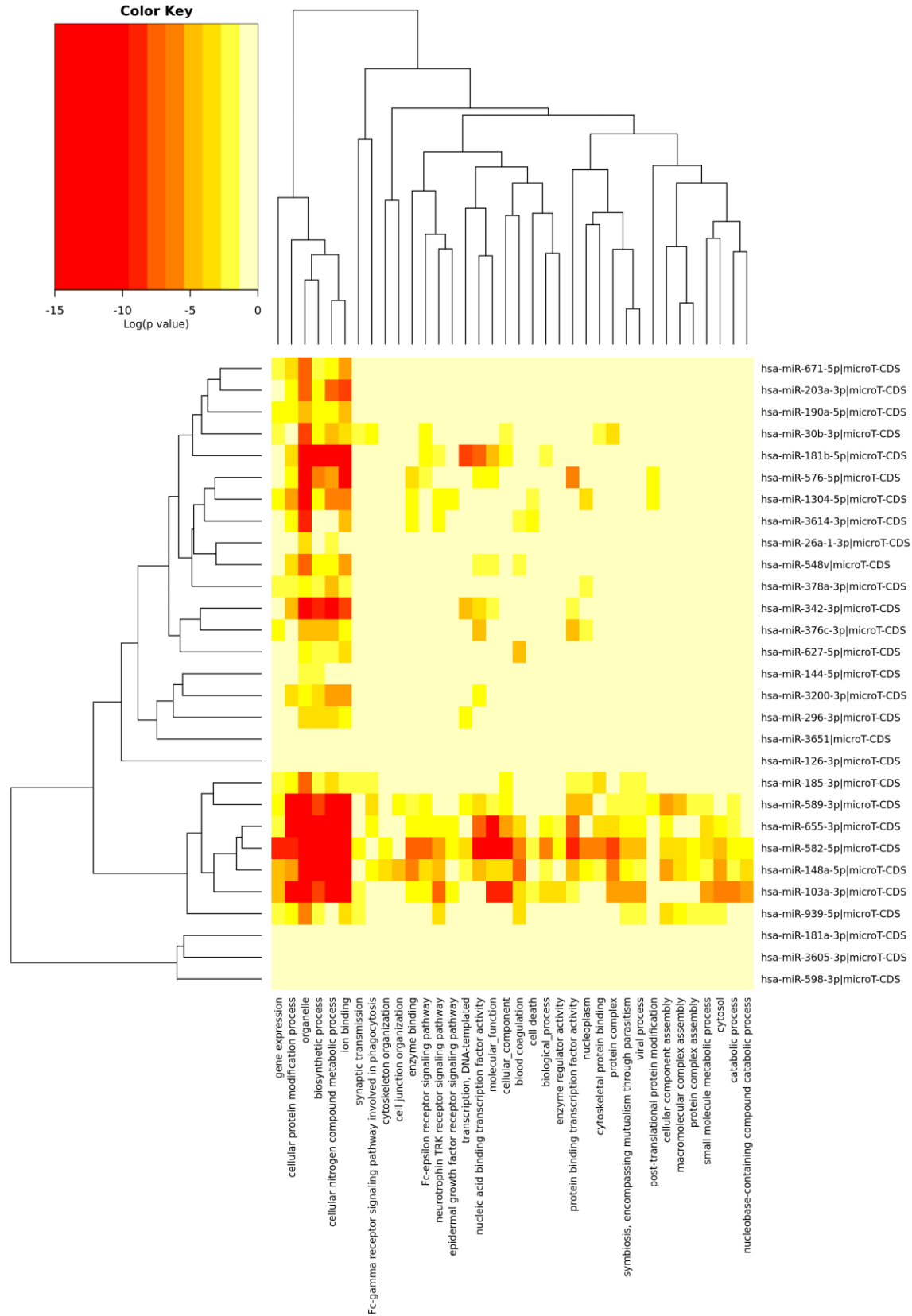
Supplementary Figure S5.5. GO category enrichment analysis of miRNA signatures in HNSC



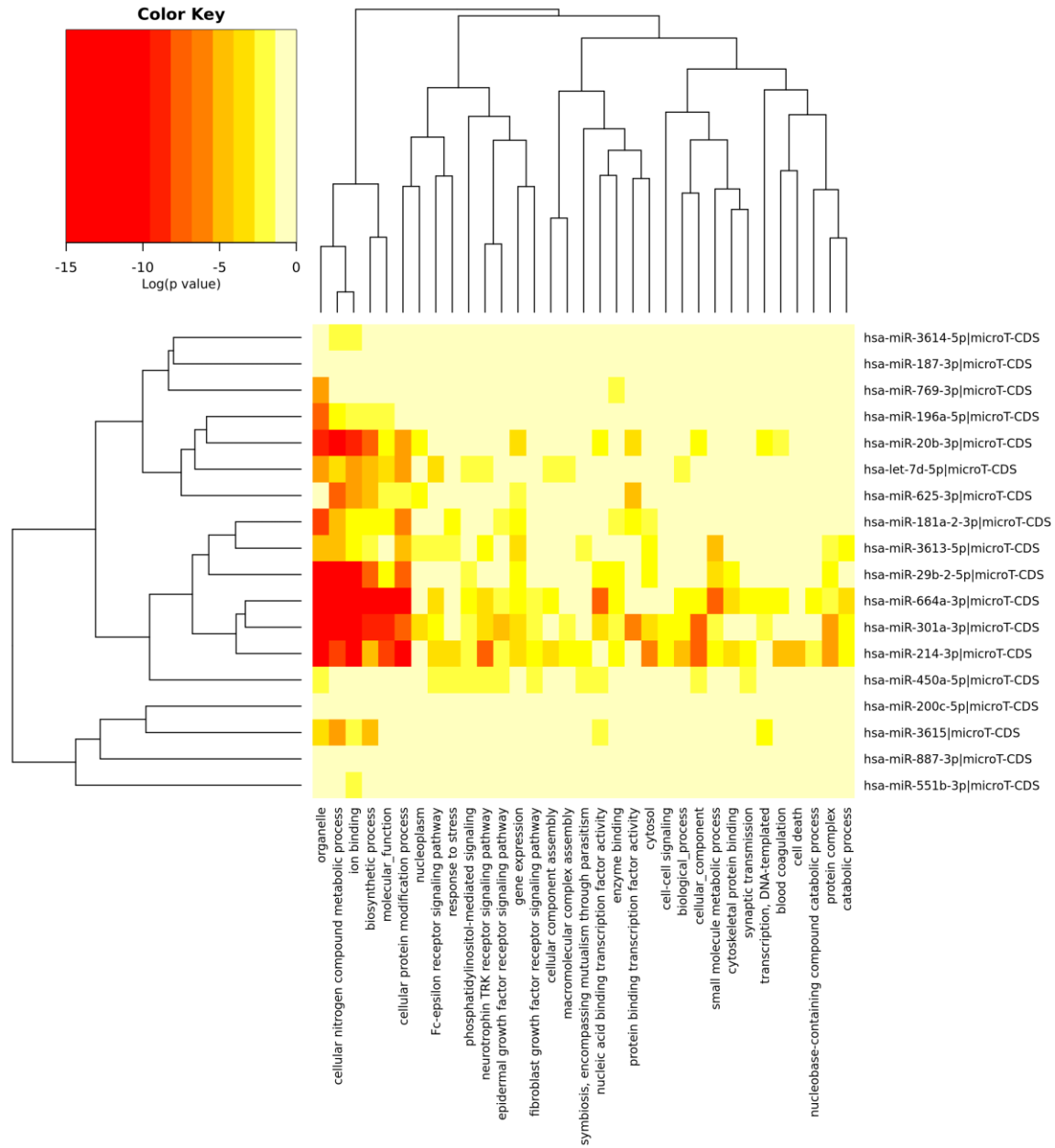
Supplementary Figure S5.6. GO category enrichment analysis of miRNA signatures in KIRC



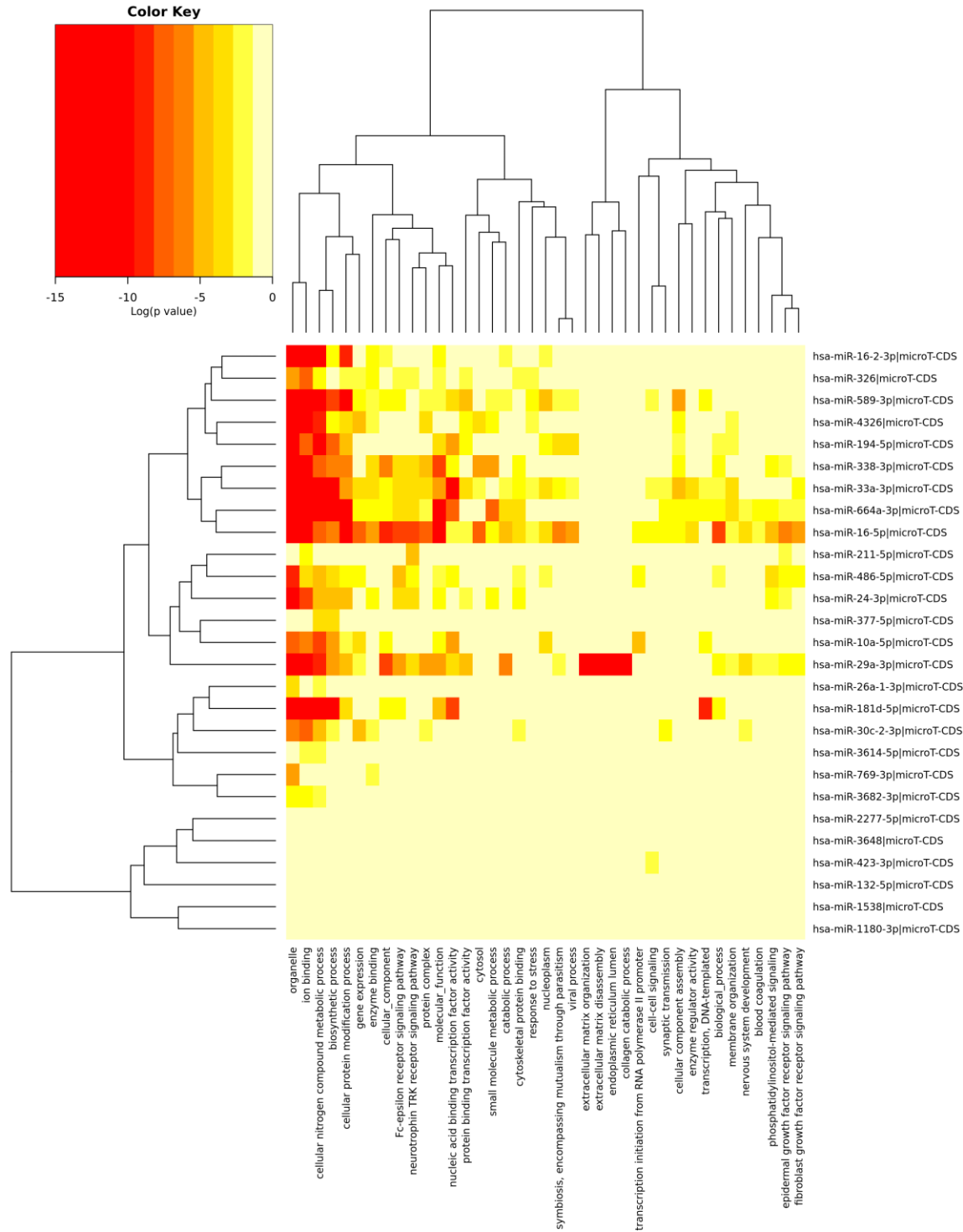
Supplementary Figure S5.7. GO category enrichment analysis of miRNA signatures in KIRP



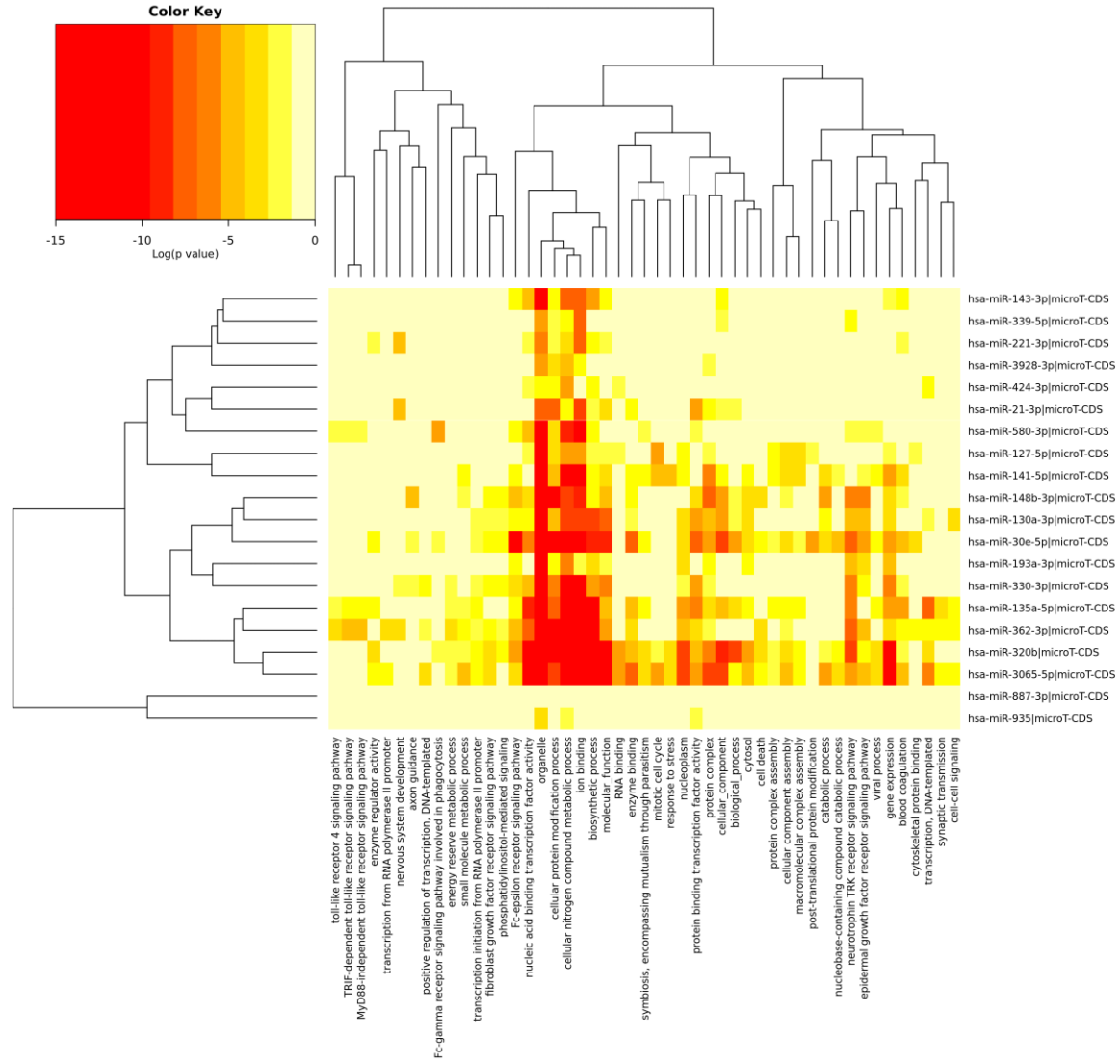
Supplementary Figure S5.9. GO category enrichment analysis of miRNA signatures in LUAD



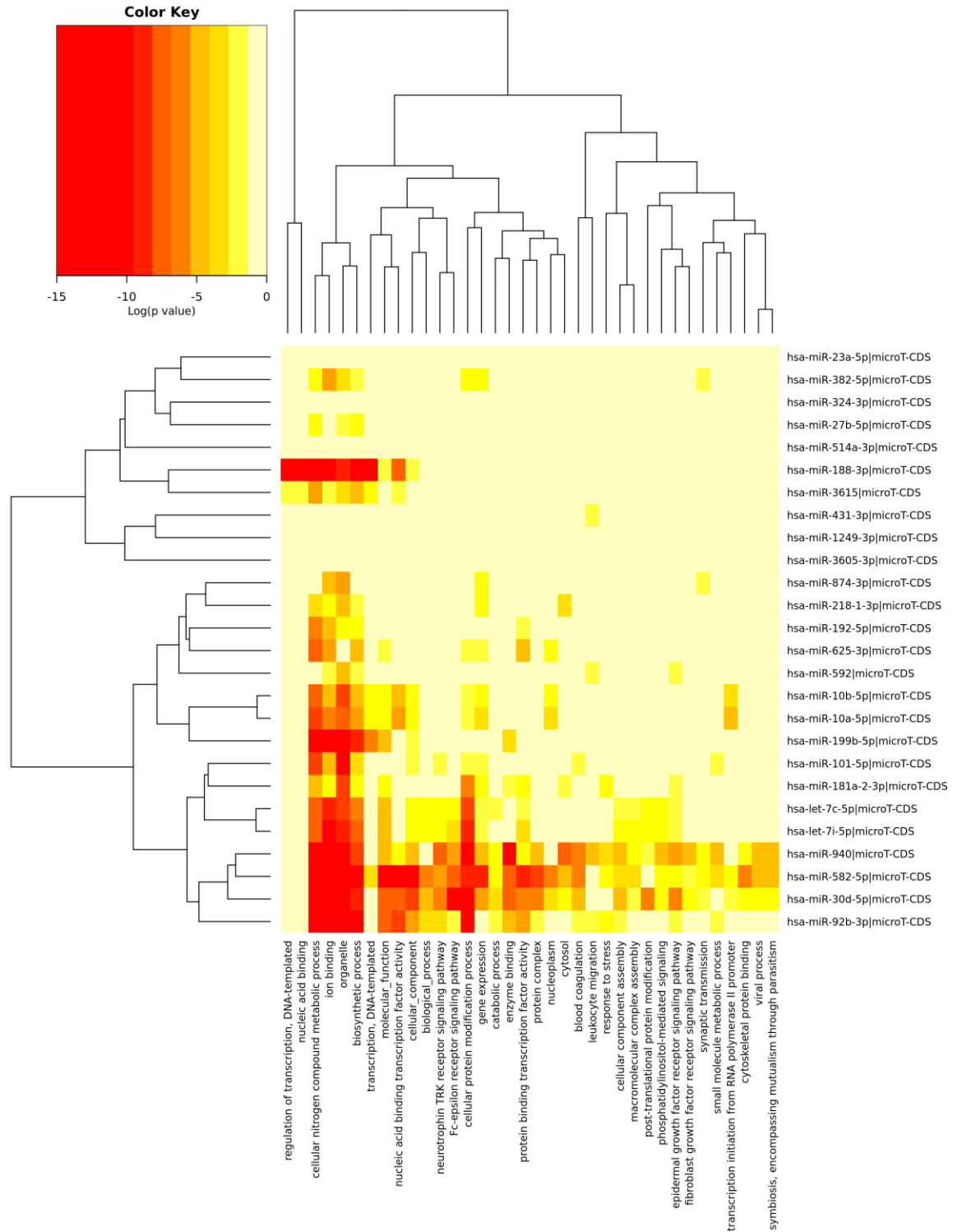
Supplementary Figure S5.10. GO category enrichment analysis of miRNA signatures in LUSC



Supplementary Figure S5.12. GO category enrichment analysis of miRNA signatures in SKCM



Supplementary Figure S5.13. GO category enrichment analysis of miRNA signatures in STAD



Supplementary Figure S5.14. GO category enrichment analysis of miRNA signatures in THCA

Supplementary Tables

Supplementary Table S1. The prediction comparison results of CancerSig with different machine learning methods.

