

Fig. S1 Proposed carotenoid biosynthetic pathways in *N. oceanica*. CDP-ME, 4-diphosphocytidyl-2-C-methylerythritol; CDP-MEP, 4-diphosphocytidyl-2-C-methyl-D-erythritol 2-phosphate; CMK, 4-diphosphocytidyl-2-C-methyl-D-erythritol kinase; CMS, 2-C-methyl-D-erythritol 4-phosphate cytidyltransferase; CRTISO, carotenoid isomerase; CYP97, cytochrome P450 beta-hydroxylase; DMAPP, dimethylallyl pyrophosphate; DXR, 1-deoxy-D-xylulose 5-phosphate reductoisomerase; DXP, 1-deoxy-D-xylulose 5-phosphate; DXS, 1-deoxy-D-xylulose 5-phosphate synthase; GAP, glyceraldehyde 3-phosphate; GGPP, geranylgeranyl diphosphate; GGPPS, geranylgeranyl diphosphate synthase; HDR, 4-hydroxy-3-methylbut-2-en-1-yl diphosphate reductase; HDS, 4-hydroxy-3-methyl-but-2-en-1-yl diphosphate synthase; HMB-PP, (E)-4-Hydroxy-3-methyl-but-2-enyl pyrophosphate; IPP, isopentenyl pyrophosphate; IPP1, isopentenyl-diphosphate Delta-isomerase; LCYB, lycopene beta cyclase; MCS, 2-C-methyl-D-erythritol 2,4-cyclodiphosphate synthase; MEcPP, 2-C-methyl-D-erythritol 2,4-cyclodiphosphate; MEP, 2-C-methylerythritol 4-phosphate; PDS, phytoene desaturase; PSY, phytoene synthase; VDE, violaxanthin de-epoxidase; VDL, violaxanthin de-epoxidase like; ZDS, zeta-carotene desaturase; ZEP, zeaxanthin epoxidase; ZISO, zeta-carotene isomerase

