

1 **Supplementary Material for**

2 **Purification and characterisation of heparin-like sulfated polysaccharides with**
3 **potent anti-SARS-CoV2 activity from snail mucus of *Achatina fulica***

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19 **Table S1.** ^{13}C NMR chemical shifts for acharan sulfate from snail mucus (AS-GAG). Carbons labeled as I and G refer to Iduronic
20 acid (IdoA) and N-acetyl glucosamine (GlcNAc) units, respectively.

Sugar residues	Chemical Shifts / ppm							
	1	2	3	4	5	6	7	8
I; $\rightarrow 4) - \alpha - \text{IdoA} \rightarrow 1$	99.233	73.281	63.721	69.743	66.397	174.801	–	–
G; $\rightarrow 4) - \alpha - \text{GlcNAc} \rightarrow 1$	93.927	53.680	68.573	77.250	71.220	59.679	174.801	22.217

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25 **Table S2.** Determination of anti-SARS-ACE2 binding for the unfractionated
 26 polysaccharide inhibitors.

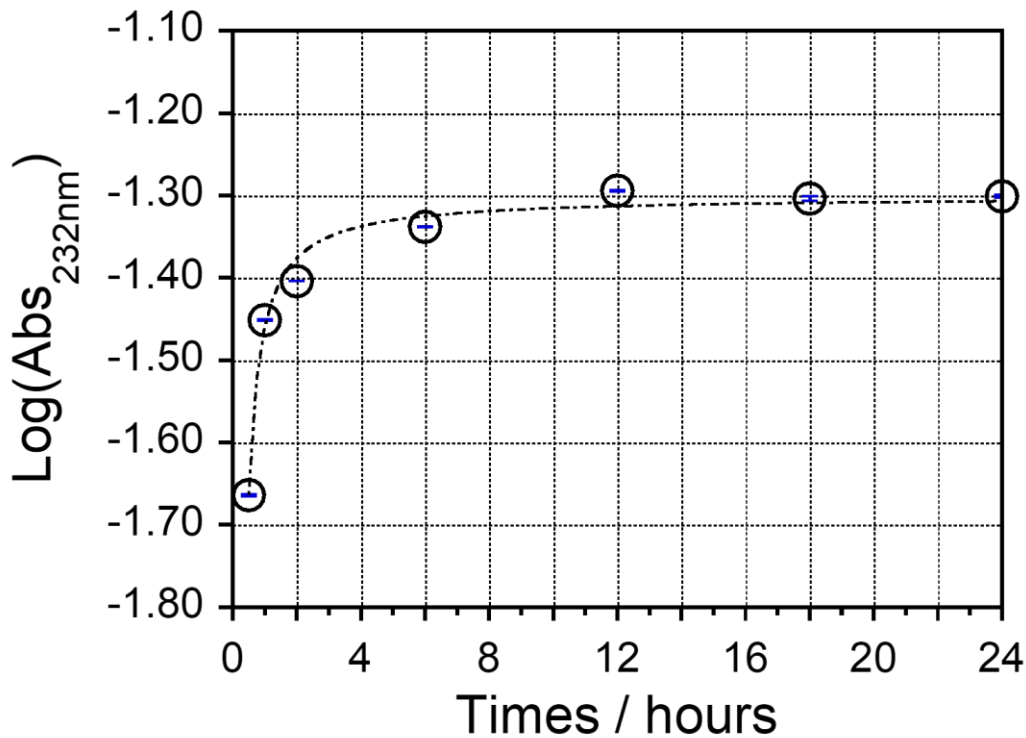
Inhibitors	IC ₅₀	IC ₅₀ 95% CI	Ref.
AS-GAGs ^a	28.4 µg mL ⁻¹	4.33 – 5.29 µg mL ⁻¹	
Enoxaparin ^a	91.5 µg mL ⁻¹	5.20 – 5.65 µg mL ⁻¹	
Pentosan polysulfate ^a	57.1 µg mL ⁻¹	4.31 – 4.84 µg mL ⁻¹	
Heparan sulfate ^a	132.4 µg mL ⁻¹	2.11 – 2.14 µg mL ⁻¹	
Heparin	0.12 µg mL ⁻¹	0.09 – 0.15 µg mL ⁻¹	[21]
Heparan sulfate	62 µg mL ⁻¹	>15 µg mL ⁻¹	[21]
LMWH	5 – 12 µg mL ⁻¹	–	[22]
UFH	1 – 4 µg mL ⁻¹	–	[22]
	5.99 µg L ⁻¹	2.90 – 11.96 µg L ⁻¹	[25]
Enoxaparin	1.08 mg L ⁻¹	247 – 4164 µg L ⁻¹	[25]

27 ^a Values from this work.

