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#### Supplementary Fig. 1: Comparison of Met-substitute incorporation rates in vitro.

MetRS\*-expressing 293T cells were grown for 8 h in Met-free medium supplemented with 4 mM L-methionine-methyl-<sup>13</sup>C,d<sub>3</sub> (Met+4), Aha or Anl, and processed for shotgun proteomics and DDA single shot analysis without selective enrichment. **a)** Average number of identified peptides with methionine-substitutions (n = 3, mean  $\pm$  SD). Cells grown in 4 mM non-isotopically labeled methionine were used to evaluate false discovery rates, and WT 293T cells incubated in Anl-supplemented media to evaluate unspecific incorporation without MetRS\*. **b)** Median intensity ratios of peptides with Met-substitution and all modified and non-modified peptides with the corresponding sequence.



Supplementary Fig. 2: Technical reproducibility of MetRS\*-based cell-selective proteomics experiments, related to Fig. 1. a) Data completeness and b) precursor coefficients of variation (CVs) of MetRS\* and Ctrl samples processed with different Anl enrichment workflows (see Fig. 1a). c) Data completeness, d) precursor CVs, and e) intensity ratios of proteins identified in both MetRS\* and Ctrl samples after processing with our enrichment workflow and MS analysis by data-dependent acquisition (DDA) or data-independent acquisition (DIA).



Supplementary Fig. 3: Specifically enriched MetRS\* 8661 PDAC cell-derived proteins with different enrichment methods, related to Fig. 1.

Exclusively identified and overlap of specifically enriched protein groups with DST-enrichment, DBCO-enrichment, and our improved enrichment workflow.



### Supplementary Fig. 4: Technical reproducibility of enrichment specificity controls.

MetRS\*-expressing (n = 3) or wild-type (Ctrl) (n = 10) 8661 PDAC cells ( $1 \times 10^7$  cells per sample) were grown for 8 h in Met-depleted medium supplemented with 4 mM Anl, and processed with our Anl enrichment workflow. **a)** Sum of identified precursor intensities per sample. **b)** Data completeness of identified protein groups in MetRS\* and Ctrl samples. **c)** Protein intensity ratios between MetRS\* and Ctrl samples analyzed using all measured Ctrl replicates or subsets of 3 Ctrl replicates. **d)** Protein intensity ratios between MetRS\* and Ctrl samples seperated by the number of missing values in Ctrl samples.



Supplementary Fig 5: Flow cytometry analysis of MetRS\*-eGFP 8661 cells, related to Fig. 2.

a) Gating strategy for MetRS\* eGFP co-expressing 8661 PDAC cells and evaluation of eGFP-expression before cell transplantation. b) Analysis of dissociated tumor cells after orthotopic MetRS\*-eGFP 8661 PDAC cell transplantation, tumor growth and Anl labeling (n = 3, biological replicates).



Supplementary Fig 6: Principal component analysis (PCA) of cell-selective proteomics samples from BMMs and PDAC cells in co-culture and isolation *in vitro*, related to Fig. 3.

PCA score plots of the first two principle components of all replicates from global proteome and secretome experiments after filtering for specifically enriched protein groups.

#### PDAC secretomes









ECM regulators



Supplementary Fig 7: Heatmaps of cell-selectively secreted proteins in co-culture experiments, related to Fig. 3. PDAC- and BMM-released proteins with cytokine function or ECM regulator function (according to Naba *et al.*<sup>75</sup>) and significant abundance differences between PDAC subtypes and culture conditions (ANOVA, FDR = 0.05, S0 = 0.1).

#### **BMM** secretomes



Supplementary Fig. 8: Immunophenotyping of PDAC tumors by flow cytometry, related to Fig. 4.

Cell numbers indicated as % of CD45+ cells. P-values were determined by a two-sided Welch's t-test (8442: n = 3, 8661: n = 5, 8513: n = 3, 9091: n = 4; biological replicates) between classical and mesenchymal tumors.



## Supplementary Fig. 9: PCA of PDAC tumor-bearing mouse serum samples, related to Fig. 6.

PCA score plots of the first two principle components of all replicates including WT (Ctrl) PDAC samples before filtering MetRS\*-samples for specifically enriched protein groups.