■ exon

— intron

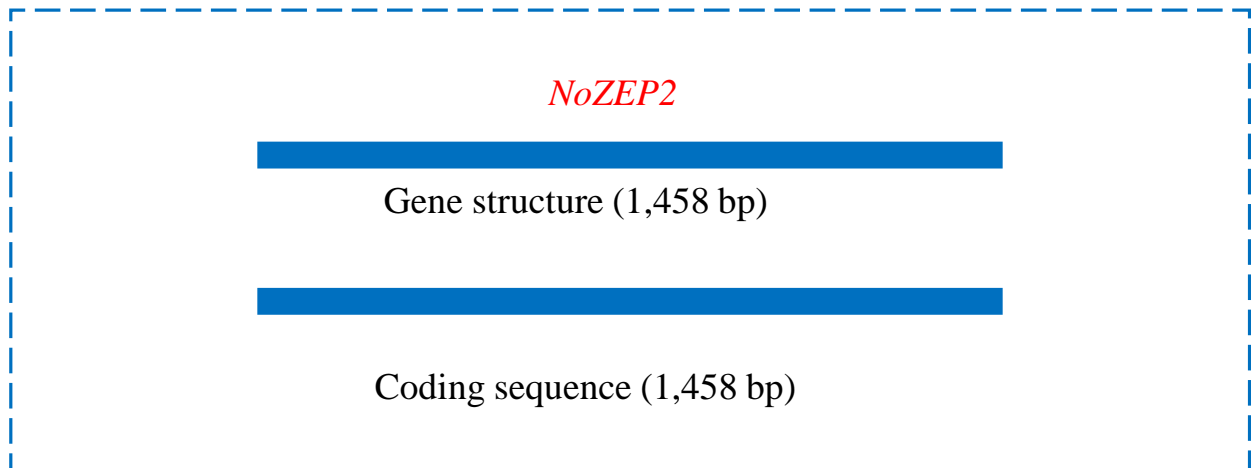
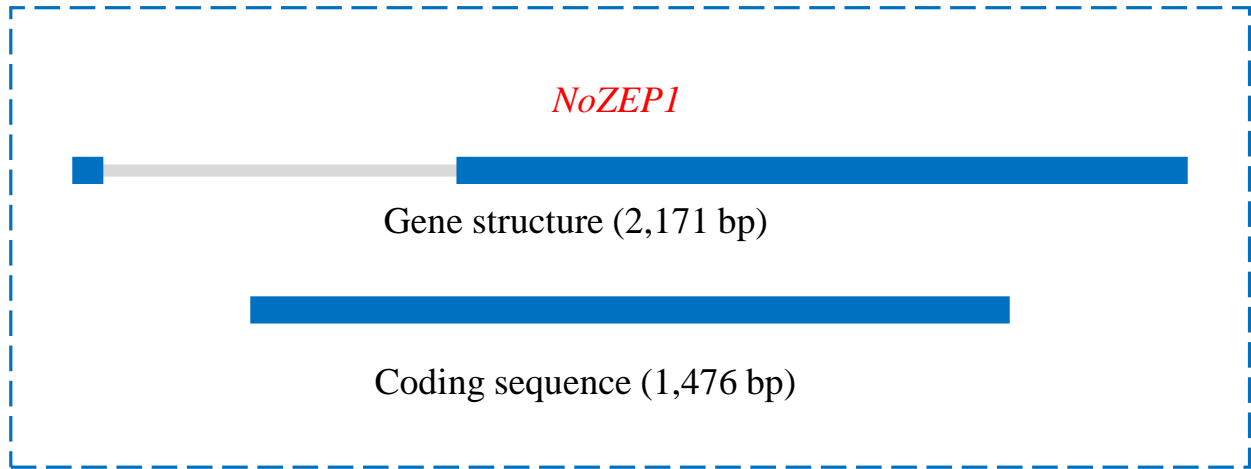


