

Supplemental Files

Three years of wastewater surveillance for new psychoactive substances in 16 countries

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Analytical Methodology

Briefly, following filtration (GF/A 1.6 μm , Whatman, Kent, UK), the pH was adjusted to 4.5–5 using aqueous ammonia (28%) prior to solid phase extraction (UCT XtracT DAU, 500 mg/6 mL; UCT Inc., Bristol, PA, USA). The samples (100 mL) were loaded under gravity and the cartridges washed with a sodium acetate buffer (20 mM, 6 mL), acetic acid (0.1 M, 2 mL), and methanol (6 mL) before being air dried for 15 min. The dried cartridges were stored at $-20\text{ }^{\circ}\text{C}$ prior to shipping to The University of Queensland for analysis. On arrival at The University of Queensland, cartridges were again stored at $-20\text{ }^{\circ}\text{C}$ for no longer than 48 h before elution. A mixture of dichloromethane:isopropanol:aqueous ammonia (80:16:4 v/v/v, 6 mL) was used to elute the analytes from the cartridges and evaporated to approximately 200 μL under nitrogen at $40\text{ }^{\circ}\text{C}$. A solution of 1% HCl in methanol (20 μL) was then added, before being evaporated to dryness. The dry residue was reconstituted with 0.1% formic acid in methanol (20 μL) and 0.1% formic acid in ultrapure water (80 μL) to give a final volume of 100 μL and a concentration factor of 1000 times.

The site in Greece used a slightly different SPE method (Oasis MCX, 3cc, 60 mg). The cartridges were conditioned with methanol (3 mL) and acidified Milli-Q water (3 mL). Following filtration (GF/A 1.6 μm , Whatman, Kent, UK), the acidified wastewater samples (10 mL) were loaded under gravity. The cartridges were then washed with 40:60 acidified methanol: MilliQ water (acidified; 3 mL) and 20:80 acetonitrile:Milli-Q water. The cartridges were then dried for 30 mins under vacuum and shipped to The University of Queensland for analysis. On arrival at The University of Queensland, cartridges were again stored at $-20\text{ }^{\circ}\text{C}$ for no longer than 48 h before elution. The elution and analysis was carried out as in Shimko *et al* (1)

Instrumentation and Data Analysis (for 2019-20 and 2020-21) from Bade *et al* 2020 (2)

