

Fig. S1. Chronic antigenic stimulation *in vitro* induces high per-cell IR expression (A) [left] Histograms and [right] summary data of longitudinal expression of BIM, BCL-2, and KI67 by *in vitro* chronically and acutely stimulated P14 cells. (B) Summary data indicating MFI (gated on IR+ population of CD44<sup>hi</sup> CD8+ live singlets) of IRs after chronic and acute stimulation of P14 cells *in vitro*. Significance calculated by unpaired two-tailed t test; \*\*\*\*p< 0.0001. (C) [left] Representative flow cytometry data of IR expression (gated on CD44<sup>hi</sup> CD8+ live singlets) and [right] cell expansion after chronic and acute stimulation, either via DCs+D<sup>b</sup>GP<sup>33-41</sup> or  $\alpha$ CD3/ $\alpha$ CD28. (D) Representative flow cytometry data of IR expression (gated on CD44<sup>hi</sup> CD8+ live singlets) after chronic stimulation for either 7 or 10 days.



## Fig. S2. In vitro chronically stimulated P14 cells develop a transcriptional signature of $T_{\text{ex}}$

(A) PCA of RNA-seq data from *in vitro* chronically and acutely stimulated P14 cells. (B) Gene expression of leading edge genes upregulated in *in vivo*  $T_{ex}$  enriched after chronic stimulation *in vitro*. (C-D) GSEA of gene sets for (C) *in vitro* activation and (D) exhaustion (41) in *in vitro* chronically stimulated cells at d4 and d7. (E) Heatmap and (F) GO analysis of genes upregulated in *in vivo*  $T_{ex}$  that were not enriched after chronic stimulation *in vitro*. (G) Gene expression of *Tox* in *in vitro* chronically and acutely stimulated P14 cells. (H) [left] TOX protein expression in *in vitro* chronically and acutely stimulated P14 cells and *in vivo*  $T_{ex}$  and  $T_{mem}$ . [right] Fold change of TOX MFI in *in vitro* chronic over acute and *in vivo*  $T_{ex}$  over  $T_{mem}$ . (I) Gene expression of leading edge of genes downregulated in *in vivo*  $T_{ex}$  enriched after chronic stimulation *in vitro*. (J-L) Representative flow cytometry data detailing co-expression of PD-1 and (J) Eomes, (K) TCF1, and (L) Granzyme B on *in vivo*  $T_{ex}$  (LCMV-CI13 30dpi), *in vitro* chronically stimulated P14 cells, Numbers in flow cytometry plots indicate percentage of parent population within each gate.



Fig. S3. In vitro chronically stimulated P14 cells develop epigenetic signatures of  $T_{\text{eff}}$  and  $T_{\text{ex}}$ 

(A) Number of DACRs (filtered on lfc>1 and p<0.05) between pairwise comparisons as indicated. (B) PCA of ATAC-seq data from *in vitro* chronically and acutely stimulated P14 cells. (C) PSEA of naive- and memory-specific ACRs in *in vitro* chronically and acutely stimulated P14 cells. (D) TF motif accessibility in DACRs between *in vitro* chronically stimulated P14 cells and *in vivo* T<sub>ex</sub>.



## Fig. S4. BHLHE40 is predicted to regulate known transcriptional networks associated with CD8 T cell exhaustion

(A) Taiji analysis of differential transcriptional networks downstream of BHLHE40 at d7 of chronic stimulation *in vitro* and d4 of acute stimulation *in vitro*. Shared downstream TFs indicated in white. (B) TF motifs that co-occur with BHLHE40 motifs. (C) ATAC-seq signal tracks for *in vitro* chronically and acutely stimulated P14 cells for the *Runx1* locus; binding motifs for BHLHE40 and BCL6 shown below.



## Fig. S5. BHLHE40 is a novel transcriptional regulator of CD8 T cell differentiation

(A) Experiment schematic of shRNA KD of Bhlhe40 in in vitro chronically stimulated P14 cells. (B) Relative mRNA expression after in vitro Bhlhe40 KD. (C) Representative flow cytometry histograms detailing IR expression by control and Bhlhe40 KD P14 cells (gated on GFP+ CD44<sup>hi</sup> CD8+ live singlets) after chronic stimulation in vitro. Mean fluorescence intensity (MFI) of each population indicated in lower right; representative of 2 experiments. (D) Representative flow cytometry plot of control and Bhlhe40 KD input P14 cells (gated on GFP+ CD44<sup>hi</sup> CD8+ live singlets) before transfer. (E,F) [left] Concatenated flow cytometry plot and [right] summary data of frequency of control and Bhlhe40 KD P14 cells at 8dpi of (E) LCMV-CI13 and (F) LCMV-Arm. (G,I) [left] Concatenated flow cytometry plots and [right] summary data of frequency and total number of KLRG1+ control and Bhlhe40 KD P14 cells at 7dpi of (G) LCMV-Arm or (I) LCMV-CI13. (H) MFI of KLRG1 (gated on KLRG1+ population) in control and Bhlhe40 KD P14 cells at 7dpi of LCMV-Arm. (J) Concatenated flow cytometry plots of effector cytokine production in control and Bhlhe40 KD P14 cells at 8dpi of [top] LCMV-Arm and [bottom] -Cl13. (K) Concatenated flow cytometry histograms detailing IR expression on endogenous (gated on GFP- gp33+ CD44<sup>hi</sup> CD8+ live singlets), control, and *Bhlhe40* KD P14 cells at 31dpi of LCMV-CI13. (L) Summary data indicating MFI (gated on IR+ population) of IRs on control and Bhlhe40 KD P14 cells at 31dpi of LCMV-CI13. (M) Protein expression of BHLHE40 after in vitro overexpression (OE) and KD of Bhlhe40; \* indicates non-specific binding. (N) Concatenated flow cytometry plots and (O) summary data of frequency of Tex subsets in control and Bhlhe40 OE P14 cells at 31dpi of LCMV-CI13. (P) Frequency and total number of progenitor and intermediate Tex in control and Bhlhe40 KD P14 cells at 37dpi of LCMV-CI13, after treatment with vehicle control or  $\alpha$ PD-L1 from 22-35dpi. (E-L.N-P) n=5 mice (Arm), n=10 mice (CI13), representative of 3 experiments. Significance calculated by paired two-tailed t test; \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001. (D-J,N) Numbers in flow cytometry plots indicate percentage of parent population within each gate.