

## SUPPLEMENTARY INFORMATION

### **Type I Interferon Response in Astrocytes Promotes Brain Metastasis by Enhancing Monocytic Myeloid Cell Recruitment**

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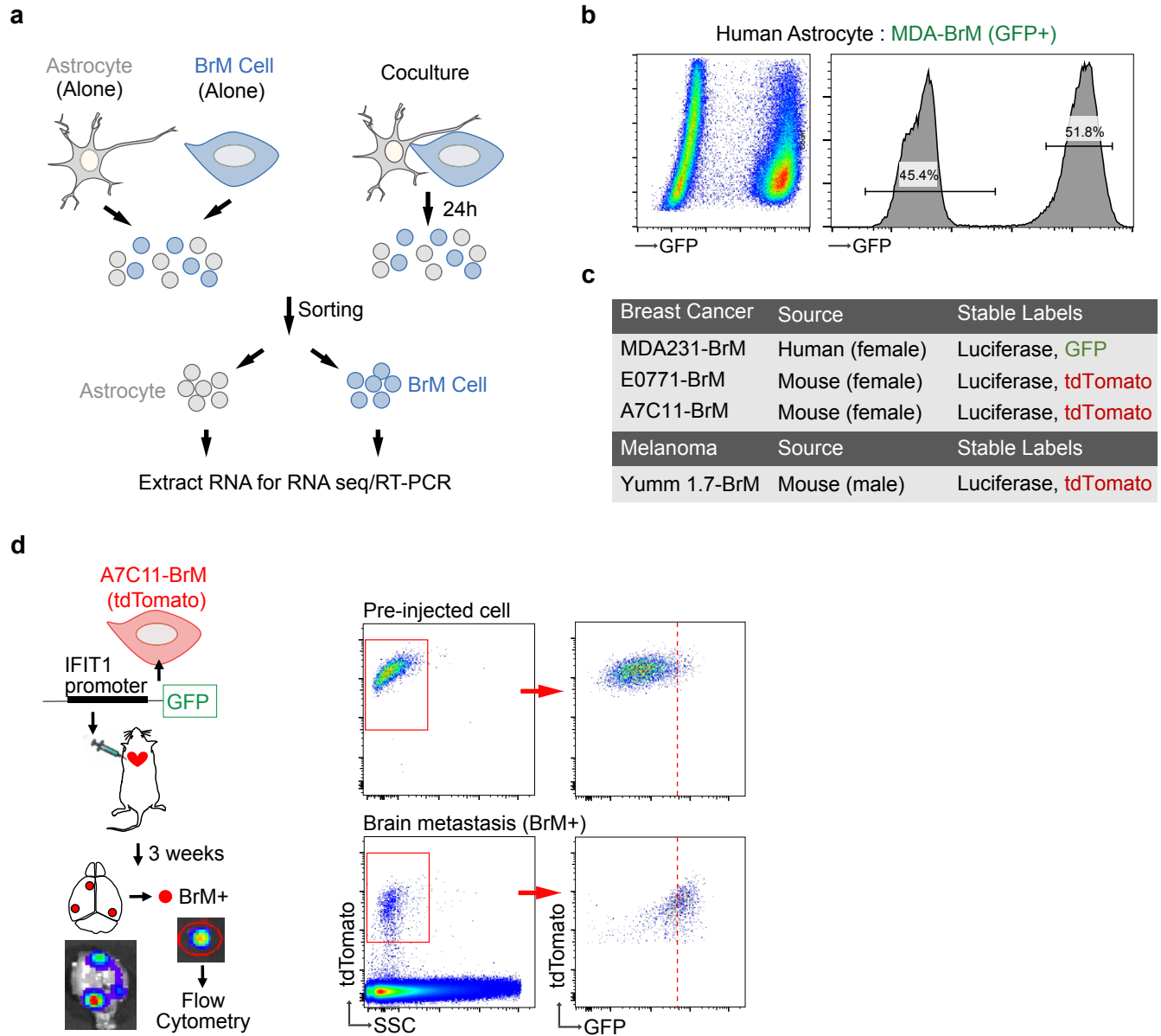
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**Supplementary Figures 1–6**