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30 **Figure S1.**

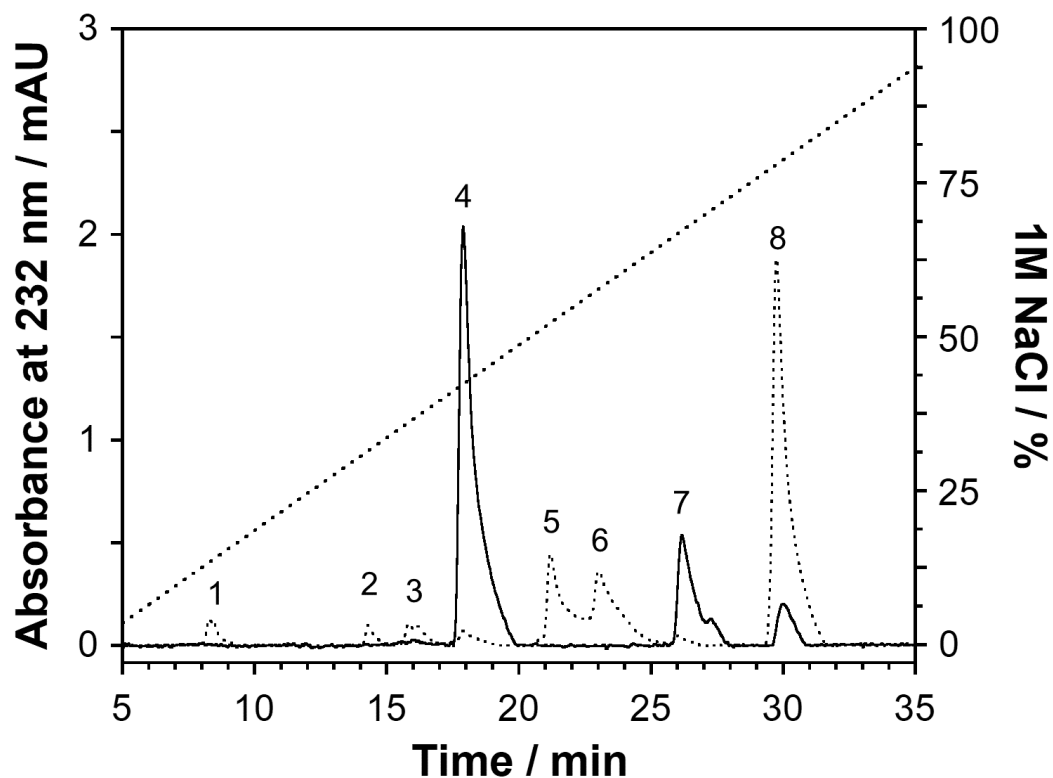


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32 **Fig. S1.** Effect of hydrolysis time on the relative change ($\text{Log}(\text{Abs}_{232\text{nm}})$) of the
33 unsaturated disaccharide content of mGAG-F3 enzymatic hydrolysis by
34 treatment with a combination of heparin lyase I (EC 4.2.2.7), II
35 (Heparitinase II), and III (EC 4.2.2.8). The digested mixture was in 0.3 mL
36 of 50 mM Tris-HCl buffer (pH 7.2) and 10 mM CaCl₂ at 37°C for 24 hrs,
37 after which the reaction samples were heated in a boiling water for 3
38 minutes. Results are reported as mean \pm SD. Error bar denotes standard
39 deviation.

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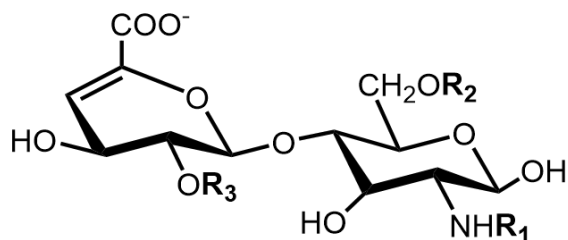
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44 **Fig. S2.** SAX-HPLC chromatogram of mGAG-F3 (solid line) and heparin (dashed
 45 line) disaccharides by treatment with a combination of heparin lyase I (EC
 46 4.2.2.7), II (Heparitinase II), and III (EC 4.2.2.8), eluting with a linear
 47 gradient of 0.1 – 1.0 M NaCl (dotted line) at a flow rate of 1.0 mL/min. The
 48 elutions were monitored by absorbance at 232 nm for detecting the
 49 unsaturated non-reducing ends of the disaccharides. The standard
 50 disaccharide positions are indicated by number labels; 1: Δ UA-GlcNAc, 2:
 51 Δ UA-GlcNSO₃, 3: Δ UA-GlcNAc(6S), 4: Δ UA(2S)-GlcNAc, 5: Δ UA-
 52 GlcNSO₃(6S), 6: Δ UA(2S)-GlcNSO₃, 7: Δ UA(2S)-GlcNAc(6S), 8: Δ UA(2S)-
 53 GlcNSO₃(6S).

