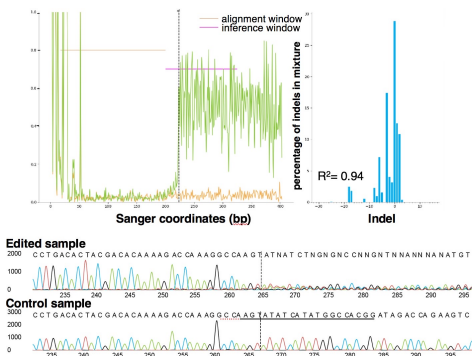


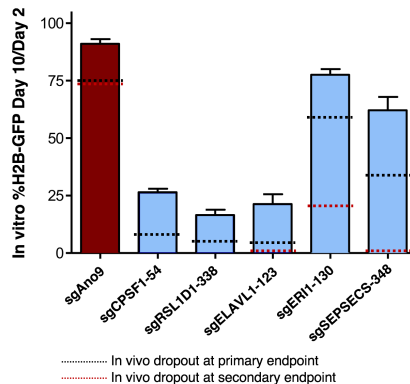
# Supplemental Figure S2

## A

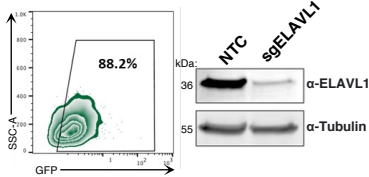


Guide	Normalized Indel %	R <sup>2</sup>
sgCpsf1-54	86	0.94
sgCpsf1-58	87	0.94
sgRsl1d1-338	63	0.97
sgRsl1d1-339	63	0.98
sgElavl1-120	61	0.97
sgElavl1-121	91	0.89
sgElavl1-122	58	0.97
sgElavl1-123	77	0.95
sgSepsecs-346	82	0.89
sgSepsecs-348	81	0.93