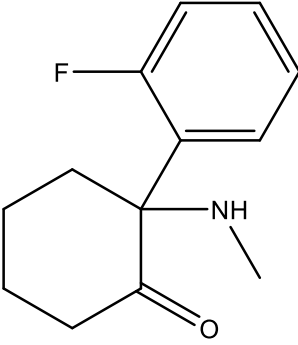
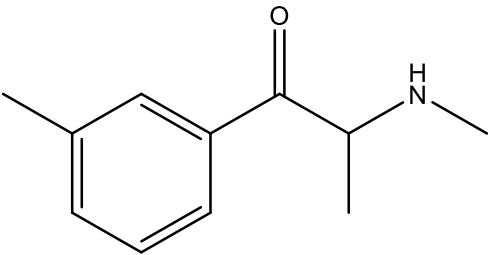
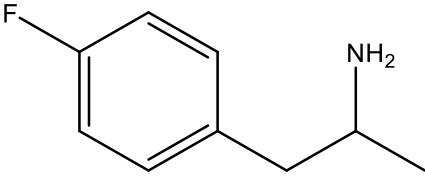
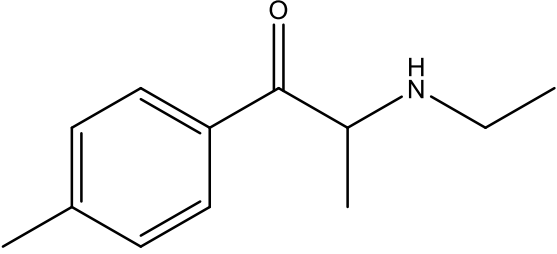
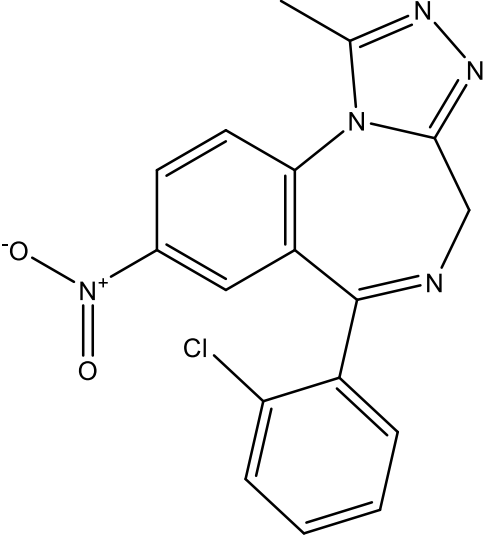
Analyses were performed using a Sciex ExionLC coupled to a Sciex 6500 + QTrap (Toronto, Canada), fitted with a TurboSpray IonDrive source. A Kinetex biphenyl column ($150 \times 2.1\text{ mm} \times 1.7\text{ }\mu\text{m}$) was used for the chromatographic separation at a flow rate of 0.3 mL/min. The injection volume was 2 μL . The mobile phases used were 95% water with 5% methanol and 0.1% formic acid (solvent A) and 95% methanol with 5% water and 0.1% formic acid (solvent B). The initial percentage of B was 2% and increased to 20% over 2 min and 45% over the following 12 min. Over the next 5 min it was increased linearly to 100% and held for 0.1 min before being brought back to the initial percentage and kept steady for the final 2.9 min to equilibrate the system to give a total run time of 22 min. The ion source parameters were as follows: $450\text{ }^{\circ}\text{C}$; curtain gas, 20 psi; collision gas, medium; ion spray voltage, 5500 V; ion source gas 1 and ion source gas 2, 50 psi. Mass spectrometric analyses were performed in positive mode using multiple reaction monitoring (MRM). The two most abundant transitions of the precursor ion for each analyte and one for the deuterated internal standards were monitored. All data were acquired with Analyst 1.7 (Sciex) and processed using MultiQuant 3.0.2.

Instrumentation and Data Analysis (for 2021-22 samples) from Bade *et al* 2022 (3)

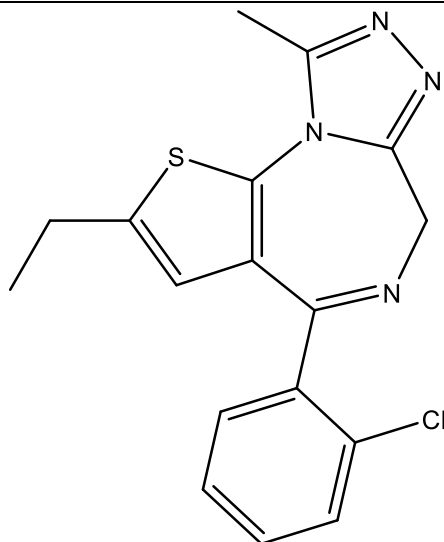
Samples were analysed using a Shimadzu Nexera LC-40 coupled to a SCIEX Triple Quad 7500 system. Chromatographic separation was achieved using a Phenomenex Kinetex Biphenyl (50 × 2.1 mm × 2.6 μm) column fitted with a SecurityGuard ULTRA Cartridges UHPLC Biphenyl 2.1 mm ID columns, at a flow rate of 0.35 mL/min and an injection volume of 2 μL. A mobile phase of 95:5 (v/v) MilliQ water: methanol with 0.1% formic acid (solvent A) and 95:5 (v/v) methanol: MilliQ water with 0.1% formic acid (solvent B) was used. The initial percentage of B was 5%, which was kept steady for the first 2 minutes. The concentration of B was linearly increased to 100% over 11 minutes and held for 2 minutes before being brought back to the starting conditions over 0.1 minutes and kept steady for the final 3.4 minutes to equilibrate the system. The total run time was 18.5 minutes. The mass spectrometer was run in scheduled multiple reaction monitoring (sMRM) mode in positive ion mode, with a 30 second retention time window around each analyte. In total, the mass spectrometric method time was approximately 15.5 minutes. The ion source gas 1 and 2 were set at 60 psi, curtain gas at 40 psi, ion source temperature at 450 °C and ion spray voltage at 2600 V.