Fig. S2 Gene structures of *NoZEP1* and *NoZEP2*

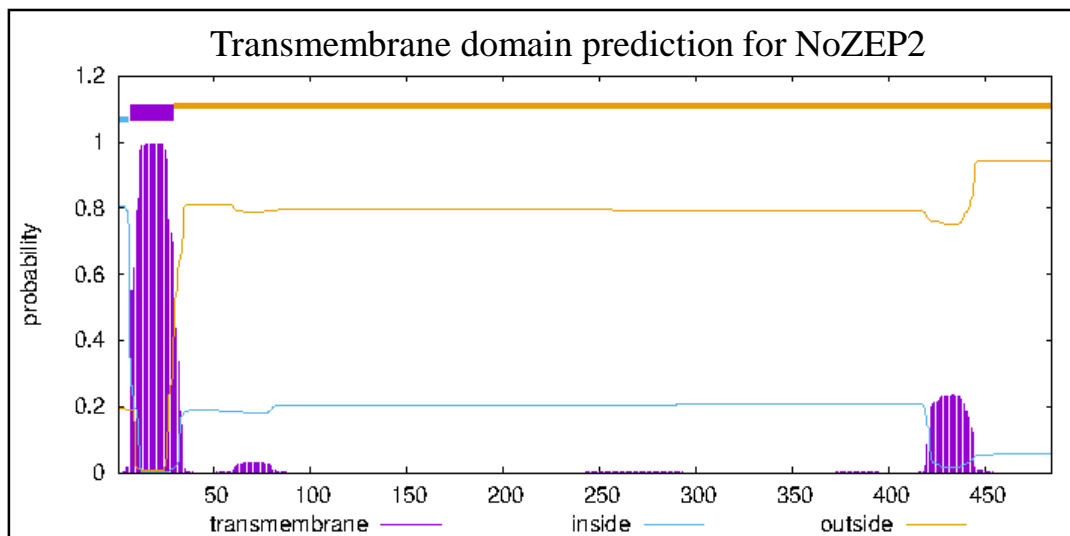
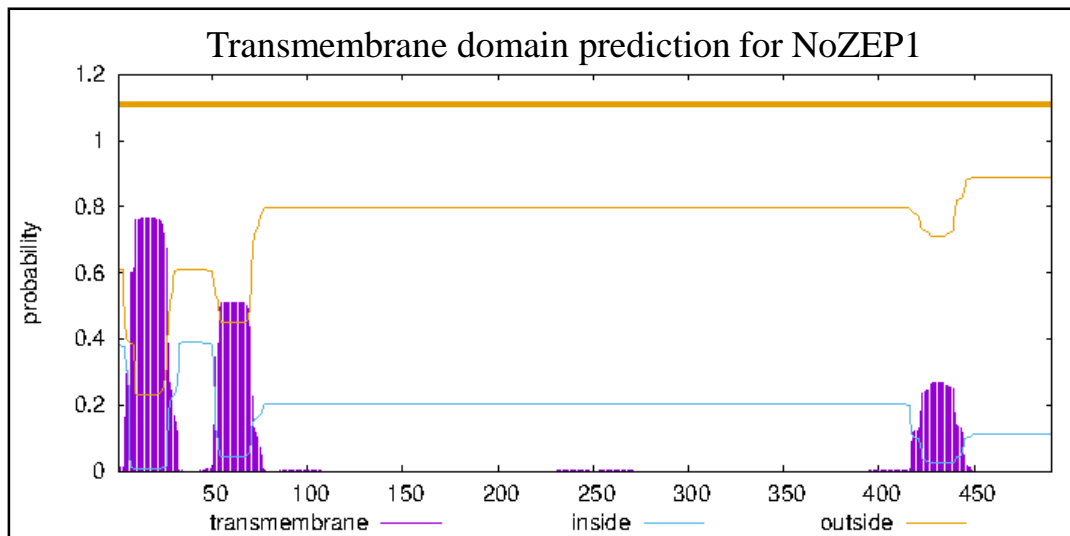
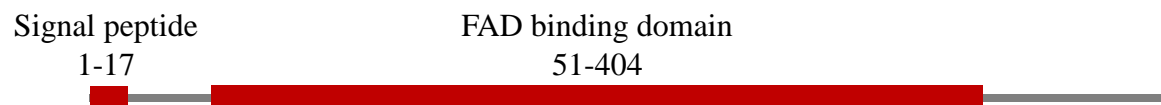


Fig. S3 Transmembrane domains prediction for NoZEP1 and NoZEP2 by TMHMM (V2.0, <http://www.cbs.dtu.dk/services/TMHMM/>)



NoZEP1



NoZEP2

Fig. S4 Predicted domains of NoZEP1 and NoZEP2 via SMART analysis (<http://smart.embl-heidelberg.de/>)