**Supplementary Tables 1-4**

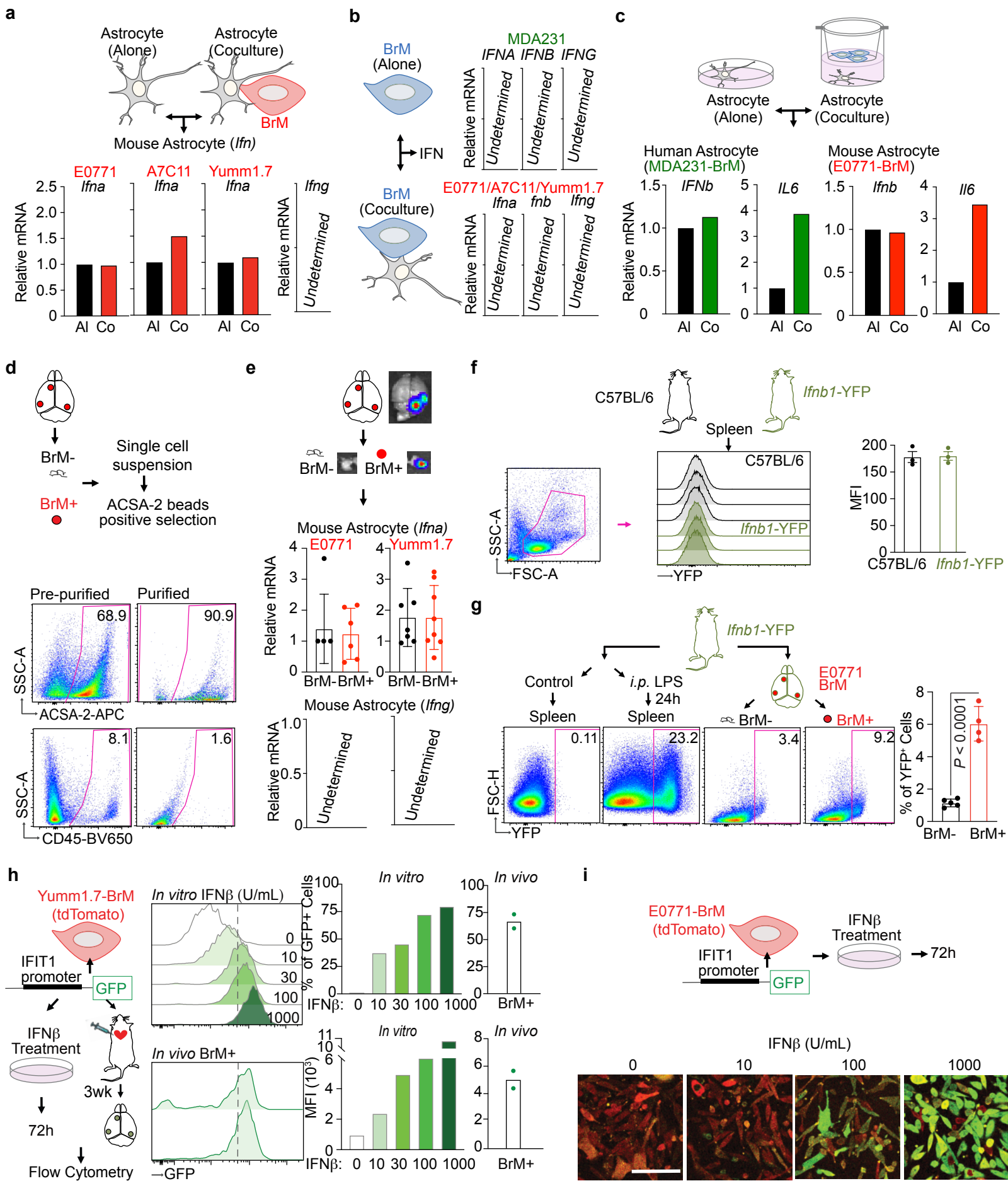


**Supplementary Fig. 1 IFN signaling is activated in brain metastasis.**

**a,b,** *In vitro* astrocyte-BrM coculture experiments. **(a)** Scheme of experiment setup. **(b)** Representative flow cytometry profiles of sorted astrocytes and BrM cells.

**c,** BrM cells used for *in vitro* coculture and *in vivo* brain metastasis experiments, including newly established E0771-BrM, A7C11-BrM and previously established MDA231-BrM and Yumm1.7-BrM.

**d,** IFIT1-GFP reporter structure is expressed in A7C11-BrM cells. 3 weeks after injecting the reporter BrM cells, macrometastatic lesions are isolated based on the luciferase signals in bioluminescent images (BLI), defined as BrM<sup>+</sup> tissues. GFP expression in tdTomato<sup>+</sup> BrM cells is detected by flow cytometry.



## Supplementary Fig. 2 Low level of type I IFN signaling in brain metastasis microenvironment.

**a**, No change of IFN $\alpha$  in mouse astrocytes after cocultured with BrM cells *in vitro*. No detectable IFN $\gamma$  expression in mouse astrocytes. RT-PCR results of *Ifna* and *Ifng* genes. Representative data shown from 3 biologically independent experiments for each BrM cell.

**b**, No detectable IFN cytokines in human or mouse BrM cells by RT-PCR.

**c**, Contact-independent coculture of astrocyte and BrM cells. RT-PCR results of the expression of IFN $\beta$  and IL-6 the astrocytes cocultured with BrM cells. Representative data shown from 3 biologically independent experiments for each BrM cell.

**d**, Astrocyte isolation. Representative results from one astrocyte purification experiment using a macrometastatic lesion established by E0771-BrM cells. ACSA-2 and CD45 are detected by flow cytometry to measure the percentage of astrocytes and immune cells, respectively, before and after purification.