Method	miRNA-signature	10-CV Accuracy	Sensitivity	Specificity	MCC	AUC
CancerSig-BLCA	35	84.40±1.27	0.64±0.04	0.93±0.01	0.65±0.02	0.82±0.01
LightGBM	35	72.22±0.04	0.83±0.06	0.51±0.14	0.35±0.10	0.72±0.06
XGBoost	35	72.70±0.06	0.88±0.06	0.41±0.16	0.33±0.18	0.73±0.05
Random Forest	35	70.98±0.05	0.77±0.07	0.59±0.16	0.35±0.13	0.74±0.05
CatBoost	35	73.93±0.06	0.91±0.07	0.39±0.12	0.37±0.15	0.74±0.05
Extra Trees	35	72.72±0.04	0.78±0.07	0.63±0.11	0.40±0.09	0.74±0.06

Supplementary Table S2.1. Contribution of individual miRNAs using MED analysis in BLCA

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-31-3p	MIMAT0004504	45.21
2	hsa-miR-29b-2-5p	MIMAT0004515	40.29
3	hsa-miR-193b-3p	MIMAT0002819	31.45
4	hsa-miR-10b-5p	MIMAT0000254	30.47
5	hsa-miR-125a-3p	MIMAT0004602	28.01
6	hsa-miR-136-5p	MIMAT0000448	20.15
7	hsa-miR-3912-3p	MIMAT0018186	19.16
8	hsa-miR-205-5p	MIMAT0000266	17.69
9	hsa-miR-337-3p	MIMAT0000754	17.69
10	hsa-miR-29a-5p	MIMAT0004503	17.20
11	hsa-miR-512-3p	MIMAT0002823	14.74
12	hsa-miR-3934-5p	MIMAT0018349	14.25
13	hsa-miR-4326	MIMAT0016888	14.25
14	hsa-miR-126-3p	MIMAT0000445	12.29
15	hsa-miR-26b-5p	MIMAT0000083	12.29
16	hsa-miR-362-3p	MIMAT0004683	10.32
17	hsa-miR-760	MIMAT0004957	10.32
18	hsa-miR-27a-3p	MIMAT0000084	6.39
19	hsa-miR-199a-5p	MIMAT0000231	5.90
20	hsa-miR-483-3p	MIMAT0002173	4.91
21	hsa-miR-181c-5p	MIMAT0000258	3.93
22	hsa-let-7i-3p	MIMAT0004585	3.93

23	hsa-miR-526b-5p	MIMAT0002835	3.93
24	hsa-miR-660-5p	MIMAT0003338	3.93
25	hsa-miR-152-3p	MIMAT0000438	3.44
26	hsa-miR-3651	MIMAT0018071	3.44
27	hsa-miR-675-3p	MIMAT0006790	2.46
28	hsa-miR-140-3p	MIMAT0004597	1.97
29	hsa-miR-3065-3p	MIMAT0015378	1.47
30	hsa-miR-191-5p	MIMAT0000440	1.47
31	hsa-miR-134-5p	MIMAT0000447	0.49
32	hsa-miR-937-3p	MIMAT0004980	0.49
33	hsa-miR-30e-5p	MIMAT0000692	0.49
34	hsa-miR-589-5p	MIMAT0004799	0.00
35	hsa-miR-411-5p	MIMAT0003329	0.00

Supplementary Table S2.2. Contribution of individual miRNAs using MED analysis in BRCA

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-200c-5p	MIMAT0004657	69.69
2	hsa-miR-503	MIMAT0002874	65.03
3	hsa-miR-1307-3p	MIMAT0005951	48.45
4	hsa-miR-361-3p	MIMAT0004682	47.93
5	hsa-miR-212	MIMAT0022695	46.89
6	hsa-miR-592	MIMAT0003260	46.89
7	hsa-miR-1185-1	MIMAT0022838	43.26
8	hsa-miR-146b	MIMAT0004766	43.26
9	hsa-miR-1468-5p	MIMAT0006789	34.46
10	hsa-miR-769-3p	MIMAT0003887	30.83
11	hsa-miR-3941	MIMAT0018357	30.31
12	hsa-miR-496	MIMAT0002818	25.65
13	hsa-miR-33b-5p	MIMAT0003301	23.58
14	hsa-miR-432-3p	MIMAT0002815	20.98
15	hsa-miR-153-1	MIMAT0000439	19.43
16	hsa-miR-1277	MIMAT0005933	16.32
17	hsa-miR-143-3p	MIMAT0000435	12.69
18	hsa-miR-3622a	MIMAT0018004	12.69
19	hsa-miR-137	MIMAT0000429	11.14
20	hsa-miR-3129	MIMAT0019202	11.14
21	hsa-miR-331	MIMAT0000760	8.03
22	hsa-miR-3651	MIMAT0018071	6.48
23	hsa-miR-676	MIMAT0018203	5.44
24	hsa-miR-627-5p	MIMAT0003296	4.40
25	hsa-miR-10b-5p	MIMAT0000254	3.89
26	hsa-miR-30b-3p	MIMAT0004589	2.03
27	hsa-miR-218-1-3p	MIMAT0004565	2.03
28	hsa-miR-379	MIMAT0000733	1.69
29	hsa-miR-642a	MIMAT0003312	1.36

30	hsa-miR-3922	MIMAT0019227	0.88
31	hsa-miR-574-3p	MIMAT0003239	0.34
32	hsa-miR-324-5p	MIMAT0000761	0.32
33	hsa-miR-374c	MIMAT0018443	0.30
34	hsa-miR-500a-3p	MIMAT0002871	0.29

Supplementary Table S2.3. Contribution of individual miRNAs using MED analysis in COAD

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-188-3p	MIMAT0004613	73.46
2	hsa-miR-1976	MIMAT0009451	70.62
3	hsa-miR-320a	MIMAT0000510	63.98
4	hsa-miR-450b-5p	MIMAT0004909	51.66
5	hsa-miR-140-3p	MIMAT0004597	35.55
6	hsa-miR-17-3p	MIMAT0000071	34.60
7	hsa-miR-301a-3p	MIMAT0000688	33.65
8	hsa-miR-582-5p	MIMAT0003247	28.91
9	hsa-miR-491-3p	MIMAT0004765	27.01
10	hsa-miR-501-5p	MIMAT0002872	26.07
11	hsa-miR-500a-3p	MIMAT0002871	24.17
12	hsa-miR-339-3p	MIMAT0004702	21.33
13	hsa-miR-127-5p	MIMAT0004604	20.38
14	hsa-miR-30c-2-3p	MIMAT0004550	19.43
15	hsa-miR-125b-2-3p	MIMAT0004603	19.43
16	hsa-miR-664a-5p	MIMAT0005948	11.85
17	hsa-miR-32-5p	MIMAT0000090	10.90
18	hsa-miR-1237-3p	MIMAT0005592	10.90
19	hsa-miR-191-5p	MIMAT0000440	5.21
20	hsa-miR-26a-1-3p	MIMAT0004499	5.21
21	hsa-miR-1247-5p	MIMAT0005899	2.37

Supplementary Table S2.4. Contribution of individual miRNAs using MED analysis in ESCA

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-708-5p	MIMAT0004926	43.83
2	hsa-miR-199a-5p	MIMAT0000231	43.83
3	hsa-miR-33b-5p	MIMAT0003301	40.12
4	hsa-miR-23a-3p	MIMAT0000078	40.12
5	hsa-miR-423-5p	MIMAT0004748	30.25
6	hsa-miR-193b-5p	MIMAT0004767	29.01
7	hsa-miR-3677-3p	MIMAT0018101	29.01
8	hsa-miR-362-3p	MIMAT0004683	21.60

9	hsa-miR-487b-3p	MIMAT0003180	17.90
10	hsa-miR-508-3p	MIMAT0002880	16.67
11	hsa-miR-31-3p	MIMAT0004504	15.43
12	hsa-miR-33a-5p	MIMAT0000091	8.02
13	hsa-miR-30a-5p	MIMAT0000087	6.79
14	hsa-miR-335-5p	MIMAT0000765	6.79
15	hsa-miR-514a-3p	MIMAT0002883	6.79
16	hsa-miR-432-5p	MIMAT0002814	5.56
17	hsa-miR-1307-3p	MIMAT0005951	5.56
18	hsa-miR-1262	MIMAT0005914	4.32
19	hsa-miR-101-5p	MIMAT0004513	3.09
20	hsa-miR-32-3p	MIMAT0004505	3.09
21	hsa-miR-195-3p	MIMAT0004615	0.62
22	hsa-miR-483-3p	MIMAT0002173	0.62

Supplementary Table S2.5. Contribution of individual miRNAs using MED analysis in HNSC

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-93-3p	MIMAT0004509	27.14
2	hsa-miR-335-5p	MIMAT0000765	18.57
3	hsa-miR-219a-1-3p	MIMAT0004567	18.10
4	hsa-miR-361-3p	MIMAT0004682	16.19
5	hsa-miR-493-3p	MIMAT0003161	15.71
6	hsa-miR-188-3p	MIMAT0004613	14.76
7	hsa-miR-3690	MIMAT0018119	12.38
8	hsa-miR-103a-3p	MIMAT0000101	11.43
9	hsa-miR-101-5p	MIMAT0004513	10.00
10	hsa-miR-206	MIMAT0000462	9.52
11	hsa-miR-151a-5p	MIMAT0004697	9.52
12	hsa-miR-187-3p	MIMAT0000262	5.24
13	hsa-miR-7-5p	MIMAT0000252	4.29
14	hsa-miR-18a-5p	MIMAT0000072	3.81
15	hsa-miR-181a-2-3p	MIMAT0004558	3.81
16	hsa-miR-365a-5p	MIMAT0009199	3.81
17	hsa-miR-125a-3p	MIMAT0004602	3.33
18	hsa-miR-30e-5p	MIMAT0000692	1.43
19	hsa-miR-3614-3p	MIMAT0017993	1.43
20	hsa-miR-409-3p	MIMAT0001639	0.48

Supplementary Table S2.6. Contribution of individual miRNAs using MED analysis in KIRC

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-144-5p	MIMAT0004600	89.06
2	hsa-miR-744-5p	MIMAT0004945	40.63
3	hsa-miR-550a-3p	MIMAT0003257	38.28

4	hsa-miR-1296-5p	MIMAT0005794	35.16
5	hsa-let-7i-3p	MIMAT0004585	28.91
6	hsa-miR-18a-5p	MIMAT0000072	26.56
7	hsa-miR-628-5p	MIMAT0004809	19.53
8	hsa-miR-197-3p	MIMAT0000227	17.97
9	hsa-miR-222-5p	MIMAT0004569	17.19
10	hsa-miR-126-3p	MIMAT0000445	14.06
11	hsa-miR-489-3p	MIMAT0002805	14.06
12	hsa-miR-27a-3p	MIMAT0000084	12.50
13	hsa-miR-3615	MIMAT0017994	11.72
14	hsa-miR-301a-3p	MIMAT0000688	5.47
15	hsa-miR-148a-3p	MIMAT0000243	4.69
16	hsa-miR-136-3p	MIMAT0004606	4.69
17	hsa-miR-675-3p	MIMAT0006790	1.56
18	hsa-miR-34a-5p	MIMAT0000255	0.78

Supplementary Table S2.7. Contribution of individual miRNAs using MED analysis in KIRP

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-491-5p	MIMAT0002807	18.39
2	hsa-miR-769-5p	MIMAT0003886	16.09
3	hsa-miR-29c-5p	MIMAT0004673	11.49
4	hsa-miR-16-5p	MIMAT0000069	9.20
5	hsa-miR-200b-5p	MIMAT0004571	7.66
6	hsa-miR-485-3p	MIMAT0002176	7.66
7	hsa-miR-22-5p	MIMAT0004495	6.90
8	hsa-miR-629-3p	MIMAT0003298	5.36
9	hsa-miR-320b	MIMAT0005792	5.36
10	hsa-miR-217	MIMAT0000274	2.30
11	hsa-miR-224-5p	MIMAT0000281	1.53
12	hsa-miR-542-5p	MIMAT0003340	1.53

Supplementary Table S2.8. Contribution of individual miRNAs using MED analysis in LIHCC

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-550a-2	MIMAT0004800	60.91269
2	hsa-miR-549	MIMAT0003333	54.72223
3	hsa-miR-518b	MIMAT0002844	51.15079
4	hsa-miR-512-2	MIMAT0002822	50.67461
5	hsa-miR-1179	MIMAT0005824	27.73809
6	hsa-miR-574-3p	MIMAT0003239	27.02382
7	hsa-miR-424-3p	MIMAT0004749	26.62697
8	hsa-miR-4286	MIMAT0016916	24.7222
9	hsa-let-7i-3p	MIMAT0004585	24.16667
10	hsa-miR-320a	MIMAT0037311	22.97617
11	hsa-miR-17-3p	MIMAT0000071	22.81745

12	hsa-miR-299	MIMAT0000687	22.02382
13	hsa-miR-3651	MIMAT0018071	17.02382
14	hsa-miR-2277-5p	MIMAT0017352	13.76985
15	hsa-miR-621	MIMAT0003290	13.61111
16	hsa-miR-181c-5p	MIMAT0000258	13.05555
17	hsa-miR-539	MIMAT0003163	12.9762
18	hsa-miR-106b	MIMAT0000680	10.83334
19	hsa-miR-1269	MIMAT0005923	8.531742
20	hsa-miR-139	MIMAT0000250	6.706364
21	hsa-miR-152-3p	MIMAT0000438	6.626987
22	hsa-miR-2355	MIMAT0016895	3.76984
23	hsa-miR-150	MIMAT0000451	2.103168

Supplementary Table S2.9. Contribution of individual miRNAs using MED analysis in LUAD

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-671-5p	MIMAT0003880	29.65
2	hsa-miR-3651	MIMAT0018071	27.88
3	hsa-miR-1304-5p	MIMAT0005892	24.78
4	hsa-miR-342-3p	MIMAT0000753	24.34
5	hsa-miR-548v	MIMAT0015020	22.12
6	hsa-miR-296-3p	MIMAT0004679	22.12
7	hsa-miR-598-3p	MIMAT0003266	21.68
8	hsa-miR-190a-5p	MIMAT0000458	21.24
9	hsa-miR-144-5p	MIMAT0004600	16.37
10	hsa-miR-185-3p	MIMAT0004611	15.93
11	hsa-miR-181b-5p	MIMAT0000257	15.49
12	hsa-miR-3614-3p	MIMAT0017993	15.04
13	hsa-miR-126-3p	MIMAT0000445	14.16
14	hsa-miR-589-3p	MIMAT0003256	14.16
15	hsa-miR-26a-1-3p	MIMAT0004499	13.72
16	hsa-miR-148a-5p	MIMAT0004549	13.27
17	hsa-miR-3605-3p	MIMAT0017982	12.83
18	hsa-miR-627-5p	MIMAT0003296	12.83
19	hsa-miR-376c-3p	MIMAT0000720	12.39
20	hsa-miR-30b-3p	MIMAT0004589	11.50
21	hsa-miR-3200-3p	MIMAT0015085	10.18
22	hsa-miR-378a-3p	MIMAT0000732	9.29
23	hsa-miR-103a-3p	MIMAT0000101	7.96
24	hsa-miR-203a-3p	MIMAT0000264	7.96
25	hsa-miR-576-5p	MIMAT0003241	7.08
26	hsa-miR-939-5p	MIMAT0004982	2.21
27	hsa-miR-655-3p	MIMAT0003331	2.21
28	hsa-miR-582-5p	MIMAT0003247	1.77
29	hsa-miR-181a-3p	MIMAT0000270	0.44

Supplementary Table S2.10. Contribution of individual miRNAs using MED analysis in LUSC

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-181a-2-3p	MIMAT0004558	28.02
2	hsa-miR-664a-3p	MIMAT0005949	20.35
3	hsa-miR-769-3p	MIMAT0003887	19.17
4	hsa-miR-29b-2-5p	MIMAT0004515	17.40
5	hsa-miR-3614-5p	MIMAT0017992	16.22
6	hsa-miR-214-3p	MIMAT0000271	13.86
7	hsa-miR-200c-5p	MIMAT0004657	12.68
8	hsa-miR-3613-5p	MIMAT0017990	12.09
9	hsa-miR-196a-5p	MIMAT0000226	10.32
10	hsa-miR-551b-3p	MIMAT0003233	9.73
11	hsa-miR-301a-3p	MIMAT0000688	9.14
12	hsa-miR-450a-5p	MIMAT0001545	9.14
13	hsa-let-7d-5p	MIMAT0000065	7.96
14	hsa-miR-187-3p	MIMAT0000262	7.96
15	hsa-miR-20b-3p	MIMAT0004752	7.96
16	hsa-miR-887-3p	MIMAT0004951	6.19
17	hsa-miR-625-3p	MIMAT0004808	0.88
18	hsa-miR-3615	MIMAT0017994	0.29

Supplementary Table S2.11. Contribution of individual miRNAs using MED analysis in READ

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-10a-5p	MIMAT0000253	30.30
2	hsa-miR-9-5p	MIMAT0000441	30.30
3	hsa-miR-135b-3p	MIMAT0004698	27.27
4	hsa-miR-491-5p	MIMAT0002807	24.24
5	hsa-miR-31-3p	MIMAT0004504	18.18
6	hsa-miR-656-3p	MIMAT0003332	18.18
7	hsa-miR-140-5p	MIMAT0000431	15.15
8	hsa-miR-1269a	MIMAT0005923	15.15
9	hsa-miR-1468-5p	MIMAT0006789	9.09
10	hsa-miR-935	MIMAT0004978	9.09
11	hsa-miR-200b-3p	MIMAT0000318	3.03

Supplementary Table S2.12. Contribution of individual miRNAs using MED analysis in SKCM