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56 **Figure S3.**



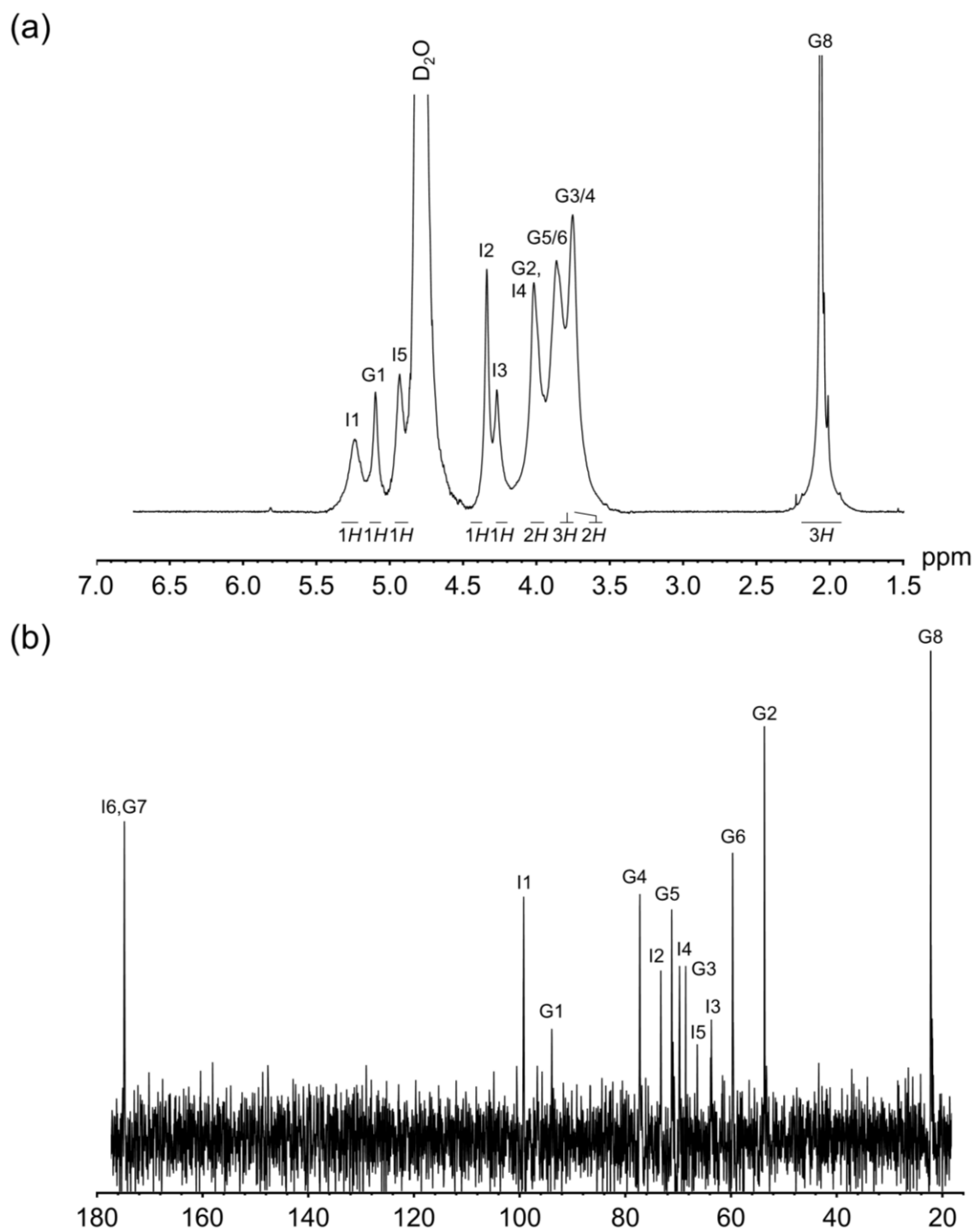
No.	Disaccharide	R1	R2	R3
1	Δ UA-GlcNAc	Ac	H	H
2	Δ UA-GlcNSO ₃	SO ₃	H	H
3	Δ UA-GlcNAc(6S)	Ac	SO ₃	H
4	Δ UA(2S)-GlcNAc	Ac	H	SO ₃
5	Δ UA-GlcNSO ₃ (6S)	SO ₃	SO ₃	H
6	Δ UA(2S)-GlcNSO ₃	SO ₃	H	SO ₃
7	Δ UA(2S)-GlcNAc(6S)	Ac	SO ₃	SO ₃
8	Δ UA(2S)-GlcNSO ₃ (6S)	SO ₃	SO ₃	SO ₃

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58 **Fig. S3.** 2D Structure of the commonly occurring unsaturated disaccharides
59 derived from digestion with heparinase enzymes. Abbreviations: Ac,
60 acetyl; Δ UA, Δ (4,5) unsaturated hexuronic acid; GlcNAc, N-acetylated
61 glucosamine; GlcNSO₃, N-sulfated glucosamine; H, hydrogen; 2S and 6S, 2-
62 O-sulfate and 6-O-sulfate, respectively.