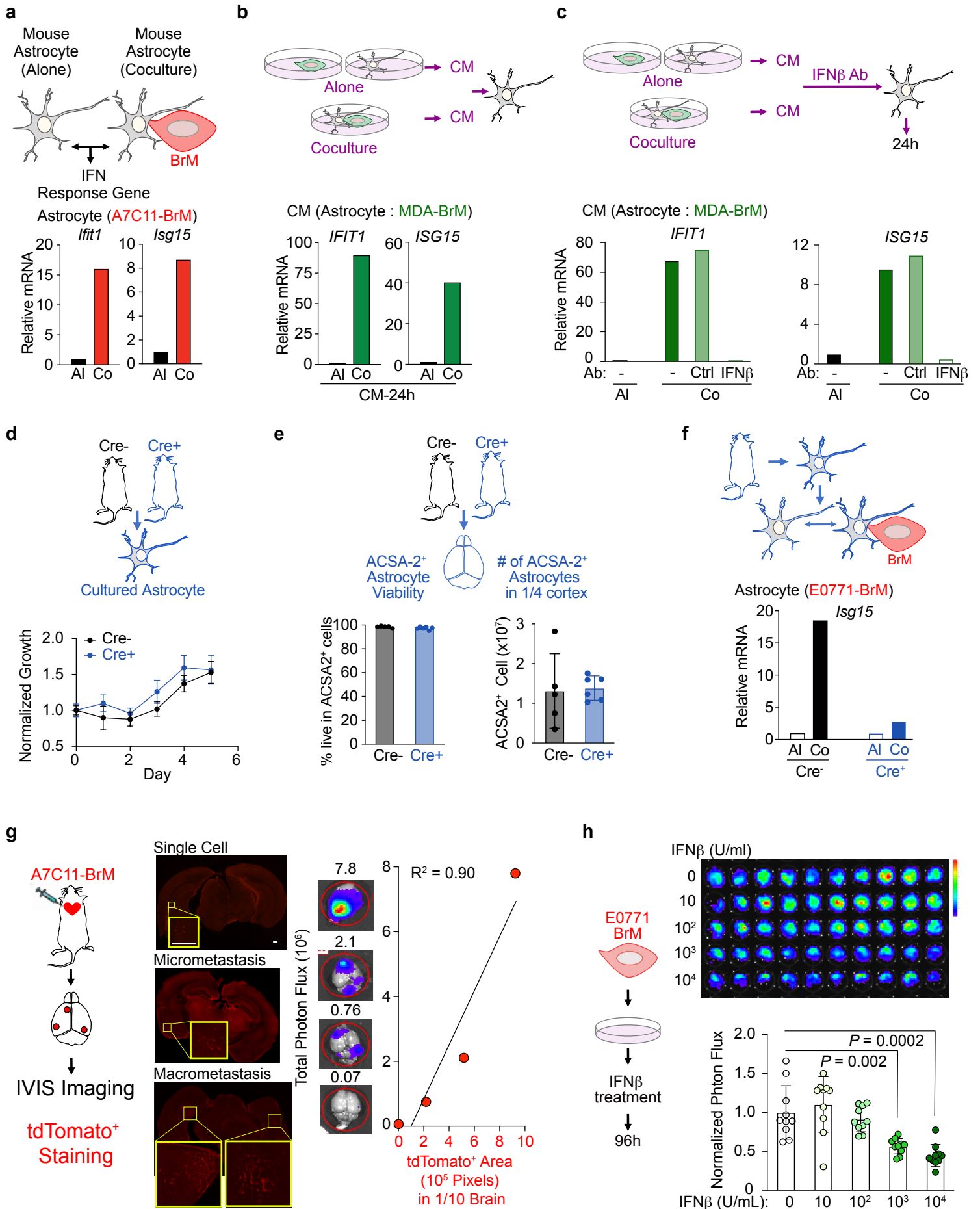
**e**, IFN $\alpha$  and IFN $\gamma$  production in metastasis-activated astrocytes *in vivo*. From the experimental mice developed with brain metastasis, metastatic lesions (BrM+) and metastasis-free tissues (BrM-) are isolated based on the luciferase signals in BLI. Astrocytes are purified by ACSA-2 magnetic beads and *Ifna* and *Ifng* expression is detected by RT-PCR. Data are from merged samples from at least 3 biologically independent experiments and presented as mean  $\pm$  S.D. E0771 model (BrM-: n = 4, BrM+: n = 6). Yumm1.7 model (BrM-: n = 7, BrM+: n = 8). Source data are provided as a Source Data file.

**f**, Basal YFP level in *Ifnb1*-YFP reporter mice. YFP signal in the splenocytes from wild type (WT) and *Ifnb1*-YFP are detected and quantified by flow cytometry. n = 3 mice. Data are presented as mean  $\pm$  S.D. Source data are provided as a Source Data file.

**g**, Defining YFP<sup>+</sup> cells in *Ifnb1*-YFP reporter mice. Splenocytes from control and LPS treated mice are used to gate YFP<sup>+</sup> cells. From the experimental mice with developed brain metastasis, metastatic lesions (BrM+) and metastasis-free tissues (BrM-) are isolated based on the luciferase signals in BLI. Percentages of YFP<sup>+</sup> cells out of total live cells are shown in the bar graph. Data are presented as mean  $\pm$  S.D. (BrM-: n = 5, BrM+: n = 4). *P* value is the result from unpaired two-tailed t test. Source data are provided as a Source Data file.

**h**, IFIT1-GFP reporter structure is expressed in Yumm1.7-BrM cells. The reporter cells treated with various concentrations of IFN $\beta$  *in vitro* and GFP expression is quantified. 3 weeks after injecting the reporter BrM cells, macrometastatic lesions are isolated for flow cytometry analyses. The GFP expression in tdTomato<sup>+</sup> BrM cells *in vivo* is quantified and compared to the IFN $\beta$ -treated cells. The percentage of GFP<sup>+</sup> cells and mean fluorescent intensity (MFI) are quantified in tdTomato<sup>+</sup> BrM cells. Data are from merged samples from 2 biologically independent experiments. Source data are provided as a Source Data file.

**i**, Representative images of IFN $\beta$ -treated IFIT1-GFP expressing E0771-BrM reporter cells observed by IVM. Scale bar, 100  $\mu$ m. Representative data shown from 3 biologically independent experiments.



### Supplementary Fig. 3 Type I IFN activation in astrocytes promotes brain metastasis.

**a**, Increased IFN response genes in mouse astrocytes cocultured with breast cancer A7C11-BrM *in vitro*. The expression of IFN response genes are measured by RT-PCR. Al, cultured alone astrocytes; Co, cocultured astrocytes. Representative data shown from 3 biologically independent experiments for each BrM cell.

**b,c**, Human astrocytes are treated with conditioned media (CM) and type I IFN response genes are detected by RT-PCR. **(b)** CM are collected from astrocyte-BrM coculture (Co) or cultured alone (AL). Representative data shown from 3 biologically independent experiments for each BrM cell. **(c)** CM are pretreated with either neutralizing antibody against IFN $\beta$  or the matched IgG control antibody (Ctrl). Representative data shown from 3 biologically independent experiments for each BrM cell.