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-211-5p	MIMAT0000268	39.07
2	hsa-miR-194-5p	MIMAT0000460	31.36

3	hsa-miR-3614-5p	MIMAT0017992	30.33
4	hsa-miR-326	MIMAT0000756	29.31
5	hsa-miR-4326	MIMAT0016888	28.79
6	hsa-miR-1180-3p	MIMAT0005825	28.79
7	hsa-miR-664a-3p	MIMAT0005949	27.25
8	hsa-miR-26a-1-3p	MIMAT0004499	21.59
9	hsa-miR-132-5p	MIMAT0004594	19.02
10	hsa-miR-1538	MIMAT0007400	16.97
11	hsa-miR-10a-5p	MIMAT0000253	15.42
12	hsa-miR-423-3p	MIMAT0001340	13.37
13	hsa-miR-338-3p	MIMAT0000763	11.83
14	hsa-miR-30c-2-3p	MIMAT0004550	11.31
15	hsa-miR-769-3p	MIMAT0003887	11.31
16	hsa-miR-2277-5p	MIMAT0017352	9.25
17	hsa-miR-16-5p	MIMAT0000069	5.66
18	hsa-miR-181d-5p	MIMAT0002821	5.14
19	hsa-miR-3648	MIMAT0018068	4.63
20	hsa-miR-589-3p	MIMAT0003256	3.60
21	hsa-miR-16-2-3p	MIMAT0004518	2.57
22	hsa-miR-29a-3p	MIMAT0000086	2.06
23	hsa-miR-24-3p	MIMAT0000080	2.06
24	hsa-miR-486-5p	MIMAT0002177	2.06
25	hsa-miR-3682-3p	MIMAT0018110	1.54
26	hsa-miR-33a-3p	MIMAT0004506	1.03
27	hsa-miR-377-5p	MIMAT0004689	0.51

Supplementary Table S2.13. Contribution of individual miRNAs using MED analysis in STAD

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-130a-3p	MIMAT0000425	34.38
2	hsa-miR-320b	MIMAT0005792	30.18
3	hsa-miR-141-5p	MIMAT0004598	29.13
4	hsa-miR-3065-5p	MIMAT0015066	25.46
5	hsa-miR-887-3p	MIMAT0004951	24.41
6	hsa-miR-362-3p	MIMAT0004683	20.73
7	hsa-miR-135a-5p	MIMAT0000428	19.69
8	hsa-miR-21-3p	MIMAT0004494	16.01
9	hsa-miR-148b-3p	MIMAT0000759	14.96
10	hsa-miR-580-3p	MIMAT0003245	12.86
11	hsa-miR-330-3p	MIMAT0000751	8.66
12	hsa-miR-143-3p	MIMAT0000435	6.56
13	hsa-miR-339-5p	MIMAT0000764	6.56
14	hsa-miR-30e-5p	MIMAT0000692	5.51
15	hsa-miR-3928-3p	MIMAT0018205	4.99
16	hsa-miR-935	MIMAT0004978	4.46
17	hsa-miR-127-5p	MIMAT0004604	3.94

18	hsa-miR-424-3p	MIMAT0004749	2.89
19	hsa-miR-221-3p	MIMAT0000278	1.84
20	hsa-miR-193a-3p	MIMAT0000459	0.26

Supplementary Table S2.14. Contribution of individual miRNAs using MED analysis in THCA

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-940	MIMAT0004983	34.80
2	hsa-let-7i-5p	MIMAT0000415	25.60
3	hsa-miR-1249-3p	MIMAT0005901	23.60
4	hsa-miR-101-5p	MIMAT0004513	23.20
5	hsa-miR-874-3p	MIMAT0004911	23.20
6	hsa-miR-3605-3p	MIMAT0017982	19.60
7	hsa-miR-592	MIMAT0003260	18.80
8	hsa-miR-188-3p	MIMAT0004613	16.40
9	hsa-miR-30d-5p	MIMAT0000245	13.60
10	hsa-miR-218-1-3p	MIMAT0004565	13.20
11	hsa-miR-23a-5p	MIMAT0004496	12.00
12	hsa-miR-324-3p	MIMAT0000762	11.20
13	hsa-miR-10a-5p	MIMAT0000253	10.40
14	hsa-miR-431-3p	MIMAT0004757	10.00
15	hsa-miR-199b-5p	MIMAT0000263	9.20
16	hsa-miR-514a-3p	MIMAT0002883	7.20
17	hsa-miR-10b-5p	MIMAT0000254	6.40
18	hsa-let-7c-5p	MIMAT0000064	5.60
19	hsa-miR-27b-5p	MIMAT0004588	5.60
20	hsa-miR-192-5p	MIMAT0000222	4.80
21	hsa-miR-382-5p	MIMAT0000737	4.40
22	hsa-miR-625-3p	MIMAT0004808	3.20
23	hsa-miR-92b-3p	MIMAT0003218	3.20
24	hsa-miR-181a-2-3p	MIMAT0004558	0.80
25	hsa-miR-3615	MIMAT0017994	0.40
26	hsa-miR-582-5p	MIMAT0003247	0.40

Supplementary Table S2.15. Contribution of individual miRNAs using MED analysis in UVM

Rank	miRNA	MIMAT-ID	MED-Score
1	hsa-miR-335-5p	MIMAT0000765	50.63
2	hsa-miR-192-3p	MIMAT0004543	45.57
3	hsa-miR-3687	MIMAT0018115	45.57
4	hsa-miR-664a-5p	MIMAT0005948	35.44
5	hsa-miR-1247-5p	MIMAT0005899	35.44
6	hsa-miR-3613-3p	MIMAT0017991	20.25
7	hsa-let-7i-3p	MIMAT0004585	20.25
8	hsa-miR-224-5p	MIMAT0000281	20.25
9	hsa-miR-3917	MIMAT0018191	20.25

10	hsa-miR-20b-3p	MIMAT0004752	15.19
11	hsa-miR-362-3p	MIMAT0004683	10.13
12	hsa-miR-581	MIMAT0003246	10.13
13	hsa-miR-222-5p	MIMAT0004569	5.06

Supplementary Table S3. Comparison of expression difference of has-let-7i-3p, has-miR-362-3p, and has-miR-3651 in cancer vs normal samples

miRNA	Cancer type	Fold change	log2(Fold change)	Mean RPM* (tumor)	Mean RPM* (normal)	p-value	adjusted p-value
hsa-let-7i-3p	Breast invasive carcinoma	-2.05	-1.04	78.73	161.63	1.21e-26	5.6e-26
	Kidney renal clear cell carcinoma	1.55	0.63	80.23	51.77	3.35e-12	8.7e-12
	Liver hepatocellular carcinoma	-1.42	-0.5	36.06	51.08	8.39e-10	3.02e-9
	Prostate adenocarcinoma	-1.54	-0.62	16.8	25.87	1.06e-8	3.56e-8
	Pan-kidney cohort (KICH+KIRC+KIRP)	1.26	0.33	60.03	47.83	0.000107	0.000158
	Uterine Corpus Endometrial Carcinoma	-1.2	-0.26	76.76	92.02	0.0022	0.00334
	Lung adenocarcinoma	-1.17	-0.23	117.97	138.33	0.0124	0.019
	Cholangiocarcinoma	1.6	0.67	53.7	33.64	0.00849	0.0219
	Lung squamous cell carcinoma	1.21	0.27	125.65	104	0.0227	0.0308
	Stomach adenocarcinoma	-1.82	-0.86	75.82	137.73	0.026	0.0386
	Bladder Urothelial Carcinoma	-1.13	-0.18	65.7	74.31	0.0328	0.0487
	Kidney renal papillary cell carcinoma	-1.33	-0.41	33.11	43.95	0.0618	0.0772
	Thyroid carcinoma	-1.09	-0.12	467.46	508.8	0.0642	0.0911
	Stomach and Esophageal carcinoma	-1.72	-0.78	66.97	115.14	0.178	0.213
	Cervical squamous cell carcinoma and	1.58	0.66	82.23	52.11	0.255	0.377

	endocervical adenocarcinoma						
	Pheochromocytoma and Paraganglioma	1.39	0.47	27.92	20.13	0.264	0.387
	Esophageal carcinoma	1.15	0.2	45.82	40.02	0.294	0.42
	Kidney Chromophobe	-1.04	-0.05	42.21	43.8	0.386	0.451
	Skin Cutaneous Melanoma	-1.58	-0.66	85.33	134.57	0.103	0.543
	Head and Neck squamous cell carcinoma	-1.05	-0.07	112.93	118.22	0.597	0.638
	Thymoma	-1.11	-0.15	69.61	77.46	0.238	0.678
	Pancreatic adenocarcinoma	-1	-0.01	76.59	76.9	0.92	0.981
hsa-miR-362-3p	Pan-kidney cohort (KICH+KIRC+KIRP)	-1.81	-0.86	2.8	5.08	7.02E-16	1.76E-15
	Liver hepatocellular carcinoma	2.33	1.22	4.23	1.82	4.02E-16	3.93E-15
	Head and Neck squamous cell carcinoma	-1.92	-0.94	2.44	4.69	7.64E-13	4.47E-12
	Lung squamous cell carcinoma	-1.67	-0.74	2.36	3.95	9.72E-12	2.99E-11
	Breast invasive carcinoma	1.94	0.96	1.63	0.84	1.98E-11	4.40E-11
	Kidney renal clear cell carcinoma	-1.67	-0.74	2.17	3.62	4.73E-09	9.64E-09
	Kidney renal papillary cell carcinoma	-2.4	-1.26	2.96	7.12	1.21E-08	3.80E-08
	Stomach adenocarcinoma	1.72	0.78	3.91	2.28	0.0000132	0.0000379
	Bladder Urothelial Carcinoma	1.76	0.82	2.83	1.6	0.0013	0.00261
	Uterine Corpus Endometrial Carcinoma	-1.4	-0.48	4.73	6.6	0.00427	0.00633

	Stomach and Esophageal carcinoma	1.31	0.39	3.77	2.88	0.00446	0.00717
	Lung adenocarcinoma	-1.22	-0.29	2.53	3.09	0.0274	0.0395
	Esophageal carcinoma	-1.4	-0.49	3.41	4.78	0.0443	0.0938
	Thyroid carcinoma	-1.1	-0.14	2.3	2.54	0.0931	0.129
	Kidney Chromophobe	1	0	5.63	5.61	0.353	0.417
	Pancreatic adenocarcinoma	-1.47	-0.56	2.44	3.59	0.121	0.438
	Thymoma	2.7	1.43	4.93	1.83	0.0376	0.509
	Skin Cutaneous Melanoma	3.8	1.93	4.69	1.23	0.12	0.574
	Pheochromocytoma and Paraganglioma	-1	0	2.51	2.51	0.459	0.59
	Prostate adenocarcinoma	-1.06	-0.08	2.32	2.45	0.603	0.663
	Cervical squamous cell carcinoma and endocervical adenocarcinoma	-1.06	-0.08	3.5	3.71	0.6	0.689
hsa-miR-3651	Lung squamous cell carcinoma	3.8	1.93	2.05	0.54	1.49E-12	4.87E-12
	Uterine Corpus Endometrial Carcinoma	3.78	1.92	2.15	0.57	5.02E-09	1.39E-08
	Stomach and Esophageal carcinoma	3.29	1.72	2.11	0.64	8.20E-08	3.21E-07
	Head and Neck squamous cell carcinoma	2.98	1.58	1.87	0.62	1.15E-07	3.23E-07
	Breast invasive carcinoma	2.18	1.12	1.35	0.62	1.42E-06	0.00000236
	Stomach adenocarcinoma	3.15	1.65	1.57	0.5	9.66E-07	0.00000342
	Kidney Chromophobe	4.68	2.23	0.47	0.1	9.91E-06	0.0000255
	Thyroid carcinoma	-1.53	-0.61	1.02	1.57	0.0114	0.0186

Esophageal carcinoma	3.11	1.64	3.38	1.09	0.0417	0.0903
Pan-kidney cohort (KICH+KIRC+KIRP)	1.62	0.7	0.44	0.27	0.211	0.235
Liver hepatocellular carcinoma	1.44	0.53	0.85	0.59	0.216	0.263
Pancreatic adenocarcinoma	-1.24	-0.31	0.27	0.34	0.498	0.757
Kidney renal papillary cell carcinoma	1.28	0.36	0.42	0.33	0.74	0.768
Thymoma	1.95	0.97	0.75	0.38	0.838	0.964

Supplementary Table S4. Co-expression analysis of miRNA signatures across 15 cancers

Cancer	BLCA-Signature	Correlated miRNAs	R
BLCA	hsa.miR.136.5p	hsa.miR.127.5p	0.85083
	hsa.miR.337.3p	hsa.miR.493.5p	0.87111
		hsa.miR.431.3p	0.86363
		hsa.miR.432.5p	0.85247
		hsa.miR.487b.3p	0.84928
		hsa.miR.654.3p	0.84251
		hsa.miR.127.5p	0.83623
		hsa.miR.409.3p	0.82585
		hsa.miR.758.3p	0.82543
		hsa.miR.379.5p	0.81228
		hsa.miR.376c.3p	0.81211
		hsa.miR.370.3p	0.80815
		hsa.miR.512.3p	hsa.miR.526b.5p
	hsa.miR.27a.3p	hsa.miR.23a.3p	0.86959
	hsa.miR.199a.5p	hsa.miR.214.5p	0.88195
	hsa.miR.526b.5p	hsa.miR.512.3p	0.87879
	hsa.miR.660.5p	hsa.miR.532.5p	0.82708
	hsa.miR.411.5p	hsa.miR.379.5p	0.90819
		hsa.miR.127.3p	0.88842
		hsa.miR.654.3p	0.84351
		hsa.miR.134.5p	0.83746
		hsa.miR.376c.3p	0.82878
		hsa.miR.381.3p	0.8251
hsa.miR.889.3p		0.8228	
hsa.miR.369.5p		0.82235	
hsa.miR.382.5p		0.81615	
hsa.miR.410.3p		0.81458	
hsa.miR.493.3p		0.8135	
hsa.miR.758.3p	0.81267		
BRCA	hsa.miR.379	hsa.miR.127	0.8525
		hsa.miR.758	0.83849
		hsa.miR.410	0.80367
hsa.miR.500a	hsa.miR.501	0.84981	
COAD	hsa.miR.450b.5p	hsa.miR.542.3p	0.86189
	hsa.miR.17.3p	hsa.miR.20a.5p	0.80638
	hsa.miR.500a.3p	hsa.miR.501.3p	0.90779
		hsa.miR.532.5p	0.86472
	hsa.miR.127.5p	hsa.miR.136.5p	0.82682
	hsa.miR.125b.2.3p	hsa.miR.99a.5p	0.8971
hsa.let.7c.5p		0.86362	
hsa.miR.125b.5p		0.81756	

ESCA	hsa.miR.191.5p	hsa.miR.425.5p	0.8532
	hsa.miR.199a.5p	hsa.miR.214.5p	0.88019
	hsa.miR.23a.3p	hsa.miR.27a.3p	0.86622
	hsa.miR.487b.3p	hsa.miR.127.3p	0.81253
		hsa.miR.889.3p	0.80128
	hsa.miR.508.3p	hsa.miR.509.3p	0.89842
		hsa.miR.514a.3p	0.8673
hsa.miR.514a.3p	hsa.miR.508.3p	0.8673	
	hsa.miR.509.3p	0.83004	
hsa.miR.432.5p	hsa.miR.431.3p	0.84021	
	hsa.miR.382.5p	0.80104	
HNSC	hsa.miR.493.3p	hsa.miR.409.3p	0.87754
		hsa.miR.382.5p	0.85962
		hsa.miR.889.3p	0.85175
		hsa.miR.758.3p	0.84649
		hsa.miR.127.3p	0.84367
		hsa.miR.493.5p	0.83876
		hsa.miR.432.5p	0.8339
		hsa.miR.379.5p	0.82919
		hsa.miR.410.3p	0.82704
		hsa.miR.370.3p	0.81859
		hsa.miR.134.5p	0.81074
		hsa.miR.431.3p	0.81054
		hsa.miR.654.3p	0.80543
		hsa.miR.206	hsa.miR.133b
	hsa.miR.1.3p		0.94774
	hsa.miR.18a.5p	hsa.miR.17.5p	0.82446
		hsa.miR.19a.3p	0.82434
	hsa.miR.409.3p	hsa.miR.382.5p	0.93486
		hsa.miR.758.3p	0.89757
		hsa.miR.134.5p	0.88559
		hsa.miR.379.5p	0.8779
		hsa.miR.493.3p	0.87754
hsa.miR.889.3p		0.87683	
hsa.miR.127.3p		0.87673	
hsa.miR.432.5p		0.87539	
hsa.miR.493.5p		0.86652	
hsa.miR.370.3p		0.85972	
hsa.miR.654.3p		0.85626	
hsa.miR.431.3p		0.84675	
hsa.miR.410.3p		0.82573	
hsa.miR.337.3p		0.81909	
hsa.miR.485.3p		0.80923	
hsa.miR.487b.3p	0.8017		

		hsa.miR.127.5p	0.80004
KIRC	hsa.miR.144.5p	hsa.miR.451a	0.94915
		hsa.miR.486.5p	0.92824
	hsa.miR.489.3p	hsa.miR.653.5p	0.81512
KIRP	hsa.miR.224.5p	hsa.miR.452.5p	0.8625
	hsa.miR.542.5p	hsa.miR.450b.5p	0.82058
LIHCC	hsa-miR-518b	hsa-miR-512	0.88368
		hsa-miR-525	0.88135
		hsa-miR-517	0.88
		hsa-miR-520a	0.87077
		hsa-miR-526b	0.86433
		hsa-miR-519a	0.85918
		hsa-miR-516a	0.84481
		hsa-miR-522	0.84263
		hsa-miR-1323	0.80711
	hsa-miR-512	hsa-miR-518b	0.87175
		hsa-miR-525	0.86591
		hsa-miR-520b	0.85993
		hsa-miR-519a	0.85658
		hsa-miR-522	0.85275
		hsa-miR-516a	0.8434
		hsa-miR-526b	0.84138
hsa-miR-517a		0.83647	
	hsa-miR-1323	0.82907	
LUAD	hsa.miR.144.5p	hsa.miR.451a	0.8937
	hsa.miR.376c.3p	hsa.miR.495.3p	0.8324
		hsa.miR.487b.3p	0.82167
		hsa.miR.381.3p	0.82055
		hsa.miR.382.5p	0.81617
		hsa.miR.654.3p	0.81027
		hsa.miR.889.3p	0.80956
		hsa.miR.493.5p	0.80489
		hsa.miR.134.5p	0.80376
		hsa.miR.379.5p	0.80133
	hsa.miR.369.3p	0.8001	
LUSC	NA	NA	NA
READ	hsa.miR.140.5p	hsa.miR.126.3p	0.82597
	hsa.miR.200b.3p	hsa.miR.429	0.85611
SKCM	hsa.miR.194.5p	hsa.miR.192.5p	0.83789
	hsa.miR.486.5p	hsa.miR.451a	0.92664
		hsa.miR.144.5p	0.80551
STAD	hsa.miR.141.5p	hsa.miR.200c.3p	0.8094
	hsa.miR.127.5p	hsa.miR.134.5p	0.89068
		hsa.miR.136.5p	0.86676

		hsa.miR.409.5p	0.83392
		hsa.miR.337.3p	0.82606
	hsa.miR.221.3p	hsa.miR.222.3p	0.89969
THCA	hsa.miR.199b.5p	hsa.miR.214.5p	0.86736
		hsa.miR.136.5p	0.84914
		hsa.miR.127.5p	0.81443
	hsa.miR.514a.3p	hsa.miR.509.3p	0.86009
		hsa.miR.508.3p	0.83453
	hsa.miR.192.5p	hsa.miR.194.5p	0.8509
		hsa.miR.409.3p	0.87451
		hsa.miR.337.3p	0.8741
		hsa.miR.134.5p	0.87061
		hsa.miR.889.3p	0.86136
		hsa.miR.381.3p	0.86114
		hsa.miR.127.5p	0.85932
		hsa.miR.127.3p	0.85734
		hsa.miR.379.5p	0.84578
		hsa.miR.136.5p	0.83969
		hsa.miR.758.3p	0.83922
		hsa.miR.136.3p	0.83552
		hsa.miR.654.3p	0.83144
		hsa.miR.409.5p	0.82336
hsa.miR.493.5p		0.82297	
hsa.miR.410.3p		0.80161	
UVM		hsa.miR.224.5p	hsa.miR.452.5p

Abbreviations: NA- Not Available.