For quantification purposes, both transitions needed to be present, while the ion ratio (within 20%) as well as retention time had to compare with the standard (within 2%). If only one transition was present, the compound was deemed at above the limit of detection (LOD) but below the limit of quantification (LOQ). For calculation purposes, this was given as the midpoint between the LOD and LOQ. As no analyte-specific internal standards were used for this method, quantification was based on the peak area ratios between native and surrogate internal standards compared to an external calibration curve. All data were acquired and processed with SCIEX OS.

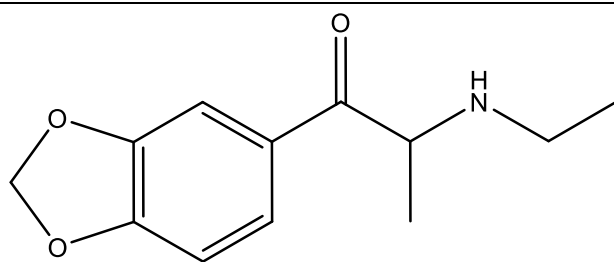
Table S1: Structures of Compounds found

Compound	Structure
2F-Deschloroketamine (2F-DCK)	 <chem>CN1CCCCC1C(=O)C2=CC=C(C=C2)F</chem>
3-Methylmethcathinone (3-MMC)	 <chem>CC(C)C(N)C(=O)c1ccc(C)cc1</chem>
4-Fluoroamphetamine	 <chem>CC(N)CCc1ccc(F)cc1</chem>
4-Methylethcathinone (4-MEC)	 <chem>CCN(CC)C(=O)c1ccc(C)cc1</chem>
Clonazepam	 <chem>CN1CCN2C(=N1)C(=C(C2)c3cc(Cl)cc([N+](=O)[O-])c3)C</chem>