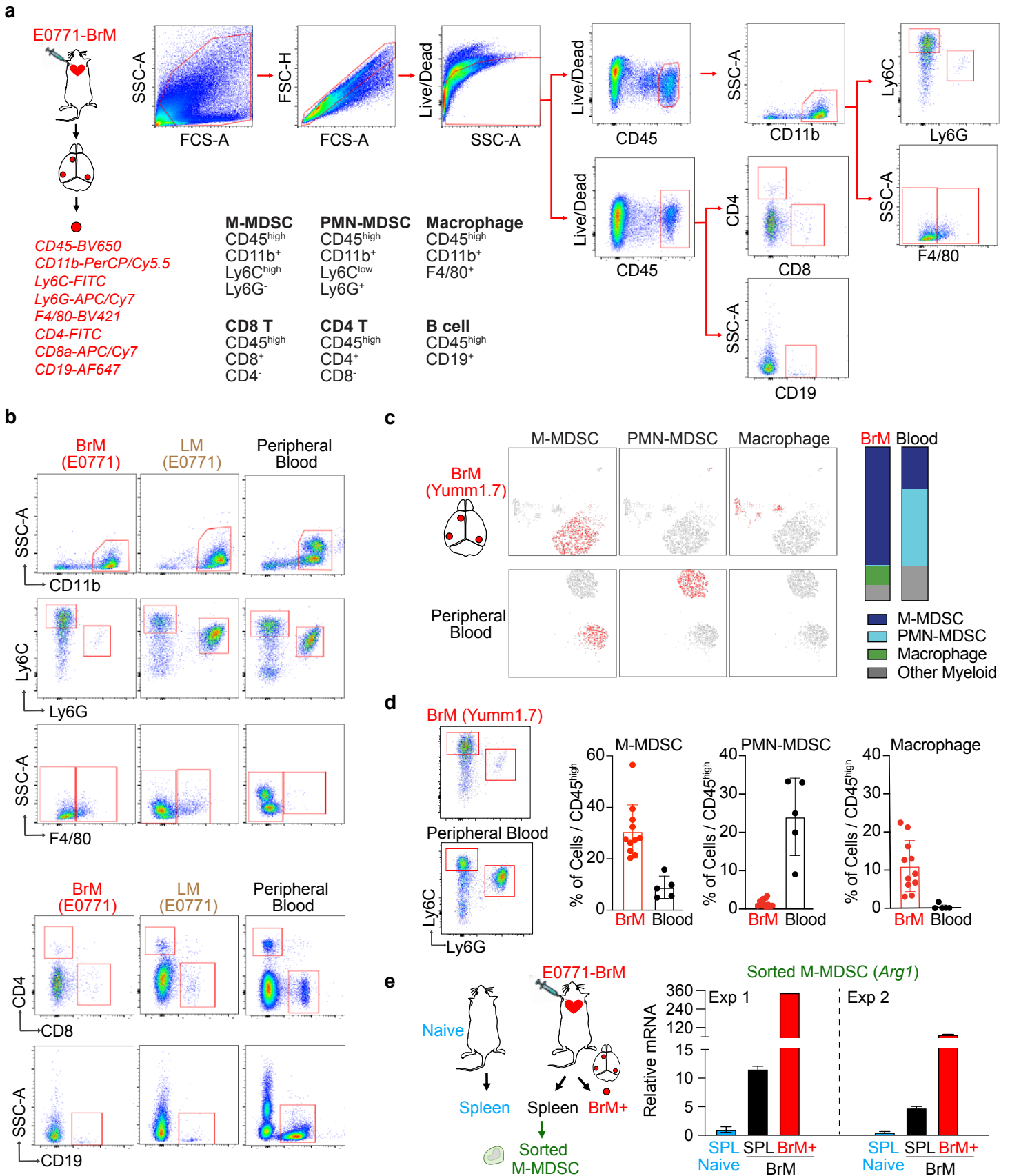
**d,e**, Characterization of astrocytes from astrocyte-specific IFNAR1 knock out mice. *Cre-*, *Gfap-Cre<sup>-/-</sup>*; *Ifnar1<sup>fl/fl</sup>* mice; *Cre+*, *Gfap-Cre<sup>+/-</sup>*; *Ifnar1<sup>fl/fl</sup>* mice. **(d)** Growth of primary cultured astrocytes.  $n = 10$  samples in each condition. 2 biologically independent experiments were conducted. Data are presented as mean  $\pm$  S.D. Source data are provided as a Source Data file. **(e)** Viabilities and numbers of astrocytes isolated from mouse brain. 1/4 of cortex of each mouse are collected. Data are from merged samples from 2 biologically independent experiments presented as mean  $\pm$  S.D. (*Cre-*:  $n = 6$ , *Cre+*:  $n = 5$ ). Source data are provided as a Source Data file.

**f**, RT-PCR results of the type I IFN response gene, *Isg15*, in the astrocytes coculture with BrM cells *in vitro*. Primary cultured astrocytes are isolated from transgenic mice. *Cre-*, *Gfap-Cre<sup>-/-</sup>*; *Ifnar1<sup>fl/fl</sup>* mice; *Cre+*, *Gfap-Cre<sup>+/-</sup>*; *Ifnar1<sup>fl/fl</sup>* mice. Representative data shown from 3 biologically independent experiments for each BrM cell. Source data are provided as a Source Data file.

**g**, Correlation between the total photon flux from IVIS imaging and immunofluorescence analyses of the brain metastatic regions in 1/10 of whole brain sections. Brain metastatic regions are defined based on tdTomato staining. Representative images of single metastatic cells, micrometastasis and macrometastasis. Bar = 400 $\mu$ m. Representative data shown from 2 biologically independent experiments. Source data are provided as a Source Data file.