Supplementary Table S5. Top-10 ranked miRNAs involvement in BLCA

Rank	BLCA	Literature
Rank-1	hsa-miR-31-3p	1
Rank-2	hsa-miR-29b-2-5p	2, 3
Rank-3	hsa-miR-193b-3p	4, 5
Rank-4	hsa-miR-10b-5p	6
Rank-5	hsa-miR-125a-3p	7, 8
Rank-6	hsa-miR-136-5p	-
Rank-7	hsa-miR-3912-3p	-
Rank-8	hsa-miR-205-5p	9, 10
Rank-9	hsa-miR-337-3p	11
Rank-10	hsa-miR-29a-5p	12

Supplementary Table S6. Top-10 ranked miRNAs involvement in BRCA

Rank	BRCA	Literature
1	hsa-miR-200c-5p	13, 14
2	hsa-miR-503	15
3	hsa-miR-1307-3p	16
4	hsa-miR-361-3p	17
5	hsa-miR-212	18
6	hsa-miR-592	19
7	hsa-miR-1185-1	-
8	hsa-miR-146b	20
9	hsa-miR-1468-5p	-
10	hsa-miR-769-3p	21

Supplementary Table S7. Top-10 ranked miRNAs involvement in COAD

Rank	COAD	Literature
1	hsa-miR-188-3p	22
2	hsa-miR-1976	-
3	hsa-miR-320a	23
4	hsa-miR-450b-5p	24
5	hsa-miR-140-3p	25
6	hsa-miR-17-3p	26
7	hsa-miR-301a-3p	27
8	hsa-miR-582-5p	-
9	hsa-miR-491-3p	28
10	hsa-miR-501-5p	-

Supplementary Table S8. Top-10 ranked miRNAs involvement in ESCA

Rank	ESCA	Literature
1	hsa-miR-708-5p	29, 30
2	hsa-miR-199a-5p	31, 32
3	hsa-miR-33b-5p	33, 34
4	hsa-miR-23a-3p	35
5	hsa-miR-423-5p	36
6	hsa-miR-193b-5p	37
7	hsa-miR-3677-3p	38
8	hsa-miR-362-3p	39
9	hsa-miR-487b-3p	-
10	hsa-miR-508-3p	40

Supplementary Table S9. Top-10 ranked miRNAs involvement in HNSC

Rank	HNSC	Literature
1	hsa-miR-93-3p	41, 42
2	hsa-miR-335-5p	43
3	hsa-miR-219a-1-3p	-
4	hsa-miR-361-3p	-
5	hsa-miR-493-3p	44
6	hsa-miR-188-3p	-
7	hsa-miR-3690	45
8	hsa-miR-103a-3p	46
9	hsa-miR-101-5p	47
10	hsa-miR-206	48, 49

Supplementary Table S10. Top-10 ranked miRNAs involvement in KIRC

Rank	KIRC	Literature
1	hsa-miR-144-5p	50
2	hsa-miR-744-5p	-
3	hsa-miR-550a-3p	-
4	hsa-miR-1296-5p	-
5	hsa-let-7i-3p	51
6	hsa-miR-18a-5p	52

7	hsa-miR-628-5p	-
8	hsa-miR-197-3p	51
9	hsa-miR-222-5p	53
10	hsa-miR-126-3p	54, 55

Supplementary Table S11. Top-10 ranked miRNAs involvement in KIRP

Rank	KIRP	Literature
1	hsa-miR-491-5p	-
2	hsa-miR-769-5p	56
3	hsa-miR-29c-5p	-
4	hsa-miR-16-5p	57
5	hsa-miR-200b-5p	58
6	hsa-miR-485-3p	59
7	hsa-miR-22-5p	58
8	hsa-miR-629-3p	-
9	hsa-miR-320b	57
10	hsa-miR-217	60

Supplementary Table S12. Top-10 ranked miRNAs involvement in LIHCC

Rank	LIHCC	Literature
1	hsa-miR-550a-2	61
2	hsa-miR-549	-
3	hsa-miR-518b	62
4	hsa-miR-512-2	63
5	hsa-miR-1179	64
6	hsa-miR-574-3p	65
7	hsa-miR-424-3p	66
8	hsa-miR-4286	67
9	hsa-let-7i-3p	68
10	hsa-miR-320a	69

Supplementary Table S13. Top-10 ranked miRNAs involvement in LUAD

Rank	LUAD	Literature
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1	hsa-miR-671-5p	70
2	hsa-miR-3651	71
3	hsa-miR-1304-5p	72
4	hsa-miR-342-3p	73
5	hsa-miR-548v	74
6	hsa-miR-296-3p	71
7	hsa-miR-598-3p	75
8	hsa-miR-190a-5p	-
9	hsa-miR-144-5p	76
10	hsa-miR-185-3p	76

Supplementary Table S14. Top-10 ranked miRNAs involvement in LUSC

Rank	LUSC	Literature
1	hsa-miR-181a-2-3p	77
2	hsa-miR-664a-3p	-
3	hsa-miR-769-3p	78
4	hsa-miR-29b-2-5p	79
5	hsa-miR-3614-5p	80
6	hsa-miR-214-3p	81
7	hsa-miR-200c-5p	82, 83
8	hsa-miR-3613-5p	84
9	hsa-miR-196a-5p	85
10	hsa-miR-551b-3p	86

Supplementary Table S15. Top-10 ranked miRNAs involvement in SKCM

Rank	SKCM	Literature
1	hsa-miR-211-5p	87
2	hsa-miR-194-5p	88
3	hsa-miR-3614-5p	-
4	hsa-miR-326	-
5	hsa-miR-4326	-
6	hsa-miR-1180-3p	89
7	hsa-miR-664a-3p	90
8	hsa-miR-26a-1-3p	91
9	hsa-miR-132-5p	92

10	hsa-miR-1538	93
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Supplementary Table S16. Top-10 ranked miRNAs involvement in STAD

Rank	STAD	Literature
1	hsa-miR-130a-3p	94
2	hsa-miR-320b	95, 96
3	hsa-miR-141-5p	97
4	hsa-miR-3065-5p	98
5	hsa-miR-887-3p	-
6	hsa-miR-362-3p	99
7	hsa-miR-135a-5p	100
8	hsa-miR-21-3p	101, 102
9	hsa-miR-148b-3p	103
10	hsa-miR-580-3p	-

Supplementary Table S17. Top-10 ranked miRNAs involvement in THCA

Rank	THCA	Literature
1	hsa-miR-940	104
2	hsa-let-7i-5p	105
3	hsa-miR-1249-3p	106
4	hsa-miR-101-5p	107
5	hsa-miR-874-3p	108
6	hsa-miR-3605-3p	-
7	hsa-miR-592	-
8	hsa-miR-188-3p	109
9	hsa-miR-30d-5p	110
10	hsa-miR-218-1-3p	111

Supplementary Table S18. Top-10 ranked miRNAs involvement in READ

Rank	READ	Literature
1	hsa-miR-10a-5p	112
2	hsa-miR-9-5p	113
3	hsa-miR-135b-3p	114
4	hsa-miR-491-5p	115

5	hsa-miR-31-3p	116
6	hsa-miR-656-3p	117
7	hsa-miR-140-5p	118
8	hsa-miR-1269a	119
9	hsa-miR-1468-5p	120
10	hsa-miR-935	-

Supplementary Table S19. Top-10 ranked miRNAs involvement in UVM

Rank	UVM	Literature
1	hsa-miR-335-5p	121
2	hsa-miR-192-3p	122
3	hsa-miR-3687	-
4	hsa-miR-664a-5p	121
5	hsa-miR-1247-5p	123
6	hsa-miR-3613-3p	-
7	hsa-let-7i-3p	-
8	hsa-miR-224-5p	124
9	hsa-miR-3917	-
10	hsa-miR-20b-3p	-

Supplementary Table S20. MiRNAs that were not previously reported in cancers

Cancer type	miRNA previously not reported	Rank of the miRNA in the signature
BLCA	hsa-miR-136-5p	6
	hsa-miR-3912-3p	7
BRCA	hsa-miR-1185-1	7
	hsa-miR-1468-5p	9
COAD	hsa-miR-1976	2
	hsa-miR-582-5p	8
	hsa-miR-501-5p	10
ESCA	hsa-miR-487b-3p	9
HNSC	hsa-miR-219a-1-3p	3
	hsa-miR-361-3p	4
	hsa-miR-188-3p	6
KIRC	hsa-miR-744-5p	3
	hsa-miR-550a-3p	4
	hsa-miR-1296-5p	5
	hsa-miR-628-5p	7
KIRP	hsa-miR-491-5p	1

	hsa-miR-29c-5p	3
	hsa-miR-629-3p	8
LIHCC	hsa-miR-549	2
LUAD	hsa-miR-190a-5p	8
LUSC	hsa-miR-664a-3p	2
SKCM	hsa-miR-3614-5p	3
	hsa-miR-326	4
	hsa-miR-4326	5
STAD	hsa-miR-887-3p	5
	hsa-miR-580-3p	10
THCA	hsa-miR-3605-3p	6
	hsa-miR-592	7
READ	hsa-miR-935	10
UVM	hsa-miR-3687	3
	hsa-miR-3613-3p	6
	hsa-let-7i-3p	7
	hsa-miR-3917	9
	hsa-miR-20b-3p	10

Supplementary Table S21. KEGG pathway analysis of miRNA signatures across 15 cancers

Cancer	KEGG pathway	p-value
BLCA	Proteoglycans in cancer	5.55E-11
	Glioma	1.08E-06
	ECM-receptor interaction	1.66E-06
	ErbB signaling pathway	2.95E-06
	Axon guidance	8.06E-06
	Signaling pathways regulating pluripotency of stem cells	8.53E-06
	Phosphatidylinositol signaling system	1.6E-05
	Renal cell carcinoma	6.8E-05
	Prion diseases	0.00011
	Neurotrophin signaling pathway	0.00015
	Acute myeloid leukemia	0.00057
	mTOR signaling pathway	0.00084
	TGF-beta signaling pathway	0.00084
	Pathways in cancer	0.00084
	Ras signaling pathway	0.00093
	Hippo signaling pathway	0.00093
	Focal adhesion	0.00093
	Colorectal cancer	0.00123
	Endometrial cancer	0.00134
	Rap1 signaling pathway	0.00178

	Choline metabolism in cancer	0.00246
	Prolactin signaling pathway	0.00248
	Circadian rhythm	0.00307
	Gap junction	0.00324
	Prostate cancer	0.00324
	Wnt signaling pathway	0.00324
	T cell receptor signaling pathway	0.004
	Melanoma	0.00468
	AMPK signaling pathway	0.00468
BRCA	KEGG pathway	p-value
	Mucin type O-Glycan biosynthesis	3.02E-15
	Proteoglycans in cancer	6.30E-13
	Hippo signaling pathway	2.01E-09
	ECM-receptor interaction	4.29E-09
	Signaling pathways regulating pluripotency of stem cells	6.02E-07
	FoxO signaling pathway	1.03E-05
	TGF-beta signaling pathway	1.25E-05
	Renal cell carcinoma	1.34E-05
	Focal adhesion	1.51E-05
	Axon guidance	1.88E-05
	Adherens junction	7.60E-05
	Wnt signaling pathway	7.60E-05
	Rap1 signaling pathway	8.12E-05
	Prion diseases	9.47E-05
	Glioma	0.00012
	Ras signaling pathway	0.00032
	MAPK signaling pathway	0.00037
	Thyroid hormone signaling pathway	0.00107
	Endocytosis	0.00144
	Pathways in cancer	0.00161
	Circadian rhythm	0.00212
	Choline metabolism in cancer	0.00212
	ErbB signaling pathway	0.00212
	Melanoma	0.0032
	Bacterial invasion of epithelial cells	0.00377
COAD	Morphine addiction	2.04E-07
	Signaling pathways regulating pluripotency of stem cells	5.03E-07
	Prion diseases	2.95E-06
	Proteoglycans in cancer	2.95E-06

	Glutamatergic synapse	6E-06
	Axon guidance	6E-06
	Transcriptional misregulation in cancer	1.8E-05
	Adrenergic signaling in cardiomyocytes	2.9E-05
	Hippo signaling pathway	0.00017
	Wnt signaling pathway	0.00027
	Long-term depression	0.00034
	Glioma	0.00034
	FoxO signaling pathway	0.00044
	Amphetamine addiction	0.00054
	Circadian rhythm	0.00063
	Estrogen signaling pathway	0.00064
	cAMP signaling pathway	0.00075
	Oxytocin signaling pathway	0.00075
	Focal adhesion	0.00085
	TGF-beta signaling pathway	0.00089
	Retrograde endocannabinoid signaling	0.00106
	Regulation of actin cytoskeleton	0.00167
	Renal cell carcinoma	0.00257
	Colorectal cancer	0.00305
	Dopaminergic synapse	0.00399
ESCA	Axon guidance	3.30E-05
	FoxO signaling pathway	3.30E-05
	Glioma	3.30E-05
	Long-term depression	5.83E-05
	GABAergic synapse	8.00E-05
	Glutamatergic synapse	0.00021
	Non-small cell lung cancer	0.00035
	Renal cell carcinoma	0.00035
	Phosphatidylinositol signaling system	0.00056
	Gap junction	0.00056
	Aldosterone-regulated sodium reabsorption	0.0007
	Morphine addiction	0.00112
	TGF-beta signaling pathway	0.0013
	Retrograde endocannabinoid signaling	0.0013
	Prolactin signaling pathway	0.00138
	ErbB signaling pathway	0.00138
	Circadian rhythm	0.00154
	Thyroid hormone signaling pathway	0.00154
	Signaling pathways regulating pluripotency of stem cells	0.00154

	Proteoglycans in cancer	0.00195
	Oxytocin signaling pathway	0.00251
	Adrenergic signaling in cardiomyocytes	0.00295
	Estrogen signaling pathway	0.00295
	Platelet activation	0.00342
	MAPK signaling pathway	0.00387
	Melanoma	0.00405
	Ras signaling pathway	0.0041
	Long-term potentiation	0.00435
HNSC	Prion diseases	9.28E-08
	Proteoglycans in cancer	9.28E-08
	Fatty acid biosynthesis	1.22E-06
	cGMP-PKG signaling pathway	0.00022
	Adherens junction	0.00038
	Adrenergic signaling in cardiomyocytes	0.00042
	Gap junction	0.00068
	AMPK signaling pathway	0.00068
	Rap1 signaling pathway	0.00068
	Signaling pathways regulating pluripotency of stem cells	0.00072
	GABAergic synapse	0.00186
	Hippo signaling pathway	0.00212
	Morphine addiction	0.00237
	Circadian entrainment	0.00237
	Oxytocin signaling pathway	0.00237
	Vascular smooth muscle contraction	0.00266
	Ubiquitin mediated proteolysis	0.00312
	Lysine degradation	0.00312
	Ras signaling pathway	0.00312
	Axon guidance	0.00392
	Pancreatic cancer	0.00467
KIRC	Prion diseases	1.98E-10
	Proteoglycans in cancer	7.53E-07
	Glioma	2.30E-06
	ErbB signaling pathway	5.38E-06
	Ras signaling pathway	6.23E-05
	ECM-receptor interaction	0.00123
	Amphetamine addiction	0.00129
	Focal adhesion	0.00129
	TGF-beta signaling pathway	0.00159
	Renal cell carcinoma	0.00159

	Rap1 signaling pathway	0.00159
	Gap junction	0.00214
	Estrogen signaling pathway	0.00214
	Long-term potentiation	0.00214
	PI3K-Akt signaling pathway	0.00214
	Axon guidance	0.00236
	Lysine degradation	0.00248
	Phosphatidylinositol signaling system	0.0027
	Dorso-ventral axis formation	0.00276
	Choline metabolism in cancer	0.00336
	Mucin type O-Glycan biosynthesis	0.00381
	Signaling pathways regulating pluripotency of stem cells	0.00442
KIRP	Fatty acid biosynthesis	1.27E-17
	Signaling pathways regulating pluripotency of stem cells	1.67E-09
	Proteoglycans in cancer	1.21E-08
	Prolactin signaling pathway	1.73E-06
	Hippo signaling pathway	1.81E-06
	Prostate cancer	0.00011
	FoxO signaling pathway	0.00013
	Rap1 signaling pathway	0.00017
	Melanoma	0.00026
	Pathways in cancer	0.0003
	Long-term depression	0.00035
	PI3K-Akt signaling pathway	0.00056
	Endometrial cancer	0.00066
	Glioma	0.00066
	Thyroid cancer	0.00084
	Colorectal cancer	0.00092
	Estrogen signaling pathway	0.00114
	TGF-beta signaling pathway	0.00208
	Non-small cell lung cancer	0.00227
	AMPK signaling pathway	0.00463
LIHCC	Fatty acid biosynthesis	2.61E-12
	ECM-receptor interaction	1.63E-11
	Fatty acid metabolism	2.79E-11
	Hepatitis B	3.58E-09
	Glioma	2.90E-08
	Proteoglycans in cancer	1.11E-07
	Lysine degradation	3.32E-07