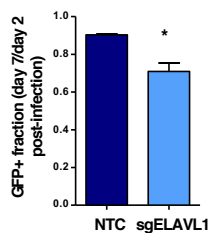
## B



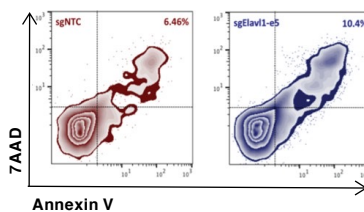
## C



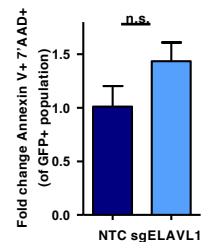
## D



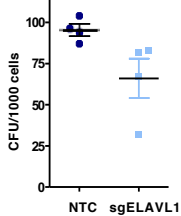
## E



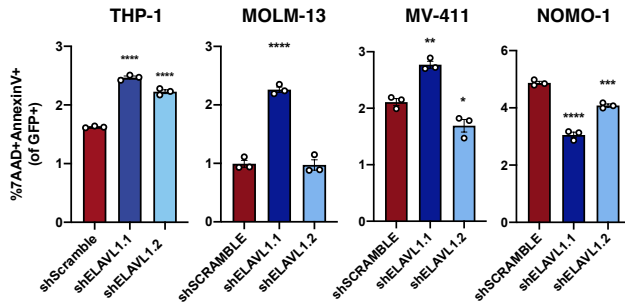
## F



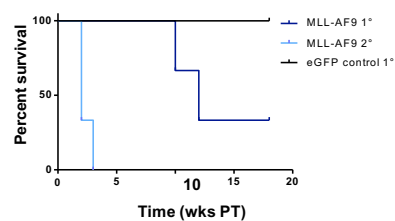
## G



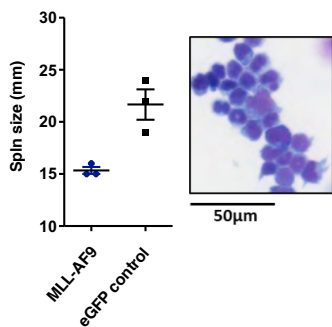
## H



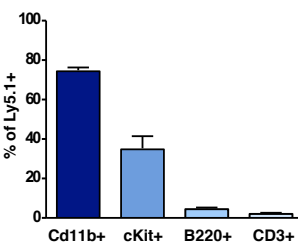
## I



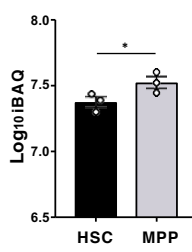
## J



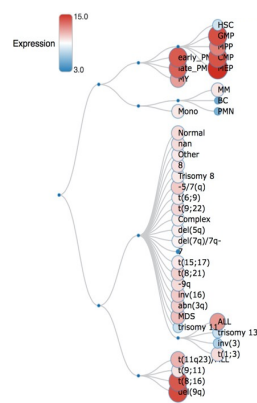
## K



## L



## M



**Supplemental Figure S2. Validation and characterization of primary and secondary screen hit-targeting sgRNAs.** (A) Representative indel plots and traces generated from ICE analysis<sup>41</sup> for individually validated sgRNAs in test RN2c cells. Level of discordance (top left) in the sample sequence (green) and control sequence (orange) before and after the expected cut-site is shown. Indel plot (top right) shows percentages of the various calculated indel sizes. Trace sequences (bottom) display the aligned sample and control sequences with the targeted cut-site. Indel percentages for individually validated sgRNAs normalized to the percent infection of each sample. R2 values were generated from Indel plots. (B) *In vitro* assessment of RN2c growth upon knockout of selected screen hits. Levels of H2B-GFP reduction achieved in parallel *in vivo* experiments in primary and secondary mice are highlighted in black and red dotted lines respectively. (C) Western blot validation of ELAVL1 knockout by pL.CRISPR.EFS.GFP-sgELAVL1 in HEK293 cells one week post-infection. Prior to lysis, cultures were analyzed by flow cytometry for the fraction of GFP<sup>+</sup> cells; sgNTC was used as a negative control. (D) sgRNA-mediated KO of ELAVL1 in THP-1 cells; GFP<sup>+</sup> fraction was followed over time. GFP<sup>+</sup> fractions of sgNTC and sgELAVL1 infected THP-1 cells at 7 days post-transduction are normalized to the percent infected at 2 days PT. (E,F) Representative flow plots (E) and quantitative analysis of the apoptotic fraction of GFP<sup>+</sup> shScramble and shELAVL1 THP-1 cells 4 days post-plating (F). Data represents 3 replicate infections. (G) CFU assays performed with FACS-purified GFP<sup>+</sup> sgNTC and sgELAVL1 THP-1 cells. (H) Cell death quantified by flow cytometric analysis in AML cell lines (THP-1, MOLM-13, MV-411, NOMO-1) 4 days post-infection with shRNA targeting ELAVL1 and Scramble (control). (I) B6.SJL mouse LSK cells were retrovirally infected with MSCV-MLLAF9-PGK-eGFP (or MSCV-PGK-eGFP, control) and GFP<sup>+</sup> cells FACS-purified 48hr post-infection transplanted into C57Blk/6 recipients. Fully engrafted primary MLL-AF9-driven leukemic BM was challenged in secondary recipients. Latency of primary (1<sup>o</sup>) and secondary (2<sup>o</sup>) transplant MLL-AF9 leukemias, as well as primary eGFP control BM (n = 3 for each arm) is shown. (J) Spleen sizes of primary engrafted MLL-AF9-driven leukemias and primary eGFP control BM (left) and Wright-Giemsa staining of peripheral blood sampled from primary engrafted recipient mouse in MLL-AF9 arm (right). (K) Flow cytometric evaluation of the immunophenotype of MLL-AF9-driven BM grafts. (L) Expression levels of *Elavl1* in mouse HSC (LSK CD34-CD150+CD48-FLK2-) and MPP (LSK CD34+CD15-+CD48-FLK2-) populations<sup>42</sup>. Intensity based absolute quantification (iBAQ) levels are shown. (M) Correlation tree of normal and malignant hematopoietic samples based on *ELAVL1* expression levels generated by the BloodPool tool, adapted from Bloodspot<sup>43</sup> showcasing low (blue) expression in HSCs relative to higher (pink to red) expression in bulk AML cells.