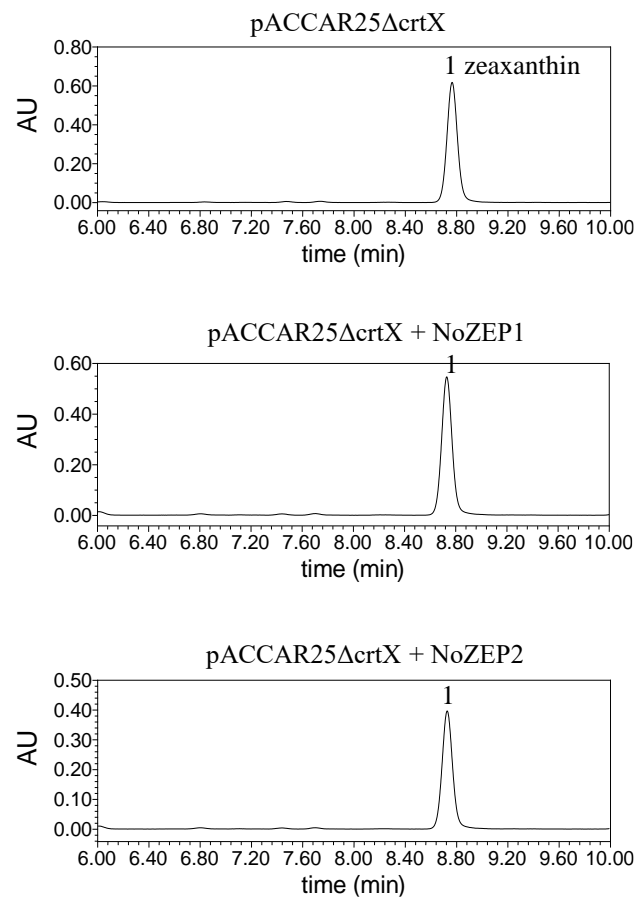


Fig. S5 HPLC elution profiles of carotenoids produced in the zeaxanthin-producing *E. coli* strain without (top) or with *NoZEPs* (middle and bottom). The absorbance was recorded at 450 nm.

HECTAR v1.3 (<https://webtools.sb-roscoff.fr/>)

Protein id	Predicted targeting category	Signal peptide score	Signal peptide cleavage site	Chloroplast score	Mitochondrion score
NoZEP1	Chloroplast	0.8244	18	0.7578	-
NoZEP2	Signal peptide	0.8271	34	0.3599	-

Cell-Ploc 2.0 (<http://www.csbio.sjtu.edu.cn/bioinf/Cell-PLoc-2/>)

Protein id	Predicted location
NoZEP1	Chloroplast
NoZEP2	Chloroplast

Fig. S6 Subcellular localization predication for NoZEP1 and NoZEP2



Fig. S7 Cultures of WT and *NoZEP*-knockdown lines from two days of HL in Fig. 4

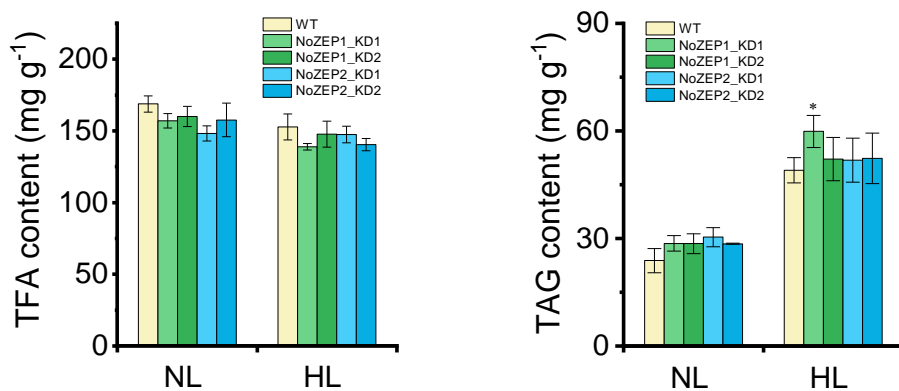


Fig. S8 The content of TFA and TAG as affected by *NoZEP1* or *NoZEP2* knockdown in *N. oceanica* under NL and HL conditions. The NL and HL samples were from day 4 and day 2, respectively. Data represent mean values \pm SD ($n = 3$). The asterisk indicates the significant difference (Student's *t*-test, $P < 0.05^*$) between WT and knockdown lines.