	Hippo signaling pathway	1.95E-06
	Pathways in cancer	4.2E-06
	Viral carcinogenesis	5.37E-06
	TGF-beta signaling pathway	9.14E-06
	Estrogen signaling pathway	1.7E-05
	Chronic myeloid leukemia	3.9E-05
	Prostate cancer	0.00011
	Renal cell carcinoma	0.00015
	Adherens junction	0.00076
	Fatty acid elongation	0.00081
	Prion diseases	0.00139
	Endocytosis	0.00141
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LUAD	Fatty acid biosynthesis	7.47E-10
	Axon guidance	9.10E-08
	Signaling pathways regulating pluripotency of stem cells	6.07E-06
	TGF-beta signaling pathway	9.26E-06
	Proteoglycans in cancer	9.26E-06
	Amphetamine addiction	4.99E-05
	Pathways in cancer	4.99E-05
	Hippo signaling pathway	9.75E-05
	Transcriptional misregulation in cancer	9.75E-05
	Thyroid hormone signaling pathway	0.00046
	Ubiquitin mediated proteolysis	0.00053
	N-Glycan biosynthesis	0.00209
	Glycosphingolipid biosynthesis - ganglio series	0.00221
	Adherens junction	0.00221
	Circadian rhythm	0.0023
	Rap1 signaling pathway	0.0027
	Arrhythmogenic right ventricular cardiomyopathy (ARVC)	0.00389
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LUSC	Glycosphingolipid biosynthesis - lacto and neolacto series	7.01E-13
	Prion diseases	1.70E-06
	Proteoglycans in cancer	2.28E-06
	TGF-beta signaling pathway	2.92E-06
	Mucin type O-Glycan biosynthesis	1.35E-05
	Signaling pathways regulating pluripotency of stem cells	1.35E-05
	ErbB signaling pathway	2.03E-05
	mTOR signaling pathway	6.97E-05

	FoxO signaling pathway	0.0009
READ	GABAergic synapse	1.25E-06
	Thyroid hormone signaling pathway	1.25E-06
	Estrogen signaling pathway	1.25E-06
	Axon guidance	5.97E-05
	Proteoglycans in cancer	5.97E-05
	ErbB signaling pathway	0.00033
	Prolactin signaling pathway	0.00033
	Renal cell carcinoma	0.00033
	Adrenergic signaling in cardiomyocytes	0.00037
	Morphine addiction	0.0012
	Ras signaling pathway	0.00208
	Phosphatidylinositol signaling system	0.0021
	Cocaine addiction	0.00238
	Pancreatic cancer	0.00238
	Signaling pathways regulating pluripotency of stem cells	0.00238
	Mucin type O-Glycan biosynthesis	0.0027
	Rap1 signaling pathway	0.00374
	Biotin metabolism	0.00421
	Long-term depression	0.00421
	Focal adhesion	0.00421
	Pathways in cancer	0.00421
	Neurotrophin signaling pathway	0.00483
	Oxytocin signaling pathway	0.00483
SKCM	Fatty acid biosynthesis	6.12E-11
	Fatty acid metabolism	6.12E-11
	ECM-receptor interaction	6.12E-11
	Proteoglycans in cancer	1.53E-07
	Signaling pathways regulating pluripotency of stem cells	1.3E-05
	FoxO signaling pathway	8.6E-05
	Glioma	0.0001
	ErbB signaling pathway	0.00014
	Endometrial cancer	0.00014
	Hippo signaling pathway	0.00037
	Pathways in cancer	0.00041
	Focal adhesion	0.00052
	Thyroid hormone signaling pathway	0.00075
	PI3K-Akt signaling pathway	0.00075
	mTOR signaling pathway	0.00084

	Colorectal cancer	0.00086
	Long-term depression	0.00086
	Wnt signaling pathway	0.00086
	Melanoma	0.00086
	Choline metabolism in cancer	0.00123
	Arrhythmogenic right ventricular cardiomyopathy (ARVC)	0.00178
	Adrenergic signaling in cardiomyocytes	0.00266
	TGF-beta signaling pathway	0.0029
	Prostate cancer	0.00298
	Thyroid cancer	0.00373
	Neurotrophin signaling pathway	0.00475
	Oxytocin signaling pathway	0.00475
STAD	ECM-receptor interaction	5.52E-10
	Renal cell carcinoma	6.37E-07
	Ras signaling pathway	7.78E-07
	Axon guidance	7.78E-07
	Pathways in cancer	8.76E-07
	Proteoglycans in cancer	3.33E-06
	Prion diseases	7.68E-06
	Adherens junction	7.68E-06
	Long-term depression	9.64E-06
	ErbB signaling pathway	9.64E-06
	Hippo signaling pathway	1.21E-05
	FoxO signaling pathway	1.67E-05
	TGF-beta signaling pathway	1.76E-05
	Focal adhesion	1.76E-05
	Glioma	5.42E-05
	Choline metabolism in cancer	0.00019
	Chronic myeloid leukemia	0.00023
	Signaling pathways regulating pluripotency of stem cells	0.00023
	Prolactin signaling pathway	0.00023
	Colorectal cancer	0.00028
	Mucin type O-Glycan biosynthesis	0.00046
	Pancreatic cancer	0.00046
	Endocrine and other factor-regulated calcium reabsorption	0.00049
	Melanoma	0.00053
	PI3K-Akt signaling pathway	0.0006
	Rap1 signaling pathway	0.00118

	Circadian rhythm	0.00174
	Thyroid hormone signaling pathway	0.0022
	Regulation of actin cytoskeleton	0.00398
	Prostate cancer	0.00398
	cGMP-PKG signaling pathway	0.00497
THCA	Prion diseases	1.49E-07
	Proteoglycans in cancer	1.48E-06
	Axon guidance	6.31E-06
	Glutamatergic synapse	6.63E-05
	Signaling pathways regulating pluripotency of stem cells	0.00016
	Long-term depression	0.0002
	Morphine addiction	0.0002
	ErbB signaling pathway	0.0002
	FoxO signaling pathway	0.00025
	TGF-beta signaling pathway	0.00033
	Transcriptional misregulation in cancer	0.00038
	Chronic myeloid leukemia	0.00099
	Ubiquitin mediated proteolysis	0.00218
	Retrograde endocannabinoid signaling	0.00242
	Renal cell carcinoma	0.0026
	cGMP-PKG signaling pathway	0.00265
	Regulation of actin cytoskeleton	0.00265
	Glioma	0.00273
	Glycosaminoglycan biosynthesis - keratan sulfate	0.00433
UVM	Hippo signaling pathway	5.37E-12
	Signaling pathways regulating pluripotency of stem cells	9.55E-11
	Pathways in cancer	2.24E-07
	Ubiquitin mediated proteolysis	3.48E-06
	Thyroid hormone signaling pathway	3.48E-06
	Transcriptional misregulation in cancer	3.48E-06
	Proteoglycans in cancer	3.48E-06
	Renal cell carcinoma	4.88E-06
	Wnt signaling pathway	1.03E-05
	Axon guidance	1.47E-05
	FoxO signaling pathway	1.58E-05
	Phosphatidylinositol signaling system	4.56E-05
	Adherens junction	0.00012
	Endocrine and other factor-regulated calcium reabsorption	0.00014

Circadian rhythm	0.00015
TGF-beta signaling pathway	0.0004
Rap1 signaling pathway	0.0004
AMPK signaling pathway	0.00071
Long-term depression	0.00103
Ras signaling pathway	0.00165
Colorectal cancer	0.00188
mRNA surveillance pathway	0.002
Morphine addiction	0.00202
MAPK signaling pathway	0.00304
Sphingolipid signaling pathway	0.00331
Cholinergic synapse	0.00485
Protein processing in endoplasmic reticulum	0.00591

Supplementary Table S22. GO category analysis of miRNA signatures across 15 cancers

Cancer	GO Category	genes	miRNAs	p-value
BLCA	cellular_component	3963	9	7.55E-19
	protein binding transcription factor activity	173	10	8.66E-18
	protein complex	1074	10	3.84E-17
	epidermal growth factor receptor signaling pathway	95	12	4.88E-17
	enzyme binding	457	12	5.25E-17
	molecular_function	5054	14	7.98E-17
	gene expression	217	14	2.00E-16
	Fc-epsilon receptor signaling pathway	91	15	2.60E-16
	nucleic acid binding transcription factor activity	379	16	5.40E-16
	neurotrophin TRK receptor signaling pathway	127	16	6.64E-16
	cellular protein modification process	927	19	8.01E-16
	biosynthetic process	1518	23	8.67E-16
	cellular nitrogen compound metabolic process	1839	23	9.21E-16
	ion binding	2413	23	9.50E-16
	organelle	3878	28	1.06E-15
	cytosol	790	11	1.67E-15
	nucleoplasm	336	9	1.37E-12
	transcription, DNA-templated	458	7	1.07E-11
	cellular component assembly	349	7	1.16E-09
	fibroblast growth factor receptor signaling pathway	71	9	1.02E-08
	symbiosis, encompassing mutualism through parasitism	130	5	3.18E-08
	small molecule metabolic process	541	8	9.61E-08
	biological_process	3279	7	9.70E-08

	viral process	119	5	1.29E-07
	catabolic process	335	4	2.13E-07
	cytoskeletal protein binding	200	7	2.00E-06
	cell death	237	6	5.99E-06
	blood coagulation	118	5	7.45E-06
	phosphatidylinositol-mediated signaling	51	8	7.85E-06
	response to stress	433	6	3.51E-05
	mitotic cell cycle	69	3	7.20E-05
	macromolecular complex assembly	217	6	0.00022
	synaptic transmission	93	4	0.004
	toll-like receptor 10 signaling pathway	26	4	0.00438
	nucleobase-containing compound catabolic process	150	3	0.00486
BRCA	mitotic cell cycle	87	5	<1E-325
	cellular protein modification process	329	5	<1E-325
	biological_process	1365	5	<1E-325
	viral process	108	5	<1E-325
	small molecule metabolic process	287	5	<1E-325
	symbiosis, encompassing mutualism through parasitism	119	5	<1E-325
	membrane organization	113	5	<1E-325
	biosynthetic process	528	6	<1E-325
	gene expression	157	6	<1E-325
	cellular nitrogen compound metabolic process	684	8	<1E-326
	catabolic process	225	3	8.33E-15
	cellular component assembly	190	7	5.48E-14
	response to stress	242	3	6.43E-14
	macromolecular complex assembly	133	6	4.60E-12
	nucleobase-containing compound catabolic process	131	5	1.16E-11
	mRNA metabolic process	52	4	1.37E-11
	RNA metabolic process	54	4	5.92E-11
	neurotrophin TRK receptor signaling pathway	43	3	1.04E-10
	cellular protein metabolic process	73	5	4.98E-09
	cellular lipid metabolic process	30	3	3.64E-07
	Fc-epsilon receptor signaling pathway	29	4	4.47E-07
	protein complex assembly	101	5	5.38E-07
	DNA metabolic process	93	3	1.52E-05
	cell death	123	4	2.56E-05
	viral life cycle	24	4	5.19E-05
	transcription, DNA-templated	119	2	0.00113
	epidermal growth factor receptor signaling pathway	25	2	0.00223

	transcription initiation from RNA polymerase II promoter	21	2	0.00406
	G2/M transition of mitotic cell cycle	23	2	0.00438
COAD	biological_process	3548	7	6.51E-22
	nucleoplasm	316	8	8.67E-22
	neurotrophin TRK receptor signaling pathway	83	8	3.40E-20
	nucleic acid binding transcription factor activity	317	9	8.46E-20
	cellular_component	3941	9	9.60E-20
	transcription, DNA-templated	611	9	5.71E-18
	enzyme binding	401	9	9.21E-18
	protein binding transcription factor activity	180	10	2.57E-17
	molecular_function	4155	11	5.57E-17
	protein complex	1102	11	2.99E-16
	gene expression	219	12	4.78E-16
	cellular protein modification process	766	13	5.09E-16
	biosynthetic process	1364	15	6.27E-16
	ion binding	1982	16	6.84E-16
	cellular nitrogen compound metabolic process	1696	18	8.43E-15
	organelle	3396	19	6.10E-14
	mitotic cell cycle	104	7	8.20E-14
	cellular component assembly	244	5	3.71E-13
	cytosol	538	5	2.85E-12
	Fc-epsilon receptor signaling pathway	52	7	3.27E-12
	cytoskeletal protein binding	218	8	5.12E-12
	catabolic process	492	9	6.00E-12
	macromolecular complex assembly	196	7	2.70E-10
	blood coagulation	103	4	4.08E-09
	nucleobase-containing compound catabolic process	198	6	6.11E-09
	epidermal growth factor receptor signaling pathway	66	6	1.15E-08
	RNA binding	399	7	3.38E-08
	small molecule metabolic process	430	6	3.94E-07
	symbiosis, encompassing mutualism through parasitism	130	5	2.19E-06
	viral process	113	5	4.88E-06
	protein complex assembly	133	5	3.17E-05
	synaptic transmission	89	4	0.0001
	fibroblast growth factor receptor signaling pathway	53	6	0.00025
	enzyme regulator activity	169	4	0.00033
	cell death	174	4	0.00068
	regulation of transcription, DNA-templated	62	1	0.00069
	microtubule organizing center	90	4	0.00104

	response to stress	357	4	0.00317
	nucleic acid binding	54	1	0.0032
	cell-cell signaling	103	3	0.00486
ESCA	cellular_component	3045	7	2.89E-23
	molecular_function	3396	8	6.40E-22
	nucleic acid binding transcription factor activity	291	9	1.99E-20
	gene expression	159	9	4.53E-18
	neurotrophin TRK receptor signaling pathway	96	10	6.16E-18
	cellular protein modification process	676	11	9.60E-18
	biosynthetic process	1067	12	5.57E-17
	ion binding	1681	12	5.69E-17
	cellular nitrogen compound metabolic process	1430	16	2.65E-16
	organelle	2841	16	3.32E-16
	transcription, DNA-templated	375	5	5.55E-16
	enzyme binding	328	7	1.48E-14
	Fc-epsilon receptor signaling pathway	50	4	6.93E-12
	protein binding transcription factor activity	131	7	3.20E-08
	protein complex	600	4	3.97E-08
	epidermal growth factor receptor signaling pathway	65	4	5.19E-08
	biological_process	2825	6	4.89E-07
	nucleoplasm	270	6	4.94E-07
	catabolic process	275	4	1.30E-06
	blood coagulation	120	7	1.60E-06
	synaptic transmission	107	6	1.65E-06
	cellular component assembly	248	4	2.02E-05
	small molecule metabolic process	299	4	3.66E-05
	cytosol	370	3	0.00036
	fibroblast growth factor receptor signaling pathway	54	5	0.00097
	enzyme regulator activity	120	2	0.00131
	post-translational protein modification	36	2	0.00211
	cell death	151	3	0.00271
	cytoskeletal protein binding	156	4	0.00433
HNSC	nucleic acid binding transcription factor activity	240	7	2.62E-23
	Fc-epsilon receptor signaling pathway	63	7	5.22E-23
	neurotrophin TRK receptor signaling pathway	91	8	5.36E-21
	gene expression	169	9	3.11E-18
	molecular_function	3534	10	4.07E-18
	cellular protein modification process	647	11	9.45E-19
	biosynthetic process	983	11	3.62E-17
	ion binding	1559	12	1.93E-17

organelle	2495	12	2.51E-16
cellular nitrogen compound metabolic process	1231	13	1.80E-16
cellular_component	3144	8	1.11E-16
enzyme binding	322	8	1.11E-16
protein complex	760	6	8.88E-16
epidermal growth factor receptor signaling pathway	72	8	5.02E-14
blood coagulation	134	8	2.18E-13
cytosol	509	6	3.68E-12
transcription, DNA-templated	117	2	1.53E-11
protein binding transcription factor activity	140	7	3.05E-11
catabolic process	321	4	5.96E-09
nucleoplasm	174	3	2.63E-08
fibroblast growth factor receptor signaling pathway	59	6	1.02E-07
symbiosis, encompassing mutualism through parasitism	118	5	2.09E-07
nucleobase-containing compound catabolic process	144	3	1.59E-06
small molecule metabolic process	264	4	2.91E-06
viral process	106	5	3.21E-06
cellular component assembly	186	3	2.09E-05
biological_process	1946	3	2.22E-05
synaptic transmission	87	5	2.38E-05
platelet activation	50	4	0.00014
phosphatidylinositol-mediated signaling	34	4	0.00045
regulation of transcription, DNA-templated	62	1	0.00051
cytoskeletal protein binding	93	2	0.00122
cellular protein metabolic process	69	3	0.00323
macromolecular complex assembly	97	2	0.00348
response to stress	180	3	0.00437
nucleic acid binding	54	1	0.00449
KIRC molecular_function	2909	7	1.56E-24
cellular_component	2662	7	2.69E-23
nucleic acid binding transcription factor activity	231	8	1.48E-20
cellular protein modification process	538	8	4.75E-20
biosynthetic process	837	10	3.86E-19
cellular nitrogen compound metabolic process	1028	11	2.96E-18
ion binding	1328	11	4.00E-17
organelle	2049	13	6.27E-18
Fc-epsilon receptor signaling pathway	55	5	2.22E-16
neurotrophin TRK receptor signaling pathway	68	5	4.44E-16
epidermal growth factor receptor signaling pathway	65	5	2.44E-15
gene expression	131	6	1.20E-12

	protein binding transcription factor activity	115	6	2.74E-10
	protein complex	500	4	1.12E-08
	enzyme binding	212	5	1.59E-07
	catabolic process	179	3	2.16E-07
	synaptic transmission	80	4	4.43E-07
	nervous system development	23	2	5.09E-07
	transcription, DNA-templated	337	5	9.21E-07
	biological_process	2416	5	1.53E-06
	fibroblast growth factor receptor signaling pathway	53	5	3.33E-06
	cytosol	380	4	4.32E-06
	homophilic cell adhesion via plasma membrane adhesion molecules	19	2	3.18E-05
	cellular component assembly	163	3	8.19E-05
	axon guidance	90	3	0.00013
	cell-cell signaling	115	4	0.00014
	blood coagulation	77	4	0.00026
	symbiosis, encompassing mutualism through parasitism	77	3	0.00032
	small molecule metabolic process	303	4	0.00085
	nucleoplasm	165	3	0.00129
	macromolecular complex assembly	129	4	0.00132
	viral process	66	3	0.00178
	phosphatidylinositol-mediated signaling	31	4	0.00209
	response to stress	235	2	0.00241
	cell death	95	2	0.00849
	cell adhesion	32	2	0.02661
	nucleobase-containing compound catabolic process	63	2	0.03911
	protein complex assembly	65	1	0.04679
	enzyme regulator activity	69	2	0.04702
KIRP	molecular_function	2322	5	1.27E-20
	neurotrophin TRK receptor signaling pathway	76	6	3.07E-20
	cellular protein modification process	515	8	2.36E-19
	gene expression	160	9	1.97E-18
	ion binding	1357	9	3.88E-17
	biosynthetic process	934	10	4.81E-18
	cellular nitrogen compound metabolic process	1137	10	4.44E-17
	organelle	2205	10	1.52E-16
	nucleic acid binding transcription factor activity	189	5	2.66E-15
	protein complex	634	5	1.09E-14
	cellular_component	2227	4	1.39E-14
	Fc-epsilon receptor signaling pathway	45	5	5.80E-14