**h**, E0771-BrM cells are treated with various doses of recombinant mouse IFN $\beta$  and cell growth is quantified by BLI after 72 hours. 2 biologically independent experiments were performed with similar results ( $n = 10$  per concentration in each experiment). Data are presented as mean  $\pm$  S.D. *P* values are the results from paired two-tailed t test. Source data are provided as a Source Data file.





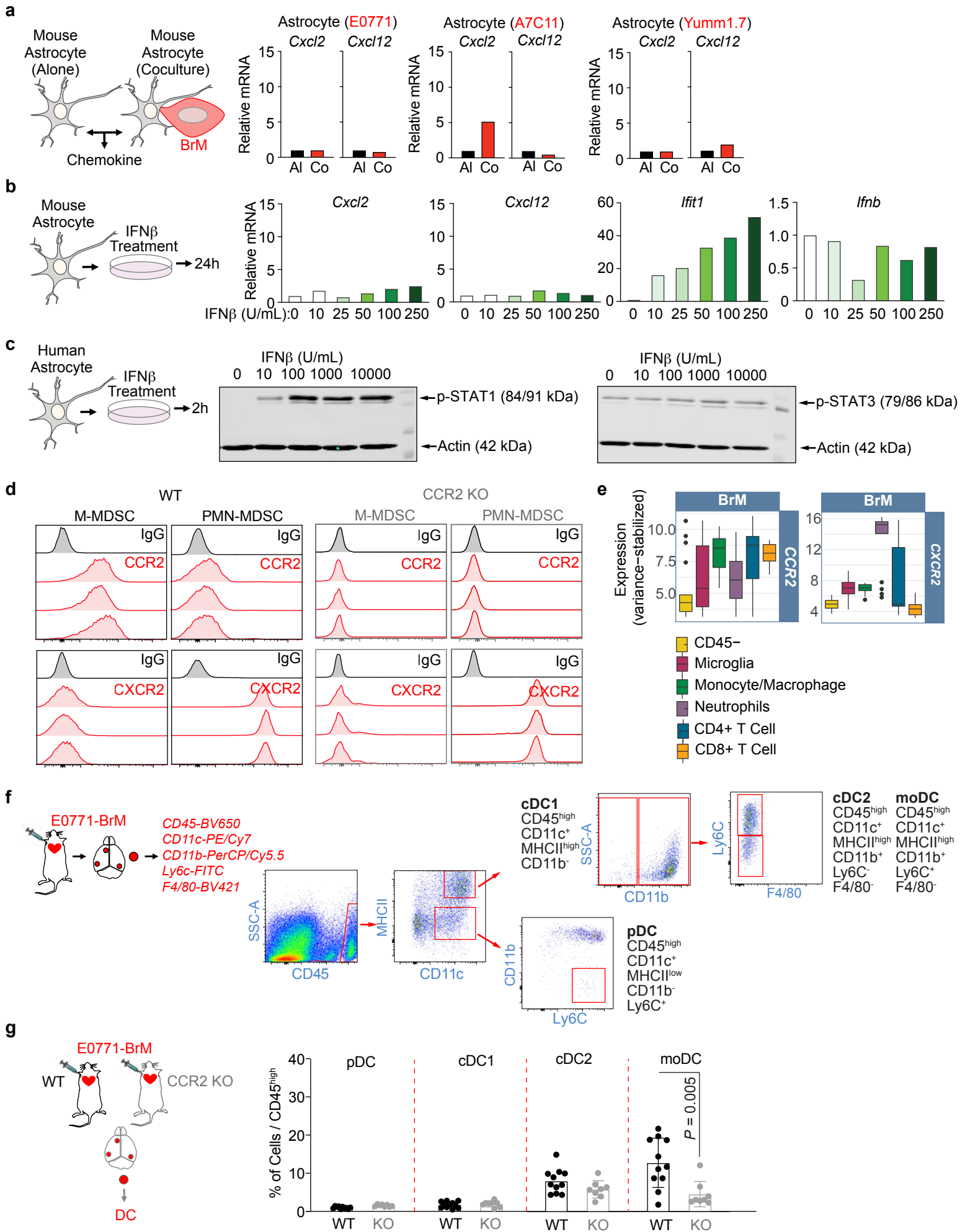
#### **Supplementary Fig. 4 Immune cell profiles in metastatic lesions.**

**a**, Representative flow cytometry profiles of immune cells in one brain metastatic lesions established by E0771-BrM cells. M-MDSC, monocytic myeloid-derived suppressor cells; PMN-MDSC, polymorphonuclear MDSC; CD8 T, CD8 T lymphocyte; CD4 T, CD4 T lymphocyte; B cell, B lymphocyte.

**b**, E0771-BrM cell are injected into experimental mice to establish brain or lung metastasis. Representative dot plots of myeloid and lymphoid subpopulations of one brain metastatic lesion, merged lung metastatic samples from one experimental mouse and the blood sample from one brain metastasis-bearing mouse.

**c,d**, Yumm1.7-BrM cells are injected into experimental mice to establish brain metastasis. **(c)** Representative tSNE files of myeloid subpopulations of one brain metastatic lesion and the blood sample from one brain metastasis-bearing mouse. Bar graphs indicate the proportions of indicated immune subpopulations in CD45<sup>high</sup> CD11b<sup>+</sup> myeloid cells (total is 100%). **(d)** Dot plots show the representative flow profiles of MDSCs. Bar graphs show the percentages of myeloid subpopulations out of CD45<sup>high</sup> cells. Data are from merged samples of 2 biologically independent experiments and presented as mean  $\pm$  S.D. (BrM: n = 11, blood: n = 5). Source data are provided as a Source Data file.