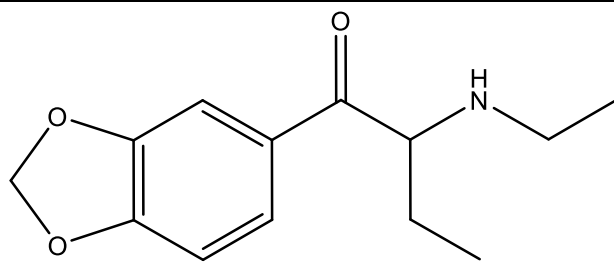
Etizolam



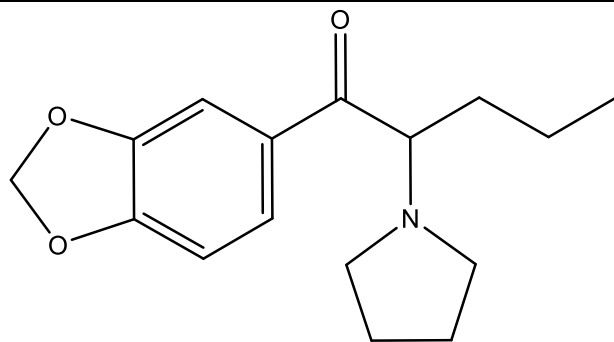
Ethylone



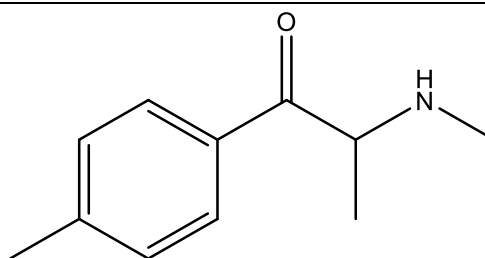
Eutylone

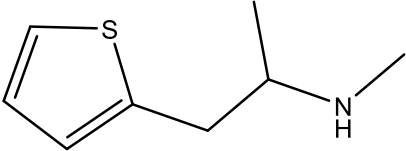
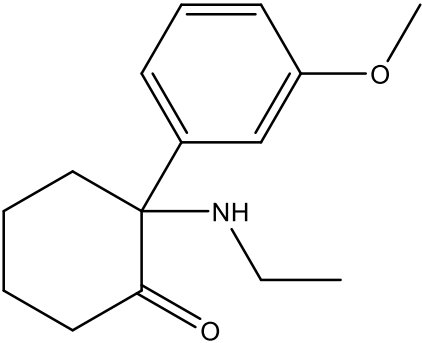
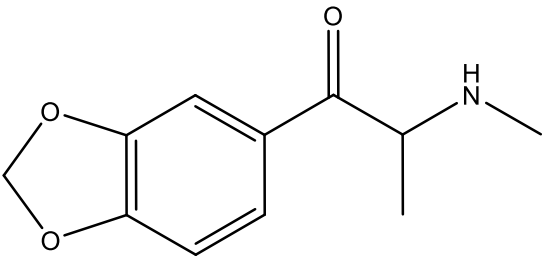
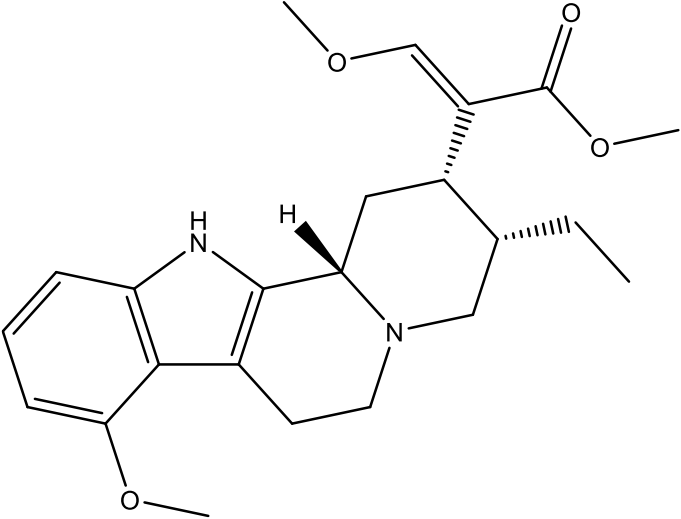
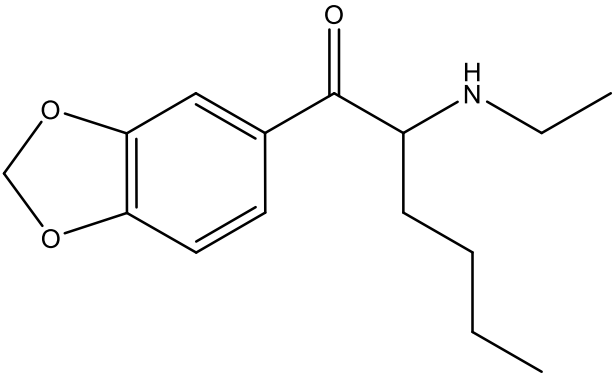


Methylenedioxypropylone (MDPV)

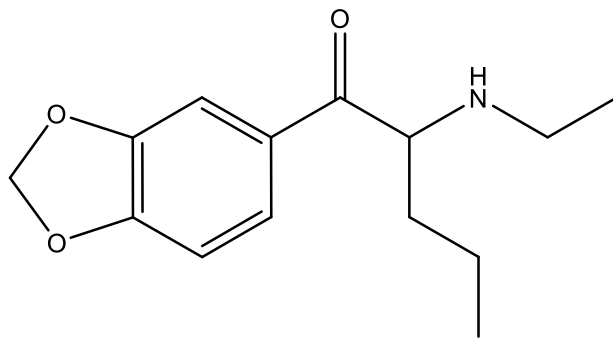


Mephedrone

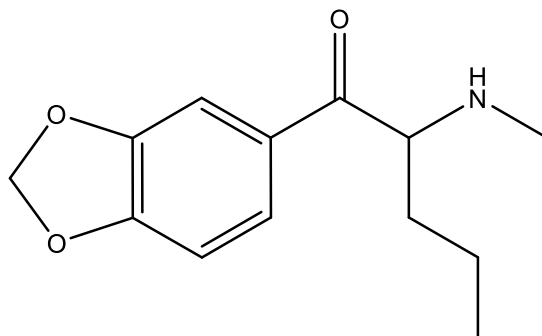


Methiopropamine	
Methoxetamine	
Methylone	
Mitragynine	
N-ethylhexedrone	

N-ethylpentylone



Pentylone



para-Methoxyamphetamine (PMA)

