Supplementary material

Minimum Reporting Standards in MR Spectroscopy checklist (accord	ling to Lin et al. NMR Biomed 2021)
1. Hardware	
a. Field strength [T]	3T & 7T
b. Manufacturer	Siemens
c. Model (software version if available)	Prisma Fit & Terra dot Plus
d. RF coils: nuclei (transmit/ receive), number of channels, type, body part	3T: 1H RX 64 channels, head, Siemens 1H TX: body coil, 7T: 1H 1 Tx/ 32 channel RX Nova Medical, 2H/1H birdcage, Stark Contrast
e. Additional hardware	N/A
2. Acquisition	
a. Pulse sequence	3T: 3D FID-CRT MRSI 7T: 3D FID-Phase Encoding MRSI
b. Volume of Interest (VOI) locations	3T: The excited 55 mm-thick slab was centered around the posterior cingulate region. 7T: whole-brain
c. Nominal VOI size [cm³, mm³]	3T: 200×200×55 mm ³ 7T: (FOV) 200x200x175 mm ³
d. Repetition Time (TR), Echo Time (TE) [ms, s]	3T: TR=950 ms / 0.8 ms acquisition delay
e. Total number of Excitations or acquisitions per spectrum	3T: 1 average 7T: 2 averages
In time series for kinetic studies	N/A
i. Number of Averaged spectra (NA) per time-point	N/A
ii. Averaging method (e.g. block-wise or moving average)	N/A
iii. Total number of spectra (acquired / in time-series)	N/A
f. Additional sequence parameters (spectral width in Hz, number of spectral points, frequency offsets); If STEAM: Mixing Time TM; If MRSI: 2D or 3D, FOV in all directions, matrix size, acceleration factors	3T: BW: 1325 Hz, 588 spectral points, 3D FOV: 200×200x130 mm ³ , grid size: 32x32x21 7T: BW: 500 Hz, 128 spectral points, 3D FOV: 200x200x175 mm ³
g. Water Suppression Method	3T: WET
h. Shimming Method, reference peak, and thresholds for "acceptance of shim" chosen	3T: Standard shim + manual adjustment, water peak < 30 Hz 7T: Standard shim + manual adjustment, water peak < 40 Hz
i. Triggering or motion correction method (respiratory, peripheral, cardiac triggering, incl. device used and delays)	N/A
3. Data analysis methods and outputs	
a. Analysis software	LCModel 6.3-1
b. Processing steps deviating from quoted reference or product	N/A
c. Output measure (e.g. absolute concentration, institutional units, ratio)	absolute concentration, ratio
d. Quantification references and assumptions, fitting model assumptions	Simulated in NMRScope-B, macromolecular background for 3T
4. Data Quality	
a. Reported variables (SNR, Linewidth (with reference peaks))	3T: SNR was calculated using the pseudoreplica method, and linewidth as FWHM of the NAA fit 7T: SNR was calculated by LCModel (peak height/residuum)
b. Data exclusion criteria	¹ H 3T: SNR<15, FWHM<0.1 ppm, CRLBs > 20% for tCr and Glu+Gln (Glx ₄), > 50% threshold for Glc ₆ ² H 7T: CRLBs > 50% for water, Glc ₆ , Glx ₄ , no CRLB threshold for first 3 time points (first 20min)
c. Quality measures of postprocessing Model fitting (e.g. CRLB, goodness of fit, SD of residual)	CRIB
d. Sample Spectrum	See Figure 8

Supplementary Table 1:

Minimum Reporting Standards for in vivo MR Spectroscopy

Note. – Parameters 3T QELT / **7T DMI (BOLD),** CRLB = Cramér-Rao lower bounds; FID = free induction decay; CRT = concentric ring trajectory; FOV = field of view; FWHM = full-width-at-half-maximum; Glu = Glutamate; Gln = Glutamine; Glc = Glucose; tNAA = total N-acetylaspartate; SNR = signal-to-noise ratio; tCr = total creatine; VOI = volume of interest.

²H DMI:

$$[M_{Abs}] = \frac{Amplitude_{M}}{Amplitude_{Water}} * \frac{f_{GM} * d_{GM} * R_{water_GM} + f_{WM} * d_{WM} * R_{water_WM} + f_{CSF} * d_{CSF} * R_{water_CSF}}{(1 - f_{CSF}) * R_{M}}$$

$$* 17.2mM * \frac{N_{water}}{N_{M}}$$

$$R_{M} = e^{-T_{E}/T_{2}} * (1 - e^{-T_{R}/T_{1}})$$

¹H QELT:

$$[M_{Abs}] = \frac{Amplitude_{glx}}{Amplitude_{tCr}} * \frac{f_{GM} * R_{tCr_GM} + f_{WM} * R_{tCr_WM} * WM_{factor}}{(f_{GM} * R_{glx_GM} + f_{WM} * R_{glx_WM})} * 7.5mM$$

$$R_M = e^{-T_E/T_2} * (1 - e^{-T_R/T_1})$$

Supplementary Figure 1:

Concentration estimation in mM units of ²H resonances detected using DMI at 7T. Metabolite amplitudes ($Amplitude_M$) were referenced to deuterated water signals ($Amplitude_{Water}$) and corrected for relaxation times R_M and voxel-wise fractional water content for GM and WM and CSF tissue (f_{GM} , f_{WM}) with d_{GM} =0.78, d_{WM} =0.65 and d_{CSF} =0.97.

Concentration estimation in mM units of ¹H resonances detected using QELT MRS at clinical 3T. Glx amplitudes ($Amplitude_{Glx}$) were referenced to total creatine signals ($Amplitude_{tCr}$) and corrected for relaxation times R_M and voxel-wise fractional GM and WM content (f_{GM} , f_{WM}). WM content was corrected by a factor of 5.7/7.5 relative to the absolute mM concentration of tCr in GM/WM.



Supplementary Figure 2:

Averaged time courses given as (mean+-SD) of deuterated water signal resonances detected using deuterium metabolic imaging (DMI) at 7T, averaged over gray matter (GM, blue) and white matter (WM, red) dominated regions. One subject was scanned only for 9 time points and with 16% increased nominal voxel volume, which could explain the smaller standard deviation and lower mean value for the last time point.



Supplementary Figure 3: Time courses of signal to noise ratio (SNR) maps of deuterated water and Cramer-Rao Lower Bounds maps of ²H Glx₄, Glc₆ and deuterated water from one representative participant for all time points acquired using ²H DMI at 7T.



Supplemental Figure 4: Time courses of signal to noise ratio (SNR) maps of total N-acetyl aspartate and Cramer-Rao Lower Bounds maps of ¹H Glx_4 , Glc_6 and total creatine from one representative participant for all time points acquired using ¹H QELT at clinical 3T.



Supplementary Figure 5: Time courses of Cramer-Rao Lower Bounds for relevant metabolites acquired with 7T ²H DMI (a) and 3T ¹H QELT (b) averaged over GM+WM voxels for each 3D dataset.



Supplementary Figure 6:

34±2 min after oral consumption of deuterium labeled glucose averaged blood plasma glucose

concentration increased significantly (p=0.033) from 86±7 mg/dl to 126±25 mg/dl before gradually

decreasing to 106±17 mg/dl at 100±1 min.