**e**, RT-PCR results of immunosuppressive gene, *Arg1*, in the sorted M-MDSC from naïve and brain metastasis-bearing mice. SPL, spleen; BrM, brain metastatic lesions. Representative data shown from 3 biologically independent experiments.



### Supplementary Fig. 5 CCR2 mediates M-MDSC migration in brain metastasis.

**a**, RT-PCR results of the expression of chemokines in astrocytes cocultured with BrM cells *in vitro*. Mouse astrocytes are cocultured with breast cancer E0771-BrM, A7C11-BrM and melanoma Yumm1.7-BrM. The expressions of CXCL2 and CXCL12 are measured. AI, cultured alone astrocytes; Co, cocultured astrocytes. Representative data shown from 3 biologically independent experiments for each BrM cell.

**b**, RT-PCR results of the expression of chemokines, IFIT1 and IFN $\beta$ -treated astrocytes *in vitro*. Mouse astrocytes cells are treated with various concentrations of IFN $\beta$  and the gene expressions. *Ifit* is used as a positive control for IFN responses. Representative data shown from 3 biologically independent experiments.

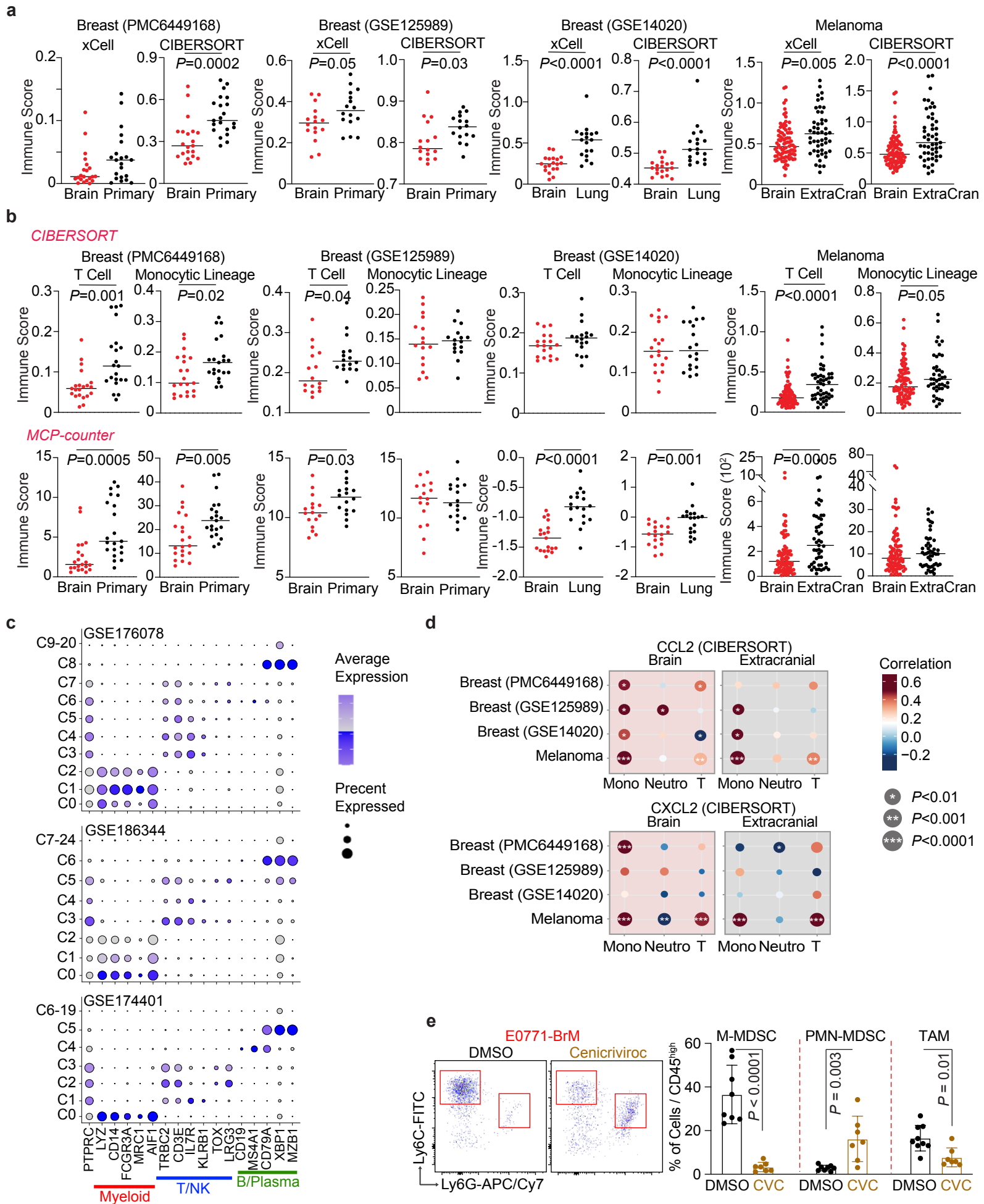
**c**, Western blot results of phosphorylated STAT1 (p-STAT1) and phosphorylated STAT3 (p-STAT3) in IFN $\beta$ -treated astrocytes *in vitro*. 2 biologically independent experiments were conducted with similar results. Source data are provided as a Source Data file.

**d**, CCR2 expression in monocytic myeloid cells. E0771-BrM cells are injected into wild type (WT) or CCR2 knock out (KO) mice. Spleens are harvested and the chemokine receptors in myeloid subpopulations are detected by flow cytometry. M-MDSC, monocytic myeloid-derived suppressor cells; PMN-MDSC, polymorphonuclear MDSC. Representative data from 3 biologically independent experiments.