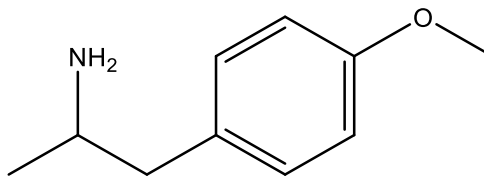


Figure S1: Regional trends in the use of NPS. Note: all sites within a country are aggregated so only a single box-and-whisker plot is shown for each country

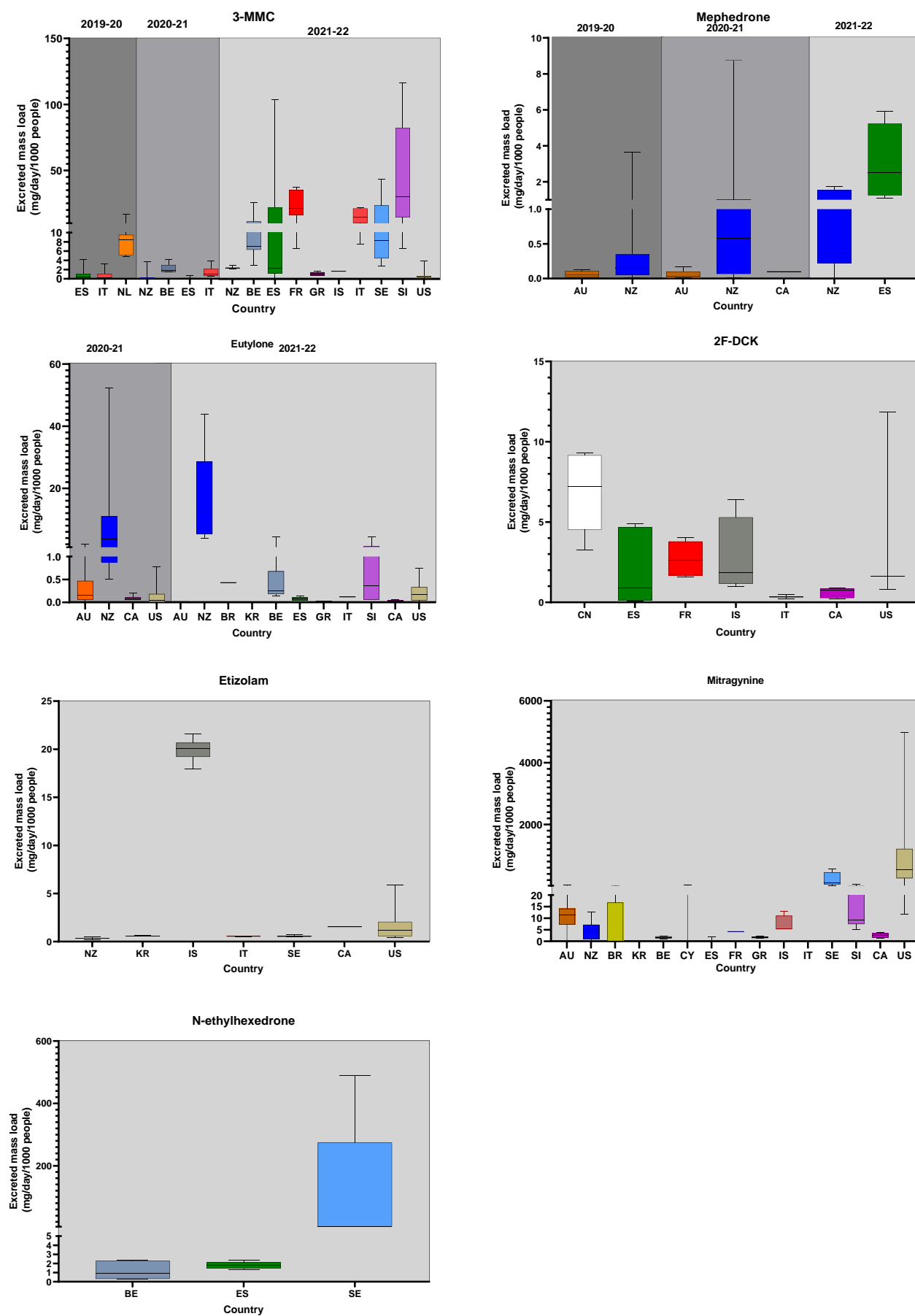


Table S2: Compounds targeted in each campaign

Compound	Sampling campaign		
	2019-20	2020-21	2021-22
25B-NBOMe	√	√	×
25C-NBOMe	√	√	√
25I-NBOMe	√	√	×
2F-Deschloroketamine	×	×	√
2-Methyl AP-237	×	×	√
2-Oxo-PCE	×	×	√
3-Ethylmethcathinone	√	√	×
3-Methylbuphedrone	√	√	×
3-Methylmethcathinone	√	√	√
4-Fluoroamphetamine	√	√	√
4-Fluoromethcathinone	√	√	×
4-Methylbuphedrone	√	√	×
4-Methylethcathinone	√	√	×
5F-EMB-PICA	×	×	√
5F-MDMB-PICA	×	×	√
5F-MDMB-PINACA	×	×	√
7-Hydroxymitragynine	×	×	√
AH-7921	√	√	×
AMB FUBINACA	×	×	√
AP-238	×	×	√
Brorphine	×	×	√
Buphedrone	√	√	×
Butylone	√	√	√
Butyryl fentanyl	√	√	×
Clonazolam	×	×	√
Cumyl pegaclone	×	×	√
Cumyl-5F-pegaclone	×	×	√
Dibutylone	×	×	√

Etizolam	×	×	√
Ethylone	√	√	×
Eutylone	×	√	√
Flualprazolam	×	×	√
Flubromazolam	×	×	√
Furanyl fentanyl	√	√	×
Isotonitazene	×	×	√
MDMB-4en-PINACA	×	×	√
MDPV	√	×	×
Mephedrone	√	√	√
Methcathinone	√	√	√
Methiopropamine	√	√	×
Methoxetamine	√	√	×
Methylone	√	√	√
Metonitazene	×	×	√
Mitragynine	×	×	√
N-Ethylheptedrone	×	×	√
N-Ethylhexedrone	×	×	√
N-Ethylpentylone	√	√	√
Pentylone	√	√	√
PMA	×	√	×
Protonitazene	×	×	√
U-47700	√	√	×
Valeryl fentanyl	√	√	×
Total analysed	26	27	34

Table S3: Flow Rates (in megalitres) and population of all sites in each campaign

	Flow rate (MegaLitres)													
	AU1	AU2	AU3	AU4	BE	BR1	BR2	BR3	CA	CN	CY1	CY2	ES1	ES2