	cytosol	370	3	3.87E-12
	biological_process	1506	2	2.76E-11
	epidermal growth factor receptor signaling pathway	62	5	9.95E-11
	nucleoplasm	189	4	4.63E-10
	enzyme binding	192	3	1.16E-09
	symbiosis, encompassing mutualism through parasitism	86	3	2.12E-07
	catabolic process	306	5	3.35E-07
	blood coagulation	95	5	5.69E-07
	fibroblast growth factor receptor signaling pathway	51	4	2.18E-06
	RNA binding	273	4	6.25E-06
	viral process	75	3	7.37E-06
	cellular component assembly	180	3	8.81E-06
	small molecule metabolic process	379	5	1.28E-05
	protein binding transcription factor activity	75	3	1.48E-05
	macromolecular complex assembly	144	4	2.13E-05
	transcription, DNA-templated	191	2	2.34E-05
	mitotic cell cycle	68	4	0.00024
	membrane organization	92	4	0.00027
	response to stress	261	4	0.00029
	phosphatidylinositol-mediated signaling	33	3	0.00034
	cytoskeletal protein binding	98	3	0.0004
	protein complex assembly	97	3	0.00112
	nervous system development	41	2	0.00189
	cell death	106	2	0.0022
LIHCC	transcription, DNA-templated	790	6	3.80E-25
	transcription initiation from RNA polymerase II promoter	90	6	8.84E-25
	protein complex assembly	250	6	1.11E-24
	cytoskeletal protein binding	249	6	1.36E-24
	enzyme regulator activity	280	6	3.16E-24
	epidermal growth factor receptor signaling pathway	95	7	4.55E-24
	blood coagulation	178	7	1.55E-23
	fibroblast growth factor receptor signaling pathway	79	7	2.61E-23
	immune system process	463	8	4.94E-22
	DNA metabolic process	271	8	1.08E-21
	mRNA metabolic process	104	8	4.73E-21
	Fc-epsilon receptor signaling pathway	79	8	7.67E-21
	mitotic cell cycle	214	9	8.60E-21
	nucleic acid binding transcription factor activity	373	9	2.00E-20
	cell death	374	9	2.64E-20

RNA metabolic process	113	9	4.37E-20
membrane organization	256	9	7.83E-20
protein binding transcription factor activity	229	10	9.93E-20
cellular component assembly	462	10	1.09E-19
cellular protein metabolic process	214	10	1.43E-19
neurotrophin TRK receptor signaling pathway	136	10	2.49E-19
response to stress	780	11	3.92E-19
nucleobase-containing compound catabolic process	368	11	7.90E-19
small molecule metabolic process	842	11	1.49E-18
macromolecular complex assembly	335	11	1.84E-18
catabolic process	755	12	2.38E-18
ion binding	2071	12	2.77E-18
biological_process	4973	13	4.40E-18
molecular_function	5196	14	5.51E-18
cellular_component	5145	14	1.01E-17
nucleoplasm	565	14	1.41E-17
cellular protein modification process	970	14	2.17E-17
protein complex	1429	14	3.83E-17
cytosol	1124	15	3.92E-17
biosynthetic process	1584	15	5.95E-17
viral process	284	15	6.04E-17
enzyme binding	572	15	1.98E-16
poly(A) RNA binding	634	15	2.17E-16
RNA binding	778	16	2.67E-16
gene expression	354	16	2.86E-16
symbiosis, encompassing mutualism through parasitism	317	16	3.13E-16
cellular nitrogen compound metabolic process	1942	17	3.53E-16
organelle	3811	19	7.66E-16
transforming growth factor beta receptor signaling pathway	82	5	1.25E-15
platelet activation	74	6	3.14E-13
TRIF-dependent toll-like receptor signaling pathway	40	6	6.62E-13
post-translational protein modification	74	6	2.35E-12
microtubule organizing center	174	6	6.78E-12
mRNA processing	185	7	2.16E-11
MyD88-independent toll-like receptor signaling pathway	41	6	4.87E-11
phosphatidylinositol-mediated signaling	51	5	8.88E-11
toll-like receptor 10 signaling pathway	33	6	1.94E-10
platelet degranulation	34	5	2.97E-10

innate immune response	202	5	3.21E-10
Fc-gamma receptor signaling pathway involved in phagocytosis	38	5	3.80E-10
small conjugating protein binding	42	4	4.73E-10
toll-like receptor TLR1:TLR2 signaling pathway	33	5	5.38E-10
toll-like receptor TLR6:TLR2 signaling pathway	33	5	5.38E-10
intrinsic apoptotic signaling pathway	34	6	6.79E-10
toll-like receptor 3 signaling pathway	43	6	8.38E-10
transcription from RNA polymerase II promoter	211	7	3.45E-09
mitotic nuclear envelope disassembly	26	5	7.33E-09
RNA splicing	116	6	9.93E-09
transcription factor binding	169	4	1.03E-08
G2/M transition of mitotic cell cycle	69	6	1.05E-08
toll-like receptor 5 signaling pathway	32	5	1.70E-08
toll-like receptor 9 signaling pathway	34	5	2.39E-08
cellular component disassembly involved in execution phase of apoptosis	28	6	7.29E-08
ribonucleoprotein complex assembly	53	4	1.10E-07
nucleocytoplasmic transport	109	5	1.15E-07
cell cycle	233	5	2.55E-07
toll-like receptor 4 signaling pathway	45	6	2.68E-07
hexose transport	24	6	2.82E-07
activation of signaling protein activity involved in unfolded protein response	39	5	4.60E-07
stress-activated MAPK cascade	25	4	6.84E-07
toll-like receptor signaling pathway	45	5	6.86E-07
protein N-linked glycosylation via asparagine	45	6	7.95E-07
viral life cycle	46	5	2.30E-06
nuclear-transcribed mRNA catabolic process, deadenylation-dependent decay	25	6	2.45E-06
insulin receptor signaling pathway	56	3	3.18E-06
toll-like receptor 2 signaling pathway	34	5	4.63E-06
regulation of transcription from RNA polymerase II promoter in response to hypoxia	18	6	4.83E-06
positive regulation of protein insertion into mitochondrial membrane involved in apoptotic signaling pathway	18	6	7.79E-06
cellular lipid metabolic process	54	4	1.01E-05
cellular component movement	44	4	1.34E-05
regulation of glucose transport	21	5	2.61E-05
cell junction organization	47	4	3.38E-05
G1/S transition of mitotic cell cycle	66	6	4.33E-05

	vesicle-mediated transport	269	4	4.58E-05
	in utero embryonic development	124	6	7.73E-05
	mRNA splicing, via spliceosome	55	2	0.00011
	protein targeting	51	1	0.00016
	focal adhesion	144	4	0.00019
	cell cycle arrest	59	4	0.00029
	chromatin organization	55	5	0.00037
	cellular response to hypoxia	45	3	0.0005
	cytoskeleton organization	144	2	0.00071
	termination of RNA polymerase II transcription	24	3	0.0015
	apoptotic signaling pathway	40	2	0.00176
	transcription coactivator activity	104	4	0.00231
	axon guidance	96	2	0.00238
	DNA damage response, signal transduction by p53 class mediator resulting in cell cycle arrest	30	4	0.00253
	extracellular matrix disassembly	31	4	0.00287
	positive regulation of apoptotic process	102	3	0.00463
LUAD	cellular_component	3483	8	1.37E-22
	protein binding transcription factor activity	155	9	2.35E-20
	molecular_function	3472	9	6.03E-19
	nucleic acid binding transcription factor activity	282	10	7.41E-19
	gene expression	207	13	7.44E-19
	cellular protein modification process	843	18	7.91E-19
	cellular nitrogen compound metabolic process	1610	21	2.76E-18
	biosynthetic process	1384	22	3.02E-18
	ion binding	2139	22	5.44E-18
	organelle	3447	24	1.25E-17
	blood coagulation	139	8	7.77E-16
	neurotrophin TRK receptor signaling pathway	87	8	1.11E-15
	protein complex	759	6	1.33E-15
	enzyme binding	327	8	4.05E-13
	Fc-epsilon receptor signaling pathway	59	8	7.57E-11
	transcription, DNA-templated	444	6	8.43E-11
	cellular component assembly	260	5	4.16E-10
	nucleoplasm	252	7	3.74E-09
	cytosol	494	5	4.75E-08
	symbiosis, encompassing mutualism through parasitism	143	7	5.81E-08
	viral process	130	7	6.02E-07
	cytoskeletal protein binding	154	5	1.04E-06
	small molecule metabolic process	456	6	1.42E-05

	biological_process	2374	5	6.28E-05
	catabolic process	339	5	0.00012
	macromolecular complex assembly	153	4	0.00015
	epidermal growth factor receptor signaling pathway	48	4	0.00251
	nucleobase-containing compound catabolic process	122	3	0.00352
LUSC	molecular_function	3331	9	2.69E-19
	cellular protein modification process	600	9	9.83E-19
	biosynthetic process	949	11	2.34E-18
	cellular nitrogen compound metabolic process	1159	12	4.31E-16
	organelle	2294	12	5.61E-16
	ion binding	1451	13	6.67E-16
	cellular_component	2483	5	2.77E-13
	gene expression	138	8	3.51E-12
	nucleic acid binding transcription factor activity	185	6	1.42E-11
	neurotrophin TRK receptor signaling pathway	67	6	1.51E-11
	small molecule metabolic process	431	5	1.19E-10
	protein binding transcription factor activity	67	4	9.02E-10
	protein complex	664	5	1.90E-09
	cytosol	392	5	1.61E-08
	Fc-epsilon receptor signaling pathway	44	6	2.15E-08
	catabolic process	309	4	1.15E-06
	biological_process	2448	4	1.69E-06
	epidermal growth factor receptor signaling pathway	50	5	5.67E-06
	enzyme binding	267	6	1.84E-05
	nucleoplasm	98	4	2.65E-05
	cytoskeletal protein binding	145	3	8.45E-05
	response to stress	191	4	9.22E-05
	blood coagulation	68	3	0.00034
	transcription, DNA-templated	325	4	0.00058
	phosphatidylinositol-mediated signaling	45	6	0.00064
	cellular component assembly	184	3	0.00147
	fibroblast growth factor receptor signaling pathway	43	4	0.00249
	synaptic transmission	67	3	0.00924
READ	molecular_function	2659	4	1.34E-19
	nucleic acid binding transcription factor activity	235	6	8.73E-19
	cellular protein modification process	558	6	2.26E-18
	biosynthetic process	796	6	3.06E-18
	ion binding	1332	8	2.05E-16
	cellular nitrogen compound metabolic process	1039	9	2.28E-16
	organelle	2166	10	2.55E-16

cellular_component	2658	4	2.20E-14
neurotrophin TRK receptor signaling pathway	61	3	1.18E-13
Fc-epsilon receptor signaling pathway	44	3	1.11E-12
gene expression	120	5	4.28E-12
protein binding transcription factor activity	98	4	7.91E-12
cytoskeletal protein binding	163	3	3.50E-11
enzyme binding	252	4	7.67E-11
transcription, DNA-templated	216	3	1.17E-10
blood coagulation	93	3	4.57E-10
protein complex	592	3	1.68E-09
nucleoplasm	181	4	8.03E-09
cellular component assembly	235	3	1.23E-08
biological_process	2416	3	1.07E-07
cytosol	440	3	2.70E-07
enzyme regulator activity	124	2	3.35E-07
epidermal growth factor receptor signaling pathway	48	3	3.44E-06
macromolecular complex assembly	74	1	2.32E-05
transcription initiation from RNA polymerase II promoter	45	3	5.12E-05
cell-cell signaling	79	2	0.00017
synaptic transmission	47	1	0.00018
protein complex assembly	66	1	0.00028
cell death	130	3	0.00061
symbiosis, encompassing mutualism through parasitism	61	2	0.00069
viral process	54	2	0.00094
platelet activation	38	3	0.00416
SKCM cellular_component	4014	9	1.04E-20
neurotrophin TRK receptor signaling pathway	101	9	3.68E-19
nucleic acid binding transcription factor activity	337	10	4.66E-19
gene expression	182	10	4.82E-19
Fc-epsilon receptor signaling pathway	85	10	9.75E-19
molecular_function	4772	13	2.00E-18
cellular protein modification process	833	14	3.73E-18
biosynthetic process	1384	15	5.97E-18
ion binding	2201	18	1.48E-16
organelle	3503	18	2.01E-16
cellular nitrogen compound metabolic process	1762	19	2.34E-16
protein complex	985	8	5.55E-16
enzyme binding	399	11	1.12E-12
small molecule metabolic process	527	6	1.19E-10

protein binding transcription factor activity	152	8	3.48E-10	
catabolic process	427	6	1.65E-09	
biological_process	3565	8	1.07E-08	
nucleoplasm	252	7	1.41E-08	
cytosol	420	4	3.16E-08	
cellular component assembly	334	7	3.17E-08	
cytoskeletal protein binding	221	8	1.84E-07	
symbiosis, encompassing mutualism through parasitism	108	5	8.55E-07	
epidermal growth factor receptor signaling pathway	71	7	8.67E-07	
transcription, DNA-templated	461	5	2.06E-06	
extracellular matrix disassembly	26	1	2.39E-06	
phosphatidylinositol-mediated signaling	51	6	2.37E-05	
viral process	84	4	2.55E-05	
fibroblast growth factor receptor signaling pathway	61	5	3.16E-05	
extracellular matrix organization	44	1	6.51E-05	
endoplasmic reticulum lumen	33	1	7.25E-05	
collagen catabolic process	23	1	0.00026	
membrane organization	147	6	0.00095	
synaptic transmission	90	4	0.0011	
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STAD	cytosol	676	8	2.11E-26
	enzyme binding	385	9	2.80E-23
	cellular_component	4268	10	4.13E-23
	nucleoplasm	368	10	4.41E-21
	epidermal growth factor receptor signaling pathway	89	10	1.02E-19
	protein binding transcription factor activity	171	11	1.11E-19
	neurotrophin TRK receptor signaling pathway	122	11	1.19E-19
	gene expression	217	12	1.33E-19
	Fc-epsilon receptor signaling pathway	85	12	3.10E-19
	nucleic acid binding transcription factor activity	393	13	7.03E-19
	protein complex	1112	13	9.97E-19
	molecular_function	4963	14	1.16E-18
	biosynthetic process	1375	16	3.33E-18
	ion binding	2170	17	1.27E-17
	cellular protein modification process	893	18	1.02E-16
	cellular nitrogen compound metabolic process	1760	19	1.18E-16
	organelle	3520	19	1.29E-16
	blood coagulation	162	10	2.22E-16
	biological_process	3373	7	1.28E-14
	cellular component assembly	360	8	1.52E-14
	transcription, DNA-templated	514	6	5.91E-12

	catabolic process	423	6	2.01E-11
	mitotic cell cycle	85	4	6.28E-10
	cell death	210	5	1.79E-09
	symbiosis, encompassing mutualism through parasitism	138	6	4.82E-09
	RNA binding	257	4	1.94E-08
	fibroblast growth factor receptor signaling pathway	67	7	4.43E-08
	macromolecular complex assembly	210	6	8.05E-08
	viral process	124	6	1.06E-07
	response to stress	323	3	2.91E-06
	small molecule metabolic process	477	6	8.06E-06
	nervous system development	58	4	2.37E-05
	cytoskeletal protein binding	114	3	2.99E-05
	phosphatidylinositol-mediated signaling	42	6	3.37E-05
	synaptic transmission	72	3	5.96E-05
	transcription initiation from RNA polymerase II promoter	68	7	6.46E-05
	nucleobase-containing compound catabolic process	177	4	9.00E-05
	cell-cell signaling	123	4	0.00011
	protein complex assembly	183	6	0.00018
	enzyme regulator activity	178	5	0.00032
	post-translational protein modification	28	2	0.00085
	axon guidance	83	3	0.00425
	Fc-gamma receptor signaling pathway involved in phagocytosis	22	3	0.00997
THCA	transcription, DNA-templated	305	6	5.65E-27
	enzyme binding	317	6	2.95E-24
	nucleic acid binding transcription factor activity	266	8	3.09E-24
	protein binding transcription factor activity	153	9	1.51E-23
	cellular_component	3623	11	2.11E-22
	molecular_function	3655	12	4.36E-21
	cellular protein modification process	651	12	1.75E-20
	gene expression	163	13	8.06E-20
	cellular nitrogen compound metabolic process	1380	18	5.87E-19
	biosynthetic process	1188	19	6.24E-19
	ion binding	1767	19	7.03E-19
	organelle	2787	19	2.12E-18
	Fc-epsilon receptor signaling pathway	61	6	1.11E-16
	neurotrophin TRK receptor signaling pathway	80	7	2.22E-16
	protein complex	658	4	4.14E-10
	blood coagulation	115	5	2.73E-09

	cytosol	470	5	4.08E-09
	nucleoplasm	175	5	1.42E-08
	epidermal growth factor receptor signaling pathway	71	8	2.18E-08
	biological_process	2523	5	2.78E-08
	cellular component assembly	278	6	1.54E-06
	catabolic process	375	5	3.80E-05
	transcription initiation from RNA polymerase II promoter	43	4	0.00029
	phosphatidylinositol-mediated signaling	41	5	0.00042
	regulation of transcription, DNA-templated	86	2	0.00044
	response to stress	315	4	0.00047
	viral process	86	3	0.00057
	post-translational protein modification	41	4	0.00064
	cytoskeletal protein binding	140	3	0.00113
	small molecule metabolic process	318	4	0.00155
	symbiosis, encompassing mutualism through parasitism	91	3	0.00236
	macromolecular complex assembly	163	5	0.0028
UVM	molecular_function	3842	3	4.49E-24
	cellular_component	3650	3	3.88E-21
	enzyme binding	364	3	1.34E-20
	protein binding transcription factor activity	175	4	1.94E-20
	nucleic acid binding transcription factor activity	338	4	5.56E-20
	nucleoplasm	390	4	1.20E-19
	Fc-epsilon receptor signaling pathway	67	4	5.73E-19
	protein complex	957	4	5.90E-19
	neurotrophin TRK receptor signaling pathway	100	4	8.05E-19
	transcription, DNA-templated	686	5	9.47E-19
	biosynthetic process	1236	6	3.20E-17
	gene expression	217	6	4.06E-17
	cellular protein modification process	766	7	7.19E-17
	ion binding	1750	7	1.60E-16
	cellular nitrogen compound metabolic process	1533	8	1.63E-15
	organelle	2867	8	1.65E-15
	epidermal growth factor receptor signaling pathway	79	3	1.96E-13
	biological_process	3261	1	1.61E-12
	cellular component assembly	324	2	2.10E-11
	small molecule metabolic process	537	3	6.44E-11
	viral process	118	1	7.00E-11
	symbiosis, encompassing mutualism through parasitism	130	1	7.84E-11

response to stress	526	3	1.23E-10
RNA binding	441	2	4.39E-10
catabolic process	446	3	1.04E-09
blood coagulation	133	4	6.46E-08
cytosol	557	1	8.22E-08
TRIF-dependent toll-like receptor signaling pathway	30	4	3.70E-07
phosphatidylinositol-mediated signaling	53	3	4.40E-07
MyD88-independent toll-like receptor signaling pathway	31	4	2.33E-06
mitotic cell cycle	92	1	2.71E-06
nervous system development	121	3	1.36E-05
post-translational protein modification	51	1	1.79E-05
cell death	211	2	2.14E-05
fibroblast growth factor receptor signaling pathway	58	2	2.45E-05
cytoskeletal protein binding	197	3	3.89E-05
macromolecular complex assembly	198	2	4.27E-05
nucleobase-containing compound catabolic process	186	1	0.00011
transcription initiation from RNA polymerase II promoter	67	2	0.00013
toll-like receptor 3 signaling pathway	30	4	0.00015
toll-like receptor 10 signaling pathway	25	3	0.00019
toll-like receptor 4 signaling pathway	34	3	0.00034
cellular protein metabolic process	100	1	0.00034
positive regulation of transcription, DNA-templated	243	2	0.00055
toll-like receptor TLR1:TLR2 signaling pathway	25	3	0.00059
toll-like receptor TLR6:TLR2 signaling pathway	25	3	0.00059
transcription from RNA polymerase II promoter	172	2	0.001
toll-like receptor 9 signaling pathway	27	3	0.00183
toll-like receptor 5 signaling pathway	25	3	0.00186
DNA metabolic process	167	1	0.00282
membrane organization	127	1	0.005