**e**, Bioinformatic analyses of chemokine receptors in myeloid immune subpopulations from 89 clinical brain metastasis samples (<https://joycelab.shinyapps.io/braintime/>).

**f**, Representative flow cytometry profiles of dendritic cells (DC) in one brain metastatic lesions established by E0771-BrM cells. pDC, plasmacytoid DC; cDC1, conventional type 1 DC; cDC2, conventional type 2 DC; moDC, monocyte-derived DC.

**g**, Percentages of DC subpopulations out of CD45<sup>high</sup> cells in brain metastatic lesions. Data are merged samples from n = 3 biologically independent experiments and presented as mean  $\pm$  S.D. *P* values are the results from unpaired two-tailed t test. (pDC: n = 10 WT, n = 8 CCR2KO. cDC1, cDC2, moDC: n = 11 WT, n = 8 CCR2KO). Source data are provided as a Source Data file.



**Supplementary Fig. 6 Correlation of CCL2 expression and monocytic myeloid immune cells score in clinical brain metastasis samples.**

**a,b**, Bioinformatic analyses of immune cells in clinical samples. We include breast cancer PMC6449168 and GSE125989 (paired primary and brain metastatic datasets), GSE14020 (unpaired brain and lung metastatic samples), and melanoma EGAD00001005046 (paired and unpaired brain metastatic and extracranial tumors) in the analyses. **(a)** Total immune scores in brain metastatic (Brain), extracranial (ExtraCran) lung metastatic (Lung) or primary tumors analyzed by CIBERSORT and xCell. Source data are provided as a Source Data file. **(b)** Immune scores of and T cell and monocytic lineage subpopulations are analyzed by CIBERSORT and MCP-counter. *P* values are the results from unpaired two-tailed t test. PMC6449168: n = 22. GSE125989: n = 16. GSE14020: n = 19 Brain, n = 18 Lung. MD Melanoma: n = 88 Brain, n = 49 Extracranial. Source data are provided as a Source Data file.

**c**, Dot plots of representative genes used for single cell RNAseq cluster annotations in each dataset. Pan-immune cells (*PTPRC*). Myeloids (*LYZ*, *CD14*, *GCGR3A*, *MRC1*, *AIF1*). T/NK (*TRBC2*, *CD3E*, *IL7R*, *KLRB1*, *TOX*, *LAG3*). B/Plasma cells (*CD19*, *MS4A1*, *CD79A*, *XBP1*, *MZB1*).

**d**, Correlations of CCL2 or CXCL12 expression and immune scores of monocytic lineage (Mono), neutrophil (Neutro) and T lymphoid (T) subpopulations in brain metastatic and extracranial tumors analyzed by CIBERSORT. Breast cancer PMC6449168 and GSE125989 (paired primary and brain metastatic datasets), GSE14020 (unpaired brain and lung metastatic samples), and melanoma EGAD00001005046 (paired and unpaired brain metastatic and extracranial tumors) are analyzed. *P* values are the results from unpaired two-tailed t test. Source data are provided as a Source Data file.

**e**, CCR2/CCR5 antagonist treatment blocks monocytic myeloid cell infiltration. Bar graphs show the percentages of myeloid subpopulations out of CD45<sup>high</sup> cells. M-MDSC, monocytic myeloid-derived suppressor cells; PMN-MDSC, polymorphonuclear MDSC. Data are merged samples from 2 biologically independent experiments and presented as mean ± S.D (M-MDSC, PMN-MDSC: n = 8 DMSO, n = 7 CEN. TAM: n = 9 DMSO, n = 7 CEN). *P* values are the results from unpaired two-tailed t test. Source data are provided as a Source Data file.

Supplementary Table 1: List of Primers used for RT-PCR (mouse genes)

Gene	Sense (5'-3')	Antisense (3'-5')	Acession No.
<i>Ccl2 (set1)</i>	CCCACTCACCTGCTGCTACT	TCTGGACCCATTCTTCTTG	NM_011333
<i>Ccl2 (set2)</i>	TCCCAATGAGTAGGCTGGAG	GCTGAAGACCTTAGGGCAGA	NM_011333
<i>lfng</i>	GCCACGGCACAGTCATTGA	TGCTGATGGCCTGATTGTCTT	NM_008337
<i>Cxcl12</i>	TGCATCAGTGACGGTAAACCA	CACAGTTTGGAGTGTTGAGGAT	NM_001012477
<i>Actb</i>	GGCTGTATTCCCCTCCATCG	CCAGTTGGTAACAATGCCATGT	NM_007393
<i>Cxcl2</i>	AGTGAAGTGCCTGTCAATG	CTTCAGGGTCAAGGCAAACCT	NC_000071.6
<i>Cxcl3</i>	CTCCAGACTCCAGCCACACT	GTCACCGTCAAGCTCTGGAT	NC_000071
<i>Cxcl5</i>	TGCCCTACGGTGGAAGTCATA	TGCATTCCGCTTAGCTTTCTTT	NM_009141
<i>Ifit1</i>	CTGAGATGTCACCTCACATGGAA	GTGCATCCCCAATGGGTTCT	NM_008331
<i>Ifnar1</i>	GGTGGTTCTGTCTCGGTGTT	GCCAGCTCCTCCAGTTAGTG	NM_010508
<i>Ifnb1</i>	CAGCTCCAAGAAAGGACGAAC	GGCAGTGTAACCTTTCTGCAT	NM_010510
<i>Isg15</i>	GGTGTCCGTGACTAACTCCAT	TGAAAAGGGTAAGACCGTCCT	NM_015783
<i>Usp18</i>	TTGGGCTCCTGAGGAAACC	CGATGTTGTGTAACCAACCAGA	NM_011909
<i>B2m</i>	GGCCCATCTTGCATTCTAGGG	CAGGCAACGGCTCTATATTGAAG	NM_009735