Population	728,759	75,225	155,604	212,309	953,987	145,000	525,000	85,133	2,004,265	250,319 ^a	31,079	205,212	178,141	1,370,216 ^a
23-Dec-21	173.4	24.93	33.3	50.59										
24-Dec-21	175.2	25.4	35.2	53.46										
25-Dec-21	147.8	23.39	30.95	45.96										
26-Dec-21	143.7	23.51	31.17	46.62										
27-Dec-21	148.6	23.43	32.15	47.73										
28-Dec-21	154.2	23.11	32.11	47.87				43.01						
29-Dec-21	157.3	23.02	31.11	48.16	294.486	40.317	105.617	48.90	1573.603	68.326	8.803	25.53	31.868	285.10
30-Dec-21	157.6	22.91	30.86	47.92	271.159	53.735	122.143	41.24	1559.779	63.838	13.264	40.99	31.354	232.53
31-Dec-21	160.4	23.48	31.09	48.37	259.077	49.898	113.999	41.24	1581.552	65.505	6.493	45.72	35.24	262.66
1-Jan-22	140.4	22.62	30.2	45.89	554.537	37.707	101.823	30.75	1719.619	67.028	5.189	26.82	33.854	228.03
2-Jan-22	144.3	23.02	31.18	46.55	590.887	37.45	98.748	41.69	1619.914	63.517	5.204	25.9	32.564	213.83
3-Jan-22					406.129	43.623	111.559	39.16	1454.112	67.953	5.078	27.11	33.063	247.81
4-Jan-22					297.354	41.529	111.945	39.16	1527.466	66.397		26.65	32.811	277.13
5-Jan-22								36.32						
	ES3	FR	GR	IS	IT	KR	NZ1	NZ2	NZ3	NZ4	SE1	SE2	SI1	SI2
Population	87,484	471,326	3,568,758	107,000	1,022,389	922,656	28,736	12,000	39,500	37,000	878,800	200,000	270,305	129,000
23-Dec-21														
24-Dec-21														
25-Dec-21														
26-Dec-21														
27-Dec-21														
28-Dec-21					356.73				10		263.52	34.619	57.27	
29-Dec-21	32.694	482.4	856.9	145.4112	304.16	294.896		2	10	10	293.76	39.624	55.226	9.68
30-Dec-21	29.818	397.9	702.6	146.5344	303.58	289.69		2	10	10	268.704	39.683	54.028	24.024