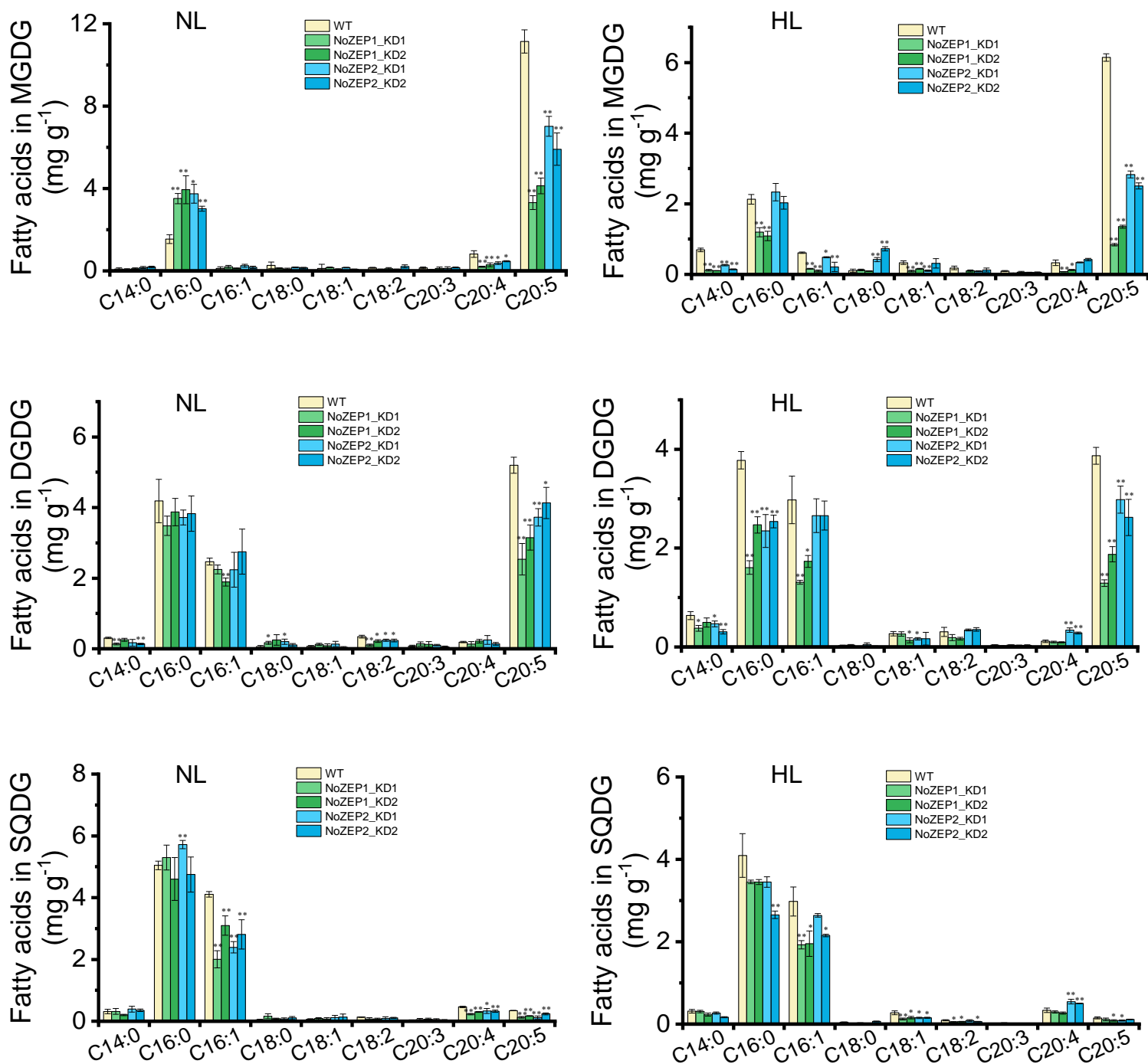


Fig. S9 The content of individual fatty acids of MGDG, DGDG, and SQDG as affected by *NoZEP1* or *NoZEP2* knockdown in *N. oceanica* under NL and HL conditions. The NL and HL samples were from day 4 and day 2, respectively. Data represent mean values \pm SD ($n = 3$). The asterisk indicates the significant difference (Student's *t*-test, $P < 0.05^*$ or $P < 0.01^{**}$) between WT and knockdown lines.