**Supplementary Table 2: List of Primers used for RT-PCR (human genes)**

<b>Gene</b>	<b>Sense (5'-3')</b>	<b>Antisense (3'-5')</b>	<b>Acession No.</b>
<i>ACTB</i>	AAACTGGAACGGTGAAGGTG	AGAGAAGTGGGGTGGCTTTT	NM_001101.3
<i>B2M</i>	CTCCGTGGCCTTAGCTGTG	TTTGGAGTACGCTGGATAGCCT	NM_004048
<i>CCL2</i>	CCCCAGTCACCTGCTGTTAT	TCCTGAACCCACTTCTGCTT	NM_002982
<i>CXCL12</i>	CTACAGATGCCCATGCCGAT	CAGCCGGGCTACAATCTGAA	NM_001178134
<i>CXCL2</i>	GAAAGCTTGTCTCAACCCCG	GTTGGATTTGCCATTTTTTCAGCA	NC_000004
<i>IFIT1</i>	GCCCAGACTTACCTGGACAA	GGTTTTCAGGGTCCACTTCA	NM_001270930
<i>IFNA1</i>	CTGTGTGATGCAGGAGGAGA	GATCTCATGATTTCTGCTCTGA	NM_024013
<i>IFNAR1</i>	AACAGGAGCGATGAGTCTGTC	TGCGAAATGGTGAAATGAGTCA	NM_000629
<i>IFNB1</i>	ATGACCAACAAGTGTCTCCTCC	GGAATCCAAGCAAGTTGTAGCTC	NM_002176
<i>IFNG1</i>	TCGGTAACTGACTTGAATGTCCA	TCGCTTCCCTGTTTTAGCTGC	NM_000619
<i>ISG15</i>	TGTCGGTGTGAGAGCTGAAG	GCCCTTGTTATTCCTCACCA	NM_005101
<i>USP18</i>	CAGACCCTGACAATCCACCT	AGCTCATACTGCCCTCCAGA	NM_017414



**Supplementary Table 3: List of Antibodies**

	<b>Company</b>	<b>Catalog#</b>	<b>Clone</b>	<b>Application</b>
CD16/CD32	BD Biosciences	553142	2.4G2	Fc blocking antibody
CD45-BV650	BD Biosciences	563410	30-F11	Flow cytometry
Ly6G-APC/Cy7	BioLegend	127623	1A8	Flow cytometry
CD11b-PerCP/Cy5.5	BioLegend	101227	M1/70	Flow cytometry
Ly6C-FITC	BD Biosciences	561085	AI-21	Flow cytometry
F4/80-BV421	BioLegend	123131	BM8	Flow cytometry
CD11c-PE/Cy7	BioLegend	117318	N418	Flow Cytometry
I-A/I-E (MHCII)-APC	BioLegend	117613	M5/114.15.2	Flow Cytometry
CD4-FITC	BioLegend	100405	GK1.5	Flow cytometry
CD8a-APC/Cy7	BioLegend	100713	53-6.7	Flow cytometry
CD19-AF647	BioLegend	115522	6D5	Flow cytometry
CCR2-AF647	BioLegend	150603	SA203G11	Flow cytometry
CXCR2-AF647	BioLegend	149305	SA044G4	Flow cytometry
Rat IgG2b-AF647	BioLegend	400626	RTK4530	Isotype control for CCR2
Rat IgG2a-AF647	BioLegend	400526	RTK2758	Isotype control for CXCR2
ACSA-2-APC	Miltenyi	130-117-535	IH3-18A3	Flow cytometry
Zombie Aqua	Thermo Scientific	L34957	n/a	Flow cytometry
Zombie Aqua	BioLegend	423102	n/a	Flow cytometry
tdTomato	OriGene	AB8181	Polyclonal	IF
Alexa Fluor 594 donkey anti-goat	Thermo Fisher	A-11058	n/a	IF
Phosho-STAT1	Cell Signaling Tech	9167	58D6	Western Blot
Phospho-STAT3	Cell Signaling Tech	9145	D3A7	Western Blot
STAT1	Cell Signaling Tech	9175	42H3	Western Blot
STAT3	Cell Signaling Tech	9132	n/a	Western Blot
Beta Actin	Santa Cruz Biotech	sc-47778	C4	Western Blot
IRDye 800CW goat anti-rabbit	Li-Cor	926-32211	n/a	Western Blot
IRDye 680RD goat anti-mouse	Li-Cor	926-68070	n/a	Western Blot
CCL2 (MCP1)	Novus Biologicals	AF-479-SP	Polyclonal	Neutralizing Ab
IFNAR1	Abcam	ab10739	Polyclonal	Neutralizing Ab
IFN $\beta$	R&D Systems	21400-1	Polyclonal	Neutralizing Ab
Mouse IgG1	R&D Systems	MAB002	11711	Isotype control for IFN $\beta$
Normal Goat IgG	Novus Biologicals	AB-108-C	Polyclonal	Isotype control for CCL2 and IFNAR1

**Supplementary Table 4: List of Primers used for genotyping**

<b>Strain</b>	<b>Sense (5'-3')</b>	<b>Antisense (3'-5')</b>
B6.Cg-Tg(Gfap-cre)77.6Mvs/2J	TTAATCCATATTGGCAGAACGAAAACG	CAGGCTAAGTGCCTTCTCTACA
B6(Cg)-Ifnar1 <sup>tm1.1Ees</sup> /J	CCGTTTATTCCATTCTACACAGGTAAGA	CTGTTTCCCATGGATAACTTCGTA