30-Dec-20	11.32	2.52	11.43	35.02	538.82									
31-Dec-20	11.03	2.57	10.87	32.48	549.03	13.02	1.4	4.66	2.12	53.03	1.67	0.06	46.07	
1-Jan-21	14.26	2.43	10.07	21.39	946.21									
2-Jan-21	12.69	2.32	10.97	19.87	1047									
3-Jan-21	10.06	2.37	9.91		1162.2	13.74	1.4	5.41	2.42	55.83	1.55	0.06	49.82	
4-Jan-21			10.39											
	AU 1	AU 2	AU 3	AU 4	CN	ES	IT	NL	NO	NZ 1	NZ 2	NZ 3	NZ 4	US
Population*	728,759	75,225	212,309	155,604	228,439 ^a	170,888	1,122,501	769,000	624,642 ^b	120,000	19,283	28,736	20,000	893,000
25-Dec-19	139.4	22	46.5	38.2										313.1
26-Dec-19	138.7	22	47.3	30.9	71.4	31.8	286.1			53	1.779	2.6	2.1	301.8
27-Dec-19	151.7	27.8	47.9	30.7	72.7	32.5	327.9			54.4	2.004	2.9	2.3	294.2
28-Dec-19	149.5	21.8	47.7	30.6	71.9	32.5	284.8			54	2.175	3.1	2.5	278
29-Dec-19	138.9	21.7	45.9	30.6	71.9	31	310.7			51.5	2.212	3.3	2.6	274.8
30-Dec-19	153.8	22	47.9	30.8	72.2	31.6	306.5	157		53.8	2.244	3.4	2.7	279.7
31-Dec-19	158.9	22.3	49.5	30.6	71.6	30.5	338.7	162		52.8	2.468	3.8	2.6	328.7
1-Jan-20	158.9	21.4	44.8	29.7	69.8	35.1	286.5	146		48.2	2.549	3.9	2.7	323.9
2-Jan-20	158.9	21.9	47.4	30.5	70.8	31.1	317.6	155		51.4	2.384	3.5	2.6	467.6
3-Jan-20					70.6	31.6	308.5	186		57	2.316	3.4	2.5	
4-Jan-20								160						
6-Jan-20								160						

^a: average population based on chemical parameters

^b: With samples collected over multiple days, population and flow rates were based on the combined days

AU: Australia; BE: Belgium; BR: Brazil; CA: Canada; CN: China; CY: Cyprus; ES: Spain; FJ: Fiji; FR: France; GR: Greece; IS: Iceland; IT: Italy; KR: Republic of Korea; NL: the Netherlands; NZ: New Zealand; SE: Sweden; SI: Slovenia US: United States

References

1. Shimko KM, O'Brien JW, Barron L, Kayalar H, Mueller JF, Tschärke BJ, et al. A pilot wastewater-based epidemiology assessment of anabolic steroid use in Queensland, Australia. *Drug Test Anal* [Internet]. 2019 Jul 2;11(7):937–49. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/dta.2591>
2. Bade R, Abdelaziz A, Nguyen L, Pandopoulos AJAJ, White JMJM, Gerber C. Determination of 21 synthetic cathinones, phenethylamines, amphetamines and opioids in influent wastewater using liquid chromatography coupled to tandem mass spectrometry. *Talanta* [Internet]. 2020 Feb 1;208(August):120479. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0039914019311129>
3. Bade R, Eaglesham G, Shimko KM, Mueller J. Quantification of new psychoactive substances in Australian wastewater utilising direct injection liquid chromatography coupled to tandem mass spectrometry. *Talanta* [Internet]. 2023;251(July 2022):123767. Available from: <https://doi.org/10.1016/j.talanta.2022.123767>