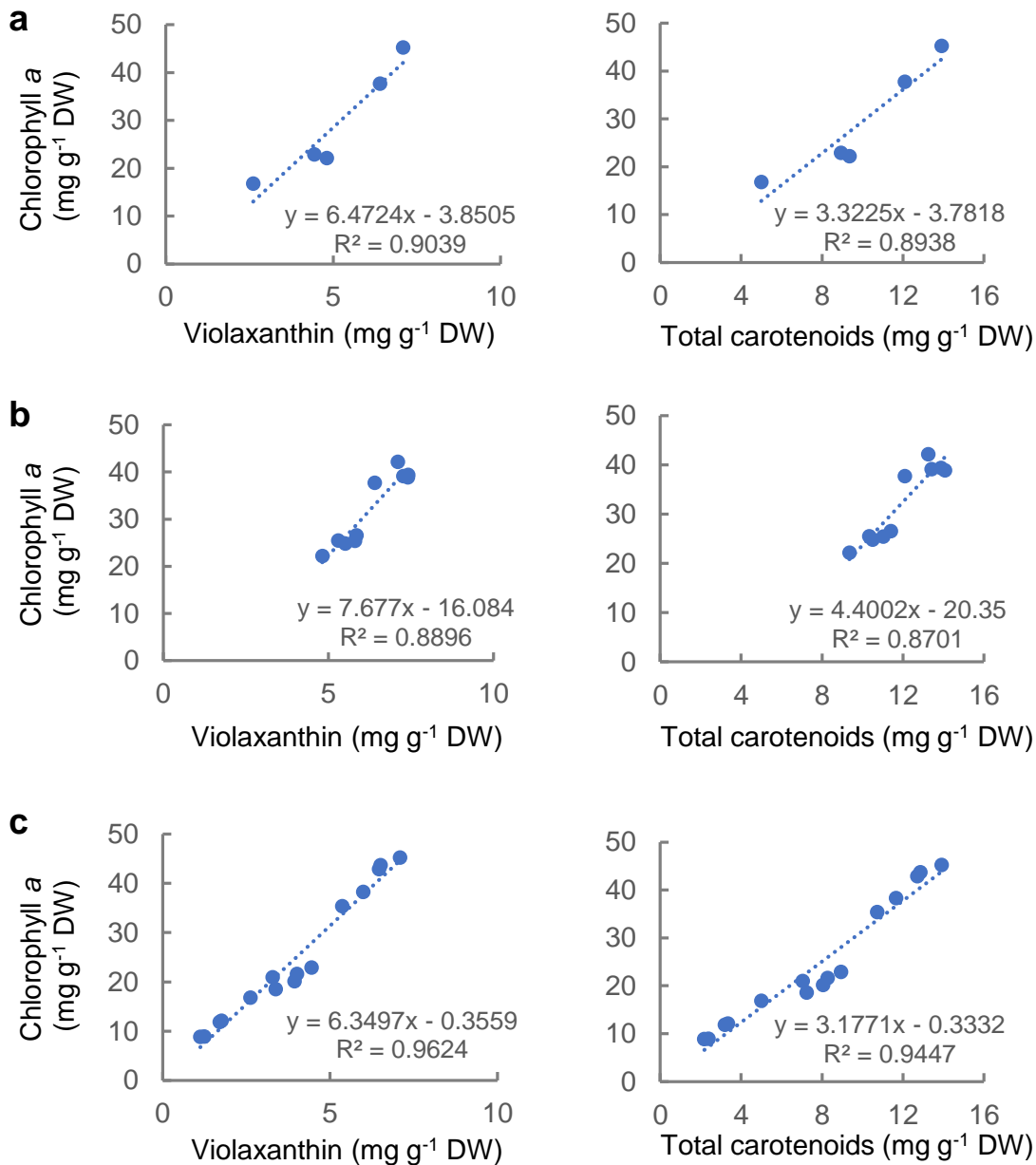


Fig. S10 Correlation between chlorophyll *a* and violaxanthin and between chlorophyll *a* and total carotenoids in *N. oceanica*. (a) WT under different light conditions. Data were retrieved from Figs 3-5. (b) WT and overexpression lines. Data were retrieved from Fig. 3. (c) WT and knockdown lines. Data were retrieved from Figs 4 and 5.