Supplementary References

1. Veerla, S, Lindgren, D, Kvist, A, Frigyesi, A, Staaf, J, Persson, H, *et al.* (2009). MiRNA expression in urothelial carcinomas: important roles of miR-10a, miR-222, miR-125b, miR-7 and miR-452 for tumor stage and metastasis, and frequent homozygous losses of miR-31. *Int J Cancer* **124**: 2236-2242.
2. Browne, BM, Stensland, KD, Patel, CK, Sullivan, T, Burks, EJ, Canes, D, *et al.* (2019). MicroRNA Expression Profiles in Upper Tract Urothelial Carcinoma Differentiate Tumor Grade, Stage, and Survival: Implications for Clinical Decision-Making. *Urology* **123**: 93-100.
3. Ratert, N, Meyer, HA, Jung, M, Mollenkopf, HJ, Wagner, I, Miller, K, *et al.* (2012). Reference miRNAs for miRNAome analysis of urothelial carcinomas. *PLoS One* **7**: e39309.
4. Lin, S-R, Yeh, H-C, Wang, W-J, Ke, H-L, Lin, H-H, Hsu, W-C, *et al.* (2017). MiR-193b Mediates CEBPD-Induced Cisplatin Sensitization Through Targeting ETS1 and Cyclin D1 in Human Urothelial Carcinoma Cells. *Journal of Cellular Biochemistry* **118**: 1563-1573.
5. Blick, C, Ramachandran, A, McCormick, R, Wigfield, S, Cranston, D, Catto, J, *et al.* (2015). Identification of a hypoxia-regulated miRNA signature in bladder cancer and a role for miR-145 in hypoxia-dependent apoptosis. *British journal of cancer* **113**: 634-644.
6. Xiao, H, Li, H, Yu, G, Xiao, W, Hu, J, Tang, K, *et al.* (2014). MicroRNA-10b promotes migration and invasion through KLF4 and HOXD10 in human bladder cancer. *Oncol Rep* **31**: 1832-1838.
7. Zhang, Y, Zhang, D, Lv, J, Wang, S, and Zhang, Q (2018). MiR-125a-5p suppresses bladder cancer progression through targeting FUT4. *Biomed Pharmacother* **108**: 1039-1047.
8. Blick, C, Ramachandran, A, McCormick, R, Wigfield, S, Cranston, D, Catto, J, *et al.* (2015). Identification of a hypoxia-regulated miRNA signature in bladder cancer and a role for miR-145 in hypoxia-dependent apoptosis. *British Journal Of Cancer* **113**: 634.
9. Gottardo, F, Liu, CG, Ferracin, M, Calin, GA, Fassan, M, Bassi, P, *et al.* (2007). Micro-RNA profiling in kidney and bladder cancers. *Urol Oncol* **25**: 387-392.
10. Wang, G, Chan, ES, Kwan, BC, Li, PK, Yip, SK, Szeto, CC, *et al.* (2012). Expression of microRNAs in the urine of patients with bladder cancer. *Clin Genitourin Cancer* **10**: 106-113.
11. Itesako, T, Seki, N, Yoshino, H, Chiyomaru, T, Yamasaki, T, Hidaka, H, *et al.* (2014). The microRNA expression signature of bladder cancer by deep sequencing: the functional significance of the miR-195/497 cluster. *PloS one* **9**: e84311-e84311.
12. Chen, L, Yuan, L, Wang, G, Cao, R, Peng, J, Shu, B, *et al.* (2017). Identification and bioinformatics analysis of miRNAs associated with human muscle invasive bladder cancer. *Molecular medicine reports* **16**: 8709-8720.
13. Korpál, M, Ell, BJ, Buffa, FM, Ibrahim, T, Blanco, MA, Celià-Terrassa, T, *et al.* (2011). Direct targeting of Sec23a by miR-200s influences cancer cell secretome and promotes metastatic colonization. *Nature Medicine* **17**: 1101-1108.
14. Shimono, Y, Zabala, M, Cho, RW, Lobo, N, Dalerba, P, Qian, D, *et al.* (2009). Downregulation of miRNA-200c links breast cancer stem cells with normal stem cells. *Cell* **138**: 592-603.
15. Zhao, Z, Fan, X, Jiang, L, Xu, Z, Xue, L, Zhan, Q, *et al.* (2017). miR-503-3p promotes epithelial-mesenchymal transition in breast cancer by directly targeting SMAD2 and E-cadherin. *J Genet Genomics* **44**: 75-84.
16. Lee, CH, Kuo, WH, Lin, CC, Oyang, YJ, Huang, HC, and Juan, HF (2013). MicroRNA-regulated protein-protein interaction networks and their functions in breast cancer. *Int J Mol Sci* **14**: 11560-11606.
17. Kastl, L, Brown, I, and Schofield, AC (2012). miRNA-34a is associated with docetaxel resistance in human breast cancer cells. *Breast Cancer Res Treat* **131**: 445-454.

18. Damavandi, Z, Torkashvand, S, Vasei, M, Soltani, BM, Tavallaei, M, and Mowla, SJ (2016). Aberrant Expression of Breast Development-Related MicroRNAs, miR-22, miR-132, and miR-212, in Breast Tumor Tissues. *J Breast Cancer* **19**: 148-155.
19. Mohammadi-Yeganeh, S, Paryan, M, Mirab Samiee, S, Soleimani, M, Arefian, E, Azadmanesh, K, *et al.* (2013). Development of a robust, low cost stem-loop real-time quantification PCR technique for miRNA expression analysis. *Mol Biol Rep* **40**: 3665-3674.
20. Bhaumik, D, Scott, GK, Schokrpur, S, Patil, CK, Campisi, J, and Benz, CC (2008). Expression of microRNA-146 suppresses NF-kappaB activity with reduction of metastatic potential in breast cancer cells. *Oncogene* **27**: 5643-5647.
21. Luo, EC, Chang, YC, Sher, YP, Huang, WY, Chuang, LL, Chiu, YC, *et al.* (2014). MicroRNA-769-3p down-regulates NDRG1 and enhances apoptosis in MCF-7 cells during reoxygenation. *Sci Rep* **4**: 5908.
22. Pichler, M, Stiegelbauer, V, Vychytilova-Faltejskova, P, Ivan, C, Ling, H, Winter, E, *et al.* (2017). Genome-Wide miRNA Analysis Identifies miR-188-3p as a Novel Prognostic Marker and Molecular Factor Involved in Colorectal Carcinogenesis. *Clinical cancer research : an official journal of the American Association for Cancer Research* **23**: 1323-1333.
23. Sun, JY, Huang, Y, Li, JP, Zhang, X, Wang, L, Meng, YL, *et al.* (2012). MicroRNA-320a suppresses human colon cancer cell proliferation by directly targeting beta-catenin. *Biochem Biophys Res Commun* **420**: 787-792.
24. Zhang, Y, Li, M, Ding, Y, Fan, Z, Zhang, J, Zhang, H, *et al.* (2017). Serum MicroRNA profile in patients with colon adenomas or cancer. *BMC medical genomics* **10**: 23-23.
25. Song, B, Wang, Y, Xi, Y, Kudo, K, Bruheim, S, Botchkina, GI, *et al.* (2009). Mechanism of chemoresistance mediated by miR-140 in human osteosarcoma and colon cancer cells. *Oncogene* **28**: 4065-4074.
26. Yu, G, Tang, J-q, Tian, M-l, Li, H, Wang, X, Wu, T, *et al.* (2012). Prognostic values of the miR-17-92 cluster and its paralogs in colon cancer. *Journal of Surgical Oncology* **106**: 232-237.
27. Weber, D, Amar, L, Gödde, D, and Prinz, C (2018). Extensive screening of microRNA populations identifies hsa-miR-375 and hsa-miR-133a-3p as selective markers for human rectal and colon cancer. *Oncotarget* **9**: 27256-27267.
28. Tao, K, Yang, J, Guo, Z, Hu, Y, Sheng, H, Gao, H, *et al.* (2014). Prognostic value of miR-221-3p, miR-342-3p and miR-491-5p expression in colon cancer. *American journal of translational research* **6**: 391-401.
29. Liu, S-G, Qin, X-G, Zhao, B-S, Qi, B, Yao, W-J, Wang, T-Y, *et al.* (2013). Differential expression of miRNAs in esophageal cancer tissue. *Oncology letters* **5**: 1639-1642.
30. Slaby, O, Srovnal, J, Radova, L, Gregar, J, Juracek, J, Luzna, P, *et al.* (2015). Dynamic changes in microRNA expression profiles reflect progression of Barrett's esophagus to esophageal adenocarcinoma. *Carcinogenesis* **36**: 521-527.
31. Byrnes, KA, Phatak, P, Mansour, D, Xiao, L, Zou, T, Rao, JN, *et al.* (2016). Overexpression of miR-199a-5p decreases esophageal cancer cell proliferation through repression of mitogen-activated protein kinase kinase kinase-11 (MAP3K11). *Oncotarget* **7**: 8756-8770.
32. Streppel, MM, Pai, S, Campbell, NR, Hu, C, Yabuuchi, S, Canto, MI, *et al.* (2013). MicroRNA 223 is upregulated in the multistep progression of Barrett's esophagus and modulates sensitivity to chemotherapy by targeting PARP1. *Clin Cancer Res* **19**: 4067-4078.
33. Sun, Q, Zong, L, Zhang, H, Deng, Y, Zhang, C, and Zhang, L (2018). A 10-microRNA prognosis scoring system in esophageal squamous cell carcinoma constructed using bioinformatic methods. *Molecular medicine reports* **17**: 5222-5228.
34. Su, H, Jin, X, Zhang, X, Xue, S, Deng, X, Shen, L, *et al.* (2014). Identification of microRNAs involved in the radioresistance of esophageal cancer cells. *Cell Biology International* **38**: 318-325.

35. Komatsu, S, Ichikawa, D, Kawaguchi, T, Takeshita, H, Miyamae, M, Ohashi, T, *et al.* (2016). Plasma microRNA profiles: identification of miR-23a as a novel biomarker for chemoresistance in esophageal squamous cell carcinoma. *Oncotarget* **7**: 62034-62048.
36. Yin, J, Wang, X, Zheng, L, Shi, Y, Wang, L, Shao, A, *et al.* (2013). Hsa-miR-34b/c rs4938723 T>C and hsa-miR-423 rs6505162 C>A Polymorphisms Are Associated with the Risk of Esophageal Cancer in a Chinese Population. *PLOS ONE* **8**: e80570.
37. Yang, H, Gu, J, Wang, KK, Zhang, W, Xing, J, Chen, Z, *et al.* (2009). MicroRNA expression signatures in Barrett's esophagus and esophageal adenocarcinoma. *Clinical cancer research : an official journal of the American Association for Cancer Research* **15**: 5744-5752.
38. Zhang, K, Wu, X, Wang, J, Lopez, J, Zhou, W, Yang, L, *et al.* (2016). Circulating miRNA profile in esophageal adenocarcinoma. *American journal of cancer research* **6**: 2713-2721.
39. Lu, TX, Sherrill, JD, Wen, T, Plassard, AJ, Besse, JA, Abonia, JP, *et al.* (2012). MicroRNA signature in patients with eosinophilic esophagitis, reversibility with glucocorticoids, and assessment as disease biomarkers. *J Allergy Clin Immunol* **129**: 1064-1075.e1069.
40. Lin, C, Liu, A, Zhu, J, Zhang, X, Wu, G, Ren, P, *et al.* (2014). miR-508 sustains phosphoinositide signalling and promotes aggressive phenotype of oesophageal squamous cell carcinoma. *Nature Communications* **5**: 4620.
41. Li, G, Ren, S, Su, Z, Liu, C, Deng, T, Huang, D, *et al.* (2015). Increased expression of miR-93 is associated with poor prognosis in head and neck squamous cell carcinoma. *Tumour Biol* **36**: 3949-3956.
42. Hui, AB, Lenarduzzi, M, Krushel, T, Waldron, L, Pintilie, M, Shi, W, *et al.* (2010). Comprehensive MicroRNA profiling for head and neck squamous cell carcinomas. *Clin Cancer Res* **16**: 1129-1139.
43. Huang, Q, Yang, J, Zheng, J, Hsueh, C, Guo, Y, and Zhou, L (2018). Characterization of selective exosomal microRNA expression profile derived from laryngeal squamous cell carcinoma detected by next generation sequencing. *Oncology reports* **40**: 2584-2594.
44. Ogawa, T, Saiki, Y, Shiga, K, Chen, N, Fukushige, S, Sunamura, M, *et al.* (2012). miR-34a is downregulated in cis-diamminedichloroplatinum treated sinonasal squamous cell carcinoma patients with poor prognosis. *Cancer Sci* **103**: 1737-1743.
45. Saad, MA, Kuo, SZ, Rahimy, E, Zou, AE, Korrapati, A, Rahimy, M, *et al.* (2015). Alcohol-dysregulated miR-30a and miR-934 in head and neck squamous cell carcinoma. *Molecular cancer* **14**: 181-181.
46. Hou, B, Ishinaga, H, Midorikawa, K, Shah, SA, Nakamura, S, Hiraku, Y, *et al.* (2015). Circulating microRNAs as novel prognosis biomarkers for head and neck squamous cell carcinoma. *Cancer biology & therapy* **16**: 1042-1046.
47. Miao, L, Wang, L, Yuan, H, Hang, D, Zhu, L, Du, J, *et al.* (2016). MicroRNA-101 polymorphisms and risk of head and neck squamous cell carcinoma in a Chinese population. *Tumour Biol* **37**: 4169-4174.
48. Koshizuka, K, Hanazawa, T, Fukumoto, I, Kikkawa, N, Matsushita, R, Mataka, H, *et al.* (2017). Dual-receptor (EGFR and c-MET) inhibition by tumor-suppressive miR-1 and miR-206 in head and neck squamous cell carcinoma. *J Hum Genet* **62**: 113-121.
49. Liu, F, Zhao, X, Qian, Y, Zhang, J, Zhang, Y, and Yin, R (2017). MiR-206 inhibits Head and neck squamous cell carcinoma cell progression by targeting HDAC6 via PTEN/AKT/mTOR pathway. *Biomed Pharmacother* **96**: 229-237.
50. Xiao, W, Lou, N, Ruan, H, Bao, L, Xiong, Z, Yuan, C, *et al.* (2017). Mir-144-3p Promotes Cell Proliferation, Metastasis, Sunitinib Resistance in Clear Cell Renal Cell Carcinoma by Downregulating ARID1A. *Cell Physiol Biochem* **43**: 2420-2433.