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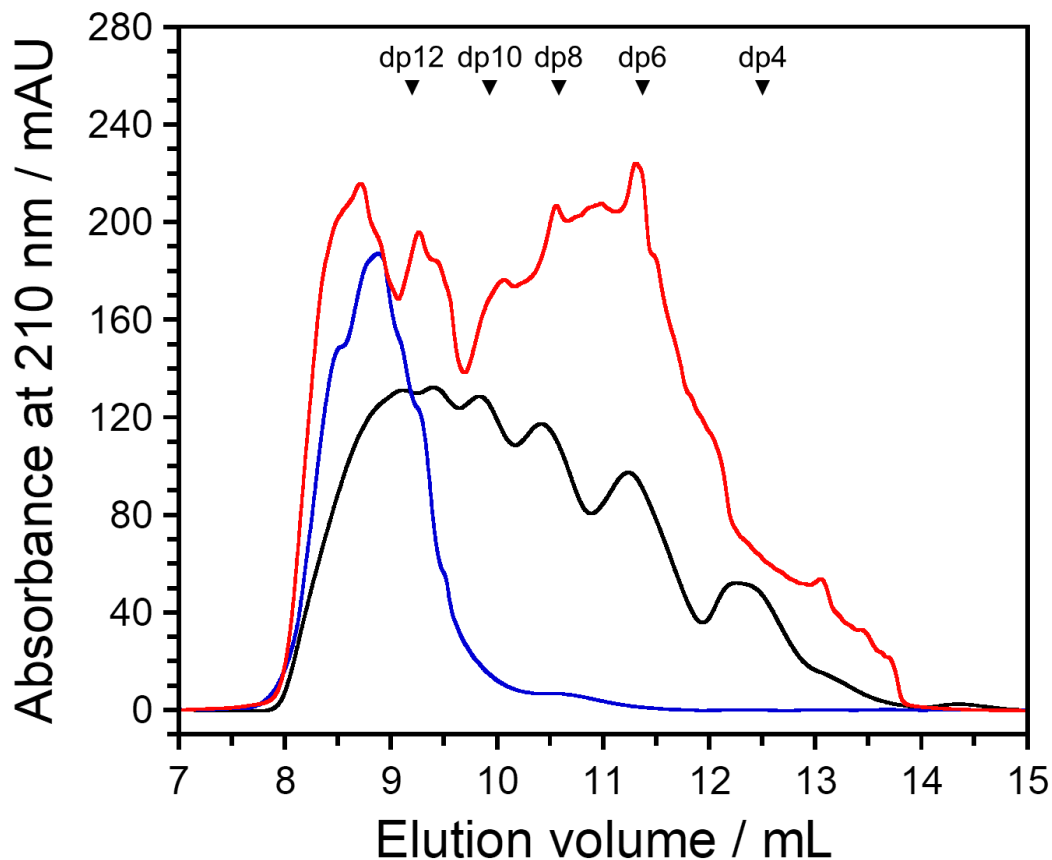
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66 **Fig. S4.** (a) ^1H - and (b) ^{13}C -NMR spectra of the AS-GAG from the mucus of *A. fulica*.
 67 The assignment was based on comparison with the previously described
 68 acharan sulfate. The peaks are labeled as I or G for Iduronic acid (IdoA)
 69 and *N*-acetyl glucosamine (GlcNAc) units, respectively. The number after
 70 these letters indicate the positions of ^1H and ^{13}C .

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74 **Fig. S5.** Size exclusion chromatography analysis of the partial digest product of
 75 sulfated polysaccharide; mGAG-F3 between pre- (blue) and post- (red)
 76 depolymerization, in comparison to LMWH (black). The mGAG-F3 was
 77 depolymerized by partial digestion with a combination of heparin lyase I
 78 (EC 4.2.2.7), II (Heparitinase II), and III (EC 4.2.2.8). The digests were
 79 chromatographed on a Superdex 30 Increase 30/100 column using 0.1 M
 80 ammonium acetate (pH 7.0) as the eluent at 1.5 CV at flow rate 0.5 ml/min.
 81 The centers of tetrasaccharide, hexasaccharide, octasaccharide,
 82 deca-saccharide, and deca-saccharide peaks are marked with arrows. A
 83 calibration curve was plotted by the logarithm of molecular weights versus
 84 retention time in each standard position. The molecular weight of mGAG-
 85 F3 fraction was estimated by compared with the calibration curve.

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