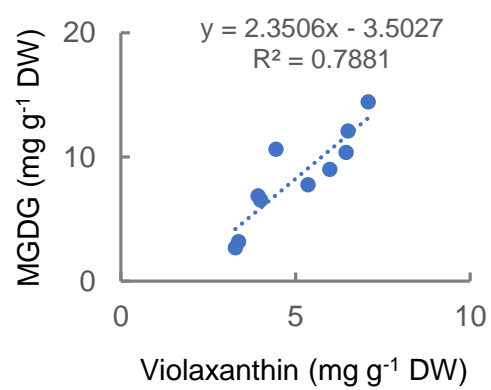
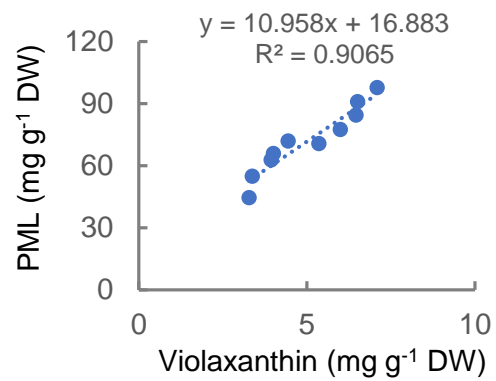
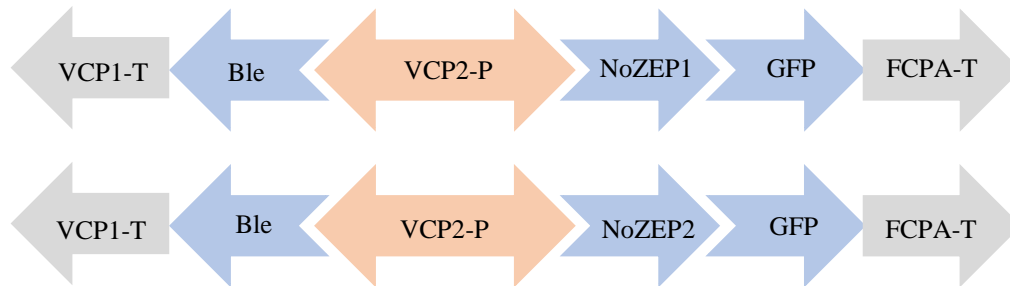


Fig. S11 Correlation between membrane lipids and violaxanthin in *N. oceanica*. Data were retrieved from Figs 4 and 6. PML, polar membrane lipids.

Vectors for *NoZEP1* and *NoZEP2* subcellular localization and overexpression



Vectors for *NoZEP1* and *NoZEP2* knockdown

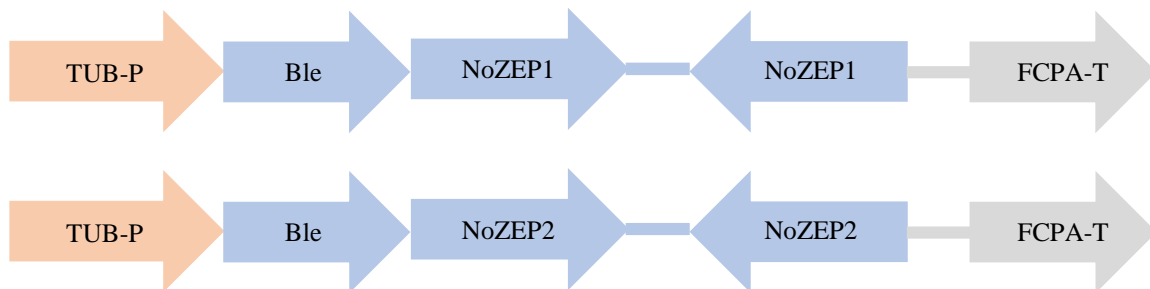


Fig. S12 Illustration of vectors used in this study for *N. oceanica* transformation