51. White, NM, Khella, HW, Grigull, J, Adzovic, S, Youssef, YM, Honey, RJ, *et al.* (2011). miRNA profiling in metastatic renal cell carcinoma reveals a tumour-suppressor effect for miR-215. *Br J Cancer* **105**: 1741-1749.
52. Zhou, L, Li, Z, Pan, X, Lai, Y, Quan, J, Zhao, L, *et al.* (2018). Identification of miR-18a-5p as an oncogene and prognostic biomarker in RCC. *Am J Transl Res* **10**: 1874-1886.
53. Khella, HWZ, Butz, H, Ding, Q, Rotondo, F, Evans, KR, Kupchak, P, *et al.* (2015). miR-221/222 Are Involved in Response to Sunitinib Treatment in Metastatic Renal Cell Carcinoma. *Mol Ther* **23**: 1748-1758.
54. Carlsson, J, Christiansen, J, Davidsson, S, Giunchi, F, Fiorentino, M, and Sundqvist, P (2019). The potential role of miR-126, miR-21 and miR-10b as prognostic biomarkers in renal cell carcinoma. *Oncology letters* **17**: 4566-4574.
55. Vergho, DC, Kneitz, S, Kalogirou, C, Burger, M, Krebs, M, Rosenwald, A, *et al.* (2014). Impact of miR-21, miR-126 and miR-221 as Prognostic Factors of Clear Cell Renal Cell Carcinoma with Tumor Thrombus of the Inferior Vena Cava. *PLOS ONE* **9**: e109877.
56. Zaravinos, A, Lambrou, GI, Mourmouras, N, Katafygiotis, P, Papagregoriou, G, Giannikou, K, *et al.* (2014). New miRNA profiles accurately distinguish renal cell carcinomas and upper tract urothelial carcinomas from the normal kidney. *PLoS one* **9**: e91646-e91646.
57. Wulfken, LM, Moritz, R, Ohlmann, C, Holdenrieder, S, Jung, V, Becker, F, *et al.* (2011). MicroRNAs in Renal Cell Carcinoma: Diagnostic Implications of Serum miR-1233 Levels. *PLOS ONE* **6**: e25787.
58. Petillo, D, Kort, EJ, Anema, J, Furge, KA, Yang, XJ, and Teh, BT (2009). MicroRNA profiling of human kidney cancer subtypes. *Int J Oncol* **35**: 109-114.
59. Heinemann, FG, Tolkach, Y, Deng, M, Schmidt, D, Perner, S, Kristiansen, G, *et al.* (2018). Serum miR-122-5p and miR-206 expression: non-invasive prognostic biomarkers for renal cell carcinoma. *Clinical Epigenetics* **10**: 11.
60. Hong, Q, Li, O, Zheng, W, Xiao, W-z, Zhang, L, Wu, D, *et al.* (2017). LncRNA HOTAIR regulates HIF-1 α /AXL signaling through inhibition of miR-217 in renal cell carcinoma. *Cell Death & Disease* **8**: e2772-e2772.
61. Tian, Q, Liang, L, Ding, J, Zha, R, Shi, H, Wang, Q, *et al.* (2012). MicroRNA-550a Acts as a Pro-Metastatic Gene and Directly Targets Cytoplasmic Polyadenylation Element-Binding Protein 4 in Hepatocellular Carcinoma. *PLOS ONE* **7**: e48958.
62. Zheng, J, Sadot, E, Vigidal, JA, Klimstra, DS, Balachandran, VP, Kingham, TP, *et al.* (2018). Characterization of hepatocellular adenoma and carcinoma using microRNA profiling and targeted gene sequencing. *PLoS one* **13**: e0200776-e0200776.
63. Augello, C, Gianelli, U, Savi, F, Moro, A, Bonoldi, E, Gambacorta, M, *et al.* (2014). MicroRNA as potential biomarker in HCV-associated diffuse large B-cell lymphoma. *J Clin Pathol* **67**: 697-701.
64. Shih, TC, Tien, YJ, Wen, CJ, Yeh, TS, Yu, MC, Huang, CH, *et al.* (2012). MicroRNA-214 downregulation contributes to tumor angiogenesis by inducing secretion of the hepatoma-derived growth factor in human hepatoma. *J Hepatol* **57**: 584-591.
65. Gui, J, Tian, Y, Wen, X, Zhang, W, Zhang, P, Gao, J, *et al.* (2011). Serum microRNA characterization identifies miR-885-5p as a potential marker for detecting liver pathologies. *Clin Sci (Lond)* **120**: 183-193.
66. Yu, L, Ding, GF, He, C, Sun, L, Jiang, Y, and Zhu, L (2014). MicroRNA-424 is down-regulated in hepatocellular carcinoma and suppresses cell migration and invasion through c-Myb. *PLoS One* **9**: e91661.
67. Morishita, A, Iwama, H, Fujihara, S, Sakamoto, T, Fujita, K, Tani, J, *et al.* (2016). MicroRNA profiles in various hepatocellular carcinoma cell lines. *Oncology letters* **12**: 1687-1692.

68. Saito, Y, Suzuki, H, Matsuura, M, Sato, A, Kasai, Y, Yamada, K, *et al.* (2011). MicroRNAs in Hepatobiliary and Pancreatic Cancers. *Front Genet* **2**: 66.
69. Xie, F, Yuan, Y, Xie, L, Ran, P, Xiang, X, Huang, Q, *et al.* (2017). miRNA-320a inhibits tumor proliferation and invasion by targeting c-Myc in human hepatocellular carcinoma. *OncoTargets and therapy* **10**: 885-894.
70. Lin, K, Xu, T, He, B-S, Pan, Y-Q, Sun, H-L, Peng, H-X, *et al.* (2016). MicroRNA expression profiles predict progression and clinical outcome in lung adenocarcinoma. *OncoTargets and therapy* **9**: 5679-5692.
71. Leidinger, P, Backes, C, Blatt, M, Keller, A, Huwer, H, Lepper, P, *et al.* (2014). The blood-borne miRNA signature of lung cancer patients is independent of histology but influenced by metastases. *Molecular Cancer* **13**: 202.
72. Othman, N, In, LLA, Harikrishna, JA, and Hasima, N (2013). Bcl-xL Silencing Induces Alterations in hsa-miR-608 Expression and Subsequent Cell Death in A549 and SK-LU1 Human Lung Adenocarcinoma Cells. *PLOS ONE* **8**: e81735.
73. Dacic, S, Kelly, L, Shuai, Y, and Nikiforova, MN (2010). miRNA expression profiling of lung adenocarcinomas: correlation with mutational status. *Modern Pathology* **23**: 1577-1582.
74. Lin, Y, Lv, Y, Liang, R, Yuan, C, Zhang, J, He, D, *et al.* (2017). Four-miRNA signature as a prognostic tool for lung adenocarcinoma. *OncoTargets and therapy* **11**: 29-36.
75. Yang, F, Wei, K, Qin, Z, Liu, W, Shao, C, Wang, C, *et al.* (2018). MiR-598 Suppresses Invasion and Migration by Negative Regulation of Derlin-1 and Epithelial-Mesenchymal Transition in Non-Small Cell Lung Cancer. *Cellular Physiology and Biochemistry* **47**: 245-256.
76. Patnaik, SK, Yendamuri, S, Kannisto, E, Kucharczuk, JC, Singhal, S, and Vachani, A (2012). MicroRNA Expression Profiles of Whole Blood in Lung Adenocarcinoma. *PLOS ONE* **7**: e46045.
77. Galluzzi, L, Morselli, E, Vitale, I, Kepp, O, Senovilla, L, Criollo, A, *et al.* (2010). miR-181a and miR-630 Regulate Cisplatin-Induced Cancer Cell Death. *Cancer Research* **70**: 1793.
78. Ma, Y, Pan, X, Xu, P, Mi, Y, Wang, W, Wu, X, *et al.* (2017). Plasma microRNA alterations between EGFR-activating mutational NSCLC patients with and without primary resistance to TKI. *Oncotarget* **8**: 88529-88536.
79. Lee, J-H, Voortman, J, Dingemans, A-MC, Voeller, DM, Pham, T, Wang, Y, *et al.* (2011). MicroRNA Expression and Clinical Outcome of Small Cell Lung Cancer. *PLOS ONE* **6**: e21300.
80. Ma, J, Mannoor, K, Gao, L, Tan, A, Guarnera, MA, Zhan, M, *et al.* (2014). Characterization of microRNA transcriptome in lung cancer by next-generation deep sequencing. *Molecular Oncology* **8**: 1208-1219.
81. Zhang, K, Zhang, M, Jiang, H, Liu, F, Liu, H, and Li, Y (2018). Down-regulation of miR-214 inhibits proliferation and glycolysis in non-small-cell lung cancer cells via down-regulating the expression of hexokinase 2 and pyruvate kinase isozyme M2. *Biomed Pharmacother* **105**: 545-552.
82. Kim, MK, Jung, SB, Kim, JS, Roh, MS, Lee, JH, Lee, EH, *et al.* (2014). Expression of microRNA miR-126 and miR-200c is associated with prognosis in patients with non-small cell lung cancer. *Virchows Arch* **465**: 463-471.
83. Liu, XG, Zhu, WY, Huang, YY, Ma, LN, Zhou, SQ, Wang, YK, *et al.* (2012). High expression of serum miR-21 and tumor miR-200c associated with poor prognosis in patients with lung cancer. *Med Oncol* **29**: 618-626.
84. Bach, DH, Luu, TT, Kim, D, An, YJ, Park, S, Park, HJ, *et al.* (2018). BMP4 Upregulation Is Associated with Acquired Drug Resistance and Fatty Acid Metabolism in EGFR-Mutant Non-Small-Cell Lung Cancer Cells. *Mol Ther Nucleic Acids* **12**: 817-828.
85. Liu, XH, Lu, KH, Wang, KM, Sun, M, Zhang, EB, Yang, JS, *et al.* (2012). MicroRNA-196a promotes non-small cell lung cancer cell proliferation and invasion through targeting HOXA5. *BMC Cancer* **12**: 348.

86. Edmonds, MD, and Eischen, CM (2014). Differences in miRNA expression in early stage lung adenocarcinomas that did and did not relapse. *PLoS one* **9**: e101802-e101802.
87. Mazar, J, DeYoung, K, Khaitan, D, Meister, E, Almodovar, A, Goydos, J, *et al.* (2010). The regulation of miRNA-211 expression and its role in melanoma cell invasiveness. *PLoS one* **5**: e13779-e13779.
88. Weber, CE, Luo, C, Hotz-Wagenblatt, A, Gardyan, A, Kordass, T, Holland-Letz, T, *et al.* (2016). miR-339-3p Is a Tumor Suppressor in Melanoma. *Cancer Res* **76**: 3562-3571.
89. Lunavat, TR, Cheng, L, Einarsdottir, BO, Olofsson Bagge, R, Veppil Muralidharan, S, Sharples, RA, *et al.* (2017). BRAF(V600) inhibition alters the microRNA cargo in the vesicular secretome of malignant melanoma cells. *Proc Natl Acad Sci U S A* **114**: E5930-e5939.
90. Leidinger, P, Keller, A, Borries, A, Reichrath, J, Rass, K, Jager, SU, *et al.* (2010). High-throughput miRNA profiling of human melanoma blood samples. *BMC Cancer* **10**: 262.
91. Qian, H, Yang, C, and Yang, Y (2017). MicroRNA-26a inhibits the growth and invasiveness of malignant melanoma and directly targets on MITF gene. *Cell Death Discovery* **3**: 17028.
92. Fattore, L, Costantini, S, Malpicci, D, Ruggiero, CF, Ascierto, PA, Croce, CM, *et al.* (2017). MicroRNAs in melanoma development and resistance to target therapy. *Oncotarget* **8**: 22262-22278.
93. Chitsazzadeh, V, Coarfa, C, Drummond, JA, Nguyen, T, Joseph, A, Chilukuri, S, *et al.* (2016). Cross-species identification of genomic drivers of squamous cell carcinoma development across preneoplastic intermediates. *Nat Commun* **7**: 12601.
94. Wang, S, Han, H, Hu, Y, Yang, W, Lv, Y, Wang, L, *et al.* (2018). MicroRNA-130a-3p suppresses cell migration and invasion by inhibition of TBL1XR1-mediated EMT in human gastric carcinoma. *Mol Carcinog* **57**: 383-392.
95. Liu, J, Ma, L, Wang, Z, Wang, L, Liu, C, Chen, R, *et al.* (2014). MicroRNA expression profile of gastric cancer stem cells in the MKN-45 cancer cell line. *Acta Biochimica et Biophysica Sinica* **46**: 92-99.
96. Li, X, Luo, F, Li, Q, Xu, M, Feng, D, Zhang, G, *et al.* (2011). Identification of new aberrantly expressed miRNAs in intestinal-type gastric cancer and its clinical significance. *Oncol Rep* **26**: 1431-1439.
97. Zuo, QF, Zhang, R, Li, BS, Zhao, YL, Zhuang, Y, Yu, T, *et al.* (2015). MicroRNA-141 inhibits tumor growth and metastasis in gastric cancer by directly targeting transcriptional co-activator with PDZ-binding motif, TAZ. *Cell Death Dis* **6**: e1623.
98. Wu, Y-C, Liu, X, Wang, J-L, Chen, X-L, Lei, L, Han, J, *et al.* (2018). Soft-shelled turtle peptide modulates microRNA profile in human gastric cancer AGS cells. *Oncology letters* **15**: 3109-3120.
99. Zhang, QH, Yao, YL, Wu, XY, Wu, JH, Gu, T, Chen, L, *et al.* (2015). Anti-miR-362-3p Inhibits Migration and Invasion of Human Gastric Cancer Cells by Its Target CD82. *Dig Dis Sci* **60**: 1967-1976.
100. Pan, Y, Ren, F, Zhang, W, Liu, G, Yang, D, Hu, J, *et al.* (2014). Regulation of BGC-823 cell sensitivity to adriamycin via miRNA-135a-5p. *Oncol Rep* **32**: 2549-2556.
101. Li, X, Zhang, Y, Zhang, H, Liu, X, Gong, T, Li, M, *et al.* (2011). miRNA-223 promotes gastric cancer invasion and metastasis by targeting tumor suppressor EPB41L3. *Mol Cancer Res* **9**: 824-833.
102. Komatsu, S, Ichikawa, D, Tsujiura, M, Konishi, H, Takeshita, H, Nagata, H, *et al.* (2013). Prognostic impact of circulating miR-21 in the plasma of patients with gastric carcinoma. *Anticancer Res* **33**: 271-276.
103. Song, YX, Yue, ZY, Wang, ZN, Xu, YY, Luo, Y, Xu, HM, *et al.* (2011). MicroRNA-148b is frequently down-regulated in gastric cancer and acts as a tumor suppressor by inhibiting cell proliferation. *Mol Cancer* **10**: 1.

104. Hu, J, Li, C, Liu, C, Zhao, S, Wang, Y, and Fu, Z (2017). Expressions of miRNAs in papillary thyroid carcinoma and their associations with the clinical characteristics of PTC. *Cancer Biomark* **18**: 87-94.
105. Mohamad Yusof, A, Jamal, R, Muhammad, R, Abdullah Suhaimi, SN, Mohamed Rose, I, Saidin, S, *et al.* (2018). Integrated Characterization of MicroRNA and mRNA Transcriptome in Papillary Thyroid Carcinoma. *Frontiers in endocrinology* **9**: 158-158.
106. Swierniak, M, Wojcicka, A, Czetwertynska, M, Stachlewska, E, Maciag, M, Wiechno, W, *et al.* (2013). In-Depth Characterization of the MicroRNA Transcriptome in Normal Thyroid and Papillary Thyroid Carcinoma. *The Journal of Clinical Endocrinology & Metabolism* **98**: E1401-E1409.
107. Wang, C, Lu, S, Jiang, J, Jia, X, Dong, X, and Bu, P (2014). Hsa-microRNA-101 suppresses migration and invasion by targeting Rac1 in thyroid cancer cells. *Oncol Lett* **8**: 1815-1821.
108. Stokowy, T, Wojtaś, B, Krajewska, J, Stobiecka, E, Dralle, H, Musholt, T, *et al.* (2015). A two miRNA classifier differentiates follicular thyroid carcinomas from follicular thyroid adenomas. *Molecular and Cellular Endocrinology* **399**: 43-49.
109. Nikiforova, MN, Gandhi, M, Kelly, L, and Nikiforov, YE (2011). MicroRNA dysregulation in human thyroid cells following exposure to ionizing radiation. *Thyroid : official journal of the American Thyroid Association* **21**: 261-266.
110. Braun, J, Hoang-Vu, C, Dralle, H, and Hüttelmaier, S (2010). Downregulation of microRNAs directs the EMT and invasive potential of anaplastic thyroid carcinomas. *Oncogene* **29**: 4237-4244.
111. Marini, F, Luzi, E, and Brandi, ML (2011). MicroRNA Role in Thyroid Cancer Development. *Journal of thyroid research* **2011**: 407123-407123.
112. Liu, Y, Zhang, Y, Wu, H, Li, Y, Zhang, Y, Liu, M, *et al.* (2017). miR-10a suppresses colorectal cancer metastasis by modulating the epithelial-to-mesenchymal transition and anoikis. *Cell Death Dis* **8**: e2739.
113. Park, YR, Lee, ST, Kim, SL, Liu, YC, Lee, MR, Shin, JH, *et al.* (2016). MicroRNA-9 suppresses cell migration and invasion through downregulation of TM4SF1 in colorectal cancer. *Int J Oncol* **48**: 2135-2143.
114. Gaedcke, J, Grade, M, Camps, J, Sokilde, R, Kaczowski, B, Schetter, AJ, *et al.* (2012). The rectal cancer microRNAome--microRNA expression in rectal cancer and matched normal mucosa. *Clin Cancer Res* **18**: 4919-4930.
115. Nakano, H, Miyazawa, T, Kinoshita, K, Yamada, Y, and Yoshida, T (2010). Functional screening identifies a microRNA, miR-491 that induces apoptosis by targeting Bcl-XL in colorectal cancer cells. *International Journal of Cancer* **127**: 1072-1080.
116. Tateishi, Y, Okudela, K, Mitsui, H, Umeda, S, Suzuki, T, Kojima, Y, *et al.* (2015). The potential role of microRNA-31 expression in early colorectal cancer. *Pathol Int* **65**: 513-518.
117. Jacob, H, Stanisavljevic, L, Storli, KE, Hestetun, KE, Dahl, O, and Myklebust, MP (2018). A four-microRNA classifier as a novel prognostic marker for tumor recurrence in stage II colon cancer. *Scientific Reports* **8**: 6157.
118. Li, J, Zou, K, Yu, L, Zhao, W, Lu, Y, Mao, J, *et al.* (2018). MicroRNA-140 Inhibits the Epithelial-Mesenchymal Transition and Metastasis in Colorectal Cancer. *Mol Ther Nucleic Acids* **10**: 426-437.
119. Bu, P, Wang, L, Chen, KY, Rakhilin, N, Sun, J, Closa, A, *et al.* (2015). miR-1269 promotes metastasis and forms a positive feedback loop with TGF-beta. *Nat Commun* **6**: 6879.
120. Huang, L, Cai, JL, Huang, PZ, Kang, L, Huang, MJ, Wang, L, *et al.* (2017). miR19b-3p promotes the growth and metastasis of colorectal cancer via directly targeting ITGB8. *American journal of cancer research* **7**: 1996-2008.

121. Robertson, AG, Shih, J, Yau, C, Gibb, EA, Oba, J, Mungall, KL, *et al.* (2017). Integrative Analysis Identifies Four Molecular and Clinical Subsets in Uveal Melanoma. *Cancer Cell* **32**: 204-220.e215.
122. Raghunath, A, and Perumal, E (2015). Micro-RNAs and Their Roles in Eye Disorders. *Ophthalmic Research* **53**: 169-186.
123. Falzone, L, Romano, GL, Salemi, R, Bucolo, C, Tomasello, B, Lupo, G, *et al.* (2019). Prognostic significance of deregulated microRNAs in uveal melanomas. *Molecular medicine reports* **19**: 2599-2610.
124. Zheng, X, Tang, H, Zhao, X, Sun, Y, Jiang, Y, and Liu, Y (2017). Long non-coding RNA FTH1P3 facilitates uveal melanoma cell growth and invasion through miR-224-5p. *PLoS One* **12**: e0184746.