Table S1. Primers used in the present study

Primer name	Sequence (5' to 3')	Purpose
NoZEP1-F1 (<i>Bam</i> HI)	<u>taggatcc</u> ATGGTTCAGCTGGGGACTTTG	Cloning NoZEP1 CDS into pUC19 vector
NoZEP1-R1 (<i>Eco</i> RI)	<u>cggaatc</u> CTAGACAGCAACCGTCTCTTT	
NoZEP2-F1 (<i>Bam</i> HI)	<u>taggatcc</u> ATGACGGGTAGGAGAAGTGGT	Cloning NoZEP2 CDS into pUC19 vector
NoZEP2-R1 (<i>Xho</i> I)	<u>gactcgag</u> CATGAGGGGAAACATACGCCT	
NoZEP1-F2 (<i>Kpn</i> I)	accactctctcaagt <u>ggtacc</u> ATGGTTCAGCTGGGGACTTTG	Cloning NoZEP1 CDS into the overexpression vector and PCR verification
NoZEP1-R2 (<i>Eco</i> NI)	accattcctcctcctcctccctaggGACAGCAACCGTCTCTTT	
NoZEP1-F3	GGGGGTCCGATTCAAGTGCA	
NoZEP1-R3	TATAGGCAACCTCCTTCACG	
NoZEP2-F2 (<i>Kpn</i> I)	accactctctcaagt <u>ggtacc</u> ATGACGGGTAGGAGAAGTGG	Cloning NoZEP2 CDS into the overexpression vector and PCR verification
NoZEP2-R2 (<i>Eco</i> NI)	accattcctcctcctcctccctaggCATGAGGGGAAACATACG	
NoZEP2-F3	CGGACCTGGAGCGTCGAGGC	
NoZEP2-R3	ACACCACGAATGGCCTGATA	
NoZEP1-F4 (<i>Bam</i> HI)	<u>cgggatcc</u> TCGACCAGAGTCGTCTGCACAC	Constructing the knockdown vector of NoZEP1
NoZEP1-R4 (<i>Xba</i> I)	<u>gctctaga</u> CGGTTCGTACAAATCCCTCTGGT	
NoZEP1-F5 (<i>Xba</i> I)	<u>gctctaga</u> TGCATTCGACCCTTCCCCACAT	
NoZEP1-R5 (<i>Eco</i> RI)	<u>cggaatc</u> TCGACCAGAGTCGTCTGCACAC	
NoZEP2-F4 (<i>Bam</i> HI)	<u>cgggatcc</u> ATGCGTAGCGACCTGTTACGCC	Constructing the knockdown vector of NoZEP2
NoZEP2-R4 (<i>Xba</i> I)	<u>gctctaga</u> GCAGGCTTCGAACTTTCTGCAG	
NoZEP2-F5 (<i>Xba</i> I)	<u>gctctaga</u> GCCTGATAACCGGCATAGTTGG	
NoZEP2-R5 (<i>Eco</i> RI)	<u>cggaatc</u> ATGCGTAGCGACCTGTTACGCC	
NoZEP1-F6	GGATTGCGTCTGACGTGTTG	RT-qPCR for NoZEP1
NoZEP1-R6	CAGAACCCAGTCGCTCTCA	
NoZEP2-F6	GCCACATGAATTCCGGTACAT	RT-qPCR for NoZEP2
NoZEP2-R6	CCGCAAGACAACCTCGGCTAT	
NoActin-F	GCCGTTATTGGATGGATATG	RT-qPCR for NoActin
NoActin-R	ACAACAACCTCTCCTTCACA	